

TOPLEY AND WILSON'S PRINCIPLES OF  
BACTERIOLOGY AND IMMUNITY

# TOPLEY AND WILSON'S PRINCIPLES OF BACTERIOLOGY AND IMMUNITY

THIRD EDITION

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IN TWO VOLUMES

*VOLUME I*



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To  
K. T. AND J. W.



## PREFACE TO THE THIRD EDITION

APART from the war this edition has been prepared under the shadow of a double misfortune. Early in 1941 Professor Topley accepted the post of Secretary of the Agricultural Research Council and thereby took a step that rendered his further participation in this book impossible. To replace him I was fortunate in enlisting the co-operation of our former colleague Professor Miles. Together we began the arduous task of revision. Our work, however, had not progressed far before the second blow fell. Quite suddenly in January 1944 Topley died. The effect of this second misfortune was almost as serious as the first. Although Topley could have made no direct contribution to the text his criticism and advice would have been constantly available and he would have helped us to maintain that uniformity of presentation for which he and I had always striven. On Miles, in particular, the burden weighed heavily since in taking over those parts of the book for which Topley had previously been responsible he was deprived of counsel that would doubtless have proved invaluable to him.

There is no call to write a long preface. We have endeavoured not merely to bring the book up to date but to present the new additions to our knowledge in a manner worthy of their importance. One chapter—that on Soil Microbiology—has been deleted but two new chapters on Chemotherapy and on the Bacteriology of Air have been added. For the sake of convenience we have divided the previous *Bacterium* chapter into three giving separate recognition to the genera *Shigella* and *Salmonella*. We have also removed the psittacosis-lymphogranuloma group of diseases from the other filtrable virus diseases with which they were associated and awarded them a chapter of their own. Except for these alterations the form of the book remains unchanged. In the first two editions we tried to ensure that scientific literature from all parts of the world was fairly represented but in the present edition we have suffered under a handicap imposed by the war which has seriously curtailed the inflow of journals from many parts of Europe as well as of course from Japan. This gap we shall look forward to filling in the future. Partly because of the necessarily one-sided picture we have been obliged to paint we have thought it wise to present our evidence in greater detail than might otherwise have been necessary and to be perhaps unduly cautious in drawing our conclusions. The bibliography has been much expanded so as to cater for the needs of those who use the book more as a work of reference than as a textbook. For the increased length of the new edition we tender our apologies. The war

has not been conducive to careful leisurely recapitulation, and our plea must be the paradoxical excuse that we have not had time to be more concise.

To those who have assisted us in various ways, we would express our thanks. We are particularly grateful to Dr N. W. Pine for his help with some chapters in Part I, to Professor A. Bradford Hill for his advice on Chapter 43 and to Professor S. P. Bedson, Lt Col R. F. Bridges, Dr A. Q. Wells, Dr Joan Taylor, Dr A. W. Stableforth, Dr R. Lovell, Miss Nancy Hayward, and many of our former helpers for information on particular problems. To Dr Stuart Mudd and his American colleagues, to Professor A. D. Gardner, Dr C. F. Robinow, Dr S. T. Cowan, Dr N. G. Heatley, Dr A. Pijper and the publishers of "Endeavour" we are indebted for a number of electron micrographs and photographs, to Professor J. R. Marreck for Fig. 32, to H. M. Stationery Office for permission to reproduce Figures 32, 34, 77, 79, 80 and 81 and to Miss Margaret Rees for the preparation of some new diagrams. We should also like to pay our tribute to the library staffs of the London School of Hygiene and Tropical Medicine, the Bureau of Hygiene, University College Hospital Medical School, and the Radcliffe Science Library, Oxford, for their unfailing courtesy and help in the tracing of numerous references.

G. S. W.

June, 1945

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# PART I

## GENERAL BACTERIOLOGY

### CHAPTER I

#### HISTORICAL OUTLINE

IN the study of any branch of science, an acquaintance with the historical development of knowledge is an important element in a clear understanding of our present conceptions. To the student of bacteriology such a basis is essential. It is almost true to say that the clue to the present position of bacteriology is the curious fact that there have been no bacteriologists. From Pasteur onwards the great majority of investigators have been more interested in what bacteria do than in what they are, and much more interested in the ways in which they interfere with man's health or pursuits than in the ways in which they function as autonomous living beings. The relations of bacteria to disease, to agriculture, and to various commercial processes have presented problems which pressed for solution, and, as a result, we have witnessed a reversal of the normal process. We have seen the development of an applied science of bacteriology, or rather its application along many divergent lines, without the provision of any general basis of purely scientific knowledge. The essential interlocking of pure and applied science has, of course, been in evidence here as elsewhere. The necessity for being able to recognize a bacterium, which has been shown to be of importance in some province of human affairs, or of determining the way in which its harmful or beneficial action is brought about, has led to an intensive study of many aspects of bacterial morphology and physiology, but, in general, it may be said that the study of bacteria themselves has been carried out *en passant*, that amount of knowledge being acquired or searched for, which would afford adequate data for the solution of some problem in applied bacteriology. Gradually the general structure of our knowledge has been added to, and gaps have been filled. Many of those who have started from some particular application have been led far afield by that desire for knowledge, altogether apart from its technical application, which is the essence of science itself. But this mode of construction has given to the general body of existing bacteriological knowledge a curious patchiness and indefiniteness which are puzzling to the student, and which must be realized and allowed for in any attempt to present the subject as a whole. There can be no question of any future reconstruction *ab initio*. The history of a science is largely a history of technique, and the foundations of bacteriological technique, which presents many peculiar difficulties, have been well and truly laid by those who have worked in this field since



the middle of the nineteenth century. The pure bacteriologist of the future will owe a lasting debt to those who have worked on the applied side, and his investigations will necessarily be based upon the knowledge gained by the medical or agricultural bacteriologist. The study of immunology, for instance, has supplied a body of facts, and an armoury of technical methods, which no bacteriologist can neglect, and which will inevitably give to future bacteriological research certain peculiarities of outlook and special methods of attack.

It is customary, in summarizing the history of bacteriology, at least in relation to medicine, to refer to the conception advanced by Fracastorius of Verona (1546), concerning a *contagium vivum* as the possible cause of infective disease, and to the views advanced by von Plenciz (1762) on the specificity of disease, based on a belief in its microbial origin. A concrete science is, however, seldom advanced to any considerable extent by arguments, however ingenious, which are propounded without appeal to experiment, or to wide and detailed observation, and the absence of all real progress until the middle of last century is sufficient evidence that the views of Fracastorius, von Plenciz and others have acquired their main significance from knowledge gathered by later generations rather than from their inherent fertility. The construction and use of the compound microscope was an essential prerequisite to the study of microbial forms, and the reported observation by Kircher (1659) of minute worms in the blood of plague patients forms, perhaps, the earliest attempt at direct microscopical observation in this field. It is, however, more than doubtful whether Kircher could have seen plague bacilli, or indeed any bacterial forms, with the apparatus which he had at his disposal. To van Leeuwenhoek (1683) must be ascribed the credit of placing the science of microbiology on the firm basis of direct observation (Dobell 1932). This Dutch maker of lenses developed an apparatus and technique (Cohen 1937) which enabled him to observe and describe various microbial forms with an accuracy and care which still serve as a model for all workers in this field. He observed, drew, and measured with considerable approximation to truth large numbers of minute living organisms, including bacterial and protozoal forms. It is perhaps somewhat surprising that this marked advance was not followed by further rapid progress in our knowledge of bacteria and their activities. Such progress was however, impossible without further developments in technique. The world of minute living things, opened to morphological study by van Leeuwenhoek, was seen to be peopled by a multitude of dissimilar forms whose interrelationships it was impossible to determine without preliminary isolation, and so far as bacteria were concerned, this isolation was not accomplished until the problem of artificial cultivation was solved, almost two hundred years later.

The real development of bacteriology as a subject of scientific study dates from the middle of the nineteenth century, and is the direct outcome of the work of Louis Pasteur (1822-95). Isolated observations of microbial parasites, by Brassi, Pollender, Davaine and others, have priority in particular instances, just as Schultze, Schroeder and Dusch and others initiated technical methods which Pasteur applied to his own researches. But it was Pasteur and his pupils who settled the fundamental questions at issue, and developed a technique which made possible the cultivation and study of bacteria.

Trained as a chemist, Pasteur was led to the study of microscopic organisms by his observations on the phenomena of fermentation. His early studies on the structure of the tartrates, and on molecular asymmetry, had led him to believe

that the property of optical activity, possessed by certain organic compounds was characteristic of substances synthesized by living things as contrasted with substances synthesized in the laboratory. It was known that small amounts of an optically active substance amyl alcohol, were formed during the fermentation of sugar, especially in association with the lactic fermentation. Since it was impossible to regard the molecule of amyl alcohol as derived from the molecule of sugar by any simple break down process he was led to the conclusion that the optically active molecule of the sugar was first broken down to relatively simple substances, which experience had shown to be without optical activity.



FIG 1—LOUIS PASTEUR (1822-1895)

and that from such inactive substances the optically active amyl alcohol was synthesized. For Pasteur this was evidence of the presence and activity of living things, and he therefore started on his study of fermentation with a strong *a priori* leaning towards the microbial theory of fermentation and away from the then dominant hypothesis of Liebig. He was prepared to adopt the theories already propounded by Cagniard Latour in 1836, and by Schwann in 1837, concerning the living nature of the yeast globules, which were always to be found in sugar solutions undergoing alcoholic fermentations and which had been described by van Leeuwenhoek in 1680.

Since, however, it was in the lactic fermentation that the production of amyl

alcohol had especially been noted, it was this reaction which Pasteur first selected for experimental study, though he had already made numerous observations on material from the vats of the breweries of Lille. He was probably influenced by the fact that the observations of van Helmholtz (1843) had already indicated that the alcoholic fermentation was due to the yeast itself or to some other organized material. Helmholtz had shown that the substance, whatever it might be, which was responsible for initiating alcoholic fermentation, would not pass through membranes that allowed the passage of organic substances in solution but held back particles in suspension. This experiment, successful with alcoholic fermentation, failed with many other ferments and fermentable liquids. Pasteur's mind was naturally addicted to generalization, and his interest lay in the phenomenon of fermentation as a general type of reaction, rather than in one kind of fermentation in particular. It was therefore natural that he should at first neglect the field in which the battle was more evenly balanced between the purely chemical conceptions of Liebig, and the biological theories of Cagniard Latour, Schwann and Helmholtz, and turn to the field in which Liebig's views had never been successfully attacked. Pasteur's first memoir was published in 1857, and in it he declared the lactic ferment to be a living organism, far smaller than the yeast-cell but which could be seen under the microscope, could be observed to increase in amount when transferred from one sugar solution to another, and had very decided preferences as regards the character of the medium in which it was allowed to develop, so that, for instance, by altering the acidity of the medium one could inhibit or accelerate its growth and activity. In this memoir Pasteur laid the first foundations of our knowledge of the conditions which must be fulfilled for the cultivation of bacteria.

These studies on fermentation occupied Pasteur almost continuously from 1855 to 1860, and he returned to them again at intervals during later years. He was able to show that the fermentation of various organic fluids was always associated with the presence of living cells, and that different types of fermentation were associated with the presence of microscopic organisms which could be differentiated from one another by their morphology and by their cultural requirements. Thus, at this early stage, the idea of specificity entered into bacteriology.

It was impossible for Pasteur to pursue these studies without facing the problem of the origin of these minute living organisms, which he regarded as the essential agents of all fermentations. At this time (1859) there were two opposed schools of thought with regard to the genesis of microbial forms of life. One school, deriving their concepts from the great naturalists of antiquity, believed in the spontaneous generation of living things from dead, and especially from decomposing organic matter. It is of little interest to remember the vague terms in which such conceptions were clothed, but one tendency may be noted, which did not escape the astute mind of Pasteur. The species of animals or plants believed to arise by spontaneous generation were diminishing in number, and the average size of those organisms still included in this category was getting smaller and smaller. In the beginning, the supporters of spontaneous generation were prepared to attribute this mode of origin to relatively large animals. Van Helmont, in the sixteenth century, offered a prescription for making mice. It needed the experiments of Redi (1688) to substitute, for the idea that worms were spontaneously generated in decomposing meat, the truth that these worms were the

larvæ of flies, and that their appearance could be very simply prevented by protecting the meat with gauze, through which the flies could not pass to deposit their eggs. The discovery by Leeuwenhoek of the world of microbial organisms gave a powerful stimulus to the somewhat decadent theory. Here, at all events were living things which obeyed no known law of reproduction, and whose existence seemed to lend support to a belief which had long been accepted by eminent authorities, and which had thereby acquired a natural prestige.

From the start of his inquiry, Pasteur leaned towards the opposing school of those who believed that spontaneous generation was a myth, that these microscopic organisms, like other living things, were reproduced in some way from similar pre-existing cells. He had already convinced himself that these organized cells were the active agents of fermentation. Clearly then they could not arise *de novo* during the changes for which they were themselves responsible, but must have been introduced from without. Their marked specificity, maintained through repeated transferences from one specimen of fermentable fluid to another of the same kind, was strong evidence in favour of their autonomous reproduction. Here again Pasteur had tentatively adopted the correct solution before starting his experimental inquiry, but the main interest of his part in the controversy lies in the consummate skill with which he developed methods which enabled him to give clear demonstrations where others had left doubt and confusion, and which determined the main rules of a technique which has made possible the cultivation and study of bacteria.

Neglecting for the moment the vaguer conceptions of the pre-experimental era, the position in 1859 was as follows. Needham, an Irish priest, had published in 1745 a memoir describing the spontaneous generation of microbial organisms in closed flasks of putrescible fluids, which had been heated to destroy pre-existing life. These views were strongly supported by the celebrated naturalist Buffon in 1749. An Italian abbot, Spallanzani, countered in 1769 with the publication of a series of admirable experiments in which he criticized Needham's results, and showed that, with longer heating, the fluid in such flasks remained clear and sterile. This controversy narrowed into a dispute as to the nature of the principle which survived short periods of heating but was destroyed by long heating in flasks hermetically sealed. For Spallanzani the principle was a living germ, for Needham it was a "vegetative force," resident in the air, or perhaps in the putrescible fluid. In any case such argument was sterile, and although it was generally admitted that the honours remained with Spallanzani, no final judgment was pronounced.

At this time oxygen was regarded as an element of quite peculiar power and significance, and the experiments of Appert (1810) on the preservation of food stuffs, by heating and hermetical closure of the containing vessels, followed by a weighty expression of opinion by Gay Lussac, had led to a general belief that the exclusion of this gas was the essential factor in ensuring the absence of fermentation. Schwann (1837) showed that the air in a flask containing a putrescible fluid, which had been sterilized by boiling, could be renewed by drawing in air which had passed through a glass tube immersed in a bath of fusible alloy kept at high temperature, and by this means he demonstrated that the presence of oxygen alone would not cause the appearance of micro-organisms in the fluid. Unfortunately, in the same memoir, Schwann reported other experiments, in which he introduced heated and unheated air into flasks, containing a sterilized solution

of sugar in a watery extract of yeast, by inverting the flasks over a mercury bath and admitting the air through the mercury seal. Here his results, as regards the occurrence of fermentation, were altogether uncertain, and his conclusions lost much of their force. Helmholtz (1844) confirmed certain of Schwann's observations. Schultze (1836) had already obtained similar results by admitting to his flasks air which had been drawn through strong potash solutions or through concentrated sulphuric acid. Schroeder and Dusch (1854) showed that the active principle could be removed from the air by drawing it through cotton wool. This last method was a real advance, since the incoming air had not been subjected to high temperatures, nor to strong chemical reagents. Unfortunately another element of doubt was introduced. Schroeder and Dusch relied, for their preliminary sterilization, on a short period of heating to the boiling point. They experimented with four kinds of material—water containing meat, malt of beer, milk and meat without the addition of water. With the first two materials their results were quite uniform: the fluids remained unaltered. With the last two materials fermentation usually occurred. They concluded that there were two kinds of decomposition, associated with the presence of living organisms, the one spontaneous, needing only the presence of oxygen, the other requiring some additional principle, which could be removed from the air by filtration through cotton wool.

This, then, was the position when Pasteur began his investigations in 1859. In a series of admirable memoirs, starting in 1860 and continuing for more than four years, he went over the ground already covered, added new and illuminating experiments of his own devising, and terminated the controversy by clear and decisive demonstrations. He showed that the material removed from air by passage through cotton wool, or through similar filters, contained organized particles which were neither crystals nor starch granules, but which were similar in appearance to the spores of moulds. By introducing these particles into flasks of sterilized organic material, he demonstrated that they were capable of giving rise to the growth of numerous kinds of living organisms. Using other methods, he showed that the air in different situations differed in its content of these germs, that they were numerous in the streets of cities, less numerous in the air of country uplands, rare in the quiet air of closed and uninhabited rooms or cellars, where the dust had deposited and remained undisturbed, and very rare in the pure air of the high Alps, above the level of human habitation. He showed that Schwann's failures were due to his use of mercury, from the surface of which his fluid had acquired the germs, which had settled on it from the air. He showed that the failures of Schroeder and Dusch were due to the inadequate sterilization of their material.

He also showed that certain animal fluids, such as blood or urine, known to be eminently liable to undergo putrefaction, could be collected in such a way as to remain permanently unaltered.

The controversy with Pouchet, Joly and Musset, which continued from 1860 to 1864, did not lead to the collection of many new facts, except those with regard to the unequal distribution of micro organisms in the atmosphere, but a later dispute with Bastian, who became a veteran in the dwindling army of the supporters of spontaneous generation, was more fertile, because it caused Pasteur to reconsider some of his ideas, and to elaborate the technical methods which he had partially developed during his re-investigation of the results obtained by Schroeder and

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While investigating the phenomenon of fermentation and the problem of spontaneous generation Pasteur had studied the behaviour of very various kinds of natural organic fluids and solutions and had succeeded in growing micro-organisms on simple synthetic media. As a result he had become assured of the fact that a medium, which is eminently suitable for the growth of one bacterium or mould, may be ill adapted for the growth of another, and that one of the primary necessities for the successful cultivation of any species of micro organism is the discovery of a suitable medium for its growth. Quick to grasp the general significance of isolated observations, he pointed out the decisive effect which must be exercised by the selective action of various environmental factors in determining the constitution of any naturally occurring bacterial flora, and he later developed these ideas in connection with the problem of infection.

As the result of these studies Pasteur had collected a mass of data which enabled him to deal successfully with bacteriological problems that could not previously have been attacked. He had learned the need for the scrupulous sterilization of everything that came into contact with material which was to be submitted to bacteriological examination. He had learned the necessary methods of sterilization, in the steamer, in the autoclave in the hot air oven, or by direct flaming, which enabled these conditions to be fulfilled. He had proved the serviceableness of the cotton wool plug for protecting media in flasks or tubes. He had realized the importance of the constitution of the nutrient material offered to a given bacterium, of the acidity or alkalinity of that medium and of the oxygen pressure to which it was subjected. Armed with this knowledge he proceeded to break new ground.

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desired greatly that his discoveries should benefit mankind in general, France in particular, and, if possible, his neighbours in the first place. Thus we find him investigating with enthusiastic care the troubles of the local vintners or brewers, or vinegar makers, and many of his memoirs are devoted to the diseases of wines or of beers, and the methods of preventing them. It was in connection with these studies that Pasteur faced a new problem of fundamental importance. He had shown that ferments were living organisms, that they were specific, that they were reproduced from parent forms and not by spontaneous generation. He was now faced with the problem as to whether one species could change into another, in particular whether *mycoderma vini* could change into the ordinary yeast of wine. Deceived on this point at first, he resorted as usual to rigorous and repeated experiments, and not only demonstrated that this mutation did not occur, but indicated clearly the conditions which led to its apparent occurrence, and the care which must be exercised before accepting any reported variation of this kind.

Anyone who reads for himself the original memoirs on fermentation and spontaneous generation (see Vallery Radot, P., 1922-1933) will realize that the possibility of applying this new knowledge to the elucidation of infective disease was already in Pasteur's mind. It needed only the spur of a request from Dumas to investigate the disease, which was then ruining the silkworm industry in the South of France, to turn his steps permanently towards the study of infective processes. We cannot follow here, even in outline, Pasteur's researches into pébrine, anthrax, chicken cholera, or hydrophobia. Some of them will be referred to in later chapters. We must, however, note certain contributions which Pasteur and his colleagues made to the fundamental data of bacterial infections. It was Pasteur who showed, in the case of anthrax, that a culture of a pathogenic organism could be passed through successive subcultures, in such a way as to dilute, beyond possibility of significant action, any other material introduced with it into the primary culture from the blood or tissues, and still produce the disease when inoculated into a susceptible animal, though it is to Koch that priority must be given as regards many points in the demonstration of the nature and action of the anthrax bacillus. It was Pasteur who introduced into bacteriology the conception of virulence and of attenuation, and who demonstrated the fact that an attenuated bacterial culture will act as a vaccine, that is, will confer immunity against subsequent infection with a virulent strain of the same bacterium. For Pasteur, indeed, a vaccine was synonymous with an attenuated culture, as opposed to a virulent culture on the one hand and to a dead culture on the other. It was Pasteur who, in the case of rabies, showed that it was possible to study the virus of an infective disease by animal passage, when the organism could not be cultivated, and even to prepare a perfectly efficient vaccine by using suitably treated animal tissue.

Thus, throughout a long scientific life, Pasteur was largely concerned with the practical application of knowledge gained during his studies on fermentation. The correct procedure for preparing good wine, good beer, good vinegar, and the methods of preserving them, the control of pébrine, of anthrax, of chicken cholera, of hydrophobia, these were the problems which occupied the last thirty years of his life, and the solution of which made his name a household word. But we shall miss the real significance of his work if we fail to realize that his fertile generalizations were of infinitely more importance for the progress of science than were his successful attacks on these isolated problems.

He had learned how to isolate and cultivate bacteria, and how to study their effect on animals, but with the minutiae of their morphology or physiology, apart from any significance these might have for the problem in hand, he was not greatly concerned. Duclaux relates that a clever and positive microscopist, who told Pasteur in very cautious language that a certain organism which he had taken for a coccus was in reality a very small bacillus, was much astonished to hear him reply "If you only knew how little difference that makes to me!"

One further point must be noted. Pasteur and his colleagues had shown how to obtain cultures of micro-organisms and propagate them indefinitely in the

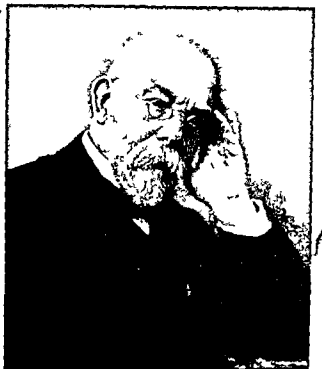


FIG. 2.—ROBERT KOCH (1843-1910)

laboratory, but the methods which they employed were not well suited to the isolation of pure strains of bacteria from an originally mixed culture, except in those relatively rare cases in which it was possible to employ a highly selective medium. Since all media were employed in the fluid state the only method of purifying a culture was to make successive transfers with very small amounts of material, in the hope that only a few bacteria, all of one kind, would be carried over. Such a technique was very uncertain in its results.

Pasteur, starting as a chemist, founded bacteriology and revolutionized medicine. At about the time when he was propounding his germ theory of disease, a young German physician some twenty years his junior was turning from clinical medicine to bacteriology. Robert Koch (1843-1910) at that time a

practising physician at Wollstein attacked the problem of anthrax and produced as his first contribution to science a demonstration of the character and mode of growth of the causative bacillus which opened a new era in bacteriological technique. This memoir he published in 1876. In the following year he published his methods of preparing fixing and staining film preparations of bacteria using the aniline dyes introduced into histology by Weigert and described his methods of photographing such preparations. In 1878 he published his memoir on traumatic infective diseases which remains a classical example of the study of experimental infections in laboratory animals. In 1881 he described his method of preparing cultures on solid media a technical advance of the first importance since it made possible the isolation of pure strains of bacteria from single colonies. Solid media prepared from naturally occurring material such as pieces of potato had previously been used for the isolation of micro-organisms particularly by mycologists and the general principles to be observed in the preparation of pure cultures had been clearly enunciated by Brefeld who had suggested the solidification of a nutrient medium by the addition of gelatin. The media and methods available for the cultivation of fungi were not however well suited for bacteria and it was left for Koch to devise in the form of his nutrient gelatin and later at the suggestion of Frau Hesse of nutrient agar a solid transparent medium easy to sterilize and handle and thus admirably adapted for obtaining isolated colonies of bacteria (see Bulloch 1930). In 1882 and 1884 he published his classical papers on the bacillus of tuberculosis. In 1883 he discovered the vibrio of cholera. Already Koch had enlisted the services of Loeffler and of Gaffky as his assistants. Later came Pfeiffer Kitasato Welch and many others and with his growing fame he began to gather round him a group of keen and able young men who were destined to introduce the methods he devised into the laboratories of many lands. In 1880 he was appointed Professor of Hygiene and Bacteriology in Berlin and in 1891 he was made Director of the newly founded Institute for Infective Diseases. His later years were devoted almost entirely to the investigation of bacteriological problems in their relation to the prevention and cure of disease and many of his contributions to our knowledge will be considered in later chapters. Koch was above all an able and careful technician. He was greatly aided by the vigour and initiative of the great German chemical and optical firms and the advances which he made in staining methods in the use of the microscope for the observation of bacteriological preparations and in the technique of cultivating bacteria revolutionized this branch of science.

The fruits of this revolution appeared with surprising rapidity. During the last quarter of the nineteenth century a succession of discoveries was reported bearing on the relation of bacteria to human and animal disease which opened a new era in medicine.

In 1874 Hansen described the bacillus of leprosy and Neisser in 1879 the gonococcus. In 1880 Pasteur recorded the isolation of the bacillus of fowl cholera and Eberth observed the bacillus of typhoid fever. In 1881 Ogston published an adequate description of the staphylococcus. In 1882 Koch discovered the tubercle bacillus and Loeffler and Schutz the bacillus of glanders. In 1883 Koch discovered the cholera vibrio. Fehleisen isolated the streptococcus of erysipelas and Klebs described but did not isolate the bacillus of diphtheria. In 1884 Loeffler isolated and subjected to thorough study the bacillus which Klebs had briefly described in the previous year, and Gaffky isolated and studied the typhoid bacillus which

Eberth had observed four years previously. In 1885 Loeffler discovered the bacillus of swine erysipelas, Kitt the bacillus of hæmorrhagic septicæmia of cattle and Salmon and Smith the bacillus associated with hog cholera. In the same year Nicolaier observed the tetanus bacillus in soil inoculation of which produced the disease in animals. In 1886 Fraenkel isolated the pneumococcus, Escherich the colon bacillus, and Loeffler the bacillus of swine plague. In 1887 Weichselbaum discovered the meningococcus and Bruce the micrococcus of Malta fever. In 1888 Gaertner described the bacillus which bears his name and Schutz the streptococcus of equine strangles. In 1889 Kitasato cultivated the tetanus bacillus which had been earlier observed by Nicolaier. In 1892 Pfeiffer isolated the bacillus which he believed to be the cause of influenza and Welch and Nuttall described the anaerobic bacillus now known as *C. welchii*. In 1894 Kitasato and Yersin independently discovered the bacillus of plague. In 1895 Moore isolated the bacillus of fowl typhoid. In 1896 van Ermengem described *C. botulinum* as the cause of a particular variety of food poisoning. In 1897 Bang discovered the bacillus of bovine abortion. In 1898 Shiga isolated the variety of dysentery bacillus which bears his name and Nocard and Roux described the minute organism of infectious pleuro pneumonia of cattle.

Thus by the close of the nineteenth century a great variety of micro organisms had been identified as occurring in definite association with human or animal disease. In many instances complete demonstration had been afforded that the relation was one of cause and effect. In others this relation was rendered highly probable. In others, again, there remained good reason for doubting whether the bacterium, whose presence had been demonstrated, played any more important role than that of a secondary invader. Beyond dispute, however, the scientific investigation of infective disease had become the province of the bacteriologist.

Another incident had done much to emphasize the importance of bacteria as the cause of disease and death although it had comparatively little influence on bacteriology itself. Joseph Lister (1827-1912) during his tenure of the Professorship of Surgery at Glasgow, was deeply interested in the post-operative sepsis, which exacted such a terrible toll on the lives of hospital patients. His attention was drawn to Pasteur's work on fermentation and the analogy between the changes which occur in fermenting organic material and the putrefaction which occurs in wounds suggested to him that in the latter as in the former the underlying cause might be the activity of minute living organisms. This led directly to the introduction of his antiseptic technique in surgery described in 1867 which opened the door to modern surgical methods. Lister's technique has since been replaced by aseptic measures, but this detracts in no way from the merit of his discovery nor from the debt which we owe to him for fighting the usual battle against the forces of ignorance and prejudice. Nor should it be forgotten that Lister made important contributions to bacteriological technique as such. He devised a method for diluting a bacterial culture and preparing a series of subcultures with so small a volume of the original fluid that many of them remained sterile the presumption being that those that grew had developed from a single bacterial cell. In this way he isolated, in 1878 a bacterium that caused the souring of milk and Bulloch (1938) expresses the view that he may perhaps have been the first bacteriologist to obtain a certainly pure culture.

But the revolution inaugurated by Pasteur and extended by Koch spread far beyond the field of medicine. Agriculturists had long been puzzling over the

problem of soil fertility, without arriving at any very helpful conclusions. One curious phenomenon was the reaccumulation of nitrates in the soil, in spite of their constant removal by the washing action of the rain. It was suspected that these nitrates might be derived in some way from the decomposition of organic material and in 1877 Schloesing and Muntz, acting on a suggestion made by Pasteur in 1862, showed by experiment that the formation of nitrates was due to the action of living organisms. Warington, at Rothamsted, confirmed these results in 1878 and 1879, and showed that two stages were involved a preliminary conversion of ammonia to nitrites, and a subsequent oxidation of nitrites to nitrates. He believed that these two stages were carried out by different organisms, but failed to isolate or identify them. This problem was solved by Winogradsky in 1890 who isolated and described both the nitrite and nitrate forming organisms. In 1888 Hellriegel and Willfarth described the nitrogen fixing bacteria which caused the formation of nodules on the roots of leguminous plants. Later Winogradsky described a free-living anaerobic organism which was able to fix atmospheric nitrogen, and Beijerinck, some ten years later, described a large, free living, nitrogen fixing aerobic bacterium, which he named *Azotobacter*, and which has since been extensively studied. The bacteriology of the soil thus became an important part of agricultural science.

In the early years of the bacteriological revolution it had been demonstrated that bacteria attacked plants, as well as animals. In 1878 Burrill described the organism of pear blight, and in 1883 Walker discovered the bacillus which causes the 'yellows' of the hyacinth. This branch of bacteriology has been pursued energetically during the last thirty years especially by Erwin Smith and his colleagues in America.

The demonstration by Pasteur of the essential nature of fermentation led, as a natural consequence, to the entry of the bacteriologist into the commercial sphere. His help was required in dairy farming in brewing in the preservation of foods, and in all those commercial processes in which bacterial activity was desired or feared.

Fig 3, which sets out in diagrammatic form the time relations of the more important discoveries associated with the work of Pasteur, Koch and Lister, may be of some service in enabling the student to follow the development of our knowledge down to the end of the nineteenth century.

This brief summary will indicate with sufficient clearness to how great an extent the bacteriologist has been occupied with applied problems. He has, by way of description, usually been satisfied if he could determine, for any given bacterium, a number of characters sufficient to differentiate it from the organisms with which he considered that it was most likely to be confused. It is in this way that our knowledge of bacterial characters has slowly grown and it is not surprising that the results should be an arbitrary and rather odd assortment of differential criteria. In such a bacterial group as that comprising the colon and typhoid bacilli, and certain nearly related organisms, it has been demonstrated that fermentation reactions form a reliable method of differentiation, and such reactions have been extensively studied. In another group morphological differences may be more distinctive, or the production of specific toxins may be a well marked feature in certain species. The soil bacteriologist employs methods which differ in important respects from those used by other workers. It is the inevitable result that systematic bacteriology has been very generally neglected, and it is only in recent

years that any real attempt has been made to survey bacterial groups as a whole, and to bring some order out of our chaos

The first two decades of the present century witnessed no such striking advances in our knowledge of the bacteriology of disease as occurred between 1875 and 1900, and the reason for this slower progress is obvious. The technique developed by Pasteur and Koch had been applied over a very wide field. Those problems which were susceptible of solution by the methods available had, to a great extent,

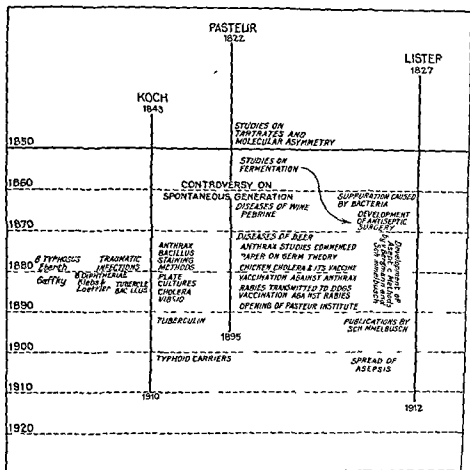


FIG. 3.

been solved, and those which remained unanswered appeared to demand new methods of attack, or at least attack along new lines

The study of immunity has absorbed the interest and energies of a large number of bacteriologists during the past forty years. This branch of bacteriology derives from Pasteur's studies on chicken cholera, anthrax and rabies, from Metchnikoff's investigations on the cellular reactions in infection, and from the work of Buchner, Nuttall, von Behring, Ehrlich, Bordet and others, who have investigated the reactions between the sera of artificially immunized animals and the bacteria, bacterial products, foreign cells, or foreign proteins with which they have been

inoculated. The development of our knowledge of these phenomena will be discussed in Chapter "

The demonstration by Loeffler and Frosch in 189 that foot and mouth disease was caused by a virus which could pass through a porcelain filter and was below the limits of direct microscopical observation opened a new field for investigation. We now know many diseases which can be transmitted by filtered suspensions of material obtained from infected animals and in which therefore a filter passing virus is presumably concerned.

There is nothing surprising in the fact that the years from 1900 to 1920 or thereabouts were for bacteriology a period of slower development as compared with the riotous growth of the eighteen-eighties. The ground won had to be consolidated. The previous advance had been a somewhat hasty affair and many secondary problems had been left for more leisurely solution. Many bacteria were very incompletely described. Many had been described independently by different investigators so that the same bacterium was masquerading under several different names. Many of the earlier descriptions especially of the anaerobic bacteria had been based on impure cultures. Little notice had been taken of resemblances between bacteria isolated at different times from different sources unless the practical application of the knowledge gained brought such resemblances forcibly to the attention of some observer. Little was known about the distribution in nature of bacteria other than those concerned in the causation of disease or in some commercial or agricultural process, and even here the data were very scanty. Bacterial ecology is indeed still almost an unexplored territory. In all these directions the past thirty years have seen a considerable advance. Bacteria have been studied more systematically. Fuller and more accurate descriptions have been recorded and errors have been corrected. The labelling of the stock strains of bacteria scattered throughout the laboratories of the world has been more closely scrutinized. Many synonyms have been suppressed and species that had received several names have been shorn of all but one. The formation of such collections as the National Collection of Type Cultures in this country has provided a much needed standard of reference.

This re-survey of the bacteriological field on its qualitative side has been associated with a great advance in our quantitative methods. The introduction of bacteriological methods of analysis in the control of water supplies, milk and so on demanded standardized tests yielding numerical answers. Those first employed were in most cases very faulty. There was a failure to realize many of the technical sources of error and there was a still more general failure to take into account the statistical principles involved in any sampling of this kind. It is indeed only within recent years that a satisfactory liaison has been established between the bacteriologist and the statistician and even now it is not as general as it should be. It is not merely a question of the kind of analytical test referred to above. The vast literature of immunity contains record after record which is rendered meaningless by a neglect of the sampling errors involved in working with small groups of animals. Reference to Chapter 43 will provide examples of the ways in which these errors may be avoided.

In the last twenty years or so there have been unmistakable signs that bacteriology is on the march again. As always in experimental science it has been a matter of technique. In this case the acceleration has followed the application to bacteriology of the more exact methods of analysis developed by the chemist and the

physicist. The facts set out in Chapter 3 show clearly the rapid advance that has followed the incursion of the biochemist into the bacteriological domain. Those described in Chapter 6 show the organic chemist and the biochemist initiating a remarkable extension of chemotherapy in relation to bacteria and the outstanding success of synthetic sulphonamide compounds and of the natural mould product penicillin and those described in Chapter 7 show the organic and physical chemist inaugurating a new era in immunology. The remarkable increase within quite recent years of our knowledge of the filtrable viruses (see Chapters 41 and 80-89) has resulted in large part from improved methods of filtration, optical examination and high speed centrifugation. It is not a rash prophecy that the years ahead of us will be the eighteen eighties over again.

## REFERENCES

- BULLOCK W. (1938) *The History of Bacteriology*. Oxford Univ. Press, London.  
 COMEY D. (1937) *J. Bact.* **34**, 343.  
 DOBELL, C. (1931) *Antony van Leeuwenhoek and his 'Little Animals'*. John Bale Sons and Danielsson, London.  
 DECLAUX E. (1920) *Pasteur the History of a Mind*. Eng. Transl. by E. F. Smith and Florence Hedges. Saunders, Philadelphia and London.  
 VALLERY RADOT P. (1900-33) *Œuvres de Pasteur*. 6 vols. Masson et C<sup>o</sup> Paris.  
 VALLERY RADOT R. (1919) *The Life of Pasteur*. Eng. Transl. by Mrs R. L. Devonshire. Constable, London.



## CHAPTER 2

### THE BIOLOGICAL CHARACTERISTICS OF BACTERIA MORPHOLOGY

**General Considerations**—With the exception of certain observations on the finer structure of the bacterial cell, which will be referred to later, our knowledge of bacterial morphology has been gained from the study of cells which have been cultivated in the laboratory under artificial conditions. The morphology of bacterial cells may be notably affected by the constitution of the medium on which the bacteria are grown, the temperature of incubation, and many other factors. In particular, the cells in a pure culture may show very striking changes with age. It is customary to regard the forms found in young cultures as typical of a given species, and the very different appearances, often met with in old cultures, as due to the occurrence of degenerative changes. How far we are justified in labelling all the morphologically atypical cells that we meet with in old cultures as degeneration or involution forms, is a controversial question which is discussed elsewhere. It must always be remembered, however, that when a description is given of the morphology of any bacterial species, such a description is supposed to apply to the cells found in a young, actively growing culture on a medium which is favourable to the growth of that particular species and incubated at the optimum temperature, unless the contrary is specifically stated. Those who are for any purpose describing the appearances met with in preparations from bacterial cultures should always recollect that such descriptions have little value unless the exact conditions of cultivation are carefully noted.

Apart from variations associated with age, variations in form, sometimes of a very striking character, may occur in young cultures of a single bacterial species. Different forms of cell may be present in a single culture, or the cells may appear to alter their form in successive subcultures, or different strains of a single bacterial species may show morphological differences, which persist in successive subcultures carried on over a considerable period of time. A description of the morphology of a given bacterial species should include the characters displayed by the modal form, and the extent to which these characters vary. Variability of form is in itself, very characteristic of certain bacterial species, while other species show only minor differences in the shape and size of the bacterial cells.

**The Size of Bacterial Cells**—Ignoring for the moment the very large bacteria which have been described by a few investigators, and those very minute organisms which will pass through a porcelain filter—the filtrable viruses—it may be said that the dimensions of most cultivable forms are of the order of low multiples or sub-multiples of  $\mu$ , i.e. of 1/1000th of a millimetre. Among the spheroidal forms, the

parasitic staphylococci and streptococci usually measure between  $0.75\mu$  and  $1.25\mu$  in diameter. Some forms of micrococci or sarcinae may show an average diameter of  $1.5$  to  $2\mu$ . Among the rod forms, a relatively large bacillus, such as *B. anthracis*, has a transverse diameter varying between  $1$  and  $1.25\mu$  and a length varying between  $3$  and  $8\mu$ . A medium-sized bacillus, such as *Bact. coli*, has a transverse diameter varying between  $0.5$  and  $1\mu$ , and a length of  $2$  to  $3\mu$ . A very small bacillus, such as the influenza bacillus, has an average diameter of  $0.2$  to  $0.1\mu$ , and a length of  $0.7$  to  $1.5\mu$ . Some bacilli, such as *Cl. tetani*, may combine a small transverse diameter,  $0.3$  to  $0.4\mu$ , with considerable length,  $3$  to  $5\mu$ . All such measurements must, of course, be taken as representing a modal size, corresponding to the modal form, and wide variations may occur. Some species of bacilli, for example, may show occasional filamentous forms, measuring  $100\mu$  or more in length. It remains true that the modal size of a bacterium is one of its distinctive characteristics and has the advantage that it can be stated in numerical terms. In such forms as the *Actinomyces*, which are normally filamentous, it is to the transverse diameter alone that we can assign a modal value.

**The Shape of Bacterial Cells**—We can recognize three main types of bacterial cell—the *coccal* or *spheroidal*, the *bacillary* or *cylindrical*, and the *spirillar*.

The coccal form is distinguished by the fact that any one axis of the bacterial cell is approximately equal to any other. Many forms approximate to true spheres, although it is doubtful whether any living cell is truly spherical. In many cases the spherical form is widely departed from, and the individual cells may be ellipsoidal or show conical distortions, flattenings or indentations, which give the cells, when observed in film preparations, shapes which may be likened to a bean, or a kidney, or a lancet, as the case may be.

The bacillary form is distinguished by the fact that one axis of the cell is markedly longer than either of the others, which are themselves approximately equal. Since it is customary to examine bacteria in film preparations, and to describe them as though they were two-dimensional bodies, it is usual to refer to the long axis and the transverse axis, ignoring any possible departure from the circular form in the cross section of the cylinder. The ratio between the length of these cylindrical cells and their transverse diameter may vary over an enormous range, so that, while some may be almost coccal in appearance, others may be filamentous. Certain modifications of the general cylindrical shape are characteristic of particular bacterial groups. The average ratio of the long axis to the transverse diameter, that is, the *thinness* or *thickness* of the cell, is one such character. The ends of the bacillus often show modifications of form which are of differential value. They may be square-cut, rounded, or acutely pointed, or may form definite clubs. Other irregularities in contour, due to the presence within the cell of structures which cause distortion of the cylindrical form, will be considered later.

The spirillar form is characterized by a bending or twisting of the cells, so that they assume curved or spiral shapes. It has become customary to speak of a bacterium which shows a single curve, thus assuming the so-called comma shape, as a *vibrio*, and of a bacterium which shows a series of curves or twists, thus assuming a corkscrew form, as a *spirillum*. *Vibrios* and *spirilla* are of necessity relatively elongated cells, and they always have rounded or pointed ends.

The study of the finer details of bacterial structure may be carried out

different ways. Each of them is subject to the limitations imposed by the optical system employed.

Stained or unstained preparations may be examined under the microscope, using light transmitted through a sub-stage condenser of the ordinary type. The limit of resolution,  $d$ , (that is, the shortest distance by which two particles must be separated in order that they may give distinct images) is determined by the wave-length of the light and the numerical aperture of the objective. It is given by the formula,

$$\frac{0.5\lambda}{\text{N.A.}}$$

where  $\lambda$  = wave length of light used and N.A. = numerical aperture of objective.

The possible range of  $\lambda$  is obviously limited to that part of the spectrum to which the human retina is sensitive and the numerical aperture is subject to the technical limitations of the optical system employed. In practice, the highest N.A. that can be employed with transmitted light is about 1.4 and this, when used with monochromatic light with a wave-length of 546 m $\mu$ , corresponding to the green mercury line, gives a resolving power of

$$\frac{546}{2} \times \frac{1}{1.4} = 195 \text{ m}\mu = 0.2 \mu \text{ (approximately)}$$

This degree of resolution is obtained only under optimal conditions, and in ordinary practice the limits of resolution are reached with particles that have a diameter of about 0.25  $\mu$ . Merling Eisenberg (1937) has modified the formula by introducing a factor measuring the intensity of illumination. By suitably reducing this intensity a limit of resolution of 0.08  $\mu$  is approached. Under ordinary conditions, however, objects smaller than 0.25  $\mu$ , though seen in the sense of being visible, do not form images that reveal the real size or shape of the particle.

Another method is that known as dark ground illumination. The bacteria, or particles, are examined unstained, and the light passing through the special sub-stage condenser is directed along a path such that only those rays that are refracted, diffracted, or scattered by the object under examination reach the eye of the observer. Bacteria so examined appear as bright images on a dark background. This method is vastly superior to the former as a means of examining living, unstained organisms, but it is subject to the same limitations in regard to resolution, and, since the highest N.A. at present available for use with illumination of this type is about 1.27, the smallest particle that can be resolved has a diameter of about 0.35  $\mu$ .

The third method, which has been developed particularly by Barnard (1910, 1925, 1930), extends the limits of resolution by decreasing the wave-length of the light used. Using ultra violet light with a wave-length of 257 m $\mu$  and an optical system of quartz lenses, Barnard has been able to photograph and resolve particles with a diameter of 0.075  $\mu$ . Beyond this point a limit is again reached, due to the lack of a refracting material that will transmit light of shorter wave-length. The degree of internal structure that may be revealed in certain bacteria by ultra-violet photography, combined with dark ground illumination, is illustrated in Fig. 4, for which we are indebted to Mr. Barnard.

The electron microscope devised by Ruska (1934) and by Marton (1934) provides the fourth method of examination (see Marton 1941). The fact that an electron beam passing through a magnetic field behaves in a manner closely analogous to a beam of light passing through a refracting medium permits the construction of a microscope generally similar to an optical microscope, and its description in the terminology of light optics. The "lenses" are circular electro-magnets whose focus varies as the strength of the applied magnetic field, and the microscope has a "condenser" coil, an "objective" coil and a "projective" coil, the last being equivalent to the eyepiece. The "wave-length" is a function of the speed of the electrons, and those used in electron microscopy are equivalent to a wave length about 1/100,000th that of visible light. Resolving powers commensurable with this wave-length cannot be attained in practice, since "chromatic" aberration in the magnetic lens cannot be corrected, and "spherical" aberration is over

a thousand times that of a glass lens. With these restrictions, a theoretical resolution of  $10\text{ m}\mu$  is possible, and in practice resolutions of  $22\text{ m}\mu$ , equivalent to a magnification of 180 000, have been attained. For biological purposes magnifications of 10 000 to 50 000 are employed. The different opacities in an object recorded on the photographic plate are due to the scattering of the electron beams, not to the absorption and refraction of light, as in the optical microscope. The intensity of the transmitted beam for a given electron speed, is roughly proportional to the thickness and density of the material examined. In a manner analogous with staining of optical microscopic objects certain biological materials may be increased in density (i.e. electron scattering power) by impregnation with salts of heavy metals (Mudd and Anderson 1942). The value of this method is at present severely limited by the necessity for completely dry specimens for examination, since the microscope works only in a high vacuum. (For a review of the recent achievements of electron microscopy, see Mudd and Anderson 1944.)

The great majority of the studies referred to in this section have, however, been carried out by the direct observation of stained preparations, and have therefore been subject to the narrower optical limitations considered above. As against these limitations must be set the advantage that, in preparations of this kind, we can study the affinity of different cell constituents for certain special stains. In most, if not in all instances, staining has been preceded by some form of fixation, so that the possibility of protoplasmic changes due to heat or other fixing agents, must be borne in mind.

#### The Nuclear Apparatus —

One of the most controversial questions in regard to the structure of the bacterial cell is the presence or absence of a nucleus, and its nature if present.

It is impossible to discuss at all fully the many conflicting statements which have been made, and the evidence on which they have been based. Much of this evidence indeed is of little value since it has been obtained by faulty or inadequate technique. Those who desire fuller information on this subject may be referred to the papers of Dobell (1911), Guilhaumon (1907) and Hollande (1934), and to the reviews of Knaysi (1935) and Lewis (1941).

Certain of the earlier workers regarded bacteria as cells possessing no nuclei. This nihilist view perhaps expressed the difficulty of demonstrating bacterial nuclei by methods that depend upon the staining of chromatin. Chromatin however is not the essential hereditary material, and its absence does not in any case establish the absence of a nucleus. Moreover as Lindgren (1935) points out, if the existence of some form of nuclear apparatus is denied an explanation is required for the constancy of transmission of multiple hereditary characters in bacteria, by a mechanism different from that in most other living forms. The hypothesis of some form of nuclear apparatus has at least the prior claim on our attention.



FIG. 4.—*B. megatherium*  
Unstained, dark ground illumination photographed with  
ultra violet light ( $\times 2500$ )

Puhka (1898-1903-1908-1909) regarded the whole bacterial cell as homologous with the cell nucleus of more highly developed organisms. There is no substantial evidence, apart from a similarity of staining reactions, for identifying the bacterial cell as nucleus, and, even if there were so the term nucleus would be a misnomer, for, as Dobell points out, a nucleus must by definition be differentiable from the cytoplasm of the cell.

Meyer (1897-1899-1908) has been a prominent exponent of the view that bacteria contain well-defined nuclei and has brought together the available evidence in his book on the bacterial cell (1912). Many other observers have shared this view (Feinberg 1909, Nakamura 1901, Ellis 1902-03, 1922, Grimme 1902, Swellengrebel 1906, Amato 1909, Vav 1909-1910) but it is reasonably certain that the structures observed differed in their nature so that the observations do not confirm one another. The observations of Mencl (1904, 1905-1909-1910) and of Ravman and Krus (1934) strongly suggested the presence of a differentiated nucleus in the bacteria they studied. Their earlier observations have not been confirmed, according to Guilhaumon (1907) the structures they describe as nuclei were stages in the formation of transverse septa. Stoughton (1920, 1937) found in a plant pathogen, *Bact. malvacearum*, a central nucleus-like body differentiable from the cytoplasm by *intravivam* staining. The constancy of its occurrence, position, and division during cell division suggested a true nucleus. Guilhaumon (1933) identified the body as a volutin granule (see below). In one case the observation by Veydovsky (1900-1904) of a nucleus in an organism he named *B. graminis*, there is a consensus of opinion that a true nucleus has been demonstrated, but there is good reason to believe that the organism itself is a fungus, not a bacterium.

There remain several possibilities, of which we may mention three—that the nuclear material is diffusely and uniformly scattered through the cytoplasm, that it is disposed in certain chromatin masses in the cytoplasm, and that there is in the cell a separate nuclear apparatus, perhaps consisting of one or more chromosomes, not usually revealed by ordinary staining methods.

The notion of the diffuse nucleus has arisen partly to explain the absence of a demonstrable discrete nuclear body (Zettnow 1918) and partly to account for the occurrence of a uniform coloration of some bacteria when treated by Feulgen's reagent. Feulgen's reagent is used as a test for thymonucleic acid. A diffuse Feulgen reaction of bacteria has been noted by a number of observers (see, for example, Patschmann and Ruppel 1932, Imšenecká 1936). Knays (1938) doubts the specificity of the reaction, and points out that if nucleic acid is demonstrated in this way, it may be merely reserve material in the cell having no necessary connection with a nuclear apparatus (see also Schaefer 1939, Mursky 1943, Stedman and Stedman 1943). As we have seen, Lundgren (1935) objects to the conception of a diffuse nucleus on genetical grounds, it is necessary to assume the existence of some form of discrete nuclear body which, by dividing into identical halves, can ensure the genetical constancy of the daughter cells. The view that nuclear material is segmented in certain chromatin masses in the cell has been developed by Schaudinn (1902) in the case of *B. butchlii*, by Guilhaumon (1907) in the case of several cultivable spore-bearing bacilli, and by Dobell (1911) in the case of bacilli parasitic in reptiles and fishes.

Schaudinn described numerous small chromatin like granules, which were scattered through the cytoplasm of *B. butchlii*, a large spore-bearing bacillus. At the start of sporulation, the granules gather into an axially situated spiral. The ends of the spiral increase in size at the expense of the chromatin granules, and develop into homogeneous masses that stain deeply with nuclear stains. These masses, as they develop into spores, cease to take up the stain, and generally appear as highly refractile unstained bodies. Guilhaumon (1907) found a similar picture in the spore-bearing bacillus studied by him. Cells eight hours old were finely vacuolated, and contained numerous granules of chromatin. At sporulation, however, no axial filament was formed and a single deeply staining mass appeared at one pole of the cell, gradually increasing in size. Dobell (1911) observed scattered chromatin that was gathered into an axial filament before sporulation.

in certain bacilli, and a permanent axial spiral in others, both of which he regarded as being nuclear in character. In certain large micrococci he observed well differentiated and homogeneous bodies which took up nuclear stains, divided before cell division occurred, and had the essential characters of nuclei. Reviewing his own results and those of other investigators, Dobell concluded that bacteria are nucleated cells, in which the nuclear apparatus is highly variable in form, appearing sometimes as scattered granules, sometimes as a spiral filament, and occasionally as a differentiated nucleus similar to that in cells of higher organisms. Lewis (1932, 1934, 1940) doubts the reality of nuclear spiral filaments, believing rather that they represent fine strands of cytoplasm compressed by an abundance of volutin granules or fat globules.

In recent years, the evidence for the existence of a discrete nuclear apparatus in bacteria has been accumulating. Hollande (1934) (see also Hollande and Hollande 1932) differentiated three structures in a number of bacilli, which he terms nucleosome, paranucleosome and metanucleosome. The paranucleosome is eosinophilic, and the metanucleosome, which surrounds it, is basophilic. Both are closely associated with the nucleosome, which is a minute spherical granule staining blue with eosinate of methylene blue. The nucleosome divides by elongation, constriction of the central portion into a thread, and finally separation into two daughter nucleosomes. Later workers confirm Hollande's work to the extent of recognizing bodies usually referred to as nucleoids, that take part in cell division. Dombrowsky (1936) described two concentrations of protoplasm in *Bact. coli*, which appeared prior to division, the line of division occurred between them. In young cells of another coliform bacillus Rosca (1937) found a chromatinic substance developing into a distinct 'nucleus' and noted the appearances resembling mitotic figures. Stille (1937) was able to demonstrate Feulgen reacting bodies in a large number of bacterial species after they had undergone gentle acid hydrolysis. Young vegetative forms contained two bodies, though cells containing four bodies appeared prior to division. Similar mild hydrolysis reveals nucleoids in *Bact. coli* and *Salm. paratyphi B* (Piekarski 1937) and in *Proteus vulgaris* (Neumann 1941). Piekarski distinguished a primary form with two bodies, and a secondary form, usually found in older cultures, with one only. The bodies were Feulgen positive, and absorbed ultra violet light of a wave length known to be strongly adsorbed by thymonucleic acid (Piekarski 1938). Later it was found that all the bacteria studied contained at least one nucleoid (Piekarski 1939, 1940, Piekarski and Ruska 1939 a, b). In some cases the nucleoids were demonstrable in electron micrographs. It may be noted here that the electron microscope has so far not revealed very much of the nuclear structure of bacteria (see for example von Bornes et al 1938, Lembke and Ruska 1940, Mudd et al 1942, Knavs and Mudd 1943). If any nuclear material is present, it must either be masked by other matter more opaque to the electron beam, or have the same electron scattering power as other cellular constituents.

Robinow (1942, 1944) has extended these studies of the nuclear apparatus revealed by preliminary acid treatment of the bacteria. Robinow's results not only confirm and extend those of Stille and Piekarski, but clarify the relation of the Feulgen positive (chromatinic) bodies to the growth of the bacteria. In old bacteria, the chromatinic bodies are small, and difficult to demonstrate, when the bacteria are transferred to a fresh nutrient medium, the bodies increase in size and stain more deeply, usually having the form of short, dumbbell shaped rods. Longitudinal divisions of these rods precedes the division of the cell, though several divisions may take place before the evidence of cellular division is evident by ordinary methods of examination. Growing bacilli may therefore contain two or more pairs of chromatinic bodies (Fig. 5). By special staining methods these multinucleate bacilli appear to be banded, indicating the existence of multiple cell units within the single bacillus. In the case of *B. megatherium* the composite nature of the cell can also be demonstrated by a plasmolysing treatment (see p. 23 below). The possibility that single bacillary rods may in fact be multicellular must be borne in mind in the interpretation not only of morphological studies on bacteria, but also of the phenomena of heredity that they display.

The fact that similar acid treatment does not destroy the chromatinic structures in other types of cell and that changes in the bodies are closely correlated with stages in the growth of the cells, makes it highly unlikely that the dumbbell bodies are artefacts. Pabinow concludes that the chromatinic bodies are comparable with the chromosomes of animal and plant cells. It may be noted that Beebe (1941) found single central Feulgen



FIG. 5.—Chromatinic bodies of bacteria, showing the various shapes taken during growth and division.

A *B. mycoides* B *Bact. coli* C, *Proteus vulgaris* D *B. mesentericus* ( $\times 4660$ ).  
(From photographs kindly supplied by Dr. C. F. Eshinow.)

positive bodies in a myxococcus, which divided with the cell and which at times developed a dumbbell shape.

The conception of the nuclear apparatus of bacteria as granules made up of one or more chromosomes has been reviewed by Lundegren (1935). Lundegren himself (1936) described a diplococcus whose nucleus, visible only at certain stages in division, consisted of a single haploid chromosome with seven chromomeres. Badian (1933) observed the longitudinal division of a single rod-like chromosome in *B. subtilis* and Chance (1938)

a mitosis like effect during the division of a bacillus resembling *B. mesentericus*. Prior to sporulation, the chromosomes of Badian's bacillus underwent division, followed by fusion end to end of the resulting two chromosomes, and division of the fused chromosome into four, only one of which was incorporated in the spore. A similar series of events was observed by Allen, Appleby and Wolf (1939) in a spore bearing bacillus. The normal vegetative cell contained one haploid chromosome. Two kinds of spore were formed, one haploid, the other diploid. In the one case a phenomenon analogous with meiosis occurred before spore formation, in the other after spore germination.

A nucleoprotein granule which persisted in all stages of growth, multiplication, ageing and starvation of the culture, and which divided before cell division was found in a staphylococcus by Knaysi (1942). The granule did not correspond to any known cellular reserve material, and was apparently the nuclear apparatus.

Taking the evidence as a whole, it appears that bacteria have a nuclear apparatus which in many respects is analogous to that of the cells of fungi, plants and animals, though the survey of bacteria by modern cytological methods is as yet too limited to permit more than a tentative conclusion that the apparatus seen in a few bacteria will prove characteristic of bacteria in general.

#### Other Intracellular Granules

**Nitrogenous Material—Volutin Granules**—In many species of bacteria there are found intracellular granules which possess a strong affinity for nuclear stains, and which are coloured a reddish purple with certain blue or violet stains, especially with polychrome methylene blue. These granules were first described by Ernst (1888, 1889, 1902) and by Babes (1889, 1895), and are frequently referred to as the Babes-Ernst granules. They have also been named volutin granules, and from their peculiar staining properties, metachromatic granules. These constituents of the bacterial cell have, from time to time, been credited with almost every conceivable function. There is now, however, very general agreement that they are relatively inert in the cell. Using a cytological term, they are metaplasmaic granules, particles of some substance concerned in cell metabolism. Their number and size vary according to the medium in which the bacterium is grown, and according to the period of growth. That they form no part of the nuclear apparatus is shown by the facts that they can be observed in yeasts and fungi, which possess well-differentiated nuclei and that they take no active part in cell division or sporulation. They contain a high proportion of nucleoprotein, and to this they probably owe their affinity for nuclear stains.

The rôle which they play in the chemistry of the bacterial cell is at present unknown; the suggestion of Marx and Worthe (1900) that the presence of these granules is correlated with the virulence of the bacterium, has been completely disproved by the work of several subsequent investigators (Ascoli 1901, Krompecher 1901, Gauss 1902, Ficker 1903, Guilhaumon 1906). Groh (1938) has recently assigned a reproductive rôle to the volutin granules of *C. diphtheriae*. His findings still await confirmation.

**Carbohydrates**—Glycogen granules and starch granules occur in many species of bacteria, and may be identified by staining with iodine.

**Fats and Lipoids**—Fat globules are frequently found in bacterial cells, and may be stained by the usual fat soluble dyes. Other lipoidal or waxy substances, which may be extracted by the usual solvents, are found in many bacteria, and are particularly abundant in certain species such as the acid fast bacilli.

These different forms of granular products of metabolism are by no means equally abundant in different bacterial species. In some, volutin granules and



to be numerous and of large size in others, they are scanty or absent. Some species which seldom contain volutin granules frequently contain fat globules. In others the presence of glycogen or starch is a characteristic feature. The data

available do not warrant us in saying that any one of these forms of granular material is the peculiar property of certain species or is uniformly absent from others, but their relative abundance or scarcity may be a striking characteristic of a particular bacterial species.

For a good and concise account of these intracellular granules see Zettnow (1918).

**Endospores** — The formation of intracellular spores by bacteria was first observed by Cohn (1875), and first studied in detail by Koch (1876) in the case of *B. anthracis*. The fact that the existence of an endospore, and its significance in reproduction, had been clearly demonstrated in the case of one of the first bacteria to be identified as the cause of an important infective disease, led many observers to approach the study of new bacterial species with a bias towards identifying any morphologically differentiated element within the bacterial cell as a spore, or its equivalent. Much of the earlier literature is, for this reason, full of mistaken allusions to spores or sporogenous granules in various bacteria. We now know that the formation of endospores is an important distinguishing characteristic of certain bacterial groups.

The great majority of sporing bacteria which have been adequately studied are monosporous, that is only one spore occurs in any one cell. A few large forms which have been observed by cytologists are disporous.

The mode of formation of the spore has been briefly referred to in discussing the nuclear apparatus of the cell. There is general agreement that it is formed by a localized accumulation of chromatin, or of a chromatin like substance, which at first stains deeply with nuclear stains, but which later becomes surrounded by a spore-wall or membrane, which is impervious to stains in the absence of special preliminary treatment, so that the ripe spore appears as an unstained, highly refractile body, spheroidal or ovoid in shape. As regards the finer details of spore-formation there is less agreement,

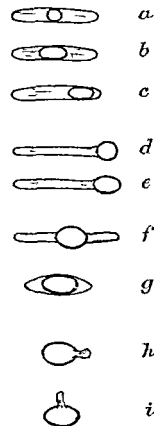


FIG. 6

- a-c Without distortion of bacterial cell  
 a Spherical equatorial  
 b Oval equatorial  
 c Oval subterminal  
 d-g With distortion of bacterial cell  
 d Spherical terminal  
 e Oval terminal  
 f Oval equatorial  
 g Oval equatorial  
 h and i Germination of spores  
 h Polar germination  
 i Equatorial germination.

and it would seem possible that the process differs to some extent in different bacterial species. Many observers have described the appearance, at the commencement of sporulation, of a clear vacuolar area, corresponding in size and shape to the space occupied by the ripe spore, and the subsequent

accumulation, within this area, of the nuclear material of which the spore is formed. Meyer and his supporters lay great stress on the appearances seen during sporulation, as evidence for the existence of differentiated nuclei. Those who have studied the internal structure of bacterial cells have, for the most part, included the process of spore formation in their observations, and the papers which have already been referred to contain full discussions of this question.

The situation of the spore within the bacterial cell, which may be terminal, subterminal, or approximately equatorial, gives a distinctive morphology to many bacterial species. In addition the diameter of the ripe spore is often greater than that of the bacillus which contains it, so that the cell is distorted and according to the position of the spore "drum stick" "barrel" or other irregular forms may be presented (Fig. 6).



FIG. 7. Terminations of spores by equatorial rupture.

A. An unidentified acid bacillus. B. *B. megaterium* (C. G. G.).  
(From photomicrographs kindly supplied by Dr. C. F. H. Davis.)

cytoplasm between the central cytoplasmic core and the spore membrane, and considers that this rather than the spore membrane itself is responsible for the refractility commonly seen in unstained preparations of spores.

Most of the available evidence indicates that the spore is simply a resting stage of the bacterial cell in which it is far more resistant to adverse environmental conditions than it is in the ordinary vegetative form. The spores of bacteria of the *Bacillus* group have a bound water content of the order of 60-70 per cent as compared with 3-21 per cent in the vegetative cell (Henry and Friedman 1937, Friedman and Henry 1938). By spectrochemical analysis Curran, Brunstetter and Myers (1913) found that the spores contained substantially more Ca, more Cu and Mn but less P and K than the vegetative cell. They suggest that heat resistance is associated with though not necessarily immediately dependent on high bound water and Ca content of the spore. The protoplasm of the spore differs materially from that of the vegetative cell both Howie and Cruickshank (1910) and Lamanna (1910) have demonstrated the antigenic dissimilarity of spores and their parent vegetative forms (see Chapter 8).



FIG. 9—The cell wall of *B. megatherium* after plasmolysis, showing transverse septa, both complete and incomplete ( $\times 3,500$ ).  
(From a photograph kindly supplied by Dr. C. F. Robinow.)

The resting spore retains its capacity to germinate for long periods. Graham Smith (1911) for example noted the survival for over seventeen years of the dry spores of *B. anthracis* kept in diffuse daylight at room temperature while Wilson and Shipp (1938) were able to grow sporing bacilli from preserved meats that had been sealed in containers for 114 years.

There is little evidence that spore-formation is associated with any sexual reproductive process. Certain appearances which have been interpreted in this sense by Schaudinn (1909) and Růžicka (1909) have been shown by Dobell (1911) to have no such significance.

However from their careful studies of the nuclear structure of a spore-bearing bacillus (see above) Allen, Appleby and Wolf (1939) concluded that the spore was not a resting stage but provided an opportunity for a re-arrangement of nuclear material. In this connection it is noteworthy that Kaplan and Williams (1941) after exploring the cultural conditions which induced spore formation in *C. sporogenes* concluded that spore formation was a natural phenomenon in the developmental cycle of the organism. Elieneberger (personal communication) has observed in several species of clostridia a process in which the dumbbell chromatonic bodies present in filamentous forms of the bacteria join into axial structures. These then divide into four chromatonic bodies, one of which matures into the nuclear body of the spore while the remaining three disintegrate. The process suggests a form of autogamy and that the spore is more than a resting stage in the life history of the bacillus.

**The Cell Wall and the Cytoplasmic Membrane**—There seems to be general agreement that bacterial cells may be differentiated with varying degrees of distinctness into two zones which have been variously called cell membrane and protoplasm, ectoplasm and endoplasm, and even cortex and medulla. Until a



A



B

FIG. 10.—Electron Micrographs of Bacteria.

A *Streptococcus pyogenes* showing the continuity of the cell wall along the length of the chain ( $\times 30,000$ ) (Mudd and Lockman 1941)

B *Bacillus cereus* at the junction of two cells, possibly an inner three layers of cytoplasm and an outer, continuous cell wall ( $\times 36,000$ ) (Johnson 1944)

(From micrographs kindly supplied by Dr. Stuart Mudd and Dr. F. H. Johnson.)

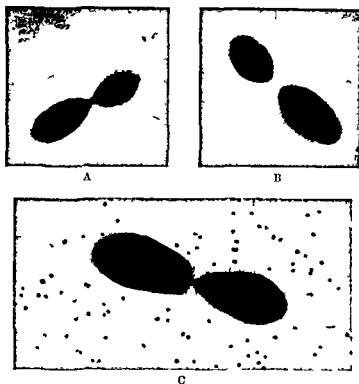


FIG. 11.—Electron Micrographs of Pneumococci ( $\times 11,000$ )

A and B showing the capsule as a delicate light halo and continuity of protoplasm in between the two cocci in A and of cell wall in B (Mudd *et al.* 1943a)  
 C showing the capsule as a demarcated halo (Mudd *et al.*, 1943b)

(From micrographs kindly supplied by Dr. Stuart Medd.)

few years ago there was some doubt whether a definite external membrane limited the ectoplasm (Legroux 1920; Legroux and Margrou 1930). The microdissection of bacteria performed by Wamoscher (1930) suggested a definite elastic outer layer distinct from the inner material of the bacterium.

Fischer (1894) studied the effect of hypertonic salt solutions on bacteria. The cell contents were observed to shrink leaving what appeared to be a rigid cell wall of the same shape as the bacterial cell before treatment. These plasmolytic experiments have since been repeated by many observers (see for example Knaysi 1930) but their significance as evidence for a definite limiting membrane in the bacterial ectoplasm has been obscured by the possible artificial nature of the structures produced. The differences of opinion (Legroux 1920; Legroux and Margrou 1930) are to a large extent resolved by results of recent demonstrations by various independent techniques of a definite cell wall.

The changes in the cell wall during the division of bacilli have been studied in detail by Knaysi (1941). Prior to the division of an elongated mother cell into two daughter cells he observed first a break in the cytoplasmic membrane and a centripetal movement of the cytoplasm. At the line of demarcation between the cytoplasmic masses of the two daughter cells a double intercellular plate was deposited which formed the two adjacent end walls of the completed daughter cell. In a yeast the deposition of the two walls preceded the cytoplasmic division.

According to Knaysi, the frequently reported constriction of the cell wall prior to division is an optical illusion, depending on a constriction of the cytoplasm. The double cell wall is well shown in the electron micrograph of *B. anthracis* made by Mudd and others (1941).

Robinow (1944) also obtained evidence that the transverse divisions of the cytoplasm of dividing bacillary cells are laid down before the bacilli divide in the accepted sense of the term. In some of his preparations (Fig. 9) the transverse walls between daughter cells appear to be growing inwards as annular diaphragms along the plane of the cytoplasmic cell boundary (personal communication).

Electron micrographs show a cell wall that retains its form even after the cells have been fractured and their contents dispersed, by ultrasonic vibration (Mudd *et al.* 1941). A rigid cell wall extending along the length of chains of cocci, and connecting adjacent cells in chains of bacilli, was observed by similar means (Figs. 10A, B, 11B) (Mudd and Lackman 1941, see also Fruhbrot and Ruska 1940, Mudd *et al.* 1942, Johnson *et al.* 1943).

The surface of the inner protoplasm takes up stain with greater avidity than the cell wall or the inner protoplasm itself. This, according to Knaysi (1938), is the cytoplasmic membrane, made up chiefly of surface active materials, lipoids and lipoproteins, which must be clearly distinguished from the rigid semipermeable cell wall.

**Capsules**—Many species of bacteria, such as the pneumococcus, the pneumobacillus and the anthrax bacillus, are characterized by the ability to develop a well marked capsule, which may be observable in stained or unstained preparations as a clear zone surrounding the bacterial cell or may after suitable fixing and mordanting, be stained by various methods. The degree to which the capsule is developed is largely determined by the environmental conditions. Thus, the pathogenic capsulated bacteria show their maximal capsule formation when growing in the animal tissues (Babes 1895, Gruber and Futak 1907, Preisz 1907, Bai 1908, Sauerbeck 1909 a, b, Eisenberg 1903, 1908, 1909). When such bacteria are grown in artificial culture there is usually a high correlation between the degree of capsulation and the content of the medium in unaltered, or slightly altered, animal protein.

It is possible that the response to a medium containing animal protein is a reaction to an unfavourable environment. For instance the pneumobacillus forms capsules in artificial culture when the phase of maximum proliferation has ceased (Hoogerheide 1938). On the other hand, the capsules of a strain of *Str. pyogenes* developed only during its active growth phase in a serum enriched medium (Morrison 1941).

The capsule is usually regarded as being formed by a thickening and alteration in consistency of the outer layer of the bacterial cell. It was however held (Meyer 1912, Zettnow 1918) that the capsular substance was an active secretion. Certain bacteria secrete a mucilaginous material in which large numbers of neighbouring cells are embedded, and some organisms, such as the Type III pneumococcus form this extracellular mucilaginous material in addition to possessing a well marked capsule (Fig. 11). It seems, however, more convenient to differentiate between the capsulated and the so called "mucoid" types of bacteria (see Ettinger Tulczynska 1933), and the fact that capsular material may frequently become separated from the cell can hardly be taken as a proof that it is a secretion in the ordinary sense of that term. But the sharp delimitation of the opaque,

rigid cell membrane from the space presumably occupied by the capsule in electron micrographs and the low density to the electron beam of the capsules themselves (Fruhbrot and Ruska 1940 Muller Heinmetts and Anderson 1943) together with the known delimitation of the cytoplasmic membrane inside the cell wall suggest that the hypothesis of a secreted capsule is the more likely to be valid. We may note that we have within recent years acquired a considerable knowledge of the chemical constitution of bacterial capsules. The capsules of the different types of pneumococci are for instance composed of complex poly-accharides (see p 279) while the capsule of the anthrax bacillus appears to be composed of a polypeptide material (see p 281).

Whether the capacity to form a capsule is really confined to those species that are typically capsulated is very doubtful. Many observers have described capsule formation by normally non-capsulated forms under particular environmental conditions and mucoid variants of normally non-mucoid species are of relatively common occurrence (see Chapter 9). The absence of a detectable capsule does not exclude the existence of thin capsules distinct from the cell wall. For example in a coccobacillus as small as *Br. nelutensis* the hypopolysaccharide material comprising 10 per cent of the whole bacillus if confined exclusively to an external covering would form a layer less than 20  $m\mu$  thick (Miles and Pirie 1939).



FIG 1<sup>a</sup>—*Zopfus enteri*  
Stained to show flagella ( $\times 1,100$ )

**Flagella.**—A large number of

bacteria including a few coccoid forms, many bacilli and most known spirilla and vibrios are more or less actively motile by means of flagella. These flagella are long thread-like processes arranged in various ways on the bacterial cell. They vary greatly in length but are often longer than the organism to which they are attached. They are extremely slender (Fig 14a) recent measurements of the flagella of *Proteus vulgaris* and *Salmonella paratyphi* B under the electron microscope indicate a thickness of 20 to 50  $m\mu$  at any rate of the electron-opaque part of the structure (Pieharski and Ruska 1939). Flagella were first effectively demonstrated by Loeffler (1890).

According to the observations of Trenkmann (1890) Ellis (1902-03) Fuhrmann (1910) and Meyer (1912) the central portion of the flagellar thread passes through the cell membrane and is in direct connection with the cytoplasm, and Fuhrmann believes that the proximal extremity is connected with a granule of chromatin, which would then be analogous to the blepharoplast of protozoan flagellates. This conclusion is adversely criticized by Zettnow (1914). Whether the optical methods at present available allow any decision on such points as these seems doubtful.

The arrangement of the flagella on the bacterial cell may take one of four forms.

There may be a single flagellum at one pole, when the arrangement is called monotrichate. There may be a single flagellum at each pole the amphitrichate condition. There may be a bunch of flagella at one pole or more rarely at each, an arrangement known as lophotrichate. The flagella may be arranged indiscriminately over the bacterial cell when they are referred to as peritrichate (see Fig. 12). The cells of any one flagellated species are characterized by one, and one only, of these possible arrangements (see Fig. 13).

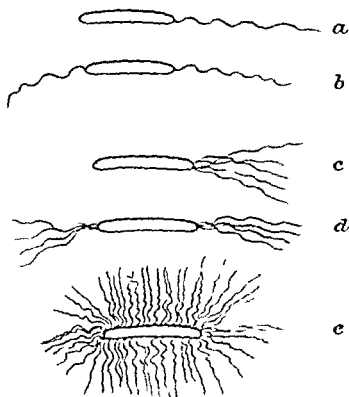


FIG. 13.—Diagram illustrating distribution of flagella

a. Monotrichate    b. Amphitrichate    c and d. Lophotrichate    e. Peritrichate

Peritrichous flagellation is regarded by Letchbourn (1934) as due to the occurrence of subpolar flagella on a chain of undivided cells. We have ourselves seen this phenomenon in the incompletely divided filamentous forms of monotrichate organisms but some exceedingly short cells have so many peritrichous flagella that it would be absurd to postulate their multiseptate nature merely to avoid the assumption of peritrichous flagellation.

Recently Wei (1937) and Papper (1938) have described the flagella of the typhoid bacillus as consisting of two equal spiral structures set up at an angle at the side of the body (Fig. 14c). Papper found that the two structures were fused into one propelling tail organ in fast-moving bacilli (B in the figure). He regards the two configurations of flagella as primary and secondary structures.



In dead bacilli the primary structures disperse into fine threads which he regards as post mortem artefacts not as peritrichous flagella which in life form a functional unit. It is however legitimate to regard the peritrichous flagellum as the primary structure for there are analogous cases of functional unity of structures that are



A



B



C

FIG. 14.—Bacterial Flagella.

- A. Electron micrograph of *Cl tetani* showing protoplasm, cell wall and flagella ( $\times 22,000$ )  
 B and C Dark ground photomicrographs of living *Salmon typhi* ( $\times 5000$ )

(From photographs kindly supplied by Dr. Stuart Mudd and Dr. A. Piper.)

made up of morphologically separate flagella. The undulating membrane of certain ciliate infusoria for instance consists of a fine row of cilia which can during life be separated by mechanical means into single cilia all moving in an unco-ordinated manner (Maier 1903 Chambers and Dawson 1925)

The nature of flagella and the modes of generating and transmitting to them the energy required for their propulsive effect is largely unknown. Under fixed cultural conditions the speed of a flagellated bacterium is considerable and to a certain extent characteristic of the strain observed. The highest speeds are recorded for the cholera vibrio, a monotrichate organism which may travel more than  $80 \mu$  per second (see Sansonetti 1919 Ogiuti 1936).

Though flagella are usually confined to vibrios and bacilli several observers have described motile cocci (Schublich 1932 Koblmüller 1935 Pownall 1935 Levenson 1938). Under the electron microscope flagella like structures have been seen in spirochetes of the genus *Treponema* (Morton and Anderson 1942).

The results that have been obtained in the detailed study of antigenic structure (see Chapter 8) have shown that the flagellar substance or at least that portion of it which forms the flagellar surface, is chemically distinct from the substances forming the cell body.

**Involution Forms**—In old cultures of many bacteria forms may be found which are quite unlike those seen in young and actively growing cultures. Some of these forms are very typical of the bacterial species in which they occur as is the case with the so-called involution forms of the plague bacillus and of some other species of *Pasteurella*, and the giant forms which occur in cultures of the meningococcus. The existence of these and other abnormal forms has long been recognized and they have been regarded as due to involution or degeneration following the ageing of the culture and the consequent unfavourable environmental conditions. This view as to their nature has been generally adopted because of the undoubted fact that under the conditions in which they usually occur the majority of the organisms are non viable while the extensive study of young actively growing cultures has in general shown no more striking departure from the normal form than the occurrence of unusually large cells, or occasionally in the rod forms of very long almost filamentous cells due apparently to delay in cell division.

**Reproduction**—Although a description of the method of reproduction in bacteria may be regarded as belonging more correctly to a discussion of their physiology than of their morphology yet the question is so closely bound up with differences in form of the cells and of the cell aggregates which result from successive division that there are many advantages in considering the question in the present chapter.

There is no doubt at all that the method of multiplication of bacteria under optimal conditions of cultivation in the laboratory, is by simple binary fission each cell dividing into two daughter cells by constriction with or without a well marked preliminary septation.

In the spheroidal forms division may occur in any diameter. In the ovoid or cylindrical forms division always occurs in the transverse diameter. It is never longitudinal.

In certain genera of bacteria such as the *Actinomyces* the formation of branching filaments is a normal phase of growth and multiplication. In a few bacillary species and notably in *C. diphtheriae* and *M. tuberculosis* rudimentary branching has frequently been described and its occurrence in actively growing cells observed under the microscope (Hill 1902). In most bacillary species such rudimentary branching is rare and is seldom observed in the examination of ordinary stained preparations. Hort (191'a b) and Gardner (1925) have however noted the occurrence of Y forms in actively growing micro-

scopic cultures of organisms of the coli typhoid group, and have observed multiplication by elongation and fission from all three points of the Y (See also Wyckoff 1934) Whether the occurrence of these forms has any special significance is a question that the future must determine

Spore formation, which has been described above, is confined to certain species of bacteria. The spore constitutes a special resting and resistant phase of the bacterial cell, its formation appears to be determined in large part by environmental factors, and there is little ground for regarding it as an integral part of any periodic or essential life-cycle (but see p. 26)

A question still at issue is the occurrence of other forms of reproduction, and in particular of the existence or non-existence of a complex life-cycle, in which sexual processes may or may not play a part. It is not possible to discuss this problem at all fully. It must, however, be noted that there are those who believe that such a life-cycle is a natural characteristic of all bacterial species, and that multiplication by binary fission, which appears to be the only mode of reproduction in young, actively growing cultures, is only one phase of the cycle, which we encourage by the particular artificial conditions under which we choose to study bacterial cells.

Jones (1913, 1914, 1920) described reproduction by gonidia-like bodies in the case of *Acetobacter*. These observations were greatly extended by Löhnis and Smith (1916) who described a far more complex life-cycle in this, and in other, species, and applied their results to the reproduction of bacteria in general (Löhnis 1921). Stoughton (1929, 1932) in describing the morphology and cytology of *Bact. malvarum*, an organism producing angular leaf-spot in the cotton plant, records the formation, by budding from the bacillary form, of coccus-like bodies that subsequently develop into bacilli, and also the fusion of bacillary cells with the formation of zygozooids.

Almqvist described various forms of reproductive bodies, mainly minute gonidial structures, in a wide range of bacteria (Almqvist 1893-1924, Almqvist and Korsen 1918, Korsen 1918). It may be noted that there was a general tendency for these appearances to be observed in old cultures, and sometimes in cultures which had been allowed to undergo a marked degree of drying. However, Allen, Appleby and Wolf (1939) observed the relatively early liberation of intracellular granules from growing vegetative cells of their bacillus. A little later, minute rods were found in the culture, which appeared to have originated in the granule, and which later developed into the modal form of the bacillus. Mellon (1917-26) has described a variety of forms which he regards as stages in the complex life-cycle of various bacterial species. Enderlein (1925) upholds a similar thesis. Kuhn (1929-1930) has described a remarkable series of morphological forms in common organisms some of which are interpreted as evidence of a life-cycle, others of which—the so-called Pettenkofer bodies—are regarded as intracellular parasites. Hadley (1927, see also Hadley, Delves and Klimek 1931) in a detailed review, which should be consulted by those who desire further information on this subject, sets out the available data with regard to the occurrence and possible significance of these various forms, and seeks to relate them to the phenomenon of bacterial dissociation, which we shall discuss in a subsequent chapter. A shorter discussion of the possible modes of bacterial reproduction will be found in papers by Bergstrand (1920, 1921, 1923).

It is, we think, impossible on the evidence at present available to come to any conclusion on the general problem at issue. That a cycle of morphological changes may characterize the life history of a particular bacterial species under natural conditions there can be no doubt. The detailed studies of Bewley and Hutchinson (1920) and of Thornton and Gangulee (1926) on *Rhizobium leguminosarum* provide a case in point, though their conclusions have been questioned by Lewis (1933).

In regard to bacteria in general, there is clearly no justification for ignoring such observations as those recorded above, but the thesis, that the bacterial forms with which we are most familiar in artificial cultures are only stages in some more complex cycle of development, has not in our view been clearly and definitely established. All bacteriologists are familiar with the so-called "involution forms" to which we have already referred, and there is little doubt that many of these forms represent dead or dying cells. It may well be that we have been too ready to refer to this category every abnormal form encountered, and few would deny that some of the appearances that are met with are strongly suggestive of some more complex form than the simple bacillus. The problem is, however, beset with technical difficulties arising from the small size of the organisms concerned. The crucial test must remain the actual observation of the reproduction of typical bacterial cells from the granules, spheres, or syncytial masses which have been described as stages in the complete life-cycle, and the unequivocal demonstration of this cyclical development demands the isolation of single cells, not only at the commencement of the cycle, but at each successive stage. No harm will be done by maintaining a severely critical attitude at the present stage of the controversy, so long as each new piece of evidence is considered on its merits.

In this connection, the work of Klineberger and of Dienes may be noted. Klineberger (1935) describes a minute pleuropneumonia like organism in symbiosis with *Streptobacillus moniliformis*, and considers that many of the morphological peculiarities that are commonly regarded as characteristic of the bacillus are, in reality, due to the presence of the symbiont. She suggests that some, at least, of the pleomorphic elements in bacterial cultures that have been described as belonging to a sexual or developmental cycle may have a similar origin. This view has been developed in subsequent publications (see Chapter 40). Dienes (1942), on the other hand, believes that the pleuropneumonia like organisms are part of a life cycle not only in the streptobacillus, but in other bacterial species. He has succeeded in inducing their appearance in cultures of *Bact. coli*, *Hemophilus influenzae*, *Flavobacterium*, the gonococcus and *F. funduliformis* (see also Klineberger 1942). Dienes (1943) and Smith (1942, 1943, 1944), have observed in *Streptobacillus moniliformis* and *F. funduliformis* the formation from the bacterial cell of 'large bodies' which may either segment and give rise to bacterial forms, or develop into colonies of pleuropneumonia like organisms, and state that the large bodies appeared in some cases to arise from the conjugation of two bacterial cells.

We do not propose to enter into the vexed controversy over the existence of filtrable forms of bacteria. The whole crux of the question seems to us to lie in the definition of the term "filtrable," and in the properties of the filter passing bodies. Bacteria of the same species are known to vary greatly in size under different environmental conditions and at different stages of growth. That certain forms may be small enough to pass through our relatively crude earthenware filters is hardly to be doubted, but whether these forms differ in any of their essential properties from larger members of the same species, and in particular whether they constitute some essential stage in the reproductive cycle of the organism, is still undecided. In our opinion the evidence that has so far been produced does not suggest that they are differentiated in any important respect other than size, and probably this differentiation is of a continuous rather than a discontinuous type.

**The Formation of Cell Aggregates**—When a bacterial cell divides the two daughter cells may at once part company, or they may remain attached to one another by their cell membranes. When this incomplete separation persists during many successive cell divisions, cell aggregates are produced, the form of which will depend upon the planes in which successive divisions take place, and upon the number of cell-divisions which occur before the cell aggregate begins to

separate. The aggregates which are formed in this way are often highly characteristic and constitute an important factor in the identification of bacterial species. In the spheroidal forms division may occur in any plane while in the cylindrical forms it always occurs in the transverse diameter. As a result the cocci may and do show a greater variety of groupings than the bacillary or spiral forms.

When the successive divisions in a coccal bacterium occur in such a way that the daughter cells remain united in pairs for a short period but these pairs separate before a further division occurs the organism in question is described as a diplococcus. When several successive divisions occur before separation of the resulting aggregate and these divisions follow no ordered sequence as regards the planes in which they take place irregular groups of cocci result which have been compared to bunches of grapes. Organisms which form this type of cell aggregate are known as staphylococci and this term is used as a generic name. When such a series of divisions without separation occurs in planes parallel to one another, the aggregates so formed have the appearance of a chain or chaplet, and organisms which behave in this way are known as streptococci and are usually classed in a single genus. In some species the cells remain attached while two divisions occur the second at right angles to the first. The resulting aggregate is a group of four cocci such an organism is sometimes referred to as a tetracoccus. In some species the typical cell grouping is produced by three divisions, the plane of each being at right angles to the other two. In this way cubes or packets each of eight cocci are produced. The species which form groupings of this type are classed in the genus *Sarcina*.

Among the cylindrical forms the only possible departure from the single-cell formation is the adherence of two or more cells in pairs or chains. Such groupings may be referred to as diplobacilli or streptobacilli but they are not sufficiently constant in a particular bacterial species to be used for purposes of classification. They occur however far more often in some species than in others and their relative frequency may be an important specific character.

Apart from the formation of such united cell aggregates the way in which a cell divides influences the grouping of the daughter cells. In certain bacillary forms such as the diphtheria bacillus, division appears to occur asymmetrically, in the sense that the daughter cells remain attached at one side of the cylinder after division has proceeded across the whole width of the organism from the opposite side. In the early stages of the subsequent growth of the daughter cells this local attachment seems to act as a hinge about which the elongating cells swing so that they come to lie at varying angles to one another depending on the period which elapses before division becomes complete. The groupings so formed, which have been compared to Chinese letters or to cuneiform characters, are very characteristic of certain species.

**Colony Formation.**—When bacteria are grown on solid media and care is taken to avoid too heavy an inoculation the individual cells multiply and form isolated colonies. The appearance of these colonies is in many cases, highly characteristic of the group or species to which a given bacterium belongs. We know little of the internal structure of bacterial colonies but that little suggests that the structural differentiation is considerable (Legroux and Margron 1920 Ravich Berger and Svinkina 1933) and it is possible that much light may be thrown on bacterial morphology by further study along these lines.

Two colonial characteristics may be noted here. Certain motile bacilli swarm in continuous films of growth on the surface of solid media, a property which is displayed characteristically by members of the *Proteus* and *Clostridium* groups (see Chapters 27 and 36). Other motile bacilli, notably among aerobic and anaerobic sporing bacilli, form rounded colonies in which the constituent bacilli are moving in a clockwise or anticlockwise direction, imparting a rotating motion to the whole colony. As a result of the rotation, the colony wanders over the surface of the solid medium, usually leaving a track in the form of a double line of bacilli, apparently thrown off from the sides of the advancing colony (see Muto 1904, Roberts 1935, Smith and Clark 1933, Russ Munzer 1933, Shinn 1938, Turner and Lake 1941, Fuller and Norman 1943).

The character of the colonies produced by a given bacterial species is of great importance in identification, and is, for this purpose, best considered in conjunction with other growth characters, such as the appearances noted in cultures in liquid media, or in the so-called stab cultures, in which inoculation is made by thrusting a platinum wire axially into a column of solid medium, such as agar or gelatin. Further description of colony structure is therefore deferred to Chapter 13.

**Staining Reactions**—The staining reactions of bacteria, and particularly their response to certain differential stains, might well be regarded as more in the nature of microchemical tests, than methods of demonstrating structure. It would, therefore, be more logical to consider staining reactions in connection with the physiology of the cell than with its morphology, but the way in which a given bacterium reacts to special stains constitutes such an important part of the general picture which we form of it, that it is convenient to deal with this matter in the present chapter.

**Reactions to General Bacterial Stains**—Quite apart from the presence of the metachromatic granules, or of other granular material, different bacterial cells may differ in the way in which they take up stains. In some cases the whole cell may be uniformly coloured. In some cylindrical forms the ends of the bacillus may be deeply stained, while the central portion may be almost colourless. This constitutes the so-called polar staining, which is very characteristic of the plague bacillus, and is found in many other bacterial species. Sometimes the cell may stain unevenly throughout its length, so that barred or beaded forms may occur, a feature which is very characteristic of the diphtheria bacillus and many allied organisms, and which occurs to a less marked extent with many acid fast bacilli. In some cases, in which barred or beaded forms are found, these conditions are associated with the presence of metachromatic granules, but these granules are not the essential cause of such irregularities in staining. It seems clear that, in such forms, there is an unequal distribution throughout the cell of those constituents of the cytoplasm which take up the stain employed, and it has frequently been suggested that they are indicative of plasmolytic changes. It seems doubtful whether plasmolysis can be invoked as a general explanation of this phenomenon, though it is true that there is a general tendency for the frequency of granular and barred forms to increase with the age of the culture, and to be notably scanty in very young cultures. They are, however, so plentiful in young and actively growing cultures of certain species that it seems impossible to regard them as resulting entirely from degenerative changes.

**The Differential Staining of Bacterial Species**—In the application of numerous stains, mordants, and differential decolorizing reagents to the study of bacteria,

the empirical discovery has been made that certain staining reactions are highly characteristic of certain groups of bacteria. Two such staining reactions are especially important in this respect.

*The Gram Stain*—Gram (1884) described a staining method which has been of the greatest service in differentiating bacterial groups, and which has been extensively studied, and frequently modified, by subsequent workers. This reaction depends on the fact that, when certain bacteria are stained with certain aniline dyes, such as gentian violet, methyl-violet and others, and are subsequently treated with a solution of iodine in potassium iodide, a mordanting action occurs which prevents the subsequent decolorization of the bacteria on treatment with alcohol. Other bacteria, after similar treatment, are readily decolorized. This difference between the retention of the stain by Gram positive bacteria and its loss by Gram negative forms is correlated with certain other characters. Thus (Kruse 1910) Gram negative organisms are more susceptible to solution in alkalies, or to digestion by enzymes, than are Gram positive organisms, they are also more susceptible to lysis by an immune serum in the presence of complement. According to Stearn and Stearn (1930), they are more resistant to the lethal action of oxidizing and perhaps of reducing agents. Brudny (1908) and Eisenberg (1909) would ascribe the difference between Gram positiveness and Gram negativeness to a difference in the permeability of the cell membrane; while Sander (1935), who has been able to render Gram positive bacteria Gram negative by treatment with a variety of reagents and to demonstrate that this change is reversible, regards the determining factor as the state of dispersion of the bacterial protoplasm. Some observers, on the other hand, and notably Deussen (1918), incline towards a more purely chemical theory, and Stearn and Stearn (1928) regard differences in intracellular pH as the determining factor.

Recently Henry and Stacev (1943) by extraction with bile salt, succeeded in removing from Gram positive organisms a substance they identify as magnesium ribonucleate, and in thereby rendering the organisms Gram negative. If the organisms are protected from oxidation, the ribonucleate will recombine and the Gram-staining property will be restored. Gram negative organisms do not yield the salt and do not combine with it to become Gram positive. The Gram positive material appears to be a high molecular complex of a reduced basic protein and magnesium ribonucleate. It was noted too that removal of the ribonucleate facilitated observation of stained nuclear bodies, a fact which supports Knaysi's contention that the Gram staining material is concentrated at the cytoplasmic membrane. The difference between Gram positive and Gram negative organisms is particularly evident in their susceptibility to various groups of antibacterial agents, both inorganic and organic (see Chapter 6), and it is probable that investigation of these biochemical differences will lead to a more precise elucidation of a staining reaction first introduced on an entirely empirical basis.

In employing Gram's method it must always be remembered that the differentiation is not absolutely sharp and specific as regards a given bacterium at all stages of its growth, or as regards all bacterial species. Those organisms which are completely Gram negative never retain the stain, but those organisms which are Gram positive frequently fail to retain the stain when preparations are made from old cultures. This is indeed easy to understand, since such cultures consist largely of dead, dying or degenerate cells, whose physical and chemical properties must be greatly altered. Certain bacterial species show reactions to this method of staining which are of an intermediate type, with the result that they are extremely sensitive to small changes in technique, sometimes appearing to be Gram positive and at others Gram negative. Due allowance must be made for all these points in determining the reaction of any given species.

*Acid fastness*—The fact that certain bacteria, after being stained with warm solutions of fuchsin, resist the decolorizing action of strong mineral acids was observed by Ehrlich (1882) and confirmed by Ziehl (1882, 1883) who modified the technique of staining. This reaction is highly characteristic of the tubercle bacillus, and of an allied group of organisms

which are collectively known as the acid fast bacilli. It is of interest to note that these bacilli are peculiarly resistant to the action of such solvents as strong solutions of alkalis or mixtures of alkalis and sodium hypochlorite (antiformin), as also to the action of digestive ferments. In this connection it may be noted that acid fast bacilli are also Gram positive, though the majority of Gram positive organisms are not acid fast. The substance which confers this property of acid fastness on the tubercle bacillus and allied forms has been studied by Klops (1896), Koch (1897), Bulloch and Macleod (1904) and Tamura (1913). It was at first regarded as an unsaturated fat, but is now generally believed to be lipoidal or waxy in nature. Bulloch and Macleod state that it has the chemical properties of an alcohol, and Tamura isolated an alcohol which had the property of acid fastness, and which he named "mykol" (For further details see Chapter 16).

## REFERENCES

- ALLEN, L. A., APPLEBY, J. C. and WOLF, J. (1939) *Zbl Bakt. Hte Abt.* 100, 3.  
 ALMQUIST, A. and KORAEV, G. (1918) *Z. Hyg. Infektkr.* 85, 347.  
 ALMQUIST, F. (1893) *Z. Hyg. Infektkr.* 15, 293. (1901) *Zbl Bakt.* 37, 18. (1905) *Z. Hyg. Infektkr.* 52, 179. (1907) *Zbl Bakt.* 45, 401. (1911) *Ibid.*, 60, 167. (1917) *Z. Hyg. Infektkr.* 83, 1. (1922) *J. infect. Dis.* 31, 483. (1924a) *Ibid.* 35, 341. (1924b) *Z. Hyg. Infektkr.* 101, 15.  
 AMATO, A. (1909) *Zbl Bakt.* 48, 385.  
 ASCOLI, (1901) *Dtsch. med. Wochr.* 20, 313.  
 BATES, V. (1889) *Z. Hyg. Infektkr.* 5, 173. (1895) *Ibid.* 20, 412.  
 BAXIAN, J. (1933) *Arch. Mikrobiol.* 4, 409.  
 BAIL, O. (1908) *Zbl Bakt.* 45, 488.  
 BARNARD, J. E. (1919) *J. R. micr. Soc.* p. 1. (1925) *Lancet* ii 117. (1930) "A System of Bacteriology," Med. Res. Council London, 1, 115.  
 BEBE, J. M. (1941) *J. Bact.* 42, 193.  
 BERGSTRAND, H. (1920) *J. infect. Dis.* 27, 1. (1921) *Johns Hopk. Hosp. Bull.* 32, 234. (1923) *J. Bact.* 8, 365.  
 BEWLEY, W. T. and HUTCHINSON, M. B. (1920) *J. agric. Sci.* 10, 144.  
 BORRIES, B. von, RUSKA, H. and RUSKA, H. (1938) *Klin. Wochr.* 17, 921.  
 BRUDNY, V. (1908) *Zbl Bakt. Hte Abt.* 21, 62.  
 BULLOCK, W. and MACLEOD, J. J. P. (1904) *J. Hyg. Camb.* 4, 1.  
 CHAMBERS, R. and DAWSON, J. A. (1925) *Biol. Bull.* 48, 240.  
 CHANCE, H. L. (1938) *J. Bact.* 35, 347.  
 COPE, T. (1875) *Beitr. Biol. Pflanz.* 1, 2.  
 CURRAN, H. R., BRUNSTETTER, B. C. and MYERS, A. T. (1943) *J. Bact.* 45, 480.  
 DEESSEN, E. (1918) *Z. Hyg. Infektkr.* 85, 235.  
 DIENES, L. (1942) *J. Bact.* 44, 37. (1943) *Proc. Soc. exp. Biol. N.Y.* 53, 84.  
 DIENES, L. and SMITH, W. F. (1942) *Proc. Soc. exp. Biol. N.Y.* 51, 237. (1943) *Ibid.* 53, 195. (1944) *J. Bact.* 48, 125.  
 DOBELL, C. (1911) *Quart. J. micr. Sci.* 56, 395.  
 DOMBROWSKY, K. H. (1936) *Zbl Bakt.* 137, 160.  
 FRIEDLICH, P. (1882) *Dtsch. med. Wochr.* 8, 269.  
 EISENBERG, P. (1903) *Zbl Bakt.* 34, 739. (1908) *Ibid.* 45, 44, 134, 638. (1909) *Ibid.* 49, 465.  
 FLEISCH, D. (1902-03) *Zbl Bakt.* 33, 1, 81, 161. (1922) *Brit. med. J.* ii 731.  
 ENDERLEIN, (1925) *Bakterien Cyclogenie*, etc. Berlin.  
 ERNST, P. (1888) *Z. Hyg. Infektkr.* 4, 25. (1893) *Ibid.* 5, 428. (1902) *Zbl Bakt. Hte Abt.* 8, 1, 31, 65, 97.  
 FTINGER-TOLOZYNSKA, R. (1933) *Z. Hyg. Infektkr.* 114, 769.  
 FEINBERG, (1900) *Zbl Bakt.* 27, 417.  
 FICKER, M. (1903) *Arch. Hyg.* 48, 171.  
 FISCHER, A. (1894) 'Untersuchungen über Bakterien' Berlin.  
 FRIDDMAN, C. A. and HEVRY, B. S. (1938) *J. Bact.* 36, 99.  
 FRÜHRRODT, L. and RUSKA, H. (1940) *Arch. Mikrobiol.* 11, 137.  
 LUBERMAN, T. (1910) *Zbl Bakt. Hte Abt.* 25, 129.  
 FULLER, W. H. and NORMAN, A. G. (1943) *J. Bact.* 46, 273.  
 GARDNER, A. D. (1925) *J. Path. Bact.* 27, 189.  
 GAUSS, C. J. (1901) *Zbl Bakt.* 31, 92.  
 GRAHAM-SMITH, G. S. (1941) *J. Hyg. Camb.* 41, 406.  
 GRAM, C. (1884) *Fortschr. Med.* 2, 185.  
 GRIMME, A. (1902) *Zbl Bakt.* 32, 1, 81, 161, 241, 321.



the empirical discovery has been made that certain staining reactions are highly characteristic of certain groups of bacteria. Two such staining reactions are especially important in this respect.

*The Gram Stain*—Gram (1884) described a staining method which has been of the greatest service in differentiating bacterial groups, and which has been extensively studied, and frequently modified by subsequent workers. This reaction depends on the fact that, when certain bacteria are stained with certain aniline dyes, such as gentian violet methyl violet and others, and are subsequently treated with a solution of iodine in potassium iodide, a mordanting action occurs which prevents the subsequent decolorization of the bacteria on treatment with alcohol. Other bacteria, after similar treatment, are readily decolorized. This difference between the retention of the stain by Gram positive bacteria and its loss by Gram negative forms is correlated with certain other characters. Thus (Krusse 1910) Gram negative organisms are more susceptible to solution in alkalis, or to digestion by enzymes, than are Gram positive organisms, they are also more susceptible to lysis by an immune serum in the presence of complement. According to Stearn and Stearn (1930) they are more resistant to the lethal action of oxidizing and perhaps of reducing agents. Brudny (1908) and Eisenberg (1909) would ascribe the difference between Gram positiveness and Gram negativeness to a difference in the permeability of the cell membrane, while Sander (1935) who has been able to render Gram positive bacteria Gram negative by treatment with a variety of reagents and to demonstrate that this change is reversible, regards the determining factor as the state of dispersion of the bacterial protoplasm. Some observers, on the other hand and notably Deussen (1918) incline towards a more purely chemical theory and Stearn and Stearn (1928) regard differences in intracellular pH as the determining factor.

Recently Henry and Stacey (1943) by extraction with bile salt, succeeded in removing from Gram positive organisms a substance they identify as magnesium ribonucleate, and in thereby rendering the organisms Gram negative. If the organisms are protected from oxidation the ribonucleate will recombine and the Gram-staining property will be restored. Gram negative organisms do not yield the salt and do not combine with it to become Gram positive. The Gram positive material appears to be a high molecular complex of a reduced basic protein and magnesium ribonucleate. It was noted too that removal of the ribonucleate facilitated observation of stained nuclear bodies—a fact which supports Knaysis' contention that the Gram staining material is concentrated at the cytoplasmic membrane. The difference between Gram positive and Gram negative organisms is particularly evident in their susceptibility to various groups of antibacterial agents, both inorganic and organic (see Chapter 6) and it is probable that investigation of these biochemical differences will lead to a more precise elucidation of a staining reaction first introduced on an entirely empirical basis.

In employing Gram's method it must always be remembered that the differentiation is not absolutely sharp and specific as regards a given bacterium at all stages of its growth, or as regards all bacterial species. Those organisms which are completely Gram negative never retain the stain, but those organisms which are Gram positive frequently fail to retain the stain when preparations are made from old cultures. This is indeed easy to understand, since such cultures consist largely of dead, dying or degenerate cells, whose physical and chemical properties must be greatly altered. Certain bacterial species show reactions to this method of staining which are of an intermediate type, with the result that they are extremely sensitive to small changes in technique, sometimes appearing to be Gram positive and at others Gram negative. Due allowance must be made for all these points in determining the reaction of any given species.

*Acid fastness*—The fact that certain bacteria after being stained with warm solutions of fuchsin resist the decolorizing action of strong mineral acids was observed by Ehrlich (1882) and confirmed by Ziehl (1882, 1883) who modified the technique of staining. This reaction is highly characteristic of the tubercle bacillus, and of an allied group of organisms

which are collectively known as the acid fast bacilli. It is of interest to note that these bacilli are peculiarly resistant to the action of such solvents as strong solutions of alkalis, or mixtures of alkalis and sodium hypochlorite (antiformin), as also to the action of digestive ferments. In this connection it may be noted that acid fast bacilli are also Gram positive, though the majority of Gram positive organisms are not acid fast. The substance which confers this property of acid fastness on the tubercle bacillus and allied forms has been studied by Klebs (1890), Koch (1897), Bulloch and Macleod (1904) and Tamura (1913). It was at first regarded as an unsaturated fat, but is now generally believed to be lipoidal or waxy in nature. Bulloch and Macleod state that it has the chemical properties of an alcohol, and Tamura isolated an alcohol which had the property of acid fastness, and which he named "mykol." (For further details see Chapter 16.)

## REFERENCES

- ALLYN, L. A., AFFLEBY, J. C. and WOLF, J. (1939) *Zbl Bakt*, IIte Abt., 100, 3  
 ALMQUIST, A. and KORAEN, G. (1918) *Z Hyg InfektKr*, 85, 347  
 ALMQUIST, E. (1893) *Z Hyg InfektKr*, 15, 293, (1904) *Zbl Bakt*, 37, 18, (1905) *Z Hyg InfektKr*, 52, 179, (1907) *Zbl Bakt*, 45, 491, (1911) *Ibid*, 60, 167, (1917) *Z Hyg InfektKr*, 83, 1, (1922) *J infect Dis*, 31, 483, (1924a) *Ibid*, 35, 341, (1924b) *Z Hyg InfektKr*, 101, 15  
 AMATO, A. (1904) *Zbl Bakt*, 48, 385  
 ASCOLI (1901) *Dtsch med Wochr*, 20, 313  
 BADES, V. (1889) *Z Hyg InfektKr*, 5, 173, (1893) *Ibid*, 20, 412  
 BADIAT, J. (1933) *Arch Mikrobiol*, 4, 409  
 BAIL, O. (1908) *Zbl Bakt*, 48, 488  
 BARNARD, J. E. (1919) *J R micr Soc*, p. 1, (1925) *Lancet* ii 117, (1930) "A System of Bacteriology," Med Res Council London, 1, 115  
 BEEBE, J. M. (1911) *J Bact*, 42, 193  
 BERGSTRAND, H. (1920) *J infect Dis*, 27, 1; (1921) *Johns Hopk Hosp Bull*, 32, 234 (1923) *J Bact*, 8, 365  
 BEWLEY, W. F. and HUTCHINSON, M. B. (1920) *J agric Sci*, 10, 144  
 BORNIES, B. von, RUSKA, F. and RUSKA, H. (1935) *Klin Wochr*, 17, 921  
 BRUDNY, V. (1908) *Zbl Bakt*, IIte Abt., 21, 62  
 BULLOCH, W. and MACLEOD, J. J. R. (1904) *J Hyg, Camb*, 4, 1  
 CHAMBERS, R. and DAWSON, J. A. (1925) *Biol Bull*, 48, 240  
 CHANCE, H. L. (1938) *J Bact*, 35, 347  
 COHN, F. (1875) *Beitr Biol Pflanz*, 1, 2  
 CURRAN, H. R., BRUNSTETTER, B. C. and MYERS, A. T. (1943) *J Bact*, 45, 485  
 DEESSEN, E. (1918) *Z Hyg InfektKr*, 85, 235  
 DIENES, L. (1942) *J Bact*, 44, 37, (1943) *Proc Soc exp Biol NY*, 53, 84  
 DIENES, L. and SMITH, W. E. (1942) *Proc Soc exp Biol*, NY, 51, 297, (1943) *Ibid*, 53, 195, (1944) *J Bact*, 48, 125  
 DOBELL, C. (1911) *Quart J micr Sci*, 56, 395  
 DOMBROWSKY, K. H. (1936) *Zbl Bakt*, 137, 160  
 EHRLICH, P. (1882) *Dtsch med Wochr*, 8, 269  
 EISENBERG, P. (1903) *Zbl Bakt*, 34, 739, (1908) *Ibid*, 45, 44, 131, 638, (1909) *Ibid*, 49, 463  
 FELLIS, D. (1902-03) *Zbl Bakt*, 33, 1, 81, 161, (1922) *Brit med J*, ii 731  
 ENDERLEIN (1925) "Bakterien Cyklogenie, etc." Berlin  
 ERNST, P. (1888) *Z Hyg InfektKr*, 4, 25, (1899) *Ibid*, 5, 428, (1902) *Zbl Bakt*, IIte Abt., 8, 1, 34, 65, 97  
 ETINGER TULCZYNSKA, R. (1933) *Z Hyg InfektKr*, 114, 769  
 FEINBERG (1900) *Zbl Bakt*, 27, 417  
 FICKER, M. (1900) *Arch Hyg*, 48, 171  
 FISCHER, A. (1894) "Untersuchungen über Bakterien." Berlin  
 FRIEDMAN, C. A. and HENRY, B. S. (1938) *J Bact*, 36, 99  
 FRÜHRODT, F. and RUSKA, H. (1910) *Arch Mikrobiol*, 11, 137  
 FUCHSMANN, F. (1910) *Zbl Bakt*, IIte Abt., 25, 129  
 FULLER, W. H. and NORMAN, A. G. (1943) *J Bact*, 48, 273  
 GARDNER, A. D. (1925) *J Path Bact*, 27, 189  
 GAUSS, C. J. (1903) *Zbl Bakt*, 31, 92  
 GRAHAM SMITH, G. S. (1941) *J Hyg, Camb*, 41, 496  
 GRAM, C. (1884) *Fortschr Med*, 2, 185  
 GRIMME, A. (1902) *Zbl Bakt*, 32, 1, 81, 161, 241, 321

- GRÖN, F (1938) *Zbl Bakt*, 141, 220
- GRUBER, M and FUTAKI K (1907) *Munch med Wochr*, 54, 249
- GUILLIERMOND, A. (1906) *Bull Inst Pasteur*, 4, 145, (1907) *Ibid.*, 5, 273, 321, (1933) *C R Soc Biol* 113, 1095
- HADLEY P (1927) *J infect Dis*, 40, 1
- HADLEY P, DELVES E and KLIMKE, G (1931) *J infect Dis*, 48, 1
- HENRY B S and FRIEDMAN, C A (1937) *J Bact* 33, 323
- HENRY H and STACEY, M (1943) *Nature Lond* 151, 671
- HILL, H W (1902) *J med Res*, 7, 115, 202
- HOLLANDE, A C (1934) *Arch Protistent*, 83, 465
- HOLLANDE A. C and HOLLANDE G (1932) *Arch Zool exp gen*, 72, No 6
- HOOGERHEIDE J C (1939) *J Bact*, 38, 367
- HORT E C (1917a) *Brit med J*, 1, 571, (1917b) *Ibid.*, 11, 377
- HOWE, J W and CRICKSHANK, J (1940) *J Path Bact*, 50, 235
- IMŠENECKI A (1936) *Zbl Bakt Ilte Abt.*, 94, 330
- JOHNSON F H (1944) *J Bact* 47, 551
- JOHNSON F H, ZWORYKIN N and WARREN, G (1943) *J Bact.*, 46, 167
- JONES D H (1913) *Zbl Bakt, Ilte Abt*, 38, 14, (1914) *Ibid.*, 40, 170, (1920) *J Bact.*, 5, 325
- KAPLAN, I and WILLIAMS, J W (1941) *J Bact*, 42, 263
- KLEBS, E (1896) *Zbl Bakt.*, 20, 488
- KLIENEBERGER, E (1935) *J Path Bact*, 40, 93; (1942) *J Hyg., Camb*, 42, 485
- KNAYSI G (1930) *J Bact*, 19, 113, (1938) *Botan Rev* 4, 83, (1941) *J Bact.*, 41, 141, (1942) *Ibid* 43, 363
- KNAYSI, G and MUDD S (1943) *J Bact* 45, 349
- KOCH, R (1876) *Cohns Beitr Biol Pflanz.*, 2, 277, (1897) *Deutsch. med Wochr.*, 23, 209
- KOBMULLER, L. O (1935) *Zbl Bakt*, 133, 310
- KORSEN G (1918) *Z Hyg InfektKr.*, 85, 359
- KRONFELDER, E (1901) *Zbl Bakt.*, 30, 385
- KRUSE W (1910) *Munch med Wochr.*, 57, 685
- KUHN, P (1929) *Med Klin*, 25, 1301, (1930) *Ibid.*, 26, 739
- LAHANNA, C (1940) *J infect Dis*, 67, 193, 203
- LEGROUX, R (1975) *Ann. Inst Pasteur*, 39, 382
- LEGROUX, R and MARGROU, J (1970) *Ann Inst. Pasteur*, 34, 417
- LEMBKE, A. and RUSKA H (1940) *Klin. Wochr.*, 19, 217
- LEVENSON S (1938) *Ann Inst Pasteur*, 60, 99
- LEWIS, I M. (1932) *J Bact*, 24, 381, (1934) *Ibid.*, 28, 133, (1938) *Ibid.*, 35, 573, (1940) *Ibid*, 40, 271, (1941) *Bact Rev.*, 5, 181
- LINDEGREN C C (1935) *Zbl Bakt, Ilte Abt*, 92, 40, (1936) *Ibid.*, 93, 389
- LOEFFLER, F (1890) *Zbl Bakt.*, 7, 625
- LÖNNIS, P (1921) *Mem. nat. Acad Sci.*, 18, 5
- LÖNNIS, P and SMITH, N P (1916) *J agric Res*, 6, 675
- MAIER, H (1903) *Arch Protistent*, 2, 73
- MARTON L. (1934) *Bull Acad Belg., Cl Sci* 20, 439, (1941) *J Bact* 41, 397
- MARY, H and WOTHE, F (1900) *Zbl Bakt*, 28, 1
- MELLOM, R R (1917) *J Bact*, 2, 81, 269 (1970) *J med Res.*, 42, 61, (1921) *Ibid*, 42, 111, (1923a) *J Bact*, 10, 481, (1923b) *Ibid*, 10, 579, (1976) *Ibid*, 12, 409
- MENCLE, E (1904) *Zbl Bakt, Ilte Abt.*, 12, 559, (1905) *Ibid*, 15, 544, (1909) *Arch Protistent*, 16, 62, (1910) *Ibid.*, 19, 127
- MERLING EISENBERG K. B (1937) *J Quekett Mus Cl.*, 1, 311, 324
- MEYER, A (1897) *Flora*, 84, 185, (1899) *Ibid.*, 88, 428, (1908) *Ibid*, 98, 335, (1912) 'Die Zelle der Bakterien.' Jena.
- MILES A A and PRIE, N W (1939) *Brit J exp Path.*, 20, 278
- MIRSKY, A. E (1943) *Advances in Enzymology* 3, 1
- MORISON J E (1941) *J Path Bact*, 53, 1
- MORTON H E and ANDERSON T F (1942) *Amer J Syph.*, 28, 560.
- MUDD, S and ANDERSON T F (1942) *J exp Med* 76, 103, (1944) *J. Amer med Ass* 126, 561
- MUDD S, HEYMETS, P and ANDERSON, T F (1943a) *J Bact*, 46, 205, (1943b) *J. exp Med.* 78, 327
- MUDD, S and LACKMAN D B (1941) *J Bact.*, 41, 415
- MUDD, S., POLEVITZKY, K. and ANDERSON, T F (1942) *Arch Path.*, 34, 199
- MUDD S., POLEVITZKY K. ANDERSON T F and CHAMBERS L. A (1941) *J Bact.*, 42, 251
- MUTO T (1904) *Zbl Bakt*, 37, 321
- NAKANISHI, K. (1901) *Zbl Bakt.*, 30, 97, 145, 193, 223
- NEUMAN, F (1941) *Zbl Bakt.*, Ilte Abt, 103, 385
- OGUTI K (1936) *Jap J exp Med*, 14, 19

- PIEKARSKI, G (1937) *Arch Mikrobiol*, 8, 428, (1938) *Zbl Bakt*, 142, 69, (1939) *Ibid*, 144, 140, (1940) *Arch Mikrobiol*, 11, 400
- PIEKARSKI, G and RUSKA, H. (1939a) *Arch Mikrobiol*, 10, 302, (1939b) *Klin Wschr*, 18, 383
- PIETSCHMANN, K (1939) *Arch Mikrobiol*, 10, 133
- PIETSCHMANN, K and RIFFEL, A (1932) *Arch Mikrobiol*, 3, 422
- PIJPER, A (1938) *J Path Bact* 47, 1
- POWELL, M (1935) *Brit J exp Path*, 16, 155
- PREISZ, H. (1907) *Zbl Bakt*, 44, 209
- RAVICH BIRGER, H D and SVINKINA, A A (1937) *G Ball Immunol*, 18, 170
- RAYMAN, B and KRUIS, K (1904) (see Guilhaumon, 1907)
- ROBERTS, J L (1935) *J Bact*, 29, 229
- ROBINOW, C F (1942) *Proc Roy Soc, B*, 130, 299 (1944) *J Hyg, Camb*, 43, 413
- ROSKA, V (1937) *Arch roum Path exp Microbiol*, 10, 207
- RUSKA, U (1934) *Z Physiol*, 87, 580
- RUSS MÜNZER, A (1938) *Zbl Bakt*, 142, 175
- RŮŽIČKA, V (1898) *Zbl Bakt*, 23, 305, (1903) *Arch Hyg*, 46, 337, (1908) *Ibid*, 64, 219, (1909) *Zbl Bakt, Ite Abt*, 23, 289
- SANARILLI, G (1919) *Ann Inst Pasteur*, 33, 569
- SANDER, F (1935) *Zbl Bakt*, 123, 385
- SAUERBECK, E (1909a) *Zbl Bakt*, 50, 239, (1909b) *Z Hyg InfektKr* 63, 313
- SCHAEDE, R (1939) *Arch Mikrobiol*, 10, 473
- SCHAUDINNY, F (1902) *Arch Protistenl*, 1, 300
- SCHIEBLICH, M (1932) *Zbl Bakt*, 124, 269
- SHIN, L E (1938) *J Bact*, 36, 419
- SMITH, N R and CLARK, F E (1938) *J Bact* 35, 69
- STEARN, A E and STEARN, E W (1928) *Univ Mo Stud* 3, No 2, 1
- STEARN, E W and STEARN, A E (1930) *J infect. Dis*, 48, 500
- STEDMAN, E and STEDMAN, E (1943) *Nature, Lond*, 152, 267
- STILLE, B (1937) *Arch Mikrobiol*, 8, 125
- STOUGHTON, R H (1929) *Proc roy Soc, B*, 105, 469 (1932) *Ibid*, 111, 46
- SWELLENGREBEL, N H (1906) *Zbl Bakt, Ite Abt*, 16, 617, 673
- TAMURA, S (1913) *Z physiol. Chem*, 87, 85
- THORNTON, H G and GANGULEE, N (1926) *Proc roy Soc, B*, 99, 427
- TRENNMANN (1900) *Zbl Bakt*, 8, 385
- TURNER, A W and CALES, C E (1941) *Aust J exp Biol med Sci* 19, 167
- VAY, I' (1909) *Zbl Bakt*, 52, 305, (1910) *Ibid*, 55, 193
- VEJDOVSKÝ, F (1900) *Zbl Bakt, Ite Abt*, 6, 577, (1904) *Ibid*, 11, 481
- WAMOSCHER, L (1930) *Z Hyg InfektKr*, 111, 422
- WEI, H (1936) *Chin med J*, Suppl I 135
- WILSON, G S and SHIFF, H L (1938) *Chem Industr*, 77, 834
- WYCKOFF, R W G (1934) *J exp Med*, 59, 381
- ZETZLOW (1918) *Z Hyg InfektKr*, 85, 17
- ZIEHL, F (1882) *Dtsch med Wschr*, 8, 451, (1883) *Ibid*, 9, 247

## CHAPTER 3

### THE BIOLOGICAL CHARACTERISTICS OF BACTERIA METABOLISM

#### The Chemical Constitution of Bacterial Cells

It would seem that bacterial cells are formed on the same general chemical pattern as the cells of other living organisms with certain characteristics that ally them closely to the fungi. The determination of the exact chemical composition of any given bacterial species or strain is rendered peculiarly difficult by variations induced by differences in the nutrient media in which they are grown (See Cramer 1891-97, Nicolle and Ahlairs 1909, Dawson 1919, Fulmer *et al.* 1921, Hunter 1923, Buchanan and Fulmer 1928-30, Eckstein and Soule 1931). Apart from the variability induced by environmental factors, any bacterial species or strain may give rise to variants that differ sharply from the parental type in certain of their metabolic activities. When due allowance is made for these disturbing factors the technical problem remains a difficult one. The collection of an adequate mass of bacterial cells for detailed chemical analysis makes large demands on time and apparatus, and the use of the chemically complex media that are necessary to secure abundant growth of certain bacterial species greatly increases the difficulty of interpreting the analytical results, particularly in regard to any constituents that are present in small amount. It is not therefore surprising that our knowledge is as yet fragmentary.

The difference in chemical constitution between different bacterial genera or species are as would be expected wider than between different strains or variants belonging to a single species. In this section we may confine our attention in the main to those chemical constituents that are shared by bacterial cells in general, noting in passing certain divergencies that serve to illustrate the kind of differences in chemical structure that have been observed.

**The Water Content of the Bacterial Cell.**—In common with all living cells bacteria contain a high proportion of water. Estimations of the water content of different bacteria carried out by different observers have varied widely and figures as high as 90 per cent. have sometimes been recorded, but those given by Nicolle and Ahlairs (1909) range, with few exceptions, from 73 per cent. (*Bact. coli*) to 80 per cent. (*Proteus vulgaris*).

**The Ash Content of Bacteria.**—The figures recorded for the ash content of bacteria vary very widely. Buchanan and Fulmer (1928-30) quote figures ranging from 2.0 to 13.94 per cent. of dry weight, omitting one widely discrepant figure. It seems probable that the ash content is particularly liable to be affected by the medium on which the bacterium is grown. Thus Fulmer and his colleagues (1921) record a reduction in the ash content of a yeast from 6.3 per cent. to 3.0 per cent. as the result of growing it in a medium free from magnesium and calcium salts.

There is general agreement that a high proportion of the total ash consists of phosphoric acid, 10-45 per cent or more reckoned as  $P_2O_5$ , among most bacterial species, 40-70 per cent or more among the acid fast forms (see Tamura 1913b, Buchanan and Fulmer 1928-30). Among the mineral constituents that have been identified are Ca, Mg, Fe, Na, K and the Cl and  $SO_4$  ions.

**The Protein and other Nitrogenous Constituents of Bacteria**—The recorded figures for total nitrogen are widely discrepant. Buchanan and Fulmer quote figures varying between 1.8 and 15 per cent of dry bacterial substance, and the percentages recorded by different observers for the same bacterial species show little agreement. The usual range would appear to be from 8 to 15 per cent. Nicolle and Alilaire's figures vary only between 8.28 and 10.79 per cent, but recent determinations carried out by Linton, Mitra and Shrivastava (1934) on the cholera vibrio give values of 12.17-15.57 per cent.

In regard to the nature and amount of the coagulable proteins of bacterial cells our knowledge is curiously scanty. Borvin and Mesrobian (1934), who record a total N figure of 13.70 per cent dry weight for *Bact. coli*, find that only 0.65 per cent of this nitrogen, reckoned on the same basis, is soluble in trichloroacetic acid, the remaining 13.05 per cent being precipitated. Their figures for other organisms vary over a considerable range, but in every case the nitrogen precipitated by trichloroacetic acid forms more than 80 per cent of the total. It should be noted that acids will precipitate most forms of macromolecular nitrogen from solutions containing precipitable proteins. The separation of proteins from such mixtures has seldom been attempted, but Linton, Mitra and Shrivastava (1934) record in the case of *V. cholerae* that the coagulable protein is almost all in the form of globulin, i.e. is precipitated by half saturation with ammonium sulphate. As regards the non coagulable nitrogen, Borvin and Mesrobian calculate that about a quarter is present as polypeptides or amino acids and about another quarter as ammonium compounds. The percentages falling in these categories vary over a considerable range.

We have more information in regard to the units of which bacterial protein is built up, and these seem, in general, to be the same as those that constitute proteins of other living cells. Numerous observers have carried out estimations by the methods of van Slyke, and the results show fair general agreement, though indicating significant differences in the proteins of different species. Thus, to take a few illustrative examples, in acid fast bacilli, Tamura (1913b) has reported that 63.62-66.74 per cent of the total nitrogen is present as mono amino acid nitrogen, 13.71-15.21 per cent as basic amino acid nitrogen, while Johnson and Coghill (1925) record 47.39-52.10 per cent and 11.35-14.43 per cent for the tubercle bacillus. For the diphtheria bacillus Tamura (1914) gives corresponding figures of 54.62 per cent and 16.89 per cent, Hirsch (1931) gives 44.80-47.41 per cent and 16.67-17.67 per cent. For *Bact. coli* Eckstein and Soule (1931) give 42.90-45.71 per cent and 16.45-19.82 per cent, and for *V. cholerae*, Linton, Mitra and Shrivastava (1934) give 54.84-57.11 per cent and 24.08-26.03 per cent. These figures it may be noted, refer to the total nitrogen present in the mono amino and basic amino acids, not to the nitrogen present in the amino form. Among the amino acids that have been identified in bacterial proteins, arginine, histidine, lysine and tyrosine appear to be almost always present, leucine and tryptophan have both been frequently demonstrated. The figures for cystine vary, some observers have failed to demonstrate its presence, others have found it in relatively small amounts. There seems little doubt that the relative proportions of different amino acids in different bacterial proteins vary significantly. Thus Tamura (1913a) records the presence of relatively large amounts of L phenylalanine in the proteins of an acid fast bacillus, while (1914) he failed to identify this acid in the proteins of the diphtheria bacillus, which contained an unusually large amount of tyrosine. The capsule of the anthrax bacillus contains a large amount of a polypeptide which on hydrolysis yields d(-)-glutamic acid in a state of almost chemical purity (Ivanovic and Bruckner

1937a b) The presence of various amino-acids in detectable amounts, and their quantitative relationship, might be largely influenced by the media employed for growth. The cultural conditions, for example, which lead to a diminution in the capsule formation of the anthrax bacillus would markedly diminish its content of  $d(-)$ -glutamic acid, and with certain strains growth in an atmosphere containing 20 per cent  $\text{CO}_2$  markedly increases capsule formation (Ivanovics 1937). But Tamura (1913b) records closely similar figures for the mono-amino-nitrogen and basic amino-nitrogen in cultures of an acid fast bacillus grown in nutrient broth and on a synthetic medium. In any case the findings recorded above give a very incomplete picture. Detailed and comprehensive analyses have, for the most part, still to be made.

There is one further point in connection with the nitrogenous constituents of bacterial cells on which all observers are agreed—their high content in nucleic proteins. (See Nishimura 1893, Galeotti 1898, Aronson 1900, Stoklasa 1908, Tamura 1913a, Schaffer *et al.* 1922, Buchanan and Fulmer 1928–30, Boivin and Mesrobian 1934, Serag, Smolens and Lackman 1940, Stokinger, Ackerman and Carpenter 1944.) The presence of nucleic acids had been suspected by many of the earlier bacteriologists because of the affinity of bacterial cells for nuclear stains, and the work of the observers quoted above, and of others, has confirmed this conclusion by direct chemical analysis. Such substances as guanine, xanthine, hypoxanthine and adenine have frequently been demonstrated, and the high content of phosphorus has been noted above. Boivin and Mesrobian (1934) and Mesrobian (1936) report that the purine nitrogen of undried cells varied from 0.18 to 0.29 per cent among six species examined, being about 10 per cent of the total bacterial nitrogen. Of this purine nitrogen 73–94 per cent was in the form of nucleic acid, the rest being made up of nucleotides, nucleosides and free purine bases.

Both ribonucleic acid (the 'yeast' type nucleic acid) and desoxyribonucleic acid (the 'thymus' type of nucleic acid) have been isolated from bacteria. Coghill (1931) isolated ribonucleic acid from *Mycophila*, Heidelberger and Kendall (1931) from streptococci and Thompson and Dubos (1938) from pneumococci of which it constituted 2–5 per cent of the dry weight. Nucleic acid of the desoxyribose type occurs in *Bact. coli* (Schaffer *et al.* 1922) and *Myc. tuberculosis* (Johnson and Brown 1922). Serag, Smolens and Lackman (1940) found 20 per cent of the dry weight of *Str. pyogenes* to be nucleic acid and identified 10–30 per cent of it as of the desoxyribose type (see also p. 306). The gonococcus also contains about 20 per cent of nucleic acid (Stokinger *et al.* 1944).

**Carbohydrate Constituents of Bacterial Cells.**—Buchanan and Fulmer quote total carbon figures ranging from a little below to a little above 50 per cent dry weight for various bacterial species, with one widely discrepant finding. The significance of this figure in relation to carbohydrate content is dubious. The actual carbohydrate content is probably a great deal lower, and the content of polysaccharides is certainly lower. In their analysis of the gonococcus, Stokinger, Ackerman and Carpenter (1944) found 5–9 per cent of the dry weight was carbohydrate. We have ourselves observed crude polysaccharide yields of between 5 and 20 per cent from over thirty different strains of *Proteus vulgaris*.

There is very little evidence that bacteria share with the majority of vegetable cells the capacity of forming a cellulose envelope, or that cellulose enters in any way into their composition (see Buchanan and Fulmer 1928–30). They do, however, form a variety of polysaccharide gums, and the studies of recent years have shown quite clearly that complex polysaccharides are of common occurrence in the surface layers of bacterial cells, and in their capsules when these are present. These polysaccharides play an important, often a dominant, part in determining antigenic specificity (see Chapter 8).

We have noted in the preceding chapter that certain bacteria may contain granules that have been variously identified, on the basis of staining reactions,

as starch or glycogen but of the significance of these intracellular granules we know little, and their exact nature is still in dispute

**Bacterial Fats, Lipins and Waxes**—Buchanan and Fulmer quote figures ranging from 1.56–10.8 per cent dry weight for the ether extractable substances in various species of bacteria. Nicolle and Ahlaire record percentages of 6.31–15.77 for acetone extractable substances, but the range of species covered is not the same. Lipin values of less than 10 per cent have been recorded for certain members of the *Salmonella* group (Williams, Bloor and Sandholzer 1939), of 5–6 per cent, for *Brucella* (Stahl and Hamann 1941) and of 10–14 per cent for the gonococcus (Stokinger *et al* 1944). There is no doubt that the production of particular lipins or waxes characterizes particular bacterial species. We have, for instance, noted in the previous chapter the production by the tubercle bacillus of a wax on which its acid fastness depends (see also Chapter 16). We know so little of the part played by fats, lipins and waxes in the economy of the bacterial cell that, for the moment, it will suffice to note their presence. For such data as are available in regard to their chemical constitution reference may be made to Buchanan and Fulmer (1928–30).

### The Study of Bacterial Metabolism

In the study of bacterial metabolism, whether we are concerned with substances utilized by bacteria as sources of food or energy, with the products of metabolism, or with the mechanisms by which dissimilation and assimilation of foodstuffs is achieved, the most striking feature is the extreme diversity of bacterial activity. It is possible to give a generalized description of metabolism that will apply with minor modifications to the nutrition of all the higher animals, but in bacteria, the diverse metabolic activities appear to have been evolved by adaptation to the widest variety of environmental conditions, so that one species or another can take advantage of almost any thermodynamically suitable type of foodstuff and any moist environment within a temperature range rather wider than that suitable for most other groups of organisms. Certain groups of bacteria, however, share a sufficient similarity of metabolic activities as to make them suitable for discussion in an introduction to the subject of bacterial metabolism. Among these are the bacteria that primarily interest the medical bacteriologist and we shall endeavour to select these for illustrative examples. The studies of the kind that concern us have, for the most part, been carried out by biochemists and mark the rapid development of microbiological chemistry as a distinctive branch of biological science. Although work in this field was initiated by Pasteur, and developed extensively by him within the limits of the chemical knowledge and technique of his time, it has been largely neglected by his bacteriological successors. These have, for the most part, been content to use the bacterial fermentation reactions as diagnostic tools, without inquiring in any systematic way into the actual chemical changes concerned. The common practice has been to map out the fermentative abilities of different bacterial species, using an arbitrarily selected series of carbohydrate and other substrates which experience has shown to possess differential value, and noting the production of acid by a colour change in a suitable indicator, or the production of gas by observing its collection within a small inverted tube contained in the culture medium. The study of the changes produced in nitrogenous substrates has been even more limited and arbitrary—the production of indole from a tryptophan-containing substrate, the production of  $H_2S$  from sulphur-containing amino acids, and so on. This neglect has been natural enough. The

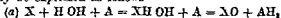


The essential factor in any oxidation then is the removal of electrons, in any reduction, their addition. Regarded in this light, bacterial respiration covers those processes of electron transfer in the mixture of substrate and bacteria which yield free energy. And just as in simple inorganic reactions the oxidation of one substance implies the reduction of another, so in a study of bacterial metabolism we must consider not only the substrate to be oxidized by the bacterium, but the substances, either in the bacterium or in its environment, which must be reduced in order that the biological oxidations may take place.

Besides the fundamental conception of oxidation as electron transfer, we are concerned in metabolic chemistry with the substances to and from which the electrons are transferred. Thus biological oxidations may be expressed simply in terms of the transfer of either hydrogen or oxygen, but in using this interpretation it must be remembered that it is a particularization of the general hypothesis of electron transfer. The addition of oxygen to a molecule, or the removal of hydrogen from it, entails a decrease in electrons, both are oxidations. Similarly the removal of oxygen from, and the addition of hydrogen to, a molecule are equally reductions. In the transfer of either oxygen or hydrogen at least two molecules are concerned. One, to supply the oxygen or hydrogen, is called the donator, the other, to which they go, is called the acceptor.

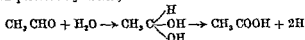
In donating oxygen a molecule is reduced, in accepting oxygen it is oxidized. In donating hydrogen a molecule is oxidized, in accepting hydrogen it is reduced. In both cases the reaction is catalysed and the catalyst is regarded as 'activating' either oxygen or hydrogen.

The theory advanced by Wieland (1913, 1921, 1922) regards hydrogen activation and consequent hydrogen transport, as the essential mechanism of cellular oxidations. The removal of hydrogen from a molecule may be preceded by the addition of a molecule of water, in which case oxygen is in fact added, or there may be no preliminary addition of water, in which case the molecule is oxidized by the simple loss of hydrogen. In both cases a suitable hydrogen acceptor must be provided. In general terms these reactions may be expressed as follows:

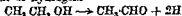


In (a) X represents the substrate to be oxidized and A the hydrogen acceptor. Water is first added to X and the hydrogen of the compound  $XH_2OH$  is then activated and passed on to A, leaving  $\Delta$  oxidized. In (b) the compound  $XH_2$  is the substrate to be oxidized. Oxidation occurs by the activation of the hydrogen and its transference to A.

An example of the first type of reaction is afforded by the oxidation of an aldehyde to an acid with previous hydration,



An example of the second type of reaction is afforded by the oxidation of an alcohol to an aldehyde by removal of hydrogen.



The hydrogen in such reactions is seldom liberated in the gaseous state. In almost all the reactions with which we are here concerned, a hydrogen acceptor must be provided. The enzyme that activates the hydrogen in the substrate to be oxidized, and so brings about its transport, is known as a dehydrogenase.

**The Mechanism of Hydrogen Transport**—The phenomena of hydrogen transport in bacteria may be demonstrated by a technique developed by Quastel and his colleagues, which consists in observing the behaviour of washed bacterial cells suspended in an appropriate buffer solution containing the necessary inorganic ions, when incubated in an evacuated Thunberg tube in the presence of a substrate and an indicator dye such as methylene blue. The reactions are usually completed within 30 minutes or less, so that although a small degree of bacterial multiplication

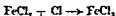
medical bacteriologist, at least, has been mainly interested in other bacterial activities but the time has quite clearly arrived when the chemical aspects of bacteriology and immunity must be mastered by all serious students of these branches of biology.

The metabolism of bacteria as of other living cells, is dependent on, and regulated by, a complex system of enzymes and catalysts, whose activity is conditioned by a variety of factors, such as temperature of incubation, the pH of the medium, the presence or absence of molecular oxygen, and the presence or absence of a particular food stuff. In order that any given bacterium may grow and multiply, these various conditioning factors must fall within a range limited by the requirements of the enzyme systems that are available. Many bacteria possess more than one enzyme system for dealing with a given type of substrate, and a change of environment is accompanied by a change in the predominating enzyme system.

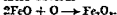
The prime needs of a bacterium are substances that it can assimilate and synthesize into protoplasm, and energy necessary for these syntheses, and for movement (if it is motile) reproduction and the maintenance of structure. Both are obtained by the decomposition of suitable substrates. No sharp dividing line can be drawn between substrates that act mainly as sources of energy and those which are needed for synthesis of bacterial proteins, carbohydrates, fats, enzymes, etc. But it is convenient to distinguish the two types of activity, and discuss the first under bacterial respiration and fermentation, and the second under bacterial nutrition. Bacterial nutrition we shall leave till a later section, but before examining the nature of respiration and fermentation in detail, we shall briefly discuss the general implications of the energy producing mechanisms in living cells.

**The Respiration of Bacteria.**—The term respiration, with its connotations for the student of mammalian physiology of the mechanical intake of oxygen for the purpose of oxidizing food substances, is apt to confuse the student of bacteriology unless he realizes that more than the direct utilization of atmospheric oxygen is concerned. Respiration covers all those metabolic mechanisms that are employed in providing energy, as opposed to those that are concerned in synthesis. It happens that many energy yielding reactions that is, exothermic reactions, are oxidations in the simple sense of the word and take place in the cell when molecular oxygen is supplied. But this concept of oxidation as the union of oxygen with a given substance is as limited in its applications to biology as it is to inorganic chemistry. In neither does the process of oxidation necessarily involve the transfer of oxygen.

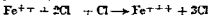
The simple reaction



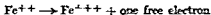
is as much an oxidation as the more obvious



The common feature of these two oxidations, and indeed of all oxidations, is change in the electronic state of the substances concerned. If the oxidation of the ferrous to the ferric chloride takes place in solution, in which the participants in the reactions are ionized, the whole process may be written



The ferrous salt is oxidized, and the chlorine atom reduced. The oxidation of the iron may be written



and the reduction of the chlorine by

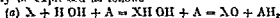


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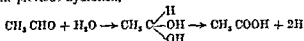
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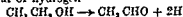


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may take place during the experiment, the enzymic activities displayed are not necessarily those of proliferating cells nor even of living cells (Cook and Stephenson 1928, Sandiford and Wooldridge 1931). In the Thunberg tube, *Bacterium coli*, for example, may be shown to possess an enzyme that is capable of oxidizing succinic acid to fumaric acid in the presence of methylene blue (Quastel and Whetham 1924, Quastel, Stephenson and Whetham 1925). A reversible equilibrium is set up

Succinic acid and methylene blue  $\rightleftharpoons$  fumaric acid and leuco-methylene blue

The enzyme responsible, succinic dehydrogenase, activates the hydrogen. The methylene blue acts as a hydrogen acceptor, and in doing so is reduced to the colourless compound leuco-methylene blue. It will be noted that the methylene blue does not require activation to become a hydrogen acceptor. In the dehydrogenase systems occurring in nature, it is probable that most of the hydrogen acceptors are activated by enzymes. The natural hydrogen acceptors may be either intermediate products of carbohydrate or protein dissimilation, or molecular oxygen itself. In the first case we have a biological oxidation in the absence of air, which we call an anaerobic oxidation, and in the second case an aerobic oxidation in which oxygen is acting as the hydrogen acceptor. Dehydrogenases may catalyse the transfer of hydrogen from the donor directly to oxygen as the acceptor but this mechanism appears to be relatively rare.

The transport of oxygen in the cell is usually brought about by a complex system of oxygen carriers and the oxygen is activated by enzymes which receive the general name of oxidases. Warburg (1920a, b) pictures the molecular oxygen uniting in the cell with some complex organic substance containing iron in the reduced, or ferrous state, and converting it into ferric iron. In the presence of an oxidizable organic molecule and a suitable oxidase oxygen is transferred and the iron returns to the ferrous condition. A natural carrier of oxygen, cytochrome, has been demonstrated in the cells of animals, yeasts and bacteria by Keilin (1925, 1926, 1928-29, 1930), and Keilin and Hartree (1930a, b, c). Cytochrome is a respiratory pigment, made up of a number of related hæmatin compounds, which plays an important part in cell respiration. Keilin's studies supply the link between the Wieland hypothesis of hydrogen transport and the Warburg hypothesis of oxidation by iron-containing compounds.

Keilin's view of cellular oxidations may be briefly summarized as follows. Organisms whose respiration demands molecular oxygen contain a widely distributed respiratory pigment cytochrome composed of three main hæmatin compounds the components *a*, *b* and *c*. The pigment also contains an unbound hæmatin compound similar to the protohæmatin of hæmoglobin, and a hæmochromogen precursor of cytochrome. Of these, the *b* component of cytochrome, the protohæmatin and the hæmochromogen precursor are autooxidizable. The *a* and *c* components are oxidized in the presence of a thermostable enzyme cytochrome oxidase, all factors that destroy or inhibit this oxidase diminish the oxygen uptake of the cell. Some bacterial cells, e.g. *B. subtilis*, have the full complement of cytochrome components, and a fourth component *a*<sub>1</sub>. *Bact. coli*, on the other hand, has no *a*, *a*<sub>1</sub>, *c* or cyto-oxidase, but only *b*, and another component *a*<sub>2</sub> which may possibly act as an oxidase to *b* (Keilin and Harpley 1941). Cytochrome therefore acts as a carrier between two activating mechanisms of the cell, the cytochrome oxidase activating molecular oxygen, and the dehydrogenase activating the hydrogen of various organic substrates, metabolic intermediaries and co-enzymes. The autooxidizable hæmatin compound *b*, the unbound hæmatin, and

the haemochromogen precursor to cytochrome may act as carriers between hydrogen donors and molecular oxygen. They may also act as direct catalysts promoting the oxidation of substrates that are not activated by specific dehydrogenases. The various haematin compounds of the cell are also responsible for the catalase and peroxidase reactions that occur in the presence of  $H_2O_2$ . The  $H_2O_2$ , which is an end product in a number of bacterial oxidations, is reduced in the presence of catalase. Catalase production is probably limited to bacterial species capable of aerobic respiration (Kluyver 1921). Callow (1923, 1924) found oxygen utilization minimal in bacteria producing no catalase (see section on Aerobiosis and Anaerobiosis, p. 70).

Substituting cytochrome for methylene blue in the succinic dehydrogenase system mentioned above, we may represent the oxidation of succinic acid in two stages, the first catalyzed by the dehydrogenase, the second by cytochrome oxidase:

Succinic acid and cytochrome  $\rightarrow$  reduced cytochrome and fumaric acid

Reduced cytochrome +  $O_2 \rightarrow$  cytochrome + water

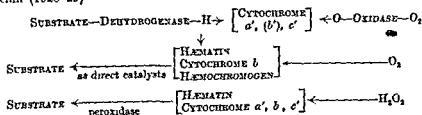
This reaction has not been directly demonstrated in bacterial cells, nor can many of the dehydrogenase systems directly reduce oxidized cytochrome. But it appears that the dehydrogenase systems capable of reacting directly with reduced cytochrome may act as intermediate links between cytochrome and other dehydrogenase systems, by which they are themselves reduced.

Many of the dehydrogenase systems, in addition to hydrogen donor, hydrogen acceptor, dehydrogenase, water, inorganic ions and the correct pH, require the presence of co-enzymes. A co-enzyme may be defined as a thermostable substance necessary in addition to enzyme and substrate to initiate a reaction. Co-enzymes are usually organic in nature, and their molecules are small enough to pass through semi permeable membranes that hold back the larger enzyme molecules. Hence existence of a co-enzyme is usually demonstrated by the inactivation of a natural enzyme preparation after it has been dialysed, and its reactivation by addition of the dialysate. Co-enzyme I, which has been identified as pyridine nucleotide diphosphate, takes part in many enzyme reactions. Co-enzyme I and its dehydrogenase constitute one of the dehydrogenase systems capable of direct reaction with cytochrome. Its relation to the cytochrome system may be represented (Dewan and Green 1933) as follows:

$A H_2 + \text{Co-enzyme I} \xrightarrow{\text{dehydrogenase for A}} A + H_2\text{-co-enzyme I}$

$H_2\text{-co-enzyme I} + \text{oxidized cytochrome} \xrightarrow[\text{co-enzyme I} + H_2\text{-cytochrome}]{\text{dehydrogenase for co-enzyme I}}$

The following diagram of the general oxidative mechanisms as they involve cytochrome itself have been taken, with slight modification, from that given by Keilin (1928-29)



### The Chemical Changes produced by Bacteria in various Substrates

On the view outlined above the oxidation of a complex substance by bacteria may be pictured as the transfer of hydrogen from one substance to another, each step being brought about by one of a graduated series of oxidation reduction systems with appropriate enzymes carriers and co-enzymes the whole resulting in an even flow of energy for cell maintenance and syntheses. We shall now examine in more detail the action of bacteria in the different types of foodstuffs.

**The Action of Bacteria on Carbohydrates and Allied Substances**—The majority of the bacteria with which we are concerned in this book are able to attack the hexose sugars. Occasionally the end result is complete oxidation to water and carbon dioxide. This is, however, an unusual type of reaction, limited to a few species and demanding a copious supply of oxygen. Various reactions leading to less complete oxidation are more common, and one of the most characteristic types of bacterial fermentation is that in which molecular oxygen plays no direct part, the reaction consisting essentially in the splitting of a complex molecule usually by hydrolysis with a rearrangement of the oxygen atoms so that one portion of the hydrolysed molecule is oxidized while the other is reduced. An example of this general type of reaction is afforded by the studies of Harden (1901-1905), and of Harden and Walpole (1906) on the fermentation of dextrose by *Bact. coli*. The main products of fermentation are lactic acid acetic acid ethyl alcohol carbon dioxide and hydrogen, and the reaction appears to be approximately represented by the formula,

$$2C_6H_{12}O_6 + H_2O = 2CH_3CHOHCOOH + CH_3COOH + C_2H_5OH + 2CO_2 + 2H_2$$

although small amounts of other substances such as succinic acid are produced.

In some instances, as in the fermentation of dextrose by the typhoid bacillus no free gas is evolved. It was suggested by Harden that in this case that part of the reaction which, with such an organism as *Bact. coli*, leads to the evolution of equal parts of  $CO_2$  and  $H_2$  stops short at the formation of formic acid  $HCOOH$  (see also Pakes and Jollyman 1901). This suggestion was strengthened by the observation of Sera (1910) while Grey (1913-14) has shown that formic acid can be identified as an intermediate product in the fermentation of dextrose by *Bact. coli*.

This type of reaction is, however, by no means the only one that occurs during the cleavage of carbohydrates and allied substances by bacteria.

The existence of wide variations is well illustrated by the results recorded by Birkenshaw, Charles and Clutterbuck (1931). Using the carbon balance sheet method employed by Raistrick and his colleagues (1931) in their extensive studies of the metabolism of moulds, they determined the relative proportions of different metabolic products formed by twenty different bacterial species growing in a synthetic medium containing glucose. In the case of *Bact. coli* and of certain nearly related organisms up to 30.4 per cent of the carbon of the glucose was recovered after fermentation in the form of lactic acid, 5.0-14.4 per cent was recovered in the form of volatile acids, while the amount present as butylene glycol ( $CH_2CHOHCHOHCH_2$ ) was negligible. On the other hand a coliform organism of a different type *Bact. anatum mobile*, yielded 26.8-31.0 per cent of the carbon as butylene glycol but none as lactic acid. Two anaerobes (*Cl. saccharobutylicum* and *Cl. pasteurianum*) yielded a large proportion of carbon

These are not the only mechanisms affecting the transfer of hydrogen to molecular oxygen. Many animal, yeast and bacterial cells, for example, contain flavoprotein (Schütz and Theorell 1933). Flavoprotein the "yellow enzyme" of Warburg and Christian (1933) is riboflavin phosphate combined with a protein group, it is spontaneously oxidizable by molecular oxygen, but acts also as a hydrogen carrier between dehydrogenase systems and the cytochrome system (Ogston and Green 1935), and anaerobically between two dehydrogenase systems (Dewan and Green 1938). It is reduced by co-enzyme I and may be related to the dehydrogenase of co-enzyme I mentioned above, which Straub, Corran and Green (1939) have shown to be a flavoprotein.

The dehydrogenase systems are reversible (Quastel and Whetham 1924, Green and Stuckland 1934). If the reaction of succinic acid and methylene blue is allowed to proceed in the presence of *Bact coli* dehydrogenase until the dye is reduced, and an excess of fumaric acid added the leuco-methylene blue gradually becomes reoxidized. The reversal is due to the action of the enzyme, for the destruction of the enzyme by heat or the addition of an antiseptic completely inhibits the reoxidization of the methylene blue. Since these reactions are reversible, it is obvious that they will not take place in a given direction unless the hydrogen donors and acceptors are in a suitable physical state for the removal of electrons from the substance to be oxidized.

**Biological Redox Systems**—In a given reversible oxidation reduction system the state of oxidation may be measured by the tendency of the system to give up electrons and the state of reduction by the tendency to take up electrons.

If a platinum electrode is immersed in a fluid containing such a system, an electrical half-cell is produced and a potential difference, depending on the availability of electrons in the system is set up at the electrode. When the half-cell is put into a circuit with the normal hydrogen electrode as a standard half-cell the electromotive force that develops will be a measure of the electrode potential in the system compared with that of the standard half-cell. This force, measured in volts, is designated the  $E_h$  of the system. A marked tendency to reduction results in a flow of electrons from the standard half-cell, i.e. the current flows from the system to the half-cell and the  $E_h$  is positive. A system with a marked tendency to oxidation, on the other hand, since electrons flow to the standard cell half-cell is indicated by a negative  $E_h$ .

The conception of  $E_h$  the oxidation reduction (or redox) potential (see Clark *et al.* 1928, McLeod 1930, Hewitt 1936) as a measure of reducing intensity may be applied to the reversible enzyme systems of the dehydrogenase type. Oxidation, for example, will not proceed in a given system except in the presence of a system with a higher  $E_h$ . Thus it will be seen that the absolute  $E_h$  value is of less importance than its value compared with other systems, for a system of a high reducing intensity, characterized by a low  $E_h$  will be further reduced by a system with an even lower  $E_h$ . For the general characterization of each system the  $E_h$  is determined in standard conditions, namely when equivalent amounts of the reduced and oxidized forms are present in equilibrium at pH 7.5 and a temperature of 30°C. This characteristic  $E_h$  is designated  $E'_0$ , and in general it may be said that, in the proper circumstances, a system of a given  $E'_0$  will in the oxidized state be reduced by a system of lower  $E'_0$  and in the reduced state will be oxidized by a system of higher  $E'_0$ . For example, the  $E'_0$  of the succinic-dehydrogenase system is about 0.00 volt and that of cytochrome-cytochrome oxidase about +0.12 volt. On electronic grounds it is to be expected that the oxidation of the succinic acid to fumaric will occur in the presence of oxidized cytochrome.

## The Chemical Changes produced by Bacteria in various Substrates

On the view outlined above the oxidation of a complex substance by bacteria may be pictured as the transfer of hydrogen from one substance to another, each step being brought about by one of a graduated series of oxidation reduction systems with appropriate enzymes carriers and co-enzymes the whole resulting in an even flow of energy for cell maintenance and syntheses. We shall now examine in more detail the action of bacteria in the different types of foodstuffs.

**The Action of Bacteria on Carbohydrates and Allied Substances**—The majority of the bacteria with which we are concerned in this book are able to attack the hexose sugars. Occasionally the end result is complete oxidation to water and carbon dioxide. This is, however, an unusual type of reaction, limited to a few species and demanding a copious supply of oxygen. Various reactions leading to less complete oxidation are more common, and one of the most characteristic types of bacterial fermentation is that in which molecular oxygen plays no direct part, the reaction consisting essentially in the splitting of a complex molecule usually by hydrolysis, with a rearrangement of the oxygen atoms so that one portion of the hydrolysed molecule is oxidized while the other is reduced. An example of this general type of reaction is afforded by the studies of Harden (1901-1905), and of Harden and Walpole (1906) on the fermentation of dextrose by *Bact coli*. The main products of fermentation are lactic acid, acetic acid, ethyl alcohol, carbon dioxide and hydrogen, and the reaction appears to be approximately represented by the formula

$$2C_6H_{12}O_6 + H_2O = 2CH_3CHOHCOOH + CH_3COOH + C_2H_5OH + 2CO_2 + 2H_2$$

although small amounts of other substances such as succinic acid are produced.

In some instances as in the fermentation of dextrose by the typhoid bacillus no free gas is evolved. It was suggested by Harden that in this case that part of the reaction which, with such an organism as *Bact coli*, leads to the evolution of equal parts of  $CO_2$  and  $H_2$  stops short at the formation of formic acid  $HCOOH$  (see also Pakes and Jollyman 1901). This suggestion was strengthened by the observation of Sera (1910) while Grey (1913-14) has shown that formic acid can be identified as an intermediate product in the fermentation of dextrose by *Bact coli*.

This type of reaction is, however, by no means the only one that occurs during the cleavage of carbohydrates and allied substances by bacteria.

The existence of wide variations is well illustrated by the results recorded by Birkenshaw, Charles and Clutterbuck (1931). Using the carbon balance-sheet method employed by Raistrick and his colleagues (1931) in their extensive studies of the metabolism of moulds they determined the relative proportions of different metabolic products formed by twenty different bacterial species growing in a synthetic medium containing glucose. In the case of *Bact coli* and of certain nearly related organisms up to 30.4 per cent of the carbon of the glucose was recovered after fermentation in the form of lactic acid. 5.0-14.4 per cent was recovered in the form of volatile acids while the amount present as butylene glycol ( $CH_2CHOHCHOHCH_2$ ) was negligible. On the other hand a coliform organism of a different type *Bact asiaticum mobile* yielded 26.8-31.0 per cent of the carbon as butylene glycol but none as lactic acid. Two anaerobes (*Cl saccharobutyricum* and *Cl pasteurianum*) yielded a large proportion of carbon



as volatile acids (31.8 per cent and 37.8 per cent) but formed little if any lactic acid or butylene glycol.

Many hypotheses have been put forward to explain the precise mechanism of the dissimilation of the relatively simple carbohydrates. It is impracticable to deal with them fully in this chapter, and the student is referred to the monograph of Stephenson (1939) and the review of Werkman (1939) for a detailed consideration of the subject. As an illustration of the complexity of the mechanisms so far elucidated and the multiplicity of enzymes and carriers that may take part we may cite the Embden-Meyerhof-Parnas scheme for the anaerobic dissimilation of glucose by animal tissues and yeast cells.

According to this scheme the hexose first reacts with adenosine triphosphate in the presence of a phosphorylase to produce a hexose diphosphate and adenosine monophosphate. The hexose diphosphate then breaks down into two molecules of triosephosphate under the influence of an aldolase. The triosephosphate combines with acetaldehyde in the presence of a mutase and co-enzyme I to form one end product of the breakdown, ethyl alcohol, and 3-phosphoglyceric acid. An enolase changes the 3-phosphoglyceric acid into phosphopyruvate by the removal of a molecule of water, and the phosphopyruvate is dephosphorylated in the presence of the adenosine monophosphate (left from the phosphorylation of the hexose) to form pyruvic acid and adenosine triphosphate. The adenosine monophosphate (adenylic acid) thus acts as a co-enzyme in the transfer of phosphate from phosphopyruvic acid to glucose. The pyruvic acid, according to the enzymic condition of the cells concerned, may react with more triosephosphate to yield lactic acid and phosphoglycerate, or be acted upon by a carboxylase in the presence of co-carboxylase to give carbon dioxide and acetaldehyde. The carbon dioxide is another end product, and the acetaldehyde is available for the reaction with triosephosphate to form alcohol.

The co-enzyme I acts as a hydrogen carrier in the third system, the full reaction being

Acetaldehyde and reduced co-enzyme  $\rightarrow$  ethyl alcohol + oxidized co-enzyme.

Triosephosphate + oxidized co-enzyme  $\rightarrow$  phosphoglyceric acid + reduced co-enzyme

The cycle, which is one of many possible cycles, is most easily demonstrated in animal tissues, and in extracts of yeast cells. For technical reasons active extracts of bacterial cells are difficult to prepare, but evidence is accumulating that the Embden-Meyerhof-Parnas scheme is applicable to anaerobic glycolysis of some bacteria. The evidence is of four kinds.

Firstly, the presence of the enzymes concerned may be inferred by comparing the action of relatively specific enzyme poisons on the dissimilation by the cells under study, with its action on defined enzyme systems.

Secondly, the presence of the various oxidative, hydrolytic and phosphorylating systems may be demonstrated by adding the hypothetical intermediaries to a suspension of the cells, and measuring its power to deal with them, but, as Stephenson (1939) points out, though this demonstrates that a particular metabolic path may be followed in experimental dissimilation, it is not necessarily followed in natural fermentation.

Thirdly, the unknown dialysable carriers may be removed from the enzyme preparation, and the need for them demonstrated by adding known carriers from other sources.

And lastly, the intermediate products postulated may be isolated from the fermentation system, particularly after the dissimilation has been interrupted by various enzyme poisons.

Virtanen (1924, 1925) and Virtanen and Karström (1931) demonstrated that phosphorylated hexoses occur in bacterial metabolism. A key intermediate, phosphoglyceric acid, has been isolated from a large number of bacteria by Werkman and his colleagues

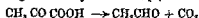
(see Werkman 1939) and from *Bact. coli* by Endo (1938), moreover it is formed during the breakdown of hexose diphosphate

The difficulty of obtaining active cell free extracts of bacteria similar to those of the much more extensively studied yeasts has been overcome by disintegrating bacteria by ultra sonic vibrations (see Chapter 5) or in special crushing mills (Booth and Green 1938). By the latter means for example Still (1940) has demonstrated that cell free preparations of *Bact. coli* will catalyse the oxidation of triosephosphate (3 phosphoglyceraldehyde) to 3 phosphoglyceric acid

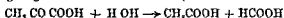
It must be emphasized that this is one example only of the possible ways of *anaerobic glycolysis*. Indeed it is probable that two or more processes of glycolysis only one of them involving phosphorylation may occur at the same time in a bacterial culture (see Tasman and Brandwijk 1938). The applicability of the Embden Meyerhof Parnas scheme to bacteria is by no means fully established but it serves to illustrate not only the complexity of the mechanisms but their dependence, in certain cases at any rate on a number of enzymes and carriers which themselves may be intermediaries between stages on other metabolic paths

Among the intermediate products of the carbohydrate metabolism of bacteria pyruvic acid holds an important place whether the metabolic path actually followed corresponds to the Embden Meyerhof Parnas scheme or to one of the alternative schemes that have been proposed. It is a source of acetic succinic butyric and fumaric acids of ethyl and iso propyl alcohol acetone glycerol acetylmethylcarbinol butylene glycol carbon dioxide and hydrogen. Werkman (1939) lists the following examples of anaerobic dismutation of pyruvic acid produced by bacteria and yeasts

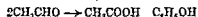
- 1 Decarboxylation, giving acetaldehyde and  $\text{CO}_2$  (*Sarcina ventriculi* Smit 1930)



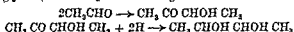
- 2 Hydrolysis giving acetic and formic acids (*Bacteriacea* Tikka 1937)



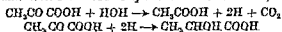
- 3 Dismutation of the acetaldehyde produced by decarboxylation giving acetic acid and alcohol (many acetic acid bacteria)



- 4 Condensation of the acetaldehyde to produce acetylmethylcarbinol and reduction giving butylene glycol (*Bact. aerogenes* Neuberg and Reinforth 1923-24)



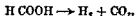
- 5 Dismutation and reduction giving acetic and lactic acids and  $\text{CO}_2$  (lactic acid bacteria Nelson and Werkman 1936 *Staph. aureus* Krebs 1937)



The metabolism of pyruvic acid is closely connected with thiamin (vitamin  $\text{B}_1$ ). The co-enzyme necessary for reaction 1, a cocarboxylase, has been identified as the diphosphate of thiamin and is necessary for the breakdown of pyruvic acid by *Bact. acidifaciens longissimus* (Lipmann 1937) *Staph. aureus* (Hills 1938) propionic acid bacteria (Silverman and Werkman 1939) and *Str. haemolyticus* and gonococci (Barron and Lyman 1939).

As noted above (p. 45) for the purposes of classification of bacteria the power of a species to break down a given carbohydrate is usually measured by the production of acid or of acid and gas. The gases produced in macroscopic quantities from small volumes of fluid medium are usually  $\text{CO}_2$  and  $\text{H}_2$ . We have noted a number of ways in which hexose sugars may be supposed to give rise to various acids and  $\text{CO}_2$ . Pakes and Jollyman (1901) tested certain members of the *Bacterium* group and showed that all those

producing gas from dextrose also did so from formate. The production of molecular hydrogen from formic acid was shown by Stephenson and Stickland (1932) to be due to hydrogenlyase which catalyses the reaction



a reaction that Woods (1930) showed to be reversible. Strains of *Bact. coli* that produce no gas have no formic hydrogenlyase (Ordal and Halvorsen 1939). Formate is not necessarily the intermediary in hydrogen production, for though *Cl. tetanomorphum* produces hydrogen from dextrose and pyruvate it will not do so from formate (Woods and Clifton 1937). As we shall see in the section on protein metabolism the hydrogen may also be produced by the hydrolytic deamination of amino-acids.

So far we have considered examples of anaerobic breakdown of carbohydrates. These substances are incompletely oxidized and the partly oxidized substrates often act as hydrogen acceptors promoting the further oxidation of substances occurring in another chain of metabolic events. The utilization of molecular oxygen results in a more complete oxidation and the liberation for cell synthesis of more free energy than can be obtained by anaerobic glycolysis.

Many bacteria are unable to utilize certain carbohydrate substances unless oxygen either molecular or combined is present. Thus *Bact. coli* supplied with organic acids such as lactic, fumaric, succinic or pyruvic as sole sources of carbon will not grow except in the presence of air (Stephenson and Whetham 1924). With the addition of nitrate anaerobic growth takes place, oxidation of the organic acid being achieved at the expense of the nitrate which is reduced to nitrite. The activation of the nitrate to become, in terms of Wieland's hypothesis, a hydrogen acceptor is due to a specific enzyme which *Bact. coli* happens to possess in common with other bacteria capable of both aerobic and anaerobic dissimilation.

The participation of oxygen as a hydrogen acceptor in glycolysis does not usually take place without the intervention either of carriers or catalysts or both. The cytochrome-cytochrome oxidase systems involving haemin compounds appear to be the most important in this respect though certain bacteria contain respiratory pigments which enable them to utilize oxygen after the cytochrome systems have been poisoned by cyanide. As an example we may cite the pigment procyanin found in *Pseudomonas pyocyanea* which Friedheim and Michaelis (1931) showed to be autooxidizable. Green, Stickland and Tarr (1934) demonstrated its ability to act as a hydrogen carrier between pairs of dehydrogenase systems studied in the test tube. Added to suspensions of pigment free strains of *Pseudomonas pyocyanea* procyanin strongly stimulates the oxygen uptake (Friedheim 1931). The stimulation may in part be due to the addition of an effective carrier between dehydrogenase and cytochrome systems but in certain circumstances (Friedheim 1934) the procyanin carries hydrogen directly to molecular oxygen. The violet pigment of *Chromobacterium violaceum* acts in an analogous manner. *Chr. violaceum* frequently gives rise to non-pigmented variants, which suspended in a buffer solution take up oxygen at a moderate rate. A solution of the pigment of *Chr. violaceum* takes up no oxygen but the addition of the dissolved pigment to the suspension results in a two- to three-fold increase in oxygen uptake (Friedheim 1932).

Many organisms contain enzyme systems for both aerobic and anaerobic glycolysis. The predominance of one system or the other in the cell at a given moment depends on the presence or absence of molecular oxygen in the environment. The mere presence of oxygen however does not necessarily impose aerobic

glycolysis on the organism. Stephenson and Whetham (1924) compared the aerobic and anaerobic breakdown of dextrose by *Bact coli* in a free supply of air. The breakdown in the early stages of growth was almost anaerobic, oxygen uptake was minimal, and lactic acid and similar incompletely oxidized substances were produced. During the later stages of growth, when the release of free energy from these incomplete oxidations was diminishing, atmospheric oxygen was freely utilized and  $\text{CO}_2$  given off.

A large number of carbohydrate substances are known to be attacked by bacteria; they range from 3 carbon compounds through pentoses, hexoses and disaccharides to polysaccharides like cellulose, starch and dextrin. Many of the bacterial species that attack hexose sugars also ferment the related hexahydric alcohols, mannitol, sorbitol and dulcitol, which are apparently broken down in an analogous manner. By analogy with yeast metabolism the breakdown of the more complex disaccharides and polysaccharides is presumably achieved by a preliminary hydrolysis to simple sugars. Saccharose is first hydrolysed to a mixture of dextrose and fructose, lactose to dextrose and galactose and maltose to dextrose; the enzymes concerned are, respectively, invertase, lactase and maltase. When hydrolysis has reached this stage the utilization of the hexose sugars proceeds along the lines indicated above. Direct evidence that this is the normal mode of cleavage of disaccharides in bacteria is scanty (see Fleming and Neill 1927). Moreover, some yeasts ferment certain complex sugars at a greater rate than they ferment dextrose, a fact difficult to reconcile with the hypothesis of a preliminary breakdown to the simple sugar (see Sobotka and Holzman 1934, Nord and Engel 1938, O'Connor 1940, Leibowitz and Hestrin 1942). Wright (1936, 1937) finding that certain milk streptococci ferment lactose and saccharose more rapidly than they ferment the constituent monosaccharides suggested that in bacteria also the disaccharides may be attacked directly without previous hydrolysis to monosaccharides. The production of acids from saccharose by a number of bacterial species, including *Bact coli*, was inhibited by low concentrations of iodoacetic acid, and yet no reducing sugar could be found in the test cultures. The concentration of iodoacetic acid was nevertheless insufficient to inhibit the hydrolysis of saccharose by yeast invertase.

Little is known of the processes of carbohydrate synthesis in bacteria. The best-established synthesis is that of 4-carbon carboxylic acids from 3 carbon compounds and  $\text{CO}_2$  (see Werkman and Wood 1942, van Niel *et al.* 1942, Evans *et al.* 1943). Among the more complex compounds, we may note the formation of the polyglucoside dextran, and the polyfructoside levan, from disaccharides, by the action of cell free extracts. These polysaccharides are contained in the capsular material of several bacteria. Hehre and Sugg (1942) produced a serologically specific (p. 279) dextran by the action on sucrose of a cell free enzyme preparation from *Leuconostoc mesenteroides*. Hestrin and Avineri Shapiro (1943, 1944) studied the formation of levan from sucrose and raffinose with enzymes extracted from a strain of *Bact aerogenes*. They suggested that the energy requisite for the synthesis of the polysaccharide might be produced by the breakdown of part of the substrate into an aldose.

**The Action of Bacteria on Proteins and other Nitrogenous Substances.**—The breakdown of proteins that occurs under natural conditions has long been known to depend on the action of bacteria. This proteolytic activity is, however, confined to particular species, many of which are anaerobic; it is at least doubtful whether even the proteolytic species are able to utilize complex proteins for growth in the absence of other sources of nitrogen.

Bainbridge (1911) found that many bacteria, including *Proteus vulgaris*, *Bact*

coli and staphylococci, were unable to grow in solutions of pure egg albumin or of serum proteins, but that *Proteus vulgaris* was able to break down such complex proteins provided that a sufficient supply of nitrogen in an assimilable form was added to the medium. Similar results have been recorded by Rettger and his colleagues (Sperry and Rettger 1915, Rettger, Berman and Sturges 1916, Berman and Rettger 1918). Of several gelatin liquefying bacteria studied by Berman and Rettger, none could utilize egg albumin in the absence of other sources of nitrogen. Only three species, *B. subtilis*, *Chr. prodigiosum* and *Proteus vulgaris*, could break down peptone. With proteases purified by precipitation, *B. subtilis* and *Chr. prodigiosum* caused complete breakdown. Only *B. subtilis*, *Chr. prodigiosum* and *Proteus vulgaris* could attack casein. *B. subtilis* and *Chr. prodigiosum* could not only liquefy gelatin, but could use it as a source of nitrogen, but such organisms as *Staph. aureus* or *Bact. cloacae*, although they caused rapid liquefaction of the gelatin when provided with another source of nitrogen for growth, showed no ability to utilize the liquefied substance.

The initial stage of liquefaction of protein gels like coagulated serum or gelatin is brought about by proteinases that by opening the peptide linkage, reduce the protein to polypeptides and dipeptides. The bacterial proteinases are readily separated from culture by filtration, and apparently act extracellularly. The breakdown into constituent amino-acids is effected by polypeptidases and dipeptidases. The peptidases have been studied in yeasts, but, being apparently for the most part intracellular they have as yet received no extensive study in bacteria, for, as noted on p. 53 adequate methods of extraction of intracellular enzymes have not long been available.

Many of the anaerobes are conspicuous for their ability to break down complex proteins, and the method of cleavage has been studied by several workers (Wolf and Harris 1918, Harris 1919, Wolf 1919a, b, see also Weil and Kocholaty 1937, Kocholaty, Weil and Smith 1938, van Hevingen 1940). From a study of the proteinases of *Chr. prodigiosum*, *Ps. pyocyanea* and *Ps. fluorescens liquefaciens*, Maschmann (1937) concluded that enzymes of the different bacteria were identical, but were not of the same type as the animal proteinase trypsin or the plant proteinase papain.

The hydrolytic degradation of protein is apparently due to the action of specialized enzymes which are produced in adequate amount when the bacteria concerned are supplied with immediately assimilable food material including nitrogen, during their consequent growth they are able to produce sufficient enzyme to initiate protein cleavage and thereby increase the available nitrogen.

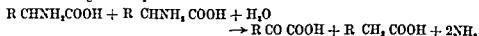
Bacteria utilize amino-acids in a variety of ways, similar in extent and complexity to those that have been described for the utilization of carbohydrates. As in our discussion of those substances, we can do no more than note some of the biochemical processes concerned, for greater detail the student is referred to the monograph of Stephenson (1939) and the review of Gale (1940). The amino-acids may be deaminated to yield the corresponding hydroxy acid by desaturation at the  $\alpha$ - $\beta$  linkage to give the unsaturated acid, by reduction to give the saturated acid by reduction and decarboxylation to give the hydrocarbon, or by oxidation. Decarboxylation results in the corresponding amine. Neither decarboxylation nor deamination are processes yielding much energy, and in bacteria with alternative energy yielding mechanisms, like the streptococci (see Ehrlichmann and Drumburg 1937) the reactions are not important in the bacterial economy. Where the amino-acid is one of the chief sources of food oxidation to compounds with fewer carbon atoms provides the requisite energy for synthesis, etc. The formation of

indole from tryptophan which is widely used as a qualitative biochemical test in the identification of bacterial species, provides a good example of the breakdown of an amino acid. Thick washed suspensions of *Bact. coli* in phosphate buffer will convert this amino acid ( $\beta$  indole  $\alpha$  amino propionic acid) to indole in the presence of oxygen. The oxygen taken up corresponds to the complete oxidation of the side chain to carbon dioxide and water (Woods 1930). In the absence of air the compound is only deaminated with the formation of  $\alpha$  indole propionic acid. The mechanism of the oxidative attack is obscure and many complex series of steps have been proposed for this aerobic dissimilation but the recent work of Fildes (1939) and Baker and Hapgood (1940) suggest that one enzyme, a tryptophanase, is responsible for the action breaking the link between the indole ring and the  $\alpha$  carbon atom of the side chain. The oxidation of the side chain to ammonia,  $\text{CO}_2$  and water follows after the liberation of the indole. A similar mechanism but one involving a disruption and resynthesis of the indole nucleus is proposed by Krebs, Hafez and Eggleston (1942).

The observation that a free supply of a fermentable carbohydrate will spare amino-acids, peptides or proteins in a medium was established by several workers (see Hirschler 1886, Smith 1897, Peckham 1897, Glenn 1911, Kendall, Day and Walker 1914, Kendall and Walker 1915, Jones 1916). The hypothesis of protein sparing formulated on the analogy with mammalian physiology was deduced from the fact that ammonia production in the medium was largely inhibited by the carbohydrate. But as Stephenson (1939) points out the analogy is imperfect for, in the mammalian case the sparing is judged by a diminished excretion of urea, an end product of metabolism, whereas ammonia may be a source of energy for a bacterium. Indeed Raistrick (1919) and Raistrick and Clark (1921) have shown that in a medium containing known amino-acids their decomposition is increased, not lessened, by the addition of the fermentable substance glycerol. The ammonia produced in these circumstances is undetectable since it is utilized by the bacteria for synthesis. It may be noted also that Berman and Rettger (1918) were unable to demonstrate any protein sparing as the result of the addition of carbohydrates except in those instances in which the rapid fall in pH resulting from carbohydrate breakdown caused an inhibition of further bacterial growth and that Heap and Cadness (1924) have shown that the presence of glucose greatly increases the rate of  $\text{H}_2\text{S}$  production from peptone by an organism that forms this gas during protein-cleavage. In regard to the cleavage of more complex protein molecules de Bord (1923) has shown that the addition of a fermentable carbohydrate to a protein containing medium causes an increase in the concentration of amino-nitrogen induced by bacterial growth. These findings are in accord with later observations by Kendall (1922) which indicate that the effect of added carbohydrate is to lessen the utilization of proteins as a source of energy not as material for synthesis.

A number of anaerobic bacteria belonging to the *Clostridium* group are incapable of gross utilization of carbohydrates (see Chapter 36) and are dependent upon amino acids as energy sources. Although bacteria have been described which find adequate energy sources in the employment of chemical mechanisms that from the formal thermodynamic point of view are comparatively unrewarding, the vigorous growth of the clostridia in media containing protein or protein digests has stimulated the search for other anaerobic energy yielding mechanisms. Stickland (1934, 1935) demonstrated a number of amino-acid dehydrogenase systems in *Cl. sporogenes*. Certain amino-acids (alanine, valine and leucine) were hydrogen donors and others (glycine, proline and hydroxyproline) hydrogen acceptors.

He noted that it would be difficult to picture any reaction by which a single amino-acid could break down anaerobically to yield energy, and that the most probable reaction would involve two amino-acids, hydrogen being transported from one to the other by the action of the bacterial enzymes. The general type of such a reaction might be represented as



Stickland demonstrated paired oxido-reductive deaminations of this kind between alanine and proline, cysteine and arginine. Woods (1936) suggests that, since these coupled reactions occur at a rate similar to those of aerobic oxidations, they may constitute an important part of the respiratory mechanism of the cell. The same type of activity has been observed in *Cl. botulinum* but not in *Cl. tetani*, which attacks amino-acids singly (Clifton 1940, 1942). Another energy yielding decomposition was described by Woods and Clifton (1937, 1938) an anaerobic oxidative deamination of amino-acids by *Cl. tetanomorphum*, in which no hydrogen acceptor is required since molecular hydrogen is produced as one of the end products. Hoogerheide and Kocholaty (1938) confirmed the evidence for coupled Stickland reactions in *Cl. sporogenes*, and found in addition that gaseous hydrogen can be utilized by the organism certain amino-acids acting as hydrogen acceptors (see also Woods and Trim 1942, Guggenheim 1944).

As with carbohydrates, our knowledge of protein metabolism is confined mainly to relatively simple systems involving decompositions, and to defining the species of bacterium, and the conditions in which these decompositions take place. Of protein synthesis we know very little except that it may be catalysed by enzymes that our methods of study have hitherto revealed as concerned only in decomposition. The enzyme aspartase in *Bact. coli*, for example, which deaminates aspartic acid to give fumaric acid and ammonia, will in suitable conditions catalyse the formation of aspartic acid from ammonia and fumaric acid (Cook and Woolf 1928).

**The Action of Bacteria on Fats**—It has long been known that many bacteria are able to decompose fats (see von Sommaruga 1894, Rubner 1900, Eijkman 1901, Carriere 1901, Schreiber 1902, Orla-Jensen 1902, Huss 1908, Sohngen 1911, Wells and Corper 1912, Kendall *et al.* 1914, Avery and Cullen 1920, Stevens and West 1922, Michels and Nakahara 1923, Neill and Fleming 1927, van der Walle 1927, Collins and Hammer 1934). The bacterial lipase induces a simple hydrolysis into glycerol and fatty acid (Trussell and Weed 1937), and, under suitable conditions, the glycerol is further decomposed and the fatty acid oxidized (see Harden 1930).

Lipolytic activity is displayed by many parasitic and pathogenic species, such as *Bact. coli*, *Staph. aureus*, streptococci, the pneumococcus and the tubercle bacillus, as well as by saprophytic organisms. But, here as elsewhere, there seem to be wide differences in activity between different bacteria. Many of the parasitic forms are feebly lipolytic, while certain saprophytic species, such as the bacterium isolated by Huss (1908) from milk, are extremely active.

The actively lipolytic bacteria are of considerable industrial importance, since they cause rancidity in butter and other fat-containing foods. It is possible, also, that such species play some part in sewage-purification.

It may be noted, in connection with the fat metabolism of bacteria, that one

species, at least, can form fat from carbohydrate (Stephenson and Whetham 1922, 1923)

### The Nature of Enzymes and Site of Enzyme Action

The elucidation of the nature of bacterial enzymes, their mode of action, and even the identity of certain enzymes, which in our present state of knowledge are often little more than names attached to something catalysing a recognizable reaction, must await the isolation of enzymes in a pure state. For the most part enzymes appear to be proteins or closely associated with proteins. They differ in their susceptibility to changes of hydrogen ion concentration, salt concentration, temperature, and to exposure to various chemicals. For example Quastel and Wooldridge (1927a, b) found that in general the dehydrogenases of *Bact coli* behaved in the same way when the bacterial suspensions were subjected to increasing temperature, pH, exposure to nitrite, benzene, toluene, phenol ether, chloroform and propyl alcohol. The first affected were those acting on alanine, glycerol glycol the sugars and glutamic acid, next, those acting on lactic, succinic and fumaric acid, and finally the formic and acetic dehydrogenases. With strong solutions of KCN and to a certain extent with  $H_2O_2$ , the picture was reversed the formic and acetic dehydrogenating systems being the least resistant. Extending these studies to non toxic inhibitors, the authors (1928) found that the lactic dehydrogenase of *Bact coli* was specifically inhibited by compounds having in common the groups  $-CO COOH$  or  $-CHOH COOH$ , and the succinic dehydrogenase by compounds with the groups  $\equiv C CHOH COOH$  or  $\equiv C CH_2 COOH$  in common. On these groups presumably depends the specific adsorption of the compounds at that part of the enzymic surface responsible for the activation of the lactic or the succinic acid, with a consequent inhibition of dehydrogenase activity.

The multiplicity of enzymic activities exhibited by a simple species—*Bact coli* for instance, is capable of activating over fifty types of dehydrogenations, and both micrococci and streptococci exhibit a wide variety of dehydrogenase activity—inspired the doubt as to whether each enzyme would prove to be a separate chemical entity, on the grounds that the single small bacterial cells would not be large enough to contain them (Grey 1924). The problem of accommodating a large variety of enzymes in a bacterium is not, however, as difficult as first appears. To some extent the production of an enzyme in quantity is conditioned by the presence of a substrate in the bacterial environment (see Chapter 9), so that in the absence of the substrate it is necessary to postulate the presence only of a few molecules of a given enzyme in order that the cell may exhibit the enzyme activity when the substrate is added. The volume of an enzyme molecule if we take pepsin, with a molecular weight of about 37,000 as a model, will be of the order of 20 to 30  $m\mu^3$ . The volume of an average cell of *Bact coli* is about  $700 \times 10^6 m\mu^3$ . Even if we assume a major part of this volume is occupied by non-enzymic material, there is ample room for several thousand enzyme molecules.

In some cases, however, the multiplicity of enzymes may be more apparent than real. Bernheim, Bernheim and Webster (1935) observed that suspensions of *Proteus vulgaris* were able to oxidize practically all the known "natural" (l form) amino-acids. Stumpf and Green (1944) found that this activity was due to a number of enzymes, all but one of which were relatively unstable, and disappeared with ageing of the suspension. The remaining stable enzyme, which they also found in *Bact aerogenes* and *Ps pyocyanea*, had nevertheless a wide



range of action and catalysed the oxidative deamination of no less than eleven amino-acids.

Quastel (1926) and Quastel and Wooldridge (1927a b) suggest that the activation of substrate molecules after their specific adsorption to the enzyme surface is due to their polarization by electrical fields which characterize the "active centres" of cellular and intracellular structures. The active centres are conceived as being developed as the result of molecular strain or distortion of certain groups or molecules brought about by the intermolecular or intramolecular forces that determine the formation of large colloidal aggregates.

As an alternative to the process of absorption and activation Woolf (1931) suggests that activation results from the distortion of the substrate molecule which occurs when enzyme donors and acceptors combine at the enzymic surface. Direct evidence of such a combination was produced by Stern (1935) who by optical means was able to demonstrate the formation of an intermediate compound not merely an absorption complex between substrate and enzyme in the decomposition of monoethyl hydrogen peroxide by animal liver catalase.

Besides the configuration of the active groups on the surface of the enzymic particle their activity may also be determined by the spatial relationship of one group with another. The oxygen uptake resulting from the action upon lactic acid of the lactic dehydrogenase and the cytochrome-cytochrome oxidase system of *Bact. coli* is not increased if cytochrome c and cytochrome oxidase from heart muscle are added to the mixture (Keilin and Harpley 1941) suggesting that the components of the bacterial dehydrogenase system are intimately bound to the protein of a single colloid particle together with the native cytochrome system to form a single oxidizing system whose efficiency depends on the mutual accessibility of the components. Succinic dehydrogenase and cytochrome oxidase (Potter 1941) and co-enzyme I and cytochrome c reductase (Lockhart and Potter 1941) have been found in similar association on particles produced by cell-disintegration. In other words, the efficiency and it might be added the specificity of an enzyme system depends not only on the integrity of the components but on their spatial distribution in the cell containing them. The various cytochrome compounds, indeed afford an excellent example of the part the so-called protein-carrier plays in determining the nature of an enzyme. Recent work has shown that a common feature of these compounds is an iron atom, held by some of its co-ordination valencies to the pyrrole nuclei forming a tetrapyrrole compound while others attach this tetrapyrrole compound to the protein. The activity of the resulting compound i.e. whether it behaves as a cytochrome cytochrome oxidase etc., is determined by the nature of the protein. It follows that the distribution of these respiratory pigments for example among the bacteria will be conditioned more by the nature of the proteins available for conjugation than by the distribution of the pyrrolic iron compounds (Keilin 1943).

Certain observations by Penrose and Quastel (1930) are of interest from this point of view. There is an organism (*Micrococcus lysodeikticus*) that is peculiarly susceptible to lysis by an active substance (lysozyme) which is contained in various tissues and secretions (see p. 1020). A suspension of this organism activates lactic acid, glucose, levulose and glutamic acid, among other substances as hydrogen donors. After dissolution by lysozyme the bacteria are found to have lost completely their power to activate the hexose sugars and glutamic acid while retaining some 30 per cent. of their activity against lactic acid.

### The Nutritional Requirements of Bacteria

Some bacterial species can live and multiply when provided only with very simple food materials; others require far more complex substances. Orla Jensen (1909) distinguished three main bacterial groups on the basis of their food requirements. The groups are

- (1) Bacteria, obtaining both their carbon and nitrogen from inorganic sources, such as  $\text{CO}_2$ ,  $\text{CH}_4$ ,  $\text{NH}_3$  or atmospheric nitrogen itself
- (2) Bacteria obtaining nitrogen from inorganic sources but requiring organic substances as a source of carbon
- (3) Bacteria demanding organic substances as sources for both carbon and nitrogen

The first group consists of autotrophic, the second and third groups of heterotrophic bacteria. The limitations of this grouping have been discussed at some length by Knight (1936), who points out that the difference between autotrophs and heterotrophs is quantitative, the autotrophs using the far more costly means of obtaining their energy, but that the quantitative difference is so large that it amounts to a qualitative difference. Thus we express in terms of the characteristic methods whereby energy is obtained for carbon assimilation—namely in the case of autotrophs by the use of radiant energy or the oxidation of inorganic compounds, in the case of heterotrophs by use of carbon compounds already partly synthesized on the paths leading to protoplasm. Knight's monograph, which has been drawn on freely in this and the following sections should be consulted for details of the argument. Like those who have previously attempted a classification of bacteria upon nutritional grounds he arranges bacterial species in classes corresponding to increasing complexity of nutritional requirements, but maintains that these are to be regarded only as stages, which merge into one another, there are, for example, heterotrophic bacteria that with training can adapt themselves to an autotrophic existence. We shall proceed to discuss the different nutritional types under the headings of Knight's four stages.

*Stage 1—Carbon is assimilated as  $\text{CO}_2$  and  $\text{N}_2$  from inorganic sources, especially ammonia. The energy required for this assimilation (the reduction of the  $\text{CO}_2$  and the synthesis of protoplasm) is derived from the oxidation of simple inorganic compounds, or from the use of radiant and chemical energy.*

The classical studies of autotrophs in Stage 1 have been made upon soil organisms, especially those concerned in the nitrogen cycle. The nitrosifying bacteria of the soil obtain energy by the oxidation of ammonia to nitrites, the nitrifying bacteria by the oxidation of nitrites to nitrates. The relation between the ammonia or nitrite oxidized, and the  $\text{CO}_2$  assimilated, is a quantitative one (Winogradsky 1890a, b, c). Among the substances oxidized by autotrophs are molecular hydrogen (Kaserer 1905, 1906, Niklewski 1908) and sulphur and sulphur compounds (Winogradsky 1889). A special class of bacteria obtains energy by a photosynthetic mechanism depending on the action of light on a cell pigment and in this respect forms a link with the blue green algae and the chlorophyll containing higher plants. These photosynthesizing species include the purple and green sulphur bacteria (see van Niel 1931, 1933, 1944, Muller 1933, Roslofson 1934, 1935, and Gaffron 1934, 1935, 1944). The purple sulphur bacteria contain a red and a green pigment, the green alone being concerned in photosynthesis,  $\text{H}_2\text{S}$  is oxidized to sulphur,

and the sulphur to sulphate The green sulphur bacteria also utilize  $H_2S$  oxidizing it to sulphur

*Stage 2*—Energy and carbon compounds for assimilation are derived by utilization of carbon compounds more reduced than  $CO_2$  which is not assimilated by the assimilation of nitrogen from simple sources ( $N_2$ ,  $NH_3$ ,  $NO_3$ ) the organisms can synthesize their protoplasm

In this group are found a few species utilizing carbon monoxide methane and other hydrocarbons Of particular interest is *B. oligocarbophilus* which though capable of obtaining its energy by the oxidation of CO can also utilize simple organic compounds like formic acetic and butyric acids (Lantzesch 1922) constituting a transitional type between Stage 2 and Stage 3

*Stage 3*—Energy and carbon compounds for assimilation are derived by utilization of carbon compounds more reduced than  $CO_2$  amino-acids are required for nitrogen assimilation some as specific components for the synthesis of protoplasm ammonia cannot be used as a nitrogen source

*Stage 4*—Energy and carbon compounds for assimilation are derived by utilization of carbon compounds more reduced than  $CO_2$  An array of amino-acids are needed for nitrogen assimilation as specific components for synthesis of protoplasm. Accessory growth-promoting substances are also required some organisms requiring more than one

Knight regards the nutritional series obtained by arranging bacteria in order of increasing complexity of nutritional requirements as a possible model for the evolutionary process that has produced the bacterial species at present in existence

The interpretation of species differences along evolutionary lines is of necessity speculative and particularly so among the bacteria

The autotroph capable of obtaining its energy from simple carbon and nitrogen compounds provides us with a possible model for the forms of life that first appeared in an environment presumably consisting of little but what we now regard as inorganic compounds But it does not by any means follow that the original bacteria had in fact a complex metabolism of this kind capable of building complex proteins carbohydrates fats and vitamins from the simplest materials. On this basis we might suppose that heterotrophs evolved from autotrophs by adaptation to environments richer in organic matter whereby they were able to dispense with enzymes concerned in the synthesis of less complex compounds With increasing parasitism the organisms would increasingly lose synthetic power until a stage was reached when the parasite was, perhaps dependent on another living organism not only for the supply of already synthesized complex food materials but for the maintenance of a strictly regulated environment to utilize the food Thus we should progress from the bacteria in Stage 4 to stricter parasites like *Myc. lepra* and finally to the rickettsiae and the viruses which need the specific environment of the cytoplasm or nucleus of certain cells for their development and have few or no demonstrable metabolic processes

The evolutionary process may in fact have gone the other way the highly complex autotroph being the final stage of a process which started in a virus like organism proceeding by the gradual acquisition of new assimilative and synthetic powers.

The study of natural and induced bacterial variation (Chapter 9) gives us no valid indication of trends in nutritional evolution Variation proceeds in both directions fastidious strains of limited synthetic powers may be trained to utilize

but an undetectable though essential trace of metal may also be present (cf the necessity for molybdenum in the fixation of nitrogen by *Azotobacter*, Burk 1934)

A particular ingredient may play no essential part in the metabolism of the bacterium but promote growth by neutralizing a toxic substance in the medium, or by changing the physical state of the medium (see, for example, O Meara 1937) to one more suitable to essential metabolic processes. Thus the addition of charcoal to a medium is said to improve its growth promoting properties for gonococci and meningococci (Glass and Kennett 1939) and for tubercle bacilli (Nassau 1942). According to Glass and Kennett, the charcoal acts as an adsorbent of inhibitory substances in the medium, perhaps toxic metabolic products, or by catalysing certain oxidations. Substances even more inert than charcoal are beneficial to growth. For example, *Bact coli* will grow better in distilled water containing CO<sub>2</sub> and ammonia if talc is added, the talc acting presumably by reason of a surface action that renders the gases more available as nutrients (Bigger and Nelson 1943), and low concentrations of agar, by reducing the rate of diffusion of oxygen from the air, stabilize redox potentials obtaining in fluid media at levels suitable for the growth of anaerobic bacteria (Reed and Orr 1943, see also Gould 1944). Zobell (1943) in a study of the growth of bacteria found in sea water, concluded that solid surfaces promoted growth by concentrating nutrients through adsorption by providing a resting place for bacteria, and by retarding the diffusion of exo-enzymes and hydrolyses away from the cell, thereby enhancing the assimilation of substances hydrolysed outside the cell. Certain proof of the participation of a given 'essential nutrient' in bacterial metabolism is difficult to obtain though the marking of food substances with radioactive elements (see CO<sub>2</sub> requirements below) may increase the certainty. Assuming that a basal medium has been established, of which separate ingredients are free of impurities, we are still faced with various difficulties. Gladstone (1939), for example, grew a strain of the anthrax bacillus in a defined medium containing a number of amino-acids. The medium ceased to support growth if either valine or leucine was removed, and supported growth feebly if isoleucine, glycine or cystine was absent. Glycine and cystine were subsequently proved to be synthesized by the organism, but the three remaining acids were apparently essential for growth. However, the medium could be made to support growth by the removal of all three amino-acids, singly, each had a toxic effect, that of valine was counteracted by leucine, *vice versa*, and that of isoleucine by valine and leucine together. None of them was an essential nutrient. (In this connection see also McLeod and Wyon 1921, Wyon and McLeod 1923, Ruggieri 1933). Again, the nutrients may be tested in too complex a form, and a precursor of the substance prove equally effective. Thus *Staph aureus* requires thiamin (vitamin B<sub>1</sub>, Knight 1937), but it can synthesize thiamin if the two constituents, thiazole and pyrimidine, are substituted in the medium (Knight and McIlwain 1938).

Considerable difficulty arises when we attempt to define the meaning of the term "essential nutrients". We may define them as essential for growth of a bacterium no matter how that growth may be achieved. In that case we must to some extent ignore the changes that the experimental procedure may induce in the bacterium, for its properties in the final medium may be markedly different from those in the starting medium. We may illustrate this by the nutrition of the typhoid bacillus. Braun and Cahn Bronner (1921, 1922) grew *Salm typhi*, *Salm paratyphi B*, and *Salm enteritidis* in media containing mineral salts, various

three and four carbon organic acids, and ammonium salts as a source of nitrogen. One strain of *Salm typhi* failed to grow on any of the media, nor would it grow when glucose was substituted for the organic acids as a carbon source, but it grew when provided with tryptophan. The variation in the tryptophan requirements of different strains of *Salm typhi* was studied by Fildes and his colleagues (Fildes, Gladstone and Knight 1933, Fildes and Knight 1933). The organism grew in a basal medium containing sodium citrate, magnesium sulphate phosphate buffer and glucose, together with fourteen amino acids: alanine, glycine, valine, glutamic acid, asparagine, tyrosine, phenylalanine, proline, histidine, arginine, leucine, lysine, cystine and tryptophan. Any one of the first ten could be omitted without affecting growth, the omission of leucine or cystine caused delay in growth, and in the absence of tryptophan there was no growth. The concentration of tryptophan necessary for growth in the presence of other sources of nitrogen was as little as 0.00061 per cent. Next, two strains of *Salm typhi* were trained to grow in the absence of tryptophan, with ammonium chloride as the sole source of nitrogen, they had evidently adapted themselves to synthesize the amino acid for it was demonstrable in the bacterial cell by the glyoxylic reaction. It would seem therefore that tryptophan is an essential constituent of the typhoid bacillus: that most strains when first removed from media that contain it are unable to synthesize it, but that they can be trained to do so by gradually accustoming them to a medium from which it is absent. Several other species were examined by the same method (see also Fildes 1935, Fildes and Richardson 1935), and from these observations Fildes and his colleagues concluded that tryptophan was essential in the metabolic processes of the bacteria studied (an "essential metabolite"). They divided bacteria into three groups: (1) Those that can synthesize tryptophan, such as autotrophic bacteria, and among the potentially pathogenic organisms examined *Bact. coli*, *Ps. pyocyanea* and *Salm typhi-murum*. (2) Those that are unable to synthesize tryptophan for themselves, but can be trained to do so, e.g., certain strains of *Salm typhi*, *C. diphtheriae*, *Staph. aureus* and probably *Myco. tuberculosis*. (3) Those that cannot synthesize tryptophan, yet for which it is an essential nutrient, to this group belong *Cl. tetani*, *Cl. sporogenes*, *Cl. botulinum* and *B. anthracis*. As a result of analyses of cultures of *Salm typhi*, Burrows (1939a, b) concludes that in no circumstances is tryptophan an essential nutrient for the organism, but that for those strains whose synthetic powers are small, tryptophan acts as a stimulant of growth.

The distinction between a stimulant and an essential nutrient is important for if we are to define bacteria studied in this manner by their synthetic abilities either manifest or latent, we may regard organisms as devoid of a particular synthetic ability, which in fact may have it but in a degree insufficient to allow growth to take place, and later researches may demonstrate this ability either by successful training of the organism, or by developing more sensitive methods of demonstrating small amounts of the metabolite in question.

Again, the medium containing minimum requirements for growth may not permit the development of the full characteristics of the parent strain. Gladstone (1937) succeeded in training certain strains of *Staph. aureus* to utilize ammonia as a sole source of nitrogen. Neither coagulase nor  $\alpha$  haemolysin (see Chapter 25) was produced in this medium, for the production of optimum quantities of haemolysin, arginine, glycine, proline, phenylalanine and valine were necessary. The trained *Staph. aureus* exhibited all its initial metabolic properties when transferred to ordinary rich laboratory media.

A nutrient may therefore be defined as a substance that is essential either for minimal growth, perhaps under restricted conditions, of an organism trained to be as unexact as possible, or for optimal growth. The studies of Mueller (1910), for example, on the nutritional requirements of the diphtheria bacillus, have been conducted on the basis of optimal growth, which he defines as the best attainable upon empirically devised media. In the first case, a medium is sought that will promote the growth of at least one of the cells originally inoculated, and the production of a trained culture is most probably due to selection of cells with the greatest synthetic powers. In the second case, not only the reproduction, but the ready and profuse growth of all the viable cells of the inoculum is aimed at. The substances required for this purpose over and above the minimal requirements are generally called stimulants of growth though Lwoff (1933) in his review of growth factors for micro-organisms distinguishes stimulants from 'starting factors' which are substances essential for the growth of an organism at the commencement of its training but which are dispensed with by the trained organism. In some cases, however the addition of a starting factor to the minimal medium enhances growth and the distinction breaks down. A true stimulant presumably acts by supplying in abundance an essential metabolite that the bacterium itself synthesizes only at a relatively slow rate.

With these reservations in mind we may note that for many heterotrophic bacteria in Stages 3 and 4 the essential nutrients, carbohydrate, nitrogenous and growth factors (vitamins) have been defined. Among bacteria pathogenic for man we have already cited *Staph aureus* and *C diphtheriae* and to these we may add *Str hemolyticus* (see McIlwain 1940), Group D streptococci (Woolley and Hutchings 1940), *Str salinarum* (Smiley Niven and Sherman 1943), *Str lactis* (Niven 1944), and *Str faecalis* (Niven and Sherman 1944), *Myc tuberculosis* (see, for example Lockemann 1942), *A meningitidis* (Frantz 1942), *A gonorrhoeae* (Gould, Hane and Mueller 1944), *Br abortus, melitensis* and *suis* (Koser, Breslove and Dorfman 1941, Koser and Wright 1942, McCullough and Dick 1942), and certain members of the *Salmonella* group (Johnson and Pettger 1943). The requirements of different strains of organisms belonging to the same species may vary widely some having greater, some lesser synthetic powers than the strain for which the essential nutrients were first defined. There is a large number of heterotrophic bacterial species for which the essential nutrients are as yet only partly defined, for the most part the carbohydrate and amino-acid requirements are readily established. For example the main body of essential nutrients is defined for certain clostridia (Elberg and Meyer 1939), pneumococcus Type III (Badger 1944), *Br tularensis* (Tamura and Gibby 1943), certain pasteurellae (Berkman 1942), and *Cl tetani* (Mueller and Miller 1942, Feeney, Mueller and Miller 1943, Pickett, 1943). These organisms are presumably in Stage 4, since the remaining unidentified factors are usually active in low concentrations and are probably accessory growth factors.

#### The Vitamin Requirements of Bacteria

In addition to foodstuffs needed for energy and synthesis, many of the heterotrophic bacteria require vitamins. A vitamin or accessory growth factor is an essential nutrient active in concentrations so small that it is unlikely to act as an important source of either carbon, nitrogen or energy. Such substances range from inorganic ions to complex organic substances like thiamin or riboflavin, and the majority so far described appear to be in some way connected with the function or the structure of enzyme-co-enzyme systems of the organism. It is customary to limit the term vitamin to organic compounds, but it should be noted that, on the ground of their activity in low concentrations, many inorganic substances have equal claim to the title. In certain conditions, for example, the

toxin production by *C. diphtheriae* is maximal with an optimal concentration of iron as low as  $2.8 \times 10^{-6}$  M (Pappenheimer and Johnson 1936), and maximal toxin production by *Cl. tetani* is similarly conditioned (Mueller *et al.* 1913).

It is impossible to review the relevant literature of the organic accessory growth factors here, summaries of recent work will be found in the reviews of Lwoff (1938) and Koser and Saunders (1938). In recent years the identification and purification of many bacterial growth factors makes it possible to assign to them the role of vitamin. As noted above, tryptophan is essential for *Salm. typhi*, whose needs are satisfied by a concentration of 0.00061 per cent, if an alternative source of nitrogen is supplied for the synthesis of protoplasm. Among other amino-acids we may note that cysteine is an essential accessory growth factor for *Staph. aureus*, being active in concentrations of less than  $10^{-6}$  M, though less complex S-compounds can replace it in a defined medium (Fildes and Richardson 1937), and glutathione is similarly essential for *V. gonorrhoeae* (Gould 1941).  $\beta$  alanine is a growth factor for *C. diphtheriae* (Mueller and Cohen 1937) though the organism is able to produce it from L-carnosine (Mueller 1938). In *C. diphtheriae*  $\beta$  alanine may be the precursor of pantothenic acid, but Evans, Handley and Hapgood (1939) found certain grass strains of *C. diphtheriae* which needed both  $\beta$  alanine and pantothenic acid. Pantothenic acid is apparently essential for certain streptococci (Mellman 1939, 1940, Woolley and Hutchings 1940, Schuman and Farrell 1941), lactic acid bacteria (Snell, Strong and Peterson 1939), *Brucella abortus* (Koser, Breslove and Dorfman 1941), *Pr. morganii* (Dorfman, Berkman and Koser 1942) and *Sh. flexneri* (Weil and Bliek 1941). Glutamine which can be synthesized by the cocci from glutamic acid is required by *Str. hemolyticus* (Fildes and Gladstone 1939, Mellman, Fildes, Gladstone and Knight 1939) it accelerates the growth of a large number of organisms. Of the recognized animal vitamins thiamin was first recognized as a growth stimulant for bacteria by Tatum, Wood and Peterson (1936) working with the propionic acid bacteria *Staph. aureus* which also needs thiamin as an essential metabolite can synthesize it from pyrimidine and thiazole (Knight 1937, Knight and Mellman 1938). Thiamin the phosphate of which acts as a co-enzyme in carboxylase systems, probably plays a fundamental metabolic role in all micro organisms (see, for example, Quastel and Webley 1941, 1942) those for which it is not an essential nutrient being able to synthesize it. Some dysentery bacilli for example, apparently synthesize thiamin in a defined medium containing nicotinic acid (Dorfman, Koser, Reames, Swingle and Saunders 1939).

The role of nicotinic acid has been studied extensively. It is an essential nutrient of *Staphylococcus* (Knight 1937, Hoiday 1937), *C. diphtheriae* (Mueller 1937), some dysentery bacilli (Koser, Dorfman and Saunders 1940), *Proteus* (Fildes 1938, Pelczar and Porter 1940), *Brucella* (Koser, Breslove and Dorfman 1941), and *Ham. pertussis* (Hornbrook 1940), though, as with other essential nutrients not all strains of a given species require it. The significance of nicotinic acid lies in its relationship with co-enzyme I and co-enzyme II, which are respectively diphosphopyridine nucleotide and triphosphopyridine nucleotide. It is supposed that nicotinic acid, or nicotinamide is synthesized into co-enzyme I by organisms that cannot themselves synthesize the pyridine ring. As we have seen co-enzyme I is essential in certain dehydrogenase systems.

*Ham. influenzae* and *Ham. para influenzae* have even more restricted synthetic powers. *Ham. para influenzae* requires co-enzyme I already synthesized for many

of its dehydrogenations (Lwoff and Lwoff 1937*a b*) According to Schlenk and Gingrich (1942) the co-enzyme can be replaced by the nicotinamide riboside, while a mixture of nicotinamide *d* ribose and adenylic acid is inactive. The utilization of the riboside may thus be considered the limit of adaptability of this organism (Gingrich and Schlenk 1944) in the direction of the synthesis of co-enzyme I. Many bacteria are capable of synthesizing co-enzyme I from simpler substances and can therefore stimulate the growth of the influenza and para influenza bacilli by reason of their co-enzyme content. For instance certain lactic acid bacteria can utilize the purine and pyrimidine bases (Stokstad 1941) and *Cl. tetani* the purine base alone. The conception of nicotinic acid simply as a building block for co-enzyme I in all bacteria has recently been questioned by Saunders, Dorfman and Koser (1941). In a deficient medium the stimulating effect of co-enzyme I on dysentery bacilli was less than that of the nicotinamide supposed to serve for its synthesis. Co-enzyme I split by hydrolysis on the other hand was as active as nicotinamide. It is possible that in this case the nicotinamide may be utilized to form a respiratory haemochromogen with haemin. The close relation of nicotinic acid with glycolysis is illustrated by the action of *Salm. paratyphi A* and *Sh. shiga* on fermentable carbohydrates. According to Kligler and Grossowicz (1940 1941) nicotinic acid is not a growth promoting substance for these organisms. In its absence there is a slow and partial aerobic glycolysis, and in consequence only a slow protein accumulation. In its presence good growth occurs and the carbohydrate is rapidly oxidized with the production of CO<sub>2</sub>. In *Staph aureus* nicotinic acid is essential for the oxidation of glucose which under aerobic conditions is converted into pyruvic and lactic acids. The addition of thiamin as well permits the oxidation of the pyruvate, glycolysis as a whole is more active and the end products are acetic and lactic and carbonic acids. In the absence of nicotinic acid in a synthetic medium glucose has an inhibitory effect on growth perhaps by interfering with the oxidation of the essential amino-acids present (Kligler Grossowicz and Bergner 1943*a b*).

Just as the influenza bacillus is incapable of synthesizing the co-enzyme needed for its dehydrogenase systems in the same way it requires a supply of haemin ready made for incorporation in the enzyme systems concerned with oxygen transport. It has long been known that *Ham. influenza* requires haematin (the X factor see Chapter 33) and that grown anaerobically the organism can dispense with it. Lwoff and Lwoff (1937*c*) concluded from respiratory studies that the haematin was needed for the synthesis of cytochrome cytochrome oxidase catalase and peroxidase i.e. it was essential to the completion of the chain of mechanisms transferring hydrogen to atmospheric oxygen.

Among other accessory growth factors of known composition needed by bacteria we may briefly mention riboflavin for lactobacilli (Snell and Peterson 1940) and streptococci (McIlwain 1940 Woolley and Hutchings 1940 and Schuman and Farrell 1941), lactoflavin for propionibacteria and streptococci (Orla-Jensen Otte and Snog Kjaer 1936) and pimelic acid for *C. diphtheria* (Mueller 1937) *p*-aminobenzoic acid (see p. 162) pyridoxin for certain streptococci (Pappenheimer and Hotte 1940, Schuman and Farrell 1941) and oleic acid for *C. diphtheria* (Cohen, Snyder and Mueller 1941) and *Erysipelothrix* (Hutner 1942).

It will be seen that the majority of bacterial vitamins are also vitamins for other organisms, including the higher mammals. Pimelic acid is of particular interest, since it has not been recognized as a vitamin in other types of organisms nor is it a compound that seems likely to play the part of a co-enzyme in bacterial metabolism. Its significance



as a growth factor, however has become a little clearer with the identification of biotin (du Vigneaud, Hofmann and Melville 1942). Withers (1931) described a growth promoting fraction in yeast extract, to which he gave the name 'bios'. Many of the individual growth promoting substances in bios have been identified and include: inositol, *l*-leucine,  $\beta$ -alanine, thiamin and pantothenic acid (see Koser and Saunders 1938) and pyridoxin (Schultz *et al.* 1939, Eakin and Williams 1939). There remained biotin (Kögl and Tonnies 1936), a remarkable growth factor active in concentrations of  $10^{-10}$  or less. Biotin proves to be a keto imidazolido thiophane valeric acid, and it is possible that pimelic acid may contribute the fatty acid chain in the bacterial synthesis of biotin by *C. diphtheriae*. In support of this, du Vigneaud, Dittmer, Hague and Long (1942) found that pimelic acid and biotin behaved alike in the growth stimulation of *C. diphtheriae*. Working with the mould *Aspergillus niger*, Eakin and Eakin (1942) showed that not only was pimelic acid a growth stimulant, but its addition to the medium increased the biotin content of the culture about twenty fold.

Biotin itself has been found essential for a number of bacteria including *Str. pyogenes* (Hottle, Lampen and Pappenheimer 1941), *Staph. aureus* (Porter and Pelczar 1941) and *Br. abortus* (Koser, Breslove and Dorfman 1941).

Biotin preparations are apparently essential for *Cl. acetobutylicum* (Oxford, Lampen and Peterson 1940), *Brucella* (Koser, Breslove and Dorfman 1941), *Str. pyogenes* (Hottle, Lampen and Pappenheimer 1941), and *Staph. aureus* (Porter and Pelczar 1941). There are also many factors as yet not fully identified, such as an apparently lipid stimulating factor for the tubercle bacillus (Boissevain and Schultz 1938). Another vitamin like substance as yet unidentified that appears to have a wide distribution in bacteria first isolated by Knight and Fildes (1933) from a yeast gum, was necessary for *Cl. sporogenes* and *Cl. botulinum* (Fildes 1935). It is probably synthesized by a number of organisms including *Salmonella typhimurium* and the tubercle bacillus.

The identity of many bacterial and animal vitamins and the discovery of large numbers of bacterial species for which the various vitamins are essential nutrients, enables us to use bacteria in the assay of these substances for therapeutic and other purposes. The vitamin for assay is added in varying concentrations to cultures of the bacterium for which it is an essential nutrient, in a medium lacking the vitamin and the growth promoting effect estimated. Clearly the assay will be accurate only with bacteria whose growth requirements are defined in every respect. Even so errors will arise, for minute traces of contaminating vitamins may be present in the materials used for the basal medium and the preparation of vitamin to be assayed may contain substances which are not essential nutrients, but nevertheless have a stimulating action on growth.

(For a discussion of the B vitamin content of media in common bacteriological use, see Stokes, Guinness and Foster 1944.)

### The Gaseous Requirements of Bacteria

Examples of autotrophic bacteria utilizing hydrogen and  $\text{CO}_2$  have been given in previous sections, but  $\text{CO}_2$  has been considered only as a main source of carbon.  $\text{CO}_2$  may also act as a subsidiary source of carbon, or as an accessory growth factor. Reference to Chapter 34 will afford a striking example of a heterotroph, which on first isolation from the tissues fails to grow in air unless 5 to 10 per cent of  $\text{CO}_2$  is added to the atmosphere. Other species flourish only in a similar excess, while others still require  $\text{CO}_2$  but in smaller concentrations.

Numerous observations of the  $\text{CO}_2$  effect suggest that this gas is necessary for the growth of many bacterial species, including anaerobes (see for instance, Chapin 1918, Cohen and Fleming 1918, Rockwell and McWhann 1921, Rockwell 1921, 1923, 1924, Rockwell and Highberger 1926, 1927, Valley and Rettger 1927, Valley 1928, McLeod, Coates,

Happold Priestley and Wheatley 1934 Atkin Barling and Miles 1936 Nye and Lamb 1936 Khairat 1939 Fleming 1941 Kemjner and Schlaver 1942)

In some cases the gas stimulates not growth but the appearance of some particular metabolite. On artificial media for example *Staph aureus* (Chap 2a) produces significant amounts of its characteristic toxin in air containing 20 per cent of  $\text{CO}_2$  and the anthrax bacillus its capsular substance (Ivanovici 1937) in the presence of an excess of  $\text{CO}_2$ .

The use of defined media has made the study of  $\text{CO}_2$  requirements more exact. Walker (1937) found that *Bact coli* failed to grow in a liquid synthetic medium when incubated in a current of  $\text{CO}_2$  free air. Walker concluded that  $\text{CO}_2$  was necessary for growth and that the lag phase (see Chapter 4) represented the time taken by the organism to produce in its immediate vicinity a growth promoting concentration of  $\text{CO}_2$ . Gladstone Fildes and Richardson (1935) demonstrated a similar inhibition for *Salmonella typhi*, *Ps. pyocyanea*, *B. subtilis*, *B. anthracis*, *C. diphtheriae*, *C. sporogenes* and *C. welchii* by the passage of  $\text{CO}_2$  free gas through culture media.

The nature of the stimulating action of  $\text{CO}_2$  is not clear. Carbon dioxide is utilized even by heterotrophic bacteria. Wood and Werkman (1936, 1938, 1940) and Krebs and Eggleston (1941) demonstrated its fixation by propionibacteria and its probable role in the synthesis of a four-carbon chain from pyruvic acid. Wood and his colleagues (1940) by using  $\text{CO}_2$  made from the radioactive carbon isotope  $\text{C}_{14}$  was able to trace the isotope as far as succinic acid. *Bact coli* and some clostridia utilize  $\text{CO}_2$  (Ruben and Kamen 1940). Hes (1938) on the other hand suggests that the  $\text{CO}_2$  acts as a catalyser in several oxido-reduction reactions. The possible interrelationship of  $\text{CO}_2$  with enzyme systems is evident from the work of Pappenheimer and Hottel (1940) who found that adenylc acid was a growth stimulant for *Str. pyogenes* if the  $\text{CO}_2$  concentration was as low as 0.25 per cent. but was dispensable if the concentration was raised to 2.5 per cent. Similarly McCullough and Dick (1942) found that the incapacity of forty-one strains of *Br. abortus* to grow in the absence of a high concentration of  $\text{CO}_2$  in the atmosphere (see Chapter 34) was associated with incapacity to grow in a certain medium containing amino-acids and four vitamins. After the strains had been trained to grow in air thirty of them grew in the basal medium.

### Aerobiosis and Anaerobiosis

It is customary to divide bacteria into three categories in regard to their behaviour towards molecular oxygen. (1) the *obligatory anaerobes*—or *anaerobes* without the qualifying adjective—which will grow only when oxygen is rigorously excluded (2) the *facultative anaerobes* which will grow both aerobically and anaerobically i.e. in the presence or absence of oxygen and (3) the *obligatory aerobes* which will grow only when supplied with molecular oxygen.

Most of the organisms in Class (2)—to which incidentally the great majority of the bacteria with which we are concerned belong—are able to grow over a very wide range of oxygen pressures but some species prefer a relatively restricted range lying well below that of ordinary atmospheric conditions.

It should be noted that in the fluid cultures commonly employed in bacteriology the degree of oxygenation in cultures of aerobic bacteria is by no means necessarily optimal. The oxygen content in the depths of a broth culture may be very low (Rahn and Richardson 1941). In many cases forced aeration of the culture greatly increases growth (see Wilson 1930, Rahn and Richardson 1942) though the effect may in some cases be due as much to removal of  $\text{CO}_2$  as to the increased oxygen supply.

In a large number of aerobic and facultatively anaerobic bacteria the capacity

to grow in the presence of oxygen and to utilize it depends on the possession of a cytochrome cytochrome oxidase system. The molecular oxygen may, however, be utilized by other respiratory mechanisms for some facultative anaerobes have no cytochrome. When the organisms are deprived of oxygen the predominant metabolic mechanisms change with a consequent change in the nutritional requirements of the organism. It was Pasteur's observations on the different metabolic products associated with the aerobic and anaerobic growth of certain bacterial species that led him to place so great an emphasis on the absence of oxygen as a determinant factor in bacterial fermentation. We have already noted some of the changes in nutritional requirements consequent on anaerobic growth. The change may concern energy sources or essential nutrients or accessory growth factors. Anaerobically *H. influenza* can dispense with haematin since it has no need of haemochromogen catalysts for oxygen transfer. The pyrimidine base uracil is required as an essential nutrient in addition to thiamin and nicotinic acid when *Staph. aureus* is grown anaerobically.

It is clear that the respiratory enzyme systems of bacteria may be adapted to aerobic or anaerobic oxidations. It is much less clear, however, why the strict anaerobes should be incapable of growth in the presence of oxygen. The explanation of this inhibitory action of oxygen so far advanced has been made in terms of oxidation-reduction potentials in the medium or the production by the anaerobes of toxic oxidation products when exposed to air.

The electrical measurement of reducing intensity developed in a culture of bacteria gives us a positive or negative value for the Eh in volts. The greater the reducing intensity the more negative this value. The Eh level in a culture is dependent upon a large number of factors. Alterations of pH for example will alter the concentration of available electrons and so alter the Eh. Moreover a particular Eh value cannot be ascribed to a particular oxidation-reduction system in the culture, but may be the resultant of several systems, each of which influences the observed level according to its characteristic E<sub>0</sub>, its relative predominance in the culture, its speed of action and the poisoning effect it has upon the culture as a whole. But though it has proved impossible to sort out all these contributory factors, or to make any valid inferences about Eh levels inside or at the surface of bacterial cells themselves, observations upon the Eh changes in culture throw some light upon the conditions governing the growth of aerobic and anaerobic bacteria. For instance the aerobic growth of *Str. pyogenes* in a medium having an initial Eh of +0.3 volt is accompanied by a gradual fall in potential to about 0.15-0.2 volt. The spores of an anaerobe like *C. tetani* will not grow in a similar medium unless the starting Eh is as low as +0.11 volt (Knight and Fildes 1930) though once established the organism itself reduces the Eh to still lower levels. The limiting potentials that will allow the growth of certain bacterial species have been determined in a number of cases (see McLeod 1930, Hewitt 1936, Gillespie and Rettger 1938a, b, Gillespie and Porter 1938, Stephenson 1939). It appears that in general anaerobic organisms unlike the aerobes are unable to reduce ordinary aerobic media to the Eh level at which they can germinate though once growth is established they can maintain it at a low level, the medium must be partially reduced for the culture of anaerobes. In bacteriological practice the reduction is usually achieved by the exclusion of molecular oxygen from the culture. The reduction of oxygen pressure by this means is not however a necessary feature of anaerobic culture (see for example, Kligler and Guggenheim 1938), anaerobiosis may be achieved even though molecular oxygen has access to the medium by the addition of reducing substances like thioglycolic acid (Quastel and Stephenson 1926), reduced iron (Hastings and McCoy 1932), granules of cooked meat (Lepper and Martin 1929, 1930a, b) or by the concomitant growth of an aerobic organism capable of bringing the Eh down to the required level (see also Reed and Orr 1913). It may be that oxygen *per se* is inhibitory.

but that the presence of reducing substances in the medium prevents its direct access to the organisms, or the inhibiting effect may be due to the formation of oxidation products through the mediation of catalysts which are inactivated in cultures at Eh levels permitting the growth of obligate anaerobes. The change over from aerobic to anaerobic growth may in fact be brought about by artificial inactivation of enzyme systems. Brokhahn and Mirsky (1938) by cyanide-inhibition of enzymes capable of reacting with molecular oxygen induced *Bact. coli* to break down glucose anaerobically in the presence of air and it is obvious that similar inhibitions may take place as a result of changes in metabolic processes that are reflected in the lower Eh levels of anaerobic cultures. The nature of the hypothetical inhibitory oxidation products is still in doubt. It has long been known that certain bacteria are sensitive to hydrogen peroxide (Traugott 1893; Freer and Norry 1909). This sensitivity is associated with absence of catalase production. The pneumococcus, for instance, produces  $H_2O_2$  to which it is sensitive, and no catalase (McLeod and Gordon 1922; Avery and Morgan 1924; Avery and Neill 1924; b; c; Neill and Avery 1925). Other organisms, like *Sh. shiga*, are moderately sensitive but produce neither  $H_2O_2$  nor catalase. Most bacterial species produce catalase more or less actively (see Gottstein 1893; Löwenstein 1903; Rywosch and Pywosch 1907). The fact that anaerobes produce no catalase led McLeod and Gordon (1925a, b) to suggest that the sensitivity of obligate anaerobes to oxygen is due to their readiness to form in its presence inhibitory concentrations of  $H_2O$ . The acceptance of this hypothesis depends on the demonstration of oxygen utilization and  $H_2O_2$  production by the anaerobes in the presence of oxygen. McLeod and Gordon grew the anaerobes in blood media, and inferred the production of  $H_2O_2$  from certain colour changes in the medium. Similar changes, however, can be produced by reducing systems in the absence of air (Anderson and Hart 1934) and do not necessarily indicate  $H_2O_2$ . Moreover Cook and Stephenson (1925) were unable to demonstrate oxygen utilization by *Cl. sporogenes* in conditions that would have detected the oxygen uptake equivalent to the formation of 1/50 000  $H_2O_2$  in their suspensions.

#### Other Factors Influencing the Growth of Bacteria.

**Hydrogen Ion Concentration.**—For any given species of bacterium there is an optimal and relatively narrow range of pH allowing vigorous growth and a wider range extending on each side of the optimum over which growth occurs less vigorously. For most of the bacteria with which we are concerned the optimal pH lies a little to the alkaline side of neutrality (pH 7.2–7.6). The range of pH over which growth is possible has not been accurately determined for many bacterial species, but for most pathogenic species it would appear to extend over some such range as pH 5.0 to pH 8.0 (see Chapter 9).

There are, however, species that show very distinctive variations from this modal range of sensitivity. *Lactobacter*, for instance, is very sensitive to acids, and will not grow in pure culture at a pH lower than 6.5 (Fred and Davenport 1915). The aciduric bacilli of the genus *Lactobacillus* are, on the other hand, highly resistant to acids, and will apparently grow to some extent at pH 4.0 or even less, though the power to grow has not, in this instance, been decisively distinguished from the power to resist the lethal action of the acid (McIntosh *et al.* 1922, 1924). The cholera vibrio is very tolerant of alkali, relatively sensitive to acid. Its optimum for growth is pH 7.6–8.0 and its limits for growth about pH 6.4–9.6. The enterococcus affords a good example of an organism with a wide growth range, extending from pH 4.8 to pH 11.0 (Downie and Cruickshank 1928; Davis and Thiel 1939).

**Temperature.**—We need only note in this section that for each species of bacterium there is an optimal temperature for growth and a range of temperature over which growth is possible. For most pathogenic bacteria the optimal temperature for growth is in the neighbourhood of 37°C. and the range of temperature over which growth occurs is approximately 15°–40°C. Here as elsewhere, however, there are wide variations, especially when we include in our survey non-pathogens as well as pathogens. In their reactions to changes in temperature bacteria as a whole display in a striking fashion their capacity for adaptation to a wide range of environmental conditions.

To take a few illustrative examples among pathogenic and potentially pathogenic species, *Bact. coli* grows best at  $37^{\circ}\text{C}$ , but will grow at any temperature within the range of  $15^{\circ}$ – $45^{\circ}\text{C}$ . Among the Gram negative cocci there are characteristic differences in reaction to temperature, thus *N. gonorrhoeae*, *N. meningitidis* and *N. catarrhalis* all show optima at about  $37^{\circ}\text{C}$ , but the range of growth of *N. catarrhalis* extends from approximately  $18^{\circ}$  to  $42^{\circ}\text{C}$ , while *N. gonorrhoeae* and *N. meningitidis* show a very restricted range of about  $30^{\circ}$  to  $38^{\circ}\text{C}$ . The different types of tubercle bacilli show temperature optima in conformity with the body temperatures of the host species that they infect thus the human and bovine types of tubercle bacillus grow best at  $37^{\circ}\text{C}$  and fail to grow below  $30^{\circ}\text{C}$ ; the avian type grows best at  $40^{\circ}\text{C}$ , and again fails to grow below  $30^{\circ}\text{C}$  while the cold blooded type grows freely at  $22^{\circ}\text{C}$ .

As examples of non pathogenic species that depart from the temperature optima given above we may note (see Buchanan and Fulmer 1928–30) that bacteria have been isolated from fish, brine and similar sources that grow well at  $0^{\circ}\text{C}$ , while from a variety of natural sources (soil, excreta, silos and especially hot springs) thermophilic species have been isolated that have optima at  $55^{\circ}\text{C}$ . or over, and are able to multiply at a temperature of  $75^{\circ}\text{C}$ . These thermophiles are of considerable economic importance, since they are a source of difficulty when it is desired to sterilize any material at a relatively low temperature (see Chapter 5).

There appears to be a definite relationship between the thermolability of enzyme systems in a given organism, and the maximum temperature at which growth occurs. In a study of eighteen species, including thermophilic organisms, Edwards and Rettger (1937) demonstrated close agreement between the minimum temperatures at which the three thermolabile respiratory enzymes—catalase, succinic dehydrogenase, and cytochrome oxidase—were destroyed and the maximum growth temperatures. The temperature at which the relatively thermostable peroxidase was destroyed was not related to growth temperatures in this way.

The relationship also appears to hold for spores, for Lamanna (1942), working with seventy two strains of the spore bearing bacillus species, found an association between the maximum temperature at which growth took place and the length of time the spore resisted heating at  $95^{\circ}\text{C}$ .

In this connection, we may note the effects of heat on bacteria prior to their growth in a given medium. If the death of a bacterium depends on the inactivation of certain enzymic systems, or the destruction of some essential nutrient, we might expect that sub lethal degrees of heating would destroy some but not all of the essential metabolic functions of the bacterium. For instance, yeast cells, increasingly exposed to heat, first lose their ability to reproduce, and then their fermentative ability (Rahn and Baroes 1932). The count of viable bacteria from cultures subjected to sub lethal doses of heat or ultra violet light may be greatly increased if the test media are enriched with blood or extra carbohydrates (Fay 1934, Curran and Evans 1937, Nelson 1943). Davis (1940) records a phenomenon he refers to as "pseudo death" among spores of *Cl. welchii*. Only a small number of originally viable spores resisted a certain exposure to heat sufficiently to produce colonies when subsequently seeded on to a standard agar medium. In some cases the area of agar seeded with heated spore suspension was free of colonies after 5 days' incubation. When a fresh culture of *Cl. welchii* was grown on the agar adjacent to the original inoculum, a number of the heated spores germinated. Presumably these "pseudo-dead" spores were without some heat labile substance essential for germination, which was supplied by the growing *Cl. welchii*.

#### Other Biochemical and Physiological Activities of Bacteria

We have endeavoured to present a broad picture of bacterial metabolism, particularly as it affects the heterotrophic organisms that constitute the greater part of the bacteria pathogenic for man and the higher animals. Many other activities of bacteria—the production of pigments and of various toxic substances,

their action on blood cells and blood pigments their particular growth requirements—are specialized activities, so characteristic of particular bacterial species that it will be convenient to consider them in later chapters. Mention may be made here of the work of Reid (1937) and Kharasch, Conway and Bloom (1936) on the effect of food and inhibitory substances on the formation of pigment by growing cultures of chromogenic bacteria, and by Baker (1933) on the influence of light on pigment production by a species of non pathogenic *Mycobacterium*. In Chapter 9 we shall consider the physiological changes that constitute one aspect of bacterial variation, and in Chapter 6 the relation of chemotherapy to the metabolic processes of bacteria.

## REFERENCES

- AITKEN, R. S., BAEHLING B. and MILES A. A. (1936) *Lancet*, ii, 780  
 ANDERSON A. B. and HART P. d. A. (1934) *J. Path. Bact.*, 39, 465  
 ARONSON H. (1900) *Arch. Kinderheilk.*, 30, 23  
 AVERY, O. T. and CULLEN, G. E. (1920) *J. exp. Med.*, 32, 571  
 AVERY, O. T. and MORGAN, H. J. (1924) *J. exp. Med.*, 39, 275, 289  
 AVERY, O. T. and NELL, J. M. (1924a) *J. exp. Med.*, 39, 347, (1924b) *Ibid.*, 39, 357, (1924c) *Ibid.*, 39, 543  
 BADGER, F. (1944) *J. Bact.* 47, 509  
 BAILEY, F. A. (1911) *J. Hyg., Camb.*, 11, 341  
 BAKER, J. A. (1935) *J. Bact.* 35, 62  
 BAKER, J. W. and HAPFOLD F. C. (1940) *Biochem. J.*, 34, 657  
 BARON E. S. G. and LYMAN C. M. (1939) *J. biol. Chem.*, 127, 143  
 BERMAN, N. and RETTGER, L. F. (1918) *J. Bact.*, 3, 367, 389  
 BERKMAN, S. (1942) *J. infect. Dis.* 71, 201  
 BERNHEIM F., BERNHEIM M. L. C. and WERTER, M. D. (1935) *J. biol. Chem.*, 110, 165  
 BIGGER, J. W. and NELSON, J. H. (1943) *J. Path. Bact.*, 55, 321  
 BIRKENSHAW, J. H., CHARLES, J. H. V., and CLUTTERBUCK, P. W. (1931) *Biochem. J.*, 25, 1522  
 BOISSEvain C. H. and SCHULTZ, H. W. (1933) *Amer. Rev. Tuberc.*, 33, 624  
 BOIVIN, A. and MESROBEANT, L. (1934) *Arch. roum. Path. exp. Microbiol.*, 7, 95.  
 BOOTH, V. H. and GREEN D. E. (1935) *Biochem. J.*, 32, 855  
 BORD G. DE. (1923) *J. Bact.*, 8, 1  
 BRAUN, H. (1931) *Zbl. Bakt.*, 122, 5  
 BRAUN, H. and CAHN BROUWER, C. E. (1921) *Zbl. Bakt.*, 88, 1, 196, 350, (1922) *Biochem. Z.*, 131, 226, 272  
 BRAUN, H. and GOLDSCHMIDT, R. (1924) *Biochem. Z.*, 146, 573, (1927) *Zbl. Bakt.*, 101, 283, 330, (1928a) *Ibid.*, 107, 329, (1928b) *Ibid.*, 109, 353  
 BRAUN, H., HOFMEIER, K., and MÜNDL, F. (1929) *Zbl. Bakt.*, 113, 530  
 BRAUN, H. and KOWDO, S. (1924) *Klin. Wochs.*, 3, 10  
 BRAUN, H. and LISCHE, H. (1928) *Zbl. Bakt.*, 107, 35  
 BRAUN, H. and MÜNDL, F. (1927) *Zbl. Bakt.*, 103, 182, (1929) *Ibid.* 112, 347  
 BRAUN, H., STAMATZAKIS, A. and KOWDO, S. (1924) *Biochem. Z.*, 145, 331  
 BRAUN, H. and VASARHELYI, J. VON (1933) *Zbl. Bakt.*, 127, 105  
 BRAUN H. and WÖRDEHOFF, P. (1933) *Zbl. Bakt.*, 128, 50  
 BROH KAHN R. H. and MIESKY, I. A. (1938) *J. Bact.*, 35, 455  
 BROOM J. C. (1929) *Brit. J. exp. Path.*, 10, 71  
 BUCHANAN, R. E. and FLYNN, E. I. (1928-30) "Physiology and Biochemistry of Bacteria." (3 vols.) New York and London.  
 BURE, D. (1934) *Ergebn. Enzymforsch.*, 3, 23  
 BURROWS W. (1933) *J. infect. Dis.*, 52, 126, (1933a) *Ibid.*, 64, 145, (1933b) *Ibid.*, 65, 134  
 CALLOW, A. B. (1923) *J. Path. Bact.*, 26, 320, (1924) *Biochem. J.*, 18, 507  
 CALLOW, A. B. and ROBINSON, M. E. (1925) *Biochem. J.*, 19, 19  
 CARRIAGE G. (1901) *C. R. Soc. Biol.*, 53, 350  
 CHAPIN C. W. (1918) *J. infect. Dis.*, 23, 342  
 CLARK, W. M. (1928) "The Determination of Hydrogen Ions." Baltimore and London.  
 CLARK, W. M., et al (1928) *Bull. U.S. Hyg. Lab.*, No 151  
 CLIFTON, C. E. (1940) *J. Bact.*, 39, 455, (1942) *Ibid.*, 44, 179  
 COGHILL, P. D. (1931) *J. biol. Chem.*, 90, 57

- COHEN, M B and FLEMING, J S (1918) *J infect Dis*, 23, 337  
 COHEN, S., SNYDER J C and MUELLER, J H (1911) *J Bact*, 41, 581  
 COLLINS, M A and HAMNER, B W (1934) *J Bact*, 27, 473, 487  
 COOK, R P (1931) *Zbl Bakt*, 122, 329  
 COOK, R P and STEPHENSON, M (1928) *Biochem J*, 22, 1368  
 COOK, R P and WOOLF, B (1928) *Biochem. J.*, 22, 474  
 CRAMER, E (1891) *Arch Hyg*, 13, 71, (1893) *Ibid*, 18, 151, (1894) *Ibid*, 20, 197, (1895) *Ibid*, 22, 167, (1897) *Ibid*, 28, 1  
 CURRAN H R and EVANS, F R (1937) *J Bact*, 34, 179  
 DAVIS H (1940) *Quart J Pharm*, 13, 14  
 DAVIS, J G and THIEL, C C (1939) *J Dairy Res*, 10, 457  
 DAWSON, A I (1919) *J Bact*, 4, 133  
 DEWAS, J G and GREEN, D E (1938) *Biochem J*, 32, 626  
 DORFMAN, A, BERKMAN, S and KOSER S A (1942) *J biol Chem* 144, 393  
 DORFMAN, A, KOSER S A, REAMES, H R, SWINGLER, K F and SAUNDERS, F (1939) *J infect Dis*, 65, 163  
 DOWNTON A W and CRUICKSHANK, J (1928) *Brit J exp Path*, 9, 171  
 EAKIN, R E and EAKIN, E A (1942) *Science*, 96, 187  
 EAKIN, R E and WILLIAMS, R J (1939) *J Amer chem. Soc* 61, 1932  
 ECKSTEIN, H C and SOULE, M H (1931) *J biol Chem*, 91, 395  
 EDWARDS, O F and REITGER, L F (1937) *J Bact*, 34, 489  
 EHRLICHMAN, O (1937) *Z Hyg InfektKr*, 119, 572  
 EHRLICHMAN, O, and DRAMBURG, K (1937) *Ibid*, 119, 623  
 ENJEMAN, C (1901) *Zbl Bakt*, 29, 841  
 ELBERG S S and MEYER, K F (1939) *J Bact*, 37, 429  
 ELSDEN S B (1938) *Biochem J* 32, 187  
 ENDO, S (1938) *Biochem Z* 296, 56  
 EVANS, E A, VENNESLAND B and SLOTIN, L (1943) *J biol Chem*, 147, 771  
 EVANS, W C HANDLEY, W R C and RAFFOLD, F C (1933) *Brit J exp Path* 20, 396  
 (1941) *Biochem J*, 35, 207  
 FAX, A C (1934) *J agric Res*, 48, 453  
 FELLEY, R E, MUELLER, J H and MILLER, P A (1943) *J Bact* 46, 509  
 FIELDS, P (1935) *Brit J exp Path*, 16, 309, (1938) *Biochem J*, 32, 1600  
 FIELDS, P and GLADSTONE, G P (1939) *Brit J exp Path*, 20, 334  
 FIELDS, P, GLADSTONE, G P, and KNIGHT, B C J G (1933) *Brit J exp Path*, 14, 189  
 FIELDS, P and KNIGHT, B C J G (1933) *Brit J exp Path*, 14, 343  
 FIELDS, P and RICHARDSON, G M (1935) *Brit J exp Path* 16, 326, (1937) *Ibid*, 18, 232  
 FISCHER, H and HASENRAUP, J (1935) *Luebig's Ann*, 515, 148  
 FLEMING, A (1941) *Lancet*, 110  
 FLEMING, W L and NEILL, J M (1927) *J exp Med*, 45, 169 947  
 FRANTZ I D (1942) *J Bact*, 43, 757  
 FREED, E B and DAVENPORT, A (1918) *J agric Res*, 14, 317  
 FRIEL, W L (1934) *Atmungs-farbstoffe bei pflanzlichen Mikroorganismen* Festschrift Zangger, 1, 805  
 FREER and NOVY (1902) *Amer chem. J.*, 27, 161  
 FRIEDHEIM, E A H. (1931) *J exp Med*, 54, 207, (1932) *C R Soc Biol* 110, 353, (1934) *Biochem J*, 28, 173  
 FRIEDHEIM E and MICHAELIS, L (1931) *J biol Chem*, 91, 355  
 FULMER, E I, NELSON, V E, and SHERWOOD, F F (1921) *J Amer chem Soc* 43, 191  
 GAYFROON, H (1933) *Biochem Z*, 260, 1, (1934) *Ibid*, 269, 447, (1935) *Ibid*, 275, 301, (1935) 279, 1  
 GALE, E F (1940) *Bact Rev* 4, 135  
 GALEOTTI, G (1898) *Z physiol Chem*, 25, 48  
 GILLESPIE, R W H and PORTER, J R (1938) *J Bact* 36, 633  
 GILLESPIE, R W H and REITGER L F (1938a) *J Bact* 36, 605 (1938b) *Ibid*, 36, 621  
 GINGRICH, W and SCHLEK, F (1944) *J Bact*, 47, 353  
 GLADSTONE, G P (1937) *Brit J exp Path*, 18, 322, (1938) *Ibid* 19, 208, (1939) *Ibid* 20, 189  
 GLADSTONE, G P, FIELDS, P, and RICHARDSON, G M (1935) *Brit J exp Path*, 16, 335  
 GLASS V and KENNETT S J (1939) *J Path Bact*, 49, 125  
 GLENY, T H (1911) *Zbl Bakt*, 58, 481  
 GOTTSTEIN, A (1893) *Virchows Arch* 133, 295  
 GOULD, R G (1944) *J biol Chem* 153, 143  
 GOULD, R G, KANE L W and MUELLER J H (1941) *J Bact*, 47, 287  
 GREEN, D E and STICKLAND L H, (1934) *Biochem J*, 28, 898  
 GREEN, D E, STICKLAND, L H, and TARR, H L A (1934) *Biochem J*, 28, 1812  
 GREY, E C (1913-14) *Proc roy Soc B* 461, (1924) *Ibid*, 96, 156  
 GUCCENHEIM, K (1944) *J Bact*, 47, 313

- HARDEN A (1901) *J chem Soc*, 79, 610. (1903) *J Hyg, Camb* 5, 483. (1930) "The Metabolism of Bacteria" "System of Bacteriology," 1, 203 *Med Res Coun.* London.
- HARDEN, A and WALPOLE, G S (1906) *Proc roy Soc, B*, 77, 399
- HARRIS J E G (1919) *J Path. Bact.* 23, 30
- HASTINGS, F G and MCCOY, E (1932) *J Bact* 23, 54
- HEAP, H and CADNESS, B (1924) *J Hyg., Camb*, 23, 77
- HEIRE, F J and SICO J Y (1942) *J exp Med*, 75, 339
- HEIDELBERGER, M and KENDALL, F E. (1931) *J exp Med*, 54, 515
- HES J W (1938) *Nature Lond.* 141, 647
- HESTRIN S and AVIVERI SHAPIRO S (1943) *Nature, Lond*, 152, 49. (1944) *Biochem. J.*, 38, 2
- HEWITT, L F (1936) "Oxidation Reduction Potentials in Bacteriology and Biochemistry" 4th Ed. L.C.C. London.
- HEYNINGER W E VAN (1940) *Biochem J.*, 34, 1540
- HILLS, G M. (1938) *Biochem J.*, 32, 333
- HIRSCH, J (1931) *Z Hyg InfektKr.* 112, 660
- HIRSCH, J and MÜLLER, A W (1933) *Z Hyg InfektKr.* 115, 443.
- HIRSCHLER (1898) *Z physiol Chem*, 10, 306
- HOLIDAY E R (1937) *Biochem J.* 31, 1299
- HOOGHEZIDE J C and KOCHOLATY, W (1938) *Biochem J.*, 32, 949
- HOPKINS F G (1921) *Biochem J*, 15, 236. (1923) *Lancet*, 1, 1251. (1929) *J. biol Chem*, 84, 269
- HORNIBROOK J W (1940) *Proc Soc exp Biol N Y.*, 45, 593
- HOSoya, S and KUROYA, M (1923) *Sci Rep Inst. infect. Dis Tokyo Univ.* 2, 233 265
- HOTTE, C A., LAMPEN J O and PAPPENHEIMER, A M (1941) *J biol Chem.*, 137, 457
- HUGHES, T P (1932) *J Bact.* 23, 437
- HUNTER, O W (1923) *J agric Res*, 24, 263
- HUSS, H (1908) *Zbl Bakt. Hte Abt.*, 20, 474
- HUTNER, S H (1942) *J Bact.*, 43, 629
- IVÁNOVICS, G (1937) *Zbl Bakt.*, 133, 449
- IVÁNOVICS, G and BRUCKNER, V (1937a) *Z Immunforsch.*, 90, 304 (1937b) *Ibid*, 91, 175
- JOHNSON, E A and RETTGER, L F (1943) *J Bact.* 45, 127
- JOHNSON, T B and BROWN E B (1922) *J biol Chem.*, 54, 721, 731
- JOHNSON, T B and COGHILL, R D (1925) *J biol Chem.*, 63, 225
- JONES, H M. (1916) *J infect Dis*, 18, 33
- KASERER, H (1900a) *Zbl Bakt. Hte Abt.* 15, 573. (1906) *Ibid.*, Hte Abt, 16, 681
- KEILIN D (1925) *Proc roy Soc, B*, 98, 312. (1926) *Ibid*, 100, 129. (1928-29) *Ibid.*, 104, 206. (1930) *Ibid*, 106, 418 (1943) *Proc. biochem Soc, Biochem. J.*, 37, xxi.
- KEILIN D and HARPLEY, C H (1941) *Biochem. J.* 35, 688
- KEILIN, D and HARTREE, E. F (1935a) *Proc roy Soc B.*, 124, 397. (1935b) *Ibid.*, 125, 171. (1935c) *Nature, Lond.* 141, 870
- KEMPNER, W and SCHLAYER, C (1942) *J Bact*, 43, 387
- KENDALL, A. I (1922) *J infect Dis*, 30, 211
- KENDALL, A. I., DAY, A A., and WALKER, A. W. (1914) *J Amer chem Soc*, 36, 1962
- KENDALL, A. I. and WALKER, A. W. (1915) *J infect Dis.*, 17, 442.
- KHAIRAT, O (1940) *J Path Bact* 50, 491, 497
- KHARASCH M. S CONWAY, E A. and BLOOM W (1936) *J Bact.*, 32, 533
- KLIGLER, I J and GROSSOWICZ N (1940) *Nature, Lond*, 146, 652. (1941) *J Bact.*, 42, 173
- KLIGLER, I J, GROSSOWICZ, N and BERGNER, S (1943a) *J Bact* 46, 399. (1943b) *Proc Soc exp Biol N Y.* 52, 332
- KLIGLER, I J and GUGGENHEIM, K (1938) *J Bact.*, 35, 141
- KLUYVER, A. J (1924) *Z physiol Chem* 133, 100
- \*KNIGHT, B C J G (1935) *Brit J exp Path.*, 16, 315. (1936) *Spec Rep Ser med. Res. Coun Lond.* No 210. (1937) *Biochem J.*, 31, 731 966
- KNIGHT, B C J G and FIELDS, P (1930) *Biochem J.*, 24, 1490. (1933) *Brit J exp Path.*, 14, 112.
- KNIGHT, B C J G and McILWAIN, H (1938) *Biochem J.* 32, 1241
- KNOER, M (1925) *Ergebn Hyg Bakt.*, 7, 641
- KOCHOLATY, W., WEIL, L. and SMITH, L. (1938) *Biochem. J.* 32, 1685
- KÖGL, F and HAAZEN SMIT, A. J (1931) *Proc Acad Sci Amst* 34, 1411
- KÖGL, F and TÖNNIS B (1936) *Z physiol Chem*, 242, 43
- KOSER, S A BRESLOVE, B B and DORFMAN, A (1941) *J infect Dis.*, 69, 114
- KOSER, S A., DORFMAN A and SAUNDERS, F (1940) *Proc. Soc exp Biol, N Y.*, 43, 391
- KOSER, S A and RETTGER, L F (1919) *J infect Dis*, 24, 301
- KOSER, S A and SAUNDERS F (1939) *Bact Rev* 2, 99
- KOSER, S A and WRIGHT M. H (1942) *J infect Dis*, 71, 86
- KREBS H A. (1937) *Biochem J.* 31, 661
- KREBS H A and EGLESTON, L V (1941) *Biochem. J.* 35, 676



- KREBS H A, HAFER, M M and EGLESTON, L V (1942) *Biochem J* 36, 306  
 LAMANTA, C (1942) *J Bact* 44, 29  
 LANTZSCH H (1922) *Zbl Bakt, Hte Abt.*, 57, 309  
 LEPPER E and MARTIN, C J (1929) *Brit J exp Path.*, 10, 327, (1930a) *Ibid.*, 11, 137, (1930b) *Ibid* 11, 140  
 LEIBOWITZ, J and HESTER, S (1942) *Biochem J*, 36 772  
 LINTON, R W, MITRA, B N, and SHRIVASTAVA D L (1934) *Indian J med Res.*, 21, 635.  
 LIPMAN, F (1937) *Enzymologia* 4, 65  
 LOCKEMAN, G (1942) *Z Hyg Infektkr* 124, 373  
 LOCKHART, E I and POTTER V R (1941) *J biol Chem* 137, 1  
 LOWENSTEIN (1903) *Wien klin Wschr.*, 16, 1393  
 LWOFF A (1938) *Ann Inst Pasteur*, 61, 580  
 LWOFF A and LWOFF, M (1937a) *Proc roy Soc B* 122, 352 (1937b) *Ibid* 122, 360 (1937c) *Ann Inst Pasteur*, 59, 129  
 McCULLOUGH N B and DICK, L A (1942) *J infect Dis* 71, 193 198  
 McILWAIN, H (1939) *Brit J exp Path.*, 20, 330 (1940) *Ibid* 21, 25  
 McILWAIN H, FIELDS, P, GLADSTONE G P and KNIGHT B C J G (1939) *Biochem J* 33, 223  
 MCINTOSH, J, JAMES W W, and LAZARUS BARLOW P (1922) *Brit J exp Path.*, 3, 138, (1924) *Ibid*, 5, 175  
 McLEOD, J W (1930) *System of Bacteriology Med Res Coun London* 1, 263  
 McLEOD J W., COATES J C., HAPFOLD F C., PRIESTLEY D P and WHEATLEY B (1934) *J Path Bact* 39, 221  
 McLEOD, J W and GORDON J (1922) *Biochem J* 16, 499 (1923a) *J Path Bact.*, 26, 326 (1923b) *Ibid*, 26, 332, (1924) *Biochem J.*, 18, 937 (1925a) *J Path Bact* 28, 147 (1925b) *Ibid*, 28, 155  
 McLEOD, J W and WYON, G A (1921) *J Path Bact* 24, 700  
 MASCHMANN, E (1937) *Biochem. Z* 294 1  
 MESROBEANU, L (1936) *Arch roum Path exp Microbiol* 9, 121  
 MEYER, K. (1934) *Zbl Bakt.*, 131, 289, 291  
 MICHAELIS L and NAKAHARA Y (1923) *Z Immunforsch* 36, 449  
 MULLER, F M (1933) *Arch Mikrobiol* 4 131  
 MUELLER, J H (1937) *J Bact.*, 34, 163 429 (1938) *J biol Chem* 123 4 1 (1940) *Bact Rev* 4 97  
 MUELLER, J H and CONY S (1937) *J Bact* 34, 381  
 MUELLER, J H and MILLER, P A (1942) *J Bact* 43, 763  
 MUELLER, J H, SCHOENBACH, E. B., JENKINWICZ, J J and MILLER I S (1943) *J clin Invest* 22, 315  
 NASSAU E (1942) *J Path Bact* 54, 443  
 NEILL, J M and AVERY, O T (1925) *J exp Med* 41, 285  
 NEILL, J M and FLEMING W L. (1927) *J exp Med.*, 45, 937  
 NELSON I E (1913) *J Bact* 45 390  
 NELSON, M E and WERKMAN C H. (1936) *Jour St Coll J Sci* 10, 141  
 NEUBERG C and REINFURTH E (1923 24) *Biochem Z* 143, 5 3  
 NICOLLE M and ALLAIRE E (1909) *Ann Inst Pasteur* 23, 547  
 NIEL, C B YAY (1931) *Arch Mikrobiol* 3, 1 (1933) *Yearb Carneg Instn No* 32 184 (1943) *Physiol Rev* 23, 338 (1944) *Bact Rev* 8, 1  
 NIEL, C B VAN RUDEN S CARSON S F, KAMEN M D and IOSTER, J W (1942) *Proc nat Acad Sci Wash* 28, 8  
 NIKLEWSKI, B (1908) *Zbl Bakt Hte Abt*, 20, 469  
 NISHIMURA, T (1893) *Arch Hyg*, 18, 318  
 NIVEN, C F (1944) *J Bact* 47, 343  
 NIVEN C F and SHERMAN J M (1944) *J Bact* 47, 335  
 NOACK, K. and SCHNEIDER, E (1933) *Naturwiss* 21, 635  
 NORD I F and DYDEL, W (1939) *Biochem Z* 296 153  
 NYE R N and LAMB, M E (1936) *J Amer med Ass* 106 1070  
 O CONNOR, R (1940) *Biochem J* 34, 1008  
 OGSTON, F J and GREEN D E (1935) *Biochem J*, 29, 1983  
 OMEARA, R A Q (1937) *J Path Bact* 45 541  
 OPPENHEIMER C. (1926) *Die Fermente und ihre Wirkung* 5th Edn., Leipzig  
 ORDAL, E. J and HALVORSON H O (1939) *J Bact.*, 33, 199  
 ORLA-JENSEN (1902) *Zbl Bakt, Hte Abt.*, 8, 11. (1909) *Ibid.*, 22, 305  
 ORLA-JENSEN S, OTTE N C and STOKHJAEV, A (1938) *Zbl Bakt., Hte Abt.*, 94, 431 447, 452  
 OXFORD A E, LAMPEN J O and PETERSON W H (1940) *Biochem J.*, 34, 1509  
 PARRY, W C C and JOLLYMAN W H (1901) *J chem Soc*, 79, 322.  
 PAPPENHEIMER, A M (1935) *Biochem. J.*, 29, 2037

- PAPPENHEIMER A M and HOTTLE, G A (1940) *Proc Soc exp Biol N Y*, 44, 645  
 PAPPENHEIMER A M and JOHNSON, S J (1936) *Brit J exp Path* 17, 335  
 PECKHAM, A W (1897) *J exp Med*, 2, 549  
 PELTZAR, M J and PORTER, J R (1940) *J Bact*, 39, 429  
 PENROSE M and QUASTEL, J H. (1930) *Proc roy Soc, B*, 107, 168  
 PESKETT, G L (1933) *Biol Rev*, 8, 1  
 PICKETT, M J (1943) *J biol Chem* 151, 203  
 PORTER, J I. and PELTZAR, M J (1941) *J Bact*, 41, 173  
 POTTER, V R (1941) *J biol Chem*, 141, 775  
 QUASTEL, J H (1925) *Biochem. J*, 19, 641, (1926) *Ibid*, 20, 166  
 QUASTEL, J H and STEPHENSON, M (1926) *Biochem J*, 20, 1125  
 QUASTEL, J H STEPHENSON, M, and WHETHAM, M S (1925) *Biochem J*, 19, 304  
 QUASTEL, J H and WELBY D M (1941) *Biochem J* 35, 192, (1941) *Ibid*, 36, 8  
 QUASTEL, J H and WHETHAM M S (1924) *Biochem J*, 18, 519, (1925a) *Ibid*, 19, 520, (1925b) *Ibid*, 19, 645  
 QUASTEL, J H and WOOLDRIDGE W R (1925) *Biochem J*, 19, 652, (1927a) *Ibid*, 21, 148, (1927b) *Ibid*, 21, 1234, (1928) *Ibid*, 22, 639  
 RABY O and BARNES M N (1933) *J gen Physiol*, 16, 579  
 RABY O. and RICHARDSON, G L. (1941) *J Bact*, 41, 223, (1942) *Ibid* 44, 321  
 RAISTRICK, H (1919) *Biochem J*, 13, 446  
 RAISTRICK, H and CLARK, A. B (1921) *Biochem J* 15, 70  
 RAISTRICK, H, *et al* (1931) *Philos Trans B*, 220, 1  
 REED G B and ORR, J H (1943) *J Bact*, 45, 309  
 REID R D (1937) *Zbl Bakt, Hte Abt.*, 95 373  
 RETTGER, L F BERMAN, N, and STURGES, W S (1916) *J Bact*, 1, 15  
 ROCKWELL, G E (1921) *J infect Dis* 28, 352 (1923) *Ibid* 32, 98, (1924) *Ibid*, 35, 381  
 ROCKWELL, G E and HIGHBERGER, J H (1926) *J infect Dis*, 33, 92, (1927) *Ibid*, 40, 438  
 ROCKWELL, G E and McHANN, C F (1921) *J infect Dis*, 28, 249  
 ROSLOFSON P A (1934) *Proc Konink Akad Wetensch, Amsterdam* 37, 660, (1935) "On the Photosynthesis of the *Thiorhodaceae*" (Thesis) Utrecht  
 RUBEN S and KAMEY M D (1940) *Proc nat Acad Sci, Wash.*, 26, 418  
 RUBNER, M (1900) *Arch Hyg* 38, 67  
 RUGGIERINI G (1933) *Biochim Terap sper* 20, 250  
 RYWOSCH, D and RYWOSCH, M (1907) *Zbl Bakt.*, 44 295  
 SANDIFORD, B R and WOOLDRIDGE, W R (1931) *Biochem J*, 25, 2172  
 SANDERS F, DORFMAN, A and KOSER, S A (1941) *J biol Chem* 138, 63  
 SCHAFFER, A J FOLKOFF C and BAYNE-JONES, S (1922) *Johns Hopk Hosp Bull.*, 33, 151  
 SCHLENG F and GINERICH W (1942) *J biol Chem*, 143, 295  
 SCHNEIDER, E (1934) *Z physiol Chem.*, 228, 231  
 SCHREIBER, R (1902) *Arch Hyg*, 41, 378  
 SCHULTZ, A S, ATKIN L and FREY, C N (1939) *J Amer Chem Soc*, 61, 1931  
 SCHEMAN, R L and FARRILL, M A (1941) *J infect Dis*, 69, 81  
 SCHUTZ, F and THEORELL, H (1938) *Biochem Z*, 295, 246  
 SERA, Y (1910) *Z Hyg Infekth.*, 66, 141, 162  
 SERGENT A. L. (1928) Les facteurs de croissances des microbes sur milieux artificiels. Paris.  
 SEVAG, M G SMOLENS J and LACKMAN, D B (1940) *J biol Chem*, 134, 523  
 SILVERMAN, M. and WERKMAN, C H (1939) *Fn ymolog a* 5, 380  
 SMILEY A L, NIVEN C F and SHERMAN, J M. (1943) *J Bact* 45, 445  
 SMITH, T (1897) *J exp Med.*, 2, 543  
 SNELL, F F and PETERSON W H (1940) *J Bact*, 39, 273  
 SNELL, F F, STRONG F M and PETERSON, W H (1939) *J Bact* 38, 293  
 SOBOTKA H and HOLZMAN, M (1934) *Biochem. J*, 28 734  
 SÖHNGEN, N L (1911) *Jber Fortschr Chem.*, 41, 788 789  
 SOMMARUGA E VON (1894) *Z Hyg Infekth.*, 18, 441  
 SÖBENSEN, S P L (1909) *Biochem Z*, 21, 131  
 SPERRY, J A and RETTGER, L F (1915) *J biol Chem*, 20, 415  
 STAHL, W H and HAMANN E E. (1941) *Mich ogr Exp Sta Bull.*, No. 177 p 17  
 STEPHENSON, M (1928) *Biochem J*, 22, 605 (1939) *Bacterial Metabolism* 2nd ed., Longmans Green & Co, London  
 STEPHENSON, M. and STICKLAND L H (1931) *Biochem J*, 25, 200, 215, (1932) *Ibid*, 26, 712  
 STEPHENSON, M. and WHETHAM, M. S (1922) *Proc roy Soc, B* 93, 262, (1923) *Ibid*, 95, 200, (1924) *Biochem J*, 18, 498.  
 STERN, A G (1936) *J biol. Chem* 114 473  
 STEVENS, F A. and WEST, R. (1922) *J exp Med*, 35, 823

- STICKLAND, L. H. (1929) *Biochem. J.*, 23, 1187, (1931) *Ibid.*, 25, 1543, (1934) *Ibid.*, 28, 1746, (1935) *Ibid.*, 29, 285
- STILL, J. L. (1940) *Biochem. J.*, 34, 1374
- STOKES, J. L. GUNNESS, M. and FOSTER, J. W. (1944) *J. Bact.* 47, 303
- STOKINGER, H. E., ACKERMAN, H. and CARPENTER, C. M. (1944) *J. Bact.*, 47, 123
- STOKLASA, J. (1908) *Zbl. Bakt., Hte. Abt.* 21, 620
- STOKSTAD, E. L. R. (1941) *J. biol. Chem.*, 139, 475
- STRAUB, F. B. CORRAN, H. S. and GREEN, D. E. (1939) *Nature Lond.* 143, 119
- STUMPF, P. A. and GREEN, D. E. (1944) *J. biol. Chem.* 153, 387
- TANURA, J. T. and GIBBY, I. W. (1943) *J. Bact.*, 45, 361
- TANURA, S. (1913a) *Z. physiol. Chem.*, 87, 85 (1913b) *Ibid.*, 88, 190 (1914) *Ibid.* 89, 289
- TARR, H. L. A. (1933) *Biochem. J.*, 27, 136
- TASMAN, A. and BRANDWIJK, A. C. (1938) *J. infect. Dis.* 63, 10
- TATUM, E. L., WOOD, H. G. and PETERSON, W. H. (1936) *Biochem. J.* 30, 1893
- THOMPSON, R. H. S. and DUBOS, R. J. (1939) *J. biol. Chem.*, 125, 65
- TIEKA, J. (1935) *Biochem. Z.*, 279, 264
- TEAUGOTT, R. (1893) *Z. Hyg. Infekthkr.*, 14, 427
- TRUSSELL, R. E. and WEED, L. A. (1937) *J. Bact.*, 33, 381
- VALLEY, G. (1928) *Quart. Rev. Biol.* 3, 209
- VALLEY, G. and RETTGER, L. F. (1927) *J. Bact.* 14, 101
- VIGNEAUD, V. DU, DITMER, K., HAGUE, I. and LONG, B. (1942) *Science* 96, 186
- VIGNEAUD, V. DU, HOFMAN, K. and MELVILLE, D. B. (1942) *J. Amer. chem. Soc.* 64, 188
- VIRTANEN, A. I. (1924) *Z. physiol. Chem.*, 138, 136 (1925) *Ibid.*, 143, 71
- VIRTANEN, A. I. and KARSTEN, H. (1931) *Acta chem. fenn.*, B, 7, 17
- WALKER, H. H. (1932) *Science* 76, 602
- WALLE, N. VAN DER. (1927) *Zbl. Bakt., Hte. Abt.*, 70, 369
- WARRBURG, O. (1925a) *Ber. disch. chem. Ges.*, 58, 1001, (1925b) *Science* 61, 575
- WARRBURG, O. and CHRISTIAN, W. (1933) *Biochem. Z.* 260, 499
- WEIL, A. J. and BLACK, J. (1944) *Proc. Soc. exp. Biol. N. Y.*, 55, 24
- WEIL, L. and KOCHOLATY, W. (1937) *Biochem. J.*, 31, 1255
- WELLS, H. G. and CORPER, H. J. (1912) *J. infect. Dis.* 11, 393
- WERKMAN, C. H. (1939) *Bact. Rev.*, 3, 187
- WERKMAN, C. H. and WOOD, H. G. (1942) *Advances Enzymology*, 2, 135
- WIELAND, (1913) *Ber. disch. chem. Ges.*, 46, 3327, (1921) *Ibid.*, 54, 2353, (1922) *Ergebn. Physiol.* 20, 477
- WILDIERS, E. (1901) *La Cellule* 18, 311
- WILLIAMS, C. H., BLOOR, W. R. and SANDHOLZER, L. A. (1939) *J. Bact.* 37, 301
- WILSON, G. S. (1930) *J. Hyg., Camb.*, 30, 433
- WINOGRADSKY, S. (1889) *Ann. Inst. Pasteur*, 3, 49, (1890a) *Ibid.* 4, 213 (1890b) *Ibid.* 4, 257, (1890c) *Ibid.* 4, 760
- WOLF, C. G. L. (1919a) *J. Path. Bact.*, 22, 270 (1919b) *Ibid.* 22, 289
- WOLF, C. G. L. and HARRIS, J. E. G. (1918) *J. Path. Bact.*, 22, 1
- WOOD, H. G. and WERKMAN, C. H. (1936) *Biochem. J.* 30, 48, (1938) *Ibid.*, 32, 1763 (1940) *Ibid.* 34, 7, 129
- WOOD, H. G., WERKMAN, C. H., HEMINGWAY, A. and NIER, A. O. (1940) *J. biol. Chem.*, 135, 789
- WOODS, D. D. (1935) *Biochem. J.*, 29, 640, 649, (1936) *Ibid.*, 30, 1934
- WOODS, D. D. and CLIFTON, C. E. (1937) *Biochem. J.* 31, 1774, (1938) *Ibid.*, 32, 345
- WOODS, D. D. and TRIM, A. R. (1942) *Biochem. J.*, 36, 501
- WOOLF, B. (1931) *Biochem. J.*, 25, 342
- WOOLLEY, D. W. and HUTCHINGS, B. L. (1940) *J. Bact.* 39, 287
- WRIGHT, H. D. (1936) *J. Path. Bact.*, 43, 487 (1937) *Ibid.* 45, 117
- WYON, G. A. and McLEOD, J. W. (1923) *J. Hyg., Camb.*, 21, 378
- YUDKIN, J. (1932) *Biochem. J.*, 26, 1859, (1933) *Ibid.*, 27, 1849, (1934) *Ibid.*, 28, 1463
- ZOBELL, C. E. (1943) *J. Bact.*, 46, 39

## CHAPTER 4

### THE GROWTH AND DEATH OF BACTERIA

LEST the title of this chapter should prove misleading, it may be pointed out that we are not here concerned with the broader aspects of growth, such as the temperature and food requirements of bacteria, or their metabolism and respiration. These have already been briefly dealt with in Chapter 3. In the present chapter we restrict ourselves to what we may describe as the dynamic aspects of growth—a study that deals essentially with the rate of change in a bacterial population.

#### Technique of Counting Bacteria

Bacteria may be counted in such a way as to obtain an estimate either of the total number of organisms alive and dead, or of the number of living organisms only. The first we shall refer to as the *Total Count*, the second as the *Viable Count*. Each method is suited to various purposes, and the choice of which to employ must depend on the type of information desired. For many purposes, such as calculation of the generation time of bacteria, and the quantitative study of bacterial metabolism, both methods should be used in conjunction.

The general principles underlying the counting of bacteria may be briefly mentioned. [For further details the reader is referred to textbooks on practical bacteriology, and for a critical review of the different methods that may be employed, with the main sources of error involved, to articles by Wilson (1922) and Wilson *et al.* (1935).]

#### (1) Total Count.

(a) *Direct Counting under the Microscope of a Stained Preparation on a Slide*—First described by Eberle (1896), this method has been used fairly extensively, and forms the basis of the Breed (1911) method for counting bacteria in milk. A drop of known volume is spread over a known area on a slide, dried, fixed, stained, and examined under a microscope. The organisms in a given number of fields are counted, and knowing the area of a given field with a particular combination of objective, tube length and ocular—this can be obtained by means of slide and eyepiece micrometers—it is possible to calculate the total number of organisms present in the original suspension. This method, though valuable for certain purposes, is open to a number of technical objections, one of the most important of which is that not all organisms—particularly when dead—stain sufficiently deeply to be visible under ordinary illuminating conditions. The method, therefore, affords an estimate of the number of stainable bacteria, not necessarily of the total bacteria.

(b) *Wright's Method* (1902)—A known volume of the bacterial suspension is mixed with a known volume of normal human blood. A smear preparation is made on a slide, dried, fixed, stained, examined under the microscope, and the number of bacteria and red cells in a given number of fields is counted. Since, in the blood of the normal adult male there are about 5.5 million red cells per c.mm., and since the numerical relationship of

of these a number of tubes of medium, usually liquid, are inoculated with suitable unit quantities. The tubes are incubated and from the number in which growth occurs, the probable number of bacteria in the original suspension may be calculated from formulae, such as those worked out by McCrady (1915) Stein (1917) Greenwood and Yule (1917) Halvorson and Ziegler (1933a, b) and Gordon and ZoBell (1938). (For useful tables see McCrady 1918 Hoskins 1934.) This method or one of its numerous modifications, is frequently adopted when an approximate estimate of the numbers of living bacteria in a suspension is required. It has the great advantage of being applicable to organisms that cannot be counted by the ordinary plating method. It is used for example, in the determination of coliform organisms in water, because no satisfactory plating medium has yet been devised for differentiating coliform bacilli in mixed culture from other bacteria though several suitable liquid media are available. It is frequently used in filtrable virus work, for this purpose quantities of the dilutions are inoculated into a susceptible animal generally by the intradermal route and the occurrence of a specific skin reaction is regarded as evidence of the presence of the organism in the corresponding dilution. As Halvorson and Ziegler (1933a, b) have pointed out the method is subject to a very large experimental error depending mainly on the numbers of tubes seeded from the different dilutions. Even with 40 tubes to each dilution the count is liable to vary between about 38 per cent. below and 47 per cent. above the true count, while with only 5 tubes to each dilution the corresponding figures are -70 and +260 per cent. The method is not therefore suitable for exact bacterial enumeration.

(b) *The Plating Method*—This is generally performed by a modification of Koch's original plating method. It consists essentially in preliminary dilution, if necessary, of the suspension, the plating out of unit quantities of suitable dilutions into a suitable solid medium and the counting of the number of colonies that develop after incubation. The average number of colonies per plate multiplied by the reciprocal of the dilution affords an estimate of the number of living organisms in the original suspension. Instead of plates, roll tubes may be used. Provided that the bacteria are homogeneously distributed in the suspension, that not more than one species of organism is present and that attention is paid to a large number of technical points, accurate counts may be obtained by this method. Departure from these provisos may often entail, however, experimental errors of considerable magnitude. (For sources and measurement of errors see Wilson *et al.* 1935 Jennison and Wadsworth 1940.)

Miles and Misra (1938) have described a method of plate counting in which the bacterial suspension instead of being mixed with the melted agar is deposited in the form of drops on the surface of the solid medium and the count estimated from the number of colonies that develop.

### The Growth Curve

If a given bacterium is seeded into a liquid medium of suitable composition, and incubated at a suitable temperature it will be found that its growth will follow a definite course. This course is most conveniently represented in graphical form (Fig. 15) the logarithms of the numbers of bacteria along the ordinates being plotted against the time in hours along the abscissae.

The growth curve may be arbitrarily divided into four phases. (1) The lag phase, *a* to *b*, lasting for a few hours, during which multiplication is slow. In the early part of this phase there may be no apparent growth, in fact many of the organisms may die, so that there is an actual diminution in their numbers. Within a short time however, growth becomes apparent, and gradually increases in pace till the beginning of the next phase. (2) The logarithmic phase, *b* to *c*, in which regular division of the organisms occurs at maximum speed. Since their increase is in geometric progression, it follows that when the logarithms of their numbers are plotted against the time in hours they fall on an ascending straight line. (3) The

stationary phase, *c* to *d*, during which the organisms cease to multiply at maximal rate, so that their increase in number becomes less and less, till ultimately it ceases, the number present in a unit volume remains approximately constant for an appreciable length of time. During this phase the number of freshly formed bacteria roughly counterbalances the number of those that are dying. (4) The phase of decline, *d* to *e*, during which the organisms gradually diminish in number till finally the culture becomes sterile.

So far we have been considering the viable bacteria only. If however, a count is performed of the total number of organisms alive and dead in the culture a different curve is obtained. It will be seen that this curve runs more or less parallel to the curve of the viable bacilli till the period of decline sets in, after which the two diverge, the total curve remaining practically stationary or rising very slightly. It will be noticed, moreover, that the total curve is throughout

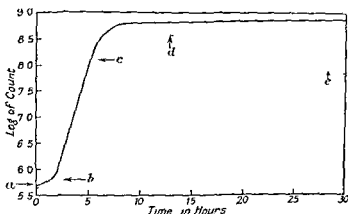


FIG. 15

Continuous line = Total number of bacteria alive or dead  
Interrupted line = Number of living or viable bacteria

somewhat higher than the viable curve. The probable explanation of this will be given later.

We must now consider each of the phases in detail.

### The Lag Phase

It is important at the outset of any discussion on growth to define our terms. Failure to do this has been responsible for much confusion in the past. Growth may occur either in size or in numbers. The individual cell may get progressively larger, or it may divide and give rise to two daughter cells. Growth in size may be referred to as cell enlargement, growth in numbers as cell multiplication.

The observations of early workers in this field (Muller 1895, Rahn 1906, Lane Claydon 1909, Coplans 1910, Penfold 1914, Ledingham and Penfold 1911, Chesney 1916, Slator 1917, Buchanan 1918) were made almost exclusively on cell multiplication. They found that the lag phase, during which little or no multiplication occurs, was affected by a number of factors such as the size and age of the inoculum, the frequency of transplantation of the parent culture, the nature of the organism, the composition of the medium, and the temperature at which the culture is incubated. Generally speaking the lag phase tends (a) to be long when a small

inoculum is used when the inoculum is taken from an old culture when the medium is unsuitable and when the culture is incubated at a temperature unfavourable to maximal growth and (b) to be short when the opposite conditions obtain. The most striking factor of all however is the stage of growth of the parent culture from which the inoculum is taken. Organisms from a culture in the lag, the stationary or the decline phase of growth fail to multiply for some time when introduced into a fresh medium whereas organisms from a culture in the logarithmic phase show no lag but continue to multiply at the same rate as in the parent culture (Muller 1935). During the logarithmic phase the organisms are known to be multiplying at maximal rapidity and since this phase lasts for not more than a few hours it follows that the organisms must all be comparatively young. By the term young we mean an organism that has been generated within a comparatively recent time from a few minutes to about an hour. The suggestion therefore is that young organisms inoculated into a fresh medium multiply without lag and that old organisms exhibit a lag phase before beginning to grow.

Numerous explanations were put forward to explain the phenomena of lag. It was suggested by Rahn (1906) that before multiplication could occur some essential substance or bios had to be excreted by the organisms into the medium. Penfold (1914) modified this view by supposing that certain bodies necessary for the synthesis of protoplasm had to accumulate within the bacteria themselves before multiplication became possible. Chesney (1916) regarded the phenomenon of lag as an expression of injury which the bacterial cell had sustained in its previous environment. Ledingham and Penfold (1914) postulated an inherent difference in the multiplication rate of the individual cells in the inoculum and suggested that during the lag phase a selection of the more rapidly multiplying cells was occurring the result of which did not become manifest till the beginning of the logarithmic stage. Experimental evidence does not support this view (Kelly and Rahn 1939, Topley and Wilson 1936).

These explanations need not be discussed further because more recent work has brought to light one supremely important fact of which the earlier observers were ignorant and which goes a long way towards explaining the difference in behaviour of young and old organisms. Briefly stated the lag phase is not a phase of rest as had been previously supposed but a phase of intense growth activity during which cell enlargement but little or no cell division occurs. The evidence for this statement must now be considered in some detail.

#### Evidence of Growth without Multiplication during the Lag Phase

(1) *Increase in Size*—If a small number of organisms from a 24-hours culture of *Bact. coli* for example are seeded into an agar medium which is then spread between a slide and a cover glass and observed on a warm stage by dark-ground illumination the first change observed is not division but a gradual and progressive enlargement affecting a certain proportion of the organisms. This enlargement becomes visible shortly after the preparation has been put up and continues till the organisms reach a certain size when they finally divide. The rate at which the enlargement occurs varies with the different organisms in the preparation. The time elapsing between the incubation of such a preparation and the first division varies from about  $1\frac{1}{2}$  to 3 hours but after the primary division has occurred subsequent generations may be produced at the rate of about one in every 30 to 60 minutes. Examination of such a preparation at the end of about five hours will reveal a small proportion of cells that

same time 1 ml portions of the culture were transferred to 100 ml of 5 per cent sodium chloride solution, the mixture was allowed to stand for 1 hour at 20° C., after which a count was performed on it, the numbers being calculated in terms of the 1 ml of added broth culture. The results are given in Table 1

TABLE 1

Time after Inoculation.	Original Culture	After 1 hour in 5 per cent. NaCl	Mortality per cent
0 hours	96,000	82,500	14.06
1 hour	80,500	60,500	24.82
1½ hours	90,500	41,000	54.70
2 hours	143,000	33,000	76.93
2½ hours	255,000	16,500	93.54

This table shows that the young bacilli, formed after the culture has entered on the logarithmic phase of growth, are very much more susceptible to the action of 5 per cent. NaCl solution than are the old bacilli inoculated at the start—a mortality of 93.54 per cent. as against 14.06 per cent. In addition it will be noticed that during the lag phase, lasting from 0 to 1½ hours, though there is no increase in the number of bacteria there is a progressive increase in their susceptibility to salt solution. Sherman and Albus interpret this as indicating that, during the lag phase, the old bacteria which have been inoculated are undergoing a process of rejuvenescence, which fits them for reproduction. Numerous instances are quoted from other fields of biology, such as the rejuvenescence of *Paramacium*, the growth curves of colonies of fruit flies, etc., that may be regarded as exemplifying the same biological law of growth. Prior to active growth, there is a latent phase of preparation, during which the old cells are being rejuvenated, and fitted for reproduction.

Sherman and Naylor (1942) made the interesting observation that cells of *Bact. coli* taken during the logarithmic phase cooled gradually to 1° C and stored at this temperature retained the characteristics of young bacteria—namely, ability to multiply without lag when transferred to fresh medium at 37° C and greater susceptibility to chemical agents (in this case cold shock)—for as long as 36 days. Cells of *Str. lactis* on the other hand rapidly acquired at 1° C the characters of senescence. What this difference was due to it is impossible to say, but the observations of Sherman and Naylor may be taken in conjunction with those of Anderson and Meanwell (1936), who found that a thermophilic milk streptococcus (1 = one capable of resisting heat at 60° C. for 30 minutes) with which they were working became less rather than more susceptible to heat during the lag phase. Generalizations, therefore based on the behaviour of coliform bacilli alone, must be accepted with reserve.

(4) *Diminution in the Electrophoretic Charge*—Working with *Bact. coli*, Moyer (1936) found that when the organisms were introduced into a fresh medium their rate of migration in a cataphoretic cell decreased during the first two hours, remained low during the third and fourth hours, and then rose towards the end of the logarithmic phase. He concluded that these changes were probably due to an alteration in the electro-chemical state of the bacterial surface during the growth cycle.

(5) *Decreased Susceptibility to Non-specific Agglutination*—Though few observations have so far been made the results recorded by Macgregor (1910) on agglutina



tion of meningococci by normal serum, Gillespie (1914) on the salt agglutination of pneumococci, and Sherman and Albus (1923) on the acid agglutination of *Bact. coli*, all suggest that young cells are less susceptible than old to non specific agglutination. This presumably is due to an alteration in the electric charge or to some other change on the surface of the cells.

Summarizing the data yielded by these observations we see that during the lag phase of growth there is a progressive enlargement of the cells before division occurs, associated with a rapid increase in their metabolic activity and their susceptibility to heat and disinfectants. Before a cell can divide it must increase in size. The so-called lag phase of growth seems to be occupied mainly by the process of cell enlargement. Up to this point all recent workers are in agreement. On two subjects, however, there is some discrepancy of opinion. Firstly do all cells, when inoculated into a fresh medium, begin to grow at once? In other words is there no such thing as a true lag phase? It is a little difficult to answer the question, but the available evidence suggests fairly strongly that under certain conditions cells introduced into a fresh medium take some time to adjust themselves to the new conditions before they start to grow even in size. The duration of this "phase of adjustment" varies with the conditions. While it lasts it constitutes a true lag phase. If the organisms in the inoculum are vigorous and unharmed and the medium into which they are introduced is favourable cell enlargement may occur at once, and no true lag phase will be seen. If on the other hand they are suffering from the toxic effects of their previous environment or the medium into which they are inoculated has an unfavourable pH or salt content, then some time may elapse before cell enlargement begins.

Secondly, are we justified in referring to the orderly change that occurs in the enlargement and division of a bacterial cell as a life-cycle? Apart from mere difference of opinion on the advisability of using such a term the answer must depend on whether there is a real differentiation of morphology and function at different stages of growth. The observations of Henri (1928) in particular leave no doubt about the differences in size, structure and staining affinity of bacterial cells at different stages of growth but the evidence in regard to their metabolic activity is a little conflicting. Hershey (1939) for instance maintains that the metabolic activity is the same for cells at all stages of growth provided that it is expressed in terms of amount of bacterial protoplasm rather than in terms of individual cells. It is true that in absolute values a cell in the late lag or early logarithmic phase uses more oxygen and produces more  $\text{CO}_2$  than a cell in the stationary or decline phase but this according to Hershey, is merely due to a difference in size of the cells. The respiration of any unit amount of protoplasm is the same with cells at all stages of growth. Huntington and Winslow (1937) and Winslow and Walker (1939) however, do not agree with this view. They found that the rate of  $\text{CO}_2$  production per cubic micron of bacterial substance was much higher during the lag and early logarithmic phases than during the stationary phase. They believe, therefore that young protoplasm is characterized by a greater rate of metabolism and that the use of such a term as life-cycle for bacteria is justifiable. Further evidence is required to settle this important difference of fact.

If we accept the view of Winslow and Walker, we shall summarize as follows

On the ground (a) that newly generated bacilli in the logarithmic phase, when transferred to a fresh medium start multiplying without exhibiting any lag phase, (b) that bacilli from cultures in all other stages show a lag phase before they start

dividing, and (c) that during this lag phase the cells are undergoing a rapid metamorphosis, resembling that perhaps described by Child (1915) in the rejuvenescence of infusoria, flat worms, and marine alga, which renders them similar to the young actively dividing bacilli of the logarithmic phase. It may be concluded that the lag phase is essentially a period in which the protoplasm of the old, but still viable, bacteria in the inoculum is acquiring the characteristics of young protoplasm. The lag phase appears to be a phase of rejuvenescence.

### Logarithmic Phase

The logarithmic phase is that period during which regular and maximum multiplication is occurring. Increase in the number of organisms is by geometrical progression, so that, if the logarithms of their numbers are plotted against time, they will fall along an ascending straight line. If counts are made at intervals, it is easy to calculate the number of generations during the phase, and the length of each generation.

Thus if  $a$  = number of organisms at the beginning of a given time

and  $b$  = " " " " " end " " " "

then at the end of the first generation,

$$b = a \times 2$$

at the end of the second generation,

$$b = a \times 2 \times 2$$

at the end of  $n$  generations,

$$b = a \times 2^n$$

To find the value of  $n$ , i.e. the number of generations, we may write

$$\log b = \log a + n \log 2$$

$$\text{or} \quad n = \frac{\log b - \log a}{\log 2} \quad . \quad . \quad . \quad (1)$$

Further, if there have been  $n$  generations in time  $T$ , the generation time  $G$  can be calculated from the formula

$$G = \frac{T}{n} \quad . \quad . \quad . \quad (2)$$

It has generally been held that during the logarithmic phase all the bacteria are alive and all are actively dividing. Thus, as represented in formula (1), 2 bacteria give rise to 4, 4 to 8, 8 to 16, and so on. If this assumption were correct, then all the organisms present during this phase should be viable, and the total count would be identical with the viable count.

This may be true when, as Kelly and Rahn (1932) have found, organisms are growing under optimal conditions and are followed for a few generations only. Observations, however, on ordinary broth cultures have shown that the total number of organisms generally exceeds the number of viable organisms, even during the logarithmic phase (Wilson 1922, 1926, Régner, David, and Kaplan 1932, Buce 1933-34, Jordan and Jacobs 1944).

The most probable explanation of this fact is that during the period of maximum growth some of the organisms that are generated fail to survive. If, for example, 80 per cent of the organisms produced during a given generation continued to live and divide, while 20 per cent died, then at the end of the logarithmic phase the total number of organisms alive and dead would exceed the number of living

The increase in the living organisms would still occur by geometrical progression and the resultant curve of plotting the logarithms against time would still fall on a straight line, the only difference would be that instead of the number of organisms being doubled in each generation their factor of increase or generation index, would be 1.6

Thus in Table 2 it is supposed that only 50 per cent of the organisms inoculated are viable. The ratio of viable to total organisms i.e.  $\frac{V}{T}$  increases with each generation. After about ten generations it rapidly approaches the value  $p - 1$  where  $p =$  the generation index. The value of  $p$  can therefore be obtained with fair accuracy towards the end of the logarithmic phase by simply calculating the viable total ratio and adding 1. In the example quoted it is approximately 1.6

TABLE 2

CALCULATIONS OF VIABLE AND TOTAL ORGANISMS WITH A 20 PER CENT DEATH RATE PER GENERATION

	No. of Viable Org. per ml	No. of Dead Org. per ml	Total No. of Org. per ml	Ratio Viable Total
At start	1 000	1 000	2 000	0.500
At end of 1st Gen.	1 600	1 400(1 000 + 400)	3 000	0.533
At end of 2nd Gen.	2 560	2 040(1 400 + 640)	4 600	0.557
At end of 3rd Gen.	4 096	3 064(2 040 + 1 024)	7 160	0.572
At end of 4th Gen.	6 554	4 706(3 064 + 1 642)	11 260	0.582

If the viable and total counts are known both at the start and at the end of  $x$  generations, then  $p$  can be ascertained at any time during the logarithmic phase since

$$\frac{V_x - V_0}{T_x - T_0} = p - 1 \quad (3)$$

or

$$p = \frac{V_x - V_0}{T_x - T_0} + 1 \quad (4)^1$$

where  $V_0$  and  $T_0 =$  number of viable and total organisms respectively at the start, and  $V_x$  and  $T_x =$  number of viable and total organisms respectively at the end of  $x$  generations

Experiments on the growth of *Salmonella typhimurium* in broth have shown that the average ratio of viable to total organisms at the end of the logarithmic phase is about 0.8. This corresponds to a death rate of about 10 per cent per generation and to a generation index of 1.8. Hence in calculating the generation time formula (1) has to be changed to

$$n = \frac{\log b - \log a}{\log 1.8}$$

Employing this formula the number of generations is greater than by the old formula and the generation time necessarily shorter.

It must be understood that the factor of increase to be used in the formula may vary with each experiment and can only be ascertained by the performance of a total and a viable count on each culture.

<sup>1</sup> For this formula we are very much indebted to Dr. H. G. W. Hoare.

TABLE 3  
GROWTH OF *Salmonella typhimurium* IN BROTH (Wilson 1922)

Time in Minutes.	Viable Count per ml.	Logarithms of Viable Count	$N_1 - N_2$ $N_2 - N_3$ etc.	$t_1 - t_0$ $t_2 - t_1$ etc.	$N_1 - N_2 - t_1 - t_0$ $N_2 - N_3 - t_2 - t_1$ etc.	M33-time Interval.
0	Not counted	—	—	—	—	—
40	828,800	5.9184	—	—	—	—
80	1,539,000	6.1872	710,200	40	17,755	60
120	4,363,000	6.6398	2,824,000	40	70,600	100
160	12,152,000	7.0845	7,787,000	40	194,675	140
200	32,490,000	7.5117	20,340,000	40	508,500	180
240	81,640,000	7.9119	49,150,000	40	1,228,750	220
280	155,900,000	8.1928	74,260,000	40	1,858,500	260
320	271,700,000	8.4341	115,800,000	40	2,895,000	300
370	334,300,000	8.5241	62,600,000	50	1,252,000	345
440	351,400,000	8.5458	17,100,000	70	244,286	405

Instead of plotting the logarithms of the bacterial numbers against time, Lemon (1933) has pointed out that interesting information may be obtained by estimating the actual rate of growth in unit intervals of time.

If for each period of time  $T$  the count is  $N_0, N_1, N_2 \dots$  etc. at times  $t_0, t_1, t_2 \dots$ , the following formula may be used—

$$\frac{(N_1 - N_0)}{(t_1 - t_0)_{(T_0)}} \text{ and } \frac{(N_2 - N_1)}{(t_2 - t_1)_{(T_1 - T_0) + T_1}} \text{ and } \frac{(N_3 - N_2)}{(t_3 - t_2)_{(T_2 - T_1) + T_2}} \text{ etc.}$$

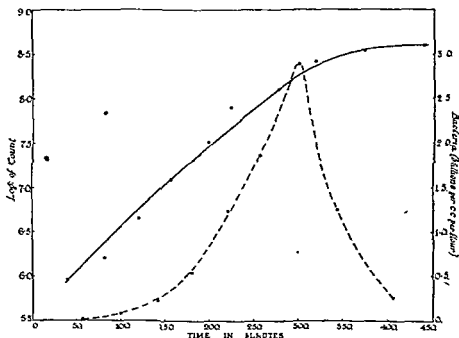


FIG 16—GROWTH OF *Salmonella typhimurium* IN BROTH, PLOTTED ACCORDING TO LEMON'S FORMULA

Continuous line = Numbers plotted logarithmically against time.

Interrupted line = Rate of arithmetic increase plotted against time.

The results are plotted on a graph whose abscissae represent  $T$  and whose ordinates indicate the rate of propagation. The numerator and the first term of the denominator in the formula represent the arithmetic increase in the number of organisms in unit time. The second term or "suffix" of the denominator is not part of the divisor but indicates the mid point of the period during which the actual increase has occurred. The use of the formula is exemplified in Table 3 and Fig. 16.

It will be observed that, while the curve of the logarithms indicates that a steady rate of multiplication is occurring during the 1-5 hour period, the curve given by Lemon's

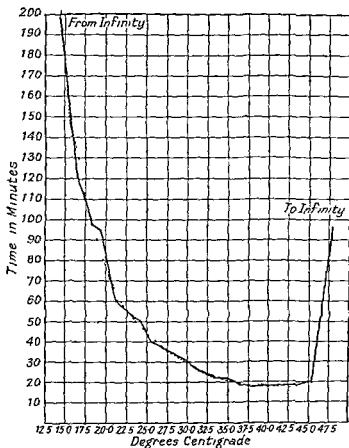


FIG. 17—THE GROWTH RATE OF *Bact. coli* AT DIFFERENT TEMPERATURES\*

Continuous line = Curve plotted from actual observations

Interrupted line = Smoothed curve  
(After Barber)

formula shows that the actual rate of arithmetic increase rises rapidly to reach a peak at 5 hours, after which it falls steeply. The reason for the rapid rise is, not that the organisms are dividing more rapidly at the end than at the beginning of the logarithmic phase, but that, owing to the fact that the actual numbers of bacteria are constantly increasing throughout this phase, the progeny of any given generation expressed arithmetically must be greater than that of any previous generation. Lemon's method of calculating the growth rate is likely to be of advantage in physiological and biochemical problems when it is desired to study the rate of change in a chemical substrate in relation to the absolute numbers of organisms produced.

The generation time during the logarithmic phase is influenced by several factors

(1) *Temperature of Incubation of Culture*—Working with *Bact coli*, Chick (1912) found that between 20° and 40° C, each rise of 1° C increased the rate of growth 1.072 times, the temperature coefficient for every rise of 10° C was 1.072<sup>10</sup>, i.e. 2.01. This agrees closely with Lane-Clayton's (1909) figure of 2.2. Barber (1908), who made an exhaustive study of the rate of growth of *Bact coli*, constructed the curve shown in Fig. 17. In this it will be seen that at a temperature of 15° C the generation time is 180, at 25° C 44', at 35° C 22' and at 40° C 17'. The curve corresponds fairly closely to the quarter of an ellipse. Further, as the increase is geometrical, by plotting the logarithms of the generation times we shall find that the points fall on a descending straight line. The maximum rate of growth of *Bact coli* occurs at about 37° C, but between 37° and 46° C there

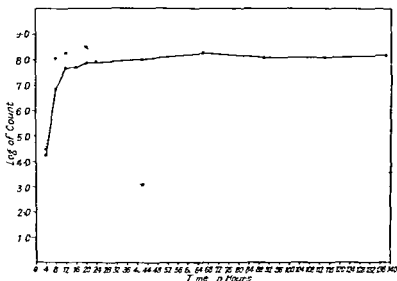


FIG. 18

A. Continuous line = Viable count of *Salm typhi-murium* in peptone water

B. Interrupted line = Viable count of *Salm typhi-murium* in peptone water + 2 per cent. glucose (After Heap and Cadness)

is little change in the generation time. For *Salm typhi* Müller (1895) found the maximum rate of growth to be between 37° and 40.4° C, at a temperature of 44.5° C the bacilli died rapidly.

(2) *Nature of Medium*—Penfold and Norris (1912) found that by increasing the concentration of peptone in a peptone water medium from 0.125 per cent to 1 per cent the generation time of *Salm typhi* was reduced from almost infinity to 40. They likewise found that the addition of glucose to the medium further reduced the generation time. Other workers have also commented on the beneficial effect of glucose, thus Heap and Cadness (1924) found that the addition of 2 per cent glucose to a 3 per cent peptone water medium greatly increased the growth of *Salm typhi murium* (Fig. 18).

(3) *Nature of Organism*—Some organisms appear to grow more rapidly than others. Mason (1935) has compiled from the literature the generation times of

after which a progressive fall occurred. It must be pointed out, however, that this might equally well be explained by assuming that the type of metabolism had changed during a period of constant growth.

Care must, of course, be exercised in drawing any conclusions on the rate of multiplication from the rate of growth, since it has already been pointed out that, owing to variations in cell size, increase in bacterial numbers may give a very fallacious idea of the true rate of growth and the converse is equally true. Further observations at frequent intervals on the numerical increase occurring during the logarithmic phase made with due attention to multiple small factors that may influence the result are required before concluding that the rate of cell-division during this phase is discontinuous.

### Stationary Phase

After multiplying at a maximum rate for a variable length of time during the logarithmic phase the organisms become less active, and divide less frequently, till finally their numbers remain practically constant. What is responsible for this decrease in the reproduction rate? The natural suggestion is that it results from an exhaustion of the food supply. Against this view, however, is the fact that if a culture that has reached the decline phase is sterilized by boiling, and then reinoculated with the same organism, growth occurs in the usual way, though the actual numbers attained may not be so great as in the primary culture (Graham Smith 1921). The same result is obtained if the culture is sterilized by filtration instead of by boiling, provided that the organism has not produced some volatile substance which would be removed during boiling as in the special case of the pneumococcus. Penfold (1914) found that if a 24 hours' culture of *Bact. coli* was centrifuged and the supernatant fluid was incubated at 37° C, fresh growth took place. It is probable, therefore, that some other factor than exhaustion of the food supply is responsible for the cessation of maximal growth.

We have already mentioned Chesney's hypothesis that toxic substances are produced during the phase of multiplication and that these so injure the bacteria as to delay their subsequent division when introduced into a fresh medium. Although this explanation may hold good for the pneumococcus which produces  $H_2O_2$  in considerable amount, it does not seem to be of more general applicability. We may similarly regard as special cases those instances in which the reaction of the medium is rendered acid during growth owing to the inclusion of a fermentable carbohydrate. In such cases neutralization of the acid by the addition of alkali often suffices to enable growth to occur (Kojima 1923).

Working with the fruit fly *Drosophila melanogaster* Pearl and Parker (1922) found that the effective reproduction rate decreased as the population density became greater.

Following on these observations Baul (1929) carried out a number of experiments on different bacteria from the results of which he concluded that in any fluid culture there was a limiting population density that could not be exceeded. This he referred to as the M-concentration. A few of his findings may be briefly recorded.

(1) Any given species of bacterium reaches in a fluid medium a particular and constant M-concentration of living organisms the value of which differs with different species.

(2) With a large inoculum the M-concentration is reached rapidly with a small inoculum more slowly

(3) Living bacteria introduced into fresh broth in M-concentration are unable to multiply

(4) Living bacteria introduced into fresh broth in a concentration greater than M die off until the M-concentration is reached

(5) If a culture that has reached its M-concentration is centrifuged and then re-incubated fresh growth will occur in the clear supernatant fluid till the M-concentration is reached. If however after centrifuging the deposit is shaken up so that the organisms are distributed once more throughout the medium no growth will occur. This seems to show that renewed growth is dependent not on the total number of bacteria in a given volume of medium but on their mode of distribution within the medium.

(6) If a culture that has reached its M-concentration is heated to  $50^{\circ}\text{C}$  for a time sufficient to destroy the majority but not all of the living bacteria and is then re-incubated fresh growth will occur till the M-concentration is again reached. This seems to indicate that heat killed bacteria do not appreciably interfere with the biological space available

(7) Ordinary meat broth can be diluted 20 times or more without affecting the level of the M-concentration showing that it is not due to exhaustion of the medium that growth ceases. In the diluted broth the turbidity i.e. the total count is much less but the number of living bacteria i.e. the M-concentration is the same as in the undiluted broth

(8) The addition of an enriching substance such as glucose to the broth increases the total number of bacteria produced but the M-concentration of viable organisms remains unchanged

(9) Growth does not cease with the attainment of the M-concentration fresh organisms continue to be produced but an equivalent number die off so that the M value remains constant

Many of these findings were confirmed by Fukuda (1929). This worker however pointed out that some of them particularly Nos 5 and 8 held true only with certain organisms. He found moreover that if broth cultures of *Ps. pyocyanea* were sterilized by heat and re-inoculated with fresh organisms growth occurred till the original M-concentration was reached. This experiment could be repeated two or three times on the same culture though with *Salm. gallinarum* only one quarter of the M-concentration was reached after the first heating.

von Wikullil (1932) brought further evidence in support of Bail's hypothesis. He found that if two organisms A and B each having the same M-concentration of 1 600 million per ml were inoculated simultaneously in the same numbers into a tube of broth the final M-concentration of the mixed culture was still only 1 600 million per ml organism A constituting 800 million and organism B 800 million. The total physical space available had now to be shared between the two organisms. If two organisms A and C having respective M-concentrations of 1 600 million and 300 million per ml were inoculated simultaneously into the same tube of broth the final M-concentration was again 1 600 million per ml but this time the whole viable population consisted of organism A. The apparent explanation was that A grew more rapidly than C and so monopolized the available space.

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after which a progressive fall occurred. It must be pointed out, however, that this might equally well be explained by assuming that the type of metabolism had changed during a period of constant growth.

Care must, of course, be exercised in drawing any conclusions on the rate of multiplication from the rate of growth, since it has already been pointed out that, owing to variations in cell size, increase in bacterial numbers may give a very fallacious idea of the true rate of growth, and the converse is equally true. Further observations at frequent intervals on the numerical increase occurring during the logarithmic phase made with due attention to multiple small factors that may influence the result, are required before concluding that the rate of cell-division during this phase is discontinuous.

### Stationary Phase

After multiplying at a maximum rate for a variable length of time during the logarithmic phase, the organisms become less active, and divide less frequently, till finally their numbers remain practically constant. What is responsible for this decrease in the reproduction rate? The natural suggestion is that it results from an exhaustion of the food supply. Against this view, however, is the fact that if a culture that has reached the decline phase is sterilized by boiling, and then reinoculated with the same organism, growth occurs in the usual way, though the actual numbers attained may not be so great as in the primary culture (Graham Smith 1921). The same result is obtained if the culture is sterilized by filtration instead of by boiling, provided that the organism has not produced some volatile substance which would be removed during boiling, as in the special case of the pneumococcus. Penfold (1914) found that if a 24 hours' culture of *Bact. coli* was centrifuged, and the supernatant fluid was incubated at 37° C, fresh growth took place. It is probable, therefore, that some other factor than exhaustion of the food supply is responsible for the cessation of maximal growth.

We have already mentioned Chesney's hypothesis, that toxic substances are produced during the phase of multiplication, and that these so injure the bacteria as to delay their subsequent division when introduced into a fresh medium. Although this explanation may hold good for the pneumococcus, which produces  $H_2O_2$  in considerable amount, it does not seem to be of more general applicability. We may similarly regard as special cases those instances in which the reaction of the medium is rendered acid during growth owing to the inclusion of a fermentable carbohydrate. In such cases neutralization of the acid by the addition of alkali often suffices to enable growth to occur (Kojima 1923).

Working with the fruit fly *Drosophila melanogaster* Pearl and Parker (1922) found that the effective reproduction rate decreased as the population density became greater.

Following on these observations Bail (1929) carried out a number of experiments on different bacteria from the results of which he concluded that in any fluid culture there was a limiting population density that could not be exceeded. This he referred to as the M-concentration. A few of his findings may be briefly recorded.

(1) Any given species of bacterium reaches in a fluid medium a particular and constant M-concentration of living organisms, the value of which differs with different species.

(2) With a large inoculum the M-concentration is reached rapidly, with a small inoculum more slowly

(3) Living bacteria introduced into fresh broth in M-concentration are unable to multiply

(4) Living bacteria introduced into fresh broth in a concentration greater than M die off until the M-concentration is reached

(5) If a culture that has reached its M-concentration is centrifuged and then re-incubated, fresh growth will occur in the clear supernatant fluid till the M-concentration is reached. If, however, after centrifuging, the deposit is shaken up so that the organisms are distributed once more throughout the medium, no growth will occur. This seems to show that renewed growth is dependent not on the total number of bacteria in a given volume of medium, but on their mode of distribution within the medium.

(6) If a culture that has reached its M-concentration is heated to 55° C for a time sufficient to destroy the majority but not all of the living bacteria, and is then re incubated, fresh growth will occur till the M-concentration is again reached. This seems to indicate that heat killed bacteria do not appreciably interfere with the biological space available

(7) Ordinary meat broth can be diluted 25 times or more without affecting the level of the M-concentration, showing that it is not due to exhaustion of the medium that growth ceases. In the diluted broth the turbidity, i.e. the total count, is much less, but the number of living bacteria i.e. the M-concentration is the same as in the undiluted broth

(8) The addition of an enriching substance, such as glucose, to the broth increases the total number of bacteria produced, but the M-concentration of viable organisms remains unchanged

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Bail and his followers were more interested in making observations than in

explaining them. The problem still remains why it is that bacteria cease dividing at maximal speed when a certain population density is reached.

A suggestion which we put forward in 1936, and which has received some support from the observations of Hershey and Bronfenbrenner (1937) and of Rahn and Richardson (1942) is that the progressive retardation in growth may often be due to a deficiency of oxygen in the culture. There are certain observations that seem to support this view. We have already seen that the addition of 2 per cent glucose to a broth medium greatly increases the growth of *Salmonella typhi* *murium*. The mechanism by which this increase is brought about is unknown, but it is not unreasonable to suppose that the glucose provides a source of energy supply, which the organisms are able to utilize in the absence of readily available oxygen. In support of this we have found that the total number of organisms per ml. in a given culture can be regulated by altering the available amount of oxygen. Thus, in a casein broth culture of *Salmonella typhi* *murium* incubated anaerobically, the total count after 24 hours is about 500 million organisms; in a culture incubated aerobically it is about 2,000 million organisms, and in a culture through which pure oxygen is bubbled continuously, it is about 8,000 million organisms (Wilson, 1930). There seems no doubt that a liberal supply of oxygen enables growth to continue for some time after it has ceased in a culture incubated anaerobically or under ordinary aerobic conditions. The suggestion is that in cultures incubated aerobically, growth continues until the increasing density of the organisms renders it impossible for each individual organism to obtain sufficient oxygen to meet its requirements. These findings apply, of course, only to aerobic and facultatively anaerobic bacteria.

This explanation, it will be observed, does not fit all of Bails' facts, though it explains many of them. The main discrepancy is in the effect on the M-concentration of adding certain substrates. Bails found that the addition of glucose, for example, led to an increase in the total number of bacteria, while leaving the M-concentration of viable bacteria unaltered. Our experience, both with glucose and with increased aeration, entirely fails to support this finding. The more favourable the conditions for growth are, the higher is the viable population that the medium can support. It is true that in a medium containing a fermentable sugar the viable population may fall rapidly after having reached a height considerably greater than in the same medium without sugar (see Fig. 18), but this appears to be due to the disinfectant action of the acid produced, and not to any specific effect of bacterial density. That this is so is shown by the effect of continuous oxygen passage in the experiments quoted above, in enabling a broth medium to support four times its usual M population.

Broom (1929) and Gildemeister and Neustat (1930) have brought evidence to show that the amount of growth of many organisms appears to depend to a considerable extent on the presence in the medium of easily assimilable carbon compounds, such as glucose. The addition of these to a culture in which growth has ceased may rapidly enable a fresh crop of organisms to be produced. The apparent ability of some organisms to inhibit the growth of others in mixed cultures seems to be due not only to their more rapid growth but also to their more active fermentative power, which enables them to break down certain compounds more readily and so deprive the weaker organisms of their requisite nutritive materials.

The fact that mere aeration of a culture may enable fresh bacterial multiplication to occur suggests that there are at least two factors leading to cessation of growth. One is the exhaustion of easily assimilable food-stuffs, the other is

oxygen starvation. It seems not improbable that, once the simpler food stuffs have been broken down, the attack on the more complex compounds can be successful only in the presence of abundant oxygen.

It must not be thought that during the stationary phase all growth has ceased. In most cases multiplication continues for some time after the cessation of the logarithmic phase, but the rate of division progressively diminishes. The time taken to reach the maximum number varies. The temperature of incubation is important, thus, Graham Smith (1921), working with *Staphylococcus aureus* found that the maximum number of viable organisms was reached on the 2nd day at 37° C, on the 5th day at 27° C, and on the 8th day at 17° C. Much depends, too, on the nature of the medium.

Not only are fresh organisms being formed during the stationary phase, but large numbers are dying. Once the maximum number of organisms has been reached, and before the phase of decline, the multiplication rate just suffices to balance the death rate, so that the number of living bacteria remains constant.

### Phase of Decline

After lasting for a variable time—from about an hour to several days—the stationary phase passes gradually into the phase of decline. The numbers decrease slowly over a period of days, weeks, or months, till all the organisms are dead. With some organisms, such as the pneumococcus this phase is short, and the culture may be sterile in 2 to 3 days, with other organisms such as *Bact. coli* it may last for months. It is possible that growth is not entirely in abeyance during this stage, for if counts are performed at daily intervals, spasmodic rises are some

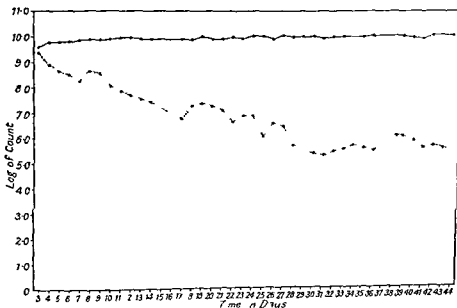


FIG. 19.—GROWTH OF *Salmonella typhimurium* IN BROTH, SHOWING THE PROLONGED DECLINE PHASE.

Continuous line = Total number of bacteria alive and dead.  
Interrupted line = Number of living or viable bacteria.

times noticed, indicative of the production of fresh organisms. Moreover, there is a gradual rise in the total count, pointing to the same conclusion (Fig. 19).

This is the curve obtained for a culture of a coliform or similar organism in a nutrient broth medium. In a medium containing a fermentable sugar, such as glucose broth, the curve is very different. Instead of descending slowly and irregularly, it passes rapidly down in an oblique straight line, till it reaches the abscissa (Fig. 18).

This difference is almost certainly due to the disinfectant action of the acid produced in the culture. The death of the organisms in Curve B is similar to the velocity of a unimolecular reaction, and can be represented by the formula

$$K = \frac{1}{t} \log \frac{n_1}{n_2}$$

where  $K$  is the velocity constant,  $t$  the interval of time between successive observations,  $n_1$  the number of living bacteria present at the beginning, and  $n_2$  the number present at the end of time  $t$ . This point will be more fully discussed in Chapter 5. For further information on the growth phases of bacteria, the reader is referred to a detailed consideration of this subject by Buchanan and Fulmer (1928).

#### Dormancy of Bacteria.

G. S. Burke (1923) inoculated a series of tubes of glucose peptic digest agar and of glucose peptic digest broth with *Cl. botulinum*, seeding one spore into each tube. She sealed the tubes to prevent desiccation, incubated them at 37° C., and noted each day in how many tubes growth had occurred. In the agar medium the majority developed in 10 days, but occasional spores continued to germinate up till the 92nd day. In the broth the majority developed in 14 days, but one or two germinated daily until the 33rd day, and thereafter at intervals till the 144th day.

V. Burke, Sprague and Barnes (1925) obtained similar results with *B. subtilis*, *B. megatherium*, and *Bact. coli*, thus showing that the phenomenon is not confined to anaerobic bacteria or the germination of spores. It is clear from these experiments that bacteria may be dormant for long periods without multiplying. Superficially, this resembles the phenomenon of lag, but it is possible that dormancy and lag are dependent on different factors. G. S. Burke (1923) draws attention to the similarity in behaviour of dormant spores and the seeds of certain of the higher plants. She suggests that in each instance the cause of the dormancy lies in the cell itself, and is connected with the degree of permeability of the wall.

It is this dormancy which appears to be responsible for the occasional failure of the intermittent process of sterilization. Probably owing to this, too, is the fact that a culture may be contaminated with an organism without signs of the contamination becoming evident till after three or four subcultures have been made.

**Mitogenetic Rays.**—The extensive work of Gurwitsch and his followers (for references see Bateman 1935) has suggested that certain plant and animal tissues under suitable conditions may emit so-called mitogenetic rays, which are able to stimulate growth in tissue cells placed in a position favourable for their absorption. According to some workers, the radiation is of the short ultra violet type, but attempts to confirm this by physical methods have not so far been successful. The rays, if they exist at all, have therefore to be detected by biological means.

So far as bacteria are concerned, the growth stimulating effects that have been reported have usually been well within the limits of the experimental error of the technique. While not denying the existence of this effect, we feel bound to adopt a strictly sceptical attitude until quantitative results that will stand the usual tests of statistical significance are forthcoming.

# REFERENCES

- ALPER, T and STERNE, M (1933) *J Hyg Camb*, 33, 497
- ANDERSON, E B and MEAKWILL, L J (1936) *J Dairy Res* 7, 182
- BAIL, O (1929) *Z Immunforsch* 60, 1
- BARBER, M A (1908) *J infect Dis*, 5, 379
- BATEMAN, J B (1935) *Biol Rev*, 10, 42
- BREED R S (1911) *Zbl Bakt, Hte Abt*, 30, 337
- BROOM, J C (1929) *Brit J exp Path*, 10, 71
- BUCHANAN, R E (1918) *J infect Dis*, 23, 109
- BUCHANAN, R E and FULMER E I (1928) *Physiology and Biochemistry of Bacteria* London
- BUICK W A (1933-34) *Zbl Bakt, Hte Abt* 89, 387
- BURKE, G S (1923) *J infect Dis*, 33, 274
- BURKE V, SPRAGUE A and BARNES L V (1925) *J infect Dis*, 36 555
- CHESNEY, A M (1916) *J exp Med* 24, 387
- CHICK, H (1912) *J Hyg, Camb*, 12, 414
- CHILD C M (1915) "Individuality in Organisms" Chicago
- CLARK, P F and RUEHL, W H (1919) *J Bact*, 4, 616
- COPLAND, M (1910) *J Path Bact*, 14, 1
- EMERLE, R (1896) *Zbl Bakt*, 19, 2
- FUKUDA, Y (1929) *Z Immunforsch*, 60, 88
- GILDEMEISTER, E and NEUSTAT, M (1930) *Zbl Bakt*, 133, 101
- GILLESPIE, L J (1914) *J exp Med* 19, 25
- GORDON, R D and ZOBELL, C E (1938) *Zbl Bakt Hte Abt* 99, 318
- GRAHAM SMITH, G S (1921) *J Hyg, Camb*, 19, 133
- GREENWOOD, M and YULE, G U (1917) *J Hyg, Camb* 16, 36
- HALVORSON, H O and ZIEGLER N R (1933a) *J Bact* 25, 101 (1933b) *Ibid* 26, 331 509
- HEAP, H and CADWELL B H E (1924) *J Hyg Camb*, 23, 77
- HELBERT, E (1904) *Dtsch Arch Klin Med*, 81, 317
- HENRICI, A T (1926) *J infect Dis*, 38, 54, (1928) *Morphologic Variation and the Rate of Growth of Bacteria* London
- HERSHEY, A D (1939) *J Bact*, 37, 280
- HERSHEY A D and BRONFENBRENNER, J (1937) *Proc Soc exp Biol N Y* 36, 556
- HIRSCH, J (1933) *Klin Wschr* 12, 191
- HOSKINS, J L (1934) *Publ Hlth Rep Wash* 49, 393
- HUNTINGTON, F and WINSLOW, C-E A (1937) *J Bact*, 33, 123
- JENNISON, M W (1935) *J Bact*, 30, 603
- JENNISON M W and WADSWORTH G P (1940) *J Bact* 39, 389
- JORDAN R C and JACOBS S E (1944) *J Bact*, 48 579
- KELLY, C D and RAHN, O (1932) *J Bact*, 23, 147
- KOJIMA, S (1923) *Sci Rep Inst infect Dis Tokyo Univ*, 2, 305
- LANE CLAYTON J E (1902) *J Hyg, Camb*, 9, 239
- LEDINGHAM J C G and PENFOLD W J (1914) *J Hyg, Camb* 14, 242
- LEMON C Q (1933) *J Hyg, Camb*, 33 495
- LIESE, W (1926) *Z Hyg Infekth*, 105, 483
- LONGSWORTH L G (1936) *J Bact*, 32, 307
- MASON, M M (1930) *J Bact*, 29, 103
- MCCRADY, M H (1915) *J infect Dis*, 17, 183 (1918) *Canad publ Hlth J*, 9, 201
- MACGREGOR, A S M (1910) *J Path Bact*, 14, 503
- MESTRE, H (1935) *J Bact*, 30, 335
- MILATZ J M W and POTTIER P B (1936) *Zbl Bakt Hte Abt*, 94, 227
- MILES A. A and MISRA, S S (1938) *J Hyg Camb* 38, 732
- MOYER, L S (1936) *J Bact* 32, 433
- MÜLLER, M (1895) *Z Hyg Infekth*, 20, 245
- PEARL, R and PARKER, S L (1922) *Proc nat Acad Sci Wash* 8, 212, quoted from MacLagan, D S (1932) *Proc roy Soc B* 111, 437
- PENFOLD, W J (1914) *J Hyg, Camb* 14, 215
- PENFOLD, W J and NORRIS D (1912) *J Hyg, Camb*, 12, 527

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## REFERENCES

- ALPER, T and STIERVE, M. (1933) *J Hyg., Camb.*, 33, 497.  
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 BAIL, O. (1939) *Z Immunforsch.*, 60, 1.  
 BARBER, M. A. (1908) *J infect. Dis.*, 5, 379.  
 BATEMAN, J. B. (1935) *Biol. Rev.*, 10, 42.  
 BREED, R. S. (1911) *Zell. Bakt., Hite. Abt.*, 30, 337.  
 BROOM, J. C. (1929) *Brit. J. exp. Path.*, 10, 71.  
 BUCHANAN, P. E. (1918) *J infect. Dis.*, 23, 109.  
 BUCHANAN, R. E. and FULMER, E. I. (1928) *Physiology and Biochemistry of Bacteria.* London.  
 BUICK, W. A. (1933-34) *Zell. Bakt., Hite. Abt.*, 89, 387.  
 BURKE, G. S. (1923) *J infect. Dis.*, 33, 274.  
 BURKE, V., SPRAGUE, A. and BARNES, L. V. (1925) *J infect. Dis.*, 36, 555.  
 CHESVEY, A. M. (1916) *J exp. Med.*, 24, 337.  
 CHICK, H. (1912) *J Hyg., Camb.*, 12, 414.  
 CHILD, C. V. (1915) "Individuality in Organisms." Chicago.  
 CLARK, P. F. and PUEHL, W. H. (1919) *J Bact.*, 4, 615.  
 COPLAND, M. (1910) *J Path. Bact.*, 14, 1.  
 ERBER, R. (1906) *Zell. Bakt.*, 19, 2.  
 FUKUDA, Y. (1929) *Z Immunforsch.*, 60, 88.  
 GILDEMEISTER, E. and NEUSTAT, M. (1935) *Zell. Bakt.*, 133, 101.  
 GILLESPIE, L. J. (1914) *J exp. Med.*, 19, 25.  
 GORDON, R. D. and ZOBELL, C. E. (1938) *Zell. Bakt. Hite. Abt.* 99, 315.  
 GRAHAM-SMITH, G. S. (1921) *J Hyg., Camb.*, 19, 133.  
 GREENWOOD, M. and YULE, G. U. (1917) *J Hyg., Camb.* 16, 36.  
 HALVORSON, H. O. and ZIEGLER, R. (1933a) *J Bact.*, 25, 101. (1933b) *Ibid.*, 26, 331, 339.  
 HEAP, H. and CADWELL, B. H. E. (1904) *J Hyg., Camb.*, 23, 77.  
 HELBER, E. (1904) *Deutsch. Arch. Klin. Med.*, 81, 317.  
 HEYRICH, A. T. (1926) *J infect. Dis.*, 38, 54. (1928) *Morphologic Variation and the Rate of Growth of Bacteria.* London.  
 HERSHEY, A. D. (1939) *J Bact.*, 37, 250.  
 HERSHEY, A. D. and BROUENBRENNER, J. (1937) *Proc. Soc. exp. Biol., N. Y.*, 36, 300.  
 HIRSCH, J. (1933) *Klin. Wochschr.*, 12, 191.  
 HOSKINS, J. H. (1934) *Publ. Hlth. Rep., Wash.*, 49, 393.  
 HUNTINGTON, E. and WINSLOW, C.-E. A. (1937) *J Bact.*, 23, 123.  
 JENNISON, M. W. (1935) *J Bact.*, 30, 603.  
 JENNISON, M. W. and WADSWORTH, G. P. (1940) *J Bact.*, 39, 359.  
 JORDAN, R. C. and JACOBS, S. E. (1944) *J Bact.*, 48, 59.  
 KELLY, C. D. and RAHN, O. (1932) *J Bact.*, 23, 147.  
 KOJIMA, S. (1923) *Sci. Rep. Inst. infect. Dis. Tokyo Univ.*, 2, 300.  
 LANE CLAYTON, J. E. (1909) *J Hyg., Camb.*, 9, 239.  
 LEDINGHAM, J. C. G. and PENFOLD, W. J. (1914) *J Hyg., Camb.* 14, 242.  
 LEMON, C. G. (1933) *J Hyg., Camb.*, 33, 493.  
 LIEST, W. (1926) *Z. Hyg. Infektkr.*, 105, 483.  
 LONGSWORTH, L. G. (1936) *J Bact.*, 32, 30.  
 MASON, M. M. (1935) *J Bact.*, 29, 103.  
 MCCRAID, M. H. (1915) *J infect. Dis.*, 17, 183. (1918) *Canad. pub. Hlth. J.*, 9, 201.  
 MACGREGOR, A. S. M. (1910) *J Path. Bact.*, 14, 503.  
 MESTRE, H. (1935) *J Bact.*, 30, 335.  
 MILATZ, J. M. W. and POTTER, P. B. (1936) *Zell. Bakt., Hite. Abt.*, 94, 227.  
 MILLS, A. A. and MISRA, S. S. (1938) *J Hyg., Camb.*, 38, 732.  
 MOYER, L. S. (1936) *J Bact.*, 32, 433.  
 MULLER, M. (1935) *Z. Hyg. Infektkr.*, 20, 245.  
 PEARL, R. and PARKER, S. L. (1922) *Proc. nat. Acad. Sci. Wash.*, 8, 212. quoted from MacLagan, D. S. (1932) *Proc. roy. Soc., B* 111, 437.  
 PENFOLD, W. J. (1914) *J Hyg., Camb.*, 14, 215.  
 PENFOLD, W. J. and NORMAN, D. (1912) *J Hyg., Camb.*, 12, 527.



- PULVERTAFT R. J. V. and LEMOV C. G. (1933) *J. Hyg., Camb.*, 33, 245  
 RAHN O. (1906) *Zbl. Bakt., Hte. Abt.*, 16, 417  
 PAHN O. and RICHARDSON G. L. (1912) *J. Bact.*, 41, 321  
 PÉGNIER, J. DAVID R., and KAPLAN A. (1932) *C. R. Acad. Sci.*, 194, 323  
 REICHENBACH, H. (1911) *Z. Hyg. Infektkr.*, 69, 171  
 ROGERS, L. A. and GREENBANK, G. R. (1930) *J. Bact.*, 19, 181  
 SCHMIDT H. (1926) *Z. Hyg. Infektkr.*, 106, 314  
 SCHMIDT H. and FISCHER, L. (1930) *Z. Hyg. Infektkr.*, 111, 542.  
 SCHULTZ, J. H. and RITZ, H. (1910) *Zbl. Bakt.*, 54, 283  
 SHERMAN J. M. and ALBUS W. R. (1923) *J. Bact.*, 8, 197 (1924) *J. Bact.*, 9, 303  
 SHERMAN J. M. and SAYLOR, H. B. (1924) *J. Bact.*, 43, 749  
 SKAN, O. (1931) *Z. Infektkr. Haus etc.*, 46, 110.  
 SLATOR, A. (1917) *J. Hyg., Camb.*, 16, 100  
 STEIN (1917) *Engng. News Rec.*, 78, No. 8, 391  
 STRAUSS, W. (1930) *Zbl. Bakt.*, 115, 228.  
 TOPLEY W. W. C. and WILSON G. S. (1936) "The Principles of Bacteriology and Immunity"  
 2nd ed., p. 70. Ed. Arnold & Co., London.  
 WALKER, H. H., WINSLOW C. E. A., HUNTINGTON E., and MOOREY M. G. (1934) *J. Bact.*  
 27, 303.  
 WIKULL, L. V. (1932) *Zbl. Bakt.*, 126, 488  
 WILSON G. S. (1927) *J. Bact.*, 7, 405 (1926) *J. Hyg. Camb.*, 25, 159 (1930) *Ibid.*, 30  
 433  
 WILSON G. S., TWIGG R. S., WRIGHT R. C., HENDRY C. B., COWELL, M. P., and MAIER, I.  
 (1935) *Spec. Rep. Ser. med. Res. Coun., Lond.*, No. 206  
 WINSLOW C. E. A. and WALKER, H. H. (1933) *Bact. Rev.*, 3, 14  
 WRIGHT A. E. (1907) *Lancet*, ii, 11

## CHAPTER 5

# THE RESISTANCE OF BACTERIA TO PHYSICAL AND CHEMICAL AGENTS    DISINFECTION

### INTRODUCTORY

THE early investigations of the problem of disinfection, which may be said to have commenced with Pringle's observations in 1750, were largely concerned with a study of the efficacy of various substances in hindering putrefaction. A century and a quarter later Bucholtz (1875) using as his medium an infusion of tobacco leaves, conducted a series of investigations on disinfectants, and Baxter (1875) working with vaccine lymph and glanders nodules, showed the influence of organic matter in diminishing the activity of disinfectants.

The next advance, illustrating the importance of technique, was made by Koch in 1881, when he introduced an exact method of comparing the germicidal power of different substances. In place of fluids swarming with different organisms of varying resistance, he tested the action of disinfectants on pure cultures of bacteria of approximately equal resistance. By drying anthrax spores on silk threads of the same length, immersing them in a solution of the substance to be tested, and subsequently transferring them to a nutrient medium in order to ascertain if the bacteria were still alive he collected a considerable quantity of information on the relative activity of different disinfectants. His work was criticized and his methods improved by Geppert (1889, 1891a & b). In 1897 Kromig and Paul published their classical paper, describing a new method for the quantitative study of disinfection and demonstrating that in a culture submitted to the influence of a germicidal agent bacteria die, not simultaneously, but in an orderly sequence. To Madsen and Nyman (1907) and to Chick (1908, 1910, 1912) must be ascribed the merit of analysing the various factors upon which disinfection depends, and of showing that the law underlying the death of bacteria is similar to that underlying a simple unimolecular chemical reaction.

Starting from empirical observations on the preservation of dead human bodies, the study of disinfection has progressed through the qualitative stage to the quantitative stage, and has now reached a point when the ultimate solution of the problem lies in the domain of the physical chemist.

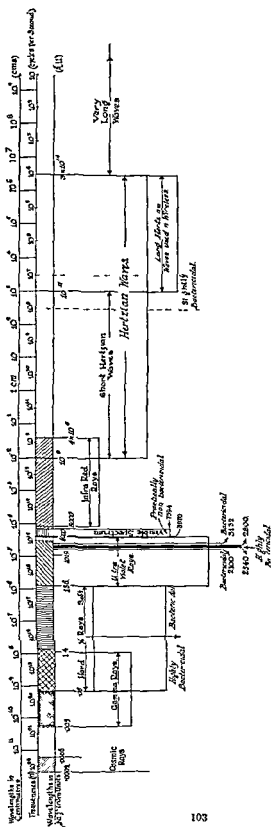
The subject of disinfection is large, and can be treated from different aspects. In the present chapter we shall make no attempt to deal with it exhaustively; on the contrary, we shall purposely neglect a considerable part of the subject dealing with the use of germicides in practice, as we consider this to fall within the province of the hygienist. Our main endeavour is to discover as far as possible the underlying principles of disinfection, to discuss the laws governing the killing of bacteria, and to point out the importance of a thorough knowledge of these

laws and principles to any one who, whether engaged in medicine, hygiene, dairy-farming, food preservation, or agriculture, is confronted with the problem of controlling bacterial activity

### Physical Agencies

**Light**—Downes and Blunt in 1877 (1877, 1878) showed that exposure of a putrescible fluid to sunlight was sufficient to sterilize it. They observed that this effect was produced only in the presence of air, and were therefore led to regard the germicidal property of light as depending on oxidation. Duclaux (1887) in 1886 showed that in sunlight vegetative bacilli were killed more rapidly than spores. The following year Roux (1887) exposed anthrax spores in a nutrient medium to the sun. Some were contained in glass tubes with plenty of air above the level of the liquid, others in glass tubes containing no free air, the former were destroyed in 29 hours, the latter survived longer than 83 hours. This was a confirmation of the work of Downes and Blunt, but Roux went further, he found that if nutrient broth was exposed to the sun in a layer 5 mm deep for 3 or 4 hours, it became changed in such a way as no longer to permit of the germination of anthrax spores, though still remaining suitable for the growth of the vegetative bacilli. This antiseptic property was lost after the broth had been allowed to stand for a time in the dark. Broth exposed to the sun in a sealed glass tube containing no free oxygen was unaffected. It was clear, therefore, that not only was sunlight in the presence of air able to destroy anthrax spores, but that it was able to produce an alteration in a nutrient medium—an alteration which was of a transient nature, suggestive of the activity of some volatile or unstable compound. This action of sunlight was reinvestigated by Burnet (1925), working with staphylococci, he found that they would not grow on agar plates that had been exposed to the sunlight, though growing quite satisfactorily on control plates that had been kept in the dark. He was able to show that the reason for this is that under the influence of sunlight hydrogen peroxide is produced, and that this substance is so powerful that its inhibitory effect is noticeable even in a dilution of 1-40 000 (see Chapter 3).

To return to the action of light on the bacteria themselves. Ward in 1892 exposed gelatin and agar plates seeded with anthrax spores to the autumn sunlight for 6 hours, each plate being shaded in such a way that only part of it received the direct rays of the sun. After incubation, a growth of anthrax bacilli was found to have occurred in the protected but not in the exposed portion. The inhibitory action of the sun was not due to the heat rays, because the temperature of the plates at no time rose above 18° C, nor was it due to dehydration or other alteration of the medium, since exposed plates seeded with fresh spores proved quite suitable for growth. Ward therefore concluded that the germicidal effect of the sun was due to its actinic rays. Further work showed that if a spectrum was thrown across an agar plate, the inhibitory effect of the light was stronger at the blue than at the red end. This was the first demonstration of the selective action of violet light. Some years later Barnard and Morgan (1903), working with the arc spectrum of carbon and of various metals, made more extensive observations, which led them to conclude that the bactericidal action of light was almost entirely due to those radiations in the ultra violet region which are included between the wave lengths 3287 and 2265 Å.U., that is, the light between the visible



# Range of Electromagnetic Waves

Uniform Velocity of 30 000 000 000 Centimetres per Second.

FIG 20 --DIAGRAM SHOWING THE RANGE OF ELECTROMAGNETIC WAVES WITH THE BACTERICIDAL ACTION OF RAYS OF DIFFERENT WAVE LENGTH

Note --Little work has been done on some parts of this range and none on others, so that the figure must be regarded as affording no more than an indication, incomplete and probably in part erroneous of the bactericidal action of these waves

The scale of this chart is taken from a diagram prepared by Vivian T Saunders, M.A., of Uppingham School, and published by John Murray, London, W 1, to whom we are indebted for permission to reproduce it

violet and the extreme ultra violet. No other portion of the spectrum had any effect whatever. Their work has been confirmed by Browning and Russ (1917), who used a quartz spectrometer illuminated by an arc of pure tungsten, which is very rich in ultra violet rays. The spectrum was thrown across a gelatin or agar plate seeded with staphylococci, the plate incubated after irradiation, and then used as an ordinary photographic negative for producing positive contact prints. An exposure of 6 minutes was sufficient to destroy the organisms in that region illuminated by rays of 2380 to 2940 Ångström units. (An Ångström unit is equal to  $1 \times 10^{-7}$  mm., i.e. 1/10,000,000th mm. One  $m\mu$  is equal to  $1 \times 10^{-6}$  mm., i.e. 1/1,000,000th mm., or 1/1,000th  $\mu$ , or 10 Ångström units.) There was a sharp line of demarcation at 2960 Å.U., rays of a longer wave length than this were almost devoid of germicidal action. It was found that the rays between 2960 and 2100 Å.U. were highly germicidal, but that the most active were those between 2800 and 2540 Å.U. As the limit of retinal sensibility is reached when the wave length falls to 3970 Å.U., it follows that the most actively germicidal portion of the spectrum is in that part which is invisible to the human eye (Fig. 20). More recent work by Gates (1930), Ehrismann and Noethling (1932), Buchholz and Jener (1933), Prudhomme (1937) and Rouyer and Servigne (1938), has shown that two of the most active wave lengths are 2650 and 2530 Å.U. Different organisms appear to vary in their susceptibility to different wave lengths.

Ultra violet light is lethal not only to bacteria, but to other unicellular organisms such as amœbæ (Barr 1923), and to the tissue cells of animals and plants. It is, in fact a protoplasmic poison. Though the short rays are undoubtedly the most active, the other rays of sunlight are not completely devoid of germicidal power, they require however, very much longer to produce their lethal effect (Thiele and Wolf 1907). Laroquette (1918) found that blue was more active than yellow, and yellow than red, green light was the poorest of all. The time necessary for destruction of micro-organisms by ultra violet light depends on the intensity of the light, the distance of the source of illumination, and the nature of the medium in which the organisms are exposed. In general, the Bunsen Roscoe law holds true, that is, within given limits the product of the intensity of irradiation and the length of exposure is constant. The temperature of the organisms at the time of exposure, provided that it is within the normal limits of viability, seems to have no effect on the action of ultra violet light (Rentschler *et al.* 1941).

The germicidal effect of sunlight under natural conditions varies greatly. Its action is complex, due partly to the actinic and partly to the caloric rays, which act by dehydration. Apart from its action on the organisms themselves, it has an action on the medium in which they are growing. In southern lands, the combined effect of the ultra violet rays and the heat rays render sunlight highly efficient as a germicidal agent, thus Semple and Greig (see Hewlett 1909) in India found that *Salmonella typhi* exposed to the sun on pieces of white drill cloth were killed in 2 hours, controls kept in the dark were still alive after 6 days. In the civilized smoke-covered towns of the north the ultra violet rays are often very weak, being largely filtered off by the impurities in the atmosphere, thus depriving sunlight of most of its activity. That light under these conditions is not entirely without effect however has been shown by Garrod (1914), who found that in hospital wards hæmolytic streptococci could be readily demonstrated in dust from the darker portions of the ward, but not from dust on window sills and other parts exposed to diffuse daylight.

Ultra violet light generated by a Cooper Hewitt Mercury vapour lamp has been used for the sterilization of drinking water Foulds (1911) found that it was quite easy to ensure a 99 per cent reduction in the bacterial count, including all *Bact coli*, by this method For further information, see Thresh and Beale (1910)

**Mode of Action of Ultra-violet Light**—Ultra violet light is peculiar in that its penetrating power is very low Even a thin layer of glass, such as a cover slip, is able to filter off a large proportion of the rays The same power is possessed by proteins It follows therefore that its action must be chiefly on the surface of the body which it irradiates Wesbrook (1896), by an ingenious series of experiments, showed that when tetanus or cholera cultures were irradiated by the sun in the presence of air, a consumption of oxygen took place D'Arcy and Hardy (1894) came to the conclusion that under the influence of ultra violet light, some oxidizing substance is produced—possibly ozone—which is responsible for its bactericidal effect This is very doubtful There seems to be no question that the destructive action of ultra violet light is manifest in the absence of atmospheric oxygen (Thiele and Wolf 1906, 1907, Blum 1932 Buchholz and v Jeney 1935), so that ozone can hardly be responsible Nor does it seem likely that hydrogen peroxide is generated in sufficient quantities to prove lethal, since Ehrismann (1930) found that, to produce a destruction of bacteria similar to that caused by ultra violet light, a concentration of 30 per cent  $H_2O_2$  was required

There is evidence that the ultra violet rays act by inducing some change in the protein molecule

Thus they are destructive, not only of organized cells, but of cellular products such as tetanus toxin (Kitasato 1891, Fermi and Pernosi 1894, Wesbrook 1896), and serum complement (Sellards 1918, Brooks 1920) Dreyer and Hanssen (1907) working with solutions of various albumins and globulins exposed in very thin layers, found that the rays caused a true coagulation of the proteins which were no longer soluble in weak acids or alkalis Corroboration of this view is found in the experiments of Tchahotins (1921) Working with the eggs of sea urchins, which are rich in lecithin, he found that if they were stained with neutral red and then irradiated with ultra violet light, the colour changed after a time to yellow, indicating the presence of alkali within the cell Further work seemed to show that the action of the rays was on the superficial membrane of the cell which was rendered more permeable to the OH ions of the medium, these, on penetrating the cell, were responsible for the change of the neutral red to yellow In support of this it was found that if the eggs were irradiated in a neutral solution no change in the colour of the dye took place From these and further experiments he comes to the conclusion that the rays act primarily on the superficial layer of the cell, coagulating its colloids, and rendering it more permeable to the ions in the surrounding medium Ehrismann (1930) exposed saline suspensions of various bacteria to ultra violet irradiation for 6 hours, and observed that a decrease in the opacity occurred, accompanied by a fall in the total count No change in pH, however, was noticed Agencies such as increased acidity, higher temperatures formol, and mercuric chloride, that tended to hasten the coagulation of protein partly or completely inhibited the clearing effect of the rays Analysis showed that the total nitrogen in the suspending fluid was increased though the amino-nitrogen figure remained unaltered

The conclusion appears to be that the rays produce a colloidal change in the protoplasm leading to the solution of certain of its constituents Whether true autolysis occurs in addition is still doubtful

Buchholz and v Jeney (1935) suggest that the reaction may possibly be of photo-chemical nature. These workers point out that the highly lethal waves 2600 and 2530 Å U correspond respectively to energy values of 4.6 and 4.8 volts and conclude that this amount of energy is required to displace sufficient electrons from the bacterial protoplasm to give rise to irreversible photo-chemical alterations and thus bring about the death of the cell. Wyckoff (1932) who has studied this aspect of the problem in the light of the quantum theory, finds that about 4 million quanta of energy are required to kill a single cocciform bacillus showing that death is not due as it appears to be with the cathode rays to a single quantum absorption but to some more generalized effect on the bacterial protoplasm. According to Lea and Haines (1940) about 100 times more energy is required to kill a bacterium when administered as ultra violet light than as X rays suggesting that the quantum yields of ultra violet light are very small. They conclude with Wyckoff that the energy of a single ultra violet light quantum is not usually sufficient to cause ionization. A similar conclusion has also been reached by Rentschler Nagy and Mouromseff (1941).

The mode of action of ultra violet light is still obscure, but the work of Gates (1930) and Ehrismann and Noethling (1932) renders it probable that it depends on the alteration of certain molecular groupings in the cell having high specific absorption spectra for these rays. The exact nature of the effect produced must await further observation.

**Photodynamic Sensitization** — We have seen that the visible rays of the spectrum have only a weak germicidal action on bacteria. It has been found however by Raab (1900) and by v Tappeiner (1900) that certain fluorescent dyes are able to sensitize infusoria to the action of these rays so that they become almost as lethal as the ultra violet rays. Thus it was shown that paramoecium suspended in a solution of acridin or eosin was killed very much more rapidly when exposed to diffuse sunlight which was itself harmless than when kept in the dark. Examining this phenomenon more closely, v Tappeiner projected a spectrum across a table and placed a culture of paramoecium suspended in 1-800 solution of eosin in the red, green violet and ultra violet parts. The culture exposed to the green rays was killed in 2 to 4 hours whereas the cultures exposed to the other rays appeared to be unharmed. This is of double interest firstly because the green rays by themselves are the least active in germicidal power and secondly, because it is in green light that eosin fluoresces most strongly. Similarly with acridin, death was most rapid when the suspension was exposed to the violet rays, these being the rays which cause acridin to fluoresce. For the sensitization to occur it was essential for the paramoecium to be in close contact with the fluorescing particles. When the cultures were exposed to light that was simply filtered through eosin they remained unharmed, the eosin had to be dissolved in the actual culture before its sensitizing action became apparent.

Later v Tappeiner and Jodlbauer (1904) and Jodlbauer and v Tappeiner (1904) demonstrated that the photodynamic action of dyes is manifest not only in relation to infusoria but in relation to bacteria toxins and to a less extent antitoxins while more recently the bacteriophage (Clifton 1931) and filtrable viruses (Perdrau and Todd 1933b) have been found susceptible.

Burge and Neill (1915) exposing various micro-organisms to ultra violet light found that the non fluorescent were killed more rapidly than the fluorescent bacteria. Their supposition is that the latter protect themselves from the coagulating

effect of ultra violet light by converting the short wave lengths into longer waves, and thus disposing of the energy of the absorbed short waves that would otherwise be spent in coagulating them. The non fluorescent bacteria are unable to do this, and hence succumb. In this connection, it is interesting to note that fluorescent bacteria may themselves exercise a sensitizing action on infusoria, thus taking the place of complex dyes. Jodibauer and v Tappeiner (1904) found that paramoecium suspended in a killed broth culture of *Ps. pyocyanea*, was killed in an hour if exposed to diffuse daylight, while surviving for 24 hours in the dark. Gram-negative organisms appear to be more resistant than Gram positive ones especially to the longer wave lengths (see T'ung 1935, Dreyer and Campbell Renton 1936).

Later work by Schmidt and Norman (1920) suggests that the photodynamic effect of certain dyes is not dependent simply on their power of fluorescence.

They showed that red blood cells mixed with eosin and exposed to sunlight were haemolysed, even if the rays that cause eosin to fluoresce were filtered off before reaching the solution. Again, in a mixture of red cells, eosin, and a protective substance, such as tyrosine, exposed to sunlight, there was no haemolysis even though the solution was fluorescing strongly. Clifton (1931) found that a staphylococcal bacteriophage suspended in 0.01-0.1 per cent methylene blue solution was inactivated by exposure to sunlight for 5 minutes or more. The reaction did not occur *in vacuo*, or in the presence of an active reducing agent such as cysteine hydrochloride (0.01 per cent). Perdrau and Todd (1933a), besides confirming these observations and finding that the optimal concentration of dye was about 1-100,000, showed that the interposition of a green screen prevented the reaction while a red screen did not.

The mode of action of dyes, in causing sensitization to light that is not itself markedly germicidal is not very clear, but it would appear that as the dye must be adsorbed on to the surface of the cell, and as oxygen is necessary for the effect to take place, the process is probably due to an activation of oxygen, or to an oxidation product of the dye (Bayliss 1924). (For a review of the whole subject see Blum 1932.)

**Electricity**—(1) *Direct Currents*. Little work has been done. Prochownick and Spaeth (1890) passed a galvanic current through simple saline suspensions of *B. anthracis*, *B. subtilis*, and staphylococci without much effect. After 2 hours *B. subtilis* had lost its motility, but was quite capable of growth. In another experiment the electrodes were coated with agar, seeded with organisms, immersed in saline, and a current passed through. No effect was noticeable at the cathode, but around the anode the organisms were killed. Thus a 60 millamp current destroyed *Staphylococcus aureus* in 15 minutes, and a 230 M.A. current destroyed *B. anthracis* in 30 minutes. They concluded that the effect was due not to the electricity *per se*, but to the nascent chlorine which was evolved at the anode from the electrolytically dissociated saline. Similar results were obtained in the same year by Apostoli and Laquerrière (1890). They employed a constant galvanic current, which was passed through a broth culture of *B. anthracis* into which the electrodes, situated a short distance apart, had been inserted. A current of 300 M.A. was fatal in 5 minutes, one of 200-250 M.A. failed to sterilize the culture in this time. They found that the action of the constant galvanic current was in direct relation to the intensity of the current, measured in milliamperes, that it depended far more on the intensity of the current than on the time for which it acted, and that the lethal effect was confined to the positive pole. They excluded



the effect of heat and were able to show that the sterilizing action of the constant current was due to the liberation of acids and of nascent oxygen at the anode.

(2) *Low-Frequency Currents* Beattie and Lewis (1920) were able to kill over 99.9 per cent. of organisms in milk by exposure for 4 minutes to an electric current, with a terminal voltage of about 4 000 and an amperage of about 2. Most of the sterilizing action appeared to be due to heat as the temperature rose to between 60 and 64° C, but the authors considered that this alone was insufficient to account entirely for the effect.

(3) *High Frequency Currents* Apart from the early experiments of D Arsonval and his colleagues in 1893 to 1896 (for references see Fabian and Graham 1933) little work has been carried out till recently on the action of high frequency currents. During the past few years however a number of workers have made observations on the effect of these currents on bacteria, bacteriophage toxins and antibodies (Szymanowski and Hicks 1932, Hicks and Szymanowski 1932, Lentze 1932, Fabian and Graham 1933, Hasche and Leunig 1933, Gale and Miller 1933). The results are not easy to summarize since the conditions of exposure used by different workers were often very different. In Fabian and Graham's experiments a gradual destruction of *Bact. coli* was brought about by exposure to a high frequency displacement current of 10 megacycles per second and an intensity of 0.8 amps, but even after 8 hours the suspension was not sterile. The higher frequencies used by most of the other workers appeared to be less harmful. Whether the current acts mainly by generation of heat in the medium, or by setting up intense electronic and ionic linear agitation within the cells, is doubtful. The observations of Bessemans and van Merhaeghe (1937), Ozzano and Re (1937) and Hasche and Loch (1937) all suggest that high frequency currents have little effect on bacteria apart from the heat generated. [1 megacycle = 1 000 kilocycles = 1 000 000 alternating cycles. Since  $V = n\lambda$ , when  $V$  = velocity of travel (186 000 miles per second)  $n$  = frequency or number of vibrations of the wave per second and  $\lambda$  = the wave length, it can be calculated that a frequency of 10 megacycles corresponds approximately to a wave length of 30 metres.] Short wave therapy using radiations of 3 to 30 metres in wave length, is now on trial in clinical medicine for the treatment of certain inflammatory processes.

*Cathode Rays*.—Wyckoff and Rivers (1930) working with *Bact. coli*, *Salmonella typhimurium* and *Staph. aureus* bombarded single bacteria on the surface of an agar plate with a known number of cathode rays. The proportion of surviving organisms was estimated from colony counts made after incubation of the plates. The cathode rays were generated in a Coolidge type electron tube working at a voltage of approximately 150 kilovolt. Destruction of the organisms occurred in the usual semi-logarithmic fashion (see p. 137). After 20 seconds 83.193.9 per cent. of the organisms were dead. Quantitative analysis rendered it evident that the absorption of a single electron was generally sufficient to cause death.

The action of cathode rays seems to depend on the release of large numbers of ions consequent on the absorption of an electron. A single 150-kilovolt electron will liberate about  $10^4$  ions within less than 0.001 c.m. The effect of such an ionic shower on organisms as small as those mentioned seems to be almost invariably lethal though with yeast cells injury and not death may result (Wyckoff and Luyet 1931).

*Röntgen Rays*.—According to Rieder (1902) the cholera vibrio when exposed on an agar plate at a distance of 10–12 cm. from the anti-cathode is killed by

Rontgen rays in 20 to 30 minutes Feistmantel (1902) found that irradiation for 50 minutes of *Actinomyces farcinica*, exposed 10 cm away from the anti-cathode, had apparently no effect Wyckoff (1930a, b) exposed *Bact coli* and *Salmonella typhimurium* on the surface of agar plates to soft X rays, obtained either from a tungsten tube operated at low voltage, or by the characteristic K radiation of copper Destruction occurred *sensu* logarithmically, but less rapidly than with cathode rays (see above) Thus after 20 seconds' exposure to filtered copper radiation only 19.6–33.3 per cent of organisms were dead Analysis showed that only about one in twenty of the absorbed quanta of these radiations proved lethal

According to Wyckoff, the X rays incident upon a cell either pass through without altering it, or else give up one or more quanta whose energy content is connected with the wave length  $\lambda$  of the rays through the relation

$$E = h\nu = \frac{hc}{\lambda}$$

where  $h$  is Planck's constant,  $\nu$  is the frequency of the rays, and  $c$  is the velocity of light

As the result of such an absorption a high-velocity electron is liberated This electron gives rise to a chain of ions in the matter through which it passes and to X rays which, in their turn, liberate more ions of less and less energy The changes caused by X rays in protoplasm are naturally identified with the physico-chemical changes induced by this ionic shower The fact that only one in twenty of the absorbed quanta proves fatal suggests that the vital elements capable of being destroyed by a direct quantum hit occupy only about one twentieth of the cell volume The harder the X rays are, the nearer do they approach in their killing effect to the cathode rays More recent work with *Bact coli* by Pugley, Oddie, and Eddie (1935) yielded results which seemed to show that, provided a correction factor was introduced for lack of uniformity of the X ray beam, the organisms died in an exponential manner Discussing these results, the authors came to the conclusion that the one quantum hit to kill explanation, first put forward by Crowther (1926), appeared to account most satisfactorily for the type of curve obtained

This explanation is supported by the observations of Lea, Haines and Bretscher (1941) Comparing the bactericidal action of X rays, neutrons, and radioactive radiations, these workers found that the effect of a given dose was independent of the temperature and of the rate at which the radiation was applied, and that the mean lethal dose was correlated with the ionization density of the radiation, being greatest for those radiations which produced their ionizations closest together These findings are explained on the assumption that each organism contains a number of sensitive "targets" and that the destruction of any one target is sufficient to impair the viability of the organism *Bact coli* is estimated to contain about 1,000 targets, each of a diameter of 8.6  $\mu\mu$ , equivalent in size to a molecule having a molecular weight of  $2 \times 10^5$

Sublethal doses of X rays may bring about changes in the morphology and growth characteristics of bacteria (see Levin and Lominski 1935, Forfota and Hamori 1937) It may be noted that bacteriophages and filtrable viruses have been shown to be susceptible to soft X rays (Wright and Kersten 1937, Moore and Kersten 1937)

Radium—Bruynoghe and Dubois (1925) found that exposure of *Leptospira icterohæmorrhagiae* for 26 hours to 8 mgm of radium, enclosed in a platinum cell  $\frac{1}{2}$  mm in thickness, rendered the organism incapable of growing *in vitro* or of giving rise

to disease in the guinea pig, but did not interfere with its motility Bruynoghe and Le Fevre de Arrie (1925) stated that they were able to deprive the viruses of rabies and of herpes of their virulence for rabbits by exposure in fairly high dilution to radon, in a dose of 5 millicuries for 48 hours. Danysz (1906) failed to produce any attenuation of the rabic virus by exposure for 20 hours to the  $\beta$ - and  $\gamma$ -emanations from 20 mgm of radium bromide Bisceglie (1926) claims to have lowered the virulence for guinea pigs of a human strain of tubercle bacillus by exposure of three successive generations to 5 mgm. of radium bromide, the exposure being maintained for 5 days. Morphologically the bacilli of the third generation had lost their acid fast properties to a considerable degree, thread forms, occasionally showing branching, were numerous, and large numbers of Gram positive Much granules were visible According to Suess (1908), exposure of tubercle bacilli to highly active radium emanations for 2 days had apparently no effect on their morphology, growth, or pathogenicity von Schroetter (1927) finds that bacilli and cocco-bacilli exposed to radon, in a dose varying for different organisms from 0.5 to 40.0 millicuries of an intensity of 5-250 microcuries, tend to elongate and become filamentous, cocci, on the other hand, swell, increasing more or less equally in size in all diameters. Spirochetes do not change their size, they are eventually killed by the rays, but they remain motile for a considerable time Spencer (1934 1935) implanted radium needles in tubes of broth inoculated with *Salm. typhi*, *Proteus* X19, or *Str. pyogenes* and incubated at 37° C There was at first a slight retardation of growth, but after 24 hours the growth was similar to that in control tubes. After 8-10 daily transfers the irradiated organisms sometimes grew more luxuriantly and tended to develop filamentous forms or, with streptococci to grow in long chains. On the other hand irradiation at 0° C proved fatal within a few days. Lea, Haines, and Coulson (1936) have recently studied the effect of  $\alpha$  and  $\beta$  rays on *Bact. coli*, *Staph. aureus*, and *B. mesentericus* exposed in very thin gelatin films Death of the organisms occurred exponentially The rate of disinfection was found to be independent of the temperature, and proportional to the intensity of the radiation All three organisms were equally sensitive to  $\alpha$  rays, but towards  $\beta$  rays *B. mesentericus* differed from the other two organisms The authors conclude that the action of the radiation can be explained best on the "target" hypothesis. In a later paper Lea, Haines and Coulson (1937) record that the death rate of *Bact. coli* and *B. mesentericus* exposed to  $\gamma$  radiation was of the exponential type, and that the mean lethal ionization doses were approximately the same as those previously observed for  $\beta$  rays.

**Sonic and Supersonic Waves**—Starting with the experiments of Wood and Loomis in 1927, several observations have been made of recent years on the destruction of organized cells by high frequency sound waves (for references up to 1932 see Chambers and Gaines 1932) Sonic waves, i.e. waves of audible frequency, of about 8,900 cycles per second, produced by a nickel tube vibrating in a strong electromagnetic field in resonance with a 2,000-volt oscillating power circuit, are said to be able to bring about a considerable destruction of coliform and certain other bacteria exposed to them for sufficient lengths of time.

The method has been used by Chambers and Floedorf (1936) for the liberation of antigenic constituents from *Salm. typhi* and hæmolytic streptococci. Death was found to occur logarithmically, the slope of the survival curve being a function of the sound intensity, i.e. of the amplitude when the frequency was constant The suspensions were not com

pletely sterile at the end of an hour, but filtrates showed the presence of the typhoid Vi antigen and the streptococcal labile antigen respectively. Rivers, Smadel and Chambers (1937) were successful in bringing about a considerable degree of destruction of vaccinia virus in the form of washed elementary bodies but not in ordinary tissue suspensions. The presence of protein in the suspension interferes with the effect of the waves, this may explain why attempts to destroy bacteria in milk, bacteriophage in cultures, and viruses in tissue suspensions have often proved a failure (Beckwith and Weaver 1936, Scherp and Chambers 1936).

Supersonic waves, i.e. waves above audible frequency, of 200 000 to 1,500,000 cycles per second, produced by connecting a piezo-electric crystal with a high-frequency oscillator, are also credited with bactericidal power. The observations of Beckwith and Olson (1932), Yen and Liu (1934) and Takahashi and Christensen (1934) suggest that a considerable destruction of bacteria, and even of filtrable viruses, may be brought about by exposure to these waves for an hour or so. Paic and his colleagues (1935a, b), however, found that ultrasonic waves of a frequency of 280,000 cycles per second had no destructive action in 2 hours on certain toxins, a coli bacteriophage, the herpes virus, or a number of different micro organisms, while completely sterilizing a culture of *Paramacium* in 5 minutes.

Too little work has yet been carried out to justify a critical discussion of the results. It is generally believed that the action of the waves, which are of course molecular and not electro magnetic, is due to the disruption of the cell as a result of the violent agitation set up in its contents. According to Liu and Yen (1934) no effect is produced on cells exposed *in vacuo*, suggesting that cavitation of dissolved gases plays an important part in the disruption of the bacteria. Probably a relationship exists between the wave length and the size of the organism or molecule exposed. Against the simple disruption explanation may be set the observations of Rivers, Smadel and Chambers (1937) on the effect of some waves on washed vaccine virus. Though the infective titre of the suspension fell from  $10^{-8}$  to less than  $10^{-2}$ , no microscopical evidence of disintegration of the elementary bodies could be obtained, nor was there any increase in the opacity of the suspension. The authors, therefore, suggest that the destructive effect may be due to the formation of some oxidizing substance from the water.

**Desiccation**—If dried on silk threads or glass slips, the proportion of organisms surviving for any given length of time varies with a great number of factors, such as the species of bacterium, the initial numbers present, the nature of the suspending medium, the rapidity of drying, and the temperature and gaseous nature of the environment (see Ficker 1898). Anthrax spores dried on silk threads may survive for over 20 years, while many of the pathogenic non sporing bacteria die in a few hours. Paul, Birstein, and Reusz (1910a), working with staphylococci dried on garnets, found that the velocity of disinfection was equal to the square root of the oxygen concentration. The lower the temperature at which the organisms were kept after being dried in this way, the smaller was the proportion that succumbed (Paul 1909).

More recent work (see Otten 1930, 1932, Elser, Thomas, and Steffen 1935, Flosdorf and Mudd 1935) has shown that even non sporing pathogenic organisms are able to survive drying indefinitely, provided that desiccation is complete and that the dried organisms are maintained in a high vacuum (0.01 mm Hg or less). Even such sensitive organisms as the meningococcus and the gonococcus remain

alive and virulent for years under these conditions. The dried organisms are resistant to quite high temperatures. Typhoid bacilli, for example, dried and sealed in *vacuo*, are said to survive exposure to a temperature of  $115^{\circ}\text{C}$  for over 30 minutes. Serum and complement can also be preserved satisfactorily by drying from the frozen state (see Hartley 1936, Floedorf and Mudd 1938).

The method is now being used extensively for the preservation of stock cultures. In practice 0.5–1.0 ml quantities of a thick suspension of the organisms in broth are distributed into suitable tubes. Drying is carried out as rapidly as possible in *vacuo* in a desiccator over phosphorus pentoxide. The tubes are then evacuated individually with an efficient pump, and sealed off in the flame. To recover the organisms, an optimal medium is desirable for primary cultivation.

**Cold.**—Very much less attention has been paid to the effect of cold on bacteria than to the effect of heat. This may undoubtedly be attributed to the fact that, although cold is an excellent means of preventing putrefaction, it has very little germicidal action. Macfadyen (1900) exposed cultures of *Bact. coli*, *Salmonella typhi*, *B. anthracis*, *V. cholerae*, *Proteus vulgaris*, and *Staphylococcus aureus* for 20 hours to liquid air at a temperature of  $-182^{\circ}$  to  $-190^{\circ}\text{C}$ . After exposure the organisms grew well on subculture, and manifested their usual biochemical activities. He noticed that photogenic bacteria, when frozen, became non luminous, but, when re-thawed, their luminosity returned with unimpaired vigour. In another experiment he exposed the same organisms in broth suspensions enclosed in fine quill tubing for 7 days to liquid air, subsequently no structural alteration could be detected in the bacteria, and all grew well on subculture. Macfadyen and Rowland (1900) found that the same organisms in sealed glass tubes withstood immersion for 10 hours in liquid hydrogen at a temperature of  $-252^{\circ}\text{C}$ , microscopically and culturally the bacteria appeared to be unaltered. Paul and Prall (1907) exposed staphylococci, which had been dried on garnets, to liquid air, and found that under these conditions they retained their viability for several months, and showed no appreciable alteration in their resistance to disinfectant agencies.

Haines (1938) found that rapid freezing with solid carbon dioxide at  $-70^{\circ}\text{C}$  killed a high proportion of some organisms, but had little effect on others. If the frozen organisms were subsequently stored at  $-20^{\circ}\text{C}$ , they died off very slowly, but if they were stored at  $-1^{\circ}$  or  $-2^{\circ}\text{C}$  they died rapidly. Evidence was brought to suggest that at the latter temperatures denaturation and subsequent flocculation of the bacterial protein occurred similar to the changes that have been observed in muscle. In practice, a temperature of  $-70^{\circ}\text{C}$  is very useful for the preservation of many bacteria and filtrable viruses.

**Dry Heat.**—We have seen that disinfection by drying is influenced by numerous small factors, in disinfection by heat, though numerous small factors may play a part, the one factor, heat, is so important that it overshadows them. We can therefore be more precise in our figures regarding this method of disinfection. Koch and Wolfhubel (1881) were the first to make exact measurements of the effect of heat on micro-organisms. They found that vegetative bacteria were killed by a temperature of just over  $100^{\circ}\text{C}$  in  $1\frac{1}{2}$  hours, many, of course, succumbed well within this interval, but this was the time necessary for complete sterilization. Spores, on the other hand, were much more resistant, requiring a temperature of  $140^{\circ}\text{C}$  for 3 hours for destruction. On what this superior power of resistance of spores depends is not known. Probably it is related to their

lower water content, since there is evidence to show that desiccation can raise the time temperature limit necessary to cause coagulation of proteins (see Hewlett 1909 Cameron 1930). The resistance of both vegetative bacteria and of spores varies considerably with the different species some being killed much more rapidly than others. The spores of moulds are intermediate in resistance between the vegetative and sporing bacteria they require a temperature of 110–115° C for 1½ hours for their destruction.

As with desiccation the higher the temperature the shorter is the survival time. Thus if the temperature is raised from 140° to 160° C spores are killed in 1 to 1½ hours. At 400° C they are killed in 20–30 seconds (Oag 1940).

Koch did not regard dry heat as an efficient method of disinfection. Though satisfactory when dealing with naked bacteria it is quite ineffectual within the times usually employed when the bacteria are protected by textile or other relatively non-conducting material. This is due to the low power of penetration of hot air. Thus when a bundle of tow measuring 55 × 50 cm was exposed to a temperature of 140–150° C the interior after 3 hours had only reached the temperature of 74.5° C—a temperature quite inadequate to kill the spores enclosed in the bundle. Moreover a temperature of 140° C is sufficient in a short time to ruin most cloth fabrics.

Flaming is a useful method of surface disinfection for non-inflammable substances its efficacy appears to depend on the temperature to which the exposed surface is raised (Mayer 1925).

**Moist Heat.**—Koch (Koch *et al* 1881) in conjunction with Gaffky and Loeffler was the first to make a quantitative study of the germicidal action of moist heat. He found that the temperature required for sterilization of spores was much lower than with dry heat. Thus anthrax spores were killed in 10 minutes at 95° C and spores present in garden earth in less than 10 minutes at 105° C. He also showed that steam under pressure is more efficient than steam at atmospheric pressure. For the disinfection of clothes too he found moist heat to be preferable to dry heat as it has a greater penetrating power. Thus after 4 hours dry heat at 140–150° C the temperature inside a roll of flannel was only 83° C and the contained spores germinated freely whereas after 1½ hours of moist heat at 120° C the temperature inside was 117° C and all the spores were dead. Koch was greatly impressed by the value of boiling water from numerous experiments he concluded that even spores seldom survive its action for more than a few minutes. We now know that Koch rather over-estimated its efficacy for there are certain bacteria the spores of which will resist the action of boiling water for hours. This is especially marked with the thermophilic bacteria thus Bigelow and Esty (1920) exposed the spores of thermophilic organisms suspended in a nutrient medium of pH 6.1 in sealed glass tubes to various temperatures in oil baths with the following results

Temperature	Killed in
100° C.	13.0 minutes
110° C	0.5
120° C	23
130° C	3.5
140° C	1.0

Thus at a temperature of 100° C they remained viable for nearly a day

Mundel (1937) has found that the addition of 2 per cent washing soda ( $\text{Na}_2\text{CO}_3$ ) greatly increases the disinfecting power of boiling water as well as reducing the tendency of metal instruments to rust. Thus, spores in a 0.35 per cent. suspension of earth in water resisted boiling for about 10 hours, but were killed in a 2 per cent. solution of soda at  $95^\circ\text{C}$ . in 10 to 30 minutes.

It is on account of the resistance of spores to boiling water that the autoclave has largely displaced the steamer in laboratory practice. Steam is still employed at atmospheric pressure for the sterilization of certain media the physical or chemical composition of which would be altered by steam under pressure but where this is necessary we take advantage of Tyndall's observation, and submit the medium to steaming for 30 minutes on 3 successive days any sporing organisms that have not been killed on the first day germinate and thus become susceptible to exposure on the second day. It must be realized that tyndallization can be successful only if the nature of the medium and the conditions to which it is subjected between successive heatings are such as to enable all the spores to germinate. It is quite inapplicable for instance to the sterilization of bacterial suspensions in a non nutrient fluid. Similarly it is unsuitable for the destruction of anaerobic spore bearing bacteria the spores of which are unable to germinate aerobically or of thermophilic spore-bearing organisms which fail to grow at a temperature below  $50\text{--}55^\circ\text{C}$ .

Steam under pressure on the other hand is so effective that a single sterilization usually suffices. In the autoclave the steam, while being submitted to pressure still remains saturated with moisture. This is most important. Steam which is superheated behaves like a gas, and condenses very slowly on objects cooler than itself. Steam that remains saturated with moisture is much more effective as it rapidly condenses on objects cooler than itself and by giving up its latent heat quickly raises them to its own temperature. All air must be expelled from the autoclave before the pressure is allowed to rise otherwise the temperature developed by a given pressure of steam will be less than that reached by saturated steam (see R. H. Worth 1931. Spooner and Turnbull 1949). Though there are a few exceptions it is safe to say that saturated steam under a pressure of 15 lbs. per square inch, i.e. with a temperature of about  $120^\circ\text{C}$  is sufficient to sterilize any medium in 30 minutes. This is therefore the exposure to which the usual media are submitted.

The higher the temperature provided the steam remains saturated, the more rapid is the sterilization. This is clear from the results of Bigelow and Esty. But there are certain factors other than temperature that affect the time necessary for sterilization by steam. One of the most important is the H ion concentration of the medium.

It will be remembered that Pasteur in his experiments on spontaneous generation (see Chapter 1) found that boiling was more lethal in an acid than in an alkaline medium. This has since been confirmed repeatedly. Bigelow and Esty (1920) for example working with the spores of thermophilic organisms found that when suspended in an acid medium of pH 4.6 they were destroyed by a temperature of  $100^\circ\text{C}$ . in 2 minutes, whereas in a less acid medium of pH 6.1 it required 9 minutes to destroy them. Chick (1910) likewise found that minute quantities of acid or alkali, too small of themselves to produce any germicidal action, had a very marked influence on the power of disinfection by hot water. With these substances the rate of disinfection was increased, but much more with the acid than with the alkali. Thus working with *Salmonella typhi* suspended in distilled water she

found that the addition of sufficient alkali to render the solution  $N/7,000$  alkaline increased the mean rate of disinfection at  $54^{\circ}\text{C}$  about 1.5 to 2 fold, a similar addition of acid increased it 5 to 7 fold. Further addition of alkali influenced the rate of disinfection but little, whereas further addition of acid rendered it too rapid for study.

Other factors are the age of the culture and the nature of the suspending medium. Young organisms are generally more susceptible to the action of heat and of chemical disinfectants than old, while the presence of protein in the suspension, or of sugar in considerable concentration (Fay 1934) tends to protect the organisms to some extent. (For general information on sterilization by steam see Underwood 1934, Konrich 1938, and on the sterilization and testing of dressings, in particular, see Hayes 1937, Savage 1940, Chisholm 1941, Report 1942.)

**Thermal Death Point of Bacteria.**—The mode of action of heat on bacteria appears to be one of protein coagulation. Chick and Martin (1910) showed that heat coagulation of proteins is an orderly process, the rate of which varies with the alteration of temperature, reaction of the medium, and other conditions. The actual process of coagulation consists of two stages: in the first, known as denaturation, the water reacts with the protein; in the second, known as agglutination, the altered protein separates out in a particulate form. In the case of hæmoglobin the coagulation occurs logarithmically, the rate at any moment being proportional to the concentration of uncoagulated protein. Very much the same law appears to be applicable to bacteria. The higher the temperature to which they are submitted, the more rapidly is their cellular protein coagulated. Between different organisms there are considerable variations; thus some vegetative bacteria, such as the gonococcus, are destroyed by heat at  $47^{\circ}\text{C}$  in a few minutes; others such as the enterococcus, withstand a temperature of  $60^{\circ}\text{C}$  for nearly an hour. It must not, however, be supposed that these temperatures are to be regarded as specific thermal death points, irrespective of the time of exposure. Chick (1910) has shown that the death of bacteria under the influence of heat is due to a protein coagulation, that this phenomenon occurs not at one definite point on the temperature scale, but over a considerable range of temperature, and that therefore the death of bacteria within a given range is mainly a function of time. To take for example *Salmonella typhi*: the thermal death point of this organism is usually given as  $55^{\circ}\text{C}$ . In experiments carried out between  $49^{\circ}$  and  $59^{\circ}\text{C}$  the temperature coefficient, i.e. the rise in the velocity of disinfection, was found to be 1.635 for  $1^{\circ}\text{C}$ . Given a value for  $k$  (see p. 137) of 0.111 at  $49^{\circ}\text{C}$ , it can be calculated that a suspension containing 100,000 bacilli per ml. would be sterilized in about 2 hours at  $47^{\circ}\text{C}$ , in 48 minutes at  $49^{\circ}\text{C}$ , in 18 minutes at  $51^{\circ}\text{C}$ , in 7 minutes at  $53^{\circ}\text{C}$ , in  $2\frac{1}{2}$  minutes at  $55^{\circ}\text{C}$ , and in 21 seconds at  $59^{\circ}\text{C}$ . If therefore a suspension was gradually heated, death might apparently take place suddenly at  $55^{\circ}\text{C}$ . But it is clear that this cannot be regarded as a point possessed of any special significance; it is merely a point near the upper end of a series of temperatures, each of which in itself can legitimately be regarded as a thermal death point. It follows that for purposes of comparison of the heat susceptibility of organisms of different species, it is essential to use suspensions of equal numbers of bacteria, and to ascertain at what temperature complete sterilization is produced within a given time. Even with these precautions, as we shall see later, there is a certain inaccuracy, due to the apparent variation in susceptibility of organisms of the same species in the same suspension, resulting in the survival of some long after the majority have been killed.



**Effect of Heat on Subsequent Multiplication**—In his studies on disinfection, Koch noticed that spores which had been heated but not quite killed required longer to germinate than unheated spores. Similar observations have been recorded by numerous workers both with spores and with vegetative bacteria. The conclusion usually drawn is that during the process of heating the organisms are damaged in some way, so that their ability to multiply when subsequently transferred to suitable conditions is interfered with. Certain figures of Eijkman (1908) lend support to this view; he heated a suspension of *Bact. coli* in saline at 52° C and after varying intervals he made duplicate plates to ascertain the number of organisms remaining alive. One set of plates was counted after 3 days' incubation, and the other set after 15 days' incubation. The results are shown in Table 4.

TABLE 4

Length of heating at 52° C	No. of organisms developing after incubation for —	
	3 days.	15 days.
0 minutes	336 000 000	336 000 000
½	144 000 000	144 000 000
1	115 200 000	198 000 000
2	51 700 000	65 600 000
3	4 000 000	33 600 000
5	800 000	2 "20,000
6	0	640 000
10	0	3 750
15	0	1 000
35	0	0

It will be noticed that during the first 30 seconds the heat though killing over 50 per cent of the organisms does not interfere with the reproduction of the remainder. Subsequently, the longer the organisms are exposed the greater is the difference between the results of the two series of plates. This suggests that a certain proportion of the remaining viable organisms are injured, and that the longer they are subjected to heat, the greater is the interference with their reproductive power.

Similar results have been obtained by Allen (1923), who, working with milk, found that the generation time of non sporing organisms which had been pasteurized was longer than that of the untreated organisms indicating an attenuation of the pasteurized organisms.

A different interpretation has however, been proposed. Eckelmann (1917) suggests that the reason why a certain proportion of heated organisms require a long time to germinate is not because they are suffering from the effects of heat but because they are provided with a more resistant cell membrane, which while allowing them to withstand temperatures that prove lethal to their fellows, interferes with their rapid reproduction. According to her, heat would act as a selective agency, killing off all the bacteria with thin cell membranes and a power of rapid reproduction, and leaving intact the bacteria with thick, relatively impermeable cell membranes and a restricted power of reproduction. Burke (1923) adheres to the same view. She found that the spores of *Cl. botulinum* frequently took several days to germinate even when placed under optimum conditions. When the spores were heated the germination period was increased generally.

in proportion to the length of exposure, she found that they might be dormant for as long as 426 days. Her conclusions are that the reason why the spores resist heat is because they are characterized by the possession of an impermeable membrane, which is also the cause of their delayed germination. The thicker the membrane, the more resistant is it to heat, and the longer does the organism take to develop.

The evidence in favour of the second view does not appear to us to be as convincing as that in favour of the first. Enkman's figures, given in Table 4, are very striking, and, assuming their general validity, it is difficult to avoid concluding that the effect of heat is to increase the lag period of such organisms as remain viable. That the escape of the few is dependent on the possession of a relatively impermeable cell membrane is quite possible, but it fails to explain why in Enkman's experiments before heating all organisms developed in 3 days, whereas after heating some failed to develop for 15 days.

Though sub lethal heat may delay germination of spores, the opposite effect has been recorded by some workers. Christian (1931a, b), for example, working with an aerobic spore bearing bacillus isolated from tainted milk, found that germination appeared to be stimulated by heating the spores to 100° C for 30 minutes at the time of inoculation. Evans and Curran (1943) also observed that heating spores of some species of aerobic spore bearing bacilli for 10 minutes at 85° C often stimulated germination. In general mild heating followed by 3 hours' incubation led to about the same degree of germination as 24 hours' incubation without pre heating.

### Chemical Agencies

**Distilled Water**—The evidence concerning the action of distilled water on the viability of bacteria is most conflicting. Spores are undoubtedly able to survive for a long time, thus Koch found that spores of the anthrax bacillus remained alive for more than 90 days. But with vegetative organisms it is otherwise. Some workers have found that they will survive for weeks, others that they are destroyed in a few hours. Such confusion can be explained only by differences in technique. One such difference of primary importance is the nature of the vessel from which the water is distilled. When a metallic still is used, traces of the metal are carried over into the distillate and undoubtedly exercise a deleterious effect on the bacteria. Ficker (1898) found for instance, that water containing copper sulphate in a dilution of 1/50,000,000 was sufficient to kill cholera vibrios in 1 hour, while Hoder (1932) found that distilled water containing 1 part of copper in 10 million sterilized a suspension of *Ps. pyocyanea* in 2 hours.

But even those who state that they used glass distilled water have obtained varied results. One reason for this discrepancy may lie in the number of bacteria inoculated. In this connection some striking figures are reported by Ficker (1898). In one experiment he seeded pure glass-distilled water with 60,000,000 cholera vibrios per ml, and found that they remained viable for several months. In another experiment, in which he reduced his inoculum to 10,000 per ml, the organisms were nearly all dead in 2 hours. He explains such a difference on the assumption that the inoculation of large numbers of organisms into distilled water converts this into a dilute nutrient medium, no longer possessing the essential purity of the initial menstruum. The H ion concentration of the water may also be an important factor. Thus Winslow and Falk (1923) give the following figures, compiled from no fewer than seventy nine tests.

VIABILITY OF *Bact coli* IN DISTILLED WATER AFTER 9 HOURS AT 37° C.\*

pH	4.0	5.0	6.0	7.0	7.5	8.0
Percentage of organisms surviving	1	82	106†	54	35	12

\* pH was adjusted by minute additions of acid or alkali

† This figure suggests that a slight increase in the numbers of surviving organisms may have occurred

The maximum viability occurs at pH 6.0 Cohen (1922) has likewise shown that when the pH of water is stabilized by the addition of buffer salts, the results are much more regular

Other factors that may influence the action of distilled water on bacterial viability are traces of alkali absorbed from the glass, the amount of CO<sub>2</sub> absorbed from the air, the quantity of dissolved oxygen, and the temperature at which the suspension is maintained Whipple and Mayer (1906) studying the length of life of *Salm typhi* in sterile tap water showed that it remained viable for nearly 2 months when the water was exposed to the air, but died out in 4 days when the water was kept under anaerobic conditions Houston (1914) likewise working with *Salm typhi*, found that when suspended in water kept at 0° C, it lived for 8 weeks, at 18° C for 3 weeks, and at 37° C for only 1 week

Summing up we may say that the length of life of vegetative bacteria in distilled water is influenced by a large number of factors When these factors are all favourable the organisms may remain viable for considerable periods, when unfavourable, they may die out in a very short time, further, the effect varies greatly with different species of organisms. There is no evidence that distilled water acts by causing disruption of bacteria, as it does of many unicellular organisms bacteria are too resistant to changes in osmotic pressure for this to be probable

Acids—Kronig and Paul (1897) were the first to show that the disinfectant action of acids in general is proportional to their degree of electrolytic dissociation, i.e. to the H<sup>+</sup> ion concentration of their solutions. Some figures of Winalow and Lochridge (1906) will make this clear Comparing the strengths of HCl and H<sub>2</sub>SO<sub>4</sub> necessary to produce a 99 per cent and a 100 per cent reduction in the numbers of *Bact coli* in 40 minutes, they observed that the disinfectant action of these two acids was in proportion to their degree of dissociation Their results are given in Table 5

TABLE 5

SHOWING PERCENTAGE REDUCTION OF *Bact coli* IN 40 MINUTES BY ACIDS OF DIFFERENT STRENGTHS

	99 per cent Reduction.		100 per cent. Reduction.	
	HCl	H <sub>2</sub> SO <sub>4</sub>	HCl	H <sub>2</sub> SO <sub>4</sub>
Normality	0.0077	0.0096	0.0123	0.0166
Degree of dissociation	97%	80%	96.4%	76%
Parts per million of dissociated hydrogen	7.49	7.68	12.8	12.6

It will be noticed that to cause a 99 per cent. reduction, the strength of HCl required was 0.0077 normal, whereas that of  $H_2SO_4$  was rather greater, 0.0096 normal. But as the degree of dissociation was greater with HCl than with  $H_2SO_4$ , the final concentration of H ions in the two solutions was practically identical.

From this experiment we may, therefore, conclude that the disinfectant action of mineral acids in high dilution is a function of their degree of dissociation, and hence of their resulting H ion concentration. Incidentally, we may notice that a considerably higher concentration of acid is necessary to sterilize a bacterial suspension completely than to reduce its numbers by 99 per cent. This point will be dealt with under the section dealing with the physical factors concerned in disinfection.

The effect of the H ion concentration of the medium on bacteria suspended in it is rather complex. There is, first of all, an optimum concentration for growth, for *Bact. coli* this is about pH 7.6. There is, secondly, an optimum concentration for survival, for *Bact. coli* this is about pH 6.0. Thirdly, there is a point at which the acid tolerance of the organism fails, this for *Bact. coli* is about pH 4.6. During growth in a medium containing a fermentable carbohydrate, *Bact. coli* produces acid, which raises the H ion concentration of the medium to about pH 5.0. This degree of acidity can be well tolerated, but if the acidity is increased beyond this point, instead of continuing to grow, the organisms cease multiplying and rapidly die. And lastly, there is evidence that the H ion concentration most suitable for certain fermentative processes is different from the optimum pH for growth (Cohen and Clark 1919, Gale 1940).

Here, however, we are dealing with the acid tolerance of micro organisms and this limit varies with different species. In Winslow and Lochridge's experiments, already referred to, the parts per million of dissociated hydrogen necessary to sterilize a suspension of *Bact. coli* in 40 minutes were 12.80 to sterilize a suspension of *Salmonella typhi* only 4.85 were required.

Apart, however, from the action of their free H ions certain acids have another disinfectant action on bacteria, which appears to be dependent on the nature of the molecule. To produce a 99 per cent reduction in the number of *Bact. coli* in 40 minutes, Winslow and Lochridge (1906) found that a 0.0812 N solution of acetic acid, or an 0.0097 N solution of benzoic acid was required. The degree of dissociation of each acid at its respective concentration is only about 1 per cent, so that the amount of dissociated hydrogen in the acetic acid was 1.2 parts per million, and in the benzoic acid 0.1 parts per million. It will be remembered, however, that when HCl was used, 7.49 parts per million were necessary. From this it is evident that the toxic action of acetic and of benzoic acid depends on some other factor than their H ion concentration. This other factor must be either the anion or the undissociated molecule. There is some evidence that the bactericidal activity of the monobasic series of organic acids increases with increase in molecular weight and decrease in surface tension, while with the dibasic organic acids the reverse holds true (Reid 1932). Halogenation of the fatty acids is said to increase their germicidal power (Tetsumoto 1937). The subject, however, is complex, and no general statement can yet be made, it will be discussed further in the section dealing with salt action.

**SUMMARY**—(1) The disinfectant action of mineral acids is proportional, not to their normal strength but to the number of free H ions per unit volume.

(2) The organic acids are only slightly dissociated, so that their H ion con-

centration is relatively low. As, however, they have a markedly germicidal effect, it must be concluded that this is a property of the whole molecule or of the anion and is specific for each acid, acetic acid has, for example, only 10-20 per cent of the toxicity of benzoic acid.

(3) Certain other acids, such as fluoric acid and nitric acid, have a specific action, which is probably due to the anion.

**Alkalies**—By similar experiments to those described in the section on acids, Krong and Paul (1897) showed that the disinfectant action of alkalies was dependent on their degree of dissociation and hence on their concentration of OH ions.

Thus of the bases KOH NaOH LiOH, and  $\text{NH}_4\text{OH}$ , KOH shows the highest degree of dissociation and is hence the most actively germicidal,  $\text{NH}_4\text{OH}$  is dissociated the least and is the least actively germicidal (Table 6)

TABLE 6. (Krong and Paul 1897)

DISINFECTION OF ANTHRAX SPORES BY ALKALIES Initial number of spores was about 6 500

	Strength.	Percentage Degree of Dissociation.	No surviving after	
			3 hrs. 20 mins.	15 hrs.
KOH	M/1	77	585	31
NaOH	M/1	72	619	33
LiOH	M/1	64	778	44
$\text{NH}_4\text{OH}$	M/1	0.4	$\infty$	$\infty$

When we turn to other bases we find exceptions. Thus,  $\text{Ba}(\text{OH})_2$  is less dissociated than KOH but is very much more toxic, similarly with the hydroxides of the other alkaline earths. The reason for this, as we shall see in the section on salts, is that the metallic ion is frequently highly toxic, and assists the hydroxyl-ion in its germicidal activities.

Summarizing, we may say that unless a toxic metallic ion is present, the disinfectant action of an alkali is proportional to its degree of dissociation, and hence to its concentration of hydroxyl ions.

Just as bacteria possess a limit of acid tolerance, so they possess a limit of alkali tolerance. Cohen (1922) found that for *Salmonella typhi* this was about pH 8.7. It is of interest to note that H ions appear to be more toxic than OH ions in similar concentration.

**Salt Action.**—Though this chapter primarily concerns the bactericidal action of various physical and chemical agencies it is convenient to introduce here the subject of salt action in general.

We have seen that distilled water cannot be considered a satisfactory medium for bacteria. Many of the vegetative organisms die rapidly in it, and few survive for long. Ficker (1898) was the first to make direct observations on the action of physiological saline on bacteria. His results showed that instead of being harmless it was actively bactericidal. Subsequent workers have confirmed his observations and have demonstrated that the bactericidal effect is due to the toxicity of the sodium ion.

Delepine and Greenwood (1914) working with a number of heavy metals—

copper, silver, zinc, cadmium, mercury—found that, though in certain concentrations they had a strong inhibitory action on bacterial growth in lower concentrations they had the reverse effect, actually stimulating growth Winslow and Hotchkiss (1922) found that the same held true for some of the lighter metals (Table 7)

TABLE 7

EFFECT OF DIFFERENT CONCENTRATIONS OF SALTS ON GROWTH OF *Bact coli* IN 1 PER CENT PEPTONE WATER

	Stimulating	Inhibiting
$\text{CaCl}_2$	0.01 M	0.5 M
$\text{MgCl}_2$	0.05 M	0.5 M
$\text{NH}_4\text{Cl}$	—	1.0 M
$\text{SrCl}_2$	0.1 M	1.0 M
$\text{NaCl}$	0.5 M	3.0 M
KCl	0.5 M	4.0 M

A further point of interest was brought out by Sherman and Holm (1922) who showed not only that NaCl stimulated growth in a concentration of 0.1 to 0.3 M but that it widened the range of H ion concentration within which *Bact coli* would grow. Taking just visible turbidity of the culture as the sign of growth, they obtained results set out in Table 8

TABLE 8

GROWTH OF *Bact coli* IN MEDIA OF DIFFERENT pH IN THE PRESENCE AND ABSENCE OF NaCl

Medium	pH	Turbidity appeared in
P W	5.3	36 hours
P W 0.2 M NaCl	5.3	4 "
P W	8.3	7 "
P W 0.2 M NaCl	8.3	3½ "
P W	4.8	No growth
P W 0.2 M NaCl	4.8	20 hours

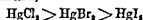
Thus the addition of 0.2 M NaCl to a 1 per cent solution of peptone in distilled water increased the rate of growth at unfavourable pH concentrations and actually enabled the organisms to grow at pH 4.8—a concentration at which in plain peptone water they refused to grow at all

It must be pointed out that a salt which is bactericidal in an aqueous solution may exert a stimulating effect when added in the same concentration to a nutrient medium

Having referred to the favourable action of many weak solutions of salts on the growth and on the survival of bacteria, we must now pass on to consider their toxic action. This is the action which appealed particularly to the early workers. Koch in 1881 drew attention to the toxic action of salts, especially to the salts of the heavy metals such as mercury and silver. There is no doubt that Koch overestimated the germicidal effect of these substances. Geppert (1889) pointed out that if the excess mercury or silver was removed at the end of the test by bubbling  $\text{H}_2\text{S}$  through the suspension, considerably higher concentrations of the

heavy metal were required to destroy the organisms than had been found by Koch. Geppert showed that minute concentrations of heavy metals were sufficient to inhibit growth, but were unable to kill the organisms. Observations by Fildes (1940) have shown that mercury is even less bactericidal than it was thought to be by Geppert. Fildes has brought some evidence to suggest that mercury acts by combining with the  $-SH$  groups of the bacterial cell, which are essential for metabolism. If the mercury is neutralized by the addition of  $-SH$  compounds, like glutathione, cysteine or thioglycollate, bacteria are able to grow after treatment with strong mercury solutions which previous workers have regarded as being actively germicidal. It seems possible, therefore, that mercury, and perhaps other of the heavy metals, act by interfering with essential metabolites of the cell. In virtue of this property its bacteriostatic power is high, but its bactericidal effect is comparatively low. In estimating the bactericidal effect in practice, it is necessary to remove the excess of mercury from the suspension at the end of the test period by treatment with  $H_2S$  or ammonium sulphide, and then to cultivate the organisms in a liquid medium containing 1 per cent thioglycollate in order to provide an adequate concentration of  $-SH$  groups. Records of the germicidal effect of mercury solutions not based on the use of this method must be regarded as unreliable. The same criticism applies to the organic salts of mercury, such as phenyl mercuric nitrate and several proprietary preparations. If thioglycollate is added to the broth used for subculture, none of these substances is able to destroy *Staph aureus* or *Bact coli* in a 1/1,000 dilution in 10 minutes at room temperature (Hoyt, Fisk and Burde 1942).

Kronig and Paul (1897), and later Paul and Prall (1907), made the very important discovery that the toxicity of solutions of  $HgCl_2$  depends not on the molecular concentration of the salt but on the concentration of free  $Hg$  ions in the solution. Thus the halogen salts of mercury were found to be active in proportion to their degree of electrolytic dissociation,



Solutions of salts in which the mercury was combined with a complex anion, and in which the degree of dissociation was poor, such as mercury acetate or cyanide, were found to be much weaker in germicidal power. The behaviour of the salts of the heavy metals is therefore analogous to that of the mineral acids, the toxicity being in proportion to the concentration of free metallic ions and of free  $H$  ions respectively.

The mode of action of heavy metals themselves, as apart from their salts, is not clear. Their toxicity may be demonstrated either by adding them to distilled water, or by placing them, in the form of a bar or coin, on the surface of an inoculated agar plate. Kling (1932) believes that the pure metal goes into actual solution. On the other hand the experiments of Hofmann (1929) and Pilod and Codvelle (1932), both of whom found that oxygen was necessary for the manifestation of toxicity, suggest that an oxide of the metal is formed which then undergoes ionization. Pure metals, of course, cannot ionize, and their failure, whether in aqueous or colloidal solution, to prove toxic under anaerobic conditions, points strongly to the necessity of preliminary salt formation followed by their ionic dissociation.

A vast amount of work has been done on the effect of different salts on bacteria. As the salts of mineral acids are electrolytically dissociated, it is clear that their action may be due either to the undissociated molecule, to the anion, to the cation, or to all three in combination. To assess the importance of each of these factors,

comparative tests have been made with salts of one metal combined with different anions, and of one anion combined with different metals. These tests have been conducted not only on various bacteria but on protozoa, and on the eggs of certain fish. On the whole the results have been reasonably concordant, as may be seen from Table 9, in which the cations are arranged in order of ascending toxicity. It must be understood that strict comparison of the action of different salts can be undertaken only in media of the same H ion concentration.

TABLE 9 (modified from Falk 1923)  
CATIONS IN SERIES OF INCREASING TOXICITY

Eisenberg (1913) Bacteria	Winslow and Hotchkiss (1922) <i>Bact. coli</i>	Woodruff and Bunzel (1909) <i>Paramecium</i>	Mathews (1904 a b) <i>Fundulus</i> Eggs
Na	K	K	Sr
H	Na	Ca	Mg
NH <sub>4</sub>	NH <sub>4</sub>	Zn	Ba
Li	Li	Sr	K
Mg	Sr	Mg	NH <sub>4</sub>
Sr	Mg	Mn	Al
Ca	Ca	Co	Ca
Ba	Ba	Ni	Na
Mn	Mn	Cd	Mo
Ce	Ti	Cu	Li
Th	Sn	Ag	Fe
Fe	Ni	Pb	Ni
Yt	Ti	Fe	Co
Cr	Zn	Hg	Zn
U	Cu		Au
Zn	Fe		Cd
Fe	Fe		Cu
Ti	Co		Fe
Be	Pb		
Al	Al		
Ne	Ce		
Pb	Cd		
Cu	Hg		
Tl			
Zr			
Ni			
Cd			
Co			
Au			
Pt			
Hg			
Ag			

From this table it will be seen that on the whole those metals of low atomic weight are less toxic than those of high atomic weight, though there are many exceptions.

To give some idea of the actual strengths necessary to cause inhibition of growth of *Bact. coli* some results of Hotchkiss (1923) are given in Table 10. She divides her salts into two groups, the more toxic ones comprising those of the heavy metals, and the less toxic comprising those of the alkali metals and of the alkaline earth metals. The salts of Group I give neutral solutions, those of Group II, owing to hydrolysis yield solutions with an acid reaction.



TABLE 10

SALT CONCENTRATIONS THAT LIMIT GROWTH OF *Bact. coli* IN 1 PER CENT PEPTONE WATER  
Incubation period, 3 days. Molar concentration.

GROUP I			GROUP II		
Salt.	No Growth	Growth	Salt.	No Growth.	Growth.
MnCl <sub>2</sub>	0.05	0.025	HgCl <sub>2</sub>	0.00001	0.000005
BaCl <sub>2</sub>	0.25	0.1	CdCl <sub>2</sub>	0.0001	0.00005
—	—	—	CeCl <sub>2</sub>	0.0005	0.0001
CaCl <sub>2</sub>	0.5	0.25	AlCl <sub>3</sub>	0.0005	0.0001
MgCl <sub>2</sub>	0.5	0.25	PbCl <sub>2</sub>	0.0005	0.001
SrCl <sub>2</sub>	1.0	0.25	CoCl <sub>2</sub>	0.0005	0.0001
—	—	—	FeCl <sub>2</sub>	0.001	0.0005
LiCl	0.75	0.5	FeCl <sub>3</sub>	0.001	0.0005
NH <sub>4</sub> Cl	1.0	0.75	CuCl <sub>2</sub>	0.001	0.0005
NaCl	2.0	1.0	ZnCl <sub>2</sub>	0.001	0.0005
KCl	2.0	1.0	NiCl <sub>2</sub>	0.005	0.001
—	—	—	SnCl <sub>2</sub>	0.005	0.001
—	—	—	TiCl <sub>3</sub>	0.005	0.001
—	—	—	TiCl <sub>4</sub>	0.01	0.0025

A further point may be noted from this table, namely, that the bivalent cations tend to be more toxic than the monovalent cations.

Very much less work has been done on the effect of *anions* on the growth of bacteria. Falk (1923) points out that the anions play an essentially different part in metabolism from the cations. The former are intimately related to the nutritive metabolism—particularly the anions that contain carbon, sulphur, nitrogen or oxygen—whereas the latter are concerned with the regulative metabolism of the organism. Nevertheless, the anions in certain concentrations do undoubtedly possess a toxic action on bacteria. Some figures of Holm and Sherman's (1921) will exemplify this point. They grew *Bact. coli* in 1 per cent peptone water, to which were added various sodium salts, the H ion concentration being kept practically constant, and compared the rate of growth in the different tubes (Table 11).

TABLE 11

		pH	First Turbidity appeared in
1 per cent peptone		7.2	4½ hours
"	0.2 M NaCl	7.3	3½ "
"	0.2 M NaI	7.3	3½ "
"	0.2 M NaNO <sub>3</sub>	7.3	3½ "
"	0.2 M Na <sub>2</sub> SO <sub>4</sub>	7.0	4 "
"	0.2 M (mixture of NaH <sub>2</sub> PO <sub>4</sub> and Na <sub>2</sub> HPO <sub>4</sub> )	7.3	4½ "
"	0.2 M Na lactate	7.0	4½ "
"	0.2 M Na oxalate	7.0	9½ "
"	0.2 M Na acetate	7.0	10½ "
"	0.2 M Na citrate	7.3	10½ "
"	0.2 M NaF	7.4	48 "

From this it will be seen that the Cl ion was the least, and the F ion the most toxic

One of the most extensive studies is that of Eisenberg (1919), who arranges the anions in order of toxicity thus  $\text{SO}_4 < \text{S}_2\text{O}_3 < \text{Tartrate} < \text{H}_2\text{PO}_4 < \text{MoO}_4 < \text{Cl} < \text{Br} < \text{NO}_3 < \text{SO}_3 < \text{Fe}(\text{CN})_6''' < \text{Acetate} < \text{ClO}_3 < \text{Citrate} < \text{HPO}_3 < \text{Oxalate} < \text{Formate} < \text{CNS} < \text{ClO}_4 < \text{BrO}_3 < \text{I} < \text{H}_2\text{PO}_4 < \text{Benzoate} < \text{Nitroprusside} < \text{HAsO}_4 < \text{CrO}_4 < \text{P}_2\text{O}_7 < \text{NO}_2 < \text{F} < \text{BF}_4 < \text{HF} < \text{BO}_3 < \text{B}_3\text{O}_6 < \text{Fe}(\text{CN})_6''' < \text{Salicylate} < \text{HSeO}_4 < \text{IO}_3 < \text{S}_2\text{O}_8 < \text{S}_2\text{O}_7 < \text{TeO}_4 < \text{SbS}_4 < \text{OsO}_4 < \text{IO}_4 < \text{Cr}_2\text{O}_7 < \text{TeO}_3$

The action of salts depends to a large extent on the medium in which they are dissolved, thus they are more active when dissolved in distilled water than when dissolved in a medium containing protein. This is an observation that has been made frequently (Behring 1890, Krong and Paul 1897, Chick and Martin 1908). Probably it is due to the fact that many cations combine with proteins to form an insoluble albuminate, hence the concentration of free ions in the medium is diminished.

Another important observation is that different bacteria vary in susceptibility to the same salt. Eisler (1909) found that *B. subtilis* was killed by N/10 LiCl whereas the El Tor vibrio was unharmed by N/5 LiCl. Eisenberg (1919) found that *B. anthracis* possesses more than the average resistance to fluorides, iodates and oxalates. *C. diphtheriae* to tellurates tellurites, Ni and Cu. *Salm typhi* to Sr salts, the pneumococcus to ferriocyanides and tellurites, and *V. cholerae* to chlorates and perchlorates. Certain organisms may be grouped together as having a similar susceptibility to the action of salts, thus, *Staphylococcus pyogenes* and *Staphylococcus candicans*, *C. diphtheriae* and the diphtheroid bacilli. *Salm typhi* and *Bact. coli*, *Chromo prodigiosum* and *Chromo kielsense* are grouped in pairs each member of the pair exhibiting a similar susceptibility to different salts.

As well as this relationship, however, there is a difference in resistance exhibited between members of the Gram positive and the Gram negative group of organisms. Eisenberg found that many salts are more toxic to the Gram positive than to the Gram negative bacteria. This holds not merely for particular salts, but for their constituent anions and cations. On the other hand, some salts, such as potassium tellurite (Fleming 1932, Fleming and Young 1940) and sodium azide (Snyder and Lichstein 1940, Mallmann *et al* 1941), are more toxic to Gram negative than to Gram positive bacteria. This property is now made use of in the preparation of selective media.

The difference in susceptibility to sodium chloride has been suggested by Schoop (1935) as a criterion for bacterial classification. He divides bacteria into three classes (1) those that grow in ordinary media but not in media containing 10 per cent NaCl—non halophiles, (2) those that grow in both media—facultative halophiles, (3) those that grow only in media containing 10 per cent NaCl—obligatory halophiles. The last group of organisms are found mainly in sea water, and in sand and mud adjacent to the sea.

**Antagonistic Effect of Salts**—Hitherto we have been considering the effect on bacteria of solutions containing one salt, we must now examine the effect of solutions containing more than one salt.

Flexner (1907) found that an 0.85 per cent solution of NaCl caused rapid disintegration of the meningococcus but that when a calcium salt was added to the solution, this disintegration no longer occurred. The conclusion he drew was that NaCl by itself is toxic to the meningococcus, but that its toxic action

can be neutralized by a salt of calcium. Students of physiology will recall the similar observations made by Ringer on heart muscle in 1880. Shearer (1919) found that living bacteria offered a considerable resistance to the passage of an electric current, depending apparently on the relative impermeability of the cell membrane. Using, therefore, electrical conductivity as his criterion of viability, he obtained evidence suggesting that a 0.85 per cent solution of NaCl was toxic to the meningococcus, but that this toxic action could be neutralized by the addition of a trace of  $\text{CaCl}_2$  or other bivalent salt. On the other hand, it appeared doubtful whether the toxic action of a bivalent could be neutralized by the addition of a monovalent salt.

Similar results have been obtained by other workers. Eisler (1909) found that the inhibitory action of LiCl on *B. subtilis* could be counteracted by the addition of a divalent, but not of a monovalent salt. Thus, N/10 LiCl was counteracted by N/20  $\text{CaCl}_2$ , by N/200 BaCl<sub>2</sub>, or by N/200  $\text{MgSO}_4$ . Further, he showed that the inhibitory effect of a divalent salt could be counteracted by either a mono or a divalent salt. Thus, N/750  $\text{MnSO}_4$  was counteracted by N/200  $\text{Ca}(\text{NO}_3)_2$ , and by N/100 KCl. This latter conclusion differs from Shearer's.

It must not be thought that the mere addition of a divalent to a monovalent salt will render the solution favourable, the two salts must be present in definite proportions. If not instead of being harmless to the organism, the solution may be actively toxic. Thus Winslow and Falk (1923) found that 0.145 M solution of  $\text{CaCl}_2$  mixed with a solution of NaCl of two or three times this strength was highly toxic to *Bact. coli*. As the proportion of NaCl was increased to four times the strength of the  $\text{CaCl}_2$  solution the toxicity of the solution diminished very markedly. That is, a solution of 0.145 M  $\text{CaCl}_2$  + 0.290 M NaCl was toxic, a solution of 0.145 M  $\text{CaCl}_2$  + 0.680 M NaCl was non toxic. A further increase of NaCl rendered the solution again toxic.

This antagonistic effect of salts brings us to the conception of a balanced solution. A balanced solution is one in which the proportion of the different salts is so ordered that their individually toxic effects are neutralized. In such a solution bacteria are able to survive very much longer than in a solution of any one of the constituent salts. Ringer's solution is of this type, and has the following composition:

NaCl	0.9 gm.
KCl	0.042 gm.
$\text{CaCl}_2$	0.048 gm.
$\text{NaHCO}_3$	0.02 gm.
Glass-distilled water	100.000 ml.

Winslow and Dolloff (1928) have, however, drawn attention to a possible fallacy in the interpretation of the antagonistic effect of salts. According to them, all cations appear to stimulate growth in a certain low concentration and to inhibit it in a certain higher concentration. So far as viability is concerned, therefore they would postulate an optimum ionic concentration for each organism, depending probably upon an alteration in the permeability of the cell wall. They would explain the apparently antagonistic effects of monovalent and divalent salts as being due not to a qualitative antagonism between the two cations, but to the production in the suspension of a more favourable ionic concentration for the survival of the bacteria. In support of this they quote experiments in which the toxic action of a given salt in dilute solution has been annulled by increasing the con

centration of the same salt. How far this explanation is of general applicability, it is as yet impossible to say.

The interaction of various salts is of considerable importance in disinfection. The germicidal action of any one salt may be increased or diminished by the addition of any other. For example, the addition of NaCl is said to lower the toxicity of  $\text{HgCl}_2$ , but to increase the toxicity of mercuric nitrate, sulphate, or acetate. In estimating, therefore, the toxicity of a salt, the saline content of the solution in which it is acting must be defined. Norton and Hsu (1916) showed that salts were able to modify the germicidal power of acids. When ammonium formate was added to formic acid, the H ion concentration of the solution decreased as a result of an increase in the concentration of undissociated acid molecules and its disinfectant power was lowered, when sodium nitrate and sodium chloride were added in very small quantities to formic acid the degree of dissociation of the acid was hardly affected, but its disinfectant power was considerably increased. They conclude therefore that the addition to an acid of a salt containing an anion common to this acid diminishes its disinfectant power, the addition of a salt which does not have any appreciable effect on the dissociation of the acid greatly increases its disinfectant power.

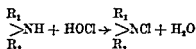
Not only do salts assist or antagonize the action of each other, they have a similar effect on disinfectants of quite different chemical constitution. Scheurlen (1895) showed, for instance, that the addition of sodium chloride in a concentration of 24 per cent to a solution of phenol increased its disinfectant power. Beckman (1896) confirmed this, and found that with staphylococci the addition of even 1 per cent NaCl to 1 per cent phenol apparently increased its activity. With anthrax spores the addition of 6 per cent NaCl to 1 per cent phenol had no effect. 12 per cent NaCl increased its activity slightly, and 24 per cent NaCl increased its activity very greatly. Thus 1 per cent phenol alone failed to kill a suspension of 24 800 000 spores in 8 days, 1 per cent phenol + 24 per cent NaCl killed them completely in between 5 and 24 hours. Romer (1898) confirmed the work of Beckman, showing that the greater the amount of salt added, the greater was the increase in disinfectant power. As a rule, the more toxic a salt is in itself, the more does it supplement the action of the disinfectant (Eisenberg and Okolska 1913).

**Mode of Action of Salts**—In endeavouring to explain the action of salts on bacteria, we must remember that we are dealing with a complex problem of which there is no simple solution. Many factors are concerned, and the most we can do here is to discuss the most important in turn.

(1) *The Osmotic Effect*—It is doubtful whether salts, except in high concentrations, exert any influence on bacteria by virtue of their osmotic pressure. Bacteria differ in this way from practically all other living cells. Thus Fischer (1900) observed that *B. subtilis* grew well in an infusion containing 9 per cent NaCl, 11 per cent KCl, or 10 per cent  $\text{KNO}_3$ , and Knaysi (1930b) found that to demonstrate plasmolysis in this organism a 25 per cent solution of NaCl was required. Though salts have little direct osmotic action on bacteria, they may exert an indirect action by causing a dehydration of the proteins on which the organisms are growing. It is this dehydrating action of salts which is relied on in many processes of food preservation.

(2) *Oxidation*—Salts and certain allied bodies that contain a high proportion of oxygen, or that are able to liberate oxygen from other compounds, have long

been known to be highly germicidal Kröning and Paul (1897) compared the disinfectant activity of certain oxidizing agents with their oxidative capacity, as measured by the method of electrical oxidation chains According to this method, oxidizing agents are arranged in order of decreasing oxidizing capacity thus  $\text{HNO}_3$ , dichromic acid, chloric acid,  $\text{Cl}_2$ ,  $\text{H}_2\text{S}_2\text{O}_8$ , and permanganic acid. This order was, with the exception of chlorine, the same as that of the germicidal action of these substances Chlorine, bromine and iodine were found to be germicidal in inverse order to their atomic weight Their action appears to depend on the liberation of nascent oxygen Ozone is another powerful oxidizing agent, like wise  $\text{H}_2\text{O}_2$ , a 3 per cent solution of which kills anthrax spores in an hour One of the most commonly used of this group of chemical substances is  $\text{K}_2\text{Mn}_2\text{O}_8$ , like  $(\text{NH}_4)_2\text{S}_2\text{O}_8$ , its action is increased by the presence of  $\text{HCl}$  Kröning and Paul prepared a mixture containing 1 per cent  $\text{K}_2\text{Mn}_2\text{O}_8$  and 11 per cent  $\text{HCl}$  dissolved in water, and found that it would kill anthrax spores in 30 seconds A similar mixture containing 37 per cent  $(\text{NH}_4)_2\text{S}_2\text{O}_8$  and 11 per cent  $\text{HCl}$  was found by Andrewes and Orton (1904) to exercise an effect very nearly as powerful Both these are extremely potent, but even more potent is  $\text{HOCl}$ , which in a concentration of 0.01 per cent kills anthrax spores in 30 seconds So far as activity is concerned, this is one of the most powerful germicides we know Bleaching powder acts by virtue of its ability, when acted upon by weak acids such as  $\text{H}_2\text{CO}_3$ , to yield nascent oxygen, which then combines to form  $\text{HOCl}$   $\text{HOCl}$  combines with organic substances containing the  $=\text{NH}$  group, to form chloramines



It is found that all bodies containing the  $\text{NCl}$  group are strongly antiseptic (Dakin 1915)

(3) *Reduction*—Certain salts, such as the sulphites and the ferrous compounds, appear to act by virtue of their reducing power Apart from such salts there are other substances that act mainly as reducing agents—sulphurous acid and formaldehyde A 5 per cent solution of formaldehyde, i.e. a 1-8 dilution of the commercial formalin, kills anthrax spores in between 1 and 2 hours

(4) *Molecular Action*—In a previous section we saw that certain acids, such as acetic and benzoic, and in fact most of the organic acids, act not by virtue of their  $\text{H}$  ion concentration but by virtue of the undissociated molecule Benzoic acid is dissociated very slightly, and its strong disinfectant power must therefore be attributed to the benzoate anion or to the undissociated molecule Probably the same explanation will account for the action of the salts of the organic acids The combination of mercury salts with the  $-\text{SH}$  compounds of bacteria described by Fildes (see p. 122) is regarded by Albert (1942) as an example of molecular action

(5) *Ionic Action*—It is clear that salts which are freely dissociated in solution owe their germicidal power to the action of the ions into which they are dissociated The way in which these ions act is a matter for speculation Bayliss (1924) points out that there are three ways in which electrolytes may exert their influence on living matter (a) They may produce effects through the electrical charges that they bear, this is specially marked with ions with valencies above

one, and bears no relation to the chemical nature of the ions. Thus the effect of  $\text{Ca}^{+}$  cannot be distinguished from Ba. These effects, especially in the case of the multivalent ions, are manifest even in very dilute solutions. (b) They may affect the nature of the solvent in which they are dissolved—the so called lyotropic effect. This action has been studied by several workers, prominent among whom are Hofmeister (1888, 1889) and Freundlich (1903). Hofmeister, working with a number of neutral salts, found that these could be arranged in a definite order relative to their action on the coagulation of colloids, and on other physical properties of proteins. Freundlich, studying the effect of electrolytes on the compressibility, surface tension, solubility, viscosity, and other properties of proteins, was likewise able to arrange them in a definite order. He concluded that the main effect was exerted not on the proteins directly but upon the solvent, modifications in the solvent thus affected the proteins. For this reason he spoke of the salt effects as 'lyotropic' effects, his series of salts hence bears the name "lyotropic series" in distinction to the 'Hofmeister series'. Both series, however, are similar in many respects. Holm and Sherman (1921) and numerous other workers have found a general concordance between the ionic series of Hofmeister or of Freundlich and the stimulative or toxic action of the salts of these series. (c) They may operate through a specific influence which is more intimately connected with the chemical properties of the ions. Thus sodium and potassium, though having the same electrical charge and exercising very similar lyotropic effects, are yet totally different in their action on heart muscle. As this action is shown by solutions so dilute that undissociated molecules are nearly absent, we know that this difference must be attributed to some specific or chemical property of the ions.

Other explanations of ionic action have been put forward, some of them modifications of the ones already given. Amongst these may be mentioned Loeb's (1899, 1900) hypothesis of ion protein combination, Mathews' (1904a, b, 1905, 1906) conception of ionic potential, and Zwaardemaker's (1918, 1919-20) radioactivity hypothesis. For these and for further information on this subject the reader is referred to an admirable summary by Falk (1923)—a summary which has been freely drawn on in this section.

There is one further point. We have treated the action of anions and of cations separately, whether both are sometimes required simultaneously for the production of the bactericidal effect is not at all clear. It seems probable that in a salt such as  $\text{HgCl}_2$ , in which a toxic cation is united to a weakly toxic anion, almost the entire action of the salt must be referred to the Hg ion. In a salt such as  $\text{K}_2\text{Mn}_2\text{O}_8$ , on the other hand, in which a weakly toxic cation is united to a powerful anion, the action must be referred to the permanganate ion.

#### SUMMARY OF SALT ACTION

(1) There is a certain concentration for nearly all salts which stimulates bacterial growth, this concentration is generally very low.

(2) There is, for nearly all salts, a limit beyond which the stimulating action passes over into a toxic action, on the whole, the higher the concentration, the more evident does the toxic action become.

(3) The toxic effect of univalent salts can be neutralized by the addition in suitable proportions of a divalent salt. In most instances, too, it is possible for a univalent salt to neutralize the toxic action of a bivalent salt. This action is known as the antagonistic action of salts.

(4) Electrolytes with bivalent cations are generally more toxic than those with univalent cations. Thus Ba is more powerful than Na.

(5) On the whole the salts of the heavier metals are more toxic than those of the lighter metals. Thus  $\text{HgCl}_2$  is more toxic than  $\text{CaCl}_2$ . But there is no strict quantitative relation between the atomic weight of a metal and its toxicity.

(6) On the other hand there is a fairly close relationship between the lyotropic and the toxic effects of a salt.

(7) The toxic action of salts is less marked in protein solutions than in distilled water. Thus the activity of  $\text{HgCl}_2$  is decreased markedly in the presence of blood serum.

(8) The more favourable the nutrient qualities of the medium in which the bacteria are suspended the less manifest is the toxic effect of salts and of germicidal agents in general upon them.

(9) Different organisms vary in their susceptibility to the disinfectant action of the same salt. Closely allied organisms respond in much the same way to the same salts.

(10) There is evidence that the Gram positive organisms with a few exceptions are more susceptible to the disinfectant action of salts than the Gram negative organisms.

(11) The addition of a salt to a solution of a germicide—whether itself a salt or not—may increase or decrease the action of the latter. This action may be due partly to the effect on the electrolytic dissociation of the germicide, partly in a colloidal solution to an effect on the dispersion coefficient of the disinfectant, and partly perhaps to the disinfectant action of the salt itself.

(12) There is little evidence that salts except in high concentrations owe their germicidal action to the osmotic pressure that they exert, since bacteria are strongly resistant to variations of osmotic pressure, but they may act by dehydrating the proteins of the medium in which they are suspended.

(13) The action of salts is complex. It may be referred to an oxidation effect, a reduction effect, a molecular effect or an ionic effect. Other effects, namely the sensitization of organisms to  $\text{CO}_2$  and their interference with proteolytic enzymes, have not been considered in this chapter. For details of these the reader is referred to an article by Rockwell and Ebertz (1924).

**Soaps and Synthetic Detergents**—Several workers have studied the germicidal effect of soaps with results that have been at times contradictory. Many of the discrepancies can be ascribed to the use of different test organisms, since certain soaps are highly bactericidal to some organisms and comparatively inert to others. For example, pneumococci are very sensitive to the soaps of the unsaturated fatty acids—oleic, linoleic, linolenic—but much less so to soaps of the saturated fatty acids—stearic, palmitic, myristic, lauric. According to Lamar (1911) virulent pneumococci are killed by a 0.5 per cent solution of sodium oleate in 15–30 minutes. Bayliss (1936) found that to kill pneumococci in 10 minutes a 0.1 per cent solution of sodium palmitate was required, but only a 0.001 per cent solution of sodium oleate. Lamar (1911) also noticed that sodium oleate even in high dilution such as 1/20,000 greatly accelerated the autolysis of pneumococci and favoured their lysis by normal or immune serum. Haemolytic streptococci, meningococci, gonococci and diphtheria bacilli resemble pneumococci in their greater sensitivity to soaps of the unsaturated fatty acids. On the contrary the Gram negative bacilli of the coliform group are fairly susceptible to soaps of the saturated fatty

acids, but are resistant to soaps of the unsaturated fatty acid series (Reichenbach 1908, Walker 1924, 1925, 1926, Belin and Ripert 1937). Staphylococci are resistant to all the common soaps in neutral and alkaline solution (Walker 1924, Bayliss 1936), but according to Eggerth (1926) they are susceptible at pH 5. The most generally useful soap in practice is sodium laurate, since it acts on pneumococci, streptococci, and typhoid bacilli, though not on staphylococci (Walker 1924). There is a suggestion that the germicidal activity of soaps increases with increase in molecular weight in the saturated fatty acid series but decreases in the unsaturated fatty acid series (Walker 1924, Bayliss 1936). Of commercial soaps Nichols (1920) found yellow or brown bar soap, such as is used in washing dishes to be effective in a 1:200 concentration in killing pneumococci and streptococci (see also Colebrook and Maxted 1933). Soap prepared from coconut oil such as salt water soap, is more germicidal than any other soap to the typhoid bacillus (Hamilton 1917, Walker 1925, 1926). If a stiff lather is made on the hands over *Bact. coli* is killed within a minute. The germicidal effect of soaps is increased by rise in temperature (Walker 1924).

The mode of action of soaps in destroying bacteria is far from clear. It is certainly not due entirely to free alkali, since this may be present in much too small an amount to have any deleterious effect at all.

Reichenbach (1908), however, thought that alkali might play a part in some soaps. He observed, for example, that with salts of the higher fatty acids the germicidal effect decreased much more slowly on progressive dilution than with salts of the lower fatty acids, and explained this by the greater hydrolysis of the former group with the consequent liberation of free alkali. Eggerth (1926) found that generally speaking soaps of the lower members of the fatty acid series were more active in acid solution, the higher members in alkaline, the point of transition varied with the test organism. He explained this result in terms of the effect of the pH on the dissociation residue and on the solubility of the soap. Lamar (1911) is of the opinion that the soap acts on the lipoidal moiety of the cells rendering them more permeable to germicidal substances in the solution. This would explain the adjuvant effect of soap on bacterial lysis by serum or by substances such as aromatic oils which are often added to commercial soap and would presuppose a germicidal effect of the soap itself.

Summarising, we may say that soaps show a strongly selective action towards bacteria, most of the pathogenic respiratory organisms being killed more readily by soaps of the unsaturated fatty acids, and most of the pathogenic intestinal organisms more readily by soaps of the saturated fatty acids. In practice, thorough washing of the hands in a stiff lather with a minimum amount of hot water, preferably using yellow bar household soap, can be relied upon to kill a high proportion of pathogenic organisms on the hands, with the exception of *Staphylococcus aureus*.

It is possible, as Noguchi (1907) suggests, that soaps in blood and lymph are responsible for some part of the natural defence mechanism of the body, since, in his experience, mixtures of soap and inactivated serum resembled complement in many respects. Burtenshaw (1942) likewise suggests that soaps and long-chain fatty acids are mainly concerned in the auto-disinfecting action of the skin. (For a study of soap derivatives, see Eggerth 1929a, b, 1931, and for a review of "germicidal" soaps, see Morton and Klauder 1914).

During recent years a large group of synthetic detergents have been used in industry. Some of these substances are highly bactericidal and are finding a place in surgery for the cleansing of skin and other surfaces. They are classified



into cationic and anionic detergents, according to whether the location of the long chain hydrophobic group is in the cationic or anionic portion of the molecule. Cetyltrimethylammonium bromide, for example is a cationic detergent, sodium cetyl sulphate an anionic detergent. Generally speaking these substances are good wetting and cleaning agents, are relatively non irritant to raw surfaces and destroy vegetative bacteria in dilutions varying from 1:100 to 1:16 000 or so. Some are toxic to leucocytes and others precipitate proteins. Their bactericidal action is often greatly diminished by the presence of organic matter and of phospholipins, their penetrating power is usually low, and some organisms like *Ps. pyocyanea*, may prove very resistant to them. On the whole, the cationic group appears to be more germicidal than the anionic group, and Gram positive are more affected than Gram negative bacteria (see Miller and Baker 1940, Baker *et al.* 1911a, b, Barnes 1912, Hoyt *et al.* 1912, Williams *et al.* 1943, Iland 1944).

**Alcohols and Ethers**—Epstein (1897) found that absolute ethyl alcohol was not a germicide, but that when diluted it became germicidal. Minervini (1898) confirmed this, and showed in addition that alcohol had little or no action on spores. For the destruction of vegetative bacteria the optimal strength depends on the degree of moisture present. A final concentration of 50–70 per cent. appears to be most effective. Thus, an equal amount of absolute alcohol should be added to an aqueous suspension of bacteria, whereas for dry bacteria a solution of alcohol already diluted to 50–60 per cent. should be used. For the disinfection of moist hands 80–96 per cent. alcohol is recommended, for the disinfection of dry hands 70–80 per cent. alcohol is better. Dry vegetative bacteria are destroyed less rapidly than moist—presumably because the penetration of alcohol takes longer (Table 12).

TABLE 12

TIME TAKEN BY DIFFERENT STRENGTHS OF ETHYL ALCOHOL TO DESTROY DRY AND MOIST STAPHYLOCOCCI  
(After Russ 1904)

Alcohol per cent.	Dry Cocci.	Moist Cocci.	Alcohol per cent.	Dry Cocci.	Moist Cocci.
98.8	>24 hrs.	1 min.	40	5 mins	0 mins
80	60 mins	1 min.	30	10 "	>60 "
70	5 "	1 min.	20	60 "	>60 "
60	5 "	1 min.	10	Not tried	>60 "
50	1 min.	1 min.	—	—	—

Note.—The dry staphylococci were dried on silk threads, the moist staphylococci were in aqueous suspension.

The presence of protein increases the disinfection time of alcohol but not to any considerable extent. The addition of a dilute mineral acid or alkali greatly increases its activity, enabling alcohol to kill spores. Thus Coulthard and Sykes (1936) found that a solution of 70 per cent. alcohol containing 1 per cent. sulphuric acid destroyed spores of *B. subtilis* in less than 24 hours, and a solution of 70 per cent. alcohol containing 1 per cent. sodium hydroxide in 24–48 hours. Alcohol lowers the germicidal effect of some substances, like the heavy metal salts, phenol and formaldehyde that are dissolved in it (Koch 1881, Krönig and Paul 1897) but is said to raise the germicidal effect of others, such as iodine. In fact, a strong

tincture of iodine—4.5 per cent iodine in 70 per cent alcohol, together with 2 per cent potassium iodide to stabilize the iodine—is one of the best skin disinfectants known. Though the bactericidal activity of alcohol is negligible below 10–20 per cent, it may prove bacteriostatic to many organisms in concentrations as low as 1 per cent (Wirgin 1902). As an antiseptic for the preservation of vaccines 25 per cent alcohol has been found to be rather more potent than 0.5 per cent phenol (see Cruickshank *et al* 1942). Commercial alcohol as a rule contains spores so that for surgical or biological use it should be filtered through a Berkefeld or similar candle (not a Sertiz, which is effective only in the presence of water) or distilled.

Ritchie (1899) showed that the germicidal action of different alcohols increased with their molecular weight, ethyl alcohol being more potent than methyl propyl than ethyl, and butyl than propyl alcohol. This has been confirmed by subsequent workers (Wirgin 1904, Tilley and Schaffer 1926, Tilley 1939, Lockemann, Baran and Totzeck 1941). For disinfection of the skin 80 per cent propyl alcohol is particularly useful, it is more bactericidal than ethyl alcohol, it is a better fat solvent and it is not so volatile. (For a detailed review of the disinfectant action of alcohol see Sobernheim 1943, and for its value as a hand disinfectant see Ahlfeld and Vahle 1896, Neufeld and Schiemann 1913).

The ethers are possessed of some degree of germicidal activity. Cultures of non sporing bacteria incubated in an atmosphere saturated with the vapour of diethyl ether— $C_2H_5OC_2H_5$ —exhibited no growth, subcultures showed that the organisms had been killed in a period varying from about 1 to 48 hours (Topley 1915). Direct immersion of *Bact. coli* in 50 per cent ether proved fatal in about 3 minutes at room temperature. On the other hand, exposure of *Cl. septicum* to pure ether failed to destroy the spores in 24 hours. According to Kronig and Paul (1897) ethereal solutions of disinfectants are almost without effect on anthrax spores.

**Phenols and Cresols**—Under this heading we shall consider the action of those bodies that are obtained from the destructive distillation of coal, and that pass over between the temperatures of 170° and 270° C. Phenol itself in certain proportions is able to pass into solution in water, but most of the bodies in this group do not do so; when mixed with water they form emulsions of varying degrees of fineness. Their mode of action is therefore different from the action of the germicides which we have so far considered. The phenols and cresols have a fairly high germicidal activity when employed in solutions above a given concentration, but it requires quite a low degree of dilution to deprive them entirely of this activity. In this respect they differ markedly from the saline disinfectants (see p. 143).

It has been supposed that phenol acts by its formation in contact with proteins of an insoluble albuminate and of other chemical compounds. Reichel (1909), however, who studied the dispersion phases of phenol between oil and water, brought evidence to suggest that the action is not so much chemical as physical, the phenol being capable of passing into solution in such substances as coagulated albumin, certain lipins, and the cytoplasm of bacteria. He suggests therefore, that its disinfectant action results from its penetration into the bacterial cell in the form of a colloidal solution.

The emulsified disinfectants, such as the cresols, probably act in much the same way as phenol, but their germicidal activity is usually somewhat higher. By virtue of their emulsoid state, their particles are adsorbed on to the surface of

suspended matter, and hence their concentration is increased in the immediate neighbourhood of the bacteria. This action is interfered with by the presence of other suspended organic matter which serves to adsorb the germicide and thus lower its effective concentration around the bacteria. Emulsoids of the cresol group are generally most active when freshly made up in solution, after a day or two probably because of an alteration in their colloidal state their activity diminishes. Some of the cresols can be employed in true solution but their solubility in water is very low. Para-chlor meta-cresol for example has a solubility of about 1/300 towards naked bacteria it is approximately ten times as active as phenol (see Withell 1912a)

According to Klarmann Shternov and Gates (1934a b) the germicidal activity of phenol derivatives is increased by halogen substitution and is still further intensified by the introduction of aliphatic or aromatic groups into the nucleus of these compounds. Their general formulæ are



para-Chlorophenol Derivative



ortho-Chlorophenol Derivative

where R is an aliphatic or aromatic group. Some of the compounds tested by these workers, such as para-chlorophenol or ortho chlorophenol derivatives with a butyl to n-octyl substituents, proved highly destructive to bacteria while being comparatively non toxic to mice on subcutaneous injection.

The *chloroxylenols* many of which form a clear solution in water have come into prominence of late years mainly for skin disinfection. They are comparatively non irritant but their bactericidal power on the whole, is considerably less than that of phenol unless employed in 30-50 or even 100 per cent. concentration they cannot be relied upon to destroy staphylococci on the skin (see Colebrook 1941) (For useful information on the use of tar derivatives in practice see Report 1942-1944)

**Dyes**—Though a few desultory observations had been made at various times on the effects of aniline dyes on bacteria Churchman (1912) was the first to investigate them thoroughly. Working with gentian violet, he found that if 5 drops of a saturated aqueous solution of this dye were added to broth cultures of different organisms the mixtures allowed to remain for an hour and transplants then made on to agar the Gram negative organisms grew satisfactorily, but the Gram positive organisms failed to develop. A similar selective property could be demonstrated by seeding the fresh unstained organisms on to plates one-half of which contained plain nutrient agar, and the other half nutrient agar containing a dilution of about 1/100 000 gentian violet. A large number of different bacteria were tested to ascertain if there was a perfect correlation between Gram positive-ness and inability to grow in media containing gentian violet. This was found not to be the case about 90 per cent of the Gram positive organisms were killed by gentian violet and failed to grow on media containing it, but the remaining 10 per cent, comprising the acid fast group in particular, were not affected. Similarly though about 90 per cent of the Gram negative organisms were resistant, the remaining 10 per cent were susceptible.

The difference between the Gram positive and the Gram negative organisms is merely one of degree. There is moreover, a considerable variation in the susceptibility of different species of Gram positive bacteria. Garrod (1933a) has shown, for example, that staphylococci are much less resistant to the violet dyes—crystal violet, methyl violet, Hofmann violet, gentian violet, Dahlia—than streptococci. The presence of 1/1 000 000 gentian violet in nutrient broth or in 5 per cent serum broth is sufficient to inhibit the growth of staphylococci, while streptococci can grow in the presence of 1/250 000, and sometimes even stronger concentrations of this dye.

Churchman (1923a) stated that, just as gentian violet had a bacteriostatic effect on most Gram positive organisms, so acid fuchsin had a similar effect on Gram negative organisms. Garrod (1933b) has recently examined this statement and concluded that it is untrue. He finds that aniline dyes generally, whether of the basic or acid type, destroy Gram positive more readily than Gram negative bacteria. On the other hand, Churchman's results gain some support from the work of Stearn and Stearn (1926, 1928). From a study of the reactions of different bacteria to different stains, these workers conclude that Gram positive bacteria have a lower isoelectric point than Gram negative bacteria. Hence Gram positive bacteria combine more actively with basic, and Gram negative with acid dyes. The subject clearly needs further investigation.

The aniline dyes have, on account of their marked germicidal effect on bacteria, been used for the treatment of wounds. Browning and his colleagues (1917) recommended flavine—diamino methyl acridinium chloride. Though they found that brilliant green sulphate, malachite green, crystal violet and flavine strongly inhibited the growth of staphylococci and *Bact. coli*, flavine was the only one that was more active in the presence of serum (Table 13). Churchman (1923b) used a mixture of gentian violet and acriflavine.

TABLE 13 (modified from Browning *et al.* 1917)

SHOWING CONCENTRATIONS OF DIFFERENT SUBSTANCES NECESSARY TO INHIBIT THE GROWTH OF *Staphylococcus aureus* AND *Bact. coli*

Substance	<i>Staph. aureus</i>		<i>Bact. coli</i>	
	Conc. in P.W.	Conc. in Serum.	Conc. in P.W.	Conc. in Serum.
Chloramine T	1/2 000	1/250	1/2 000	1/50
Cl <sub>2</sub> water	1/2 500	>1/1 000 *	1/2 500	>1/1 000 *
Phenol	1/250	1/250	1/500	1/500
HgCl <sub>2</sub>	1/1 000 000	1/10 000	1/1 000 000	1/10 000
Brilliant green sulphate	1/10 000 000	1/30 000	1/130 000	1/3 500
Malachite green	1/10 000 000	1/40 000	1/20 000	1/1 000
Crystal violet	1/4 000 000	1/400 000	1/8 000	1/8 000
Flavine	1/20 000	1/200 000	1/1,300	1/100 000

\* These concentrations were insufficient to prevent growth.

Table 13 is of interest in showing not only that flavine is much stronger in its inhibiting action in ox serum than in peptone water, but that HgCl<sub>2</sub> and the chlorine group of germicides are markedly diminished in activity in the presence of serum, whereas phenol remains unaffected. Thus diminution of activity in the presence of organic matter will be referred to later.

Hitherto the dye treatment of wounds has not fulfilled the expectations of its advocates probably because the dyes can seldom be present continuously in sufficient concentration in every part of the wound to inhibit bacterial growth completely. As Browning (1933) points out the destruction of organisms in the centre of masses of necrotic tissue or blood clot is probably beyond the power of any disinfectant. Moreover many of the dyes inhibit leucocytic activity and cause damage to the tissues (see Fleming 1940 Russell and Falconer 1941 1943 Manifold 1941 Rubbo *et al* 1942 Russell and Beck 1944). They may however have some value in the temporary prevention or control of infection (see McIntosh and Selbie 1942 Browning 1943).

**Essential Oils**—Chamberland (1887) tested the disinfectant action of a large number of essential oils by exposing anthrax spores and anthrax bacilli to their vapours in closed tubes. After 4 days exposure at 37° C only one oil was successful in killing the spores—namely oil of Ceylon cinnamon. Anthrax bacilli, contained in blood were killed by oil of vespetro in 18 hours at 37° C in 40 hours by oil of angelica and in 65 hours by oil of Ceylon cinnamon. Other oils the vapours of which were germicidal though less actively so were oil of geranium and oil of marjoram.

He then tested the effect of the oils in a solution of alcohol and saponin. By this method he found the most active in killing anthrax bacilli were oils of marjoram, cinnamon sandal wood clove juniper and *Artemisia annua*. He draws attention to the fact that cinnamon and marjoram oils are strongly active both in the gaseous and in the liquid state. Similar observations were made by Cadeac and Meunier (1889). They worked with *Salm typhi* and *Pf mallei* which were allowed to remain in contact with the pure oil for a given time and then seeded on to agar. Table 14 shows some of their results:

TABLE 14  
TIME NECESSARY TO KILL *Salm typhi*.

	1° minutes
Ceylon cinnamon oil	25
Clove oil	35
Wild thyme oil	50
Oil of geranium	..

Many other oils did not kill for 24 to 48 hours some not for 4 to 10 days and some not even in 10 days. Garlic vapour has quite a strong bacteriostatic and even a moderately germicidal, effect (see Bocker 1933).

It will be seen that certain of the essential oils if applied pure are fairly active germicides. The majority however are more valued for their antiseptic than for their disinfectant action. For this purpose they were used extensively by the ancient Egyptians in the process of embalming, with results which can be seen at the present day (see Risler 1936).

Vegetable oils that have no germicidal action themselves deprive other germicides which are dissolved in them of most of their activity, in this respect they resemble alcohol. Koch for example found that phenol dissolved in vegetable oils such as olive or cotton-seed oil was only slightly active. McMaster (1919) has since confirmed this but has pointed out that mineral oils do not have this effect. Phenol dissolved in paraffin oil for example is nearly as active as when dissolved in water.

With regard to animal oils Harris, Bunker, and Milas (1932) find that some, such as seal oil and tuna oil, give off vapours which are germicidal, while others, such as cod liver oil and sardine oil, become germicidal only after exposure to sunlight or ultra violet light. It is possible that  $H_2O_2$  is given off by the animal oils, and that its rate of evolution is accelerated by irradiation.

**Sulphonamides and Mould Products**—These two groups of substances, whose chief interest lies in their ability to control infection in the body will be more conveniently discussed in Chapter 6.

### The Dynamics of Disinfection

**Reaction Velocity**—The figures obtained by Kronig and Paul (1897) in their work on the disinfection of anthrax spores by  $HgCl_2$  were submitted by Madsen and Nyman (1907) to a mathematical analysis, with the result that the reaction velocity of disinfection was found to be similar to that obtaining in a unimolecular reaction. Madsen and Nyman themselves made fresh experiments using the garnet method, and were able to confirm the findings of Kronig and Paul. In the following year Chick (1908), working independently, reached the same conclusions with regard to the analogy between disinfection and a unimolecular reaction (Fig. 21).

In a unimolecular reaction only one of the reacting substances need be regarded as undergoing change, the rate of change being proportional to the concentration of this substance. Examples in chemistry are the inversion of cane sugar by acids, the decomposition of  $AsH_3$  into  $As$  and  $H_2$  and the disintegration of radio-active substances. When only one of the reacting substances is undergoing change, the velocity of this change according to the Law of Mass Action will depend upon the concentration of this substance at any given moment, the temperature and other conditions remaining constant. This statement may be expressed by the relation

$$V = Cl$$

in which  $V$  represents the velocity of the reaction,  $C$  the concentration of the substance, and  $l$  a constant depending on the nature of the substance. The velocity may be expressed by  $\frac{dx}{dt}$  in which  $x$  represents the amount of substance changed in time  $t$ , if the original amount of substance is designated by  $a$ , then  $a-x$  will represent the amount remaining after time  $t$ . The equation may now be written

$$\frac{dx}{dt} = l(a-x)$$

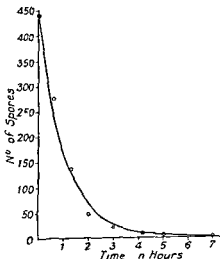


FIG. 21

Disinfection of anthrax spores with 5 per cent phenol at  $33.3^\circ C$ . The curve is drawn through a series of calculated points; the circles represent the experimental observations.

(After Chick.)

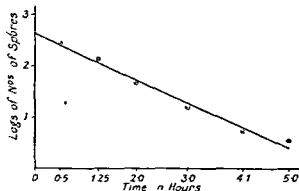


FIG. 22.

Disinfection of anthrax spores with 5 per cent. phenol at 33.3° C. The curve is drawn through a series of calculated points—the circles represent the experimental observations. This curve is constructed from the same observations as those used for Fig. 21 but the numbers of organisms are expressed logarithmically.

(After Chick.)

surviving bacteria per unit volume. For example, let us suppose that there are 100,000 organisms being submitted to disinfection, and that the rate is one at which 90 per cent. of the organisms are killed in each minute. Then,

Time	Nos. Surviving.
After 0 minutes	100,000
" 1 "	$1/10 \times 100,000$ or 10,000
" 2 "	$1/10 \times 10,000$ or 1,000
" 3 "	$1/10 \times 1,000$ or 100
" 4 "	$1/10 \times 100$ or 10

Supposing that  $B$  represents the initial number of living organisms, and  $b$  the final number, then the reaction velocity may be expressed by the equation:

$$k = \frac{1}{t} \log \frac{B}{b}$$

Chick, using the drop method, made experiments on the disinfection of anthrax spores by 5 per cent. phenol. Her results are given in Table 15 and Figs. 21 and 22.

TABLE 15  
ANTHRAX SPORES 5 PER CENT PHENOL. 33.3° C.

Time.	Mean No. of Bacteria per Drop	Value of $k^1$
0 hours	439.0	—
0.5 "	275.5	0.40
1.25 "	137.5	0.40
2.0 "	46.0	0.49
3.0 "	15.8	0.48
4.1 "	5.45	0.46
5.0 "	3.6	0.42
7.0 "	0.5	0.42

<sup>1</sup> Values of  $k$  are calculated to base 10 and not to base  $e$ , and in hours, not in minutes as in Table 18.

which, on integration, becomes

$$k = \frac{1}{t} \log \frac{a}{a-x}$$

This is the equation representing the velocity of a unimolecular reaction and is often spoken of as the logarithmic law. If  $\log(a-x)$  be plotted against time in this equation, the resulting graph will be a straight line (Fig. 22).

We may adapt the unimolecular reaction definition to the process of disinfection by saying that at any moment the reaction velocity is proportional to the number of

It will be seen from Table 15 that  $k$  has a mean value of 0.44, from Fig 21 that the velocity of the reaction becomes slower and slower, till it is almost negligible (in theory the reaction never reaches completion) and from Fig 22 that the logarithms of the numbers of surviving organisms plotted against time in hours fall along a descending straight line

In the case of vegetative bacteria, she (Chick 1908, 1910) found that though the disinfection of some organisms such as *Salmonella typhi* and *Bacterium coli* conformed to the unimolecular reaction formula, with others there was a slight departure from it. Thus with *Staphylococcus aureus* exposed to 0.6 per cent phenol at 20° C, there was invariably a lag period, lasting about 4 minutes before the rate became strictly proportional to the number of bacteria (Fig 23). Paratyphoid bacilli behaved in the opposite way. Instead of there being a lag phase, there was a preliminary rush during which the rate of disinfection proceeded faster than it should have done according to the equation

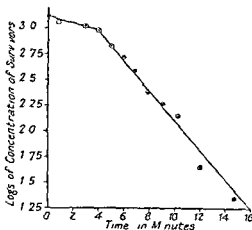


FIG 23

Disinfection of *Staph aureus* with 0.6 per cent phenol at 20° C. The numbers of organisms are expressed logarithmically.  
• (After Chick)

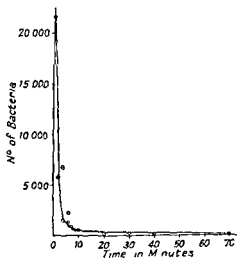


FIG 24

Disinfection of a 24 hours' culture of paratyphoid bacilli with 0.6 per cent phenol at 20° C.  
(After Chick)

Chick confirmed her work on disinfection by phenol by showing (1) that the death of *Bacterium coli* under the influence of a bactericidal serum conforms to the unimolecular reaction law (Chick 1912), and (2) that the same law holds in the process of disinfection by hot water (Chick 1910), she pointed out the close parallel that exists between disinfection by hot water and the heat coagulation of proteins. Paul, Burstein and Reusz (1910a) found that the killing of *Staphylococcus aureus* by drying proceeds in accordance with the law of a unimolecular reaction. The results obtained by Clark and Gage (1903) in the study of disinfection by sunlight may also be interpreted in the same way. Robertson (1914) has adduced

mathematical evidence in favour of this view.

It does not, of course, follow from such results as these that a unimolecular reaction, in the chemical sense, is actually taking place, the recorded observations



merely show that a logarithmic curve describes the death of bacteria under the action of a disinfectant, just as it describes a chemical reaction whose rate is governed by the concentration at any moment of one of the reacting substances. Two alternative explanations have been offered to account for the form of the curves which Chick has described. The first of these suggests that the varying resistance of the bacteria in any given suspension can be described in the form of a frequency curve, as is almost certainly the case, and that the survival curves described by Chick are simply an expression of this difference in resistance. The obvious objection to this hypothesis, as Chick has pointed out, is that the form of the frequency curve describing the distribution of resistance must be supposed to be of the extreme skew form, if it is to account for the experimental results. Withell (1912a), however, has shown that the distribution of bacterial resistance is normal if the survival times are plotted on a logarithmic instead of an arithmetic scale. Such a logarithmic distribution of a characteristic has been noted in pharmacological and zoological work (see Gaddum 1933, Hemmingsen, 1931), and its occurrence in bacteria need not therefore occasion surprise.

An alternative explanation (Chick 1910) is that the death or survival of any given bacterium during any interval of time is determined by a multitude of small and independent causes—by "chance" in the statistical sense—the presence of the disinfectant weighting the chance of survival against each bacterium to a constant degree, for any given concentration of the disinfectant, and with other controllable conditions held constant. If the chance of each bacterium dying during any unit of time is  $x$ , and remains  $x$  over the whole period of the experiment, then the death rate will be the same during each unit of time, the survivors at the end of any one time interval will suffer the same proportionate decrease in their numbers during the time interval which follows, and a logarithmic curve of decrease will result. This explanation does not, of course, mean that variations in resistance of individual bacteria play no part in disinfection. With vegetative bacteria, the rate of death is often represented by a sigmoid curve rather than by a straight line, suggesting that differences in resistance dependent on the age of the individual organisms are responsible for the deviation. The real question is whether in the disinfection of spores that do not differ materially in age the exponential type of curve is due to chance in the statistical sense, or to a frequency distribution of resistance of the logarithmic type. As Irwin (1912) points out, it would require very accurate data to distinguish between the two. (For a further discussion of this subject see Eijkman 1908, Hewlett 1909, Reichel 1909, Reichenbach 1911, Loeb and Northrop 1917, Brooks 1919, Cohen 1922, Knaves 1930a, Knaves and Gordon 1930, Bancroft and Richter 1931, Jordan and Jacobs 1944.)

**Concentration of Disinfectant.**—Chick (1908) found that the relationship between the concentration of a disinfectant and the time taken for disinfection is not a simple but an exponential one, the exponent of the concentration being a factor varying with each disinfectant. That is to say, doubling the concentration of phenol does not halve the time necessary for the completion of the reaction as might be expected, but diminishes it to a far greater extent. Watson (1908), working on Chick's figures, found that the relation could be expressed by the formula

$$C^n t = \text{a constant}$$

where  $C$  is the concentration,  $n$  a constant varying with each disinfectant, and  $t$

the time necessary for disinfection. This equation represents the relation when one molecule of one substance reacts with an excess of molecules of a second. For purposes of calculation it may be written

$$n \log C + \log t = \text{a constant,}$$

that is, the relation between  $\log C$  and  $\log t$  is a linear one

An example will make this clear (Table 16)

TABLE 16  
DISINFECTION OF PARATYPHOID BACILLI BY PHENOL AT 20° C

Parts of Phenol per 1 000	Time taken for Disinfection	$5.5 \log C + \log t$
8.0	45 minutes	6.62
7.5	75 "	6.69
7.0	105 "	6.67
6.5	125 "	6.58
6.0	275 "	6.64
5.5	440 "	6.71
5.0	690 "	6.68

In this table the value of  $n$  is taken as 5.5, the method of calculating this we shall consider presently. It will be seen that the values of the constant are closely similar. If the logarithms of the concentrations are plotted against the logarithms of time, the resulting curve is found to be linear (Fig. 25)

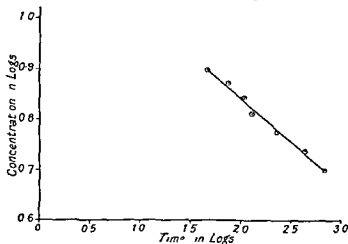


FIG. 25

Disinfection of paratyphoid bacilli with varying concentrations of phenol. Both the numbers of the bacteria and time are expressed logarithmically.  
(After Watson from observations by Chick.)

For dealing with the salts of the heavy metals a slight modification of the formula is required, due to the fact that these salts are dissociated in solution, and their action depends not on their molecular but on their ionic concentration. If the concentration of  $Hg$  ions is substituted for concentration of  $HgCl_2$ , for example, then the formula holds good (see Table 17)

Paul Birstein and Peusz (1910b) found that the value of the velocity constant  $k$  for aqueous solutions of HCl was approximately proportional to the square root of its concentration.

TABLE I  
DISINFECTION OF PARATYPHOID BACILLI BY  $\text{HgCl}_2$  AT  $^{\circ}\text{C}$ .

Parts of H-Cl per 1 000	Conc of Hg ions.	Time taken for Disinfection.	$3.6 \log C + \log t$
1.0	63.0	15 minutes	7.03
0.5	5.5	7.0	7.54
0.1	4.5	10.0	31
0.05	3.0	13.0	6.85
0.01	23.0	65.0 "	6.99
0.005	16.5	230.0 "	6.95

Calculation of Exponent  $n$ —The exponent  $n$  may be regarded as a concentration coefficient varying with each disinfectant. To calculate its value we use the formula

$$1 \text{ } KC^*t = \log \frac{B}{b}$$

$k_1$  is determined for concentration  $C_1$  and  $k_2$  for concentration  $C_2$  in a given experiment. Then

$$n = \log \frac{k_2}{k_1} = \log \frac{C_2}{C_1}$$

Taking the figures in Table 16

let 7.0 parts of phenol per 1000 =  $C_2$  and  $t = 105$

let 5.0 parts of phenol per 1000 =  $C_1$  and  $t = 690$

$$k_2 = \frac{1}{t} \log \frac{B}{b}$$

B in this experiment was 30 000 000  $b$  can be taken as 1. Then

$$k_2 = \frac{1}{105} \log 30\,000\,000$$

$$= 0.0712$$

$$k_1 = \frac{1}{690} \log 30\,000\,000$$

$$= 0.0105$$

$$n = \log \frac{k_2}{k_1} = \log \frac{C_2}{C_1}$$

$$= \log \frac{0.0712}{0.0105} = \log 5$$

$$= \log 6.5 - \log 1.4$$

$$= 5.5$$

For  $\text{HgCl}_2$ , when the concentration of Hg ions only was considered the value of  $n$  was found to be 3.8 for the Ag ions of  $\text{AgNO}_3$  0.86. If the molecular concentration of  $\text{HgCl}_2$  is considered then  $n$  is equal to about 1.

The value of  $n$  for any given disinfectant is very important because it gives us

$1 \text{ } K$  is the true velocity constant of the disinfectant being independent of the concentration, and thus differing from  $k$  which is constant only at a given concentration.

information that is not conveyed by the simple reaction velocity. For phenol let us take  $n = 6$ , and for  $\text{HgCl}_2$ ,  $n = 1$ . Then a doubling of the concentration of  $\text{HgCl}_2$ , i.e.  $C^a$  or  $2^1$ , will halve the time taken for completion of the reaction, doubling the concentration of phenol, i.e.  $C^a$  or  $2^6$ , will diminish it 64 times. Conversely, halving the concentration of  $\text{HgCl}_2$  doubles the time of the reaction, halving the concentration of phenol increases it 64 times.

A substance with a high value of  $n$  is actively germicidal above a given concentration, it requires, however, but a low degree of dilution to abolish its germicidal activity entirely. In contrast, a substance with a low value of  $n$  while being actively germicidal in solutions above a given concentration, exercises an inhibiting effect on the growth of bacteria even when employed in high dilution.

One further point may be dealt with here, namely the question of whether the numbers of bacteria present in a suspension affect the reaction velocity. Working with  $\text{HgCl}_2$  and anthrax spores, Madsen and Nyman (1907) found the numbers of spores to be of no importance, a suspension containing 124 800 was sterilized as rapidly as one containing only 7,750. Eisenberg and Okolska (1913) however divided the disinfectants into 3 classes: in the first, comprising alcohol, phenol and formaldehyde, the numbers of bacteria had but little effect, i.e. a concentration of disinfectant that would destroy a given number of bacteria would also destroy 100 times that number. In a second group, comprising acetone,  $\text{HgCl}_2$ , and  $\text{K}_2\text{Mn}_2\text{O}_8$ , the numbers of bacteria proved to be of importance: thus a concentration that destroyed a given number failed to destroy 10 times that number. A third group comprising  $\text{HCl}$ ,  $\text{H}_2\text{SO}_4$ , oxalic acid,  $\text{KOH}$ , and other bodies occupied an intermediate position between the first two classes.

**Temperature Coefficient**—As the temperature increases in antiseptical progression, the velocity of the reaction increases in geometrical progression or mathematically expressed

$$\frac{k'}{k} = \theta^{(T - T')}$$

in which  $k'$  and  $k$  are the velocity constants of the reaction at temperatures  $T'$  and  $T$  respectively, and  $\theta$  is the temperature coefficient.

In the disinfection of paratyphoid bacilli by 0.6 per cent phenol at  $20^\circ$  and at  $30^\circ \text{C}$ , Chick (1908) obtained the following figures (Table 18).

TABLE 18  
PARATYPHOID BACILLI PHENOL 6 PER 1000

Time in Minutes.	Average No. of surviving Bacteria	$k$
At $20^\circ \text{C}$		
1	539	—
2	276.6	0.29
3	137.5	0.30
4	80.1	0.28
5	42	0.22
At $30^\circ \text{C}$		
1	1368	—
2	162	0.93
3	65.5	0.66
4.1	15.1	0.63
5	1.5	0.59

The mean value of  $k$  at  $20^\circ \text{C} = 0.27$ .

The mean value of  $k'$  at  $30^\circ \text{C} = 0.7$

Paul Birste n and Reusz (1910b) found that the value of the velocity constant  $k$  for aqueous solutions of HCl was approximately proportional to the square root of its concentration

TABLE 1  
DISINFECTION OF PARATYPHOID BACILLI BY  $\text{HgCl}_2$  AT  $20^\circ \text{C}$

Parts of $\text{HgCl}_2$ per 1 000	Cont. of Hg ions.	Time taken for Disinfection.	$2.3 \log C + \log t$
1.0	63.0	1.5 minutes	7.49
0.5	57.5	7.0	7.54
0.1	42.5	10.0 "	7.31
0.05	3.0	13.0	6.90
0.01	23.0	63.0 "	6.99
0.005	16.5	230.0 "	6.93

Calculation of Exponent  $n$  — The exponent  $n$  may be regarded as a concentration coefficient varying with each disinfectant. To calculate its value we use the formula

$$k = KC^n t = \log \frac{B}{b}$$

$k_1$  is determined for concentration  $C_1$  and  $k_2$  for concentration  $C_2$  in a given experiment. Then

$$n = \log \frac{k_2}{k_1} = \log \frac{C_2}{C_1}$$

Taking the figures in Table 16

let 7.0 parts of phenol per 1000 —  $C_2$  and  $t = 100$

let 5.0 parts of phenol per 1000 =  $C_1$  and  $t = 690$

$$k_2 = \frac{1}{t} \log \frac{B}{b}$$

$B$  in this experiment was 30 000 000  $b$  can be taken as 1. Then

$$k_2 = \frac{1}{105} \log 30\,000\,000$$

$$= 0.012$$

$$k_1 = \frac{1}{690} \log 30\,000\,000$$

$$= 0.0108$$

$$n = \log \frac{k_2}{k_1} = \log \frac{C_2}{C_1}$$

$$= \log \frac{0.0712}{0.0108} = \log \frac{7}{5}$$

$$= \log 6.5 = \log 1.4$$

$$= 0.5$$

For  $\text{HgCl}_2$  when the concentration of Hg ions only was considered the value of  $n$  was found to be 3.8 for the Ag ions of  $\text{AgNO}_3$  0.87. If the molecular concentration of  $\text{HgCl}_2$  is considered, then  $n$  is equal to about 1.

The value of  $n$  for any given disinfectant is very important because it gives us

$K$  is the true velocity constant of the disinfectant being independent of the concentration and thus differing from  $k$  which is constant only at a given concentration.

To find the value of  $\theta$

$$\frac{k'}{k} = \theta^{(T - T_1)}$$

$$\theta^{10} = \frac{0.7}{0.27}$$

$$= 2.592$$

$$\theta = 2.592^{0.1}$$

$$= 1.1$$

$$\theta = 1.1 \text{ for } 1^\circ \text{C}$$

$$= 1.1^{10} \text{ or } 2.6 \text{ for } 10^\circ \text{C}$$

A similar experiment is shown diagrammatically in Fig. 26

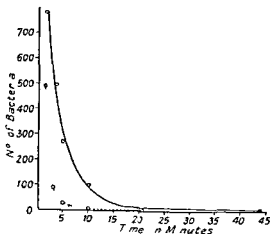


FIG. 26.

Disinfection of a 24 hours culture of paratyphoid bacilli with 0.6 per cent phenol at different temperatures

Continuous curve = 11°C

Interrupted curve = 21°C

(After Chick)

Phenol 6 per 1000

11°C

21°C

Paratyphoid bacilli  
Time elapsing in minutes-

2.2

1.0

Then  $\theta = \frac{2.2}{1.0} = 2.2$  for 10°C, or  $2.2^{0.1} = 1.08$  for 1°C

In disinfection by hot water a very much higher value is obtained for  $\theta$ . Thus Chick (1910) working with *Salmon typhi* at temperatures of 49°C and 51.1°C, found that the velocity constant of the reaction was increased 13.1 times for the 5°C rise in temperature, i.e.  $\theta = 1.67$  for 1°C, or about 170 for 10°C.

The consistent effect of rise of temperature on the velocity of disinfection points to a close analogy with an ordinary chemical reaction. Arrhenius elaborated a formula, which has been found to be applicable to many chemical re-

actions and it is interesting therefore to ascertain whether it applies also to the reaction of disinfection. The formula is

$$A = \frac{T_0 T_n}{T_0 - T_n} \log \frac{K_0}{K_n}$$

where  $K_0$  and  $K_n$  are the velocity constants of the reaction in question corresponding to the absolute temperatures  $T_0$  and  $T_n$  respectively and  $A$  is a constant. As the time taken for the completion of a reaction may be considered as inversely proportional to the velocity of the reaction this equation may be re-written thus

ferred to a solid medium in order to ascertain whether all the spores had been killed. This method is open to certain fallacies to which attention was first drawn by Geppert (1889-1891a b). Geppert found that if after a short time in mercuric chloride solution the spores were transferred to a culture medium they failed to grow but that if they were inoculated into a guinea pig they gave rise to anthrax. The reason for this appeared to be that sufficient  $\text{HgCl}_2$  was carried over by the thread into the broth to prevent germination of the spores occurring in the tissues of the guinea pig however this quantity was neutralized and the spores that were still alive were able to develop. The truth of this explanation was shown by the fact that cultivation of the spores proved successful provided the threads were treated with ammonium sulphide to neutralize the mercury before being inoculated into the broth. The thread method, therefore yielded higher values than the disinfectant actually possessed. Geppert's work was of considerable importance particularly in relation to disinfection by salts of the heavy metals. These substances have a low concentration exponent and therefore act as antiseptics even in high dilution. Since it is impossible by this method to remove all traces of the disinfectant from the interstices of the thread Kröning and Paul (1894) replaced the threads by garnets. They introduced a further improvement which rendered the test quantitative by plating out the washings from the garnets and counting the number of colonies that developed. The two tests that are chiefly used at the present time either in their original or in a modified form are the Rideal Walker and the Chick Martin methods.

**The Rideal Walker Drop Method (Rideal and Walker 1903)**—In this method similar quantities of organisms are submitted to the action of varying concentrations of phenol and of the germicide to be tested. Subcultures are made into broth every 2½ minutes up to 15 minutes and the tubes incubated at 37°C for 3 days. That dilution of disinfectant X which sterilizes the suspension in a given time is divided by that dilution of phenol which sterilizes the suspension in the same time and a phenol coefficient obtained. Thus

TABLE 20

*Salmon typhi* 4 HOURS BROTH CULTURE AT 37°C TEMPERATURE AT WHICH TEST WAS CONDUCTED 60°F

Disinfectant.	Dilution.	Time in minutes of exposure of suspension to disinfectant.					
		1	5	10	15	1.1	15
Phenol	1:110	+	—	—	—	—	—
"	1:120	+	+	—	—	—	—
"	1:130	+	+	+	—	—	—
"	1:140	+	+	+	+	—	—
X	1:225	+	—	—	—	—	—
"	1:250	+	+	—	—	—	—
"	1:275	+	+	+	—	—	—
"	1:300	+	+	+	+	+	—

+ = growth

— = no growth

The phenol coefficient of X is therefore  $\frac{225}{130} = 1.73$

(For a full discussion of the Rideal Walker method see *Lancet* 1909 ii, 1516)



**The Chick-Martin Test**—In the Rideal Walker method the disinfectant acts in pure solution. But, in practice, disinfectants have usually to act in solutions containing organic matter. As the presence of organic matter seriously lessens the activity of most disinfectants, Chick and Martin (1908) suggested that the disinfectant should be tested on the organisms not in distilled water but in water containing a suspension of 3 per cent dried human feces. Further instead of allowing the time to vary, as in the Rideal Walker method they fix a time limit of 30 minutes for the action of the disinfectant, making subcultures at the end of this time. The phenol coefficient of disinfectants, especially those of the emulsified disinfectants is distinctly lower by this method. Thus whereas the activity of phenol was reduced about 10 per cent in the presence of 3 per cent feces that of the commercial cresols was reduced 30-50 per cent.

of time. Nor does it give us any information about the toxicity of the disinfectant to the tissues, its anti-leucocytic power, its activity in the presence of protein or lipins, or its selective action on different bacteria. Any attempt to express by a single index a number of dissociated properties is clearly illogical. The phenol coefficient as given by the Rideal-Walker method is at best a grossly over-simplified answer to a very difficult problem, and at worst little short of bacteriological prostitution. To scientific workers and to those interested in human and animal therapy the result is meaningless. Were it not for the big financial interests concerned in the commercial production of disinfectants, the test would doubtless die of disuse.

What is the remedy? Several have been proposed but none is altogether satisfactory. Phelps (1911) suggests that a method of standardization should comprise the determination of  $k$ ,  $n$  and  $\theta$  for each disinfectant. By the formula

$$K\theta^{\alpha} = \log \frac{B}{b}$$

we can calculate the reaction velocity at any concentration and by the formula

$$K_T = K_{20} \times \theta^{(\alpha - 20)}$$

where  $K_T$  represents the velocity to be calculated at the temperature desired and  $K_{20}$  the velocity actually determined at a temperature of 20°C (or any other convenient temperature) we can determine the value of the temperature coefficient  $\theta$ .

One objection to this method is that the reaction velocity often varies during the progress of disinfection. To meet this difficulty Hobbs and Wilson (1942) suggested that the value of  $k$  should be taken in the middle stage of the reaction. Withell (1942a, b) has improved on this by proposing that the time taken to destroy 50 per cent. of the organisms— $L_{50}$ —should be selected as a comparative measure of bactericidal efficiency, and has pointed out that this value can be most easily determined by the use of probit logarithm of time graphs (see Bliss 1933, 1941). Withell (1942b) has also drawn attention to the danger of attempting to compare two bactericidal agents that yield different types of time-survivor curve. Many of these are at variance with that yielded by phenol and a phenol coefficient is therefore inapplicable.

If Phelps' method is adopted then it is advisable to test the disinfectant against two or three different organisms as suggested by Eisenberg (1919) since it is known that some germicidal agents have a strongly selective action on certain bacteria. The information derived from the use of this method is considerably greater and more valuable than that supplied by the Rideal-Walker coefficient. Nevertheless, it still leaves us ignorant of a number of important properties of the disinfectant, particularly its toxicity to body tissues and its activity in the presence of cellular material. For this reason the behaviour of the disinfectant in tissue suspensions or cultures can be studied (see Bronfenbrenner *et al.* 1939, Sallie *et al.* 1939, Welch and Brewer 1942).

In conclusion we may say that work of recent years, particularly on the sulphonamides, has shown how inadequate any laboratory test of germicidal activity is to indicate the behaviour of a given substance in the animal body. Attention in the future is therefore likely to be devoted far more than in the past to a study of disinfectant agents in the presence of living tissues.

**Practical Application of Germicides**—We shall not give more than a brief account of the use of germicides in practice, and shall confine ourselves to general remarks, illustrating the application of the principles that we have already considered.

**Gaseous Disinfectants**—The gaseous disinfectants most commonly employed are sulphur dioxide, chlorine, and formaldehyde. The first two are active only in a moist atmosphere. Thus sulphur dioxide combines with water to form sulphurous acid, and chlorine to form hypochlorous acid. To be successful in the destruction of vegetative bacteria, sulphur dioxide should be present in a concentration of 2–3 per cent., chlorine of 1 per cent., and formaldehyde 1–2 per cent. of the atmosphere. Ozone is sometimes used for the sterilization of water and for meat preservation. Heise (1917) found that concentrations by volume of about 1/1000 destroyed 90 per cent. of coliform organisms on the surface of an agar plate in 1 minute, concentrations of about 1/270 000 in 1 hour and concentrations of about 1/720 000 in 3–4 hours. The gas has little penetrating power and is of value only for the destruction of organisms unprotected by colloidal material (see Elford and van den Ende 1912). The use of aerosols and vapour disinfectants for sterilization of the air—as apart from fumigation—is considered in Chapter 91.

**Liquid Disinfectants**—In actual practice it is inevitable that disinfectants should be employed more or less empirically, it is impossible, from knowledge gained in the laboratory, to predict exactly the length of time requisite for the complete sterilization of any material. Realizing this, we err on the safe side and arrange our conditions so as to obtain sterilization in a time much shorter than that which is actually allowed. To do this, however, it is necessary to take into consideration the principles that we have already considered so far as they are known and pay particular attention to such variables as the nature of the organism, the material in which it is contained, the H<sup>+</sup> ion concentration, the salt content, and the temperature at which the reaction is to proceed. Having considered these, the disinfectant to be chosen, the concentration in which it shall be allowed to act, and the time for which its action shall continue may be determined. For general purposes, we may lay down a few simple rules:

- (1) Spores are more resistant than vegetative bacteria.
- (2) Bacteria possessing a high content of lipins, such as the acid fast bacilli are very resistant to liquid disinfectants. Tubercle bacilli in sputum may withstand 5 per cent. phenol for 24 hours, but they are killed by boiling in 1 minute.
- (3) For the destruction of spores and acid fast bacilli heat is preferable to chemical disinfectants.
- (4) Bacteria suspended in a protein medium are more resistant than those in a non protein medium.
- (5) If the protein medium is also a good nutrient medium the organisms are even more resistant.
- (6) Disinfection by nearly all germicides proceeds more quickly in an acid than in an alkaline medium. There is evidence that a given concentration of H<sup>+</sup> ions is more bactericidal than of OH<sup>-</sup> ions.
- (7) The effect of salts in the medium depends on their nature and on their concentration. In general, salts increase the action of phenol and of the emulsified disinfectants, but diminish that of HgCl<sub>2</sub>.

(8) The higher the temperature at which a disinfectant is allowed to act the more rapid is the process of sterilization

(9) Some germicides dissolved in alcohol or in vegetable oils are deprived of the greater part of their power but an alcoholic solution of iodine is a potent skin disinfectant.

(10) Doubling the concentration of  $\text{HgCl}_2$  halves the time taken for sterilization, doubling the concentration of phenol diminishes it about 64 times.

(11) For use in a protein medium the acid disinfectants, such as hypochlorous acid bleaching powder, and the disinfectants that can be employed combined with acids, are most effective. The alkalis are also reliable.

(12) In the presence of organic matter, whether in solution or in suspension, the activity of certain disinfectants is markedly lowered, especially with the emulsified disinfectants, with oxidizing agents, and with the salts of the heavy metals. Phenol is much less affected and likewise certain dyes, such as flavine.

(13) Salts of the heavy metals—mercury, silver, and copper—are mainly of value for their bacteriostatic effect, their ability to kill bacteria has been grossly overestimated in the past and is in reality comparatively low.

(14) For the disinfection of varnished or greasy surfaces, an emulsified disinfectant or a bactericidal wetting agent is to be recommended, aqueous solutions are of little value.

(15) For the disinfection of metal instruments, all substances that act on the metal causing rust or other change, must be avoided.

(16) For references to disinfection of the hands and skin, the following papers may be consulted. Ahlfeld and Vahle (1896) Colebrook (1930-1941), Colebrook and Marted (1933) Neufeld and Schiemann (1943).

(17) For instructions in the use of liquid disinfectants in hospitals, see Report (1944).

**Solid Disinfectants**—These are generally made up in the form of powders, with a basis of lime silicious matter, or vegetable fibre. Phenol is the commonest disinfectant incorporated. To destroy bacteria they must pass into solution, in the dry state they act merely as deodorants.

## REFERENCES

- AHLFELD F and VAHLE, F (1896) *Disch med Wochr.*, 22, 81  
 ALBERT A (1947) *Lancet* ii 633  
 ALLEN P W (1923) *J Bact.*, 8, 500.  
 ANDREWS F W and ORTON K. J P (1904) *Zbl Bakt.*, 35, 645-811  
 APOSTOLI G and LAQUERRIERE, A. (1890) *C R Acad Sci* 105 918  
 D ARCY R F and HARDY W B (1894) *J Physiol* 17, 370.  
 BAKER, Z. HARRISON R W and MILLER, B F (1941a) *J exp Med* 73, 249 (1941b) *Ibid.*, 74, 611-691  
 BANCROFT W D and RICHTER, G H (1931) *J phys. Chem.*, 35, 511  
 BARVARD J E. and MORGAN H DE R (1903) *Proc roy Soc.*, 72, 126  
 BARNES J M (1947) *Lancet* i 531  
 BARR, C. E. (1923) *J med Res* 44, 79  
 BAXTER, (1875) *Rep loc Govt Bd. pub. Hlth.*, New Ser., No 5 appendix p. 216  
 BAILLIS M. (1930) *J Bact.*, 31, 43  
 BAYLIS W M (1924) *Principles of General Physiology* 4th ed. London.  
 BEATTIE, J M and LEWIS F. C. (1930) *Spec Rep med. Res Coun Lond.*, Ser No 49  
 BECKMAN J W (1896) *Zbl Bakt* 20 5  
 BECKWITH T D and OLSON A R. (1932) *Proc. Soc exp Biol.*, N Y., 29, 362.  
 BECKWITH T D and WEAVER, C F (1930) *J Bact* 22, 361  
 BEHRING (1890) *Z Hyg Infektkr.*, 9, 395

- BELEHRÁDEK, J. (1920) *Nature*, 118, 117
- BELIN, M and RIFERT, J. (1937) *C R Soc Biol.*, 124, 612
- BESSEMAN, A and MEIRHAEGH, A VAN (1937) *Bull Acad Med*, 118, 263
- BIGELOW, W D and ESTY, J R. (1920) *J infect Dis*, 27, 602
- BISCEGLIE, V. (1926) *Z Immunforsch.*, 49, 272
- BLISS, C I. (1938) *Quart J Pharm*, 11, 192, (1941) *J Amer pharm Ass*, 29, 465
- BLUM, H T. (1932) *Physiol Rev*, 12, 23
- BÖCKER, O E. (1938) *Z Hyg Infekthkr*, 121, 160
- BROUVERENNER, J, HERSHEY, A D, and DOUBLY, J. (1931) *J Bact*, 37, 583
- BROOKS, S C. (1919) *J gen Physiol*, 1, 61, (1920) *J med Res*, 41, 411
- BROWNING, C H. (1933) *Brit dent J*, 54, 389, (1913) *Brit med J*, i, 311
- BROWNING, C H and RUSS, S. (1917) *Proc roy Soc, B*, 90, 33
- BROWNING, C H, KENNAWAY, E L, GULBRANSEN, R, and THORNTON, L H D. (1917) *Brit med J*, i, 73
- BRUYNOGHE, R and DUBOIS, A. (1925) *C R Soc Biol*, 93, 849
- BRUYNOGHE, R and LE FÈVRE DE ARRIC, M. (1925) *C R Soc Biol*, 93, 852
- BUCHHOLZ, J and JENKY, A V. (1935) *Zbl Bakt*, 133, 299
- BUCHOLTZ, L. (1875) *Arch exp Path Pharmacol*, 4, 1
- BURGE, W E and NEILL, A J. (1915) *Amer J Physiol*, 38, 399
- DURKE, G S. (1923) *J infect Dis*, 33, 274
- BURNET, F M. (1925) *Aust J exp Biol med Sci*, 2, 65
- BURTONSHAW, J M L. (1942) *J Hyg Camb*, 42, 184
- CADÉAC and MEUNIER, A. (1889) *Ann Inst Pasteur*, 3, 317
- CAMERON, A T. (1930) *Trans roy Soc Can*, 3rd Series, 24, Section V, 53
- CHAMBERLAND, M. (1887) *Ann Inst Pasteur*, 1, 153
- CHAMBERS, L A and FLOSDORF, E W. (1936) *Proc Soc exp Biol N Y*, 34, 631
- CHAMBERS, L A and GAINES, N. (1932) *J cell comp Physiol*, 1, 451
- CHICK, H. (1908) *J Hyg, Camb*, 8, 92, (1910) *Ibid*, 10, 237, (1912) *Ibid*, 12, 414
- CHICK, H and MARTIN, C J. (1908) *J Hyg, Camb*, 8, 693, (1910) *J Physiol*, 40, 404
- CHRISTOLM, J. (1941) *Brit med J*, i, 135
- CHRISTIAN, M I. (1931a) *J Dairy Res*, 3, 113, (1931b) *Nature Lond*, 127, 584
- CHURCHMAN, J W. (1912) *J exp Med*, 18, 221; (1923a) *Ibid*, 37, 1, (1923b) *Ibid*, 38, 1
- CLARK and GAGE. (1903) 34th Rep State Bd Hlth, Mass
- CLIFTON, C E. (1931) *Proc Soc exp Biol, N Y*, 28, 745
- COHEN, B. (1922) *J Bact*, 1, 183
- COHEN, B and CLARK, W M. (1919) *J Bact*, 4, 409
- COLEBROOK, L. (1930) *Interim Rep depart Comm Maternal Mortality and Morbidity*
- Appendix D, Min Hlth, Lond (1941) *Bull War Med*, 2, 73
- COLEBROOK, L and MATTED, W R. (1933) *J Obstet Gynecol*, 40, 966
- COULTHARD, C F and SYKES, G. (1936) *Pharmaceut J*, 83, 79
- CROWTHER, J A. (1926) *Nature*, 118, 86
- CRUICKSHANK, J C, HOBBS, B C, MCFARLAN, A M and MAIER, I. (1942) *Brit med J*, ii, 182
- DARIN, H D. (1915) *Brit med J*, ii, 318
- DANYSZ, J. (1906) *Ann Inst Pasteur*, 20, 206
- DELÉPINE A S and GREENWOOD, A. (1914) *J R sanit Inst*, 35, 317
- DOWNES, A and BLUNT, T P. (1877) *Proc roy Soc*, 26, 489, (1878) *Ibid*, 28, 199
- DRYFER G and CAMPBELL-RENTON, M L. (1936) *Proc roy Soc, B*, 120, 417
- DREYER and HANSEN. (1907) *C R Acad Sci*, 145, 274
- DUCLAUX, E. (1887) *Ann Inst Pasteur*, 1, 88
- ECKELMANN, F. (1917) *Zbl Bakt, Hto Abt*, 48, 140
- LOGFORTH, A H. (1926) *J gen Physiol*, 10, 147, (1927a) *J exp Med*, 49, 53, (1927b) *Ibid*, 50, 299, (1931) *Ibid*, 53, 27
- EHRLICH, O. (1930) *Z Hyg Infekthkr*, 111, 618
- EHRLICH, O and NOETHLING, W. (1932) *Z Hyg Infekthkr*, 113, 597
- ELJEMAN, C. (1908) *Biochem Z*, 11, 12
- EISENBERG, P. (1919) *Zbl Bakt*, 82, 69
- EISENBERG, P and OKOLSKA, M. (1913) *Zbl Bakt*, 69, 312
- EISLER, M V. (1909) *Zbl Bakt*, 51, 546
- FLYORD W J and FENDE J, VAN DEN. (1942) *J Hyg, Camb*, 42, 240
- ELSEN, W J, THOMAS, R A, and STIFFEN, G I. (1935) *J Immunol*, 23, 433
- FELSTEIN, F. (1897) *Z. Hyg Infekthkr*, 24, 1
- EVANS F R and CURRAN H R. (1943) *J Bact*, 46, 513
- FABIAN, F W and GRAHAM, H T. (1933) *J infect Dis*, 53, 76
- FALE, I S. (1923) *Abst Bact*, 7, 33, 87, 133
- FAY, A C. (1934) *J agric Res*, 48, 453
- FRISTMANTEL, C. (1902) *Zbl Bakt*, 31, 433
- FERRI, C and PEROSSI, L. (1894) *Z Hyg Infekthkr*, 16, 385

- FICKER M (1898) *Z Hyg InfektKr* 29 1  
 FILDES P (1940) *Brit J exp Path* 21, 67  
 FISCHER. (1900) The Structure and Functions of the Bacteria Trans by A C Jones  
 Oxford  
 FLEMING A. (1932) *J Path Bact.* 35 831  
 FLEMING A. (1940) *Proc R Soc Med* 33, 487  
 FLEMING A and YOUNG M Y (1940) *J Path Bact.*, 51, 99  
 FLEXNER, S (1906) *J exp Med* 9 103  
 FLOSDORF E. W and MCDD S (1935) *J Immunol.*, 29 39 (1938) *Ibid* 34, 469  
 FORBATA E and HASORI A. (193) *Zbl Bakt* 139 163  
 FOTLIS M (1911) *J R Army med. Cps.*, 16 167  
 FREUNDLICH H (1903) *Z phys Chem.*, 44, 199  
 GADDUM J H (1933) *Spec Rep Ser med Res Coun., Lond.*, No. 183  
 GALE, C K and MILLER, D (1935) *J lab clin Med.*, 21 31  
 CALE L F (1940) *Bact Rev.*, 4, 133  
 CARROD L P (1933a) *St. Bart's Hosp med Rep.*, 66 903 (1933b) *Brit J exp Path.*, 14, 187 (1944) *Brit med J.*, 1 243  
 GATES, F L (1930) *J gen Physiol* 14, 31  
 GEFFERT J (1889) *Bevl Klin Wchr* 28 89 819 (1891a) *Dtsch med Wchr.*, 17 97  
 875 883 (1891b) *Ibid.*, 17 1065  
 HAINES I B (1938) *Proc roy Soc., B.*, 124 451  
 HAMILTON H C (191) *Amer J publ Hlth.*, 7 289  
 HARRIS P S BUCKER, J W M and MILAS A. (1939) *J Bact* 23, 499  
 HARTLEY P (1936) *Quart Bull Hlth Org., L o N.*, 5 735  
 HASCHÉ, E and LECNIG H (1935) *Dtsch med Wchr* 61, 1193  
 HASCHÉ, F and LOCH P (193) *Z Hyg InfektKr* 120 909  
 HAYES S N (193) *Brit med J* 1 911  
 HEISE, P (1917) *Arch Peischgesundh.Amt.* 50 904 418  
 HENNINGSEN A W (1934) *Udensk Medd Dansk naturhist Foren København.*, 98 123  
 HEWLETT R T (1909) *Lancet* 1 41 815 889  
 HICKS P A and SZYMANOWSKI W T (1939) *J infect Dis.*, 50 466  
 HOBBS B C and WILSON G S (1949) *J Hyg., Camb.*, 42, 436  
 HODER, F (1939) *Z Immunforsch.*, 74, 453  
 HOFMANN P (1939) *Zbl Bakt.*, 114, 916  
 HOFMEISTER, F (1888) *Arch exp Path Pharmac.*, 24, 94 (1889) *Ibid.*, 25 1  
 HOLM G E and SHERMAN J M (1931) *J Bact.*, 6, 511  
 HOTCHKISS M. (1923) *J Bact* 8, 141  
 HOUSTON A C (1914) *10th Res Rep metrop Wat. Bd*  
 HOYT A FISK R T and BURDE, G (1942) *Surgery* 12, 786  
 ILAND C N (1944) *Lancet* 1 49  
 IRWIN J O (194) *J Hyg Camb* 42, 3 8.  
 JODLBACHER, A and TAPPEINER, H v (1904) *Munch med Wchr.*, 51, 1096  
 JORDAN P C and JACOBS S E (1944) *J Hyg Camb* 43, 995  
 KITASATO S (1891) *Z Hyg InfektKr.*, 10 96  
 KLAHMANN E SHTERNOV A and GATES, L W (1934a) *J Lab clin Med.*, 19 835  
 (1934b) *Ibid* 20 40  
 KLING A. (193) *C R Acad Sci* 194 1409  
 KNAYSI G (1930a) *J infect Dis.*, 4 293, 392, 398 (1930b) *J Bact.*, 19 113  
 KNAYSI G and GORDON M (1930) *J infect. Dis* 47 303 318.  
 KOCH I (1881) *Mitt Reichsgesundh.Amt* 1, 234  
 KOCH P and WOLFFHÜGEL, G (1881) *Mitt Reichsgesundh.Amt.*, 1, 301  
 KOCH P GAFFKY and LOEFFLER. (1881) *Mitt Reichsgesundh.Amt* 1, 322  
 KOHNICH F (1938) Die bakterielle Keimtotung durch Wärme Ferd. Enke Verlag  
 Stuttgart  
 KRONIG B and PAUL, T (189) *Z Hyg InfektKr* 25 1  
 LANAR, R V (1911) *J exp Med* 13 1 340  
 LARDEQUETTE, M DE. (1918) *Ann Inst Pasteur* 32, 1 0  
 LEA D E. and HAINES R B (1940) *J Hyg Camb.*, 40 169  
 LEA D E., HAINES P B and BRETSCHER, F (1941) *J Hyg., Camb.*, 41, 1  
 LEA D E HAINES R B and COLLISON C A (1936) *Proc. roy Soc., B.*, 120 47 (193) *Ibid* 123, 1  
 LENTZ, F A. (193) *ZW Bakt.*, 126, 508  
 LEVY B S and LOMTANIKI, I (1935) *C R Acad Sci.*, 200 863  
 LIT S C and YEN A C H (1934) *Proc Soc exp Biol.*, N Y., 32, 485  
 LOCKEMAN G BAR, F., and TOTZECK W (1941) *Zbl Bakt* 147 1  
 LOEB, J (1899) *Pflug Arch ges Physiol.*, 75 303 (1900) *Amer J Physiol.*, 3, 327  
 LOEB, J and NORTROP J H (1917) *J Biol Chem.*, 32, 103  
 MACFADYEN A (1900) *Proc roy Soc B* 66 180 339

- MACFADYEN, A and ROWLAND S D (1900) *Proc. roy Soc. B*, 66, 433  
 McINTOSH, J and SELBIE, F R (1942) *Lancet* ii 750  
 McMASTER P D (1919) *J infect Dis*, 24, 378  
 MADSEN, T and NYMAN M (1907) *Z Hyg Infektkr*, 57, 388  
 MALLMANN, W L, BOTWRIGHT W L and CHURCHILL, E S (1911) *J infect Dis* 63  
 MANFOLD M C (1941) *Brit J exp Path* 22, 111  
 MATHEWS, A P (1904a) *Amer J Physiol*, 10, 290, (1904b) *Ibid* 11, 45. (1905) *Ibid* 12, 419, (1906) *J infect Dis*, 3, 572  
 MAYSER, H (1920) *Zbl Bakt*, 94, 238  
 MILLER, B T and BAKER, T (1940) *Science* 91, 604  
 MINERVINI R (1898) *Z Hyg Infektkr*, 29, 117  
 MOORE, H N and KERSTEN, H (1937) *J Bact* 33, 815  
 MORTON, H C and KLAUDER, J V (1944) *J Amer med Ass* 124 114  
 MÜNDEL, O (1937) *Z Hyg Infektkr*, 120, 267  
 NEUFELD F and SCHIFMANN O (1943) *Z Hyg Infektkr* 124 751  
 NICHOLS, H J (1920) *J Lab clin Med* 5, 502  
 NOGUCHI H (1907) *Biochem Z*, 6, 327  
 NORTON, J F and Hsu, P H (1916) *Z Hyg Infektkr*, 18, 180  
 OAO, R A (1910) *J Path Bact* 51, 137  
 OTTEN, L. (1930) *Zbl Bakt*, 116, 199, (1932) *Trans Far East Ass trop Med Soc Bangkok* 1930, p 89  
 OZZANO T and RE C (1937) *G Batt Immun*, 19, 535  
 PAIC, M, DEUTSCH V, and BORCHIA I (1935a) *C R Soc Biol*, 119, 1063  
 PAIC, M, HABER, P, VOET, J, and ELIASZ, A (1935b) *C R Soc Biol* 119, 1081  
 PAUL, T (1909) *Biochem Z*, 18, 1  
 PAUL, T and PRALL, F (1907) *Arb Reichsgesundh.Amt*, 28, 73  
 PAUL, T, BIRSTEIN, G, and REUSZ A (1910a) *Biochem Z*, 25, 367 (1910b) *Ibid* 29 20.  
 PEDRAU, J R, and TODD C (1933a) *Proc roy Soc B*, 112, 277 (1933b) *Ibid* 112 285  
 PHELPS, E B (1911) *J infect Dis*, 8, 27  
 PILOD and CONVELLE (1932) *C R Acad Sci*, 194, 497  
 PRINGLE (1750) *Philos Trans*, 46, 480 525  
 PROCHOWNICK, L and SPARTH, F (1890) *Dtsch med Wochr*, 16, 564  
 PUDHOMME, B O (1937) *C R Soc Biol* 128, 289  
 PUGSLEY, A T, ODDIE, T H, and EDDY, C E (1935) *Proc roy Soc. B* 118, 276  
 RAAB, O (1900) *Z Biol*, 39, 524  
 REICHEL, H (1909) *Biochem Z*, 22, 149  
 REICHENBACH, H (1908) *Z Hyg Infektkr*, 59, 296, (1911) *Ibid*, 69 171  
 REID, J D (1932) *Amer J Hyg*, 16, 540  
 REITSCHLER H C NAGY, R and MOUROMSEFF, G (1941) *J Bact*, 41, 745  
 Report (1942) 'The Prevention of 'Hospital Infection' of Wounds. *Med Res Coun.*  
*London War Memo.* No 6  
 Report (1944) 'The Control of Cross Infection in Hospitals' *Med Res Coun.* *London.*  
*War Memo* No 11  
 RIDEAL, S and WALKER, J T A. (1903) *J R sanit Inst*, 24, 424  
 RIEDER, H (1902) *Munch med Wochr*, 49, 402  
 RISHWORTH H R (1938) *Brit med J* ii 574  
 RISLER, J (1936) *C R Acad Sci*, 203, 517  
 RITCHIE, J (1899) *Trans path Soc Lond.*, 50, 256  
 RIVERS T M, SMADEL, J E and CHAMBERS, L A (1937) *J exp Med* 65, 77  
 ROBERTSON, T B (1914) *J Hyg, Camb*, 14, 143  
 ROCKWELL, G E and EBERTZ, F G (1924) *J infect Dis*, 35, 573  
 RÖMER, C (1898) *Munch. med Wochr*, 45, 298  
 ROUX, E (1897) *Ann Inst Pasteur*, 1, 445  
 ROUYER, M and SERVIGNY, M. (1934) *Ann Inst Pasteur* 52, 565  
 RUBBO S D ALBERT A and MAXWELL, M (1942) *Brit J exp Path*, 23, 79  
 RUSSELL, V (1904) *Zbl Bakt*, 37, 115 250  
 RUSSELL, D S and BECK, D J K (1914) *Brit med J*, i 112  
 RUSSELL, D S and FALCONER, M A (1941) *Brit J Surg*, 28, 472 (1943) *Lancet* i 550  
 SALLE, A J, McOMIE, W A, SHECHMISTER, I L and FORD D C (1939) *J Bact.* 37, 639  
 SAVAGE, R M (1940) *Quart J Pharm.*, 13, 237  
 SCHERR, H W and CHAMBERS, L A (1936) *Proc Soc exp Biol.* vi 35, 493  
 SCHERLEN F (1895) 'Die Bedeutung des Moleculargewichtes der wassergerästen  
 Desinfektionsmittel für ihren Wirkungswert' Straßburg  
 SCHMIDT, C L A and NORMAN G F (1920) *J infect Dis*, 27, 40  
 SCHOOF, G (1935) *Zbl Bakt*, 134, 14  
 SCHROETTER, H V (1927) *Zbl Bakt*, 104, Beiheft p. 205  
 SELLEARDS, A W (1918) *J med Res*, 38, 293

- FICKER M (1895) *Z Hyg InfektKr* 29 1  
 FIELDS P (1940) *Brit J exp Path* 21, 67  
 FISCHER (1900) *The Structure and Functions of the Bacteria*. Trans. by A. C. Jones  
 Oxford  
 FLEMING A (1930) *J Path Bact* 35 831  
 FLEMING A (1940) *Proc P Soc Med* 33, 45  
 FLEMING A and YOUNG M A (1940) *J Path Bact.* 51, 29  
 FLEXNER S (1907) *J exp Med.* 9 105  
 FLOSDORF E. W. and MUDD S. (1935) *J Immunol.* 29 389 (1938) *Ibid* 34, 479  
 FORBOTA, E. and HAMORI A. (1913) *Zbl Bakt.* 139 163  
 FORTD., M. (1911) *J P Army med Cps.* 16, 167  
 FREUNDLICH H (1903) *Z phys Chem.* 44, 109  
 GADDUM J H (1933) *Spec Rep Ser med Res. Coun., Lond.* No. 183  
 GALE, C. K. and MILLER, D. (1935) *J lab clin Med.* 21, 31  
 CALE, E. F. (1940) *Bact. Rev.* 4, 135  
 CARROD L P (1933a) *St. Bart's Hosp med Rep* 66, 203 (1933b) *Brit. J exp Path.* 14, 180 (1944) *Brit med J.* 1 115  
 GATES, F. L. (1930) *J gen Physiol* 14, 31  
 GERBERT J (1889) *Berl. klin Wochr* 26 80 819 (1931a) *Deutsch med Wochr.* 17, 97  
 80 830 (1931b) *Ibid* 17 1065  
 HAINES R B (1938) *Proc. roy Soc., B.* 124 451  
 HAMILTON H C (1917) *Amer J publ Hlth.* 7 522  
 HARRIS P S, BURKER, J W M., and MILLAS A. (1930) *J Bact.* 23, 409  
 HARTLEY P (1936) *Quart Bull Hlth Org., L.* 5 5 735  
 HASCHÉ, E. and LUTVIG H (1935) *Deutsch med. Wochr.* 61, 1193  
 HASCHÉ, F. and LOCH P (1931) *Z Hyg InfektKr.* 120 209  
 HAYES S (1913) *Brit med J.* 1, 911  
 HEISE, R. (1917) *Arch. PeichgerundhAmt.* 50 204 418.  
 HENNINGSEN A M (1934) *Vdenok Medl Dansk naturhist Foren Koenhar.* 98, 125  
 HEWLETT R T (1909) *Lancet* 1 41 815 859  
 HICKS P A. and SEYMANOWSKI, W T. (1932) *J infect. Dis.* 50, 466.  
 HOBBS B C and WILSON G S (1942) *J Hyg., Camb.* 42, 436  
 HODER, F. (1937) *Z Immunforsch* 74, 455.  
 HOFMAN P (1919) *Zbl Bakt.* 114, 216  
 HORMEISTER, F. (1885) *Arch exp Path Pharmac* 24, 24 (1889) *Ibid.* 25 1  
 HOLM G E. and SHERMAN J M (1911) *J Bact.* 6, 511  
 HOTCHKISS M (1923) *J Bact.* 8 141  
 HORTON A C. (1914) *10A Res Exp metrop Wat. Ed*  
 HOYT A., FISK, P T and BURDE, G (1910) *Surgery* 12, 56.  
 ILAND C N (1944) *Lancet* 1, 49  
 IRWIN J O (1941) *J Hyg., Camb* 42, 323  
 JODLBAUER, A. and TAPPEINER, H v (1904) *Munch med Wochr.* 51, 1096  
 JORDAN P C and JACOBS S E. (1944) *J Hyg., Camb.* 43, 255  
 KITAYATO S (1891) *Z Hyg InfektKr.* 10 267  
 KLARMAN E., SHTERNOV V A., and GATES, L. W. (1934a) *J Lab clin. Med.* 19, 835.  
 (1934b) *Ibid.* 20 40  
 KLING A. (1931) *C R Acad Sci.* 194, 1402.  
 KRAYSL, C (1930a) *J infect Dis.* 47 293 322, 308 (1930b) *J Bact.* 19 113  
 KRAYSL, G. and GORDON M. (1930) *J infect. Dis.* 47, 303 318  
 KOCH, R. (1881) *Mitt. PeichgerundhAmt.* 1, 234  
 KOCH, P. and WOLFFHUEL, G. (1881) *Mitt. PeichgerundhAmt.* 1, 301  
 KOCH R., GARTKY and LOETTLER. (1881) *Mitt. PeichgerundhAmt.* 1, 322  
 KONRICH F (1938) *Die bakterielle Keimtötung durch Wärme.* Ferd. Enke Verlag  
 Stuttgart  
 KRONIG B. and PAUL, T. (1897) *Z Hyg InfektKr* 25, 1  
 LAMAR, R V (1911) *J exp Med.* 13 1 340  
 LARDETTE, M. DE. (1918) *Ann Inst. Pasteur.* 32, 170  
 LEA D E. and HAINES P B (1940) *J Hyg., Camb.* 40, 162  
 LEA, D E., HAINES, P B. and BRETSCHER, F. (1941) *J Hyg., Camb* 41, 1  
 LEA D E., HAINES, P B. and COULSON C. A. (1936) *Proc roy Soc., B.* 120, 4 (1931)  
*Ib d.* 123, 1  
 LENTZ, F A. (1930) *Zbl Bakt.* 126, 508  
 LEVIN B S. and LEVIN EL I. (1935) *C R Acad. Sci.* 200 563  
 LIT S C. and YEN A. C. H. (1934) *Proc Soc. exp Biol., U.S.* 32, 485.  
 LOCKEMAN G BAR, F. and TOTZICK, W. (1941) *Zbl Bakt* 147 1  
 LOEB J (1899) *Pflug Arch ges Physiol.* 75 303 (1900) *Amer J Physiol.* 3, 32  
 LOEB J. and NORTROP J H (1917) *J biol Chem.* 32, 103  
 MACADAM A (1900) *Proc roy Soc., B* 66, 180 339

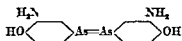


- MACFADYEN, A and ROWLAND S D (1900) *Proc roy Soc, B*, 66, 488
- McINTOSH, J and SELBIE, F R (1942) *Lancet*, ii, 750
- McMASTER, P D (1919) *J infect Dis*, 24, 378
- MADSEN, T and NYMAN, M (1907) *Z Hyg Infektkr*, 57, 398
- MALLMANN, W L, BOTWRIGHT, W I and CHURCHILL, E S (1911) *J infect Dis* 69, 215
- MANFOLD, M C (1941) *Brit J exp Path*, 22, 111
- MATHEWS, A P (1904a) *Amer J Physiol*, 10, 290, (1904b) *Ibid*, 11, 455, (1905) *Ibid* 12, 419, (1906) *J infect Dis*, 3, 572
- MAYER, H (1925) *Zbl Bakt*, 94, 238
- MILLER, B I and BAKER, Z (1940) *Science*, 91, 624
- MIVERVINI R (1898) *Z Hyg Infektkr*, 29, 117
- MOORE, H N and KIRSTEN, H (1937) *J Bact*, 33, 615
- MORTON, H E. and KLAUDER, J V (1944) *J Amer med Ass* 124, 1195
- MÜNDEL, O (1937) *Z Hyg Infektkr*, 120, 267
- NEUFELD, F and SCHREMAN, O (1943) *Z Hyg Infektkr*, 124, 751
- NICHOLS, H J (1920) *J Lab clin Med*, 5, 502
- NOGUCHI, H (1907) *Biochem Z*, 8, 327
- NORTON, J F and HSU, P H (1916) *Z Hyg Infektkr*, 18, 180
- OAG, R A (1940) *J Path Bact*, 51, 137
- OTTEV, L (1930) *Zbl Bakt*, 116, 199, (1932) *Trans Far East Ass trop Med 8th Congr Bangkok*, 1930 p 89
- OZZANO, T and RE, C (1937) *G Batt Immun*, 19, 535
- PAIC, M, DEUTSCH, V, and BORCHLA, I (1935a) *C R Soc Biol*, 119, 1003
- PAIC, M, HABER, P, VOET, J, and FLIASZ, A (1935b) *C R Soc Biol*, 119, 1001
- PAUL, T. (1909) *Biochem Z*, 18, 1
- PAUL, T and PRALL, F (1907) *Arb ReichsgesundhAmt*, 26, 73
- PAUL, T, BIRSTEIN, G, and REUSZ, A (1910a) *Biochem Z*, 25, 367, (1910b) *Ibid* 29, 202
- PERDRAU, J R, and TODD C (1933a) *Proc roy Soc, B*, 112, 277, (1933b) *Ibid* 112, 288
- PHELPS, E B (1911) *J infect Dis*, 8, 27
- PILOD and CONVELLE (1932) *C R Acad Sci*, 194, 497
- PRINOLE (1750) *Philos Trans*, 46, 480 525
- PROCHOWNICK, L and SPAETH, F (1890) *Dtsch med Wschr*, 16, 564
- PRUDHOMME, R O (1937) *C R Soc Biol*, 126, 259
- PEGGLEY, A T, ODDIE, T H, and EDDY, C E (1935) *Proc roy Soc, B*, 118, 276
- RAAB, O (1900) *Z Biol*, 39, 524
- REICHEL, H (1909) *Biochem Z*, 22, 149
- REICHENBACH, H (1908) *Z Hyg Infektkr*, 59, 296, (1911) *Ibid*, 69, 171
- REID, J D (1932) *Amer J Hyg*, 16, 540
- RENTSCHLER, H C, NAGI, R and MOCROMSEFF, G (1941) *J Bact*, 41, 745
- Report (1942) "The Prevention of 'Hospital Infection' of Wounds" *Med. Res Coun., Lond., War Memo*, No 6
- Report (1944) "The Control of Cross Infection in Hospitals." *Med Res Coun., Lond., War Memo*, No 11.
- RIDEAL, S and WALKER, J T A (1903) *J R sanit Inst*, 24, 421
- RIEDER, H (1902) *Münch med. Wschr*, 49, 402
- RISHWORTH, H R (1939) *Brit med J*, ii, 574
- RISLER, J (1936) *C R Acad Sci*, 203, 517
- RITCHIE, J (1899) *Trans path Soc Lond*, 50, 250
- RIVERS, T M, SHADFL, J E and CHAMBERS, L A (1937) *J exp Med*, 65, 677
- ROBERTSON, T B (1914) *J Hyg, Camb*, 14, 143
- ROCKWELL, G E and EBERTZ, F G (1924) *J infect Dis*, 35, 573
- RÖMER C (1898) *Münch med Wschr*, 45, 298
- ROUT, E (1887) *Ann Inst Pasteur*, 1, 445
- ROUYER, M and SERVIGNE M (1938) *Ann Inst Pasteur* 61, 565
- RUSSO S D, ALBERT, A and MAXWELL, M (1942) *Brit J exp Path*, 23, 67
- RUSS, V. (1904) *Zbl Bakt*, 37, 115, 220
- RUSSELL, D S and BECK, D J K (1941) *Brit med J*, i 112
- RUSSELL, D S and FALCONER, M A (1941) *Brit J Surg*, 28, 472 (1943) *Lancet*, i, 580
- SALLP, A J, McOMIE, W A., SHECHMISTEF, I L and FOORD, D C (1933) *J Bact*, 37, 630
- SAVAGE, R M. (1910) *Quart J Pharm*, 13, 237
- SCHERP, H W and CHAMBERS, L A (1936) *Proc Soc exp Biol, N Y*, 35, 495
- SCHREIBLER, E (1895) "Die Bedeutung des Moleculärzustandes der wassergetrennten Desinfektionsmittel für ihren Wirkungswert" *Strassburg*
- SCHMIDT, C. L. A and NORMAN G F. (1920) *J infect Dis*, 27, 40
- SCHOOF, G (1935) *Zbl Bakt*, 134, 14
- SCHROETTER, H V (1927) *Zbl Bakt*, 104, Beibl p. 205
- SELLARD, A W (1918) *J med. Res*, 38, 293

- SHEARER, C (1919) *J Hyg., Camb.*, 18, 337  
 SHEPHERD J M and HOLM G E. (1922) *J Bact.*, 7, 465.  
 SNYDER, M L and LICHSTEIN H C. (1940) *J infect Dis.*, 67, 113.  
 SOBERNHEIM, G (1943) *Schweiz. med Wochr.*, 73, 1250 1304 1333.  
 SPENCER, R. R. (1934) *Publ. Hlth Rep Wash.*, 49, 183 (1935) *Ibid.*, 50, 1642.  
 SPOONER, E T C and TURNBULL, L. H. (1943) *Bull War Med.*, 2, 345  
 STEARN E W and STEARN A. E. (1926) *J Bact.*, 11, 345 (1925) *Univ Mo Stud.*, 3, No 2 1  
 SUESS E (1908) *Z Tuberk.*, 12, 450  
 SZYMANSKI, W T and HICKS P A. (1932) *J infect Dis.* 50, 1  
 TAKAHASHI, B \ and CHRISTENSEN, R. J. (1934) *Science* 79, 413.  
 TAPPEINER, H v (1900) *Munch med Wochr.*, 47, 5  
 TAPPEINER, H v and JONLEBAUER, A. (1904) *Munch med Wochr.*, 51, 737  
 TCHAHOTINE, S. (1921) *Ann Inst Pasteur* 35 321  
 TETSUMOTO S. (1937) *Jap J exp Med.*, 15 1  
 THIELE, H and WOLF K. (1906) *Arch Hyg.*, 57, 29. (1907) *Ibid.*, 60, 29  
 THRESE, J C. and BEALE, J F. (1910) *Lancet*, ii, 1849  
 TILLEY F W (1939) *J Bact.*, 38 499  
 TILLEY F W and SCHAFER, J M. (1936) *J Bact.*, 12, 303  
 TOPLEY W W C (1915) *Brit. med J.*, i, 237  
 TUNG T (1935) *Proc Soc exp Biol.*, \ Y., 33, 328  
 UNDERWOOD W B (1934) "Textbook of Sterilisation. Amer Sterilizer Co., Erie Pa  
 WALKER, J E. (1934) *J infect Dis.* 35 55 (1925) *Ibid.*, 37 181. (1936) *Ibid.*, 38, 127  
 WARD H. M. (1897) *Proc roy Soc.*, 52, 393  
 WATSON H. E. (1908) *J Hyg., Camb.*, 8, 536  
 WELCH H and BREWER, C M. (1942) *Amer J publ Hlth.*, 32, 261  
 WESBROOK, F F (1896) *J Path. Bact.*, 3, 70.  
 WHIFFLE, O C and MAYER, A. (1906) *J infect. Dis.*, Supp 2, p. 76.  
 WILLIAMS, R., CLAYTON-COOPER, B., DUNCAN J M and MILES, E. M. (1943) *Lancet*, i, 522.  
 WINSLOW C E. A and DOLLOFF A. F. (1928) *J Bact.*, 15, 67  
 WINSLOW C E. A. and FALK, I S. (1923) *J Bact.*, 8, 23  
 WINSLOW C E. A. and HOTCHKISS, M. (1922) *Proc. Soc. exp Biol.*, \ Y., 19, 314  
 WINSLOW C E. A. and LOCHBRIDGE, E. E. (1906) *J infect. Dis.*, 3, 54  
 WIRGIN G (1902) *Z Hyg InfektKr.*, 40 307 (1904) *Ibid.*, 48, 149  
 WITHELL, E. P. (1942a) *J Hyg., Camb.*, 42, 124. (1942b) *Ibid.*, 42, 339  
 WOODRUFF L. L. and BENZEL, H. H. (1909) *Amer J Physiol.*, 25, 190  
 WRIGHT E. V. and KERSTEN H. (1937) *J Bact.*, 34, 639  
 WYCKOFF R. W G (1930a) *J exp Med.*, 52, 435 (1930b) *Ibid.*, 52, 769 (1932) *J gen. Physiol.*, 15 351  
 WYCKOFF R. W G and LUTY B J. (1931) *Endocrinology* 17, 1171  
 WYCKOFF R. W G and PETERS T M. (1930) *J exp Med.*, 51, 921  
 YEN A. C H. and LIT S C. (1934) *Proc. Soc. exp Biol.*, \ Y., 31, 1250  
 ZWAARDEMAKER, H. (1918) *Amer J Physiol.*, 45, 147 (1919-20) *J Physiol.*, 53, 273

to a fairly constant pattern, the ratio is a valuable index of therapeutic efficiency. For example, using rats suffering from an experimental trypanosomiasis, the ratio of the minimal lethal dose to the minimal curing dose is unity for arsenic acid and 4 for atoxyl. Arsenic acid is clearly useless, and atoxyl not very satisfactory. Under the same conditions, arsphenamine has a ratio of 37.

Although a great deal of precise chemical investigation accompanied the search for chemotherapeutic agents, we have to-day little knowledge that will enable us to predict what type of substance is likely to prove effective against a given parasite. The successful discoveries have been largely the result of trial and error. Widely different substances prove to be similar in their efficacy on one type of infection, though once a type of substance has proved efficacious it is usually possible to relate its molecular structure to its chemotherapeutic activity. For example, trivalent arsenic attached to a benzene ring is most effective against trypanosomes when there is an amino group in the *para* position, and against spirochaetes when there is a hydroxyl group in the *para* position and an amino group in the *ortho* position. Thus, arsphenamine has the formula



Another general point of importance is that the drug does not necessarily act in the form in which it is administered. Atoxyl,  $\text{H}_2\text{N} \text{---} \text{C}_6\text{H}_4 \text{---} \text{AsO}_3\text{HNa}$ , for example, is a pentavalent arsenical, but is reduced to the active trivalent form in the body.

It is supposed that chemotherapeutic drugs act either by weakening the parasite so that it is easily eliminated by the defence mechanisms of the tissues, or by killing the parasite outright. Experience with experimental trypanosomiasis shows that, if small doses of the drug are given, the "weakening" process may not only be ineffective, but that subsequent generations of the parasite may develop immunity to the drug and become "drug fast."

### CHEMOTHERAPEUTIC AGENTS ACTIVE AGAINST BACTERIA

The development of chemotherapy in spirochetal and protozoal infections in the early years of the century had no parallel in bacterial infections. In 1911 Morgenroth and Levy (1911) were able in a certain proportion of cases to protect mice against pneumococcal infection by ethyl dihydrocupreine, but apart from this there is little to note until 1930, when Domagk (1930a, b) reported that the compound sulphonamido-crysoidin (Prontosil) would cure streptococcal infection in mice. The curing dose was between one-tenth and one-fiftieth of the tolerated dose for the acute infection that followed the intraperitoneal injection of ten lethal doses of *Str. pyogenes*, and between one hundredth and one five-hundredth for a less acute infection. The interval between infection and the start of successful therapy was in some cases as long as three days. Domagk's observations were quickly confirmed by French and British workers (see Levaditi and Vaisman 1935a, b; Nitti and Boret 1930, Buttle 1930, Colebrook and Kenny 1936). Sulphonamido-crysoidin, however, was active only in the animal body, and had no effect on streptococci *in vitro*. Trefouel and his colleagues (1930) supposed that the *in vivo* action was due to a breakdown product of the drug, and showed that the

sulphonamide half of the molecule was active both in the test tube and the infected mouse (see also Gosselinet *et al* 1936, Buttle, Gray and Stephenson 1936). That sulphonamido-crysoidin was broken down to yield the active sulphonamide in the body was demonstrated by Fuller (1937) and *para*-aminobenzene sulphonamide was thereby established as the active principle of prontosil.

to be proved active, and was successful in the treatment of pneumococcal, meningococcal and gonococcal infections. Sulphathiazole and sulphadiazine were improvements particularly in respect of *Staph aureus* infection, and as an example of the selectivity that a drug may display it is interesting to note that sulphadiazine

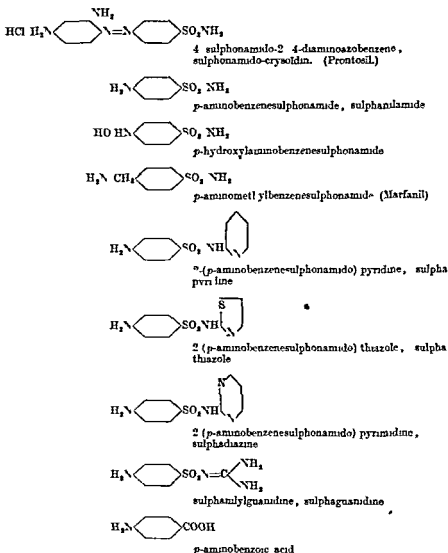


FIG. 27

proved to be effective against Friedlander's bacillus, which had hitherto proved insusceptible to sulphonamides

Sulphaguanidine is included as an example of a modification designed to be antibacterial soluble and yet absorbed with difficulty from the alimentary tract, suitable, that is, for the treatment of infections of the intestinal tract

**Definition of Some Terms**

The terminology of antibacterial effects produced by all kinds of compound is confused. The failure of an inoculum of bacteria to grow after exposure to a given compound is referred to sometimes as a bacteriostatic effect, sometimes as a bactericidal effect. The agent producing this effect may be called variously an antiseptic, a bacteriostatic, a bacterial antagonist, an inhibitor or a suppressor of growth, and so forth. Agents which protect the bacterium from the antibacterial action may be referred to as a deviating substance, an antagonist, a reversing substance, or an inhibiting substance.

The bacteriostatic action varies directly as the concentration of the drug and inversely as the concentration of the bacteria. This latter relation cannot be simply explained in terms of the amount of sulphonamide available per bacterium, for in any given test only a small fraction of the drug is taken up by the bacteria (McIntosh and Whitby 1939, Kohn and Harris 1941, Rose and Fox 1942, Pike and Acton 1942). With organisms that grow optimally at 37° C a rise in temperature above this level greatly increases the sulphonamide activity. White (1939) found that only 1/100th of the amount active at 37° C. was required at 39° C. The degree of bacteriostatic action is greatly influenced by the medium used for the test. In general, the richer the medium the less effective the drug. From the point of view of precision work in bacterial metabolism the variability of common ingredients of routine media like peptone, tissue extracts and blood fluids is enormous. Nevertheless, in carefully prepared routine media, comparative tests of sulphonamide inhibition are valid, though absolute measures are unreliable, owing to the variable amounts of substances that antagonize the action of the drug (see for example MacLeod 1940, Lewis and Snyder 1940, MacLeod and Mirick 1942). Antagonizers are present in peptone (Lockwood 1933, Weld and Mitchell 1939) in serum, and in extracts and hydrolysates of animal tissue (Landy and Dicken 1942, Lewis 1942). Reliable results are possible only in defined media where the antagonistic effect of each ingredient may be determined with certainty. It is generally believed that in low concentration the sulphonamide drugs exert only a bacteriostatic effect but the careful observations of Wolff and Julius (1939) suggest that they are lethal to actively dividing organisms, and that at other stages of the culture cycle bacterial growth is merely inhibited. The analogy to penicillin is very striking (see p. 183).

#### The *In Vitro* Resistance of Bacteria to Sulphonamides.

The resistance of bacteria to a sulphonamide is measured in terms of the minimum concentration of the drug that prevents the growth in a defined medium of a standard inoculum of the strain under test. Without careful standardization of the medium and conditions of test (see, for example, Strauss *et al* 1941) these measures serve only as rough indications. Comparative tests, however show that, although bacteria as a whole are generally susceptible to the action of one or other of the sulphonamides there are wide variations in susceptibility, both from one species to another, and from strain to strain within a species. Thus although the species *Str. pyogenes* is relatively susceptible to sulphanilamide, and *Staph. aureus* relatively resistant yet in an examination of a large number of strains of both species, there would be a wide overlap of susceptible strains of *Staph. aureus* and resistant strains of *Str. pyogenes*. Strain variations in natural resistance have been noted by a number of observers (see for example, Green 1940, Green and Bielschowsky 1942, Poston and Organ 1942, Felsenfeld 1943) and must be taken into account in the therapy of infections by sulphonamides, since clearly the assumption of a general level of susceptibility for a given species may have serious consequences if applied consistently to the treatment of all infections by that species.

Of even greater importance is the demonstration that resistance to sulphonamides can be acquired in the test tube. Bacteria may be trained by serial subculture in media containing non bacteriostatic concentrations of the sulphonamide (Green 1940, MacLeod 1940, Strauss *et al* 1941, Schmidt *et al* 1942, Kirby and Rantz 1943, Kirby 1943, Lankford *et al* 1943, Harris and Kohn 1943, McIntosh

and Selbie 1943b) The degree of resistance acquired appears to depend mainly upon the strain, but partly upon the culture medium in which the training takes place The concentration of drug required to induce resistance is proportional to the bacteriostatic potency of the drug used, and the resistance, once acquired, is sufficiently established as a character of the strain to withstand serial subculture in media free from the sulphonamide.

Resistance can also be induced *in vivo* (MacLean *et al* 1939, Schmidt and Hilles 1940, Frisch and Price 1941, Vivino and Spink 1942) For example, Schmidt, Sesler and Dettweiler (1942) infected sulphapyridine-treated mice with virulent pneumococci, pooled the cultures from the heart blood of the mice which died and with the pooled cultures infected another group of sulphapyridine-treated mice Two to three repetitions of this process caused an appreciable increase of *in vitro* resistance, and three to nine repetitions produced a maximal degree of resistance The acquired resistance was sufficiently established to withstand 215 passages through normal mice The effect of the drug does not appear to be one of selective breeding out of naturally resistant variants, but a direct action on all the metabolizing cells in the culture



an aromatic amino compound led Woods (1940) to test the antagonizing action of *p*-aminobenzoic acid. It proved to be a powerful antagonist, in a defined basal medium containing streptococci, one molecule of the acid antagonized the bacteriostatic action of 26 000 molecules of sulphanilamide. The probable presence of *p*-aminobenzoic acid in yeast was later established by Rubbo and Gillespie (1940) Blanchard (1941) and McIlwain (1942b). (See also Ratner *et al* 1944.)

Subsequent observers confirmed Woods' findings (see Green and Bielschowsky 1942 Rubbo and Gillespie 1942, Wood 1942) and showed that the ratio of molar bacteriostatic concentrations of sulphanilamide to the antagonizing concentration of *p*-aminobenzoic acid varied between 26 000:1 and 1000:1. The acid antagonizes all the sulphonamides and Wood (1942) observed that the greater the bacteriostatic activity of the sulphonamide, the smaller is the ratio, i.e. the greater its ability, molecule per molecule, to counteract the antagonizing effect of *p*-aminobenzoic acid.

### The Woods-Fildes Hypothesis

The structural similarity of sulphanilamide and *p*-aminobenzoic acid led Woods (1940) and Fildes (1940a) to propound an hypothesis of sulphonamide action that has proved exceedingly fruitful in the investigation of antibacterial substances. The Woods Fildes hypothesis made precise the previously held notion that sulphonamides in some way interfered with an essential metabolic function of the bacterium. Woods and Fildes contend that *p*-aminobenzoic acid is an essential metabolite for susceptible bacteria whose growth is inhibited when sulphonamides by reason of their structural similarity to the natural metabolite, block the enzyme system concerned with this particular part of the essential metabolism of the organism. Inhibitor and antagonist compete for the enzyme system, and bacteriostasis results when the inhibitor is successful in the competition.

The degree of competition in a given organism may be expressed by the ratio  $C_i/C_M$ ,  $C_i$  being the molar concentration of inhibitor that is just bacteriostatic in the presence of a molar concentration  $C_M$  of the corresponding metabolite (McIlwain 1942b). Thus the ratio, for which McIlwain proposed the name "antibacterial index", was found by Woods to be 26 000 for *Str. pyogenes*, sulphanilamide and *p*-aminobenzoic acid. Among other things, it can be used to predict the chemotherapeutic efficiency of a drug, if the concentration of the metabolite (i.e. the antagonist) in the tissues of the animal is known (see below, p. 164).

The facts of *p*-aminobenzoic acid antagonism to the sulphonamides have been amply confirmed both *in vitro* and in the living animal. The Woods Fildes hypothesis of its action is widely accepted. However, it is not consistent with all the phenomena of sulphonamide action, and for this reason we shall examine both the corroborative evidence and the inconsistencies in some detail.

### *para*-aminobenzoic Acid as Essential Metabolite

In two species of bacteria there is no doubt that *p*-aminobenzoic acid is an essential metabolite for it is demonstrably an essential nutrient. Both *Cl. acetobutylicum* (Rubbo *et al* 1941 Rubbo and Gillespie 1942 Lampen and Peterson 1941) and *Acetobacter suboxydans* (Lampen *et al* 1942) require the acid for growth and both are susceptible to an inhibiting action of sulphonamides that is reversed by excess of the acid (see also Kuhn and Schwartz 1941 Wiedling 1941 Isbell 1942 Landy and Dicken 1942). In one other species a soil bacillus isolated by Mirick (1943) an enzyme system capable of oxidizing *p*-aminobenzoic acid has been demonstrated, the growth of the organism was inhibited by sulphapyridine.

and pantoyltaurine in which the  $-\text{COOH}$  group is replaced by  $-\text{SO}_2\text{H}$  were found to be similarly related (see also Barnett and Rolinson 1942). Recently McIlwain (1943a) has further elucidated the action of pantoyltaurine by comparing the metabolism of resistant and susceptible strains of *C. diphtheriae* and *Str. pyogenes*. Pantothenic acid is an essential nutrient for susceptible strains of both species. Resistance could be induced in *C. diphtheriae* by serial subculture either in media containing sub-inhibiting amounts of pantoyltaurine or in media lacking pantothenate, and all resistant strains were shown to have developed the power to use  $\beta$ -alanine which they presumably synthesized into pantothenic acid. Resistant *Str. pyogenes* had developed no ability to synthesize pantothenate. They were however susceptible to salicylate and pantoyltaurine suggesting that resistance to pantoyltaurine alone is due to the possession of an alternative metabolic process which in turn is susceptible to salicylate (see also Ivanovics 1942). McIlwain and Hughes (1944) have also shown that pantothenate is metabolized by *Str. pyogenes* during glycolysis but that neither absence of pantothenate nor its antagonization by pantoyltaurine affects the glycolysis. On the other hand either the absence of pantothenate or the addition of pantoyltaurine or the inhibition of glycolysis will inhibit growth. These facts are most conveniently explained by assuming that both glycolysis and pantothenate are necessary for the formation of an essential growth metabolite in the cell.

Pantoyltaurine is of particular interest since McIlwain (1942a) was able to collect data from which he predicted its *in vivo* action. The antibacterial index for *Str. pyogenes* was as low as 500, the molar concentration of pantothenic acid likely to be found in animal tissues lay between  $10^{-6}$  and  $10^{-5}$ , and the molar concentration of pantoyltaurine required to inhibit the streptococci in the presence of this amount of pantothenic acid was well below the maximum tolerated concentration. The prediction that pantoyltaurine would be chemotherapeutic *in vivo* was amply confirmed by McIlwain and Hawking (1943). Rats were protected against 10 000 lethal doses of *Str. pyogenes*, and the protective effect was abolished by artificially raising the pantothenate concentration of the blood. The drug was ineffective in mice in whose tissues the natural pantothenate content is higher.

Woolley and White (1943) have also demonstrated a resistance to an antibacterial substance of the Woods-Fildes type which, like that of *Str. pyogenes* to pantoyltaurine, does not depend on an ability to synthesize increased amounts of the essential metabolite. Pyrimthiamine the pyridine analogue of thiamin inhibited the growth of yeast and bacteria in direct proportion to their natural thiamin requirements. But the resistant strains produced neither thiamin nor any other pyrimthiamine antagonist in detectable amounts. Failure to demonstrate increased synthesis of the essential metabolite by drug resistant organisms does not however invalidate the Woods-Fildes hypothesis. The resistant organism may develop a different unrelated path along which to carry its essential metabolic processes.

Further examples of antagonism are described in connection with other antibacterial agents. We may note here for example that polyamines like triethylenetetramine and tetraethylenepentamine antagonize the bacteriostatic effect of mepacrine on *Bact. coli* (Silverman and Evans 1943) and of propamidine on *L. casei* and *Str. lactis* (Snell 1944). McIlwain (1941b) has applied the Woods-Fildes hypothesis to the antagonism of nucleic acid and related substances to acriflavine. As in the sulphonamides, there was a constant ratio between inhibitor and antagonist. Amino-acid concentrates, especially in the presence of artificial hydrogen carriers like methylene blue were also antagonistic, but with increasing concentrations of inhibitor, increasing concentrations of antagonists became ineffective. McIlwain concludes that the acriflavine competes with nucleic acid

and like substances for an essential enzyme system and that the amino acid concentrates are effective because they are substrates or products of the essential enzyme, which to some extent can be replaced by artificial hydrogen carriers.

#### Other Sulphonamide Antagonists

There is a large group of substances capable of antagonizing sulphonamides. For example, antagonizers are found in necrotic tissue and abscesses (Lockwood *et al* 1938), in pus (MacLeod 1940), and, as we have already noted in the complex organic ingredients of routine culture media (see Strauss and Finland 1941). In some of these the active principle may be *p*-aminobenzoic acid though it is unproven. Among other substances structurally unrelated to *p*-aminobenzoic acid that are reported as antagonists are methionine (Kohn and Harris 1941), co enzyme I (West and Coburn 1940), nicotinic acid and nicotinamide (Dorfman *et al* 1940), urethane (Johnson 1942), purine bases (Martin and Fisher 1942, Snell and Mitchell 1942) and certain amino acids.

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### Other Sulphonamide Antagonists

There is a large group of substances capable of antagonizing sulphonamides. For example, antagonizers are found in necrotic tissue and abscesses (Lockwood *et al* 1933), in pus (MacLeod 1910), and, as we have already noted, in the complex organic ingredients of routine culture media (see Strauss and Finland 1941). In some of these the active principle may be *p*-aminobenzoic acid, though it is unproven. Among other substances structurally unrelated to *p*-aminobenzoic acid that are reported as antagonists are methionine (Kohn and Harris 1941), co-enzyme I (West and Coburn 1940) nicotinic acid and nicotinamide (Dorfman *et al* 1940), urethane (Johnson 1942), purine bases (Martin and Fisher 1942, Snell and Mitchell 1942) and certain amino acids.

Kohn and Harris postulated that a methionine phase of metabolism was secondary to a phase for which *p*-aminobenzoic acid was a catalyst and that when the latter was blocked by sulphanilamide, added methionine enabled the essential metabolism to carry on. With regard to co-enzyme I and nicotinamide Strauss, Dingle and Finland (1941) could not confirm West and Coburn's observation that co-enzyme I antagonized the inhibition of staphylococci by sulphapyridine but found a partial antagonism of sulphaguanidine by a combination of uracil, pyruvate and adenylic acid. Koser and his colleagues (Dorfman and Koser 1942, Berkman and Koser 1943) have defined some of the conditions in which co-enzyme I and nicotinamide are active. Using a strain of *St. aureus* in a basal medium they found that these substances antagonized the inhibition of its respiratory activity by sulphapyridine and sulphathiazole but not inhibition by sulphanilamide, sulphydrazine, sulphaguanidine or sulphacetamide. *p*-Aminobenzoic acid was antagonistic to all the sulphonamides tested though its effect on sulphapyridine and sulphathiazole was less than that of nicotinamide. They concluded that the radicle attached to the sulphonamide part of the molecule could also affect the enzyme systems of the cell, and that in this case the structural similarity of the pyridine or thiazole rings in sulphapyridine and sulphydrazine with nicotinic acid was responsible for inhibition of respiratory activity dependent upon nicotinamide (pyridine-carboxamide) and co-enzyme I (diphospho-pyridine nucleotide) (see also v. Furk 1943, Teplov *et al* 1943). Antagonism by amino-acids is illustrated by a recent report of Savag and Green (1944) who reversed the sulphonamide inhibition of *Staph. aureus* in a medium containing glucose and certain amino acids by the addition of tryptophan.

### Alternative Hypotheses of Sulphonamide Action

The success of the Woods Fildes hypothesis in predicting the inhibitory activity of analogues of growth factors does not necessarily confirm the truth of the hypothesis in relation to sulphonamide-action, though it provides a strong incentive to give the hypothesis priority of place. The student is referred to the review of Henry (1943) for a full discussion of the objections to the hypothesis. We shall do no more than summarize the main points.

Neither the competitive relationship between sulphonamides and *p*-aminobenzoic acid, nor their structural similarity, necessarily signifies a competition for an enzyme system utilizing *p*-aminobenzoic acid. In the first place *p*-aminobenzoic acid antagonizes sulphonamide inhibition of systems in which it can play no essential part; e.g. the carboxylase system of *Staph. aureus* (Savag *et al* 1943), the digestion of starch by diastase, and the adsorption of methylene blue to charcoal (Eyster 1943). Moreover, the rates of inhibition and antagonization may differ

markedly, a fact difficult to reconcile with the hypothesis of direct competition between the two (Hirsch 1944) (See p 165, and Johnson *et al* 1944, for competition of relatively unrelated structures.) In the second place, sulphanilamide may be antagonized as we have seen, by substances that have no structural relation to it.

The hypothesis postulates that *p*-aminobenzoic acid is an essential metabolite in a wide variety of plants and animals (see Fildes 1940, 1941). That it is a growth factor for certain bacteria and animals is no more than suggestive, at present its wide distribution and a similar distribution of enzyme systems that utilize it remain unproven. Two other of Henry's points the limited value of the evidence of (a) increased *p*-aminobenzoic acid production by sulphonamide-resistant strains and (b) analogous systems of growth factors and inhibitors, we have already dealt with. There remains the most cogent objection, that animal and bacterial respiratory systems are inhibited by sulphanilamide. In two systems, the sea urchin's egg (see Henry 1943) and perhaps *Br. tularensis* (Tamura 1941) the inhibition is not reversed by *p*-aminobenzoic acid. Sulphonamides appear to have a direct action on the respiratory enzymes of bacteria, both aerobic and anaerobic (Sevag and Shelburne 1942, Dorfman and Koser 1942, Berkman and Koser 1943) on bacterial dehydrogenase (Macleod 1939, Fox 1942), and on cocarboxylase (Sevag, Shelburne and Mudd 1942, Sevag *et al* 1943). Sevag and his colleagues conclude that sulphonamides inhibit oxidative enzymes (see also Sevag and Green 1944) and therefore the growth of the bacteria. Henry groups the sulphonamides with 'indifferent' cell inhibitors like narcotics, inhibiting a specific fraction of the total oxidative reactions of the cell upon which cell division depends. Like the narcotics, the sulphonamides stimulate in low, and inhibit in higher, concentrations (Finklestone-Savill *et al* 1937, Green and Bielschowsky 1942, Lamanna and Shapiro 1943). Like narcotics, they act upon a wide variety of tissues. For example, they inhibit the growth of tissue cultures of tomato plants (Bonner 1942) and wheat and oat seedlings (Brian 1944, Jones 1944), and they inhibit the reproductive division of flagellates (Lwoff *et al* 1941). The specificity of *p*-aminobenzoic acid as an antagonist does not imply that this substance, or any other antagonist, necessarily acts by specific interference. The antagonist may act as a non specific growth stimulant (see, for example, Rantz and Kirby 1944a), though it is pertinent to note that Lynch and Lockwood (1941) distinguished clearly between the antagonistic action of peptone in a human serum medium which was due to growth stimulation and that of *p*-aminobenzoic acid, which was not. Alternatively, antagonizers may combine directly with the inhibitor, forming an inactive complex. This is unlikely to be the case with *p*-aminobenzoic acid and a sulphonamide, for they do not react in the absence of bacteria. The antagonism of mercapto compounds to the disinfectant action of HgCl<sub>2</sub> (Fildes 1940b) may be of this nature. Again, cationic antiseptics of the long-chain fatty acid type, which presumably act by disorganizing the lipid membrane of the bacterial cell, are antagonized by the addition of phospholipins (Baker, Harrison and Miller 1941). Finally, antagonizers may shield the susceptible enzyme system from the inhibition, without blocking the enzyme action. Here again cationic antiseptics provide a model for the hypothesis. (See also Penicillic Acid, p 178). Valko and DuBois (1944) reversed the antibacterial action of a highly toxic cation, *N*-dodecyl dimethyl ammonium chloride, displacing it by the addition of a relatively non toxic cation like *N*-hexadecyl dimethyl ammonium

The necessity for the *para* position holds also for the antagonistic effect of *p*-aminobenzoic acid. As Rubbo and Gillespie (1942) showed, *o*- and *m*-aminobenzoic acid are poor antagonizers, and very poor essential nutrients for *Cl. acetobutylicum* as compared with the *para* form.

The S of the radicle can be replaced by arsenic, carbon phosphorus, selenium and tellurium though not all the compounds so formed are antibacterial, and not all the antibacterial compounds are antagonized by *p*-aminobenzoic acid (Rosenthal *et al* 1939, Green and Bielschowsky 1942 Hirsch 1942). The substance 4-(4-aminobenzyl) (H<sub>2</sub>N C<sub>6</sub>H<sub>4</sub> CO CO C<sub>6</sub>H<sub>4</sub> NH<sub>2</sub>) is sulphur free, and yet behaves as an active sulphonamide, being antagonized by *p*-aminobenzoic acid (Kuhn Moller and Wendt 1943).

As will be seen from Fig. 27, most of the sulphonamides in general use are derived from sulphanilamide by the introduction of substituents—notably heterocyclic radicals—into the amide group. Methylation of the pyrimidine ring of sulphadiazine in the 4 and 4-6-position yields sulpha merazine and sulpha mezathine respectively which have a similarly high degree of activity. The compound N-(3,4-dimethyl benzoyl) sulphanilamide



is also active but alteration of the position of the methyl groups in the benzoyl residue completely destroys the activity (Langer and Martin 1943).

The bacteriostatic potency of the sulphonamides varies directly with their ability to counteract the antagonistic effect of *p*-aminobenzoic acid, i.e. the stronger the drug the lower its antibacterial index. At about pH 7, that of sulphanilamide lies between 26 000/1 and 1000/1. At this pH, however, *p*-aminobenzoic acid is almost fully ionized, sulphanilamide very little, and the ratio in terms of anions may be near unity (Fox and Rose 1942). The acid dissociation constants (*K*<sub>a</sub>) of the drugs increase with activity, that of sulphanilamide being  $2.2 \times 10^{-11}$ , sulphapyridine  $5.1 \times 10^{-9}$ , and sulphathiazole  $6.2 \times 10^{-8}$ , the last approaching most closely to *p*-aminobenzoic acid, whose *K*<sub>a</sub> is  $1.2 \times 10^{-3}$  (Schmelkes *et al* 1942). Albert and Goldacre (1942) on the other hand suggest that the activity of sulphanilamide and *p*-aminobenzoic acid may be a function of their feeble basicity, both have a *K*<sub>b</sub> of about  $10^{-11}$ . The association between concentration of anions and activity is not, however, constant. Maximum activity may be displayed at a pH at which the solution must contain a mixture of dissociated and undissociated forms (Cowles 1942 Brueckner 1943). It is suggested that both forms are necessary for activity, the non ionized form alone being capable of penetrating the cell but the ionized form being an active inhibitor. A similar explanation may apply to Bell and Roblin's (1942) data, which showed that the relation of activity with a high *K*<sub>a</sub> value held good only for a given range of compounds. Above a certain point, activity fell off with increasing *K*<sub>a</sub>. Bell and Roblin, however, postulated that activity depended on the electronegativity of the SO<sub>2</sub> group, as well as on ionizing capacity. Compounds with the highest *K*<sub>a</sub> are those with substituents on the amide nitrogen possessing a high electron-attracting power and as a consequence increasing activity due to increasing acid strength may be counteracted by decrease in the electronegativity of the SO<sub>2</sub> group. Kumler and Daniels (1943) discuss in some detail the association of activity with the polarity of the molecule and suggest that it is immaterial whether the sulphona-

mide acts as an anion, cation or neutral molecule so long as the structure of the compound permits a separation of charge with the formation of a quinonoid structure by movement of bonds after ionization of the amino group. The essence of the polarity in this connection is the electropositivity of the  $\text{NH}_2$  group.

The general form of the sulphonamides is  $\text{H}_2\text{N}-\text{C}_6\text{H}_4-\text{SO}_2\text{NH}_2$ . If the substituent  $\text{R}$  (e.g., a thiazole or a pyridine ring compound) is an electron acceptor the negative charge on the  $\text{SO}_2$  increases and the positive charge on the  $\text{NH}_2$  becomes greater. Substitutions in other parts of the *p*-aminobenzene sulphonamide molecule that interfere with the induction of this positive charge will produce inactive compounds (see also Jensen and Schmuth 1942).

The conception of activity in terms of the highly polar amino group is supported by the work of Bradbury and Jordan (1942) who found that sulphathiazole, sulphathiazole, sulphapyridine and *p*-aminobenzoic acid all modified the electrophoretic mobility of *Bact. coli* in the same way. Inactive benzene ring compounds and non-resonating isomers of the active compounds did not produce the characteristic change in mobility. As the result of testing a large number of derivatives and analogues of *p*-aminobenzoic acid, Johnson, Green and Pauli (1944) confirmed the importance of the amino group in the *para* position and concluded that inhibitory activity may be determined primarily by the chemical reactivity of the functional group rather than by structural similarity of the substance to *p*-aminobenzoic acid.

In conclusion it should be noted that though sulphonamides like sulphathiazole and sulphadiazine approach the maximum attainable activity *in vitro* as judged by their physicochemical characters they are not necessarily the best attainable for chemotherapy, which must take into consideration absorption, excretion and toxicity in the host, as well as susceptibility to antagonism by products of the host's metabolism.

### The Relation of Chemical and Antibacterial Activity in other Compounds

It will be clear from the foregoing that though *within a group* of chemically related compounds activity and chemical structure are closely connected no ready generalizations in this respect about antibacterial substances are possible in the present state of our knowledge. There are however certain noteworthy features which widely different types of compounds, whether antiseptic or chemotherapeutic, have in common. It is beyond the scope of this book to deal with them in detail. The student is referred to the paper by Albert (1942) upon which we have drawn in the following paragraphs.

We are not for the moment concerned with antiseptics which act by an immediate and extensive disruption of the economy of the bacterial cell such as we see in the coagulative antiseptics. Apart from these antibacterial substances range from the frank antiseptics that are general protoplasmic poisons to the highly selective chemotherapeutic agents with all grades of selectivity of action in between.

The life of the cell depends on the smooth working together of many systems and any agent that interferes with this will be an antiseptic. Since all the enzymes about which there is chemical information have proved to be proteins it is not surprising that agents affecting all proteins are normally antiseptics. Oxidizing agents, halogens and formaldehyde fall into this category. Their mode of action on a protein is seldom understood but this is a general problem of protein chemistry rather than specifically the concern of bacteriology. Agents that upset the relations of lipins and proteins to one another will kill the cell for it is on these relations



that the structure of the interfaces generally depends. The lipid solvents (chloroform toluene etc.) fall into this category as do the phenols cresols and soaps, for although these can act as protein coagulants they are generally antiseptic at higher dilutions than are necessary for coagulant action. Of greater interest are the selective agents that are not classifiable in these general terms.

Crystal violet a triphenylmethane dye used for the surface treatment of wound infections is bacteriostatic in low concentrations—an effect according to a number of observers which is due to the poisoning of the Eh of the bacterial environment at a level too high for cell development (Dubos 1929 Ingraham 1933 Fildes 1940b Hoffman and Rahn 1944 but see Stearn 1930 for a contrary view). In a like manner quinones are antibacterial and redox active. There is however little evidence either in synthetic quinones or in natural quinones like citrinin and penicillic acid (see below) that they owe their activity to interference with optimal Eh values for bacterial growth though Page and Robinson (1943) found that the  $E_h$  values of a series of quinones which were markedly active against *Staph aureus* clustered round a potential of about  $-0.03$  volt.

Antibacterial activity in many compounds may be interpreted in terms of their capacity to neutralize acidic or basic groups in the bacterial cell forming feebly ionized complexes (see Stearn and Stearn 1944). The anionic antiseptics like soaps and the acid dyes combine with basic groups. The cationic antiseptics which include basic dyes like brilliant green or crystal violet the amino-acridine antiseptics and the higher aliphatic amines like cetyltrimethylammonium bromide combine with acidic groups and since acidic groups preponderate in most bacteria particularly in Gram positive organisms it is to be expected that the cationic antiseptics will be more active than anionic antiseptics and that Gram positive organisms will be more readily killed than Gram negative, and this is indeed the case.

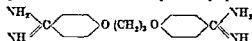
Another aspect of the difference in susceptibility of Gram positive and Gram negative bacteria was pointed out by Miller and his colleagues (1942). Proceeding from the fact that gramicidin and the anionic detergents acting only on Gram positive bacteria were antagonized by certain phospholipins they suggested that Gram negative bacteria owed their insusceptibility to the presence of such phospholipins. In support of this hypothesis they record experiments in which *Bact coli* was made susceptible to tyrothricin (a mixture of gramicidin and tyrocidin) by the addition of protamine sulphate a precipitant of phospholipins. Thus a slightly inhibitory concentration of the protamine together with a non inhibitory concentration of tyrothricin completely inhibited the respiration of *Bact coli*.

In the mono-aminoacridine series of compounds antibacterial activity increases with basic strength up to a certain point but compounds with a basic strength greater than this are no stronger as antiseptics (Albert Rubbo and Goldacre 1941). It is probable that the compounds of extreme basicity are too active to form non ionized neutralization compounds in the bacteria. We have already noted Albert and Goldacre's view (1942) that basicity and activity of the sulphonamides are associated and it is of interest that in both series of drugs those whose structure permits resonance and induction of polarity tend to be the most active.

In addition to basicity surface activity may be a feature of a compound contributing to the bactericidal power of a cationic antiseptic. Albert suggests that the distinction between the acridines and the cationic detergent antiseptics, the one group mainly bacteriostatic, the other bactericidal, may possibly lie in

Two sulphones, *p*-methylsulphonylbenzamidine ( $\text{CH}_3\text{SO}_2\text{C}_6\text{H}_4\text{C}(\text{NH})\text{NH}_2$ ) and *p*-methylsulphonylbenzylamine ( $\text{CH}_3\text{SO}_2\text{C}_6\text{H}_4\text{CH}_2\text{NH}_2$ ) have a marked *in vitro* action against clostridia and *Str. pyogenes*. The former designated V 187 has proved highly effective in the treatment of experimental gas gangrene of the guinea pig. It is not antagonized to any marked extent by body fluids. The latter (V 33a) is also effective against gas gangrene in the guinea pig but is more readily antagonized by the body fluids. Neither substance is antagonized by *p*-aminobenzoic acid, and both act equally well on normal and sulphonamide-resistant bacteria (Evans *et al.* 1944).

**Propamidine** Propamidine 4,4'-diamidino-diphenoxv propane first proved an effective



therapeutic agent for animal trypanosomiasis (Lounie and Yorke 1939) with a therapeutic index of about 30. It is also bacteriostatic in low concentrations, *Str. pyogenes* for example being inhibited in dilutions of  $1:2 \times 10^6$ , and non-toxic for leucocytes in dilutions of 1:1000, it has been used successfully in the local treatment of infections of open wounds and burns (Thrower and Valentine 1943) and of conjunctivitis due to the *Morax Axenfeld* bacillus (Valentine and Edwards, 1944).

It is noteworthy that propamidine bears a structural resemblance to stilbene and that a number of stilbene derivatives have a high bactericidal activity. For example deoxy diethyl stilboestrol ( $\text{HO C}_6\text{H}_4\text{C}(\text{C}_2\text{H}_5) \text{C}(\text{C}_2\text{H}_5)\text{C}_6\text{H}_5$ ) inhibits *Str. pyogenes* in a dilution of  $1:2 \times 10^6$  (Brownlee, Copp, Duffin and Tonkin 1943, see also Faulkner 1943).

**Amino-acridine Compounds (Flavines)**

The claim of the amino-acridine compounds to chemotherapeutic rank is at present not clear. Their toxicity is such that they are suitable only for application to the surface of infected tissues. The degree of tissue intoxication that justifies submitting infected tissues to the undoubted *in vitro* antibacterial action of the drugs is in dispute. McIntosh and Selbie (1942, 1943a) employed proflavine successfully in conjunction with the sulphonamides in the treatment of experimental gas gangrene. For a full discussion of these drugs from the chemotherapeutic point of view, the reader is referred to the review of Browning (1943).

## NATURAL ANTIBACTERIAL SUBSTANCES FROM MICRO-ORGANISMS

Since the early days of bacteriology and mycology, the inhibition of growth of one micro-organism by the growth of another has been a familiar phenomenon, and its possible significance in the therapy of infectious diseases commented upon. The application of these natural bactericidal or bacteriostatic substances to the treatment of infections was usually limited by the low therapeutic index of the materials available, the doses tolerated by the animal were ineffective against the infecting organism. In recent years however, improvements in methods of isolation, and a clearer understanding of the antibacterial effects, have led to the description of a large number of antibacterial substances from bacteria and moulds, some of which are therapeutically active in experimental infection, and one at least penicillin, which is highly effective in the treatment of infective disease in the human subject.

The reader is referred to the comprehensive reviews of Porter and Carter (1938) and of Waksman (1941) for a discussion of antagonisms between micro-organisms and of the earlier work on natural antibacterial substances, which is beyond the

scope of this book. We shall confine the discussion to a few illustrative examples, and note some of the more recently described substances, with especial emphasis on those that have proved their therapeutic value.

### Antibacterial Substances from Bacteria

*Ps. pyocyanea* was among the first bacteria to be studied from the point of view of its elaboration of antibacterial substances. Cultures of the organism yield a number of them—pyocyanase, first prepared by Emmerich and Low (1899), and Emmerich, Low and Korschun (1902) depends upon its content of fatty acids of high molecular weight for its activity (Birch Hirschfeld 1931, Hettche 1931) and affects a large variety of Gram positive and Gram negative bacilli including pathogenic cocci, *Salmonella* bacilli, the diphtheria bacillus (Wagner 1929, Kramer 1935) and *Br. abortus* (Kocholatz 1912). pyocyanin, a respiratory pigment (see Ehrsmann 1931 and Chapter 3),  $\alpha$ -hydroxyphenazine and a colourless compound having a strongly lytic action on *V. cholerae* (Schoental 1941). Iodium, an antibacterial substance from *Chromo. iodinum* is a N oxide of a hydrophenazine, and is apparently specifically antagonized by naphthoquinone (McIlwain 1913b).

Bacteriolytic enzymes of bacterial origin were described by Malfitano (1900, 1903). Nicolle (1907) isolated from *B. subtilis* a lytic agent active against saline suspensions of staphylococci, pneumococci, the anthrax plague and glanders bacilli, and salmonellae (see also Jobling and Petersen 1911, Jobling, Petersen and Fggstein 1915, Sartorius 1921, Much 1925).

The inhibitory actions of certain bacteria were not obviously due to separable enzymes. Thus, certain strains of *Bact. coli* which inhibited the *in vitro* and *in vivo* growth of other bacteria would act only if whole, living cells were present. In culture, they overgrew and eliminated *Salmonella* and dysentery bacilli, streptococci, staphylococci, *B. anthracis* and *C. diphtheriae* (Nussle 1930, 1932, 1933, Gundel and Khewé 1932, Koch and Krämer 1932, Besta and Kuhn 1931, Makowsky 1936, Rejlof 1937, Maner 1939). The *in vivo* inhibition is not marked. For example, Gundel and Khewé infected mice subcutaneously with mixtures of *Bact. coli* and a lethal dose of *B. anthracis*. A number of the mice survived and the survival time of those that died was increased, as compared with mice receiving the lethal dose alone. Other observers have reported the inhibition of *C. diphtheriae* by various streptococci. Inhibition by the live cultures was observed in the subcutaneous tissue of the guinea pig (Besta and Kuhn 1931, Weigmann and Hölzl 1940, Hölzl 1941), and Dulacquet (1939) reported on the successful treatment of throat carriers of *C. diphtheriae* by killed cultures of an inhibitory organism.

Two of the most interesting developments in the isolation of antibacterial substances we owe to the work of Dubos. The first was directed to a specific end, the isolation of a bacterium whose enzymes would destroy the polysaccharide substance in the capsules of Type III pneumococcus. This was accomplished by testing large numbers of bacterial strains, mostly spore-bearing aerobic bacilli in a medium containing the polysaccharide as the sole source of carbon. A bacillus which grew in this medium was trained to produce large amounts of a polysaccharide splitting enzyme, which in a purified form was subsequently used with success in the treatment of experimental Type III pneumococcus infection of the rat (see Chapter 74). The enzyme was specific, and affected only Type III pneumococci. Dubos extended this search to bacilli with a wide range of antibacterial action by serial subculture of samples of soil in a medium containing large quantities

of dead cocci. The medium favoured the growth of the soil bacilli most capable of utilizing the coccal protoplasm as a source of food, and, by selection certain spore-bearing bacilli were finally obtained from which antibacterial substances could be isolated (Dubos 1939) (see also Stokes and Woodward 1942). Similar bacilli were isolated from manure, sewage and cheese. Working mainly with one of them *B. brevis*, Dubos and Hotchkiss (1941) obtained a mixture of bactericidal substance, tyrothricin, from which they separated two crystalline substances, gramicidin and tyrocidin.

Tyrocidin had the following properties. It was a polypeptide with a free basic  $\text{NH}_2$  group, a weakly acidic or phenolic group, and contained tryptophan, tyrosine and dicarboxylic amino-acids—about one fifth of the amino-acid produced on hydrolysis was composed of *d*-amino acids. The molecular weight was in the region of 2,000. It was bactericidal *in vitro* for Gram positive and Gram negative organisms, but was also markedly toxic for animal tissues and leucocytes losing much of its antibacterial activity when in contact with them. It appeared to act as a general protoplasmic poison. (See also Downs 1943.)

Gramicidin was also a polypeptide, but with no free acid or basic groups, about half the amino-acids being of the *d* form. The molecular weight was about 1,400. Gramicidin killed only Gram positive organisms *in vitro*. It protected mice against infections by pneumococci, streptococci and staphylococci, one  $\mu\text{g}$  of the substance for example was effective when injected intraperitoneally along with 10,000 lethal doses of pneumococci. Though highly toxic when given by the parenteral route the curative dose was well below the dose tolerated by the animal.

The high content of the unnatural *d*-amino-acids in both substances suggested that their peculiar properties might in part be dependent on these bodies. Tyrocidin on the whole behaved like an antiseptic, gramicidin acted in a more specific manner like the sulphonamides (Hotchkiss and Dubos 1940; Dubos and Hotchkiss 1941; Hotchkiss 1941; Lipmann, Hotchkiss and Dubos 1941; Dubos, Hotchkiss and Coburn 1942). (See also Tishler *et al.* 1941.) It is noteworthy that Fox, Fling and Dollenback (1944) found that *d* leucine inhibited the growth of *L. arabinosus*, an organism for which *l* leucine is an essential nutrient, and suggested that the *d* isomer might act by interfering with the uptake of the *l* isomer (cf. the antagonism of *p*-aminobenzoic acid and sulphanilamide.)

As the result of a study of the products of acid hydrolysis, Gordon, Martin and Svyne (1943) conclude that the structural unit of gramicidin is a cyclopeptide made up of 24 amino-acid residues—6 *d* leucine, 6 *l* tryptophan, 5 *d* valine, 3 *l* alanine, 2 glycine and 2 unknown hydroxyamino compounds. Tyrocidin they found to contain eight different amino-acids of which phenylalanine was in the *d* form, the rest being mainly in the *l*-form. (See also Christensen 1943, 1944 and Svyne 1944.)

Gramicidin is hæmolytic, and lowers surface tension, though destruction of its antibacterial activity by heat does not alter the surface activity (Heilman and Herrell 1941a, b). Tyrocidin lyses both red cells and leucocytes and both substances are toxic when injected into animal tissues. Administered by mouth to dogs and mice neither is toxic and neither has any effect on infections (Rammelkamp and Weinstein 1942; Robinson and Molitor 1942). For successful therapy, gramicidin and tyrocidin must be brought into immediate contact with the infecting bacteria. Thus oral, subcutaneous or intravenous administration has no effect on intraperitoneally infected mice, and mice infected systemically are not protected by intraperitoneal administration (Robinson and Grassle 1942). Serial subculture in low, but increasing concentration of gramicidin induced resistance in a susceptible strain of *Staph. aureus* and the development of resistance was accompanied by the production of colony variants (Phillips and Barnes 1943). The antibacterial activity of gramicidin is destroyed by gentle hydrolysis in dilute alkali insufficient to destroy its polypeptide structure.

The substances are nevertheless of great interest apart from their possible use in therapy. Those with a selective action on certain bacterial species are useful for incorporation into culture media for suppressing the growth of unwanted bacteria (see Chapter 12) the properties of some of them afford an explanation of microbial antagonisms observed in nature, and, where their constitution is known many of the substances are valuable in relating antibacterial activity to molecular structure or as reagents for exploring the metabolism of the organisms upon which they act. It is noteworthy that most of the substances with a high therapeutic index act predominantly or exclusively upon Gram positive bacteria, with the exception of streptothricin. Those acting on both Gram positive and Gram negative bacteria appear, like the antiseptics, to be general protoplasmic poisons. None is predominantly or exclusively active against Gram negative bacteria.

Numerical measurements of antibacterial powers, and of toxicity to animal tissues afford only a crude means of comparing the various substances. Apart from the varying criteria of a given type of activity, the measures are dependent on the degree of purity of often undefined substances, the strains of bacteria tested, the type of culture medium, and the animal or type of animal tissue employed. It may, however be said that most of the substances are bacteriostatic in concentrations lower than 1:5,000, and many are bactericidal in higher concentrations.

For a general comparison of a number of these substances, with a number of antiseptics, see Wakman and Woodruff (1942).

**Actinomycin.** The bacteriolytic substance produced by sporulating cultures of a *Streptothrix* like mould (*Actinomyces albus*) has already been mentioned (Gratia and Dath 1924). Welch (1936, 1937, 1938, 1939, 1940 Welch and Elford 1937) has studied the lysin, to which he gave the name actinomycin in some detail. It appeared to be a proteolytic enzyme and was inactivated by heating to 54°-69° C. by ultra-violet radiation and by strong acids. It induced bacteriolysis over a wide range of pH, with an optimum at pH 7.5 to 8.5. Lysis appears to depend on at least two factors, a lethal substance which kills the bacteria, and a bacteriolytic enzyme either from the *Actinomyces* or from the bacteria themselves (see also Welch 1942).

**Actinomycin A and B.** These substances were isolated from cultures of *Actinomyces antibioticus* a soil organism, by Wakman and Woodruff (1940a & 1941). Staphylococci and streptococci are inhibited in dilutions of over 1:25 × 10<sup>4</sup>, and *Salmonella bacilli* in 1:25,000.

Actinomycin A is a water-soluble, red pigment, with a molecular weight of about 800 and appears to be a polycyclic nitrogen compound having a reversible redox system of the quinone type (Wakman and Tishler 1942). It is both bactericidal and bacteriostatic inhibiting Gram positive organisms in a dilution of 1:10<sup>4</sup>, and Gram negative organisms at dilutions of 1:5,000 to 1:10<sup>5</sup>. It is active against aerobic and anaerobic bacteria and against certain fungi. Actinomycin A is highly toxic to animals. Doses, either oral or parenteral of 1 mgm per kilogram body weight are lethal for mice, rats and rabbits the animals dying in 15-20 hours with respiratory failure. Post mortem there are gross pathological changes in liver, kidney and spleen. Sublethal doses have no protective effect in mice infected with *Str. pyogenes* or Type I pneumococci (Robinson and Wakman 1942). Actinomycin A inhibits fibrinolysis by *Str. pyogenes* the production of coagulase by *Staph. aureus* but does not affect either tetanus or diphtheria toxin (Neter 1942, 1943) or staphylococcal toxin (Blair and Hallman 1943).

Actinomycin B is a water insoluble substance. It is markedly bactericidal, but not bacteriostatic and is highly toxic to animals.

In experimental streptococcal and staphylococcal infections, it is about 25% as to protein content as compared to penicillin G (Cham. Fever Journal and Williams 1943).

**Penicillin (Novatin).** Penicillin as well as penicillin is produced in cultures of *P. notatum*. An acid medium favours the production of penicillin, which can be detected by using *Portia* (which is insensitive to penicillin) as the indicator organism in a medium of bacteriostatic power. Penicillin is relatively stable water-soluble substance active against Gram-positive and Gram-negative bacteria - dilutions of 1:125 x  $10^4$  to 1:250 x  $10^4$  serum does not interfere with its action. It is active only in the presence of glucose which it decomposes with the production of  $H_2O_2$ . In animals it is inactivating form, with local resistance at the site of action (Kochubski 1942, 1943a, b). The same substance was described by Gribbhard and his colleagues (1942) under the name of novatin. It is a transacetylaminopentamide with the presence of a carboxylic acid. The presence of serum and glucose and the absence of a catalyst are necessary for its antibacterial activity which is mainly due to the  $H_2O_2$  produced in these circumstances (Rohdendorf and Rastbach 1945). A similar substance Novatin R was extracted from cultures of *P. notatum* by Roberts and his colleagues (1945). Like novatin, it was a derivative of penicillin,  $H_2O_2$  from glucose, but unlike novatin, the glucose could be replaced by sucrose, mannose or caliche (van Rensburg et al 1945). It is noteworthy that a milk bacteria which has been described, which is antibacterial action also depends on its power to produce  $H_2O_2$  from the medium containing the bacteria (Green and Paley 1945).

**Penicillin.** Penicillin is a naturally occurring substance which is found in a number of penicillia, bactericidal for many strains 1:1000. Among the penicillins, it is considered there appears to be a relative association between the capacity to produce penicillin and penicillin-like substances. Penicillin activity unlike that of penicillin is not suppressed more than the penicillins, but is suppressed by -SH compounds (Johnson 1942, 1945; Atkinson and Smith 1945).

**Penicillin Acid.** A chemically defined water-soluble substance which is found in *Penicillium notatum*. In its natural form it has about the same activity as penicillin, but has about 10 times as good an antibacterial action on *Staph. aureus* and *Portia*, and is not therefore prone to be chemically destroyed as is (Oxford, Rastbach and Smith 1942; Oxford 1942b). Penicillin acid and certain bacteriostatic compounds (see footnote below) are associated by peptide and certain amino acids as the result of combination between the antibacterial agent and the amine (Oxford 1942).

**Penicillin.** A water-soluble substance from *P. notatum*, active mainly against Gram-positive bacteria (see below).

**Penicillin.** A chemically defined water-soluble substance from a *Penicillium notatum*. Bactericidal activity range from 1:15 x  $10^4$  to penicillin 1:5 x  $10^4$  to *Staph. aureus* and *St. typhimurium* to 1:2 x  $10^4$  to Gram-negative bacterial bacilli. It is more active than penicillin in bacteria (Johnson and Cham 1942).

**Penicillin.** A substance isolated from *Penicillium notatum* by Rohdendorf and Rastbach (1945). It is a derivative of penicillin, which is found in nature in the presence of an additional hydroxyl group in the 6 position. It has the molecular weight 366,000 and is water-soluble. It is water-soluble and is stable in the presence of glucose in comparison with penicillin, where the additional hydroxyl group is associated with a marked increase in antibacterial activity (Oxford and Rastbach 1942). A study of a number of relationships and relationships revealed that the structure of the methoxy group  $OCH_3$  in the penicillin structure resulted in an increase of the hydroxyl group in a decrease in activity. The substitution of the methoxy by hydroxyl in an active compound also decreased its activity (Oxford 1942b).

**Streptomycin.** A chemically defined water-soluble substance from an unidentified species of the genus *Streptomyces* related to *Streptomyces*. Like some others, it acts on Gram-positive and Gram-negative bacteria (Schell, Brey and Wakeman 1944).

In experimental streptococcal and staphylococcal infections its chief effect was to prolong survival time but not to prevent death (Chain Florey Jennings and Williams 1943)

**Penatin (Notatin)** Penatin as well as penicillin is produced in cultures of *P. notatum*. An acid medium favours the production of penatin which can be detected by using *Bact. coli* (which is insusceptible to penicillin) as the indicator organism in titrations of bacteriostatic power. Penatin is a relatively stable water-soluble substance active against Gram positive and Gram negative bacteria in dilutions of  $1:12.5 \times 10^4$  to  $1:2.50 \times 10^6$ . Serum does not interfere with its action. It is active only in the presence of glucose which it decomposes with the production of  $H_2O_2$ . In animals it is moderately toxic with local reactions at the site of injection (Kocholatzky 1942 1943; b). The same substance was described by Coulthard and his colleagues (1942) under the name of notatin. It is a glucose aerodehydrogenase with the properties of a flavoprotein. The presence of oxygen and glucose and the absence of a catalase are necessary for its antibacterial activity which is mainly due to the  $H_2O_2$  produced in these circumstances (Burkinshaw and Raistrick 1943). A similar substance Penicillin B was extracted from cultures of *P. notatum* by Roberts and his colleagues (1943). Like notatin, it was a flavoprotein, producing  $H_2O_2$  from glucose but unlike notatin the glucose could be replaced by xylose, mannose or galactose (van Bruggen *et al.* 1943). It is noteworthy that a milk flavoprotein has been described whose antibacterial act on also depends on its power to produce  $H_2O_2$  from the medium containing the bacteria (Green and Pauli 1943).

**Penicidin** Penicidin is a relatively stable diffusible substance found in a number of penicillia bactericidal for *Salmonella typhi* at  $1:100,000$ . Among the penicillia so far tested there appears to be a negative association between the capacity to produce penicidin and penicillin-like substances. Penicidin activity unlike that of penatin is not suppressed under anaerobic conditions but it is suppressed by —SH compounds (Atkinson 1942 1943b; Atkinson and Stanley 1943).

**Penicillic Acid** A chemically defined water soluble aliphatic keto acid from *Penicillium cyclopium*. In the animal body it has about the same toxicity as phenol but has about 100 times as good an antibacterial action on *Staph. aureus* and *Bact. coli* and it may therefore prove to be chemotherapeutically useful (Oxford Raistrick and Smith 1947; Oxford 1942a). Penicillic acid and certain bacteriostatic quinones (see spinulosin below) are antagonized by peptone and certain amino acids as the result of combination between the antibacterial agent and the antagonizer (Oxford 1942c).

**Penicillin** A water soluble substance from *P. notatum* active mainly against Gram positive bacteria (see below).

**Proactinomycin** A chemically undefined relatively stable substance from a *Proactinomyces*. Bacteriostatic titres range from  $1:1.5 \times 10^4$  for pneumococci  $1:5 \times 10^5$  for *Staph. aureus* and *Str. pyogenes* to  $1:2 \times 10^3$  for Gram negative intestinal bacilli. It is more toxic than penicillin to leucocytes (Gardner and Chain 1942).

**Spinulosin** A substance isolated from *Penicillium spinulosum* by Burkinshaw and Raistrick (1931). It is a toluquinone like fumigatin differing from it only in possessing an additional hydroxy group in the 6 position. It has the molecular structure 3,6-dihydroxy-4-methoxy-2,5-toluquinone. It is weakly antibacterial, and is chiefly of interest in comparison with fumigatin, since the additional hydroxy group is associated with a marked decrease in antibacterial activity (Oxford and Raistrick 1942). A study of a number of toluquinones and benzoquinones revealed that the introduction of the methoxy group  $OCH_3$  into the quinone often resulted in an increase or of the hydroxy group in a decrease in activity. The substitution of the methoxy by hydroxy in an active compound also decreased its activity (Oxford 1942b).

**Streptomycin** A chemically undefined water soluble substance from an unidentified species of *Actinomyces* apparently related to streptothricin. Like streptothricin it acts on Gram negative and Gram positive bacteria (Schatz, Bugie and Wakeman 1944).

or by the addition to the medium of traces of zinc, which, by catalysing the complete oxidation of the dextrose prevents the accumulation of gluconic acid, a product of incomplete oxidation of the dextrose (Foeter Woodruff and McDaniell 1943). *P. notatum* throws variants during its growth in stock cultures, and it is essential to maintain a master culture of high penicillin producing capacity. In fluid cultures, *P. notatum* grows as a felt on the surface of the medium, with consequent limitations in the exposure of the growing mould to air and to the nutrients in the fluid. Clifton (1943) introduced a method resembling that used in the commercial process for the rapid production of vinegar, in which the culture is kept continuously trickling over wood shavings packed in an aerated columnar container. Improved methods of fractionation, including chromatographic methods, have also improved the yield from crude culture filtrates (Abraham, Chain and Holiday 1942).

**Bacteriological Aspects.** Fleming's original observations on the range of *in vitro* activity of penicillin have been amply confirmed. *Staph. aureus*, *Str. pyogenes*, *A. meningitidis* and *A. gonorrhoeae*, the gas-gangrene clostridia and *Actino. boris* are among the most susceptible pneumococci and *Str. viridans* a little less so. If we take the amount of penicillin required to inhibit a certain number of susceptible *Staph. aureus* as unit, the inhibiting dose for these other bacteria lies between 0.25 and 4. *C. diphtheriae* is slightly less susceptible the dose being from 10-30, for *Str. faecalis*, *Salm. enteritidis* and *Br. abortus* and *melitensis*, it is from 30-100, for *Proteus vulgaris*, *Sh. shiga*, *Past. pestis*, 150-500, for *Bact. coli*, *aerogenes* and *friedlanderi*, 1 000-16 000, and for *Myc. tuberculosis*, *Ps. pyocyanea*, *L. icterohamorrhagiae*, *V. cholerae*, *Hamophilus*, yeasts and moulds, the inhibiting dose is of this order, or higher (Bornstein 1940, Abraham *et al.* 1941, Florey and Jennings 1942, Hobbv, Meyer and Chaffee 1942a, McKee and Rake 1942a, McKee, Hamre and Rake 1943, McKee, Rake and Menzel 1944, Helmholtz and Sung 1944). *Ery. monocytogenes* (*Listerella*) resists doses of 40 (Foley, Epstein and Lee 1944). These figures are for the most part determined for a few strains only, and as might be expected the inhibiting dose varies from strain to strain in a species. Thus, it may vary up to 25-fold for pneumococci, and up to 100-fold for *Staph. aureus*.

Bacteria may be trained to grow in gradually increasing concentrations of penicillin. By this method the resistance of *Staph. aureus* may be increased up to a 1 000-fold (Abraham *et al.* 1941). Rammelkamp and Maxon (1942) reported a 64-fold increase in strains habituated to penicillin *in vitro* and up to a 100-fold increase in strains from four of fourteen patients treated with penicillin (see also McKee and Houck 1943). Penicillin resistant pneumococci have been induced *in vivo* by passage through penicillin treated mice. The resulting strains were resistant *in vitro*, and remained so after 30 passages through normal mice (Schmidt and Seiler 1943). By similar means, Rake and his colleagues (Rake *et al.* 1944) induced resistance in *Staph. aureus* and pneumococci. It should be noted that resistance induced *in vivo* is not necessarily due to an increased insusceptibility to penicillin, for Rake and his colleagues found that one strain of *Staph. aureus*, when passed through penicillin treated mice, gained in mouse-virulence but its *in vitro* resistance was unchanged, the strain had clearly adapted itself to the antibacterial action of the mouse tissues, but not to the antibacterial action of penicillin.

Solutions of penicillin exposed to air rapidly lose their potency owing to contamination by bacteria, micrococci, and bacilli of various kinds that elaborate enzymic substances destroying the penicillin. Abraham and Chain (1940) described a penicillinase in *Esch. coli* and a similar substance has been described in paracolon bacilli (Harper 1943); in certain naturally insensitive strains of *Staph. aureus* (Kurby 1944) and in *B. subtilis* (Ungar 1944, Duthe 1944). These enzymic substances are found in filtrates, cultures and extracts of the bacteria. They are relatively stable. Their relation to the structure of penicillin and of penicillin like substances is not yet known. At present, their interest



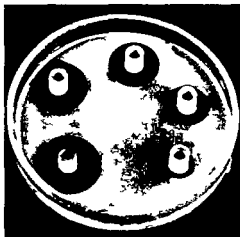


FIG. 25. The assay of penicillin by the plate method. With falling concentrations of penicillin in the cups, the zone of inhibition of *Staph. aureus* is progressively less.

(From a photograph kindly supplied by Dr. N. G. Healey.)

or by the absence of some characteristic growth product of the test organism, for example the absence of haemolysin of *Str. pyogenes* detectable in slide-cells (Wright, Colebrook and Storer 1923) or in culture media (Rake and Jones 1943; Wilson 1943) containing red blood cells, or the absence of acid production in a medium containing an indicator and a carbohydrate fermented by the test bacterium (Fleming 1942). Strains of species other than *Staph. aureus* are suitable for assay. Rammelkamp (1941) for example employed a highly sensitive strain of *Str. pyogenes* for the measurement of low concentrations of penicillin in small quantities of fluid. (See also Lochhead and Timonin 1943.)

**Laboratory Use of Penicillin.** The selective action of penicillin *in vitro* has been fruitfully employed in developing selective media for isolating resistant organisms from mixtures. Following Fleming's original observations on this aspect of penicillin (Fleming 1929) we may note its use for isolation of *H. influenzae* and other haemophilic bacteria (Fleming and Maclean 1930; Fleming 1937; Schoenbach and Seidman 1942), *H. pertussis* (Maclean 1937; Buxbaum and Fiegoh 1943; Cruckshank 1944) and the acne bacillus (Craddock 1942).

**Mode of Action and Constitution.** Little is known of the mode of action of penicillin on bacteria. It is bacteriostatic in low concentrations and appears to be bactericidal in high concentrations. In low concentrations its action is relatively slow (Fleming 1929; Heilman and Herrell 1942). In completely inhibitory concentrations bacterial respiration may continue for some hours and some bacteria survive for 24 hours or longer (Abraham *et al.* 1941). The rate of killing depends to some extent on the bacterium employed and for any one strain is relatively constant until some 99 per cent. of the bacteria are dead, after which the numbers of survivors may remain constant or even increase slightly. Antibacterial activity is marked at 37°C, slight at 18°C, absent at 4°C. The rate increases with diminishing inoculum and with increasing concentrations of penicillin, though only to a certain point beyond which the rate does not increase. Within the limits of experimental error no utilization of penicillin is detectable (Hobby, Meyer and Chaffee 1942b). Though a bacterial strain may be trained to

lytic enzyme that is released from penicillin inhibited cells. Smith and Hay observed that swelling and lysis were associated with active growth since fully grown suspensions showed neither effect. Rantz and Kirby suggest that staphylococci must divide if penicillin is to be effective—a suggestion also made by Rammelkamp and Keefer (1913b) to explain the prolonged survival of a few bacteria in cultures exposed to inhibitory concentrations of penicillin. Bigger (1944b) concluded from a study of the death rate of large inocula of *Staph aureus* in penicillin broth that growing organisms were killed even by low concentrations of the drug. A few cocci however survived prolonged exposure to penicillin but on removal to penicillin free broth grew freely and in this state proved to be fully susceptible to the lethal action of penicillin. Bigger suggests that penicillin is bactericidal only for growing organisms—on organisms in the lag phase of growth it may have a purely bacteriostatic action which by preventing growth renders the organisms insusceptible to the lethal effect of the drug.

In the absence of more extensive data it is unprofitable to speculate on the mode of action of penicillin *in vitro*. If it is generally confirmed that only dividing bacteria are susceptible it is possible that penicillin may act directly on an essential part of the mechanism of fission—but it is equally possible that it interferes with the metabolism of a substance essential for fission—but one of which the bacterium possesses a reserve—the reserve not being used up until the bacterium has multiplied for a few generations.

Little work has been made public on the constitution of penicillin. The earlier reports (Abraham, Chum and Holday 1942; Catch *et al* 1942; Meyer *et al* 1942) deal with the analysis of impure preparations. More recently two breakdown products of highly active preparations have been described: penicillamine, an amino-acid accounting for over half the total nitrogen of penicillin (Abraham *et al* 1943) and penillic acid produced on acid inactivation of penicillin (Duffin and Smith 1943). Meyer, Hobby and Dawson (1943) have prepared methyl, ethyl and *n*-butyl esters of penicillin in the hope of obtaining products more stable and less rapidly excreted than penicillin. The esters were less active than penicillin *in vitro* but were active in experimental streptococcal infection in mice both by the subcutaneous and oral routes. They were more toxic than penicillin and presumably owed their activity to hydrolysis in the animal body with liberation of penicillin.

**Action in the Animal Body.** The toxicity of penicillin in therapeutic concentrations appears to be due to impurities since it varies inversely as the number of arbitrary units per milligram of preparation (Robinson 1943; Hamre *et al* 1943) (see also Hobby *et al* 1942c; Herrell and Helman 1943). The mode of administration to man and animals, and the absorption, distribution and excretion have been studied in some detail. Penicillin is ineffective by mouth and *per rectum*: it is inactivated by the acid of the gastric juice and by the bacteria of the rectal contents. Given parenterally it is rapidly excreted and a variable amount—sometimes up to 75 per cent.—is found in the urine. It is not clear whether the remainder is destroyed in the body. Penicillin does not pass easily between the blood stream and serous cavities and for treating infections of the meninges, pleural cavity, peritoneum and joints it must be introduced directly (Fleming 1943; Florey and Florey 1943; Pilcher and Meacham 1943).

Like that of sulphonamides, the therapeutic action of penicillin is apparently purely antibacterial, serving to increase the efficacy of the natural defence mechanism.

isms of the body by inhibiting the growth of the infecting organisms. In general, except when the lesions produced by the infecting organisms are inaccessible to the penicillin introduced, the *in vivo* potency of penicillin parallels the *in vitro* potency. The first experimental infections successfully treated with penicillin were of mice infected intraperitoneally with *Str. pyogenes* and *Staph. aureus*, and intramuscularly with *Cl. septicum* (Chain *et al.* 1940). It was later shown to be effective in mice infected with virulent pneumococci (Holby *et al.* 1942c), with *N. meningitidis* (Dawson *et al.* 1943), with *Cl. welchii* (McIntosh and Selbie 1943a, McKee, Hamre and Rake 1943, Hae and Hubert 1943-1944), and with *Staph. aureus* (Powell and Jamieson 1942b). Robinson (1943) was unable to demonstrate any action on mice infected with *Myc. tuberculosis*, *Trypanosoma equiperdum* or influenza virus.

Little information is yet available on the effect of penicillin on virus infections. The evidence so far obtained suggests that most viruses are insusceptible, but that two viruses belonging to the lymphogranuloma group (see Chapter 85), namely mouse pneumonia virus and the lymphogranuloma virus itself, constitute possible exceptions to this rule.

- BURTON, H., McLEOD, J. W., McLEOD T. S., and MAYR HARTING, A. (1940) *Brit. J. exp. Path.* 21, 288
- BUSH, M. T. and GOTH, A. (1943) *J. Pharmacol.* 78, 164
- BUTTLE, G. A. H. (1935) *Lancet*, ii, 840, (1937) *Proc. roy. Soc. Med.*, 31, 154
- BUTTLE, G. A. H. GRAY W. H., and STEPHENSON, D. (1936) *Lancet* i, 1286
- BUTTLE, G. A. H. PARISH, H. J., McLEOD, M., and STEPHENSON D. (1937) *Lancet*, i, 681
- BUTTLE, G. A. H., STEPHENSON D., SMITH, S., DEWING T., and FOSTER, G. E. (1937) *Lancet* i, 1331
- BUXBAUM L. and FIEGOLI, V. F. (1943) *J. Bact.*, 46, 543
- CALLOMON F. F. T. (1943) *Amer. Rev. Tuberc.* 47, 97
- CATCH, J. R. COOK, A. H. and HEILBRON I. M. (1942) *Nature, Lond.*, 150, 633
- CHAIN, E. FLOREY, H. W., GARDNER, A. D., HEATLEY, N. G., JENNINGS M. A., ORR EWING J. and SANDERS A. G. (1940) *Lancet*, ii, 226
- CHAIN E., FLOREY H. W., and JENNINGS, M. A. (1941) *Brit. J. exp. Path.* 23, 202, (1944) *Lancet* i, 112
- CHAIN J. FLOREY H. W., JENNINGS M. A., and WILLIAMS T. I. (1943) *Brit. J. exp. Path.*, 24, 108
- CHALLINOR, S. W. and MACNAUGHTAN J. (1943) *J. Path. Bact.*, 55, 441
- CHRISTENSEN H. M. EDWARDS R. R., and PIERSMA H. D. (1941) *J. biol. Chem.* 141, 187
- CHRISTENSEN H. M. (1943) *J. biol. Chem.*, 151, 319, (1944) *Ibid.*, 154, 427
- CHRISTENSEN M. H. (1934) *Zbl. Bakt.* 131, 193
- CLIFTON C. E. (1943) *Science* 98, 69
- CLITTERBUCK, P. W., LOVELL, R. and PAISTRICK, H. (1937) *Biochem. J.*, 26, 1907
- COCKKINS, A. J. (1938) *Brit. med. J.*, i, 1151
- COLEBROOK, L. (1938) *Spec. Rep. Ser. med. Res. Coun., Lond.* No 119
- COLEBROOK, L. and KENNY M. (1936) *Lancet* i, 1279
- COLEBROOK L. and PURDIE, A. W. (1937) *Lancet* ii, 1237, 1291
- COOPER, M. L. and KELLER, H. M. (1943) *J. Pediat.* 22, 418
- COULTHARD C. E. et al. (1942) *Nature, Lond.*, 150, 634
- COWLES P. B. (1942) *Jale J. Biol. Med.* 14, 599
- COYNE F. P., PAISTRICK H., and ROBINSON P. (1931) *Philos. Trans. B.*, 220, 207
- CRADDOCK S. (1942) *Lancet* i, 558
- CRUICKSHANK, J. C. (1939) *Lancet* ii, 681
- CRUICKSHANK, R. (1944) *Lancet*, i, 176
- DAWSON M. H., HOBBS G. L., MEYER, K., and CHAFFER, E. (1943) *Ann. intern. Med.*, 19, 707
- DOMAG G. (1933a) *Dtsch. med. Wochschr.* 61, 250 (1933b) *Ibid.*, 61, 829 (1937) *Klin. Wochschr.* 16, 1412, (1943) *Dtsch. med. Wochschr.*, 69, 379
- DORFMAN A. and KOSER, S. A. (1942) *J. infect. Dis.*, 71,
- DORFMAN A., RICE, L., KOSER, S. A. and SAUNDERS, F. (1940) *Proc. Soc. exp. Biol. & Y.*, 45, 750
- DOUGLAS, S. R. and COLEBROOK L. (1916) *Lancet* i, 181
- DOWNS, C. M. (1943) *J. Bact.* 45, 137
- DUBOS R. (1939) *J. exp. Med.*, 49, 575 (1939) *Ibid.*, 70, 1, 11
- DUBOS R. J. and HOTCHKISS R. D. (1941) *J. exp. med.* 73, 679
- DUBOS R. J., HOTCHKISS R. D. and COBURN, A. F. (1941) *J. biol. Chem.*, 148, 421
- DEFFIN, W. M. and SMITH S. (1943) *Nature, Lond.* 151, 251
- DELSCOUET (1939) *Arch. Med. Pharm. nov.* 129, 410
- DUTHIE E. S. (1944) *Brit. J. exp. Path.*, 25, 96
- EBERHARDT O. (1934) *Z. Hyg. Infektkr.*, 116, 209
- EMMERICH, P. and LÖW, O. (1930) *Z. Hyg. Infektkr.*, 31, 1
- EMMERICH, R., LÖW O., and KORSCHUN, A. (1907) *Zbl. Bakt.*, 31, 1
- EULER, H. von (1943) *Ber. dtsch. chem. Ges.* 75, 1876
- EVANS D. G. FULLER, A. T. and WALKER, J. (1944) *Lancet* ii, 523
- EYSTER, H. C. (1943) *J. cellular comp. Physiol.*, 21, 191
- FAULKNER G. H. (1943) *Lancet* ii, 38.
- FEINSTONE, W. H. WILLIAMS R. D., WOLFE R. T. HUNTINGDON E., and CROSSLEY, M. L. (1940) *Johns Hopk. Hosp. Bull.* 67, 497
- FELDMAN, W. H. and HINSHAW, H. C. (1943) *Amer. Rev. Tuberc.*, 48, 256
- FELDMAN W. H. MANN, F. C. and HINSHAW, H. C. (1942a) *Amer. J. Path.* 18, 750, (1942b) *Amer. Rev. Tuberc.*, 48, 187
- FELKE, H. (1938) *Klin. Wochschr.* 17, 13
- FELSENFIELD O. (1943) *J. Bact.*, 45, 9.
- FILDES P. (1940a) *Lancet* i, 955, (1940b) *Brit. J. exp. Path.* 21, 67, (1941) *Ibid.*, 22, 293
- FINKLESTONE-SAYLES H., PAINE, C. G., and PATRICK L. B. (1937) *Lancet*, ii, 792.
- FITZGERALD, R. J. and FEINSTONE W. H. (1943) *Proc. Soc. exp. Biol. & Y.*, 52, 97

- BURTON H., McLEOD J. W., McLEOD T. S. and MAYE HARTING A. (1940) *Brit. J. exp. Path.*, 21, 208.
- BUSH, M. T. and GOTH, A. (1943) *J. Pharmacol.* 78, 164.
- BUTTLE, G. A. H. (1935) *Lancet*, ii, 840 (1937) *Proc. roy. Soc. Med.* 31, 154.
- BUTTLE, G. A. H., GRAY W. H., and STEPHENSON D. (1936) *Lancet* i, 1<sup>st</sup> 6.
- BUTTLE, G. A. H., PARISH, H. J., McLEOD M., and STEPHENSON D. (1937) *Lancet* i, 681.
- BUTTLE, G. A. H., STEPHENSON D. SMITH, S. DEWING T. and FOSTER, G. E. (1937) *Lancet* i, 1331.
- BYSTROM L. and FIEGOLL, V. F. (1943) *J. Bact.*, 46, 543.
- CALLOWAY F. F. T. (1943) *Amer. Rev. Tuberc.* 47, 9.
- CATCH, J. R. COOK, A. H., and HEILBRON I. M. (1942) *Nature Lond.*, 150, 633.
- CHAIN E., FLOREY H. W., GARDNER, A. D., HEATLEY N. G., JENNINGS M. A., ORR EWING J., and SANDERS A. G. (1940) *Lancet*, ii, 206.
- CHAIN E., FLOREY H. W. and JENNINGS M. A. (1942) *Brit. J. exp. Path.*, 23, 200 (1944) *Lancet* i, 112.
- CHAIN F. FLOREY H. W. JENNINGS M. A., and WILLIAMS, T. I. (1943) *Brit. J. exp. Path.*, 24, 108.
- CHALLENOR, S. W. and MacNAUGHTAN J. (1943) *J. Path. Bact.*, 55, 441.
- CHRISTENSEN H. M. EDWARDS, R. R. and PIERESMA H. D. (1941) *J. Biol. Chem.* 141, 19.
- CHRISTENSEN H. M. (1943) *J. Biol. Chem.*, 151, 319 (1944) *Ibid.*, 154, 4<sup>th</sup>.
- CHRISTINSON M. H. (1934) *Zbl. Bakt.*, 131, 193.
- CLIFTON C. E. (1943) *Science* 98, 60.
- CLUTTERBUCK, P. W. LOVELL, P. and RAISTRICK, H. (1934) *Biochem. J.*, 28, 190.
- COCKINIS, A. J. (1938) *Brit. med. J.*, i, 1151.
- COLEBROOK, L. (1938) *Spec. Rep. Ser. med. Res. Coun. Lond.*, No. 119.
- COLEBROOK, L. and KEVY M. (1936) *Lancet* i, 1<sup>st</sup> 9.
- COLEBROOK, L. and PURDIE, A. W. (193) *Lancet* i, 123 1<sup>st</sup> 91.
- COOPER, M. L. and KELLER, H. M. (1943) *J. Pediat.* 22, 418.
- COULTHARD C. E. et al. (194) *Nature Lond.* 150, 634.
- COWLES P. B. (1942) *Yale J. Biol. Med.*, 14, 599.
- COYNE, F. P. RAISTRICK H., and ROBINSON R. (1931) *Philos. Trans. B.*, 220 20.
- CRADDOCK, S. (1942) *Lancet*, i, 508.
- CRICKSHANK, J. C. (1939) *Lancet* ii, 681.
- CRICKSHANK, R. (1944) *Lancet* i, 16.
- DAWSON M. H. HOBBS G. L., MEYER, K., and CHAFFER, E. (1943) *Ann. intern. Med.*, 19, 07.
- DONAGE, G. (1933a) *Dtsch. med. Wschr.* 61, 200 (1933b) *Ibid.* 61, 809 (1937) *Kl. u. Wschr.*, 16, 141<sup>st</sup> (1943) *Dtsch. med. Wschr.*, 69, 30.
- DORFMAN A. and KOSER, S. A. (194) *J. infect. Dis.* 71.
- DORFMAN A. RICE, L., KOSER, S. A., and SAUNDERS, F. (1940) *Proc. Soc. exp. Biol. N. Y.*, 45, 50.
- DOUGLAS, S. R. and COLEBROOK L. (1916) *Lancet* i, 181.
- DOWNS, C. M. (1943) *J. Bact.* 45, 137.
- DUBOS P. (1939) *J. exp. Med.* 49, 50 (1939) *Ibid.*, 70, 111.
- DUBOS, R. J. and HUTCHISS P. D. (1941) *J. exp. med.*, 73, 679.
- DUBOS R. J., HUTCHISS R. D. and COBURN A. F. (1942) *J. Biol. Chem.*, 148, 421.
- DEFFIN W. M. and SMITH, S. (1943) *Nature Lond.*, 151, 201.
- DULLCOUET (1939) *Arch. Med. Pharm. nat.* 129, 410.
- DETHLE, E. S. (1944) *Brit. J. exp. Path.* 25, 96.
- EHRLICHMAN O. (1934) *Z. Hyg. Infektkr.* 116, 200.
- EMMERICH P. and LOW O. (1899) *Z. Hyg. Infektkr.*, 31, 1.
- EMMERICH R., LOW O. and KORSCHUN A. (1907) *Zbl. Bakt.*, 31, 1.
- EULER, H. VON (1943) *Ber. dtsch. chem. Ges.* 75, 18, 6.
- EVANS D. C. FULLER, A. T., and WALKER, J. (1944) *Lancet*, ii, 523.
- EYSTER, H. C. (1943) *J. cellular comp. Physiol.* 21, 191.
- FATLENER, G. H. (1943) *Lancet* ii, 38.
- FEINSTONE, W. H., WILLIAMS, R. D. WOLFF R. T., HUNTINGDON E., and CROSSLEY M. L. (1940) *Johns Hopk. Hosp. Bull.* 67, 427.
- FELDMAN W. H. and HINSHAW H. C. (1943) *Amer. Rev. Tuberc.*, 48, 206.
- FELDMAN W. H., MANN F. C., and HINSHAW H. C. (1944a) *Amer. J. Path.* 18, 50 (1944b) *Amer. Rev. Tuberc.*, 48, 18.
- FELKE, H. (1938) *Kl. u. Wschr.*, 1<sup>st</sup> 13.
- FELSENFIELD O. (1943) *J. Bact.*, 45, 20.
- FILDES P. (1940a) *Lancet* i, 900 (1940b) *Brit. J. exp. Path.* 21, 67 (1941) *Ibid.* 22, 203.
- FINKLESTONE-SAYLISS H., PAINE, C. G., and PATRICK, L. B. (1937) *Lancet* ii, 9.
- FITZGERALD R. J. and FEINSTONE, W. H. (1943) *Proc. Soc. exp. Biol. N. Y.*, 52, 27.

- JANCSÓ, N VOV and JANCSÓ, H. VOV (1936) *Z ImmunForsch.*, 88, 275
- JENSEN K. A. and SCHMITH, K. (1942) *Z ImmunForsch.*, 102, 261
- JOBLING, J. W. and PETERSEN, W. (1914) *J exp. Med.*, 19, 251, 459, 20, 37, 321, 452
- JOBLING, J. W., PETERSEV, W., and EGGSTEIN, A. A. (1915) *Ibid.*, 21, 239, 22, 129, 141, 401, 568, 590, 597
- JOHNSON, F. H. (1942) *Science*, 95, 104
- JOHNSON, O. H., GREEN, D. E., and PAULI, R. (1944) *J biol Chem.*, 153, 37
- JONES, H. RAKE, G., and HAMRE, D. M. (1943) *J Bact.*, 45, 461
- JONES, R. F. (1944) *Nature, Lond.*, 153, 379
- KATZMAN, P. A. et al. (1944) *J biol Chem.*, 154, 475
- KENNY M., JOHNSTON F. D. and HAEBLER, T. VOV (1937) *Lancet* ii 119
- KING, J. T. and HENSCHEL, A. F. (1941) *Proc. Soc exp Biol.*, A 1, 47, 400
- KIRBY W. M. M. (1943) *Proc Soc exp Biol.*, A 1, 52, 175, (1944) *Science* 99, 452
- KIRBY, W. M. M. and PANTZ L. A. (1943) *J exp Med.*, 77, 29
- KLARER, J. (1941) *Klin Wochr.*, 20, 1250
- KOCK, F. E. and KRÄMER, E. (1932) *Zbl Bakt.*, 123, 308
- KOCHOLATY W. (1942) *J Bact.*, 44, 469, (1943a) *Science*, 97, 186, (1943b) *Arch Biochem.*, 2, 73
- KOHV, H. I. and HARRIS, J. S. (1941) *J Pharmacol.*, 73, 343
- KOLLE, W., LEUFOLD H., SCHLOSSBERGER H., and HUNDENRAGEN K. (1931) *Arch. Inst exp Therap Frankfurt*, 14, 43.
- KOLMER, J. A. (1926) *Principles and Practice of Chemotherapy* Philadelphia
- KRAMER, H. (1935) *Z ImmunForsch.* 84, 500
- KUHN R., MÖLLER, E. F., and WENDT, G. (1943) *Ber chem Ges., Frankfurt*, 76, 400
- KUHN R. and SCHWARZ, K. (1941) *Ber chem Ges., Frankfurt*, 74, 1617
- KUMLER, W. D. and DANIELS, T. C. (1943) *J Amer chem Soc.*, 65, 2190
- LAMAYNA C. and SHAFIRO, I. M. (1943) *J Bact.*, 45, 385
- LAMPEN, J. O. and PETERSON, W. H. (1941) *J Amer chem. Soc.*, 63, 2283
- LAMPEN J. O., UNDERKOFER, L. A. and PETERSON W. H. (1942) *J biol Chem.*, 146, 277
- LANDY, M. and DICKEN D. M. (1940) *J biol Chem.*, 146, 109
- LANDY, M., LARKUM, N. W. OSWALD, E. J. and STREIGHTOFF, F. (1943) *Science*, 97, 265
- LANKFORD C. E., SCOTT, V. and COOKE, W. R. (1943) *J Bact.* 45, 201
- LAUER, P. and MARTIN, H. (1943) *Schweiz. med Wochr.* 73, 399
- LEVADITI, C. and VAISMAN, A. (1930a) *C. R. Acad. Sci.*, 200, 1694, (1930b) *C. R. Soc. Biol.*, 119, 946, (1931) *Ibid.* 127, 1478
- LEWIS J. C. (1942) *J biol Chem.*, 146, 441
- LEWIS, K. H. and SVYDER, J. F. (1940) *J Bact.*, 39, 28
- LIBBY, R. L. and JOYNER, A. L. (1940) *J infect Dis.*, 67, 67
- LIESKE, R. (1921) 'Morphologie und Biologie der Strahlenpilze' Leipzig
- LIPMAN, F., HOTCHKISS R. D., and DUBOS, R. J. (1941) *J biol Chem.*, 141, 163
- LOCHHEAD, A. G. and TIMONIN, M. (1943) *Canad. J. publ Hlth.*, 34, 236
- LOCKWOOD, J. S. (1938) *J Immunol.*, 35, 105
- LOCKWOOD, J. S., COBURN, A. F., and STOKINGER, H. E. (1938) *J Amer med. Ass.*, 111, 2209
- LONG, P. H. and BLISS E. A. (1937a) *Canad med Ass J.*, 37, 457, (1937b) *J Amer med Ass.*, 108, 32
- LOURIE E. M. and LORKE, W. (1939) *Ann trop. Med Parasit.* 33, 289, 300
- LUCCHESI, P. F. and GULDERSLEEVE, N. (1941) *J Amer med Ass.*, 116, 1506
- LWOFF, A., NITTI F., TRÉFOUET, J., and HAMON V. (1941) *Ann Inst Pasteur.*, 67, 9
- LYNCH, H. M. and LOCKWOOD, J. S. (1941) *J Immunol.*, 42, 430
- McILWAIN, H. (1940) *Brit J exp Path.*, 21, 136, (1941a) *Ibid.*, 22, 148 (1941b) *Biochem. J.*, 35, 1311, (1942a) *Brit J exp Path.*, 23, 90, (1942b) *Ibid.*, 23, 260, (1942c) *Biochem. J.*, 36, 417, (1943a) *Brit J exp Path.*, 24, 203, 212, (1943b) *Biochem. J.*, 37, 260, (1944) *Proc biochem Soc., Biochem. J.*, 38, viii.
- McILWAIN, H. and HAWKINS, F. (1943) *Lancet*, i, 449
- McILWAIN H. and HUGHES D. E. (1944) *Biochem. J.*, 38, 187
- McINTOSH J. and SELBIE F. R. (1942) *Lancet*, ii, 700, (1943a) *Ibid.*, ii, 224, (1943b) *Brit J exp Path.*, 24, 246
- McINTOSH J. and WHITEY, L. E. H. (1939) *Lancet*, i, 431
- McKEE, C. M., HAMRE, D. M. and RAKE, G. (1943) *Proc Soc exp Biol.*, A 1, 54, 211
- McKEE, C. M. and HOUCK C. L. (1943) *Proc. Soc exp Biol.*, A 1, 53, 33
- McKEE, C. M. and MACPHILLAMY, H. B. (1943) *Proc Soc exp Biol.*, A 1, 53, 247
- McKEE, C. M. and RAKE, G. (1942a) *J Bact.*, 43, 640 (1942b) *Proc Soc exp Biol.*, A 1, 51, 273
- McKEE, C. M., RAKE, G., GREEP R. O., and DYKE H. B. VAN (1939) *Proc Soc exp Biol.*, N Y, 42, 417

- RAMMELKAMP, C. H. and WEINSTEIN, L. (1942) *J infect Dis.*, 71, 166
- PANTZ, L. A. and KIRBY, W. M. M. (1944a) *J Immunol.*, 48, 29. (1944b) *Ibid.*, 48, 335
- RATNER, S., BLANCHARD, M., COBURN, A. F., and GREEN, D. E. (1941) *J biol Chem.*, 155, 659
- PEFLOH, H. (1937) *Z Immunforsch.*, 90, 29
- Report of Patulin Trials Committee (1944) *Lancet*, ii, 373
- RIST, V. (1939) *C R Soc Biol.*, 120, 972
- RIST, V., BLOCH, F. and HAMON, V. (1939) *Ibid.*, 120, 976
- ROBERTS, E. C. *et al.* (1943) *J biol Chem.*, 147, 47
- ROBINSON, H. J. (1943) *J Pharmacol.*, 77, 70
- ROBINSON, H. J. and GRAESSLE, O. E. (1942) *J Pharmacol.*, 76, 316
- ROBINSON, H. J. and MOLITOR, H. (1942) *J Pharmacol.*, 74, 75
- ROBINSON, H. J. and WAKSMAN, S. A. (1942) *J Pharmacol.*, 74, 25
- ROBLIN R. O., WILLIAMS J. H., WENNER, P. S., and ENGLISH J. P. (1940) *J Amer chem. Soc.* 62, 3002
- POSE, H. M. and FOX, C. L. (1942) *Science* 95, 412
- ROSE, S. B. and MILLER, R. E. (1939) *J Lact.*, 38, 525
- ROSENTHAL, S. M., BAUER, H., and FLYOVE, E. (1939) *Publ Hlth Rep., Wash.*, 54, 1317
- RUBBO, S. D. and GILLESPIE, J. M. (1940) *Nature, Lond.*, 146, 838 (1942) *Lancet*, i, 3f
- RUBBO, S. D., MAXWELL, M., FAIRBRIDGE, R. A., and GILLESPIE, J. M. (1941) *Aust. J exp Biol. med. Sci.*, 19, 185
- SANTORIUS, F. (1924) *Zu Bakt.*, 93, 162
- SCHATZ, A., BUGIE, E., and WAKSMAN, S. A. (1944) *Proc. Soc. exp Biol., N.Y.*, 55, 66
- SCHIEFMAN, O. (1915) *Z Immunforsch.*, 24, 167
- SCHMELKE, F. C., WYSS, O., MARKS, H. C., LEWIS, R. J., and STRANDSKOV, F. B. (1942) *Proc Soc exp Biol N.Y.*, 50, 145
- SCHMIDT, L. H. and HILLES, C. (1940) *Proc Soc exp Biol N.Y.*, 43, 225
- SCHMIDT, L. H. and SESLER, C. L. (1943) *Proc Soc exp Biol N.Y.*, 52, 353
- SCHMIDT, L. H., SESLER, C. L., and DETTWILLER, H. A. (1942) *J Pharmacol exp Therap.*, 74, 175
- SCHWITZER, P. J., CAMAGNI, L. J., and BUCK, M. (1943) *Proc Soc exp Biol, N.Y.*, 53, 75
- SCHOENBACH, E. B. and SEIDMAN, L. P. (1942) *Proc. Soc. exp Biol., N.Y.*, 49, 108
- SCHOENTAL, R. (1941) *Brit J exp Path.*, 22, 137
- SEVAG, M. G. and GREEN, M. H. (1944) *Amer J med. Sci.*, 207, 686
- SEVAG, M. G., HENRY, J., and RICHARDSON, R. (1943) *Amer J med Sci.*, 205, 877
- SEVAG, M. G. and SHELBERNE, M. (1942) *J Lact.*, 43, 411, 421, 447
- SEVAG, M. G., SHELBERNE, M., and MADD, S. (1942) *J. gen. Physiol.*, 25, 805
- SILVERMAN, M. and EVANS, E. A. (1943) *J biol Chem.*, 150, 265
- SMITH, L. D. and HAY, T. (1942) *J Franklin Inst.*, 223, 598
- SMITH, M. I., ROSENTHAL, S. M., and JACKSON, E. L. (1942) *Pub Hlth Rep., Wash.*, 57, 1534
- SNELL, E. E. (1941) *J biol Chem.*, 141, 121. (1944) *Ibid.*, 152, 475
- SNELL, E. E. and MITCHELL, H. K. (1942) *Arch. Biochem.*, 1, 93
- SPINK, W. W., WRIGHT, L. D., VIVINO, J. J., and SKEGGS, H. R. (1944) *J exp Med.*, 79, 331
- STAMP, T. C. (1939) *Lancet*, ii, 10
- STANSFELD, J. M., FRANCIS, A. E. and STUART HARRIS, C. H. (1944) *Lancet*, ii, 370
- STEARN, A. E. (1930) *J Bact.*, 19, 133
- STEARN, A. E. and STEARN, E. W. (1924) *J Bact.*, 9, 491
- STOKES, J. L. and WOODWARD, C. R. (1942) *J Bact.*, 43, 253
- STOKINGER, H. E., CHARLES, P. C. and CARPENTIER, C. M. (1942) *J Lact.*, 44, 261
- STRAUSS, E., DISGLE, J. H. and FINLAND, M. (1941) *J Immunol.*, 42, 313
- STRAUSS, E. and FINLAND, M. (1941) *Proc. Soc. exp Biol., N.Y.*, 47, 428
- STUART HARRIS, C. H., FRANCIS, A. E., and STANSFELD, J. M. (1943) *Lancet*, ii, 684
- SYNGE, R. L. M. (1944) *Biochem. J.*, 38, 285
- TAMURA, J. T. (1944) *J Bact.*, 47, 529
- TEPLY, L. J., AXELROD, A. E., and ELYENJEN, C. A. (1943) *J Pharmacol.*, 77, 207
- THROWER, W. R. and VALENTINE, F. C. O. (1943) *Lancet*, i, 123
- TILLET, W. S., CAMBER, M. J., and HARRIS, W. H. (1943) *J clin Invest.*, 22, 249
- TIMONIN, M. I. (1942) *Science*, 96, 494
- TISHLER, M., STOKES, J. L., TREYNER, N. R. and COVY, J. B. (1941) *J biol Chem.*, 141, 197
- TRÉFOUËL, J., TRÉFOUËL, J., VITTI, F., and BOVET, D. (1935) *C P Soc. Biol.*, 120, 756
- TYTLER, W. H. and LAPP, A. D. (1942) *Brit. med J.*, ii, 748
- UGAR, J. (1944) *Nature, Lond.*, 154, 236
- VALENTINE, F. C. O. and EDWARDS, A. M. (1944) *Lancet*, i, 753
- VALKO, E. L. and DUBOIS, A. S. (1944) *J Bact.*, 47, 15
- VIVINO, J. J. and SPINK, W. W. (1942) *Proc. Soc. exp Biol., N.Y.*, 50, 336

- WACHS, W. (1932) *J. Immunobiology*, **63**, 483.
- WAKSMAN, S. A. (1941) *Proc. Roy. Soc.*, **5**, 231. (1943) *J. Biol.*, **46**, 279. (1944) *Sci. res.* **92**, 279.
- WAKSMAN, S. A. and B. CIL, L. (1943) *Proc. nat. Acad. Sci.*, **29**, 279.
- WAKSMAN, S. A. and GLEGER, W. B. (1944) *J. Biol.*, **47**, 291.
- WAKSMAN, S. A., B. KUNG, L. S. and SPENCER, L. L. (1943) *J. Biol.*, **45**, 233.
- WAKSMAN, S. A. and TISHLER, M. (1947) *J. Biol. Chem.*, **142**, 519.
- WAKSMAN, S. A. and WOODRUFF, H. H. (1944) *J. Biol.*, **40**, 31. (1947) *Proc. Soc. exp. Biol. N. Y.*, **45**, 609. (1947) *J. Biol.*, **42**, 231. (1947) *Ibid.*, **44**, 272.
- WEIGMANN, F. and B. LIL, H. (1943) *Z. Hyg. Inf.krankh.*, **122**, 673.
- WEINLING, R. (1937) *J. Hyg. pathology*, **27**, 117. (1941) *Ibid.*, **31**, 991.
- WEINLING, P. and F. M. S. O. H. (1935) *Ibid.*, **26**, 1008.
- WELD, J. T. and MITCHELL, L. C. (1937) *J. Biol.*, **28**, 235.
- WELSH, M. (1936) *C. P. Soc. Biol.*, **123**, 1013. (1937) *Ibid.*, **124**, 1401. (1938) *Ibid.*, **125**, 244. (1939) *Ibid.*, **126**, 347. (1940) *Ibid.*, **127**, 1173. (1941) *Ibid.*, **128**, 134. (1942) *Ibid.*, **129**, 131.
- WELSH, M. and L. F. W. J. (1937) *C. P. Soc. Biol.*, **125**, 1013.
- WENGATZ, H. F., B. A. R. A., and CARPENTER, C. M. (1938) *J. Biol.*, **25**, 30.
- WEST, R. and COOPER, A. F. (1947) *J. exp. Med.*, **72**, 91.
- WHITBY, L. L. H. (1934) *Lancet*, **1**, 1710.
- WHITE, F. C. (1940) *Science*, **92**, 127. (1943) *Proc. Soc. exp. Biol. N. Y.*, **54**, 27.
- WHITE, F. C. and HILL, J. H. (1947) *J. Biol.*, **45**, 433.
- WHITE, H. J. (1937) *J. Biol.*, **28**, 249.
- WITKING, S. (1941) *Science*, **94**, 389.
- WILKINS, W. H. and HARRIS, C. C. M. (1947) *Dis. J. exp. Path.*, **23**, 166. (1948a) *Ibid.*, **24**, 141. (1948b) *Ann. appl. Biol.*, **30**, 279.
- WILK, S. L. (1943) *Nature Lond.*, **152**, 478.
- WINE, H. (1947) *J. Pharmacol.*, **78**, 141.
- WINT, L. K. and J. L. C. H. W. (1937) *Ann. Inst. Pasteur*, **62**, 616.
- WOLFE, D. D. (1940) *Proc. J. exp. Path.*, **21**, 74.
- WOLF, W. B. (1947) *J. exp. Med.*, **73**, 307.
- WOODRUFF, H. W. and WHITE, S. G. C. (1943) *J. exp. Med.*, **78**, 481.
- WRIGHT, A. F., G. L. K. S. L., and STOKER, J. J. (1937) *Lancet*, **2**, 417-473.



## CHAPTER 7

### THE ANTIGEN ANTIBODY REACTIONS

THE problems of infection and resistance—the nature of the mechanisms which determine the course of events when a potentially pathogenic parasite gains access to an animal host—will be discussed in later chapters

Very early in the development of this field of research investigators began to study the reactions which occur when blood, blood serum, or other body fluids are allowed to react in the test-tube with bacteria or with bacterial products. Almost from the first the study of these reactions was pursued by many workers without particular reference to the role, if any, which they played in the combat between parasite and host and it soon became clear that they provided a new technique, which could be applied to a variety of biological problems quite apart from the study of disease.

The development of our knowledge of the various reactions which may occur when the blood or serum of a given animal is mixed with various bacteria, bacterial products, foreign cells or foreign proteins, may be briefly summarized as follows.

In 1888 Nuttall demonstrated that the defibrinated blood of certain animals had the power of killing certain bacteria. Buchner (1889*a, b, c*) showed that this bactericidal power was possessed by the cell free serum, and that it was lost when the serum was heated to 55° C for 1 hour.

In 1890 von Behring and Kitasato showed that the serum of animals which had received repeated injections of non lethal doses of tetanus toxin, or of diphtheria toxin, had acquired the property of specifically neutralizing these toxins, and thus preventing their poisonous effect.

Between 1893 and 1895 Pfeiffer (see also Pfeiffer and Issaëff 1894) recorded the occurrence of bacteriolysis, or granular degeneration followed by partial dissolution, in cholera vibrios and some other bacteria when these were introduced into the peritoneal cavity of guinea pigs which had previously received inoculations of killed cultures of the particular bacteria in question. They showed, also, that the substances which determined this bacteriolysis were present in the blood serum, and in other body fluids.

In 1895 Bordet published his classical paper on the properties of the sera of immunized animals. We may note here that although the idea of increased resistance to infection was implicit in the early conception of the process of immunization this term soon came to possess a wider meaning. Any animal, into whose tissues has been introduced any antigenic foreign substance, dead or living and whose serum has in consequence, gained the property of reacting in some way with that particular substance, is spoken of as having been immunized against it, and such a serum is referred to as an *immune serum* or an *antiserum*. Bordet, extending the observations of Buchner, showed that two different substances are

involved. In 1903 Wright and Douglas, in an extensive series of experiments, demonstrated that this action of normal serum was due to a thermolabile substance which acted directly upon the bacteria and not upon the leucocytes. To this substance they gave the name of *opsonin*. Neufeld and Rimpau (1904-1905) demonstrated the presence of thermostable substances in the blood serum of animals immunized against streptococci and pneumococci which acted specifically on these bacteria in such a way as to increase the degree to which they were ingested by phagocytic cells. To these substances they gave the name of *bacteriotropins*.

Thus, in the earliest years of the present century, we had at our disposal a considerable body of facts with regard to the action of the sera of normal and of immunized animals on bacteria, foreign cells and foreign proteins. Little indeed has been added during the intervening years, to our knowledge of what may happen when a normal or immune serum is mixed *in vitro* with the material against which it is active. Investigators during this period have been mainly engaged in trying to discover how these reactions are brought about. On this aspect of the problem a mass of information has been collected, and although the correlation of the ascertained facts has been a difficult matter, and our understanding of the underlying mechanism of the serum reactions is still far from complete, we seem during the last two decades to have made appreciable progress towards an orderly arrangement of evidence and a generalization of theory, which has resulted in a clearer conception of the processes involved.

**Terminology**—With the gradual development of our knowledge of the serum reactions new names have been invented to describe the phenomena observed and to designate the substances which are assumed to be the essential reagents. As in other terminologies which have grown naturally and have never yet been systematized, the terms employed are often ill-defined, and there is much overlapping. There has indeed been a riotous creation of hypothetical entities in the discussion of the available data, and this redundant growth is responsible for many of the obstacles encountered by the student in mastering the facts of a subject, which in its main outlines is not intrinsically difficult. It is however, quite impossible to dispense with the use of some kind of scientific shorthand and it is therefore necessary to gain an adequate acquaintance with the terms in common use, and especially to realize their limitations.

It is usual to refer to the substances which make their appearance in the blood serum, in response to the inoculation of foreign substances of various kinds, and which react with these substances, *in vitro* or *in vivo*, in some observable way, as *antibodies*. The foreign substances which, on inoculation into the tissues, stimulate the formation of these antibodies, are referred to as *antigens*. These two terms may be defined as follows.

✓An **ANTIGEN** is any substance which, when introduced parenterally into the animal tissues, stimulates the production of an antibody, and which, when mixed with that antibody, reacts specifically with it in some observable way.

✓An **ANTIBODY** is any substance which makes its appearance in the blood serum or body fluids of an animal, in response to the stimulus provided by the introduction of an antigen into the tissues, and reacts specifically with that antigen in some observable way.

The introduction of the antigen into the tissues is usually made by the parenteral route, since the antigen must reach the tissues in an unaltered state. Antigens

recent years they are now rarely employed. It is usually better, for reasons that are discussed in later sections of this chapter to rely on the term *antigen* allowing the context to make clear which particular antigen or antigenic component is indicated.

These general rules of nomenclature have certain exceptions. When the antigen itself possesses some well marked biological activity, which is neutralized by the antibody the antibody is named by adding the prefix *anti* to the name of the antigen. Thus the antibody which neutralizes a toxin is called an *antitoxin*, the antibody which neutralizes snake-venom is called *antivenin*, and so on. In many cases descriptive terms have been retained for the designation of antibodies instead of coming new names and this practice has much to recommend it in the present state of our ignorance.

It will be observed that the thermolabile substance which is present in normal serum and which appears to be the active agent in bringing about the lysis of sensitized cells and the death of sensitized bacteria demands a separate name. This substance is not an antibody, since it is not increased in amount as the result of immunization and it must for the moment be regarded as in a class by itself. Its original name of alexine has become very generally replaced by the name complement which was introduced by Ehrlich. It may be defined as follows.

✓ **COMPLEMENT (or ALEXINE)** is a thermolabile substance, or complex of substances present in varying concentrations in the blood serum of most normal animals which has the property of bringing about the lysis of certain cells and bacteria in conjunction with certain antibodies which render the cells or bacteria sensitive to its action.

This definition is incomplete for it begs the question as to whether complement, acting by itself produces any significant reaction whatever. It describes, however with sufficient accuracy, the part which complement plays in the particular reactions with which we are concerned.

### Theories of the Mechanism of the Serum Reactions

Before entering on a detailed discussion of the various serum reactions, it is well to have a general idea of the theories which have been propounded to account for the phenomena observed.

One theory, which has played a prominent part in immunological studies, was propounded by Ehrlich (1898-1900). It has been developed and modified with extreme ingenuity by himself and by his colleagues to meet the demands made upon it by the continuous accumulation of new and often disconcerting facts, which had to be fitted into a structure growing more and more complex, and obviously becoming a little strained, as one new hypothesis after another was added to the central conception.

According to this side-chain theory we should regard the cell as being built up of highly complex chemical aggregates, with attached groupings, or side chains, the normal function of which is to anchor nutrient substances to the cell, and in some cases to act upon and modify them, as a preliminary to their incorporation into the essential cell substance. These side-chains or *receptors*, thus form the point of contact between the cell substance and any other materials which gain access to the fluids in which the cell is bathed. It is only by gaining attachment to these receptors that substances of the class to which antigens belong can exert any action on the cell, and so stimulate it to activity. In Ehrlich's view the antigens, whatever their nature attach themselves to these cell receptors. Since the antigens are in all cases foreign substances, which have no part in the

or precipitation, Ehrlich postulated the existence of another grouping in the receptor, which determined the particular change in the condition of the antigen after the antibody was anchored by its haptophore group. This second, active group was named by Ehrlich the *ergophore group*. In certain cases, which we discuss more fully below, it became necessary to postulate the existence of receptors which, while themselves inactive, served to unite an antigen to a second active substance—the complement or alexine to which we have referred above. To meet

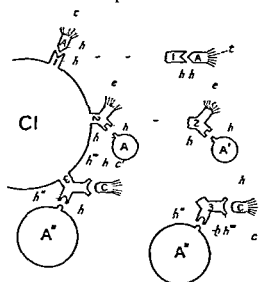


FIG. 31

## Cl. Cell.

- 1 Receptor of 1st order with its haptophore group  $h$
- 2 Receptor of 2nd order with its haptophore group  $h$  and its ergophore group  $e$
- 3 Receptor of 3rd order with its two haptophore groups,  $h$  the cytophilic group, and  $h$  the complementophilic group
- A, A' Various antigens, each with its haptophore group  $h$ . A has in addition an ergophore or toxophore group  $t$
- c. Complement, with its haptophore group  $h$  and its ergophore group  $e$

this case Ehrlich postulated the existence of receptors with two haptophore groups, one of which became attached to the antigen to be acted upon, and one to the complement which was the acting substance. Both these groups were to be regarded as strictly specific in their chemical affinities. The one which combined with the cell or other antigen to be acted upon was known as the *cytophilic group*, the one which combined with the complement was referred to as the *complementophilic group*. Ehrlich named this type of receptor an *amboceptor*, because both groups were supposed to be of the haptophore type. He also referred these three types of receptors to three orders, the first having a single haptophore group, the second one haptophore and one ergophore group, and the third two haptophore groups.

Fig. 31 gives a diagrammatic representation of Ehrlich's general conception.

Ehrlich regarded these receptors as definite chemical entities which entered into firm union with antigens, or with complement, by linkage of the corresponding haptophore groups. When it was found that such antigens as the toxins could be so treated as to lose their toxicity without losing their power of combining with antitoxin, he assumed that the toxophore group had been altered or destroyed, while the haptophore group remained intact. Similarly he postulated the existence of a modified complement, in which an intact haptophore group was associated with an ergophore group which had lost its functioning power. It will be realized that such a theory lent itself readily to schematic representation of the various reacting substances and their assumed modifications, and that there was a natural tendency to elaborate the assumed structure, and vary the functional activity of the various groups, to account for new phenomena not readily explicable by the unmodified hypothesis. This is in fact what happened.

not analogous to that between a strong acid and a strong base, but is a reversible action of the type which occurs between weak acids and weak bases, the equilibrium attained in any particular case being determined by the concentration of the reacting substances in accordance with the law of mass action

Our present concept of the mechanism of the antigen antibody reactions is derived, in varying degree, from each of these three theories. Its essential core is provided by Ehrlich's classical hypothesis, simplified and made more precise by the pioneer chemical studies of Landsteiner and his colleagues (see pp 252-258). It follows Bordet in postulating the union of antigen with antibody in varying proportions. It adopts the assumption of Arrhenius and Madsen that the antigen antibody compound is dissociable, at least in some cases and under certain conditions

Before discussing how this synthesis of conflicting theories has been brought about it will be convenient to give a brief description of each of the antigen antibody reactions in turn

### The Precipitin Reaction.

Our knowledge of the mechanism of this reaction has, in the main, been obtained during recent years but it forms so convenient a starting point for a general description of the *in vitro* reactions between antigens and antibodies that we may with advantage ignore historical succession. In this reaction the antigen as well as the antibody is initially in a state of colloidal solution. We can therefore work with chemically purified antigens such as crystalline egg albumin, or one of the bacterial polysaccharides that will be described in future chapters. In this way we can eliminate the complexities that are introduced by the presence in the reacting system of several different antigenic components. We can also as Landsteiner and his colleagues have shown (see pp 252-255), prepare synthetic antigens, the specificity of which is determined by known chemical groupings, and study their flocculation by antisera prepared against them. Moreover, in the precipitin reaction our data in regard to the quantitative relationships in our reacting system have attained their greatest precision

Specific serological precipitation occurs when any antigen in solution is allowed to react with its corresponding antiserum in the presence of electrolytes provided that the concentration of each of these reagents, and the experimental conditions, such as temperature of incubation and time of incubation, are suitably arranged

The rate of formation of the antigen antibody compound, and of its flocculation varies according to the conditions of the experiment. It is hastened by any procedure that increases the frequency of impact between the molecules, or particles, of antigen and antibody, or between the first formed particles of the antigen antibody complex (see Eagle 1932). Such procedures include any decrease in the total volume of fluid in the reacting system, with a consequent decrease in the space between the reacting particles, and any factor that increases their rate of movement, such as shaking or convection currents in a tube partially immersed in a water bath. It is also hastened by increasing the concentration of electrolytes up to an optimal point, beyond which a further increase may cause a retardation. An increase in temperature up to an optimum also increases the speed of flocculation and it seems doubtful whether this effect is wholly due to an increased frequency of impact. With some reacting systems the higher water bath temperatures (37-55° C) may be found unsuitable, since the antigen antibody compound may be partly soluble in this range

These observations have since been confirmed by many other workers, and with a number of different antigen-antibody systems. In these systems the optimal ratio as determined by Dean and Webb's method is a constant, and the method affords a reliable method of determining (a) the antibody content of two or more solutions in terms of a fixed amount of antigen or (b) the antigen content of two or more solutions of antigen in terms of a fixed amount of antibody.

A few examples may make these points clear.

Dean and Webb titrated 33 samples of rabbit and horse antisera, and found the optimal ratio varied from 1:177 to 1:14. Taking the two sera exhibiting these extreme ratios, we see that one part by volume of antigen (horse serum) gave optimal flocculation with 177 and with 14 parts by volume of the two antisera. Since one part by volume of antigen requires a constant amount of antibody for optimal flocculation, this amount must be contained in the one in 177 volumes, and in the other in 14 volumes, and the second serum contains  $177/14 = 12.6$  times the concentration of antibody in the first.

Taylor, Adair and Adair (1932) have used this method to determine the amount of a given antigen in a naturally-occurring mixture. Thus, they have estimated the percentage of egg albumin in egg white, and the amount of globulin in horse serum, using in each case chemically purified antigens for their titrations. The results obtained were in close agreement with previous determinations, carried out by the usual methods of chemical analysis, as shown by the following comparative figures.

Percentage of crystalline egg albumin in egg white.	Per cent
Hopkins (1900)	6.0
Wu and Ling (1927)	8.5
Optimal ratio method	7.29

Percentage of globulin in horse serum.	Per cent
Hammarsten (1878)	4.5
Gibson and Banzhaf (1910)	4.0
Optimal ratio method	4.46

The validity of the calculations depends among other things on the assumption that there is one type of antibody reacting with a single antigen in solution. With solutions containing two antibodies and two antigens the titration series may yield two distinct zones, or a single zone that is narrow or broad according to the degree of coincidence of two optimally reacting mixtures in the test tubes. (See for example Dean, Taylor and Adair 1932; Goldsworthy and Rudd 1935; Pochon 1936.)

It will be noted that Dean and Webb in determining the optimal ratio for flocculation kept the amount of antiserum constant, and varied the amount of antigen. The ratio determined by this method may conveniently be referred to as the *constant-antibody optimal ratio* or *constant-antibody O.P.*

Prior to these studies Ramon (1922) had shown that diphtheria antitoxin could be titrated by mixing falling amounts of antitoxic serum with constant amounts of toxin and noting which tube in such a series first showed flocculation. The fact that this technique was introduced as a practical method of standardizing an important therapeutic reagent and that controversy centred round its relation to the current *in vivo* methods of standardization probably delayed the recognition of its wider theoretical significance. We may conveniently refer to the optimal antigen-antibody ratio as determined by Ramon's technique, as the *constant antigen optimal ratio* or *constant-antigen O.R.*

Considering first the rows in the table, we see that the constant-antibody O R is  $1/5$   $1/200$   $1/75$   $1/300$ , etc., that is,  $1/40$ . Considering columns, the constant antigen O R is  $1/5$   $1/1,200$ ,  $1/75$   $1/1,600$  etc., that is  $1/240$ . In this instance therefore the constant antigen optimal mixture contains six times as much antibody as the constant antibody optimal mixture. The two optima are on the face of it quite arbitrary, depending on which reagent we vary in comparing reactions taking place in a constant volume. For a  $1/30$  dilution of antibody,  $1/1,200$  antigen precipitates faster than any other dilution, but this particular dilution of antigen can be made to precipitate even more quickly by increasing the concentration of antibody to  $1/5$ , when precipitation occurs in 10, as compared with 20 minutes.

### The Two Optimal Ratios and Chemical Equivalence

If the precipitates are removed by centrifugation the supernatant fluid remaining may be tested, by the addition of more antibody or more antigen, for the presence of residual antigen or antibody. Dean and Webb (1926) found that at constant antibody optimum there was neither antigen nor antibody in the supernatant fluid and confirmatory results have been recorded by Taylor (1931, 1933) Smith (1932) and Duncan (1932a). Duncan also found that at the constant-antigen O R of the system with which he was working the supernatant fluid contained a gross excess of antibody. It seems that in many systems the amounts of antibody and antigens in an optimally reacting mixture as defined by the constant antibody titration are equivalent. For this reason, the constant antibody optimal ratio is obviously important in determining the nature of the combination between antigen and antibody. However, the Ramon method of titrating antitoxin gives a constant-antigen O R and the close agreement between *in vitro* and *in vivo* tests indicates that the ratio corresponds to a mixture in which toxin is first neutralized by antitoxin—that is an equivalent mixture. The fact is, however not irreconcilable with the generally observed equivalence at the constant antibody O R for the two ratios in diphtheria toxin-antitoxin titrations happen to differ very little from one another (see Miles 1933, Boyd 1941, Boyd and Purnell 1944).

The second problem that falls for discussion at this stage is that raised by Bordet's hypothesis. Do antigen and antibody combine in constant proportions, or may they combine in varying proportions, according to their relative concentrations in the reacting mixture? The precipitation reaction affords particularly favourable opportunities for attacking this important problem.

Confining ourselves to the constant antibody titration, we may distinguish three zones—the central equivalence zone, the zone of antigen excess and the zone of antigen deficiency which is usually referred to as antibody excess, though this term should properly be used for the constant antigen titration series. The zone of antigen excess may be further divided into two—a region where a marked excess of antigen totally inhibits precipitation and a region of moderate excess, where some precipitation occurs. The equivalence zone may be narrow and confined to mixtures in the optimal ratio, or the range of complete precipitation of both reagents may be so broad that the antigen concentration at the antigen excess end of the equivalence zone may be eight times, and is often two or three times that at the antibody excess end of the zone.

These are the kind of results obtained with rabbit and horse antiserum. With other animals, notably the guinea pig, it is sometimes impossible to obtain any zone of complete

to the amount and composition of the precipitate. The antibody excess zone is to the left, the equivalence zone or point is indicated by the arrows, and the antigen excess zone is to the right. As the amount of antigen added is increased, the amount of antibody in the precipitate rises to a maximum. The maximum is not, as might be expected, attained in the equivalence zone but increases in the antigen

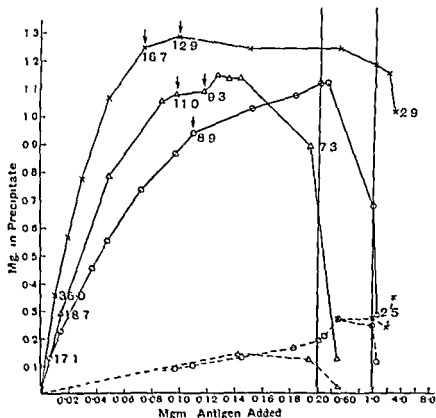


FIG. 3. Am units (antibody nitrogen and antigen in the precipitate formed from 1 ml. of antibody solution) on the addition of the amounts of antigen shown as abscissa. (Continue as lines antibody nitrogen, broken lines antigen, Type III pneumococcus polysaccharide (x), III) in mgm. of polysaccharide.

x Type III and homologous antibody

O Azoprotein and homologous antibody

Δ Egg albumin and homologous antibody

The arrows show the points at which neither antigen nor antibody was detectable in the supernatant fluids. The figures against points on the curves show the ratio of antibody to antigen in the precipitate at these points.

The scale of the abscissa

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is twice in order to bring the whole curve into the figure as in Kenall (1935).

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excess zone. According to the amount removed from the extra forming a antigen

constant fluid, all the available antibody zone, so that the antibody represents the antigen excess zone must be high capable of entering into com-. The existence of such antibody



extracted rabbit rat guinea pig and sheep antisera and lecithin that of extracted horse man mouse cat dog and goat antisera. Although it is possible that a small amount of lipin is an essential constituent of antibody globulin (Chow and Coebel 1935), the large amounts often found in specific precipitates are adsorbed non specifically during precipitation and are essential neither to the complete antibody molecule nor to precipitation.

Native antibody may be a lipo globulin complex containing this small amount of lipin (see Chow and Coebel 1935). Heidelberger (1939) suggests that the action of lipin is essentially mechanical providing nuclei round which specific precipitation is initiated.

### The Inhibiting Effect on Precipitation of Antigen or Antibody Excess

We may now return to the inhibitory effect exerted by antigen or antibody excess and to the observed difference between the constant antibody O R and the constant antigen O R.

Boyd and Hooker (1934-1938) pointed out that the ratio of antibody to antigen in optimal precipitates varied inversely as the molecular weight of the antigen. For example with the Type III pneumococcal polysaccharide with an estimated molecular weight of 4 000 the mean ratio was 60 with serum albumin of molecular weight 70 200 the mean ratio was 7.4 and with a mollusc haemocyanin of molecular weight 3 000 000 the mean ratio was 1.53. The relation holds with sufficient accuracy over a whole series of precipitation systems. Since the molecular weight of the rabbit and horse antibodies used in the majority of the systems was in the region of 150 000 it follows from the size of the ratio that the antigen molecules concerned must each possess a number of similar reactive groups capable of combining with antibody. At the optimum the amount of antibody taken up is consistent with the hypothesis that it forms a layer of contiguous units of antibody protein (see also Kleczkowski 1941a).

We may call the reactive groupings on antigen and antibody valencies though unlike the valency of orthodox chemistry they are not simple atomic valencies but combining complexes. On this basis antigens are multivalent a fact which may also be deduced from the reaction of antigens with artificially introduced reactive groups. (See p. 262).

There is more doubt about the valency of antibody. Analysis of precipitates formed in the presence of a great excess of antigen should help to decide this point by telling us the maximum number of antigen molecules that combine with a single molecule of antibody (see Hooker and Boyd 1942). Unfortunately for technical reasons the composition of precipitates in this region is difficult to determine with accuracy but the available figures indicate that usually one and in rare cases two antibody molecules combine with one of antigen. On this basis, therefore antibody is usually monovalent or at most divalent.

### The "Two-Stage" and the "Lattice" Hypotheses

If we accept the hypothesis of the monovalent antibody we are committed to an explanation of the phenomena of precipitation along the lines of Bordet namely that there is first a stage of specific combination followed by a second stage of non specific flocculation depending on the presence of electrolytes and serum lipins. The union of antibody globulin with antigen renders it hydrophobe, and consequently salt sensitive. The antigen antibody complexes are forced out of solution and having more attraction for one another than each separate particle has for water they cohere and eventually form a visible precipitate. This is the two-stage hypothesis.

On the other hand there are phenomena in precipitation and agglutination

hydrophile groups remain to slow or totally to inhibit precipitation. The compounds are schematically represented in Fig 33 where D E and F represent an antigen molecule after combination in the zones of antibody excess, equivalence and antigen excess.

The compounds formed according to Marrack's lattice hypothesis are also illustrated in Fig 33 A-C which represent two-dimensionally the reaction of a hypothetical hexavalent antigen and trivalent antibody. In gross antigen excess, all three antibody valencies are occupied by antigen, the solution consists of quadri-molecular complexes which cannot combine with one another to form larger complexes because only antigen valencies remain unsatisfied. In a lesser antigen excess complexes as large as A<sub>1</sub> are formed, but these cannot aggregate more fully since again only antigen valencies are unsatisfied. B and B<sub>1</sub> illustrate similarly two complexes formed in antibody excess, and C represents an early stage in lattice formation at the equivalence point: there are free antigen and antibody valencies for the extension of the lattice indefinitely.

On this view the hydrophobe character of the larger aggregates is ascribed not only to a loss of attraction for water but also to a specific attraction between the particles or molecules of which the aggregates are composed. This conception as Marrack points out is entirely compatible with the dependence of flocculation on the presence of salt in moderate concentration. The action of the electrolyte in reducing the negative charge on the molecules or particles will assist aggregation whether this be due to a loss of attraction for water or to mutual attraction between one molecule or particle and another.

Similar arguments apply to the constant antigen titration. As antibody concentration increases the antigenic particles are more and more rapidly and fully sensitized, and the speed of flocculation increases. In antibody excess, the complexes formed are more soluble than those at the optimum. The marked inhibitory effect of excess antigen, however in the constant-antibody titration is not usually paralleled in the constant-antigen titration by inhibition in antibody excess. As we have seen (p 203) the addition of antibody in excess of constant antigen equivalence produces even more rapid flocculation so that the constant-antigen optimal ratio of antigen to antibody is higher than the equivalent ratio (see also Duncan 1934). Marrack (1934) suggested that if antibody but not antigen were denatured during the combination of the two compounds like A (Fig 33) would be soluble, since the antibody is protected by the fully hydrophile antigen whereas compounds like B would be hydrophobe since partial denaturation of the antibody at the surface would take place. Moreover since it appears that antigen has a greater valency than antibody in antibody excess there will be a greater packing of antibody round the antigen than of antigen round antibody in antigen excess. We have already noticed that in constant-antigen titrations of systems like diphtheria toxin and horse antitoxin antibody excess has a sharply inhibiting effect. Thus precipitation may be completely inhibited by doubling the concentration of antibody that gives optimal precipitation (Healer and Pinfield 1935; Pappenheimer and Robinson 1937). The work of Pappenheimer (1940) suggests that the narrow flocculating zone sharp inhibition by antibody excess, and the easily elicited Dancy phenomenon of horse antitoxic sera are dependent on the nature of the antibody. Horse antisera to diphtheria toxin and ovalbumin exhibited all three properties, whereas the corresponding rabbit antisera to the two antigens did not. Boyd (1941) came to a similar conclusion from a study

of the antigen antibody compound, but it is possible that some of them for instance the lessened absorption of agglutinins by typhoid bacilli at a pH below 4.0 observed by de Kruijff and Northrop may be due to the destruction or inactivation of the bacterial antigen concerned (see Duncan 1933a)

We are in the study of such reactions as these, dealing with systems of great physical and chemical complexity, about which we know relatively little. We should therefore be very cautious in ascribing any observed phenomenon to the influence of a particular physical or chemical factor until other possibilities have been satisfactorily excluded.

Before considering the effect of electrolytes and other factors on the actual flocculation of bacteria it will be convenient to note certain quantitative data in regard to the absorption of agglutinins by bacterial cells when these ancillary factors are kept constant. Here, as elsewhere, the observed relationships are inexplicable on the hypothesis of firm union in simple multiple proportions. For instance Eisenberg and Volk (1902) showed that, when a constant amount of bacterial suspension was allowed to react with varying concentrations of an agglutinating serum proportionately more agglutinin was absorbed from the more dilute serum though absolutely more was absorbed from the more concentrated serum. When a constant dilution of antiserum was absorbed with varying amounts of bacteria the amount of agglutinin bound did not bear a linear relation to the absorbing dose of bacterial cells. With increasing doses of bacteria the amount of agglutinin removed became proportionately less. Such a relationship is analogous to that observed in any adsorption process. Craw (1906) noted, in studying the absorption of agglutinin by bacteria a phenomenon analogous to that described by Danyasz in the toxin antitoxin reaction (see p. 216).

### The Zone Phenomenon in Agglutination

The system with which we are dealing in the agglutination reaction differs sharply from that concerned in precipitation in that the floccules consist mainly of the antigen carrying material the concentration of which in the initial mixtures can be judged from measures of opacity or from actual counts, of the bacterial suspension. For this reason quantitative measures of the agglutination reaction are most conveniently performed by the constant antigen method varying the concentration of antisera. The constant-antibody technique is possible but has the disadvantage that the initial heavy opacity of concentrated antigen suspensions may mask moderate degrees of flocculation. Titration of bacterium agglutinin systems by both techniques yields optimally flocculating zones though they are less well defined than precipitin zones. The failure of some bacterial suspensions to agglutinate in the higher concentrations of the dilution series of homologous antiserum the pro-zone has been noted by many workers. In other systems, the inhibition of agglutination is not absolute but can be observed particularly with light bacterial suspensions (Heuer 1922 da Costa Cruz 1939).

Duncan (1932b) and Miles (1933) studied the constant antigen optimum and its relation to the constant antibody optimum. As in precipitin systems the antibody content of the constant-antibody optimal mixture was greater than that in the constant antigen optimal mixture. In one system Duncan found the difference to be six fold. Equivalence zones cannot be fully defined since removing a precipitate by centrifugation in the zones of antigen-excess also brings down unagglutinated antigen, but the equivalence zone appears to cover a wide region of the constant-antigen series, including the optimal mixture.

Shibley (1929) observed the pro-zone phenomenon in antisera that had been exposed to moderate heat, and attributed it to a modification of antibody whereby it was preferentially absorbed by the bacteria, but had lost its power to sensitize. The probable

pH over which acid agglutination occurs may vary considerably from one bacterial species to another, and has sometimes been employed as a differential criterion, but its usefulness in this respect would seem to be very limited (see Sgaitzer 1914, Buchanan 1919)

The relation of salt agglutination to agglutination by specific antisera has been studied in some detail by Northrop and de Kruij (1922a, b). Working with *Salmonella typhi*, they showed that, with those kations that caused a great reduction in surface potential in low concentrations, flocculation of either sensitized or unsensitized bacteria tends to occur when the surface potential is reduced below about 15 mvt. With kations that reduce the surface potential only in higher concentrations unsensitized bacteria are not agglutinated, even when the surface potential is reduced to zero, while sensitized bacteria flocculate when the potential falls below 15 mvt. The effects produced on sensitized and unsensitized *Salmonella typhi* by the trivalent kation  $Al^{+++}$ , the bivalent kation  $Ba^{++}$ , and the monovalent kation  $Na^{+}$ , are shown in Fig. 34 (Northrop and de Kruij 1922a).

In an attempt to determine why unsensitized bacteria fail to agglutinate at relatively high salt concentrations, even when their surface potential is reduced to zero, Northrop and de Kruij (1922a) studied the effect of salt concentration on the "cohesive force" of bacterial cells, by measuring the forces required to separate two cover slips coated with bacteria. They found that the mutual attraction of unsensitized bacterial cells was reduced with increasing salt concentration, whereas this effect was not produced when the bacteria had previously been covered with serum protein. It would seem then that one of the ways in which serum protein or a specific antibody, exerts its effect is by so altering the bacterial surface that it reacts to a reduction of surface potential in relatively high salt concentrations in the same way that unsensitized bacteria react to an equal reduction of potential in very low salt concentrations.

Before leaving this question of the influence of electrolytes on agglutination, or on other antigen antibody reactions, we may note briefly the effect of salt concentrations higher than those that we have considered in the previous paragraphs. High concentrations of sodium chloride (2N or above) inhibit both precipitation and agglutination. These reactions may be delayed in concentrations of 0.2–0.5 N sodium chloride. (See Streng 1909, Friedberger and Goldschmidt 1910, Landsteiner and Welecki 1911, Eagle 1932.) Schmidt (1930) noted that the reaction was delayed in high concentrations of various salts, and that the order of activity of the anions was approximately that of the Hofmeister series, namely  $(ClO_4^{-} > SCN^{-} > ClO_3^{-} > NO_3^{-} > Br^{-} > IO_3^{-} > SO_4^{2-}/2 > Cl^{-} > NO_2^{-} > F^{-})$

and Marrack and Smith (1930) found that toxin-antitoxin floccules were dispersed by strong salt solutions the order of activity being  $I^{-} > SCN^{-} > Br^{-} > NO_3^{-}$ . (See also p. 230.) It is probable that the effect is due to a direct action of the salt on the serum proteins.

The effect of specific antibody itself on the surface charge of bacteria, at the pH (7–8) and the salt concentration (0.9 per cent. = 0.15 N NaCl) at which agglutination reactions are usually carried out, may be negligible (Shibley 1926, Marrack 1934), though some workers (see Brown and Broom 1929) have ascribed great importance to this effect.

There seems little doubt that the essential effect of sensitization is not its direct action on the surface charge, but the fact that sensitized bacteria react as hydrophobe colloids, even in moderately high salt concentrations, while unsensitized bacteria do not. These hydrophobe particles remain dispersed only when their surface potential is maintained at a level greater than about 13–15 mvt. When the charge is reduced below this level by the action of electrolytes the bacteria flocculate. It may be noted (Streng 1909, Northrop and de Kruij 1922b) that when the amount of antibody combined with the bacteria is very small, agglutination may occur only over a very narrow range of salt concentration.

### The Reversibility of the Union between Antigen and Antibody.

The formation and precipitation of insoluble antigen antibody complexes imply a certain degree of irreversibility, a degree which is illustrated by the Danyesz phenomenon referred to above. Danyesz (1902) showed that if a constant amount of diphtheria toxin is added to a constant amount of antitoxin, the toxicity of the mixture varies according to the way in which the addition is made. If, for instance, to a given amount of antitoxin, the equivalent amount of toxin is added all at once, the mixture is non toxic. If the same amount of toxin is added in two or more fractions, with an interval of 15 minutes or more between each addition the mixture will be highly toxic. The fraction of toxin added earlier unites with more than its equivalent amount of antibody, so that there is insufficient antibody to neutralize the later fractions, and there is, during the period of the experiment, no detectable redistribution of antibody among the toxin molecules. The solubility of the compounds formed in antigen or antibody excess, however suggests that if the union was reversible, a precipitate formed from equivalent amounts should dissolve in excess of antigen or antibody. The evidence on this point is conflicting.

Little solution in these circumstances could be detected in precipitates of the following antigens with their respective antisera. Types I and II pneumococcal polysaccharides (Sobotka and Friedlander 1928), hæmoglobin (Brain and Haurowitz 1930). On the other hand the following specific precipitates are soluble: diphtheria toxin in antibody and antigen excess (Healey and Pinfield 1935), Type III polysaccharide (Heidelberger and Kendall 1935a), hæmocyanin (Hooker and Boyd 1936) and ovalbumin (Boyd 1940).

Boyd found that the ovalbumin precipitates were as readily soluble after 10 months' storage in the cold. In other cases the antigen antibody union appears to become less reversible with time. For instance, both Enders and Shaffer (1937) and Morris (1940) found that dilute mixtures of living Type I pneumococci and protective antibody were at first infective but became less so if the mixtures were allowed to stand before injection into mice. It should be noted that the fact of decreased dissociability with age is not necessarily due to increasing firmness of combination, an irreversible denaturation of the antibody protein in the compound may take place (see also Chapter II, p. 342).

The solubility of antigen-antibody complexes in high salt concentrations and, in some cases in antigen or antibody excess, suggests that dissociation of the complex takes place. The establishment of this fact is important in deciding which of the better known chemical and physical processes best describes the phenomena of precipitation and agglutination. With Bordet, we may interpret these reactions as adsorption phenomena, or with Arrhenius and Madsen, as obeying the mass law. In the equations both for the mass law and the adsorption isotherm of Freundlich (see below) the amount of absorption complex, or of compound present in the system at equilibrium depends on the concentration of one or more of the reactants. However, in most precipitin systems studied (see p. 219) the composition and amount of the antibody antigen complex is dependent, not on the final, but on the initial concentration of the reactants, a fact that implies a relatively high degree of irreversibility of the antigen antibody union. The union takes place probably within a few seconds, in systems that precipitate visibly in a few minutes (see Burnet 1931, Boyd and Hooker 1938, Follensby and Hooker 1939, Heidelberger, Treffers and Mayer 1940, and Boyd, Conn, Gregg, Kistiakowsky and Roberts 1941). In this case the laws will apply only in the earlier stages of the reaction and their application to data obtained mainly from

When we come to the results obtained in partial neutralization or partial adsorption experiments we find that they are not compatible with the concept of firm union in constant proportions but that they afford no secure basis for distinguishing between an adsorption reaction and a combination due to intramolecular forces that is subject to the law of mass action

The law which describes the relation between the amount of a given substance adsorbed from a solution, and the concentration of that substance remaining in solution when equilibrium has been reached has been formulated by Freundlich (see Freundlich 1906 1922 Freundlich and Neumann 1909) and expressed in the form of the following equation

$$\frac{x}{m} = aC^n$$

where  $x$  is the amount of the substance adsorbed by the surface  $m$   $C$  is the final concentration of the substance in the fluid  $a$  is a constant depending on the units of measurement, and  $n$  is a constant less than unity

If we let  $x$  represent the adsorption on unit surface, the equation will become

$$x = aC^n$$

or 
$$\log x = \log a + n \log C.$$

The curves of the two equations are shown in Figs. 35 and 36

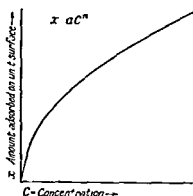


FIG 35

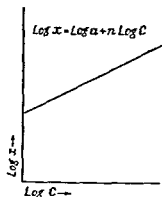


FIG 36.

It is clear that this formula expresses the fact that the amount of substance adsorbed by unit surface increases with increasing concentration, not in direct proportion to this increase but to some root value of it. If for instance, the value of  $n$  were 0.5 the amount adsorbed by unit surface would increase as the square root of the concentration. It follows that proportionately more of a dissolved substance will be adsorbed from a weak solution than from a strong one

The figures recorded in many antigen antibody reactions and particularly in the neutralization of toxin by antitoxin, fit a curve of this type very closely over a considerable part of their range but the calculated and observed values usually differ significantly with a very great excess of either reagent (see for instance von Krogh 1911) This discrepancy is not surprising (see Marrack 1933) The Freundlich isotherm, in its classical form, does not approach to a maximum at high concentration of one reagent but rises continuously though at a decreasing rate. This seems very unlikely to describe the course of any antigen antibody reaction. The adsorbing reagent would be expected to become fully saturated

When we come to the results obtained in partial neutralization, or partial adsorption experiments we find that they are not compatible with the concept of firm union in constant proportions, but that they afford no secure basis for distinguishing between an adsorption reaction and a combination due to intramolecular forces that is subject to the law of mass action

The law which describes the relation between the amount of a given substance adsorbed from a solution, and the concentration of that substance remaining in solution when equilibrium has been reached has been formulated by Freundlich (see Freundlich 1906 1922, Freundlich and Neumann 1909) and expressed in the form of the following equation

$$\frac{x}{m} = aC^n$$

where  $x$  is the amount of the substance adsorbed by the surface  $m$   $C$  is the final concentration of the substance in the fluid  $a$  is a constant depending on the units of measurement, and  $n$  is a constant less than unity

If we let  $x$  represent the adsorption on unit surface, the equation will become

$$x = aC^n$$

or  $\log x = \log a + n \log C$

The curves of the two equations are shown in Figs. 35 and 36

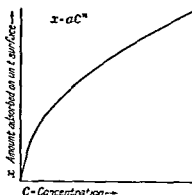


FIG 35

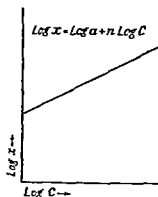


FIG 36

It is clear that this formula expresses the fact that the amount of substance adsorbed by unit surface increases with increasing concentration not in direct proportion to this increase, but to some root value of it. If, for instance, the value of  $n$  were 0.5, the amount adsorbed by unit surface would increase as the square-root of the concentration. It follows that proportionately more of a dissolved substance will be adsorbed from a weak solution than from a strong one.

The figures recorded in many antigen-antibody reactions, and particularly in the neutralization of toxin by antitoxin, fit a curve of this type very closely over a considerable part of their range, but the calculated and observed values usually differ significantly with a very great excess of either reagent (see, for instance von Krogh 1911). This discrepancy is not surprising (see Marrack 1935). The Freundlich isotherm, in its classical form, does not approach to a maximum at high concentration of one reagent but rises continuously though at a decreasing rate. This seems very unlikely to describe the course of any antigen-antibody reaction. The adsorbing reagent would be expected to become fully saturated

at some point. Modified equations have in fact been evolved that allow for such a saturation limit.

With this proviso, the demonstration that there is a close correspondence over most of the range between observed values and values calculated on the basis of Freundlich's isotherm clearly suggests the possibility that antigen antibody reactions are examples of adsorption phenomena. But an equation derived from the law of mass action gives a curve that has a close resemblance to the Freundlich isotherm over a great part of its range. It differs from it in rising rather less steeply at low concentrations of the reagent undergoing adsorption and asymptoting towards a maximum at high concentrations. Arrhenius and Madsen (see Arrhenius 1915) give several examples in which there is a clear correspondence between the observed values and those calculated on the mass action hypothesis over a considerable range of antigen antibody reactions though here again there are usually discrepancies at high concentrations of one or other reagent. Biltz (1910) found that the curve of neutralization of tetanolyisin by an antitetanic serum would fit the Freundlich isotherm, but would also fit a curve based on the mass action equation. It does not in fact seem possible, on the basis of the recorded data, to decide between adsorption and mass action by invoking the form of the neutralization curves obtained.

The agreement of experimental data with a theoretical equation does not guarantee its validity, whether the equation is derived from the mass law or Freundlich's isotherm. The adsorption isotherm can be used to describe a number of different statistical phenomena such as association between death rates and overcrowding (Brownlee 1925). It describes, in fact, the statistical behaviour of many complex systems of interrelated events, and the agreement of data with it should probably be regarded as evidence of a complex reaction, and not as an indication of the nature of the factors producing the complexity.

The mass law has been applied by Heidelberger and his colleagues to the extensive data they obtained from an analysis of various specific precipitin reactions. Type III pneumococcal polysaccharide, azoprotein dye, egg albumin (Heidelberger and Kendall 1935a & c, 1937), thyroglobins (Stokinger and Heidelberger 1937) and serum albumin (Kabat and Heidelberger 1937).

The hypothesis assumes that antibody may be considered as homogeneous, that antigens and antibody are multivalent with respect to one another, and that before precipitation there must occur a competing set of bimolecular reactions, whose nature depends on the relative concentrations of antigen and antibody. If antigen and antibody combine in equimolecular proportions an equation applicable to all cases in which there is excess of antibody is

$$y = 2x - x^2/A,$$

where  $y$  is the number of molecules of antibody precipitated when  $x$  molecules of antigen are added, and  $A$  is the number of antibody molecules precipitated at the equivalence point of a constant antibody titration. There are a number of objections to this derivation. Among them is the fact that there are not always equimolecular amounts of antigen and antibody at the equivalence point (see Marrack, 1938; Heidelberger 1939). Without assuming equimolecular combination at equivalence the following equation derived from experimental data is found applicable to many systems

$$y = 2Rx - R^2x^2/A,$$

where  $x$ ,  $y$  and  $A$  are expressed in milligrams and  $R$  is the ratio of antigen to antibody at the equivalence point. This and other similar equations, are empirically successful



in characterizing antisera in terms of standard antigenic preparations and *vice versa*. Antibody content may be expressed in terms of maximally precipitated nitrogen and degrees of antigenic similarity as the proportion of homologous antibody precipitable by different antigenic preparations. Heterologous systems are not so readily characterized by this method. For instance Pennell and Huddleson (1938) found a good fit with the precipitation of *Brucella abortus* and *melitensis* endo-antigens by homologous antisera, but the fit was poor with the cross-reacting systems, *abortus* antigen and *melitensis* anti-serum. This may be due to peculiarities of the antigenic relationship between these two organisms (see Chapter 34). A more important objection is that the relation does not hold for the region of antigen excess (Malkiel and Boyd 1934) though Heidelberger (1939) points out that this region is characterized by the co-existence of soluble and insoluble antigen-antibody complexes in equilibrium and his original assumptions do not necessarily apply.

Hershey (1941a & 1942, 1943a) has expressed the main features of specific precipitation in a descriptive theory that differs from those of Heidelberger and Kendall in a number of respects. The mass law is not invoked. Hershey adopts the notion of competing bimolecular reactions but unlike Heidelberger and Kendall assumes that the initial antigen-antibody combination is reversible. The compounds may be characterized in terms of valency of antigen and of antibody and a dissociation constant  $k$ . Of special interest is his restricted theory in which the lattice hypothesis is assumed. That is both antigen and antibody are multivalent and not only the initial combination but the later aggregation is determined by antigen and antibody valencies. The original papers must be consulted for the details of the theory. It may be noted that by assuming that aggregates as well as initial compounds have a determined lattice structure the assumption of irreversibility of reaction is not only unnecessary but is irreconcilable with the lattice hypothesis (1943a). The agreement of the theory with available data is as good as that of Heidelberger and Kendall's and the theory is applicable over a wide range of the precipitin reaction. The consequences of the restricted theory are compatible with a large number of observed facts including insolubility of precipitates, the different effects of antigen and antibody excess and the varying relation of optima with various points of reference in the equivalence zone. Taken in conjunction with the experimental data, the success of the restricted theory at least provides an additional reason for giving the lattice hypothesis the first place among the working hypotheses of serology. (See also Ghosh 1935, Kendall 1942 and Pauling, Pressman, Campbell and Ikeda 1942.)

### Tests of the Lattice Hypothesis

The description of the precipitin reaction in terms of Heidelberger and Kendall's mass law equations corresponds in essentials with the lattice hypothesis of Marrack. Both depend on the assumption that antibodies are multivalent. The main objections advanced against the conception of multivalent antibody are firstly the difficulty of conceiving the mode of formation of multivalent antibodies in the animal and secondly that the assumption of multivalence is unnecessary to describe the phenomena of precipitation. The first point is dealt with more fully in a later section (p. 251). Hooker and Boyd (1937) object to the assumption of multivalence on the grounds that inhibition by antibody excess is rare, that non-specific adjuvants like electrolytes and lipins are necessary for precipitation, that the surface properties of bacteria sensitized by antibody are those of a hydrophobic salt sensitive denatured globulin and that in the presence of excess of antibody the rate of flocculation of an antigen is similar to a simple colloid undergoing non-specific flocculation. The same authors (1942) point out that analysis of precipitates formed in the zone of antigen excess where antibody is not likely to have all its valencies satisfied seldom reveals a molecular ratio of antigen to antibody of more than unity.

Eagle (1938) subjected diphtheria antitoxin and antipneumococcal serum to progressively intensive treatment with formaldehyde. At one stage the antibodies lost their precipitating power but were still protective. A separation of activities that suggests a non-specific element in aggregation. Kleczkowski (1941b) and Byden and Kleczkowski (1942b) made complexes of antibody globulin with albumin, in which the antibody retained its specific combining power, but was incapable of *in vitro* flocculation. Phenomena of this kind do not necessarily invalidate the lattice hypothesis but they give an indication of the degree to which non-specific factors may be operative in aggregation.

Tooley, Wilson and Duncan (1935) devised a test of the multivalence of antibody using mixed agglutination systems. Two antigens are chosen of approximately equal particle size, and in similar concentration. The amount of antibody which in each case will flocculate the antigen in a given time is determined and the reaction of the two antigens and antibodies allowed to take place in a mixture. If the lattice hypothesis holds the aggregates that form will consist of one antigen or the other since they are held together by specific antibody. If flocculation is non-specific, the aggregates will be mixed. Moreover in the first case the speed of flocculation of each antigen will be independent of the presence of the other, but if antibody merely sensitizes the antigens to the action of electrolyte, then the concentration of flocculable material in the mixtures will be doubled and the speed of the mixed reaction much greater than that of either reaction separately. Tooley, Wilson and Duncan found that pneumococci, and the microscopically distinguishable coliform bacilli formed separate aggregates. With mixtures of two distinguishable types of red blood cell, Atkinson (1935) and Hooker and Boyd (1937) found mixed aggregates. Wiener and Herman (1939), on the other hand found separate aggregates with mixtures containing red cells and a precipitating antigen red cells and pneumococci and two types of red cells. Mixed aggregates were formed with the latter when antibody was in excess. Non-specific flocculants also gave mixed aggregates. With precipitating systems, Hooker and Boyd (1937) found an increased speed of flocculation in mixtures indicating non-specific aggregation. Duncan (1939) confirmed this but with agglutinating systems, found no increase in speed of agglutination. He suggested that both mechanisms were operative, but that one or the other might predominate according to the system and the circumstances of its reaction. Heidelberger and Kendall (1937) saturated pneumococci with horse antibody. The addition of more pneumococci to the over-sensitized cells resulted in prompt agglutination of all the pneumococci a result strongly suggesting that, in addition to the valencies employed in binding antibody to the original pneumococci there were on the bound antibody free valencies available for combination with the added cocci. Boyd and Hooker (1938) suggest that in mixed systems specific combination may operate in the second stage, and a lattice may form in antigen excess. In antibody excess, the "two stage" reaction appears to take place, for they found that red cells saturated with a very marked excess of antibody were still susceptible to the flocculating action of salts. Hershey's (1943b) recent experiments, made on somewhat similar lines with bacteriophage, also support the lattice hypothesis. A living *Bact. coli* bacteriophage adhered firmly to a precipitate consisting of dead *coli* bacteriophage and homologous antibody. Other specific aggregating systems with *eg.* staphylococcal phage, serum proteins, pneumococcal polysaccharide, absorbed only a small amount of the phage. He obtained no evidence that the particles of antigen were rendered non-specifically "sticky" by their coating of antibody, a condition to be expected if aggregation was non-specific.

Another experimental approach to the problem of antibody valency is possible. No safe assumption can be made of the precise valency of natural antigens. But, as we shall see later (p. 258), certain fractions of natural antigens will react with antibody, provided they possess the chemical groupings that characterize the full antigen. These reacting fractions are called haptens, and in some cases can be prepared synthetically. The reaction of monovalent and divalent synthetic haptens with antibody should provide a clue to the valency of antibody. Mono-

valent haptens do not precipitate with homologous antibody, though they can be shown to combine with it. It is argued (Marrack 1933, Pauling 1940) that if antibody is divalent, divalent haptens should aggregate with it in long chains, with tri or multi valent antibody, divalent haptens could form a lattice and precipitate.

Precipitation of divalent haptens was observed by Landsteiner and van der Scheer (1932b) and by Pauling, Campbell and Pressman (1941), and Pauling, Pressman and Ikeda (1942) showed that the hapten-antibody ratio was constant throughout the titration range of a divalent hapten with homologous antiserum. The observed molar ratio was 0.75 as against the ratio of unity to be expected if both the antigen and the antibody were bivalent. Hooker and Boyd (1941b) and Boyd (1942) obtained no evidence of precipitation or of the formation of long chains of alternating antigen and antibody. Landsteiner and van der Scheer's haptens may have polymerized in solution and so formed multivalent hapten particles, though there is no positive evidence that the same criticism is applicable to the results of Pauling and his co-workers. Boyd (1942) from a study of specifically precipitable trivalent arsonic haptens, concluded that their precipitability depended, not on their power to form a lattice, but on the closeness of their combining groups to one another. When the arsonic combining groups were attached to a compound so that they were separated by a relatively large molecule, the hapten was not precipitable by antibody. Boyd suggests that after the arsonic groups of this latter type of hapten have united with antibody, the number of hydrophilic groups remaining free is sufficient to keep the compound in solution. When these groups were rendered less hydrophilic by acetylation, the haptens became precipitable. Precipitation on this basis is due to the lowering of solubility by mutual neutralization of polar groups in antibody and hapten, and by crowding of antibody molecules together so that other polar groups are occluded. This 'occlusion' hypothesis is an extension to haptens of the 'two-stage' theory as applied to the reaction of full antigens in antibody excess. The serological reactions of haptens, however, are not strictly parallel to those of full antigens, and this lack of parallelism may invalidate any conclusions that we may draw about the antigen-antibody reactions. For example, Woolf (1941), working with antigens synthesized by conjugating protein to hapten, observed that the antigen-antibody ratios were dependent, as in other systems, on the amounts, and not on the concentrations of the reactants. Inhibition of the reaction by hapten, on the other hand, depended on its concentration, a fact which suggested that hapten-antibody compounds were far more dissociable than antigen-antibody compounds. Moreover Hershey (1942) points out that experiments with haptens are unlikely to indicate anything about the valency of antibody, since the discrepancy in size between the reactive groups of the haptens and a reactive patch (i.e. unit valency) on the antibody is such that a modified lattice will form in the early stage of precipitation, whether antibody is monovalent or not. In a later paper (1944) he shows on theoretical grounds that the combination of antibody with divalent hapten would not result in specific precipitation, since it is highly unlikely that sufficiently long chains of alternating antigen and antibody molecules could form, precipitation in such circumstances would imply either a non-specific effect, or irreversible antigen-antibody linkages, or, as already noted, either an increase in the valency of hapten by its polymerization, or an effective multivalence of unit reactive patches on the antibody molecule.

It is impossible at present to decide between the lattice and the two-stage hypotheses, at any rate as mutually exclusive hypotheses. The proponents of both are united in assuming that the initial combination is determined by specific chemical factors and that antigens are multivalent. It should be noted that the demonstration of the multivalence of antibody will not alone prove that a specific lattice is formed, it will be necessary also to show that the antigen-antibody linkages are specific, and that the bonds between the molecules are reversible.

(Hershey 1944) The resolution of the controversy either by the establishment of one or the other hypothesis, or their merging into a more general hypothesis, must await further research

#### The Microscopic Appearance of Specific Aggregates

There are few observations of specific aggregates, either during or after their formation which yield any information about the nature of the antigen antibody union. Pijper (1939, 1941a, b) records an interesting study of the agglutination of *Salmonella typhi* made by dark ground cinematography. He confirms the well-established fact that flagellar antibody first immobilizes the organisms by a direct action on the flagella after which aggregates are formed by the adhesion to one another of the motionless flagella of bacteria fortuitously brought into contact by Brownian movement or convection currents (Fig. 37). Agglutination by antibody to the somatic antigens results in the formation of closely packed clumps of bacteria, which retain their motility since the flagella are not immobilized. The cinematograph record shows the motile, sensitized bacteria swimming in all directions across the field along relatively straight lines, and many of them are seen swimming directly towards already formed aggregates, a phenomenon that Pijper attributes to



FIG. 37.—Dark ground photomicrograph of *Salmonella typhi* aggregated by flagellar antibodies (From a photograph kindly supplied by Dr. A. Pijper)

attraction exerted by the aggregate. It is difficult to imagine that the union of antigen and antibody could generate a field of force capable of acting at distances of several microns, and it appears to us that the apparent attraction can be explained by postulating a straight-line course for the motile sensitized bacilli over short distances. If an aggregate is in the line of travel, a head-on collision with the aggregate and adhesion to it will give the impression of attraction.

The bacterial aggregates formed during agglutination varied with the antibody used. Sensitization by flagellar antibody gives the fortuitous pattern already noted; by somatic O antibody, an end-to-end arrangement (Fig. 38), and by Vi antibody (see Chapter 8), the characteristic packing depicted in Fig. 39. The factors that determine these patterns are matters for speculation. It may be noted in this connection that Miles and Pine (1939) observed specific precipitates of antibody and native antigen from *Brucella melitensis* which appeared to be built up of bacterial fragments adhering to each other end-to-end.

During the action of flagellar antibodies, Pijper observed the deposition of granules on the flagella. Mudd and Anderson (1941) found a patchy distribution of thickenings on the flagella of specifically sensitized *Salmonella typhi*. Measured in electron micrographs, the increase in thickness varied from 1 to 17  $\mu\mu$ . Similar thickenings occurred on the cell wall. The dimensions of the thickenings were of an order of size compatible with those of a unimolecular layer of rabbit antibody.

The sensitized patches appeared to be sticky, not only for each other, but for non specific particles. No estimate of the degrees of stickiness is possible from an electron micrograph but if we assume that the non specific and specific stickiness were of the same order, the observation provides evidence in favour of the two-stage rather than the lattice hypothesis.

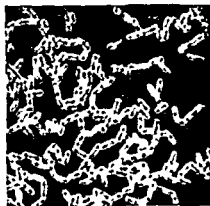


FIG. 38.—Dark ground photomicrograph of *Salm typhi* aggregated by somatic antibodies.

(From a photograph kindly supplied by Dr A. Eljper.)



FIG. 39.—Dark ground photomicrograph of *Salm typhi* aggregated by Vi antibodies.

(From a photograph kindly supplied by Dr A. Eljper.)

In a mixture of a tobacco mosaic virus having a particle width of  $15\text{ m}\mu$ , and its homologous antibody Anderson and Stanley (1941) demonstrated that, in antibody excess the increase in width of the virus particles was compatible with the hypothesis that highly elongated rabbit antibody molecules, about  $4 \times 27\text{ m}\mu$  were attached radially to the surface of the antigen. The authors point out that electron micrographs of the aggregates suggest a rough lattice formation. Such a resemblance is irrelevant in a consideration of Marrack's hypothesis in which 'lattice' is used more in a crystallographic, than in a secular sense of the term.

### The Lysis of Red Blood Cells

Although the original observations on the phenomenon of lysis were made on bacteria, Bordet's demonstration that a similar reaction could be obtained with red blood corpuscles, and the numerous and detailed investigations of Ehrlich and his school into the interactions between red cells and their corresponding antisera, turned the attention of the great majority of workers from bacteriolysis to hæmolysis, and the greater part of our knowledge of the mechanism of lytic reactions in general is based on the data obtained in studying the lysis of red cells. For this reason we shall discuss hæmolysis before considering the lysis of bacteria.

It is not possible, in the space at our disposal, to give any account of the historical development of our knowledge of this subject. Reference must be made to textbooks dealing particularly with immunological reactions, or to the collected papers of Bordet and of Ehrlich, who have been the protagonists in this particular controversy. The main facts, which are not in dispute, are as follows.

The phenomenon of hæmolysis consists in a laking of the red blood corpuscles, that is, in a setting-free of their contained hæmoglobin, and not in a true solution. The cell stromata remain undissolved, though altered in size and shape, and probably in other physical and chemical characteristics.

As has already been noted, it was shown by Bordet (1898) that hæmolytic sera, like the bactericidal sera studied by Buchner, are inactivated by heating for 30

minutes at 56° C. He also showed that this inactivation concerned not the hæmolytic antibody itself but a second non specific thermolabile factor, which caused the lysis of the red cells when these had been sensitized by the specific hæmolysin. This non specific, thermolabile factor, which is present in all normal, fresh, unheated sera, was named *alexine* by Buchner. It is now generally known by the name *complement* employed by Ehrlich.

The fundamental reactions that demonstrate the nature of the lytic reaction may be briefly summarized as follows. The defibrinated or citrated blood of a suitable animal, such as the sheep, is centrifuged, and the deposited red cells are separated and washed several times in saline to free them from the last traces of serum. The washed cells are then made into a 5 per cent suspension in saline. The serum from some convenient animal—usually a rabbit—that has received repeated injections of washed sheep corpuscles and has in consequence produced a specific hæmolysin to high titre, is heated at 56° C for 30 minutes to inactivate the normal complement. The fresh, unheated serum of some other animal, usually the guinea pig, is used as a source of complement. When these reagents are mixed in various combinations, and the mixtures incubated at 37° C, the following results are obtained.

- |  |   |                    |
|--|---|--------------------|
| (1) Red cells + Hæmolysin              | → | No hæmolysis       |
| (2) Red cells + Complement             | → | No hæmolysis       |
| (3) Red cells + Hæmolysin + Complement | → | Complete hæmolysis |

If mixtures (1) and (2) are centrifuged, and the deposit and supernatant fluid examined separately for the presence of hæmolysin and complement, the following results will be noted, provided that the proportions of the reagents in the original mixtures have been suitably adjusted.

- |   |   |                    |
|---|---|--------------------|
| (4) Deposit from (1) + Complement                 | → | Complete hæmolysis |
| (5) Supernatant from (1) + Red cells + Complement | → | No hæmolysis       |
| (6) Deposit from (2) + Hæmolysin                  | → | No hæmolysis       |
| (7) Supernatant from (2) + Red cells + Hæmolysin  | → | Complete hæmolysis |

Reaction (4) shows that hæmolysin has combined with the red cells in (1) and sensitized them to the lytic action of complement. Reaction (5) confirms this by demonstrating the absence of hæmolysin from the supernatant fluid. Reaction (6) shows that complement has not combined directly with the red cells in (2). Reaction (7) confirms this by demonstrating the presence of complement in the supernatant fluid.

The controversy that arose between Bordet and his co-workers on the one hand, and Ehrlich and his school on the other, concerned the mode of union between the complement and the sensitized red cells. As already indicated, Ehrlich's conception of the hæmolytic antibody was that of a special type of side chain, which he referred to as a receptor of the third order, or an amboceptor. He endowed this hypothetical receptor with two haptophore, or combining groups, one of which united with the red cell, and was named by him the cytophilic group, while the other united with the complement, and was named the complementophilic group. Ehrlich's conception of the structure and mode of action of an amboceptor may be represented as in Fig 40. In the diagram, R C represents the red cell, and R one of its receptors, A the amboceptor, or hæmolysin, attached to the receptor of the red cell by its cytophilic haptophore group Cy, and to the complement by its complementophilic haptophore group Cm. Crepe

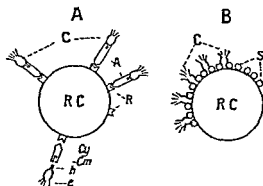


FIG 40

sents the complement, with its haptophore group *h*, by which it is attached to the complementophilic group of the amboceptor, and *e* its ergophore group, in virtue of which it brings about lysis of the red cell, when united to it by the hæmolytic amboceptor.

Bordet's conception of the action of the hæmolytic antibody, which he refers to as the "*substance sensibilisatrice*," or sensitizer, is essentially different. He

denies that the sensitizer acts as a link between the red cell and the complement, or that the complement ever unites with the sensitizer as such. He regards the combining affinities of the hæmolytin as being directed entirely upon the red cell. The complement, in his view, unites not with the hæmolytin, but with the complex, red cell + hæmolytin, or, to put it in another way, with the red cell as altered by union with the hæmolytin. Bordet's conception may be represented diagrammatically in Fig 40B. RC represents the red cell, S the sensitizer or hæmolytin, which has united with the red cell, and altered the physical conditions at the surface, and C the complement, uniting with the red cell which has been so altered.

It will be seen that the controversy can be narrowed down to the question what evidence is there for the existence of a complementophilic group of the hæmolytin? We have not space to present all the experimental data which have been advanced by each side in turn, nor the arguments which have been based upon them. It may, we think, be safely asserted that Ehrlich and his supporters have, in this respect, failed to substantiate their position, and that the weight of evidence strongly upholds Bordet's view.

The quantitative aspects of the union between red cells and hæmolytin have been studied by many observers, and the results recorded have been of the same general kind as those observed with mixtures of soluble antigens and the corresponding antibodies, or with bacteria and their homologous agglutinins. They are not compatible with the view that red cells and hæmolytin unite in constant proportions, in a firm chemical union. They are compatible with the view that the union obeys the laws of an adsorption process—for instance, Bordet was able to demonstrate a reaction analogous to the Danysz phenomenon. They show (see Muir 1909) that the red cell hæmolytin compound can be dissociated under certain conditions, though the dissociation is not of the type that occurs on simple dilution of a dissociable chemical compound. There is, indeed no reason to suppose that the laws that determine the union of a hæmolytin with the specific antigenic components of a red cell differ in any way from those that determine the union of precipitins or agglutinins with their corresponding antigens.

Not all the antigenic groups on the surface of the red cell appear to take part in the sensitization of the cell to the action of complement. Heidelberger, Weil and Treflors (1941) calculate that, in a red cell suspension sensitized by one minimal hæmolytic dose of hæmolytin (see below) at most only 14 per cent of each cell surface is occupied by antibody. A figure of the same order, 6 per cent, was estimated by Haurowitz and Yenson (1943).

The effect of electrolytes and hydrogen ion concentration on the union of hæmolytin with red cells has been referred to on p. 211, in relation to the union of agglutinins with

bacteria. It may be noted (see, for instance, Markl 1902, Topley 1915) that electrolytes in high concentration have an inhibitory effect on the union of complement with sensitized red cells, and on the resulting hæmolysis.

The effect of temperature on hæmolysis differs somewhat from its effect on precipitation and agglutination, because we have an additional reagent to consider—the complement—and this reagent is thermolabile. Union between red cells and hæmolyisin occurs readily at 0° C (see, for example, Ueno 1938). At this temperature complement also unites with sensitized red cells though very much more slowly, but lysis fails to occur over any ordinary period of observation, unless excess of hæmolyisin and complement is present. At room temperature lysis occurs but so slowly that this temperature is unsuitable for experimental purposes. The optimal temperature for hæmolysis is in the neighbourhood of 37° C. At temperatures much above this the complement is inactivated, the rapidity of inactivation increasing as the temperature is raised.

The problem of the titration of a hæmolytic serum differs from that of titrating precipitins or agglutinins in that we are faced with three dependent variables instead of two. Our essential reagents are red cells, hæmolyisin and complement. Of these reagents, it is natural to keep our red cells constant, regarding as our end point the lysis of a specified quantity of red cells, in a specified time under specified conditions. It is a common practice to use a 5 per cent suspension of red cells in saline, and to employ some convenient volume of this suspension, such as 0.5 ml. in our tests. We have two reagents left, hæmolyisin and complement. The natural, and usual, plan is to vary the amount of the reagent that we want to measure—the hæmolyisin—while keeping the complement constant. But here we meet a difficulty. We can measure our reagents only in terms of their activity and they are dependent variables, the more complement we add, up to a point the less hæmolyisin we need, and *vice versa*. We get out of our difficulty by making use of the kind of relation that exists between our variables. However much complement we add, we need a certain amount of hæmolyisin. However much hæmolyisin we add, we need a certain amount of complement. Moreover, the limit at which further additions of one reagent make no appreciable difference to the required amount of the other is not a high multiple of the minimal dose. So we define our units of measurement as follows.

*The Minimal Hæmolytic Dose (M H D) of hæmolyisin is the smallest amount that will cause complete lysis of an arbitrarily selected amount of red cells, in the presence of excess of complement, in 1 hour at 37° C.*

*The Minimal Hæmolytic Dose (M H D) of complement is the smallest amount that will cause complete lysis of an arbitrarily selected amount of red cells, in the presence of excess of hæmolyisin, in 1 hour at 37° C.*

In an actual titration we proceed as follows. We first titrate our complement which must be used fresh, adding varying dilutions of our complement containing serum to mixtures of red cells and a hæmolytic serum of known titre. A hæmolytic serum, it may be noted, remains stable over a considerable period of time. In this titration we use excess of hæmolyisin say 5 or 6 M H D. We note the last tube that shows complete hæmolysis, and this gives us the M H D of our complement. We now dilute our complement so that the volume we intend to use, commonly 0.5 ml., contains 3–5 M H D, and to a series of tubes we add 0.5 ml. of our red cell suspension. 0.5 ml. of our diluted complement, and 0.5 ml. of the hæmolyisin under test, the dilution of the latter increasing from tube to tube. After 1 hour at 37° C. we note the last tube that shows complete hæmolytic and this gives us the M H D of our hæmolyisin.



### The Lysis of Bacteria.

Although we talk of bacteriolytins, antibodies that lyse bacteria and bactericidins antibodies that kill them, the two effects are not differentiated in practice, and though we often talk of the bacteriolytic titre of a serum it is the bactericidal effect that we actually measure. It is true that in many instances, as in Pfeiffer's classical experiments on the lysis of the cholera vibrio, gross degenerative changes in the bacteria have been observed and there is no doubt that those bacteria that are susceptible to the lethal action of complement in the presence of a specific antibody undergo a change in structure that is analogous to the lysis of red blood corpuscles. The change is not however, of the same dramatic kind, and the naked-eye observation of the changes in a turbid bacterial suspension is not a practical method of observing bacteriolysis. The principles involved in the reaction do not differ from those concerned in haemolysis and the method employed in titration is essentially similar.

A very light bacterial suspension is employed and the serum under test after its natural complement has been inactivated by heat is mixed in increasing dilution with a constant amount of the bacterial suspension and a constant amount of complement. The surviving bacteria in the mixtures are counted after varying intervals by some suitable cultural method and the highest dilution of serum that produces a significant killing effect is noted. The method contains many possible sources of technical error. Gengou (1899), for instance, drew attention to the fact that bacteria are agglutinated as well as lysed by a specific antiserum, and that this effect may greatly reduce the number of colonies in a plate count, since a clump of bacteria will produce a single colony. When the original mixtures are incubated for long periods before plating this effect will be counterbalanced by the multiplication of those bacteria that have been agglutinated but not killed, but this possible source of error must be considered in estimating the significance of any reduction of the viable count over a short period, and a control mixture containing the bacteriolytic serum without complement must always be included in the test.

It was noted by Neisser and Wechsberg (1901) that a marked pro-zone often occurred in tests of this kind. A particular dilution of serum might exert no bactericidal action while a much higher dilution resulted in a complete killing of the bacteria. This phenomenon was seized on by the Ehrlich school as an example of the union of complement with free amboceptor, the hypothesis being that complement united indifferently with all the amboceptor present and that, when this was present in excess, chance would favour the union of all the complement, or the greater part of it with the amboceptor that was not attached to bacteria. We know, however from our experience with precipitation and agglutination, that a similar inhibitory effect is produced by a great excess of antibody in reactions in which complement plays no part and it is clear that the use, in bactericidal tests, of very thin bacterial suspensions will favour the frequent occurrence of zones of gross antibody excess. The Neisser-Wechsberg phenomenon cannot, therefore, be regarded as affording any support to the view that complement combines directly with antibody.

A point of considerable importance in relation to the bactericidal, or bacteriolytic, reaction is that its occurrence depends in large part on the nature of the bacterial cell. Certain bacteria such as the cholera vibrio the typhoid bacillus, and most Gram-negative bacilli are readily killed and lysed when acted on by complement after sensitization by a specific antibody. Other bacteria, such as the Gram-positive cocci, are insusceptible to the direct killing action of antibody and com-

plement. And it should be noted that this insusceptibility is not due to any failure of the sensitized cell to combine with complement (see Buxton 1905a, b, c, Muir and Browning 1909).

### The Nature of Complement

We have seen that complement is a non specific substance present in all normal sera, and not increased in amount as the result of immunization. It does not follow that complement is a single substance, or a single system. Different kinds of antibodies, for instance the antibodies acting on different species of red cells might require different kinds of complement all, or many of which might be present in any specimen of normal serum. This question may be analysed into at least three components. (1) As regards one kind of lysis for instance that of red cells, is the complement present in a given specimen of serum a single entity or are there separate complements corresponding to the different hæmolytic antibodies? (2) If only one kind of complement is concerned in hæmolysis is it the same, or a different complement which brings about the lysis of bacteria or of other organized cells? (3) If the complement in a given specimen of serum is one and the same, irrespective of the kind of cells which are lysed or of the particular antibodies which are sensitizing them, is the complement in different sera and particularly in sera from different animal species always the same?

This problem afforded one of the most closely debated points in the long controversy between Ehrlich and Bordet, and the question has been investigated by many other workers. Space does not allow us to reproduce the arguments employed, nor the experimental evidence on which these arguments were based. For this reference is best made to the original papers which contain interesting examples of the complexity of the hypothetical receptor apparatus employed by Ehrlich to describe his experimental results. The provisional answers to these questions which are, in our view, afforded by the available evidence are as follows.

There is no evidence that more than one kind of complement is concerned in hæmolysis.

The evidence strongly suggests that the complement, in a given serum which causes lysis of one type of cell, for instance the red cell is identical with that which causes lysis of another type of cell, for instance a bacterium.

It is clear that sera derived from different animal species show qualitative differences in their complementary activities, and the available evidence indicates that the differences are due to both qualitative and quantitative variations in the components of the complement concerned.

Accepting the view that there is little, if any, evidence in favour of the existence of a multiplicity of complements in the sense employed above it remains to inquire whether complement is a single chemical substance or is a name for a property of normal serum that is dependent on a number of different factors. Here the answer is not in doubt. The complementary action of fresh unheated serum depends on the interaction of a number of separate components of which there appear to be four.

Complement becomes inactive if the englobulins are precipitated by removal of electrolytes. The remaining fluid is inactive. If the precipitate is removed, and dissolved in saline it also is inactive, but a mixture of the two fluids will cause hæmolysis of sensitized red cells (Ferrata 1907, Brand 1907, Liefmann 1909, Skwirsky 1910, Amako 1911, Gengou 1911). The 'globulin' fraction is known as "mid piece," and the soluble albumin fraction as "end piece," because

mid piece unites directly with sensitized cells whereas end piece unites with such cells only in the presence of mid piece. In various complement fixation reactions, mid piece is mainly absorbed, the greater part of the end piece being left in solution. Both mid piece and end piece are inactivated by heat.

It has long been known that a serum loses its complementary power after absorption by yeast. Such a serum is rendered active again by adding serum in which mid piece and end piece have been destroyed by heating to 56° C. (Coca 1914). This third heat stable component is present in both mid piece and end piece, as prepared by Liefmann's method of mid piece precipitation by CO<sub>2</sub> (Whitehead, Gordon and Wormald 1925).

Gordon, Whitehead and Wormald (1926a, b) demonstrated a fourth component of complement which is heat stable, but not absorbed by yeast. It is specifically inactivated by treating the serum with ammonia. Though this component is necessary for the hæmolytic action of complement, it is not necessary for the complementary effect of normal serum in the opsonic reaction (see p. 236). The existence of the fourth component was confirmed by Deisler (1932) and many subsequent workers.

The demonstration of components by inactivation with specific chemicals must be viewed with caution for activity is associated with proteins and agents which denature the proteins will also destroy complement. The postulation of a benzene inactivated component by Tokano (1936), a dialysable component by Chow and Zia (1935) and of the necessity for ionized calcium (Ottolenghi and Mori 1905) were probably based on observations of denaturation (see Jones and Ecker 1940, Pillemer and Ecker 1941a, Ecker and Pillemer 1942).

Complement is most active at a pH of about 7.3 in the presence of physiological saline. Its activity is progressively reduced by increases and decreases of pH and salt concentration. Its inactivation by various salts was studied by Gordon and Thompson (1933a, b) who found that the activity of various ions fell into the general order of the Hofmeister series (SCN<sup>-</sup> > I<sup>-</sup> > Br<sup>-</sup> > NO<sub>3</sub><sup>-</sup> > SO<sub>4</sub><sup>-2</sup> > Cl<sup>-</sup>). Some of the salts produced a reversible others an irreversible inactivation. It loses activity in a few days at 0° C. and in a few minutes at 56° C. According to Wehmer (1941) the inactivation of complement at any given temperature between 4° C. and 56° C. proceeds like one unimolecular reaction between 0° C. and 37° C. and another between 50° C. and 56° C. At the lower temperatures only mid piece is labile; at the higher both mid piece and end piece are labile; end piece predominantly so. In the 30–50° C. range the rate depends on the velocities of two unimolecular reactions.

Dry complement that has been kept for months is still active when reconstituted (Craigie 1931). Filtration through a porcelain candle (Strong and Culbertson 1934) inactivates complement mainly by retention of mid piece and end piece.

Many attempts have been made to isolate the components of complement (see Ecker and Pillemer 1942). The association of mid piece activity and end piece activity with 'globulin' and 'albumin' fractions of serum is to a large extent illusory for as Pillemer and Ecker (1941b) have shown, the electrophoretic patterns of both fractions show four distinct proteins, two of those in "globulin" fraction differing from any observed when normal guinea pig serum is analysed by this method. Inactivation of the third and fourth components has little effect on these patterns. These authors propose the symbols C'1, C'2, C'3 and C'4 for the active principles in mid piece, end piece, third and fourth component respectively. By carefully controlled methods, Pillemer, Ecker, Oncley and Cohn (1941) have isolated three fractions with high C'1, C'2 and C'4 activity from

guinea pig serum C'1 is a  $\gamma$  globulin containing 2.7 per cent carbohydrate. A muco globulin fraction of carbohydrate content 10.3 per cent contains C'2 and C'4. The C'2 is heat labile and the C'4 relatively heat stable. According to Pillemer, Seifter and Ecker (1941) C'4 is a carbohydrate, whose carbonyl groups are specifically attacked during inactivation by ammonia and other compounds, and which is normally carried by C'2, the two forming a globulin carbohydrate complex. C'1 accounts for 0.6 per cent, and C'2 and C'4 for 0.17 per cent of the total serum protein.

More recently Ecker and his colleagues have made a similar analysis of human complement. Four components are present with properties analogous to the four in guinea pig sera. C'1 has been isolated, and closely resembles guinea pig C'1, both have similar electrophoretic mobilities and sedimentation constants. The carbohydrate content of human C'1 was slightly higher, 3.2-3.7 per cent, and so was the apparent isoelectric point, pH 6.0-6.4 as against pH 5.2-5.4. In human serum, C'1 constituted about 0.6-0.8 per cent of the total serum protein. Of all four components in the two species, only C'3 was completely interchangeable in haemolytic tests. Human C'1 and C'4 could replace the guinea pig C'1 and C'4 and guinea pig C'2 could replace human C'2, but the corresponding converse replacements were either less effective or ineffective (Ecker, Pillemer and Seifter 1943, Seifter, Pillemer and Ecker 1943, Pillemer, Seifter, San Clemente and Ecker 1943).

#### Complement and Vitamins.

The relation of complement to vitamin C has received much attention in recent years. The reduced complement titres in guinea pig sera during the winter months (Tokunaga 1928, Marsh 1936, Horgan 1936) suggests a dietary deficiency associated with a lack of green food. In the guinea pig Harde and Thomson (1935) demonstrated an association between vitamin C intake and complement titres, Ecker, Pillemer, Wertheimer and Grodis (1938) and Chu and Chow (1938) a correlation between complement and serum vitamin C levels. The connection between the two substances may lie in the capacity of ascorbic acid to take part in redox mechanisms in the blood (Ecker, Pillemer, Martensen and Wertheimer 1938). Ascorbic acid reactivates both complement deprived of its C'3 and complement inactivated by oxidation, and increases the titre of complement from scorbutic guinea pigs. Hexoxidase, which oxidizes ascorbic acid, will reduce complement activity *in vitro*, apparently by acting on C'3.

On the other hand, Zilo (1915) and Koch and Smith (1924) found high complement titres in scorbutic animals, and, like Heinicke (1934) and Kapnick and Cope (1940) were unable to demonstrate any relation between complement and ascorbic acid levels (see also Crandon, Lund and Dill 1940, Feller *et al* 1942).

These results have been confirmed (Spink *et al* 1942, Agnew *et al* 1942) in human subjects with low serum ascorbic acid the injection of ascorbic acid did not raise the complement titre, nor was oxidation of ascorbic acid with  $\text{CuCl}_2$  accompanied by a reduction of the complement titre. In guinea pigs, variations in serum ascorbic acid induced by diet were not reflected in complement titres. Similarly Kodicek and Traub (1943) could not alter complement levels in any significant manner by variations of ascorbic acid intake in guinea pigs, they suggested that since there was a wide normal variation in complement titres, previously observed associated variation in the levels of the two substances may have been fortuitous. Moreover, the *in vitro* bactericidal power of human blood was not altered by an increase or decrease of ascorbic acid (Spink, Agnew, Nickelsen and Dahl 1942). These authors, however, noted differences between the efficacy of synthetic ascorbic acid and native vitamin C, and the discrepancies may perhaps depend on the action of substances associated with native vitamin C. The vitamin is certainly

among the reducing agents that reactivate oxidized complement and there appears to be a variable relation between the two, whose nature is as yet obscure.

The hypothetical dietary deficiency in complement poor guinea pigs associated with lack of green food may conceivably be a vitamin K deficiency, for Busing and Zuzack (1943) report a marked correlation between the complement titre and vitamin K intake in the young chick.

#### Complement in Other Animals

The greater part of the work on complement has been concerned with guinea pig serum. The quantitative and qualitative variations in complement among different species of animal have received little study. Shrigley and Irwin (1937) found no association between hæmolytic and bactericidal activity in guinea pig, sheep or rabbit sera but noted a wide species variation of both. Ecker, Pillemer and Kuehn (1942) fractionated the sera of man, dog, cat, guinea pig, monkey, rabbit and sheep and found wide species differences in the effect on opsonic powers of measures designed to inactivate or remove each of the four components. No generalization about complement titres in mammals is possible without specifying the components present and the method of measuring the complement—e.g. bactericidal or hæmolytic—and it must be remembered that in all cases the titre is determined by the amount of the least plentiful component. In human sera, for example, it is C'2 (Hegedus and Greiner 1935; Heidelberger and Mayer 1942) and to some extent C'3 (Dozou, Seifter and Ecker 1944). Mouse serum, which has no complementary activity, lacks C'2 (Brown 1943).

#### Complement Fixation.

We have noted that the absorption of complement is a general property of bacterial cells that have been sensitized by a specific antibody, though some bacteria undergo lysis as a result of this absorption while others do not. That complement fixation apart from the observation of any resulting change in the bacterial cells could be used as a general method for the detection and titration of specific antibodies was first demonstrated by Bordet and Gengou (1901). In these experiments a suspension of a given bacterium was allowed to react with a specific antiserum in the presence of complement. After time had been allowed for the reaction to take place red cells and a suitable dilution of hæmolytin were added and the mixture was incubated again for 1 hour at 37° C. It was found that in these circumstances no lysis took place, showing that no free complement was present and it was reasonably inferred that it had been absorbed by the bacterium-sensitizer complex.

The reaction may be summarized as follows

(a) Bacteria + Sensitizer + Complement = Fixation ✓

(b) (a) + Red Cells + Hæmolytin = No hæmolysis

It will be noted that (b) is simply an indicator reaction. It has no connection with the fixation of complement by the sensitized bacteria. That reaction has already occurred in (a).

Using this technique Bordet and Gengou demonstrated sensitizers for *Past. pestis*, *B. anthracis*, *Salmonella typhi*, the bacillus of swine plague and *Proteus vulgaris* in the corresponding antisera.

The following year Gengou showed that the same phenomenon occurs when soluble proteins are allowed to react with their specific antisera in the presence of complement. He demonstrated specific complement fixation, using as antigens cow's milk, egg white, horse fibrinogen and heated dog serum and as antibody in each case the serum of a rabbit which had been immunized against the corresponding protein.

It thus became clear that complement fixation was a general reaction liable to occur when any antigen was allowed to react with its specific antibody in the presence of complement.

The study of the relation between complement fixation and precipitation has yielded results of great theoretical importance. Gay (1905) showed that the precipitate formed by the interaction of an antiserum with the corresponding antigen frequently had the power of absorbing complement. This suggested that precipitation and complement fixation might be two aspects of a single reaction. Mur and Martin (1906b) found that although there was a close correlation between precipitation and complement fixation the correlation was not absolute. They showed that complement fixation might occur in the absence of precipitation and that in the presence of a constant amount of antiserum there was a particular amount of antigen that gave maximal complement fixation while amounts much greater or much less than this might fix little or no complement.

Dean (1912) elucidated the cause of these earlier discrepancies. In constant-antibody titration series he found that mixtures giving maximal complement fixation contained less antigen than mixtures giving maximal precipitation: in the one antibody was in excess in the other antigen was in excess. He showed also that complement was fixed in the early stages of precipitation before visible flocculi had formed and concluded that fixation was maximal in slowly aggregating mixtures of antigen and antibody where the relative persistence of small aggregates ensured a large total absorbing surface. The two maxima were not due to the reactions of different antibodies but were different secondary results of a single antigen-antibody reaction. Goldsworthy (1928) confirmed these results but showed that the relation of the maximal complement fixing and precipitating mixtures described by Dean were determined by the particular system he used. With slowly reacting antisera the maximal fixing mixture coincided with the optimal ratio, with rapidly reacting antisera it was in the region of antibody excess. The determining factor appeared to be the exposure to complement of a maximal absorbing surface of antigen-antibody particles for the maximum period. Later studies show that there is no constant relation between the point of maximal fixation and the established reference points of constant antibody titrations. With an ovalbumin system the point coincided with the equivalence ratio (Maltaner and Maltaner 1940) with pneumococcus carbohydrate Goodner and Horsfall (1936) found it equal to the ratio for maximal precipitation in the antigen excess region. Rice and Sickles (1942) and Rice (1943) in a more extensive study of the pneumococcal carbohydrate system record a close relation between the ratio for optimal fixation and the antibody content of the serum and note that the ratio usually falls in the region of antibody excess. Apparently the size of the antigenic particle employed appears to affect the ratio. Both Spooner and Dawden (1935) and Platt (1936) working respectively with tobacco mosaic virus and pneumococci found the ratio in the antigen excess region. They also noted that the ratio was higher with low than with high serum concentrations. According to Platt the result is due to the fact that with a particulate antigen the complement-absorbing surface is determined by the concentration of antigen and a large number of cocci sub-optimally sensitized for agglutination would absorb more complement than fewer fully sensitized cocci.

### The Role of the Components in Fixation

During the fixation of complement very little protein is removed from guinea pig serum. Haurowitz (1910) found little was added to the weight of specific precipitates formed in its presence. Heidelberger (1911) estimated that 0.25-0.40 mgm. per ml. of complement protein mainly C1 was added to specific precipitates. Similar proportional weights are added when hæmolytic antibody combines with washed red-cell stromata (Heidelberger and Treffers 1911). In the sheep red-cell hæmolytic system from 2 to 7 molecules of C1 were fixed for every molecule

of hemolytic antibody according to the initial concentrations of the various reagents (Heidelberger, Weil and Treffers 1941). The regularity of the molecular ratio in a given system and the fact that C1 is in excess of hemolysis provides quantitative confirmation of the view that the action of complement is not enzymic. However Haurowitz and Yenson (1943) in confirming the quantitative findings of Heidelberger and his colleagues estimated that while the complement required for hemolysis if spread in a unimolecular film on the surface of the red cell would cover 6 per cent. of it the amount of saponin or oleic acid required for non-specific lysis of a red cell was over 1 000 times the amount required for a complete unimolecular film. In their view the high activity of a small amount of complement suggested an enzymic nature. It should be noted that heat inactivation of complement does not destroy all its combining powers (Heidelberger 1941, Pillemer, Chu, Seifter and Ecker 1949).

By testing the complementary activity of all possible recombinations of the various fractions and mixtures of fractions of guinea pig complement Pillemer, Seifter and Ecker (1949) have shown that C1 is not the only combining component of complement. C4 and C2 are also fixed but contribute little in the absence of C1. C1 and C3 are fixed in the absence of C3 and C4 (see also Leno 1935). Little C3 is fixed but this acts in a manner similar to a catalyst to produce the complementary effect (e.g. hemolysis) on systems with which C1, C2 and C4 have combined.

With human complement, they found the fixation of C2 and C3 varied with the nature of the bacterium employed as antigen. In the absence of C1 or C4 very little fixation occurred. In the absence of C2, C4 was fixed in large amounts, and in the absence of C3 both C2 and C4 were fixed. The influence of the different components upon fixation of others was, as in the guinea pig, curiously arbitrary. As with guinea pig complement, all four components were necessary for the two specific complement effects, lysis of sensitized red cells, and death of sensitized bacteria. Neither purified C1 and C2 nor fractions containing C3 or C4 were active alone though if fixed to the cell they were active on the subsequent addition of the remaining components. Bactericidal and hemolytic power of single components and of various mixtures ran exactly in parallel, indicating a close similarity of action on the respective cell surfaces (Dozou, Seifter and Ecker 1943, 1944; Seifter, Dozou and Ecker 1944).

The nature of the fixation is still in doubt. Heidelberger, Weil and Treffers (1941) suggest that complement-combining components differ from normal globulin in being able to form loose dissociable unions with antibody molecules and that these unions are stabilized when the components are surrounded by antibody molecules in specific antigen-antibody aggregates. In cases where excess of the component combines in presence of minimal amounts of antibody a similar loose combination with antigen is postulated.

Complement is inhibited by various ant-coagulants but they appear to act by reason of their acidic or basic nature or of their large molecular size and their action does not indicate an immediate connection between coagulation and complement (Wadsworth, Maltaner and Maltaner 1937; Ecker and Pillemer 1941). For example C2 and C4 are unstable in acid solutions which at the same time depress the ionization of calcium necessary for coagulation (Pillemer and Ecker 1941a). Again, the action of lipid solvents in reducing complementary activity may be attributed to their independent action on hydrated complement for the dehydrated complement may be extracted by alcohol and ether without loss of activity on reconstitution with water (Ecker, Pillemer and Grabill 1935). The tanning agent sodium hexametaphosphate inactivates complement apparently by an alteration of the groups on which the activity depends, as a result of combination with the basic groups of the proteins concerned (Gordon and Atkin 1941).

### Opsonins and Bacteriotropins

Wright and Douglas (1903, 1904) named and described the *opsonins*—thermolabile, relatively non specific substances, occurring in normal serum, acting on a variety of bacteria and rendering them liable to phagocytosis by leucocytes. Neufeld and Rimpau (1904, 1905) named and described the *bacteriotropins*—thermostable antibodies occurring in the serum of immunized animals, acting specifically on the bacteria against which the animals had been immunized, and rendering them liable to phagocytosis.

The mode of action of the bacteriotropins is clearly analogous to that of the precipitins, agglutinins and lysins. Like these antibodies they are relatively thermostable. Like them they unite specifically with an antigen carried by the bacterial cell. We can safely assume that the anchoring of the antibody globulin to the antigen alters the condition at the cell surface in such a way as to make it easier for the leucocytes to engulf the bacteria, just as it makes the bacteria salt sensitive, and, in certain cases, renders the cell membrane more permeable. It would appear that one important factor is a lowering of the negative charge and hence of the difference in electrical potential between the bacteria and the surrounding fluid.

Falk and Matsuda (1926) found that alterations in the charge carried by pneumococci induced by the addition of lanthanum nitrate or sodium oleate had a striking effect on the phagocytosis of these organisms and Broom and Brown (1930) were able to decrease the phagocytosis of staphylococci previously sensitized with serum by preventing the usual reduction in surface charge by the addition of potassium ferrocyanide. Mudd and others (1929) studied the changes induced by specific antisera in four strains of acid fast bacilli. They found that sensitization (a) increased the cohesiveness of the bacilli, (b) decreased the electric charge as evidenced by a decrease in velocity of cataphoresis, (c) decreased the wettability of the bacilli by oil as evidenced by their distribution at an interface between tricaprui and normal saline, and (d) increased their susceptibility to phagocytosis.

Bacteria may be opsonized by non specific substances: tanning agents like gallotannic acid (Peiner and Fischer 1929, Reiner and Kopp 1929), tannic acid (Freud 1929) and alum, chrome and ferric salts (Neufeld and Ettinger Tulczynska 1929). Gordon and Thompson (1936) and Gordon and Atkin (1938) made a systematic study of the artificial opsonization of *Salmonella typhi* and *Staphylococcus aureus*. Metallic tanning agents appeared to act by combination with carboxyl groups, the vegetable tannins with the basic groups of the bacterial proteins. The two organisms differed in their susceptibility to certain agents, *Staphylococcus aureus* perhaps by virtue of having on its surface more acidic groups than *Salmonella typhi*; was more susceptible to protamine a highly basic protein. These authors noted that while potassium oxalate and distilled water reversed opsonization, opsonization by specific serum was more difficult to reverse.

There remains the problem of the relation of the normal opsonins of Wright and Douglas to the bacteriotropins of Neufeld and Rimpau. In their thermolability the opsonins resemble complement, and their identity with this serum constituent seemed at first to be rendered probable by the observation of Muir and Macleod (1906a) that such complexes as red cells and haemolysin, a protein antigen and its corresponding antibody or bacteria and a specific antibacterial serum, all removed the opsonin from a normal serum at the same time as they removed the complement. The observation of Neufeld and Hune (1907) that absorbing a normal serum with yeast had a similar double effect seemed to point in the same direction. But there were difficulties in this simple conception. The normal opsonins



do not show the same strict specificity as the bacteriotropins—normal unheated serum promotes the phagocytosis of a wide variety of antigenically unrelated bacteria—but from the first there was evidence that suggested the presence of specific factors of one kind or another. Thus it has been the general experience that the serum of any one person varies in its opsonic effect on different bacteria, that the sera of different individuals may show striking differences when tested against the same bacterium, and that any one person may show a variation in the opsonizing power of his serum for a particular bacterium, especially as the result of infection with that organism or of artificial immunization. Variations of this kind have been studied extensively by Wright and his colleagues (see Wright 1909). Moreover, evidence pointing in the same direction was obtained by *in vitro* experiments. Bulloch and Western (1906) succeeded in removing the opsonic power of normal serum for particular bacteria by selective absorption. Other workers were unable to confirm these results, but the careful studies of Hektoen (1908) on the normal opsonins acting on the red blood corpuscles of different animal species afforded strong support for the correctness of Bulloch and Western's contention.

The explanation of these anomalous findings would appear to lie in the fact that the opsonic effect of normal serum resembles its hemolytic and bacteriolytic effects in being dependent on both antibody and complement. Chapin and Cowie (1907) showed that normal serum may have its opsonic power for a staphylococcus removed by absorption with that organism in the cold, but that such absorbed serum may still have the power of reactivating normal serum that has been inactivated by heat. The cocci that have been used for absorption when washed and resuspended in saline show little if any increased susceptibility to phagocytosis in the absence of unheated serum. Later (Cowie and Chapin 1907) they showed that normal serum loses almost all its opsonic power when diluted fifteen times with saline. If, however, such diluted serum is added to another sample of serum that has been inactivated by heating at 55°–60° C for 10 minutes the mixture is almost as active in promoting phagocytosis as was the original unheated, undiluted serum. It would appear that normal serum contains specific sensitizing antibodies, present in amounts so small that they are ineffective in a dilution of 1:15 or more. Even in undiluted serum these antibodies are unable, by themselves, to alter the bacterial surface sufficiently to promote phagocytosis, but when complement is adsorbed by the incompletely sensitized bacteria the necessary change in surface conditions is produced. It would seem also that complement is not without effect on the action of the bacteriotropins, for G. Dean (1907) has shown that the action of a heated antiserum is increased by the addition of a little unheated normal serum. Sleeswijk (1908) has confirmed Dean's results, and concludes that sensitization, as a preliminary to phagocytosis, is primarily dependent on a specific antibody. When this antibody is present in adequate concentration it can produce its effect in the absence of complement, though added complement may enhance it. When the antibody is present in very small amount complement is necessary to induce adequate sensitization. (See also Ward and Enders 1933.)

It is of interest to note that the action of complement in opsonization appears to differ in some way from its action in hemolysis, since as Gordon, Whitehead and Wormald (1926b) have shown the addition of ammonia renders complement inactive as a hemolytic agent but removes none of its opsonic activity.

Gordon and Thompson (1933) inactivated complement by ammonia, Congo red, acid

alkali and hypotonic solutions and certain sodium and potassium salts. In each case by adjusting the conditions of treatment it was possible to destroy the haemolytic activity without loss of opsonic activity. They concluded that the opsonic system differs markedly from the complement system. Maltaner (1935) suggested that ammonia treated serum opsonized because the fourth component was supplied by the leucocytes employed in the opsonic mixture. Gordon (1937) confirmed Maltaner's observations but maintained that the opsonin was not closely related to complement since specifically sensitized red cells that have absorbed from ammonia treated serum all the components of complement excepting fourth component are not lysed in the presence of leucocytes—a source of fourth component. It is possible, however on the basis of these experiments to conclude that all but the fourth component is concerned in opsonization, especially since Pillemer, Seifter and Ecker's demonstration (1949) that no haemolysis takes place unless C4 is fixed previously or simultaneously with C1. On the other hand the experiments of Ecker, Pillemer and Kuehn (1942) clearly demonstrate the lack of any constant association in the sera of a number of mammals of opsonization with one or other of the four recognized components of complement.

Ecker, Weisberger and Pillemer (1942) and Ecker, Pillemer and Kuehn (1942) measured the opsonic action of large numbers of sera from various mammals on staphylococci both alone and in conjunction with staphylococcal antibody (bacteriotropin). In general mixtures of heat labile opsonin and specific bacteriotropin had less opsonic activity than either acting separately. There was no enhancement of opsonic activity except with weak concentrations of opsonin and antibody. Though there was no relation between opsonic power and haemolytic complement titre in any of the species studied yet the processes inactivating complement also reduced opsonic power. All the normal opsonins were heat-labile, those of the guinea pig, monkey and sheep were inactivated by ammonia, those of the monkey and sheep by absorption with yeast. The opsonic powers were variously distributed between mid piece and end piece fractions, mid piece predominating markedly in man and guinea pig and moderately in monkey and cat, little in sheep and rabbit. In the dog neither fraction was opsonic but a mixture of the two was.

The observation of Gordon and Thompson (1937) that protamine could opsonize bacteria led Gordon and Atkin (1939) to test globin. Globin occurring naturally in the body as haemoglobin is a distinctly basic protein, forms salts with acid proteins like casein and proved to be a good artificial opsonin. This observation not only offers a possible analogy for a natural opsonin but clearly shows that body proteins other than those known to be components of complement can act as opsonins.

The quantitative measurement of opsonic or bacteriotropic action is a technical problem of great difficulty. The system is a very complex one including living phagocytic cells the uniform distribution of which is exceedingly difficult to ensure. It is impossible, even when the tubes are rotated by one of the various mechanisms that have been devised, to obtain results of the same order of accuracy as in the precipitin agglutination or haemolytic reactions (see for example Hanks 1940).

In the method employed by Wright and his colleagues serum leucocytes and bacterial suspensions are mixed in capillary tubes and incubated at 37° C for 15–30 minutes. Their contents are then expelled on to slides. Films are prepared and stained and the bacteria contained in the first 50–100 leucocytes encountered are enumerated. The relative opsonic effect of two sera is expressed as the ratio between the numbers of bacteria taken up under their influence by the same number of leucocytes. As might be supposed the experimental error is a high one (see Greenwood 1913). An alternative method suggested by Klein (1907), is to fix on some particular degree of phagocytosis as an arbitrary end point and to dilute the serum under test until this end point is reached, but it seems probable that the experimental error will still be large.

The test has recently been revived by Huddleson and his colleagues (1933) as the opsono-cytophagic reaction and used as a measure of the immune response in brucella infections and pertussis. No attempt is made to measure a precise ratio of control and test opsonic effects, but bloods are arbitrarily graded according to the number of bacteria ingested by a standard number of leucocytes.

### The Toxin-Antitoxin Reaction

This reaction hardly falls into the same category as those we have been considering since the titrations are carried out *in vivo*. There is, however, no doubt at all that the neutralization of toxin by antitoxin depends on the same factors, and is governed by the same laws as any other antigen-antibody reaction, and we have seen that when toxin and antitoxin are mixed in the test tube specific precipitation occurs. It is convenient to discuss in this chapter some of the peculiarities recorded in the *in vivo* tests since many of these were noted during the pioneer studies of Ehrlich (1897) on the standardization of diphtheria antitoxin and so formed the basis of the controversy in regard to antigen-antibody reactions in general.

The starting point of any method of standardization is the definition of units of measurement. When these units have to be defined in terms of some *in vivo* reaction the resulting measurements are liable to errors of a kind different from those involved in *in vitro* titrations. These errors and the ways in which they can be avoided or allowed for are discussed in Chapter 43. For the moment we are concerned only with the general nature of the quantitative results that have been recorded.

The first reagent to which a unit was assigned was the toxin. This unit, the *Minimal Lethal Dose* may be defined as follows:

*The Minimal Lethal Dose (M.L.D.) of diphtheria toxin is the least amount that will, on the average, kill a guinea pig of 250 gm weight within 96 hours after subcutaneous inoculation.*

Ehrlich (1897) defined the unit of antitoxin in terms of the M.L.D. as *the smallest amount of antitoxin that will neutralize 100 M.L.D. of toxin*. This left the M.L.D. as the fundamental unit but it was soon discarded. Toxin on storage or on treatment with a variety of physical or chemical reagents, has a tendency to lose its toxicity while retaining its combining power for antitoxin. It is converted into *toxoid*. Under these conditions the definition of a unit of antitoxin, A.U., in terms of the number of M.L.D. of toxin that it will neutralize clearly becomes impossible, since no standard stable toxin can be preserved. An antitoxic serum, when dried *in vacuo* and stored at 0° C. remains stable over long periods of time, and hence provides an excellent standard of reference. Ehrlich's original antitoxin has been adopted as an international standard, and the correct definition of a unit of diphtheria antitoxin is as follows:

*One unit of Diphtheria Antitoxin (1 A.U.) is contained in that amount of antitoxic serum that has the same total combining capacity for toxin and toxoid together, as one unit of Ehrlich's original antitoxin.*

The fact that one unit of Ehrlich's original antitoxin happened to neutralize 100 M.L.D. of the particular toxic filtrate with which he was working has now only a historical interest. The units of other antitoxins are defined in a similar way some particular specimen of the antitoxic serum in question being selected as an arbitrary standard against which all subsequent samples are measured.

The actual procedure consists in first determining the quantity of a given toxic

filtrate that is neutralized, or nearly neutralized, by one unit of the standard anti-toxin, and then determining the amount of the antitoxic serum under test that will neutralize, or nearly neutralize, this amount of toxic filtrate. This amount of the antitoxic serum will contain 1 A U. Since the two tests are performed within a few days of one another there will be no significant change of toxin to toxoid during the interval, and the proportions of the two reagents in the toxic filtrates will remain constant.

This method of titration led to the definition of two other doses of toxin—"toxin" here, as in the case of the M L D, referring in practice to a toxic filtrate, containing both toxin and toxoid.

*The Limes Nul ( $L_0$ ) dose of diphtheria toxin is the largest amount of toxin that, when mixed with one unit of antitoxin and injected subcutaneously into a guinea-pig of 250 gm weight, will, on the average, give rise to no observed reaction.*

Actually, the  $L_0$  dose is usually recorded as the dose that when tested in this way, gives rise to a minimal local oedema.

*The Limes Tod ( $L_+$ ) dose of diphtheria toxin is the smallest amount of toxin that, when mixed with one unit of antitoxin and injected subcutaneously into a guinea pig of 250 gm weight, will, on the average, kill that guinea pig within ninety six hours.*

Other doses of toxin, determined by other methods of testing, have been defined in terms of their combining power for antitoxin, and are now commonly employed for standardization purposes (see Chapter 61), but the relation between the  $L_0$  and  $L_+$  doses is the matter that concerns us here.

If toxin combined with antitoxin in constant proportions giving firm chemical union it would be expected that

$$L_+ \text{ toxin} - L_0 \text{ toxin} = 1 \text{ M L D}$$

In fact it does not. The difference between the  $L_+$  and the  $L_0$  dose has been found, with different toxic filtrates, to vary from 10 M L D to 100 M L D or more.

Ehrlich attempted to account for this phenomenon, to which his name has often been attached, by postulating the existence, in toxic filtrates, of a special modification of toxoid, *epitoxoid*, having less affinity than toxin, or unmodified toxoid, for antitoxin. Over the range between  $L_0$  and  $L_+$  doses he assumed that the additional toxin added merely displaced epitoxoid, and that only when all epitoxoid had been displaced from its union with antitoxin did the added toxin remain free to exert its lethal effect. Whether toxoid differs from toxin in its affinity for antitoxin, or whether varieties of toxoid exist that differ from one another in this respect, we do not know with any certainty. Since we have many reasons other than the observed difference between the  $L_0$  and the  $L_+$  doses of toxic filtrates for discarding the hypothesis of chemical union in constant proportions between antigen and antibody, we are not faced with Ehrlich's dilemma, and have no need to postulate the existence of epitoxoid, or of any of the other special varieties of toxin and toxoid that he evolved to explain the results observed in his later studies on partial neutralization.

These studies, and those of many subsequent observers, made it clear that, when varying amounts of antitoxin are added to a constant amount of toxin, the curve of neutralization is not linear, as it would be if we were studying the neutralization of a strong acid by a strong base (See Arrhenius and Madsen 1902, 1904, Arrhenius 1915, von Krogh 1911, Glennie *et al* 1925). The observed departure from linearity is not peculiar to diphtheria toxin and antitoxin, but is characteristic of toxin antitoxin reactions in general (see, for instance, Burnet 1931), and, as we

have seen, is equally well exemplified by the results obtained when studying the absorption of any antibody by any antigen.

We have already discussed the Danysz phenomenon (p. 216). It was first observed in mixtures of diphtheria toxin and antitoxin, but a similar phenomenon has been shown to occur in other antigen-antibody reactions, and analogies have been drawn with chemical reactions of other kinds. Moriyama (1937), for instance, noted that the precipitating effect of tannic acid on proteins varies according to the way in which the reagents are mixed. He was able to elicit Danysz effects in urease-anti-urease and ricin-antiricin systems when tannin replaced the specific antibody in the systems. He attributed the effects to the capacity of the complex proteins concerned to form several quantitatively different compounds according to the proportions present at first mixing—compounds which, being incompletely reversible, do not contribute to a redistribution of two reagents when more of one reagent is added. Pappenheimer (1942), in a recent review, points out that both the discrepancy between  $L_0$  and  $L_+$  doses of diphtheria toxin and the Danysz effect can be adequately explained on the assumption that toxin (T) and antitoxin (A) form compounds varying in constitution from  $T_2A$  to  $TA_n$  (see Pappenheimer, Lundgren and Williams 1940). In the neutral mixture, the average composition of the complexes may be  $TA_2$ . The addition of an excess of T would result in complexes  $TA$  and  $T_2A$ , and only with toxin in marked excess would enough be left free to kill the guinea pig. Similarly in the Danysz effect, the first fraction of toxin would form slowly reversible complexes, like, for example,  $TA$  and its polymers, leaving insufficient antitoxin for union with the remainder of the toxin added later.

The state of toxin and antitoxin in over-neutralized, neutral and under-neutralized mixtures of the two has been studied by Eagle (1937), who precipitated toxin-horse-antitoxin mixtures with an antiserum to horse globulin. Assuming that free toxin was not carried down in the precipitate, Eagle was able to show that antitoxin unites with excess of toxin to form a *toxic* complex, and toxin with an excess of antitoxin to form a complex with *antitoxic* properties. Only in one proportion was a precisely neutral complex found. Here again we have recorded analogous findings in the reactions of other antigens with other antibodies. This phenomenon is more readily explicable on the adsorption hypothesis than on the theories advanced by Ehrlich, or by Arrhenius and Madsen.

#### Other Manifestations of the Antigen-Antibody Reaction.

The addition of antigens to antibody even in optimal proportions is not always followed by a visible precipitation. Either of the two reagents may be too weakly reactive or too dilute. The classical method of detecting the union of antigen and antibody in these circumstances is by the complement fixation reaction (see p. 232), but the reaction may be made manifest by other less elaborate means. The high sensitivity of the various flocculation tests for syphilis (see Chapter 81) is due to the addition of agents such as lipins which shift the precipitating system more to the hydrophobe state. Again dyes may act both as sensitizers and as indicators in precipitation reactions. For instance Dean (1937) in a study of non-specific precipitation of proteins by the electropositive dye isamine blue, found that appropriate dilutions of the dye added to constant-antibody titration series markedly increased the sensitivity of the titration. Berger (1943) made use of Janus green and Victoria blue for the same purpose in the flocculation test for syphilis. Cannon and Marshall (1940), taking advantage of the fact that the amount of antibody required for the surface sensitization of large particles is smaller than that required for the surface sensitization of the same bulk of material in a more finely divided

state (see Zinsser 1930), devised a method of detecting small non precipitating amounts of antibody by titrating them against suspensions of colloidal particles coated with the appropriate antigen (see also Lowell 1913)

The fine turbidity which precedes flocculation in strongly reacting antigen-antibody systems and which may be the end result in weakly reacting systems can be measured by photoelectric and other types of nephelometer, and the measures made the basis of titrations of antigen or antibody (see, for example Lobb 1938 Pope and Healey 1939 Bukantz, Cooper and Bullock 1941) Advantage may also be taken of physicochemical properties of antigen-antibody compounds other than precipitability in the presence of electrolyte Thus, du Nouy and Hamon (1935, 1936) found a well marked maximum of viscosity to occur in mixtures containing optimal proportions of diphtheria toxin and antitoxin

After initial sensitization, which is relatively rapid the agglutination of bacteria depends on their rate of collision under the influence of Brownian movement and convection currents in the suspending fluid If the cells are brought together rapidly by centrifugation, agglutination may be rapidly detected by the resistance to resuspension of the centrifuged mass as compared with that of a control preparation of bacteria without antibody

An interesting consequence of an antigen-antibody reaction is the swelling of pneumococcal capsules in the presence of specific anti capsular rabbit serum first described by Neufeld 1902, Neufeld and Etinger-Tulezyska 1931 (see Chapter 24) The swelling reaction can be elicited in both living capsulated pneumococci and in decapsulated organisms (Brown 1938) The swelling may be reversed if the antibody is destroyed by heat (Etinger-Tulezyska 1933) or by digestion of serum proteins with papain (Kalmanson and Bronfenbrenner 1942) It is reversed if the antibody is removed by repeated washing or by adding an excess of specific polysaccharide to a suspension of washed swollen cells (Hemmelt and Nungester 1942)

According to Hershey (1940) the capsule does not appear to present a smooth surface to the action of antibody, for the amount absorbed is greater than such a surface could accommodate as a closely packed unimolecular layer, it is possible that the capsular polysaccharide is in the form of a loose fibrous gel whose surface and interstices provide ample surface for the adsorption of large amounts of antibody The electron micrographic studies of Mudd and his colleagues support this view As we have already noted homologous antibody is deposited on *Salmonella typhi* in an electron opaque layer about as thick as a unimolecular film of globulin (Mudd and Anderson 1941) Under the influence of rabbit antibody, the pneumococcal capsule which is relatively transparent to the electron beam and appears to be a low density gel becomes opaque, and increases in thickness The increase in thickness is more than 25 times that of a unimolecular layer of antibody, even assuming that the globulin is packed with its maximum length perpendicular to the surface of the capsule (Mudd, Hemmelt and Anderson 1943) There is clearly some adsorption of non-specific material for the opacity of swollen capsule is increased in the presence of non-specific serum proteins It seems probable that the swelling is not purely an antigen-antibody effect Johnson and Dennison (1944) measured the volume increase during swelling by two independent methods it varied from 2.9 to 9.5  $\mu^3$  by one method, and from 7.4 to 10.1  $\mu^3$  by the other They concluded that the volume increase could not be accounted for solely by the molecular volume of antibody absorbed, but might be due to a hydration of the capsule following the antigen-antibody reaction

We may refer to the 'adhesion phenomenon' in which spirochaetes specifically sensitized by antibody adhere readily to bacterial cells added to the reacting mixture Advantage is taken of this behaviour in the serological study of spirochaetes (p. 912)

## THE QUALITATIVE ASPECTS OF THE ANTIGEN-ANTIBODY REACTIONS—IMMUNOLOGICAL SPECIFICITY

This problem has been attacked mainly from the side of the antigens but it will facilitate our discussion of the recorded data to deal first with the probable structure of the serum antibodies with which they react

### The Nature and Properties of the Serum Antibodies

We have already made frequent references to the view that the serum antibodies are specialized globulins. The properties of normal serum globulins have been established by a variety of methods. The fractionation of serum by precipitants like ammonium sulphate yields a mixture of proteins differing from one another physically and chemically in lipid and carbohydrate content (see Tiselius 1937a, Stenhagen 1937, Hewitt 1938). It is difficult to decide how far these fractions are the results of the chemical handling the native protein receives in its preparation. The measurement of the rate of migration of serum proteins in an electrical field (Tiselius 1937a) provides convincing evidence that native serum contains several well-defined components. After a given period of exposure in the Tiselius electrophoretic apparatus the proteins of native serum separate into a number of zones each containing particles having approximately the same electrophoretic mobility. In normal horse and rabbit sera the albumin forms a single well-defined zone and there are three components of globulin  $\alpha$ ,  $\beta$  and  $\gamma$  the  $\alpha$  being the most and the  $\gamma$  the least mobile.

The current hypotheses of protein structure are none of them sufficiently developed to warrant description in a book of this kind. The student may consult the reviews of Pauling and Niemann (1939), Svedberg (1939) and Synge (1943) for details of the controversies regarding the precise architecture of the proteins. It is sufficient for our purpose to note those features of protein chemistry that are relevant to the understanding of serology in its present state. In the first place it is generally accepted that the proteins are built up of polypeptide chains of amino-acids and that the pattern of recurrences whether regular or irregular of the various amino-acids in the chains will vary from protein to protein according to the relative proportion of constituent amino-acids. A secondary structure is imposed on the protein molecule by the folding of the chains so that several of them are held together by hydrogen bonding and other interatomic electrical forces. In the so-called fibrous proteins, the arrangement of chains as revealed by X-ray analysis is markedly regular (see Astbury 1943). Keratin and silk fibroin are examples of this type of protein. The globulins belong to the class of non-fibrous proteins, and here it is supposed that polypeptide chains are primarily folded into a lamina that has approximately the thickness of the polypeptide chain and that these laminae are built up into a more or less globular protein molecule being held together by interatomic forces similar to those that hold the chains in laminae. Whether the laminar structure exists or not it is a fact that certain proteins can be spread on an air-water interface into films whose thickness is much less than the known diameter of the protein molecule. Some proteins are denatured by this process but others including serum proteins will re-form into globular molecules after spreading.

That is to say, although protein molecules compared with smaller molecules

are relatively rigid structures, by reason of the large number of interatomic forces which, acting together, stabilize the geometric form of the molecule, they are also capable of some distortion, which may or may not be reversible. In much of the earlier work on protein molecules, the assumption was made that protein molecules behaved as spheres, i.e. were isodimensional. It is now evident that some molecules, including the globulins, are markedly anisodimensional, behaving as though they were rod shaped, or at any rate larger in one dimension than in any other. The term "globular," applied to the non fibrous proteins, does not imply that they necessarily behave as spherical or nearly spherical units. For example, in the serum reactions the shape of the molecule, and the degree to which it may be changed, will affect the range of permissible speculation about the structure of the antigen antibody complexes.

The molecular weight of proteins may be determined from their sedimentation rate in the intense gravitational fields of high speed ("ultra") centrifuges. Those of non fibrous proteins range from 17,500 for lactalbumin to over 6 millions for mollusc haemocyanins. The molecular weight of the normal serum globulins and the majority of antibody globulins in the horse and rabbit are of the order of 150,000 (see Svedberg 1937, Heidelberger and Pedersen 1937). Neurath (1939) estimated that the physicochemical properties of rabbit antibody globulin were consistent with those of an ellipsoid, 27  $\mu\mu$  in length, and a maximum width of 4  $\mu\mu$ . Svedberg has pointed out that the molecular weights of the non fibrous proteins cluster round certain values which are whole-number multiples of the lowest weight recorded, 17,500, and suggests that they are built up from units having a molecular weight of approximately 35,000. This relationship of the molecular weights of proteins, whether animal or vegetable in origin, presumably reflects a similarity of the enzyme systems that take part in protein synthesis. On this view, the rabbit antibody globulin molecule would correspond to four Svedberg "units." As we shall see, this possibility has been raised in connection with the valency of antibody by Hooker and Boyd who argue that if the four units each carried an antibody receptor, and were arranged linearly, the whole molecule would be multivalent, and markedly anisodimensional (p. 251).

### The Purification of Antibody.

Antibodies are precipitated from serum by agents that bring down the serum globulin. Precipitation of antisera with sodium or ammonium sulphate with alcohol at various temperatures or by dialysis, brings down the greater part of the contained antibody with the globulin fractions. (See Banzhaf and Gibson 1907, Gibson and Collins 1907, Ledingham 1907, Mellanby 1908, Hartley 1914, Felton 1926, 1923, 1933, Maxfield and Burbury 1927, Barr, Glenny and Pope 1931, Barr and Glenny 1931, Laidlaw and Dunkin 1931, Barr 1932 and many others.)

A considerable degree of purification of antibody solutions is possible where the antibody is confined to one particular fraction. By fractionation Felton (1926, 1933) purified the antibody to the Type I pneumococcus and Laidlaw and Dunkin (1931) the antibody to distemper virus.

Specific precipitation of serum globulins by antisera prepared in an animal of another species also brings down the antibodies (Landsteiner and Prásek 1911, Eisler 1920, Smith and Marrack 1930). Moreover, the inactivation of antibodies by heat corresponds in general with the denaturation of proteins by heat (see Streng 1909, Madsen and Streng 1910, Marrack 1938). Many of the earlier studies of



this phenomenon are complicated by the fact that the heat inactivation was carried out in the presence of other serum proteins and as the work of Kleczkowski (1911b) van der Scheer Wyckoff and Clarke (1941a) Bawden and Kleczkowski (1942b) Jennings and Smith (1942) and Krejci Jennings and Smith (1947) shows heating mixed solutions of antibody globulin and proteins like albumin or casein in the presence of salts (Kleczkowski 1943) produces globulin albumin and globulin casein complexes which display no direct antibody activity but which cannot be considered as denatured in the ordinary sense of the word.

The study of the antigen antibody complex offers a more direct approach to the nature of antibody since protein united with antigen has presumably been selected from the antiserum by reason of its specific combining powers. A certain amount of non specific material may be adsorbed by the complex—such adsorption occurs in complement fixation—but the amounts are relatively too small to invalidate the assumption that the bulk is in fact antibody. For instance Heidelberg and Landsterner (1923) Marrack and Smith (1931b) and Haurowitz and Breinl (1933) have shown that non specific coloured proteins are not carried down in the precipitate when an antigen reacts with its specific antibody and Dean Taylor and Adair (1935) using the optimal proportions technique and examining a sample of antiserum containing antibodies to purified egg albumin and purified horse serum albumin found that either antibody could be specifically precipitated by the corresponding antigen without affecting the titre of the other. It has been established that antigen antibody complexes contain considerable quantities of protein from the antiserum and that this protein has the general characters of serum globulin. Thus the precipitate formed when 2.0 mgm. of Type II pneumococcal polysaccharide which contains no nitrogen and is therefore particularly suitable for work of this kind reacts with its homologous antiserum has been found to contain 37 mgm. of serum protein (Felton and Bailey 1926) and the antibody titre of a given antiserum may be measured with considerable accuracy by determining the amount of protein that is removed by a soluble or particulate antigen (Heidelberg and Kendall 1929 Heidelberg *et al* 1930 Heidelberg and Kabat 1934).

That the protein so bound has the general properties of globulin is attested by such observations as those of Marrack and Smith (1930 1931a) who found that the diphtheria toxin antitoxin compound showed the same ultra violet absorption spectrum as serum globulin or those of Breinl and Haurowitz (1930) who found that a precipitate containing about 10 per cent. of hæmoglobin and 90 per cent. of material derived from a homologous antiserum showed the same proportions of tyrosine cystine histidine and arginine as did serum globulin when examined by the same technique.

Antigen antibody complexes act as antigens and induce in animals anti antibodies that are specific for antibody in that they combine with the antigen antibody complex but not with antigen alone. The antisera will also react with normal serum globulins (Ando 1937 see also Marrack and Duff 1938). Another noteworthy feature of such antisera to which we shall refer later is their capacity to react with specific precipitates of other antigens provided that the antibody in these precipitates was produced in the same species of animal (Treffers and Heidelberg 1941). However the properties of antibody protein as revealed in the antigen-antibody complex are open to the objection that the antibody may have been altered by its union with antigen.

Many attempts have been made to dissociate antibodies from an antigen antibody complex in a protein free condition. The dissociation of the antigen antibody compound may be accomplished in a variety of ways, and certain of the antibody solutions so obtained have contained very little protein (see Landsteiner and Jagic 1903, Muir 1903, Bail and Tsuda 1909, Spät 1910, Kosaki 1918, Huntoon and Ffris 1921, Huntoon *et al* 1921, Locke and Hirsch 1925), but in none of these instances has it been satisfactorily demonstrated that the amount of protein present was insufficient to account for the antibody action observed (see Eagle 1930). Similar attempts have been made by adsorption on to various reagents other than the specific antigen, followed by elution with various fluids at different pH levels. Some of these attempts have been claimed as successful (see Frankel and Olitzki 1930, Olitzki and Frankel 1931, Frankel 1932, Olitzki 1932), but these claims have not been confirmed by subsequent workers (see Narrack 1931, Rosenheim 1935).

Northrop (1911) and Rothen (1911) prepared purified diphtheria antitoxin from toxin antitoxin precipitates by digestion with trypsin. The purified antitoxin had a molecular weight in the region of 90,000. By fractional precipitation Northrop isolated a crystalline antitoxic protein which appeared to be antigenically distinct from normal horse proteins.

Later studies of the effect of electrolytes on specific aggregation (see p. 215) led to the preparation of dissociated antibody solutions of a high degree of purity (Heidelberger and Kendall 1936, Heidelberger, Grabar and Treffers 1938). Heidelberger and his associates were able to purify both rabbit and horse antisera by salt dissociation of agglutinated pneumococci, Types I, II and III. Dissociated and native antibody behaved alike in serological reactions. Dissociated antibody had the properties of a globulin. The molecular weight of dissociated rabbit antibody, calculated from its sedimentation rate in the ultracentrifuge, was like that of antibody in native antiserum and of globulin in normal serum, in the region of 150,000 to 195,000.

#### Varieties of Antibody Globulin

Horse antibody to the Type III pneumococcus has a molecular weight over 4 times that of the corresponding rabbit antibody. A similar component was found in other preparations of horse antisera, but not in normal horse serum (Heidelberger and Pedersen 1937). Later, Kabat (1939) showed that these heavier antipneumococcal globulins, of molecular weight 910,000 to 930,000 were formed also in the cow and pig.

These antibodies of high molecular weight differ in certain respects. For instance 'heavy' antibody to pneumococcal polysaccharide will not fix guinea pig or human complement in the presence of homologous antibody, whereas ox antibody of similar weight does so (Heidelberger and Treffers 1941). Heavy antibodies do not appear to be quite so firmly constituted as antibodies of a lower molecular weight for they are disaggregated by relatively mild treatment with barium hydroxide without much reduction of precipitating power. Not all horse antibody has this high molecular weight: for example, diphtheria, scarlet fever, tetanus and *Cl. welchii* antitoxins from the horse when diluted, contain no abnormally heavy protein molecules (Fell Stern and Coghill 1940).

The significance of the large antibody molecules is not clear, but it is evident that response to immunization on the part of certain animals is more than the production of normal globulins modified only in configuration, a new type of globulin may in fact be formed.

The serum globulins as a whole behave in the ultracentrifuge as though they were homogeneous but they are separable as we have already seen into fractions of differing solubilities. Antibodies are distributed variously among the fractions (see for example Adair and Taylor 1936). The distribution depends on the animal immunized, the nature of the antigen and the mode of fractionation.

It is still questionable whether the euglobulins and pseudoglobulins are artefacts produced during fractionation of serum. Rabbit antisera to horse and human serum proteins react to some extent separately with euglobulins and pseudoglobulins suggesting that the two exist as such in the native serum (Harris and Eagle 1935).

Marrack and Duff (1938) were unable to demonstrate an absolute serological distinction between euglobulin and pseudoglobulin which neither singly nor in mixture had the full serological reactivity of native globulin. Treffers and Heidelberger (1941a) also found only a partial serological distinction between the water-soluble and water-insoluble globulins.

The increase in serum globulin during immunization has been noted by several observers (see Marrack 1938) though not all of it is attributable to the formation of antibodies (Liu, Chow and Lee 193; Boyd and Bernard 1937). In Boyd and Bernard's experiments with a variety of antigens the antibody never accounted for more than 35 per cent of the increase though as the authors point out some of the serologically inert proteins may have been antibodies of a reactivity too weak to be detected by their methods.

Ando and his colleagues (Ando, Kee and Komiyama 1937; Ando, Kee and Manako 1937; Ando, Manako, Kee and Takeda 1937; Ando, Manako and Takeda 1938; Ando, Takeda and Hamano 1938) by immunizing rabbits with antigen-antibody aggregates, distinguished two types of globulin A and B in horse sera. A predominating in the water-soluble B in the water-insoluble fractions. Both normal sera and antisera contained A and B globulins. Immunization with diphtheria toxin or Shiga's dysentery bacilli, increased the A globulins. B globulins were increased during immunization with pneumococci, *Salmonella typhi* and other bacterial antigens.

The globulin components revealed by Tiselius's electrophoretic apparatus undoubtedly exist as such in native serum. Protein particles of similar molecular weight may prove to be heterogeneous by electrophoresis, and Tiselius (1937b) showed that antibody preparations of Heidelberger and Pedersen (1937) which resembled normal globulins in the ultracentrifuge differed strikingly from the normal in their rate of migration in an electrical field. By comparing antibody to egg albumin and to pneumococci before and after absorption with the homologous antigens, Tiselius and Kabat (1939) demonstrated that in the rabbit and the monkey antibody was electrophoretically identical with the  $\gamma$  component of normal globulin. Horse antibody migrated as a new component, midway in mobility between the  $\beta$  and the  $\gamma$  components. In the hands of Moore, van der Scheer and Wyckoff (1940) and van der Scheer, Lagedin and Wyckoff (1941) the high molecular weight antibody in horse antipneumococcal sera migrated with the normal  $\gamma$  globulin. Van der Scheer, Wyckoff and Clark (1940, 1941b) described in some antisera an increase in  $\gamma$  globulin, in others a new component T both associated with an increase in antibody. The T component was midway in mobility between the  $\beta$  and  $\gamma$  components (see also Pöthen 1941). It appears more readily in antitoxic as distinct from antibacterial sera, though not regularly so. Kekwick and Record (1941) found two antibody fractions in diphtheria antitoxin from the horse: one associated with the  $\beta$ , the other with the  $\gamma$  component. With optimal amounts of toxin, the  $\beta$  fraction flocculated slowly and formed a compound with the composition  $(TA)_2$ . The  $\gamma$  fraction flocculated 20 times as rapidly and formed compounds of the type  $(TA)_4$ . The  $\gamma$  fraction, which is precipitated with the euglobulin during salt fractionation, predominated in early bleedings from immunized horses; in the later bleedings, the  $\beta$  fraction constituted the main antitoxin content (Kekwick, Knight, MacFarlane and Record 1941).

We may note one more variety of antibody recently described by Race (1944). The

erythrocytes of certain human subjects contain an antigen D, which is one of the natural Rh antigens (Chapter 49). An antibody to D occurs naturally in the sera of other human subjects and, like other natural antibodies to the blood group antigens, it agglutinates erythrocytes containing the corresponding antigen. In certain subjects, however, Race found "incomplete" antibodies to D, which combined specifically with D containing erythrocytes, but failed to agglutinate them. The properties of this naturally occurring "incomplete" antibody in many respects resemble the reacting but non flocculating albumin antibody globulin complexes made artificially by Bawden and Hrczowski (1942) (see pp. 213, 214).

**The Modification of Antibody by Physical and Chemical Means.**—Although there is a general parallelism between gross alterations of the globulin nature of antibodies, and their activity with respect to antigens, yet the protein can in many cases be altered considerably without destroying its efficacy as an antibody.

If for instance, Type II pneumococcal antibody is "unfolded" in a mono layer on an air water interface, it loses its capacity to combine with the specific carbohydrate of the Type II pneumococcus (Danielli, Danielli and Marrack 1938).

A similar unfolding of normal proteins appears to take place in urea solutions (see Bernheim *et al.* 1942), possibly by the weakening of the hydrogen and other bonds responsible for maintaining the association of adjacent polypeptide chains. In urea solutions at pH 7.8 diphtheria antitoxin slowly loses its neutralizing power for toxin, and more rapidly at a higher or a lower pH (Wright 1944). These results suggest that the antibody activity is closely associated with integrity of the general molecular structure and that destruction of the antibody configuration is associated with ionization of basic or acidic groups.

Chemical modification by moderate treatment with formaldehyde, which reacts irreversibly with the protein at the surface of the molecule, has little effect on antibody activity (see Mudd and Joffe 1933, Ivanoff 1936, and Eagle 1938). As Eagle showed stronger treatment with formaldehyde, or with a reactive diazo compound, removed first the precipitating and then the protective power of antipneumococcal sera and diphtheria antitoxins, a fact we have already noted in discussion of the lattice hypothesis. Again, the amino groups of an antibody globulin molecule may be treated with ketene gas until one third of them are acetylated, without marked loss of the power to combine with antigen (Goldie and Sandor 1937). The chemical groups on the surface of globulin molecule responsible for the antigenic power of the protein do not appear to form part of the specific combining groups of the antibody. The destruction of activity of horse antipneumococcal sera by photodynamic oxidation in the presence of methylene blue only reduces the power of the proteins to precipitate with anti horse serum (Ross 1935). The independence in the antibody molecule of the antibody-combining group and the groups that determine antigenic specificity has been demonstrated serologically by several workers. Smith and Marrack (1930), and later Eagle (1936) demonstrated that diphtheria antitoxin from the horse, which has combined with a rabbit anti horse precipitin would react with toxin. Eagle in addition showed that antitoxin combined with toxin could still react with the rabbit antibody. Treffers and Heidelberger (1941a, b) immunized animals with specific precipitates formed by the combination of various antigens with their homologous antibodies, both horse and rabbit. The reaction of the resulting anti antibodies was conditioned solely by the species of animal from which the antibody was obtained, any difference between various antibodies that might be expected from the fact that they each possessed different combining groups for their homologous antigens, was not demonstrable by serological methods.

**The Enzymic Digestion of Antibodies.**—Hydrolysis by enzymes ultimately destroys the antibody activity of serum globulin (see Marrack 1938). Partial hydrolysis under controlled conditions is selective in its action on the various serum proteins. For instance, Pope (1939a, b) investigating the Parfentjev patent for the refinement of antitoxin, con-

cluded that by heat treatment in an acid solution, diphtheria antitoxin globulin was partially denatured and could be refined by proteolytic enzyme hydrolysis leaving an undenatured portion which carried most of the antitoxic activity. Similarly Posenheim (1942) demonstrated that diphtheria antitoxin was split into approximate halves of different solubilities by papain digestion, one half carrying the antitoxic activity the other being inert. If digestion was prolonged so that the halves split into quarters, no antitoxic activity remained. The process appeared to be one of enzyme cleavage and not a denaturation. The antibody loses most of its original antigenic activity as a result of its "proteolytic refinement" (Weil, Parfenyev and Bowman 1935). The change is reflected in the distribution of electrophoretic components. Refined antisera show a diminution or absence of albumin, disappearance of the T component in diphtheria antitoxin and its replacement by a new  $\gamma$  component (van der Schuer, Wiskoff and Clarke 1941c), or a relative increase in the  $\mu$  and  $\alpha$  components (Fell, Stern and Cochill 1941).

From this fact, and the varying distribution of antibody among the components, it is clear that the characterization of an antibody globulin by its electrophoretic mobility has no necessary connection with the fact that it carries immunologically active groups.

An interesting example of partial digestion of antibody is recorded by Kalmanson and Paffenheimer (1947). A bacteriophage just neutralized by homologous rabbit antibody was restored to activity by papain digestion of the globulin on its surface. The digestion of over neutralized bacteriophage, on which there were many more antibody molecules, did not reverse the neutralization. Some part of the antibody apparently remained on the phage particles after digestion, for just neutralized phage after digestion would sensitize guinea pigs to rabbit globulin, and over neutralized phage after digestion was found to be incapable of stimulating phage antibody on injection into a rabbit, presumably because the antibody remains formed an obscuring layer on its surface.

Posenheim (1937) has drawn attention to an increase in the resistance of antibodies to pepsin and trypsin hydrolysis that takes place as immunization proceeds. She found that "H" bacterial agglutinins (see p. 276) from late bleedings of an immunized rabbit were resistant to a degree of digestion that destroyed 80 per cent. or more of agglutinins from the first bleeding. There did not, however, appear to be a corresponding increase in the resistance of the serum proteins to digestion. "O" antibodies did not become resistant in this way. Both Paffenheimer and Polson (1937) and Pope (1939a) noted that the resistance of diphtheria antitoxin to proteolytic refinement varied with the stage and the method of immunization of the horse.

### Unity and Diversity of Antibodies.

Immunization with a single antigen yields an antiserum which can display a variety of specific phenomena for instance, precipitation, neutralization of toxins, protection of an infected animal. In the interpretation of antibody reactions there are two questions about the variety of antibodies that need an answer. In the first place are all these manifestations due to a single kind of antibody or does a precipitin for example, differ fundamentally from an antitoxin, when the toxin series are unknown in both reactions? In the second place in a solution that displays a given kind of serological activity are all the molecules of antibody similar? In other words in a monospecific antibody solution are there antibodies with different functions and are those of the same function homogeneous?

The view that precipitins of various antitoxins, etc. differed in kind held the field for a long time, but the unitarian hypothesis that these were due to the same kind of antibody in different circumstances has now gained almost universal acceptance. This has been due mainly to the work of Dean (1917) in this country and of Luzzati (1921) in America and to Nicolle and Cesar (1922) in France. The unitarian view had been admitted by Bail and Hoke (1908). We have seen how it was established the essential nature of precipitation and complement

fixation. We have noted that precipitation is a general form of reaction between all antigens and all antibodies, including toxin and antitoxin. And we have seen that the union of antigen and antibody at the surface of a bacterial cell sensitizes it alike to the flocculating action of electrolytes, to the lytic action of complement, and to the phagocytic action of leucocytes.

There are too in records of the assay of the protective power of antisera, many instances of complete parallelism between the protective power and some *in vitro* agglutination, antihæmolytic or antitoxic reaction. For instance, the type specific precipitin content of antimeningococcal sera was found to parallel their protective power in mice (Pittman 1943).

The detailed evidence has been reviewed by Marrack (1938). It remains to note that Delves (1937) found that precipitating antibodies to pure human albumin and pseudoglobulin would agglutinate and opsonize colloidal particles coated with homologous antigen, that Gerlough, Palmer and Blumenthal (1941) were able to establish the identity of precipitins, agglutinins and protective antibodies in antisera to six different types of pneumococcus, and finally, by their method of dissociation of agglutinated bacteria, Heidelberger and Kabat (1936) and Alexander and Heidelberger (1940) were able to demonstrate directly the identity of the agglutinin and precipitin to Type I pneumococcus and to *Haemophilus influenzae* Type b.

This conception of the serum reactions does not, of course, in any way modify our belief in a multiplicity of antibodies corresponding to a multiplicity of antigens. A red cell, a bacillus or a crude protein solution such as horse serum contain many antigens and give rise to many antibodies. The unitarian hypothesis as Zinsser (1921) has emphasized, implies simply that the injection into the tissues of a chemically pure antigen will lead to the formation of one antibody capable of producing all the various manifestations of antigen antibody union.

When we discuss, in the next chapter, the antigenic structure of bacteria we shall see that there is a sense in which it would be correct to differentiate one antibody from another in terms of function—to say, for instance, that a particular antibody is an agglutinin but not a lysin. This difference in functional activity however, is determined not by a difference in the nature of the antibody but by a difference in the structural position of the antigen to which it is attached.

**The Homogeneity of Monospecific Antibodies**—It is often assumed that the behaviour of antibody in solution, provided that it has been formed in response to a single antigen, is the sum of the effects of a homogeneous collection of antibody globulin molecules. We have already discussed a number of phenomena that indicate a heterogeneity of the antibody in a given serum. Thus in monospecific sera there is an antibody that precipitates with antigen only in the presence of fully reacting antibody (Marrack and Smith 1931b, Heidelberger and Kendall 1935a, b, c, Heidelberger, Treffers and Mayer 1940). Heidelberger postulated univalence for this low grade antibody, and multivalence for the more avidly reacting antibody. Landsteiner and van der Scheer (1940) found in an anti hen ovalbumin serum two kinds of antibody both precipitating with hen ovalbumin, turkey albumin precipitated readily with one, and failed to precipitate with the second, though there was evidence of combination. Other evidence of heterogeneity of antibody is provided by the change in quality of antibody during the course of immunization. The broadening of reactivity and the increase in avidity of antiserum with prolonged immunization has been observed many times. Hooker

and Boyd (1941a) suggest that they may be due to (a) the formation of antibodies to minor antigenic determinants (b) the increase in the number of reactive groupings on the antibody or (c) the wider affinity of the combining groups due to the formation of a larger and more complex reactive patch on the antibody surface. The broadening may be accompanied by a change in the nature of the globulins. Raffal Pait and Terry (1940) found the earlier, less avid antibody to be associated with the water insoluble globulins the later, avid antibody with water soluble globulins. The evidence of heterogeneity found among fractions of antiserum made by precipitation with salts is less convincing because it is difficult to exclude artefacts but the differences noted above in reactivity displayed by electrophoretic fractions of antisera cannot be lightly dismissed.

Goodner and Horsfall (1937) obtained other evidence of heterogeneity in antipneumococcal sera. In several rabbit antisera the ratio of precipitin content to protective power was constant, but in horse antisera it was inconstant. Moreover in individual sera of both species the protective power of antibody left after some of it had been precipitated by antigen varied with the amount of antigen added, though the variation was much greater in the horse. When horse antiserum was fractionated the pseudoglobulin was found to contain antibody with a protective power only one-seventh that in the euglobulin fraction though the pseudoglobulin antibody was far more reactive as precipitin. In an electrophoretic fractionation of diphtheria antitoxin, the antibody  $\gamma$  component combined with twice as much nitrogen per flocculating unit as that from the  $\beta$  component. Goodner, Horsfall and Bauer (1938) also demonstrated heterogeneity in the size of antibody particles in both rabbit and horse antisera. For example, in horse antisera very large particles, corresponding to a pore size of 170  $m\mu$  in a collodion membrane filter were found in concentrated serum and of 83  $m\mu$  in native serum. Dilution in saline reduced the particle size in both types of serum.

The antibodies formed in response to the injection of what is apparently a single antigen may also display varying kinds of serological specificity. Thus, Landsteiner and van der Scheer (1939) working with proteins modified by coupling to pentapeptides (made up of various combinations of glycine and leucine) showed that all the antibodies produced in response to any one peptide-protein were absorbed by the same peptide coupled to an antigenically unrelated vehicle namely erythrocyte stromata. Nevertheless varying amounts of the total antibody could be removed from the sera prepared against one peptide by absorption with stromata to which one of the other peptides had been coupled. Absorption occurred even when the absorbing peptide could not conceivably have been a breakdown product of the antibody producing peptide.

It is clear that in horse sera at any rate, apparently mono-specific antibodies display heterogeneity both with regard to their reactivity as antibodies, and to their other physicochemical properties. Monospecific rabbit antibodies appear to be less heterogeneous at least functionally. Indeed the degree of homogeneity may be surprisingly high. Thus, Hershey, Kalmanson and Bronfenbrenner (1943) record an analysis of a number of rabbit anti-phage sera with a 25-fold range of combining power in which there was full correlation between three varying qualities—specific rate of phage neutralization neutralizing capacity, and precipitating capacity—and no tests revealed any heterogeneity of antibody in any one serum.

### The Valency of Antibody

It is obvious that the conception of mono-specific antibody molecules of different valencies will help to explain some of the observed heterogeneity of antibody in the natural state. As we have seen, the assumption of multivalent antibody is

an essential part of the lattice hypothesis, and on p. 220 we have indicated a number of objections to the lattice hypothesis which imply the rejection of a multivalent antibody. Nevertheless variations in "avidity" and precipitating power of antibody obtained during a prolonged course of immunization are conveniently explained in terms of variations in valency. But, as we should expect from our ignorance of globulin metabolism in the animal body, there are no impressive data to support the speculation. The most cogent objection to the theory of multivalence (see Hooker and Boyd 1912) lies in the fact that immunization with a known mixture of distinct antigens does not result in polyspecific antibody (see, for example, Hektoen and Boor 1931, Dean, Taylor and Adair 1935). It is argued that, if during the synthesis of antibody in the animal there was in the growing antibody globulin molecule more than one region that could be influenced by antigen, there is no good reason why two or more regions on one molecule should not be influenced by separate antigens, and thus give rise at least to a dispecific antibody.

The conception of dispecific antibody offers a simple explanation of the results of antibody response to certain natural complex antigens (Meyer 1936, Miles 1933) but since the antigenic constituents of the mixtures were not fully defined the success of the hypothesis is not good evidence of its validity. The difficulty may in part be resolved by assuming that monovalent Svedberg units of molecular weight 35 000 are in fact formed in the body, and that these are later united into larger molecules which are multivalent. But here again unless we postulate that the units are always built on to one another in conjunction with a single antigen particle there is no good reason for assuming that Svedberg units of diverse specificity should not be joined to form a polyspecific antibody.

Molecules of antigen having on the surface two antigenic determinants may give rise to apparently dispecific antibodies. For example Heidelberg and Kendall (1934) immunizing rabbits with ovalbumin modified by an azo-dye produced antibodies to ovalbumin alone and to azo protein alone and also antibody that reacted equally well with azo protein or ovalbumin (see also Singer 1942). Landsteiner and van der Scheer (1938) on the other hand could find no evidence of a double specificity even in antibodies prepared against a single azo antigen having two haptene groups. They prepared antigens with aminosuccinylacetic acid (S) with aminophenylarsenic acid (A) and with a compound of the two (SA). Antisera to SA precipitated with SA, A and S but sera absorbed with A precipitated only with S and sera absorbed with S only with A. The various antisera contained one or two kinds of specific antibody, but never dispecific antibodies. Haurowitz and Schwerin (1943) obtained analogous results by immunizing rabbits with a globulin antigen containing both *p*-azo phenylarsenic (A) and *m*-phenylsulphonic groups (S) or with a globulin containing both A groups and diiodotyrosine (T) groups. The resulting antisera contained antibodies against A and S and against A and T respectively but there was no evidence of dispecific antibodies against (A + S) or (A + T).

With regard to the apparent variations in combining power displayed by monospecific antibody, Hershey (1941b) points out that it is not necessary to invoke differences in valency to explain the properties of "low grade" or of highly avid antibody. The "affinity" of a unit reactive patch on the antigen molecule for unit reactive patch on the antibody may be described in terms of a dissociation constant  $k$ , and Hershey has ingeniously subsumed most of the observed individual variations of antibody under variations in  $k$ . For example, a small  $k$  explains large maximal antigen antibody ratios, high avidity, and broadened reactivity in sera taken late in immunization. Heidelberg's univalent antibody, and non-precipitable antibody left after absorption of precipitable antibody, would have a large  $k$ . Variations in  $k$  would be independent of the nature of the immune



response, and of valency, and variations in  $k$  would explain the effect of salts, lipins and other non specific substances on the progress of immune reactions.

The analysis of specific precipitates in the region of antigen excess should provide the best direct evidence for or against the multivalence of antibody. In their review Hooker and Boyd (1942) point out that in only two systems have the recorded estimates of the molecular ratio of antigen to antibody been more than unity, namely, with diphtheria toxin (Pappenheimer, Lundgren and Williams 1940) and with Heidelberger and Kendall's (1934) azo-dye ovalbumin antigen—and in both cases the value of 2 is in doubt. We have already noted that the value of 2 accords well with Hershey's descriptive theory of the lattice hypothesis. It is clear that precise analyses of a large number of antigen antibody compounds of this kind are required to settle this point.

We may summarize the position with regard to the unity or diversity of antibodies as follows.

The antibodies in a monospecific antiserum prepared against a single antigen may vary in physicochemical properties, such as particle size, electrophoretic mobility, readiness to dissociate from union with antigen, and perhaps in valency, in combining power and "avidity", in resistance to destructive agents, and in capacity to form complexes with antigens that are susceptible to other substances in the reacting system such as electrolyte, complement, or a phagocyte. In no case however is there good evidence of association between a given set of physicochemical properties and a particular type of serological reactivity. The heterogeneity in a collection of antibody particles present in a certain preparation may affect the different serological reactions in different degrees, but there is no reason to suppose that opsonins, precipitins, antitoxins and so forth differ in kind, either constantly or fundamentally. In this sense then we may accept the hypothesis of the unity of antibodies.

### The Nature and Properties of Antigens

The two outstanding characters of antigens, their power to stimulate antibody production and their specificity may be to some extent explained in terms of their chemical constitution. The basis of specificity is the more clearly understood, and it will be convenient to discuss it first.

#### The Basis of Specificity

Specificity is not absolute. There is no one natural antigen of which it would be safe to predict that it would not react with antibody prepared against another antigen. The variety of syntheses of which living tissues are capable, though wide is limited and many tissue constituents in one organism are similar to those in another organism. Where these constituents are antigenic, we often find the similarity reflected in the serological reactions of the antigens with homologous and heterologous antibodies.

Thus the lens proteins of mammals are antigenically similar, though they differ markedly from other proteins found in the animal body. This type of specificity is referred to as 'organ' specificity, and has been noted in antigens from brain, testes and placenta, and in mammalian proteins like keratin, fibrinogen and thyroglobulin (see Landsteiner 1936).

On the other hand antigens are often 'species' specific. We can differentiate by serological tests between the egg albumin of the duck and of the hen (Dakin and Dale 1919) between the haemoglobins of different animal species (Higashi

1922 Landsteiner and Heidelberger 1923 Hektoen and Schulhof 1923) and between different vegetable proteins (Wells and Osborne 1911 Wells 1915 Jones and Gersdorff 1923 Lewis and Wells 1925 Wells *et al* 1927)

As the classical researches of Nuttall (1904) indicated in the first place this degree of specificity may be closely correlated with biological classifications so that we might expect cross reactions between the serum proteins of man and monkey but none between those of man and ox. But though this holds to a large extent later studies have revealed relationships between the antigens of organisms as widely separated as man and Shiga dysentery bacillus (see Chapter 8). A serological relationship therefore between two organisms can strengthen a relationship already established on other biological grounds but it cannot by itself be considered as good evidence of anything more than a similarity of perhaps a small part of the metabolism of the two organisms.

Analysis in many cases reveals chemical differences between native antigens but the complexity of the antigenic particles concerned is such that analyses are necessarily crude. Far more significant information has been gained by altering the chemical of protein antigens along certain limited and well-defined lines and noting the resulting changes in their immunological reactions.

Obermayer and Pick (1906) (see also Pick 1912) showed that the nitration or halogenation of proteins—that is, the introduction of the nitro group or of a halogen element such as iodine—profoundly altered the antigenic reactions of the treated protein. Serum proteins so treated lost their species specificity but they gained a new specificity shared by normally unrelated serum proteins that had been chemically altered by the same procedure. Thus an antiserum prepared against the nitrated serum of a particular animal species failed to react with the unaltered serum of that species but reacted with a wide range of nitrated sera from other animals. Since it was known that the nitro group and the halogen elements entered into the benzene ring of certain of the amino acids which build up the complex protein molecule Obermayer and Pick were led to attach particular importance to these chemical groupings as factors determining immunological specificity (see also Wormall 1930 Johnson and Wormall 1937 Snapper and Grunbaum 1936 Shahrokhi 1943).

Landsteiner and his colleagues however have shown that the salt-forming groups of the amino-acids (the carboxyl, hydroxyl and amino groupings) play an equally important part. By esterification methylation and acetylation they have succeeded in altering the immunological specificity of proteins (see Landsteiner and Prašek 1914 Landsteiner and Lampl 1917a Landsteiner 1917).

This relatively simple modification of active groups on the surface of the antigen was largely extended by Landsteiner and his colleagues (see Landsteiner 1930 1933 1936) who utilized the diazo reaction to introduce larger modifying groups into the protein molecule. The substances for introduction contain an  $-NH_2$  group usually attached to a ring structure in the molecule. The  $-NH_2$  group is converted to  $-N=NCl$  in presence of  $NaNO_2$  and  $HCl$  and the resulting diazo compound mixed with the protein. The diazo compound reacts with the phenolic groups of tyrosine (see Fig. 41) with the imidazole ring of histidine and probably with the indole group of tryptophan the  $-NH$  group of proline and hydroxyproline and all the aliphatic  $-NH_2$  groups on the protein surface (Eagle and Vickers 1936). By varying the conditions of the reaction it is possible to control the amount of modifying substance introduced into the protein. Fig. 41 illustrates the coupling of atoxyl (*p*-amino benzene arsenic acid) to a protein on the assumption that it unites with the phenolic group of the tyrosine residues in the protein molecule.

The results obtained with azo-proteins were criticized by Clutton, Harington and Mead (1937) on the ground that the diazo link is highly artificial and is never found in natural substances. They devised means of coupling carbohydrate to tyrosine, by an O-glucosido linkage and attaching the compound to the  $\text{NH}_2$  groups of protein. Subsequent work (Clutton, Harington and Yuill 1937) however, showed that the nature of the linkage to the protein had no observable effect on the modification of antigenic specificity induced by the glucosido-tyrosyl groups themselves. Another objection to the diazo method is the drastic chemical treatment to which the proteins are subjected during diazotization. Hopkins and Wormald (1933a, b) were able to modify the specificity of proteins by the much gentler action of phenyl isocyanate in alkaline solution. The isocyanate reacts with free  $-\text{NH}_2$  groups to form a substituted urea. The results of chemical

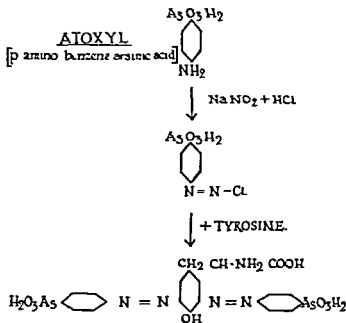


FIG. 41

modification by all these methods are essentially similar, and we may take those obtained by the extensively employed azo-proteins as valid.

In general the protein to which such a grouping has been attached will retain its species specificity, though usually with some loss of potency. Thus an antigen prepared by coupling atoxyl to horse globulin will react with an antiserum prepared against any atoxyl azo-protein in virtue of the atoxyl grouping, and with an antiserum prepared against horse globulin or against an azo-protein prepared from horse globulin in virtue of the specific horse-globulin groupings. This difficulty is overcome by coupling any grouping that it is desired to study to two immunologically unrelated proteins. One of these is used as the antigen for the preparation of the antiserum, the other is used for the *in vitro* tests. Thus, atoxyl may be coupled with horse globulin, and the atoxyl azo-protein so prepared may be used for the immunization of a rabbit. The serum so prepared will react with this antigen

in the test-tube in virtue of antibody groupings that unite specifically with unaltered horse globulin. But if atoxyl is also coupled to chicken globulin, and the atoxyl azo-protein so prepared is used in the *in vitro* tests, the precipitation that occurs will depend solely on antibody groupings acting specifically on atoxyl, since horse globulin and chicken globulin show no antigenic relationship.

Using these methods, it has been possible to prepare antisera that give specific precipitation with synthetic antigens in which the active groupings are provided by such substances as metanilic acid, atoxyl, levo-, dextro- and meso-tartaric acid glucosides, galactosides, and so on (Landsteiner and Lampl 1917*b*, Landsteiner 1919, 1930, Landsteiner and van der Scheer 1928, 1929, 1931, 1932*a*, *b*, 1934*a*, *b*, Avery and Goebel 1929, Avery, Goebel and Babers 1932, Goebel, Avery and Babers 1934), dipeptides and pentapeptides (Landsteiner and van der Scheer 1932*a*, 1939), pyrazolon compounds (Erlenmeyer and Berger 1934, Harte 1938), pyridine (Landsteiner and Pirie 1937), strychnine (Hooker and Boyd 1940), thyroxine (Clutton Harington and Yull 1938), aspirin (Butler Harington and Yull 1940) and histamine (Fell *et al.* 1943).

These results clearly demonstrate an immunological specificity dependent upon chemical groups of known structure. In the last three examples the specific action of the antibodies concerned could be shown by a marked reduction of pharmacological activity of the thyroxine, aspirin and histamine in animals treated with antisera prepared against thyroxyl protein, aspiryl protein and histamine protein respectively (see also Singer p. 259).

The comparison of antigens modified by optically active isomers, as, for instance, with dextro-, meso- and levo-tartaric antigens, or with glucoside and galactoside antigens (see also Woolf, Marrack and Downie 1936), in which the isomers differ only in the arrangement round a single carbon atom, shows clearly that, as might be expected, stereo-chemical differences in structure are important determinants of immunological specificity.

Specificity depends in part on the nature of any grouping in a complex organic molecule, and in part on the position in the molecule which the grouping occupies. Fig. 42 gives an example—the cross precipitations observed with an antigen prepared from *o*-amino benzene sulphonic acid (see Landsteiner 1919, Marrack 1934).

In the centre of the figure, labelled [G], is represented the active grouping of the azo protein used as an antigen in the precipitation tests. Around it, labelled [A], are placed antibodies prepared against antigens synthesized by coupling the groupings shown to some unrelated protein. The double-headed arrows in

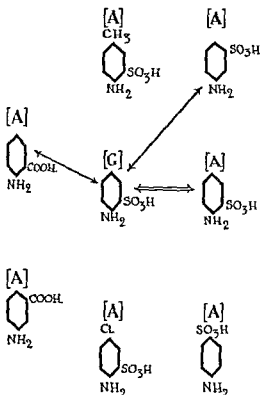


FIG. 42.—PRECIPITATION

When this procedure is adopted, it is found that relatively simple substances may function as incomplete, or partial antigens in the sense that they combine specifically with corresponding antibodies. Thus, diazotized atoxyl coupled with tyrosine, instead of with an intact protein molecule, does not form a precipitate when mixed with the corresponding antiserum but it inhibits the precipitation that would normally occur when this antiserum is mixed with an atoxyl azo protein. A similar inhibition may be demonstrated with atoxyl itself in higher concentrations, or even with arsenic acid when this is added in sufficient amount (Landsteiner 1920).

When the inhibition reaction is used to determine the immunological behaviour of non identical but structurally related organic compounds it is found that the specificity is less strict than that displayed in cross precipitation tests. Fig 43 illustrates the findings in relation to *o* amino benzoic acid (see Landsteiner and van der Scheer 1931, Marrack 1938). The antibody prepared against an azo protein containing this grouping, indicated at the top of the figure, gives specific precipitation with another azo protein containing the same active grouping but a different protein component, indicated at the bottom of the figure. Inhibition by various other groupings is indicated by the horizontal broken arrows. The reaction is inhibited not only by benzoic acid, or benzoic acid substituted either in the ortho or meta position by the methyl group but by compounds showing considerable structural differences, such as thiophene carboxylic acid and naphthoic acid.

The specific combination of certain azo dyes with the corresponding antibodies, without the formation of a precipitate, has also been demonstrated by Marrack and Smith (1932), using an ingenious colorimetric technique.

We have already noted that the partial antigens that inhibit precipitation do not generally themselves yield a precipitate *in vitro*. Neither will they stimulate the production of precipitins *in vivo*. In the definition of an antigen that was given at the beginning

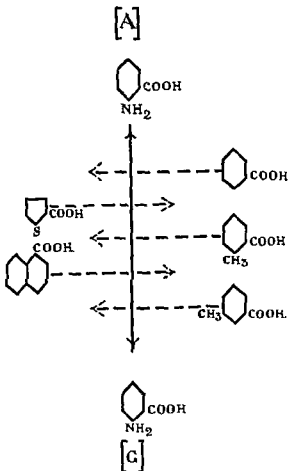


FIG 43—INHIBITION

dicates the instances in which precipitation occurs. The reaction is strongest with the homologous antigen, containing the  $\text{C}_6\text{H}_4\text{SO}_3\text{H}$  group, but precipitation occurs when the  $\text{SO}_3\text{H}$  group is shifted to the meta position ( $\text{C}_6\text{H}_3\text{SO}_3\text{H}$ ), or when it is replaced, in the ortho position, by  $\text{COOH}$  ( $\text{C}_6\text{H}_4\text{COOH}$ ). It does not occur when the  $\text{SO}_3\text{H}$  group is shifted to the para position ( $\text{C}_6\text{H}_3\text{SO}_3\text{H}$ ), or when the  $\text{SO}_3\text{H}$  group in the ortho position is replaced by a  $\text{COOH}$  group in the meta position ( $\text{C}_6\text{H}_3\text{COOH}$ ). The reaction is also abolished by introducing a methyl or chlorine group in the para position, even though the  $\text{SO}_3\text{H}$  group is retained in the ortho position.

Another example, illustrating the importance of spatial configuration and the immunological equivalence of different chemical groups so long as this spatial configuration is maintained is afforded by the studies of Erlenmeyer and Berger (1932). They found that azo-proteins prepared from the compounds  $\text{NH}_2\text{C}_6\text{H}_4\text{O}\text{C}_6\text{H}_4\text{NH}_2$ ,  $\text{NH}_2\text{C}_6\text{H}_4\text{NH}\text{C}_6\text{H}_4\text{NH}_2$  and  $\text{NH}_2\text{C}_6\text{H}_4\text{CH}_2\text{C}_6\text{H}_4\text{NH}_2$  behaved similarly in precipitation reactions though  $\text{NH}_2\text{C}_6\text{H}_4\text{C}(=\text{O})\text{C}_6\text{H}_4\text{NH}_2$  reacted differently.

The studies of Landsteiner and van der Scheer (1932a, 1934b) on peptides also illustrate the effect of relative position on active groups in a complex molecule.

They prepared synthetic azo proteins from the dipeptides glycyl glycine, leucyl leucine, glycyl leucine and leucyl glycine. It was found that the terminal amino-acid carrying the carboxylic group had the greater influence on immunological specificity. Thus a glycyl leucine antiserum reacted best with the corresponding antigen, less effectively with a leucyl leucine antigen and much less effectively with a glycyl glycine or leucyl-glycine antigen. The nature of the penultimate amino acid also influenced the reaction and in later studies (1939) with pentapeptides made from various combinations of glycine and leucine it was found that the arrangement of all five members of the polypeptide chain affected specificity.

The influence of the whole substituted group apart from that of active acidic or other sub groups, is well shown in the studies of Erlenmeyer and Berger (1932). Mutsaers and Gregoire (1935) and Jacobs (1937), with compounds containing more than one benzene ring. Here the nature of the whole compound was prominent in determining specificity.

To obtain precipitation with these synthetic antigens and the corresponding antisera it is in most cases necessary to employ the complete antigen, i.e. the active group coupled to a suitable protein.

The specificity of an antiserum however, can be tested by adding to it the active substance, alone or coupled to a substance simpler than protein. Combination of the group with the antibody is then tested by adding the complete antigen. If precipitation is inhibited, we may conclude that the simpler fragment of antigen has combined with the antibody, forming a soluble compound and blocking its combining groups.

When this procedure is adopted, it is found that relatively simple substances may function as incomplete, or partial antigens, in the sense that they combine specifically with corresponding antibodies. Thus, diazotized atoxyl coupled with tyrosine, instead of with an intact protein molecule, does not form a precipitate when mixed with the corresponding antiserum, but it inhibits the precipitation that would normally occur when this antiserum is mixed with an atoxyl azo protein. A similar inhibition may be demonstrated with atoxyl itself in higher concentrations, or even with arsenic acid when this is added in sufficient amount (Laudsteiner 1920).

When the inhibition reaction is used to determine the immunological behaviour of non identical but structurally related organic compounds, it is found that the specificity is less strict than that displayed in cross precipitation tests. Fig 13 illustrates the findings in relation to *o* amino benzoic acid (see Landsteiner and van der Scheer 1931, Marrack 1938). The antibody prepared against an azo protein containing this grouping, indicated at the top of the figure, gives specific precipitation with another azo protein, containing the same active grouping but a different protein component, indicated at the bottom of the figure. Inhibition by various other groupings is indicated by the horizontal broken arrows. The reaction is inhibited not only by benzoic acid, or benzoic acid substituted either in the ortho or meta position by the methyl group but by compounds showing considerable structural differences, such as thiophene carboxylic acid and naphthoic acid.

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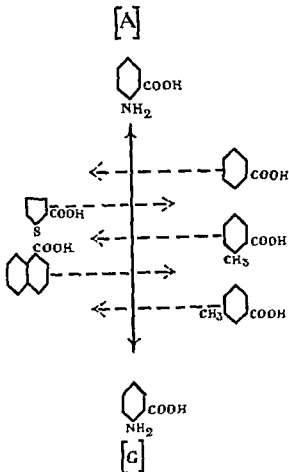


FIG 43—INHIBITION

of this chapter the capacity to act as a stimulant of antibody production was included among its essential properties—it is, indeed, the property from which its name is derived. The difference in behaviour between substances of varying complexity, all bearing the same active specific grouping led Landsteiner to formulate the conception of the *hapten*, or partial antigen, which, while bearing the specific grouping and therefore combining with the homologous antibody, lacks certain of the properties of the complete antigen.

In subsequent chapters frequent reference will be made to various polysaccharide components derived from different types and species of bacteria. Many of these polysaccharides from the immunological point of view, occupy a position intermediate between Landsteiner's simpler haptens and complete antigens. They react specifically with the homologous antisera *in vitro*, giving precipitation, but they do not induce antibody formation *in vivo*, or, if they do so at all, the stimulus they provide is of an altogether different order from that provided by the complete antigen as it occurs in the bacterial cell. It seems likely that the range of activity of a partial antigen is determined in the main by its molecular size. Landsteiner and van der Scheer (1932b) coupled succinic, adipic and suberic acids, through their aniline compounds, to proteins, and prepared antisera against the complete antigens so produced. Partial antigens were prepared by coupling the same compounds to resorcinol. These partial antigens gave precipitation with the corresponding antisera, and it was noted that the partial antigen derived from suberic acid gave particularly strong precipitin reactions, probably on account of its long aliphatic side chains.

It is convenient to classify the antigenic complexes we have described under the following headings

- (a) Simple haptens, possessing immunological specificity, combining with the homologous antibody and so preventing precipitation, but neither forming a precipitate *in vitro*, nor stimulating antibody production *in vivo*
- (b) Complex haptens, possessing immunological specificity, combining with the homologous antibody and forming a precipitate, but not stimulating antibody production *in vivo*
- (c) Complete antigens, possessing immunological specificity, combining with the homologous antibody and forming a precipitate, and stimulating antibody production *in vivo*

It must not, however, be supposed that there is any sharp natural demarcation between our named classes. It seems likely that the characters that determine full antigenic potency are high molecular weight, associated with a large molecular surface and a multiplicity of active specific groupings, and the relative activity of these groupings themselves. If we were in a position to give a detailed description of the chemical structure and immunological properties of a very large number of antigenic substances, ranging from the simplest to the most complex, we should probably find that they formed a continuous rather than a discontinuous series, the falling-off in immunological activity as we passed from large and complex to small and simple molecules being marked by the gradual loss first of one immunological property, then of another, until we were left only with the ability to combine specifically with the antibody under optimal conditions of concentration and other relevant factors.



### The Basis of Antigenicity

The distinction between haptens and antigens brings us to the second of our inquiries the basis of antigenicity. It seems likely that the characters determining full antigenic potency of a substance are high molecular weight and the presence on the molecular surface of groupings that are distinctive either by reason of their molecular shape or by reason of high electrochemical activity of certain sub groups within the group. There are no rigid criteria by which we may judge whether a substance is likely to be antigenic or not. There are only a number of features associated with antigenicity.

It is sometimes stated that substances are antigenic because they are foreign to a given species of animal. This criterion is not very helpful. If we define foreign as harmful to the animal we shall cover antigens like snake venom or diphtheria toxin but not non toxic antigens like ovalbumin. If we define it as derived from a different species of organism we shall expect for example insulin from the ox to be antigenic in the rabbit which is not the case. If quite unjustifiably we define it as not normally present in the tissues of the animal we assume either that we have a complete specification of the normal constituents of these tissues or that we can always recognize an abnormality and we shall find in a number of cases that our only criterion of abnormality of a substance is the response of the tissues to it as an antigen. That is the term foreign can be correctly used to describe all antigens only when it is synonymous with the term antigenic.

We have already noted that antibody response may be conditioned by various distinguishable groupings on the surface of the antigenic particle and we may reasonably expect that antigenicity itself will be determined by certain combinations of chemical groups. These groups and combinations of them are called antigenic "determinants" but in seeking to relate antigenicity to the determinants of an antigen we must not be misled by a too restricted notion of a determinant. To call atoxyl a determinant in atoxyl azo protein does not necessarily imply an additive effect when atoxyl is linked to the protein. The true determinant is atoxyl in certain positions on the surface of the protein. The atoxyl covers some native groups or renders them in some way inaccessible to antibody and the reactivity of other surface groups may be altered by the presence of the new group. Moreover, the determinant effect that we attribute to atoxyl may depend on its electrochemical and spatial relations with the rest of the surface, whether antigenic or not.

We may illustrate this distinction by reference to the determinant effect of organic arsenic compounds. Singer (1942) prepared antisera to arsenic ox globulin and used it to protect mice against lethal doses of the synthetic antigen. Successive adsorption of the sera with arsenic rabbit globulin and ox globulin did not remove all protective power clearly antibodies to the protein-determinant complex were left in the serum.

The sphere of influence of the determinant will clearly depend on the chemical structure of the antigen. A protein antigen like horse globulin could not be spread in a unimolecular film without losing reactivity with antibody (Danielli, Danielli and Marrack 1938). Here the relationship of amino acids in the complete protein molecule was a determinant of specificity. On the other hand the films of nucleoprotein and other fractions of streptococci spread in thin expanded films on the surface of slides still retained a conspicuous degree of specific reactivity (Bateman, Calkins and Chambers 1941; Chambers, Bateman and Calkins 1941).

Similarly with the fractionation of a native antigen, the loss of specificity that follows the removal of a recognizable chemical compound does not necessarily mean that the compound as such is the determinant and even if the specificity is restored when the compound is reintroduced the essential determination of specificity lies in the restoration of the relations between the compound and the remainder of the molecule.

Full proteins are usually antigenic though among the larger mammals the antibody response to protein antigens of other mammalian species is partly conditioned by the function as well as the structure of the protein. As Harington (1940) has pointed out the metabolic proteins like serum globulin are fully antigenic the storage proteins like lens proteins and casein less so and hormonal proteins like insulin and thyroglobulin apparently shared by a large number of animal species have little or no antigenic power.

The early successes in the study of protein antigens the early failures to demonstrate antigenic response to the injection of carbohydrates and fats, and the fact that most of the known antigens contained at least a substantial amount of protein led to the supposition that all antigenicity was conferred by protein and that in the alleged examples of non protein antigens the substances were contaminated with chemically undetectable but immunologically active traces of protein. It is in many cases difficult to exclude the possibility of antigenic contaminants in a purified preparation. The scanty work for example (see Marrack 1938) on the antigenicity of lipins suffers from this defect. There is another way in which a substance may be erroneously called antigenic. It may combine with the native proteins of the animal into which it is introduced and convert them into 'foreign' proteins that stimulate antibody formation. We shall discuss examples of this instructive phenomenon in connection with hypersensitivity (Chapter 51). But carbohydrate substances have been prepared whose protein content is so low that the possibility of an effective antigenic stimulus may be neglected, but which act as full antigens and the unique efficacy of proteins in conferring full antigenicity can no longer be upheld.

Among the proteins, gelatin is not antigenic. It lacks tyrosine and tryptophan and has only a little phenylalanine a fact which suggests that the aromatic radicles are necessary for antigenicity (see Wells 1925). Hopkins and Wormald (1933b) failed to make gelatin antigenic by introducing aromatic radicles. Hooker and Boyd (1933) however demonstrated that antibodies were formed to a diazo arsanilic acid gelatin complex and Clutton Harington and Mull (1938) had a similar success with glucosido-tyrosyl gelatin. The absence of an aromatic radicle is not, however the sole reason for non-antigenicity. Insulin which is a full protein containing aromatic radicles, is in no sense a full antigen. Various workers have reported anaphylactic sensitization with insulin preparations (Barral and Roux 1931, Lewis 1933, Bernstein, Kirsner and Turner 1938) and Wasserman, Brohahn and Mirsky (1940) record the production of complement fixing antibodies in the rabbit. Insulin nevertheless does not stimulate frankly precipitating antibodies, but the introduction of a glucosido-tyrosyl residue converted insulin into a full antigen (Clutton Harington and Mull 1938). That tyrosine and other aromatic radicles are not all important is also shown by the masking of original specificity that occurs when determinants are linked to amino-acid residues containing no aromatic rings. From the alterations induced in serum pseudoglobulin by different degrees of substitution of azo-atoxyl, Haurowitz, Sarafian and Schwerin (1941) concluded that specificity depended on a determinant arrangement of tyrosine, free amino and perhaps other groups on the molecular surface. In native lens proteins, the sulphydryl groups are apparently important determinants (Ecker and Pillemer 1940).

A single protein molecule may possess more than one set of determinants, in the sense that on injection into an animal two distinct types of specific antibody are produced (Hooker and Boyd 1936). Haurowitz (1937) also was able to demonstrate distinct antibodies to globulin, arsanyl azo-globulin and to the arsanylic group alone, in rabbits immunized with an apparently homogeneous preparation of arsanyl azo-globulin.

Further, Haurowitz, Tunca and Schwerin (1943) have shown that non antigenicity may in fact be due to failure of retention on the part of the animal. One hour after injection of arsanyl azo gelatin, rabbit liver contained 4.4 per cent of the total dose, as compared with 34 per cent arsanyl azo globulin, while the corresponding figures in urine were 66 per cent and 92 per cent. Their observation obviously indicates a reconsideration of many examples of non antigenicity that have been attributed to the absence from the antigenic molecule of substances supposed to render it capable of stimulating antibody production. A failure of adsorption on to certain cells, or of combination with body substances may also underlie the varying response of different species of animals to the Type I pneumococcal polysaccharide. It is antigenic, or at least capable of stimulating a specific immunity, in man, horse, cat, dog and mouse, but not in the sheep, rabbit, guinea pig or rat (see Horsfall and Goodner 1936, Downie 1937).

**The Antigenicity of Denatured and Degraded Proteins**—Denaturation by heat or by prolonged exposure to alkali, which lead to racemization, changes the antigenic behaviour of proteins (see Hartley 1931, Wells 1929, Marrack 1938). Of particular interest is the effect of reversible denaturation. When horse or ox serum albumin was denatured by strong urea solution, and regenerated by dialysis the protein was found to be less antigenic but apparently had lost none of its specificity (Erickson and Neurath 1943, Martin, Erickson, Putnam and Neurath 1943). The loss of antigenicity was attributed to breakdown of internal structure of the molecule, the retention of specificity to the preservation of a characteristic arrangement of amino-acid residues in the polypeptide chains.

These authors also noted an association between antigenicity of various protein preparations and their carbohydrate content. Crystalalbumin, which was the most feebly antigenic of the preparations studied, contains no carbohydrate. Lack of carbohydrate cannot be the sole reason for low antigenicity, for the regenerated, feebly antigenic albumins had lost none of their original carbohydrate, though it is possible that the mode of linkage may have been drastically altered. The carbohydrate in some proteins may be loosely attached, but in others, like serum and egg white protein, it forms an integral part of the molecule and is not separable until polypeptide chains are broken. Some influence on antigenicity is to be expected. Indeed, the accumulating evidence that a number of native antigens consist of complexes of polypeptides, lipins and carbohydrates suggests that the antigenicity of a protein may depend on the presence of its non protein components. The striking example of the dependence of antigenicity upon a complex of protein and non protein substances is provided by Morgan and Partridge's (1941) analysis of a bacterial antigen in *Shigella shiga*. This was separable into a feebly antigenic polypeptide and a non-antigenic polysaccharide, recombination of the two restored the full antigenic properties. On the other hand there is no evidence that the carbohydrates associated with some of the fractions separable from serum albumins and globulins (Coghill and Creighton 1938, Rumington and van den Ende 1940) and from egg albumin and ovomucoid (Ferry and Levy 1934, Sevag and Seastone 1934, Neuberger and Yuill 1940) have any effect on antigenic specificity. The extent to which these carbohydrates are integral parts of native protein however, is not yet clear.

The immunological reactivity of proteins decreases with progressive hydrolysis. The stage at which the hydrolysed protein ceases to react serologically varies with the protein

(see Stull and Hampton 1941) Landsteiner (1942) found that polypeptide chains of 8-10 amino-acids from hydrolysed silk fibroin were haptens in that they inhibited the reaction between fibroin and specific antibody and concluded that the determinant units of the protein were of this size

A number of mechanisms can be used by an animal to remove or neutralize a substance that has been introduced into it such as excretion, if the substance is small enough to get through the kidney hydrolysis if suitable enzymes are present absorption by phagocytes or by the cells of the reticulo-endothelial system (see Chapter 50) or in the case of substances like insulin whatever mechanisms normally remove indiffusible hormones etc. after they have produced their physiological effect. In these circumstances a substance will be antigenic only if it is too big to be excreted too stable to be hydrolysed and so unusual that it is more readily handled by the mechanism resulting in antibody formation than by the more direct physiological modes of destruction and elimination.

*In summary* then it appears that to be an antigen a substance must have certain minimum chemical properties, but it must also be so constructed that it cannot be handled by one of the readily available mechanisms of elimination and thus removed before it has time to exert an antigenic stimulus.

#### The Effective Number of Determinants, and the Valency of Antigen.

The valency of native antigens may be estimated from analysis of specific precipitates in the region of antibody excess. This figure gives only the *minimal* number of combining groups on the antigen molecule since spatial considerations may not permit their full saturation with antibody in the precipitate. The values range from 5 for crystalline ovalbumin (molecular weight about 40 000) (Heidelberger and Kendall 1936; Heidelberger 1938) to 231 for the hæmocyanin (molecular weight  $5-6 \times 10^4$ ) of the crab *Variparus* (Malkiel and Boyd 1937).

The number of valencies is correlated with the surface area of the antigenic protein molecules (Hooker and Boyd 1947) and it appears that the maximal size of a determinant supposing it to be protein in nature is of the order of 30 amino-acids.

By coupling arsanilic acid in varying proportions to casein and testing the compounds against antisera prepared against another arsanil azo-protein Hooker and Boyd (1937) calculated that an average of at least 13 introduced groups per molecule of casein was necessary for precipitation. Haurowitz Kraus and Marx (1936) similarly estimated 10-20 groups for arsanil azo-globulin. The assumption is made that the protein particles are of full molecular weight. During the preparation of azo-proteins however the average particle size may decrease (Hooker personal communication) and the minimal number of reacting groups may in fact be lower than the calculated 10-13.

#### Unrecognized Antigens

Our definition of an antibody implies that it should react in a *detectable* manner with the corresponding antigen and we consequently infer antigenicity when a substance reacts specifically in a detectable manner with the antiserum prepared against it. It is nevertheless possible for a substance to be antigenic in the formal sense of stimulating antibody production and yet be non reacting. As we saw in the preceding section 10-20 arsanilic acid determinants were necessary for the precipitation of arsanil-azo-protein by its antibody. Yet the introduction of an average of one arsanilic acid group per molecule of protein gives

a preparation that stimulates the production of antibodies to the azo compound (Haurowitz 1937)

The important observation of Bawden and Kleczkowski (1941, 1942a) that, when antigens like bushy stunt virus or human serum globulin are heated in the presence of a serum albumin, complexes are formed which, though non precipitating are still antigens, demands a similar modification of the conception of antigenicity. The antiserum to the albumin globulin complex precipitated with globulin alone; the precipitation was inhibited by the complex, which in this respect behaved like a hapten. There is no good reason to suppose that antigenic, but non reacting substances do not occur in nature. The examples from experimental serology suggest that they may be discovered in the first case by reaction between the "antiserum" and a heterologous antigen that happens to possess a large number of the determinant groups characterizing the "antigen," and in the second case by reactions between "antiserum" and degradation products of the apparently non antigenic substance, though it would be hard to predict the kind of degradation processes likely to yield positive results. Bawden and Pirie (1944) have provided a natural example of the first kind. Extracts of the leaves of tomatoes infected with the bushy stunt virus contain a substance which on injection into rabbits, yields an antiserum that does not precipitate with the substance but precipitates with bushy stunt virus obtained from sap. In this case the non precipitating antigenic complex consisted of virus and a chromoprotein; on separation, the former precipitated with the antibody, and the latter had an inhibiting effect on this precipitation.

#### The Nature of the Antigen-antibody Union

The large body of evidence makes it quite clear that the antibody receptor is adapted with a high degree of precision to the surface of native antigens and that the adaptation is changed by imposing additional determinants on the antigenic surface. The nature of the adaptation is not clear. The surface of a large molecule which may as a whole be electrically neutral is characterized by positively or negatively charged atomic groupings, which create localized electric fields in the immediate neighbourhood of the molecule, operative over a very short distance.

When two molecules approach each other closely these fields if of opposite sign, will cause an attraction. In large molecules—and it will be remembered that it is with reactions between large molecules that we are mainly concerned—the electric fields set up may be very numerous, and they will have a quite definite spatial arrangement. Taken in conjunction with the shortness of the distance over which these intermolecular forces are operative, this complexity and constancy of pattern provide just the conditions required for specificity. Two molecules possessing at their surface electric fields so arranged that, when they come into close contact, there will be multiple points of attraction between them, will tend to adhere to one another. Two molecules possessing the same number of electric fields, but with these so arranged that close contact cannot be made at many points simultaneously, will show no tendency to adhere. The better the fit, in this specialized sense, the greater will be the total force of attraction.

One of the best available examples of the action of these intermolecular forces is the selective formation of mixed crystals. The molecules involved must conform to the required pattern in regard to their dimensions and the relative position of their active groupings, or atoms. As a result most crystals consist of one kind of molecule alone, but molecules of different kinds may be built into a single crystal, provided

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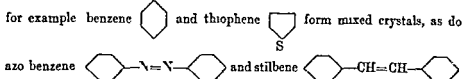
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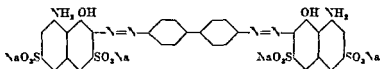
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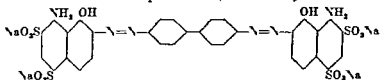
that the structural differences between them do not exceed certain limits. Thus



The formation of mixed crystals might, perhaps, be regarded as analogous to the formation of a precipitable antigen-antibody compound, particularly if we adopt Marrack's lattice hypothesis. It is of interest to note that the phenomenon of crystallization also presents an analogy to the inhibition of precipitation by haptens. Many crystals will specifically adsorb on to their surfaces the molecules of another substance, with which they will not form mixed crystals. Such adsorption inhibits the growth of the crystal by the addition of further molecules of the substance of which the crystal is composed. The conditions, in regard to molecular size and position of active groupings, that determine the specificity of adsorption on the surface of a crystal, are less strict than those that determine availability for building into a complete crystal lattice. This would be expected, since only one aspect of the adsorbed molecule need conform to the distribution of active groupings on the surface of the molecule. Relatively small differences may, however, be sufficient to determine the occurrence or non-occurrence of adsorption. Thus (France 1930) the dye



is adsorbed on the cube faces of potash alum, while the dye



is not

These analogies cannot be pressed far for many of the substances which are known to act as antigenic determinants could not find in any of the geometrically possible rearrangements of the polypeptide chains of an antibody globulin, configurations that are specific in the sense that a surface of a crystal fits molecules of its own kind. Moreover Pauling, Campbell and Pressman (1943) point out that the attractions between permanent electric charges and dipole moments on the molecule are small in water, and have probably little effect in antigen-antibody reactions, the attractions are more likely to be due to van der Waals' forces and to hydrogen bonding. Van der Waals forces are due to momentarily induced charges between molecules having no permanent dipole moments. Considered singly these forces are small and would be strong enough to account for attraction of antigen and antibody only if large areas of each molecule were approximated sufficiently to give a summation effect of hundreds of van der Waals attractions. Hydrogen bonding occurs when the hydrogen attached to a strongly electronegative atom is attracted to the unshared electron pair of another electronegative atom (co-ordinate covalency). Since all these forces are non specific,



specificity must depend on the existence of an area on the surface of the antibody molecule that is in structure complementary to an area on the antigen molecule, enabling large surfaces to approach close enough for the forces to operate over a high proportion of the area of contact. Van der Waals' forces, for example, decrease as the 7th power of the distance and are active only at distances of 0.1 to 0.3  $\mu\mu$ .

The complementary nature of the two surfaces must include adaptation on the part of the antibody to different determinants, their spatial relationships, and the relative accessibility of the various groups. Cross reactions between heterologous antigens and antibodies, when there is good reason to believe they are due to a monospecific antibody, and not to other types of antibody in the antiserum, would result from the juxtaposition of less complementary areas, so that the attractions are feebler, the dissociation of the antigen antibody link greater, and, as has been observed, the inhibition with haptens more effective, than in homologous reactions (Landsteiner and van der Scheer 1936).

Since it is likely that added determinants protrude from the surface of 'synthetic' antigens, Hooker and Boyd (1942) suggest that the corresponding surface of the antibody is a depression. Indeed it may be that a spatial protrusion is a necessary condition for determinant action. This speculation is supported by the failure of all attempts to produce specific anti antibodies, for if the antibody receptors were depressions on the surface of the globulin molecule they would on this basis have no determinant action. The anti antibodies, as we have seen react equally well with normal globulins from the same animal, and when they react with the antibodies, do not interfere with the original antibody receptors.

## REFERENCES

- ABRAMSON, H. A. (1935) *Nature Lond.* **135**, 995.  
 ADAIR, M. E. and TAYLOR, G. L. (1936) *J. Hyg. Camb.* **36**, 564.  
 AGNEW, S., SPINK, W. W. and MICKELSEN, O. (1942) *J. Immunol.* **44**, 297.  
 ALEXANDER, H. E. and HEIDELBERGER, M. (1940) *J. exp. Med.* **71**, 1.  
 AMAKO, T. (1911) *Z. Immunforsch.* **8**, 168.  
 ANDERSON, T. F. and STANLEY, W. M. (1941) *J. biol. Chem.* **139**, 339.  
 ANDO, K. (1937) *J. Immunol.* **33**, 41.  
 ANDO, K., KEE, P. and KOMIYAMA, T. (1937) *J. Immunol.* **32**, 181.  
 ANDO, K., KEE, R. and MANAKO, K. (1937) *J. Immunol.* **32**, 83.  
 ANDO, K., MANAKO, K., KEE, R. and TAKEDA, S. (1937) *J. Immunol.* **33**, 27.  
 ANDO, K., MANAKO, K. and TAKEDA, S. (1938) *J. Immunol.* **34**, 295.  
 ANDO, K., TAKEDA, S. and HAMANO, M. (1938) *J. Immunol.* **34**, 303.  
 ARRHENIUS, S. (1904) *Ark. Kemi, Fysik. Astr.*, **20**, 559, (1915) *Quantitative Laws in Biological Chemistry*. London.  
 ARRHENIUS, S. and MADSEN, T. (1902) *Festskrift. Skaatsens Serum Inst.*, No. 3, (1904) *Zbl. Bakt.* **38**, 612, (1904) *Ibid.* **37**, 1.  
 ASTBURY, W. T. (1943) *Advances in Enzymology* **3**, 63.  
 AVERY, O. T. and GOEBEL, W. F. (1929) *J. exp. Med.* **50**, 533.  
 AVERY, O. T., GOEBEL, W. F., and BARNES, F. H. (1932) *J. exp. Med.* **55**, 769.  
 BAIL, O. and HOKE, E. (1908) *Arch. Hyg.* **64**, 313.  
 BAIL, O. and TSUDA, K. (1909) *Z. Immunforsch.* **1**, 546.  
 BANTHAY, E. J. and GIBSON, R. B. (1907) *J. biol. Chem.* **3**, 253.  
 BARR, M. (1932) *J. Path. Bact.* **35**, 913.  
 BARR, M. and GLENNY, A. T. (1931) *J. Path. Bact.* **34**, 539.  
 BARR, M., GLENNY, A. T., and POPP, C. G. (1931) *Brit. J. exp. Path.* **12**, 217.  
 BARRELL, P. and ROY, J. (1931) *C. R. Soc. Biol.* **106**, 292.  
 BATEMAN, J. B., CALKINS, H. E. and CHAMBERS, L. A. (1941) *J. Immunol.* **41**, 321.  
 BAWDEN, F. C. and KLECZKOWSKI, A. (1941) *Brit. J. exp. Path.* **22**, 208 (1942a) *Ibid.* **23**, 169, (1942b) *Ibid.* **23**, 178.  
 BAWDEN, F. C. and PIRIE, A. W. (1944) *Brit. J. exp. Path.* **25**, 68.

- BECHHOLD H. (1904) *Z phys Chem.*, 48 385  
 BEHRING ROY and KITASATO (1890) *Dtsch med Wochr.*, 16, 1113.  
 BENIASCH M. (1911) *Z Immunforsch.*, 12, 268  
 BERGER, F M (1943) *Brit J exp Path.*, 24, 252.  
 BERNHEIM F, NEURATH, H., and ERIKSON J O (1947) *J Biol. Chem.*, 144, 259  
 BERNSTEIN C., KIRSNER, J B and TURNER, W J (1935) *J Lab. clin. Med* 23, 938  
 BILTZ, W (1910) *Biochem Z.*, 23, 27  
 BORDET J (1935) *Ann Inst Pasteur* 9 46° (1893) *Ibid.*, 12, 688 (1899) *Ibid.*, 13, 25  
 (1903) *Ibid.*, 17 161  
 BORDET J and GENGOU O (1901) *Ann Inst. Pasteur* 15 290  
 BOYD W C. (1940) *J Immunol.*, 28, 143 (1941) *J exp. Med* 74, 369 (1947) *Ibid.*, 75 407  
 BOYD W C and BERNARD H (193) *J Immunol.*, 33, 111  
 BOYD W C, CONY J B, GREGG D C., KINTIAKOWSKY C B. and ROBERTS R. M. (1941) *J Biol. Chem.*, 139 87  
 BOYD W C. and HOOKER, S. B. (1934) *J gen. Physiol.*, 17, 341 (1938) *Proc. Soc exp. Biol. N. Y.*, 39 491  
 BOYD W C and FURVELL, M A. (1944) *J exp Med.*, 80 289  
 BRAND E. (1907) *Berl klin Wochr.*, 44, 1075.  
 BREKID, F and HAUBOWITZ, F (1930) *Z physiol. Chem* 192 45  
 BROOM J C and BROWN H. C (1930) *Brit J exp Path.*, 11, 302.  
 BROWN A M. (1935) *Brit. J exp Path.*, 16 504.  
 BROWN G C (1943) *J Immunol* 46, 319  
 BROWN H C and BROOM J L. (1939) *Brit J exp Path.*, 10 387  
 BROWN H R. (1938) *J Lab. clin. Med.*, 23, 127  
 BROWNLEE, J (1935) *J Hyg., Camb.*, 23, 437  
 BUCHANAN P E. (1919) *J Bact.* 4, 73.  
 BUCHNER, H (1889a) *Zell Bakt.*, 5 81 (1889b) *Ibid.*, 6, 1 (1889c) *Ibid.*, 6 571  
 BUKANTZ, S C, COOPER, A and BULLOWA J G M. (1941) *J Bact.*, 42, 15  
 BULLOCH W B and WESTERN G T (1906) *Proc roy Soc., B* 77 531  
 BURNET F M (1931) *J Path. Bact.* 34, 41  
 B SING K. H and ZUZACK, H. (1943) *Z Immunforsch* 102, 401  
 BUTLER, G C, HARRINGTON C P and YCILL, M. E. (1940) *Biochem J.*, 34, 838.  
 BUXTON B H. (1900a) *J med Res* 13, 305 (1900b) *Ibid* 13, 431 (1900c) *Ibid.*, 13, 461  
 CANNOY P R and MARSHALL, C E. (1940) *J Immunol.*, 38, 365  
 CHAMBERS, L A., BATEMAN J B., and CALKINS H E. (1941) *J Immunol* 40 453  
 CHAPIN W S. and COWIE, D M (1907) *J med Res* 17 213  
 CHOW B F and GORREL, W F (1935) *J exp Med* 62, 19  
 CHOW B F and ZIA S H. (1938) *Proc. Soc. exp. Biol. N. Y.* 38, 695.  
 CHU F and CHOW B F (1938) *Proc Soc exp. Biol. N. Y.* 38, 679  
 CLUTTON R F, HARRINGTON C R., and MEAD T H. (1937) *Biochem J.*, 31, 764.  
 CLUTTON R F, HARRINGTON C P and YCILL, M. E. (1938) *Biochem J.*, 32, 1111  
 1119  
 COCA A F (1914) *Z Immunforsch.*, 21, 604  
 COGHILL, R D and CREIGHTON M (1938) *J Immunol.*, 35 477  
 COLWELL C A. and YOUNG G P (1941) *J infect Dis.*, 63, 226 *J Immunol.*, 42, 9.  
 COSTA CRUZ, J DA. (1939) *C P Soc Biol* 100 932.  
 COULTER, C B (1930-31) *J gen Physiol* 3, 513  
 COWIE, D M and CHAPIN W S (1907) *J med Res.*, 17 579.  
 CRAIGIE, J (1931) *Brit J exp Path* 12, 75  
 CRANDON J H, LUND C. C., and DILL, D B (1940) *New Eng med. J.*, 223, 353.  
 CRAW J A. (1905) *J Hyg., Camb* 5 113.  
 DAKIN H D and DALK, H. H (1919) *Biochem J.*, 13, 248  
 DALE, H H and HARTLEY P (1916) *Biochem J.*, 10 110  
 DANIELLI J F, DANIELLI, M., and MARRACK, J R. (1938) *Brit J exp Path* 19 395.  
 DASTYS, J (1907) *Ann. Inst. Pasteur* 16, 331  
 DEAN G (1907) *Proc roy Soc B* 79 399  
 DEAN H P (1910) *Z Immunforsch.*, 13, 84 (1917) *Lancet*, L 45 (1937) *J Path. Bact* 45 45  
 DEAN H. R., TAYLOR, G L. and ADAMS, M. E. (1935) *J Hyg., Camb.*, 35 69  
 DEAN H P and WEBB, P A. (1936) *J Path. Bact.*, 29 43  
 DESSLER, K. (1939) *Z Immunforsch.*, 73, 365.  
 DELVES E. (1937) *J infect Dis* 60 55.  
 DENYS, J and LECLEF J (1935) *La Cellule* 11, 177  
 DOWNIE, A W (1937) *J Path. Bact.*, 45, 149  
 DOZOIS, T F., SEITZER, S., and ECKER, E. E. (1943) *J Immunol.*, 47 215 (1944) *Ibid*

- DREYER, G and INMAN, A C (1917) *Lancet*, i 305
- DUNCAN, J T (1932a) *Brit J exp Path*, 13, 499, (1932b) *Ibid*, 13, 499, (1934) *Ibid*, 15, 23, (1933) *Ibid*, 16, 40, (1937) *Ibid*, 18, 108, (1938) *Ibid*, 19, 328
- EAGLY, H (1930) *J Immunol*, 18, 393, (1932) *Ibid*, 23, 153, (1936) *Ibid*, 30, 339, (1937) *Ibid*, 32, 119, (1938) *J exp Med*, 67, 490
- EAGLE, H and VICKERS P (1936) *J biol Chem*, 114, 193
- ECKER, E F and PILLEMER, L (1940) *J exp Med*, 71, 585, (1941) *J Immunol*, 40, 73, (1942) *Ann N Y Acad Sci*, 43, 63
- ECKER, E E, PILLEMER, L, and GRABILL F J (1938) *Proc Soc exp Biol, N Y*, 38, 318
- ECKER, E E, PILLEMER, L, and KUERN, A O (1942) *J Immunol*, 43, 245
- ECKER, F F, PILLEMER, L, MARTINSEN, E W, and WERTHEIMER, D (1938) *J biol Chem*, 123, 311
- ECKER, F E, PILLEMER, L, and SEIFTER, S (1913) *J Immunol*, 47, 181
- ECKER, F F, PILLEMER, L, WERTHEIMER, D, and GRADIN, H (1938) *J Immunol*, 34, 19
- ECKER, F F, WEISBERGER, A S, and PILLEMER, L (1942) *J Immunol*, 43, 227
- FABRIC, P. (1897) *Klin. Jb*, 6, 299, (1898) *Dtsch med. Wschr*, 24, 597, (1900) *Proc roy Soc, B*, 66, 424
- FISCHBERG, P and VOLK, R (1902) *Z Hyg Infektkr*, 40, 153
- EISLER, M (1920) *Zbl Bakt*, 84, 46
- ENDERS J F and SHAFFER, W F (1937) *J Immunol*, 32, 379
- ERICKSON, J O and NEURATH, H (1943) *J exp Med*, 78, 1
- ERLENMEYER, H and BERGER F (1932) *Biochem Z*, 252, 22, (1934) *Arch exp Path Pharmacol*, 177, 116
- FINGER TILLYNCKA R (1933) *Z Hyg Infektkr*, 114, 769
- FULER, H von, and BRUNER, E (1931) *Z Immunforsch*, 72, 65
- FALK, I S and MATSUDA, T (1926) *Proc Soc exp Biol, N Y*, 23, 781
- FELL, N, RODVY, G, and MARSHALL, D E (1913) *J Immunol*, 47, 237
- FELL, N, STERN, K G, and COCHILL, R D (1940) *J Immunol*, 39, 223
- FELLER, A F, ROBERT, L B, RALLI F P, and FRANCIS, T (1942) *J clin Invest*, 21, 121
- FELTON, L D (1926) *Bull Johns Hopk Hosp*, 38, 33, (1928) *J infect Dis*, 43, 543 (1932) *J Immunol*, 22, 453
- FELTON, L D and BAILEY, G H (1926) *J infect Dis*, 38, 131
- FERRATA, A (1907) *Berl Klin Wschr*, 44, 366
- FERRY, R M and LEVY, A H (1934) *J biol Chem*, 105, xxvii
- FOLLENSBY, F M and HOOKER, S B (1934) *J Immunol*, 37, 367
- FRANCE, W G (1930) *Coll Symp Monogr*, 7, 69 (See Marrack 1934)
- FRANKEL, M (1932) *Proc roy Soc, B*, 111, 165
- FRANKEL, M and OLITZKI, L (1930) *Nature, Lond*, 126, 723
- FREUND, J (1929) *Proc Soc exp Biol, N Y*, 28, 876
- FREUNDLICH, H (1906) *Z phys Chem*, 57, 385, (1922) "Kapillarchemie" Leipzig
- FREUNDLICH, H and NEUMANN, W (1907) *Z phys Chem*, 67, 538
- FRIEDBERGER, E and GOLDSCHMIDT, E (1910) *Z Immunforsch*, 6, 299
- GAY, F P (1905) *Zbl Bakt*, 29, 603
- GENGOU, O (1899) *Ann Inst Pasteur*, 13, 642, (1902) *Ibid*, 16, 731, (1911) *Z Immun Forsch*, 11, 143
- GERLOUGH, T D, PALMER J W and BLUMENTHAL, R R (1941) *J Immunol*, 40, 53
- GROSH, B N (1935) *Indian J med Res*, 23, 285
- GIBSON, R B and BANZHAF, E J (1910) *J exp Med*, 12, 411
- GIBSON, R B and COLLINS K R (1907) *J biol Chem*, 3, 233
- GLENNY, A T, POPE C G, and WADDINGTON, H (1925) *J Path Bact*, 28, 279
- GORBEL, W F, AVFRY, O T, and BARBERS F H (1934) *J exp Med*, 60, 599
- GOLDIE H and SANDOR, G (1937) *C R Soc Biol*, 126, 291
- GOLDSWORTHY, N E (1928) *J Path Bact*, 31, 290
- GOLDSWORTHY, N E and RUDD, G V (1935) *J Path Bact*, 40, 169
- GOODYER K and HORSFALL, T L (1936) *J exp Med*, 64, 201, (1937) *Ibid*, 66, 413 425, 437
- GOODYER, K, HORSFALL, F L, and BAUFER, J H (1938) *J Immunol*, 35, 439, 451
- GORDON, J (1937) *J Immunol*, 32, 375
- GORDON, J and ATRIN, W R (1938) *Brit J exp Path*, 19, 204, (1939) *J Path Bact*, 48, 477, (1941) *Brit J exp Path*, 22, 226
- GORDON, J and THOMPSON F C (1933a) *Brit J exp Path*, 14, 33, (1933b) *Ibid*, 14, 277, (1935) *Ibid*, 16, 101, (1936) *Ibid*, 17, 159, (1937) *Ibid*, 18, 390
- GORDON, J, WHITEHEAD, H R, and WORMALL, A (1926a) *Biochem J*, 20, 1028, (1926b) *Ibid*, 20, 1036

- GREENWOOD, M (1913) *Lancet*, 158
- GRUBER, M and DURHAM, H E (1896) *Munch med Wochr*, 43, 283
- HAMMARSTEN, O (1878) *Arch ges Physiol*, 17, 413
- HANKS J H (1940) *J Immunol*, 38, 159
- HARDE, E and THOMSON, A F (1933) *C R Acad. Sci*, 200, 1425
- HARINGTON, C R (1940) *J Chem Soc*, 119
- HARRIS, T and EAGLE, H (1935) *J. gen Physiol*, 19, 353.
- HART, R A. (1938) *J Immunol*, 34, 433.
- HARTLEY, P (1914) *Mem Dept Agric India* 4, 179, (1925) *Brit J exp Path.*, 6, 180  
(1931) *Med Res Coun Lond 'System of Bacteriology'* 6, 224
- HALPOWITZ, F (1937) *Klin Wochr*, 16, 257, (1940) *3rd int Congr Microbiol*, 819
- HAUROWITZ, F and BREINL, F (1933) *Hoppe Seyl Z*, 214, 111
- HAUROWITZ, F, KRAUS, F, and MARX, F (1936) *Z physiol. Chem.*, 245, 23
- HAUROWITZ, F, SARAFIAN, K, and SCHWERIN, P (1941) *J Immunol*, 40, 391
- HAUROWITZ, F and SCHWERIN, P (1943) *J Immunol*, 47, 111
- HAUROWITZ, F, TUNCA, M and SCHWERIN, P (1943) *Biochem. J.*, 37, 249
- HAUROWITZ, F and YENSON, M M (1943) *J. Immunol*, 47, 399
- HEALEY, H and PITFIELD, S (1935) *Brit. J. exp Path.*, 16, 535.
- HEGEDÜS, A and GREIFER, H (1938) *Z Immunforsch*, 92, 1
- HEIDELBERGER, M. (1933) *J Amer chem Soc.*, 60, 242, (1939) *Bact Rev*, 3, 49, (1941) *J exp Med*, 73, 681
- HEIDELBERGER, M. GRABAR, P, and TREFFERS, H P (1938) *J exp Med*, 63, 913
- HEIDELBERGER, M and HABAT, E. A. (1931) *J exp Med*, 60, 643, (1936) *Ibid.*, 63, 737
- HEIDELBERGER, M and KENDALL, F E. (1929) *J exp Med*, 50, 809, (1934) *Ibid*, 59, 519  
(1935a) *Ibid*, 61, 559, 563 (1935b) *Ibid*, 62, 467, (1935c) *Ibid*, 62, 697, (1936) *Ibid.*, 64, 161, (1937) *Ibid*, 65, 647
- HEIDELBERGER, M., KENDALL, F E, and TROELL, T (1936) *J exp Med*, 63, 819
- HEIDELBERGER, M. and LANDSTEINER, K (1923) *J exp Med*, 38, 561
- HEIDELBERGER, M. and MAYER, M (1942) *J exp Med*, 75, 285
- HEIDELBERGER, M and PEDERSEN, K O (1937) *J exp Med*, 85, 393.
- HEIDELBERGER, M., SIA, R H P, and KENDALL, F E. (1930) *J. exp Med.*, 52, 477
- HEIDELBERGER, M. and TREFFERS, H P (1941) *J gen Physiol*, 25, 523
- HEIDELBERGER, M., TREFFERS, H P., and MAYER, M (1940) *J exp Med.*, 71, 271
- HEIDELBERGER, M., WEIL, A J., and TREFFERS, H. P (1941) *J exp Med.*, 73, 695
- HEINICKE, A. (1934) *Z Immunforsch.*, 83, 245.
- HEKTOEN, L. (1908) *J infect. Dis*, 5, 249
- HEKTOEN, L. and BOOR, A H. (1931) *J infect Dis*, 43, 508
- HEKTOEN, L. and SCHULHOF, K (1923) *J. infect. Dis.*, 5, 249
- HERSHEY, A D (1940) *J Immunol*, 39, 383, (1941a) *Ibid*, 42, 455-455, (1941b) *Ibid.*, 42, 515, (1942) *Ibid*, 45, 39, (1943a) *Ibid*, 46, 249, (1943b) *Ibid*, 47, 77, (1944) *Ibid*, 48, 381
- HERSHEY, A D, KALMANSON, G, and BROVFEVERENYER, J (1943) *J Immunol*, 46, 267, 281
- HEUER, G (1922) *Z Hyg InfektKr.*, 95, 100
- HEWITT, L F (1938) *Biochem J.*, 32, 26, 1534
- HIGASHI, S (1922) *J Biochem, Tokyo*, 2, 315
- HOOKE, S B and BOYD W C. (1932) *J Immunol*, 23, 463, (1933) *Ibid.*, 24, 141, (1936) *Ibid*, 30, 33, 41 (1937) *Ibid*, 33, 337, (1940) *Ibid*, 38, 479, (1941) *Proc. Soc. exp Biol.*, 11, 47, 187, (1941b) *J Immunol*, 42, 419, (1942) *Ibid*, 45, 127
- HOPKINS, F G (1900) *J Physiol.*, 25, 306
- HOPKINS S J and WORMALL, A. (1933a) *Biochem J*, 27, 740, (1933b) *Ibid.*, 27, 1706.
- HORGAN, E. S (1936) *Nature, Lond*, 137, 872
- HORSFALL, F L and GOODNER, K. (1935) *J exp Med*, 62, 485, (1936a) *J Immunol*, 31, 135, (1936b) *J exp Med*, 64, 583
- HUDDLESON I F, JOHNSON, H W and HAMANN, E E (1933) *Amer J publ Hlth.*, 23, 917
- HUNTOON, F M. and ETRIS, S (1921) *J Immunol*, 6, 123
- HUNTOON, F M, MASUCCI, R., and HANUM, E. (1921) *J Immunol.*, 6, 185
- HYDE, R R (1923) *J Immunol*, 8, 267
- IWANOFF, K. (1936) *Z InfektKr*, 118, 197
- JACOBS J (1937) *J gen Physiol*, 20, 353
- JENNINGS, R K. and SMITH, L D (1942) *J Immunol.*, 45, 103.
- JOHNSON, F H. and DENNYSON, W L. (1944) *J Immunol.*, 48, 317
- JOHNSON, L R and WORMALL, A. (1932) *Biochem. J.*, 26, 1202
- JONES, C B and ECKER, E E (1940) *Proc Soc exp Biol*, 11, 44, 264

- JONES, D B and GERSDORFF, C E F (1923) *J biol Chem*, 56, 79
- JOOS, A (1901) *Z Hyg InfektKr*, 36, 422, (1902) *Ibid*, 40, 203
- KABAT, E A (1939) *J exp Med*, 69, 103
- KABAT, E A and HEIDELBERGER, M. (1937) *J exp Med*, 66, 229
- KAPNICK, I and COPE, O (1940) *Endocrinology*, 27, 543
- KALMANSON, G M and BROUVERENYER, J (1942) *Science*, 96, 21, (1943) *J Immunol* 47, 387
- KEKWICK, R A, MACFARLANE, M G KNIGHT, B C J G, and RECORD B R (1941) *Lancet*, 1, 571
- KEKWICK, R A and RECORD, B R (1941) *Brit J exp Path*, 22, 29
- KENPF, A H and NUGESTER W J (1942) *J infect Dis*, 71, 50
- KENDALL, F E (1942) *Ann N Y Acad Sci* 43, 85
- KLECZKOWSKI A (1941a) *Brit J exp Path*, 22, 44, (1941b) *Ibid*, 22, 188 192, (1943) *Biochem J*, 37, 30
- KLEIN, H (1907) *Johns Hopk Hosp Bull*, 18, 245
- KOCH, M L and SMITH, A H (1924) *Proc Soc exp Biol N Y* 21, 366
- KODICK, E and TRAU, B (1943) *Biochem J*, 37, 456
- KOSAKI, M (1918) *J Immunol*, 3, 109
- KRAUS, R (1897) *Wien klin Wochr*, 10, 736
- KREJCI, L E, JENNINGS, R K, and SMITH, L D (1942) *J Immunol* 45, 111
- KROGH, M VON (1911) *Z Hyg InfektKr*, 68, 201
- KRUIP, P H DE and NORTHERP, J H (1922-23) *J gen Physiol*, 5, 127
- LAIDLAW, P P and DUNKIN, G W (1931) *J comp Path*, 44, 1
- LANDSTEINER, K. (1917) *Z ImmunForsch*, 26, 122, (1919) *Biochem Z* 93, 108 (1920) *Ibid*, 104, 280, (1930) *Naturwissenschaften*, 18, No 29, 653, (1933) Die Spezifität der serologischen Reaktionen " Julius Springer, Berlin, (1936) The Specificity of Serological Reactions " Baltimore, (1942) *J exp Med*, 75 269
- LANDSTEINER, K and HEIDELBERGER M (1923) *J gen Physiol*, 6, 31
- LANDSTEINER, K. and JAGIO, N (1903) *Munch med Wochr* 50, 764
- LANDSTEINER, K and LAMPL, H (1917a) *Z ImmunForsch*, 26, 133 208 (1917b) *Biochem Z*, 86, 343
- LANDSTEINER K and PIRIE, N W (1937) *J Immunol* 33, 260
- LANDSTEINER, K and PRÁSEK, E (1911) *Z ImmunForsch* 10, 68 (1914) *Ibid* 20, 211
- LANDSTEINER, K and SCHREIBER, J VON DER (1928) *J exp Med*, 48, 315 (1929) *Ibid* 50, 407, (1931) *Ibid*, 54, 293, (1932a) *Ibid*, 55, 781, (1932b) *Ibid* 56, 399 (1934a) *Ibid* 59, 751, (1934b) *Ibid*, 59, 769, (1936) *Ibid*, 63, 310 (1938) *Ibid* 67, 709 (1939) *Ibid*, 69, 705, (1940) *Ibid* 71, 445
- LANDSTEINER, K and WELECKI, S (1911) *Z ImmunForsch*, 8, 393
- LEDINGHAM, J C G (1907) *J Hyg. Camb*, 7, 65
- LEISHMAN, W B (1902) *Brit med J*, 1, 73
- LEWIS, J H (1937) *J Amer med Ass* 108, 1336
- LEWIS, J H and WELLS H G (1925) *J biol Chem*, 66, 37
- LIBBY, R L (1938) *J Immunol*, 34, 269, 35, 289
- LIEPMAN, H (1909) *Munch med Wochr*, 56, 2097
- LIU, S C, CHOW B F, and LEE, K H (1937) *Clin J Physiol*, 11, 201
- LOCKE, A and HIRSCH, E F (1925) *J infect Dis*, 37, 449
- LOWELL, F C (1943) *J Immunol*, 46, 177
- MADSEN, T and STRENG O (1910) *Z phys Chem*, 70, 263
- MATLAND, H B and BURBURY, Y M. (1927) *J comp Path*, 40, 93
- MALKIEL, S and BOYD W C (1937) *J exp Med* 66, 383
- MALTANER, E (1935) *Proc Soc exp Biol, N Y*, 32, 1550
- MALTANER, F and MALTANER, E (1940) 3rd int Congr Microbiol, 781
- MAREL (1902) *Z Hyg InfektKr*, 39, 86
- MARRACK, J R (1934) *Spec Rep Ser med Res Coun Lond*, No 194, (1938) *Ibid*, No 230
- MARRACK, J R and DUFF D A (1938) *Brit J exp Path*, 19, 171
- MARRACK, J and HOLLERING, H F (1938) *Brit J exp Path* 19, 424
- MARRACK, J and SMITH, F C (1930) *Proc roy Soc, B*, 106, 1, (1931a) *Brit J exp Path*, 12, 30, (1931b) *Ibid*, 12, 182, (1932) *Ibid*, 13, 394
- MARSH, F (1936) *Nature Lond* 137, 618
- MARTIN, D S, ERICKSON, J O, PUTNAM, F W, and NEURATH H (1943) *J gen Physiol*, 26, 533
- MELLANBY, J (1908) *Proc roy Soc, B* 80, 399
- MEYNES, I (1897) *Z Hyg InfektKr*, 25, 413
- MEYER, K. (1936) *Ann Inst Pasteur*, 56, 684
- MICHAELIS, L. (1911) *Dtsch med Wochr*, 37, 969
- MILES, A A (1933) *Brit J exp Path*, 14, 43, (1939) *Ibid*, 20, 63.

- MILES, A A and PIRIE N W (1939) *Brit J exp Path.*, 20, 109
- MOORE, D H VAN DER SCHEER, J, and WYCKOFF, R W G (1940) *J Immunol.*, 38, 221
- MORIYAMA H (1937) *J Shanghai Sci Inst Sect IV*, 2, 279
- MORGAN, W T J and PARTRIDGE S W (1941) *Biochem J.*, 35, 1140
- MORRIS, M C. (1940) *J Immunol.*, 39, 369
- MUDD, S and ANDERSON, T F (1941) *J Immunol.*, 42, 251
- MUDD S, HEINMETS, F., and ANDERSON, T F (1943) *J exp Med.*, 78, 327
- MUDD, S and JOFFE, E W (1933) *J gen Physiol.*, 18, 947
- MUDD, S, LUCKE, B, McCUTCHEON M., and STARCIA, M. (1929) *J exp Med.*, 49, 779
- MUIR, R (1903) *Lancet*, ii 100, 446, (1909) 'Studies on Immunity' London
- MUIR, R and BROWNING, C. H (1909) *J Path. Bact.*, 13, 78
- MUIR, R and MARTIN, W B M (1906a) *Brit med J.*, ii, 1783, (1906b) *J Hyg., Camb.*, 6, 265
- MUTSAERS W and GRÉGOIRE, P F (1936) *C R Soc Biol.*, 123, 144
- NEISSER, M and WECHSBERG, F (1901) *Munch med Wochr.*, 48, 697
- NEUBERGER, A and LILL, M E. (1940) *Biochem J.*, 34, 109
- NEUFELD F (1902) *Z Hyg Infekthkr.*, 40, 54
- NEUFELD F and ETINGER TULZINSKA, R (1909) *Zbl Bakt.*, 114, 202, (1931) *Z Hyg Infekthkr.* 112, 492
- NEUFELD, I and HÖVE. (1907) *Arb Reichsgesundh.Amt.*, 25, 164
- NEUFELD F and RIMPAU, R (1904) *Dtsch med Wochr.*, 11, 1458 (1905) *Z Hyg Infekthkr.*, 51, 283
- NEURATH, H (1939) *J Amer chem Soc.*, 61, 1841
- NEURATH, H. and SAUM, A M (1939) *J Biol Chem.*, 128, 347
- NICOLLE, M and CÉSARI E. (1922) *Ann Inst Pasteur* 38, 463
- NORTHROP J H (1922) *J gen Physiol.*, 4, 629, (1928) 'The Newer knowledge of Bacteriology and Immunology' Jordan & Fall, Chicago, p 782, (1941) *J gen Physiol.*, 25, 463
- NORTHROP, J H and KRUIJ, P H DE. (1922a) *J gen Physiol.*, 4, 639, (1922b) *Ibid.*, 4, 655
- NOBY P L DE and HAMON V (1935) *C R Acad Sci.*, 200, 1200, (1936) *Ann Inst Pasteur* 58, 309
- NUTTALL, G (1888) *Z Hyg Infekthkr.*, 4, 303
- NUTTALL, G H F (1904) *Blood Immunity and Blood Relationships* Cambridge
- OBERMAYER, F and PICK, E P (1900) *Wien Lin Wochr.*, 19, 327
- OLITZKI, L (1932) *Z Immunforsch* 76, 206
- OLITZKI L and FRANKEL, M (1931) *Proc Soc exp Biol.*, 11, 28, 492
- OTTOLENGHI D and MORI V (1905) *Zbl Bakt.*, 33, 338 463
- PAPPENHEIMER, A M (1940) *J exp Med.*, 71, 263, (1942) *J Bact.*, 43, 273
- PAPPENHEIMER, A M LUNDGREN H P, and WILLIAMS, J W (1940) *J exp Med.*, 71, 247
- PAPPENHEIMER, A M and ROBINSON, E S (1937) *J Immunol.*, 32, 291
- PAULING L (1940) *J Amer chem Soc.*, 62, 643
- PAULING L CAMPBELL, D H, and PRESSMAN, D (1941) *Proc nat Acad Sci Wash.*, 27, 120 (1943) *Physiol Rev* 23, 203
- PAULING L and NIEMANN C (1939) *J Amer chem. Soc.* 61, 1860
- PAULING L PRESSMAN D CAMPBELL, D H, and IKEDA, C. (1942) *J Amer chem. Soc.*, 64, 3003
- PAULING L PRESSMAN D and IKEDA C (1942) *J Amer chem. Soc.*, 64 3010.
- PENNELL P B and HUDDLESON I F (1938) *J exp Med.*, 68, 73, 83
- PETERMANN M L (1941) *J Biol Chem.*, 144, 607
- PFEIFFER, R (1893) *Z Hyg Infekthkr.*, 11, 393, (1894a) *Ibid.*, 16, 268, (1894b) *Ibid.*, 18, 1 (1895) *Ibid.*, 19, 70
- PFEIFFER, R and ISSAIEFF (1894) *Z Hyg Infekthkr.*, 17, 300
- PICK E P (1912) *Kolle und Wassermanns 'Hdb d path Mikroorg'*, IIte Aufl., 1, 680
- PILGER A (1938) *J Path Bact.*, 47, 1, (1941a) *Ibid* 53, 431 (1941b) *J Bact.*, 42, 390.
- PILLEMER, L CHU, F SEIFTER, S, and ECKER, E E (1942) *J Immunol* 45, 51
- PILLEMER, L and ECKER, E E (1941a) *J Immunol* 40 101 (1941b) *Science* 94, 437
- PILLEMER L ECKER, E E, ONCLEY, J L, and COHN, E J (1941) *J exp Med.*, 74, 297
- PILLEMER, L SEIFTER, J and ECKER, F E (1941) *J Immunol.*, 40, 89, (1942) *J exp Med* 75, 421
- PILLEMER, L SEIFTER S, SAN CLEMENTE C L, and ECKER, E E. (1943) *J Immunol.*, 47, 200
- PITTMAN M (1943) *Publ Hlth Rep.* Wash 58, 139
- PLATT A E (1936) *Aust J exp Biol med Sci* 14, 101, (1938) *Ibid.*, 16, 275
- POCHOV J (1936) *C P Soc Biol* 121, 387
- POPE, C G (1939a) *Brit. J exp Path.*, 20, 132, (1939b) *Ibid.*, 20, 201

- POPE, C G and HEALEY, M. (1938) *Brit J exp Path.*, **19**, 397
- PORGES, O (1908) *Zbl Bakt.*, **40**, 133
- PORGES, O and PRANTSCHOFF, A (1908) *Zbl Bakt.*, **41**, 466, 546, 658
- POTTER, E (1921) *Z ImmunForsch.*, **32**, 538
- RACE, R R (1914) *Nature Lond.*, **153**, 771.
- RAFFEL, S, PAIT, C F, and TERRY, M C (1940) *J Immunol.*, **39**, 317
- RAMON, G (1922) *C R Soc Biol.*, **86**, 661, 711, 813
- REIVER, L and FISCHER, O (1929) *Z ImmunForsch.*, **61**, 317
- REIVER, L and KOPP, H (1929) *Z ImmunForsch.*, **61**, 397
- RICE, C E (1943) *J Immunol.*, **46**, 427
- RICE, C E and SICKLES, G R (1942) *J Immunol.*, **43**, 319
- RIMINGTON, C and ENDE, M VAN DEN (1910) *Biochem J.*, **34**, 941
- ROSENHEIM, A H (1935) *J Path Bact.*, **40**, 75, (1937) *Biochem J.*, **31**, 54
- ROSS, V (1938) *J Immunol.*, **35**, 351, 371
- ROTHEN, A (1941) *J gen Phys.*, **25**, 487
- SCHER, J VAN DER, LAGSDIN, J B, and WYCKOFF, R W G (1941) *J Immunol.*, **41**, 209
- SCHER, J VAN DER WYCKOFF, R W G, and CLARKE, F H (1940) *J Immunol.*, **39**, 65  
(1941a) *Ibid.*, **40**, 39, (1941b) *Ibid.*, **40**, 173, (1941c) *Ibid.*, **41**, 349
- SCHMIDT, S (1930) *C R Soc Biol.*, **103**, 101
- SEIFTER, S, DOZORS, T F, and ECKER, E E (1944) *J Immunol.*, **49**, 45
- SEIFTER, S, PILLEMER, L, and ECKER, E E (1943) *J Immunol.*, **47**, 195
- SEVAG, M G and SKATONE, V (1934) *Z ImmunForsch.*, **83**, 464
- SOALITZER, M (1914) *Z Hyg InfektKr.*, **76**, 209
- SHAHROKH, A B (1943) *J biol Chem.*, **151**, 659
- SHIRLEY, G S (1926) *J exp Med.*, **44**, 667 (1929) *Ibid.*, **50**, 825
- SHRICLEY, E W and IRWIN, M R (1937) *J Immunol.*, **32**, 281
- SINGER, E (1942) *Ann J exp Biol med Sci.*, **20**, 209
- SKWIRSKY, P (1910) *Z ImmunForsch.*, **5**, 538
- SLEESWIJK, J G (1908) *Zbl Bakt.*, **46**, 513
- SMITH, F C and MARRACK, J (1930) *Brit J exp Path.*, **11**, 494
- SMITH, W (1932) *J Path Bact.*, **35**, 509
- SNAPPE, I and GRUNBAUM, A (1936) *Brit J exp Path.*, **17**, 361
- SOBOTKA, H and FRIEDLANDER, M (1928) *J exp Med.*, **47**, 57
- SPÄT, W (1910) *Z ImmunForsch.*, **7**, 712
- SPINK, W W, AGNEW, S, and MICKELSEN, O (1942) *J Immunol.*, **44**, 289, 297
- SPINK, W W, AGNEW, S, MICKELSEN, O, and DAHL, L M (1942) *J Immunol.*, **44**, 303
- SPOONER, F T C and BAWDEN, F C (1935) *Brit J exp Path.*, **16**, 218
- STENHAGEN, E. (1938) *Biochem. J.*, **32**, 714
- STORINGER, H E and HEIDELBERGER, M (1937) *J exp Med.*, **66**, 201
- STRENG, O (1909) *Z Hyg InfektKr.*, **62**, 281
- STRONG, P. S and CULBERTSON, J T (1934) *J Hyg. Camb.*, **34**, 522
- STULL, A and HAMPTON, S F (1911) *J Immunol.*, **41**, 143.
- SVEDBERG, T (1937) *Nature, Lond.*, **139**, 1001, (1939) *Proc roy Soc B.*, **127**, 1
- SYNOR, R L M (1943) *Chem. Rev.*, **32**, 135
- TAKANO, Y (1930) *Z ImmunForsch.*, **67**, 29
- TAYLOR, G L (1931) *J Hyg, Camb.*, **31**, 56, (1933) *Ibid.*, **33**, 12
- TAYLOR, G L, ADAIR, G S, and ADAIR, M E (1932) *J Hyg, Camb.*, **32**, 310
- TELIELIUS, A (1937a) *Biochem J.*, **31**, 1464, (1937b) *J exp Med.*, **65**, 641
- TELIELIUS, A and KABAT, E A (1939) *J exp Med.*, **69**, 119
- TOKUNAGA, H (1929) *Zbl Bakt.*, **107**, 283
- TOPLEY, W W C (1915) *Proc roy Soc. B.*, **88**, 396
- TOPLEY, W W C, WILSON, J, and DUNGAN, J T (1935) *Brit J exp Path.*, **16**, 116
- TREFFERS, H P and HEIDELBERGER, M (1941a) *J exp Med.*, **73**, 120, (1941b) *Ibid.*, **73**, 293
- UENO, S (1938) *Jap J med Sci*, **11**, *Soc Med Hyg*, **2**, 201, 225
- WADSWORTH, A, MALTANER, F, and MALTANER, E (1937) *J Immunol.*, **33**, 297
- WARD, H K and FIDERS, J F (1933) *J exp Med.*, **57**, 527
- WASSERMAN, P, BROTHMAN, R H and MURSKY, I A (1940) *J Immunol.*, **38**, 213
- WEHMEYER, P (1941) *Z ImmunForsch.*, **100**, 179
- WEIL, A J, PARFENTJEV, I A, and BOWMAN, K L (1918) *J Immunol.*, **35**, 399
- WELLS, H G (1915) *J infect Dis.*, **16**, 209, (1920) 'The Chemical Aspects of Immunity' New York
- New York, (1929) 'The Chemical Aspects of Immunity' 2nd Edit New York
- WELLS, H G, LEWIS, J H, and JONES, D B (1927) *J infect Dis.*, **40**, 326
- WELLS, H G and OSBORNE, T B (1911) *J infect Dis.*, **8**, 66
- WHITEHEAD, H R, GORDON, J, and WORKMALL, A (1925) *Biochem J.*, **19**, 618.
- WIEFNER, A S and HERMAN, M (1939) *J Immunol.*, **36**, 205
- WINSLOW, C E A, FALK, I S, and CAULFIELD, M F (1923) *J gen. Physiol.*, **6**, 177

- WOOLF, B. (1941) *Proc roy Soc., B.*, 120, 60  
WOOLF, B., MAREACK, J. R., and DOWNIE, A. W. (1936) *Chem Ind.*, 14, 156.  
WORMALL, A. (1930) *J exp Med*, 51, 295  
WRIGHT, A. E. (1909) "Studies in Immunization" London.  
WRIGHT, A. E. and DOUGLAS, S. R. (1903) *Proc. roy Soc., B.*, 72, 364, (1904) *Ibid.*, 73, 156  
WRIGHT, G. G. (1944) *J exp Med*, 79, 455  
WU, H. and LING, S. M. (1927) *Chin J. Physiol.*, 1, 431  
YOUNG, G. P. and COLWELL, C. A. (1910) *J. infect Dis.*, 68, 235, 67, 48, (1913) *J Immunol.*, 46, 217  
ZILIO S. (1919) *Biochem J.*, 13, 172  
ZENESER, H. (1921) *J Immunol.*, 6, 289. (1930) *Ibid.*, 18, 483



## CHAPTER 8

### THE ANTIGENIC STRUCTURE OF BACTERIA

A COMPLETE description of the antigenic structure of a bacterial cell would include (a) the number and kind of different antigens present, (b) their relative proportions, and (c) their position in the cell or cell appendages

Though in no instance has a complete description of this kind been achieved, yet for certain bacterial species we can construct working models of the antigenic structure, which, though crude, have at least enabled us to solve a number of immunological puzzles, and it seems certain that further advance along the same lines will enable us to solve many more

We may start our discussion by considering the methods that are available for studies of this kind

#### Antigenic Analysis by Selective Qualitative Absorption of Antibodies

The fact that a single bacterial cell contains many antigens was established during the earliest studies on the agglutination reaction. The original observations of Gruber and Durham (1896) showed that the specificity of this reaction was not peculiar to each species of bacterium. For instance, while the colon bacillus was sharply differentiated from the typhoid bacillus and both from certain vibrios these vibrios were not so clearly differentiated from one another. Durham (1901) as the result of a more detailed study of these group agglutinations enunciated quite clearly the hypothesis of a multiplicity—as he called it, a *mosaic*—of antigens within a single bacterial cell. Employing small letters to denote the antigenic bacterial components, and capitals to denote the corresponding agglutinins in the antisera, he suggested that the actual agglutinins and agglutinogens involved might be represented as follows

	Agglutinogens		Agglutinins
Bacterium 1	a, b, c, d, e	Serum 1	A, B, C, D, E
Bacterium 2	c, d, e, f, g, h	Serum 2	C, D, E, F, G, H
Bacterium 3	e, f, g, h, j, k	Serum 3	E, F, G, H, J, K

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- WOOLF, B. (1941) *Proc. roy. Soc., B*, 130, 60.  
WOOLF, B., MARRACK, J. R., and DOWDIE, A. W. (1936) *Chem. Ind.*, 14, 156.  
WORMALL, A. (1930) *J. exp. Med.*, 51, 295.  
WRIGHT, A. E. (1909) "Studies in Immunization." London.  
WRIGHT, A. E. and DOUGLAS, S. R. (1903) *Proc. roy. Soc., B*, 72, 364; (1904) *Ibid.*, 73, 176.  
WRIGHT, G. G. (1944) *J. exp. Med.*, 79, 455.  
WU, H. and LING, S. M. (1927) *Chin. J. Physiol.*, 1, 431.  
YOUNG, G. P. and COLWELL, C. A. (1940) *J. infect. Dis.*, 68, 235; 67, 43; (1943) *J. Immunol.*, 48, 217.  
ZILBO S. (1919) *Biochem. J.*, 13, 172.  
ZINSSER, H. (1921) *J. Immunol.*, 6, 280; (1930) *Ibid.*, 18, 493.

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- WOOLF, B (1941) *Proc. roy Soc., B.*, 130, 60  
 WOOLF, B, MARRACK, J R., and DOWDIE, A W (1936) *Chem. Ind.*, 14, 156.  
 WORMALL, A. (1930) *J exp Med.*, 51, 295  
 WRIGHT, A E. (1909) "Studies in Immunization." London.  
 WRIGHT, A. E. and DOUGLAS, S R (1903) *Proc. roy Soc., B*, 72, 364, (1904) *Ibid.*, 73, 156  
 WRIGHT, G G (1944) *J exp Med.*, 79, 455.  
 WU, H and LING, S M (1927) *Chin J Physiol.*, 1, 431  
 YOUNG, G P and COLWELL, C. A. (1940) *J infect Dis.*, 63, 230, 67, 49, (1943) *J Immunol* 48, 217  
 ZILIO S (1919) *Biochem J.*, 13, 172.  
 ZINSSER, H (1921) *J Immunol.*, 6, 289, (1930) *Ibid.*, 18, 493

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haemolytic antibodies When an agglutinating serum is exposed to an excess of bacterial cells that contain some or all of the corresponding agglutinogens the agglutinins that find their counterparts in the antigenic structure of the bacteria are bound, or absorbed, and thus removed from the fluid in which the bacteria are suspended If the bacteria and the antibodies adsorbed to them are separated by centrifugation, the supernatant fluid can be tested as regards its remaining agglutinins For instance, taking the illustrative example given above, if Serum 1 were absorbed with an excess of Bacterium 1 all the agglutinins A, B, C, D, and E would be removed and the supernatant fluid would have no agglutinating action. If it were absorbed with Bacterium 2, C, D and E would be removed, but A and B would remain The supernatant fluid would not agglutinate Bacterium 2—the absorbing strain—nor Bacterium 3 (since that bacterium does not contain agglutinogens *a* or *b*), but it would still agglutinate Bacterium 1 in virtue of the remaining agglutinins A and B If it were absorbed with Bacterium 3, the single common agglutinin E would be removed The supernatant fluid would (as always) fail to agglutinate the absorbing strain, but it would still agglutinate Bacterium 1 in virtue of agglutinins A, B, C, and D, and Bacterium 2 in virtue of agglutinins C and D

A note may be interpolated here with regard to two terms that are sometimes loosely applied. The antiserum that is produced by the inoculation into a suitable animal of a particular bacterium is frequently referred to as a *homologous* serum A serum that agglutinates the same bacterium but has been produced by the inoculation of some other bacterium, differing in one or more of its antigenic components, is termed a *heterologous* serum Used in this sense the terms are useful and logical, and may be applied either to an antiserum in relation to a bacterium, or to two antisera or two bacteria in relation to one another, implying in either case complete correspondence between active combining groups. Thus, a serum that contains the active antibody groupings A, B, C, D is homologous with a bacterium containing the active antigenic groupings *a, b, c, d*, two bacteria each containing the active antigenic groupings *d, e, f, g* are homologous with one another, and so on. But an antiserum containing the active groupings A, B, C, D is not homologous, either with a bacterium containing the active groupings *a, b, c, e*, or *a, b, c, d, e*, or *a, b, c* The terms cannot, logically, be applied to single antigens or single antibodies. They describe the relation between a group of different antibodies in a serum and a group of antigens attached to a bacterial cell, or between the group of antigens attached to one bacterial cell and the group attached to another We are also, it should be noted, using the terms qualitatively, not quantitatively

Suppose that we have an unknown bacterium *x*, which we suspect to be antigenically homologous with a known bacterium *y*, and suppose that we have available an anti *y* serum. We absorb the anti *y* serum with *x* and then test it against *y* If it now fails to agglutinate *y* we may assume that all the active antigenic groupings in *y* are also present in *x* But we have not excluded the possibility that *x* has additional active antigenic groupings, not present in *y* So we prepare an anti *x* serum, and then absorb it with *y* If, after absorption, it fails to agglutinate *x*, we assume that all the active antigenic groupings present in *x* are also present in *y* That is, *x* and *y* are antigenically homologous. This double cross-absorption method is often spoken of as the "murder test."

The results of agglutination and adsorption tests with related strains of bacteria and their respective antisera, when interpreted in the light of these principles, have in many cases provided detailed pictures of the antigenic make-up of certain bacteria. These pictures are for the most part qualitative, though it is usually possible to guess roughly which of the antigens predominate The validity of these qualitative interpretations depends on the assumption that the separate antigenic components react only with

corresponding antibodies in the serum and that the various antibodies in the serum reflect the various antigens in the bacterium.

We have seen on p. 251 that certain complex antigens appear to possess more than one kind of determinant, and stimulate the production of an antibody with two or more specificities (see Burnet 1931, Heidelberger and Kendall 1934, Morgan 1936a, Meyer 1938, 1939, Miles 1939). If then we postulate a bacterium  $x$  with an antigenic complex  $ab$  and a second bacterium  $y$  with an antigen  $a$ , the results of absorption tests would indicate that  $x$  and  $y$  were antigenically identical since  $ab$  would remove all A agglutinins from the anti- $y$  serum and  $a$  would remove the AB agglutinins from the anti- $x$  serum. Indeed the only way we should ever discover the existence of the complex  $ab$  would be in comparison with two other bacterial species which happen respectively to possess  $a$  only and  $b$  only. If on the other hand we postulate that the response to bacterium  $x$  (with the complex  $ab$ ) consists of separate antibodies A and B then absorption tests would lead us to postulate that  $x$  and  $y$  had an antigen in common and that  $x$  had in addition a distinct antigen peculiar to itself. The extraction and separation of the various antigens provides a way out of this impasse. The different antibodies may then be removed from the bacterial antisera in specific precipitates and the precise specificity of the remaining antibodies for the other antigens determined.

### Quantitative Techniques in Antigenic Analysis.

The methods of antigenic analysis discussed above yield qualitative answers. Estimation of the relative amounts of the various antigens is possible only if certain assumptions are made. These are that there is a quantitative correspondence between the different kinds of antibodies in a serum and the different kinds of antigens present in the homologous bacterium; that all the antigens have the same antigenic capacity, weight for weight; and that each of the antigens reacts with its corresponding antibody with the same degree of intensity. None of these assumptions is warranted and they are indeed seldom made explicitly though they are sometimes implicit in the inferences drawn from analytical data. It will be clear from Chapter 7 that the last two assumptions are unjustified. The first requires a little amplification.

If a bacterium  $y$  removes half the antibody from anti- $x$  serum the fact that a large amount of antibody peculiar to  $x$  remains is sometimes interpreted as indicating a large amount of corresponding antigen in  $x$ . But as Miles (1939) pointed out, the antibody response to many repeated doses of a small amount of antigen may be as great as that to many doses of a large amount so that after prolonged immunization with a bacterium containing a major and a minor antigen the corresponding antibodies may be in equal concentration in the serum. Removal of the antibody to the major antigen would in this case leave a large amount of antibody representing what was in fact a minor antigen.

Nevertheless though quantitative correspondence of antibody and antigens is unlikely ever to occur, it is clearly important to know how much of the various antigens and antibodies are contained in the two reagents: a bacterial suspension and an antiserum at our disposal. A knowledge of the antibody concentrations is particularly useful. For example the quantitative relationship in antigen-antibody reactions described in Chapter 7 permits the measurement of the concentration of one antigen in a mixture of antigens derived from a bacterium.

Heidelberger and his colleagues (see Heidelberger and Kabat 1934, 1936, 1937, 1938, Alexander and Heidelberger 1910, Henriksen and Heidelberger 1941) have measured specific agglutinin content in terms of maximum antibody nitrogen adsorbed by known doses of bacterial suspensions. The data so obtained permit

comparison of bacterial strains in terms of the proportion of total antibody they will absorb from a specific antiserum. Duncan (1932a, b) and Miles (1933 1939) attacked the problem by means of the optimal proportions technique. The equivalence point between a bacterial suspension and an antiserum may be the resultant of the reaction of a number of different antigens and their corresponding antibodies which may to some extent be distinguished by appropriate variations of the experimental conditions.

This leads us to the problem of differentiating two types of bacteria, both of which contain the same antigenic components, but in different proportions. Using the qualitative differential absorption technique, with maximal absorbing doses of bacteria, the two types would appear identical since each would absorb all the agglutinins from both the homologous and heterologous serum using those terms in this case to include quantitative as well as qualitative relations. By careful adjustment of the absorbing dose it is, however, sometimes possible to demonstrate a quantitative difference of this kind. An example dealing with the differentiation of *Br melitensis* from *Br abortus*, will be found on p. 824

### The Study of Bacterial Variation as a Guide to Antigenic Structure

Bacterial variation as a general phenomenon, is dealt with in the succeeding chapter but the detailed study of the antigenic differences displayed by bacterial variants has played so large a part in the formation of our present views in regard to antigenic structure that we must anticipate a little and discuss in the present section certain examples that will help to illustrate the kind of antigenic changes that occur

Smith and Reagh (1903) isolated a non motile variant of the hog cholera bacillus and compared its agglutination reactions with those of the normal motile, flagellated type of this organism. They found that a serum prepared against the motile strain agglutinated both the motile and the non motile strains, but the titre was much higher for the motile than for the non motile, and the clumps formed by the motile strain were fluffy and formed rapidly, while the clumps formed by the non motile strain were tight, small and granular, and formed far more slowly. A serum prepared against the non motile strain agglutinated the motile and non motile strains to the same degree, giving in each case the slow, granular type of agglutination. The serum prepared against the motile strain, when absorbed with the non motile strain, lost its power of agglutinating the non motile bacilli but retained its power of agglutinating the motile strain. The serum prepared against the non motile strain lost its agglutinating power for both strains when absorbed with the motile strain. Smith and Reagh concluded that the normal motile hog cholera bacillus has two kinds of antigens one contained in the flagella the other in the cell body. The non motile type has lost the flagellar antigens and retains the body antigens only. Beyer and Reagh (1904) extended these observations and showed that the flagellar antigens were so altered by heating to 70° C for 15 minutes that the heated bacilli no longer gave the flagellar type of agglutination, but reacted like the non motile strain. The somatic, or body, antigens were not affected by this treatment and still gave the characteristic slow, granular agglutination (see also Orcutt 1924, Craigie 1931)

These observations of Smith and Reagh and of Beyer and Reagh demonstrated all the essential points of difference between the flagellar and somatic types of agglutination, but they passed almost unnoticed until similar findings were recorded by Weil and Felix (1917) in connection with their work on the diagnostic significance of the agglutination of a particular strain of proteus bacillus by the blood of a patient



suffering from typhus fever. The proteus bacillus, in its normal, flagellated form, grows on the surface of nutrient agar as a thin spreading film which resembles the mist produced by breathing on glass, and was named the *Hauch* form—the 'breath' form, by Weil and Felix. A non flagellated variant grew in isolated colonies with no thin spreading growth between them, this was the *Ohne Hauch* form—the form without an exhalation. As is the way in laboratory shorthand these soon became the 'H' and 'O' forms, and so the thermolabile, flagellar antigens are now the H antigens, and the thermostable, somatic antigens are the O antigens.

Arkwright (1920, 1921, 1924) working with bacteria of the typhoid paratyphoid dysentery group described a type of variation that has proved particularly instructive from this point of view. He noted that a particular type of variant was of relatively common occurrence among these bacilli, and this variant differed from the parent form in several characteristic ways. Thus, the normal parent form gave smooth colonies on a solid medium and a diffuse growth in broth and was not auto agglutinable in normal saline (0.85 per cent). The variant form gave rough or granular colonies on solid media and a granular growth in broth and was auto agglutinable in normal saline, though a stable suspension could usually be prepared in distilled water or in a saline solution with a greatly decreased salt content (0.2–0.4 per cent). These differences in colony form, growth in broth and salt sensitiveness were associated with a profound change in antigenic structure. The normal parent form reacted specifically as regards its agglutinability by immune sera, the variant was agglutinated not only by its own antiserum but by antisera prepared against many other bacteria in the rough state, some of which were only distantly related to the parent form according to the ordinarily accepted bacteriological criteria. By the usual transition—through shorthand to symbols—the normal form, giving smooth colonies, became the *Smooth* (or S) form, the variant giving rough colonies, became the *Rough* (or R) form. And here again we have become entangled in the web of our words for we have never defined exactly the criteria of roughness and smoothness. The implicit meaning is certainly not the meaning we want, since, as we shall see, the correlation between colonial form and antigenic structure is by no means constant. If we give *Smooth* and *Rough* their obvious or commonsense meaning then we shall want new terms for the underlying changes that really interest us. If we employ the legitimate licence of scientific terminology, and say that by *Smooth* we mean one sort of antigenic structure and by *Rough* another, then we must at least define quite clearly what we mean. Up to the present we have evaded our difficulties and so we find that the rough type of one species is recorded as having the more important characters of the smooth type of another species, as in the case of the anthrax bacillus (see Preisz 1904, Eisenberg 1912, Bordet and Renaux 1930, Tomcsik and Szongott 1932, 1933) or it becomes necessary to invent new descriptive terms, such as *matte* and *glossy*, as in the case of the hæmolytic streptococcus (see Todd 1928a, b, Todd and Lancefield 1928). Probably the most convenient convention is to use the symbols S and R to distinguish the antigenic variation and retain, as far as is necessary, the words *Smooth* and *Rough* as descriptions of colonial variants. There will thus be no contradiction in asserting that the S form of *B. anthracis* grows as a rough colony.

In most of the cases with which we are familiar the change from S form to R form (the S → R variation) is associated with a loss of virulence—an association of obvious importance to the immunologist—as well as with a particular kind

of change in antigenic structure. We have very good reasons for believing that the change in antigenic structure determines the loss of virulence, and it certainly determines the change in antigenic value of the variant strain regarded as an immunizing agent. We may, then, for our immediate purpose and to avoid confusion, explicitly limit ourselves to this antigenic variation and its consequences, noting that if we define the  $S \rightarrow R$  variation in this way we are acting in defiance of general usage.

In terms of antigenic structure we may say that this variation consists in the *loss of the heat stable somatic antigen that characterizes the surface of the normal virulent bacterial cell*. This loss may be associated with an uncovering of some other somatic antigen, which then dominates the antigenic behaviour of the strain.

Variation of this type has now been described in a wide range of different bacterial species—in the *Pasteurella* group (de Kruif 1921, Webster 1925), in pneumococci (Griffith 1923, Reimann 1925, 1927), in staphylococci (Bigger, Boland and O'Meara 1927) in streptococci (Todd 1923a) and in a host of other organisms. We are probably justified in regarding it as a type of variation to which pathogenic bacteria are inherently liable.

Sometimes (see White 1932, 1933, Henderson 1939) this variation may proceed still further, and another antigenic constituent may be lost. The antigenic behaviour of the organism may then be dominated by a component that, in the normal state, was altogether latent. It may be noted (a) that the  $S \rightarrow R$  variation is quite independent of the  $H \rightarrow O$  variation (rough variants are often flagellated), and (b) that loss variations of the  $S \rightarrow R$  or more deeply seated types are often irreversible.

There is another kind of antigenic variation to which reference must be made before we pass to the next section of our discussion.

Andrewes (1922, 1925) described a curious *phasic variation* in the H antigens of certain flagellated species. Taking two bacteria,  $x$  and  $y$ , that showed the flagellar type of agglutination when tested either against an anti- $x$  or against an anti- $y$  serum, and therefore possessed at least one H antigen in common, he absorbed the anti- $x$  serum with bacillus  $y$  and thus obtained a serum that agglutinated  $x$  but not  $y$ . He then took a broth culture of  $x$  and plated it on a solid medium, thus obtaining separate colonies. Subculturing from several of these, he obtained different cultures, each representing a single bacillus in the original broth culture. When he tested these different cultures against the anti- $x$  serum, rendered specific by absorption, and against the anti- $y$  serum, containing the common, or "group" antibody, he found that his subcultures fell sharply into two classes. Those of one class were agglutinated to titre by the specific anti- $x$  serum, but not at all by the anti- $y$  serum. Those of the other were agglutinated to titre by the anti- $y$  serum, but not at all by the specific anti- $x$  serum. The only possible conclusion would seem to be that the original culture of  $x$  showed cross agglutination with the anti- $y$  serum not because each bacillus in the culture possessed two antigenic components, say  $a$  and  $b$ , in their flagella,  $a$  being specific for  $x$  and  $b$  being shared by  $y$ , but because some of the bacilli possessed  $a$  alone, and others  $b$  alone. We need not, for the moment, worry as to whether the  $a$  bacilli do or do not possess a trace of  $b$  and vice versa. Similarly Andrewes was able to show that the  $y$  culture, which agglutinated with anti- $x$  and anti- $y$  sera, contained some bacilli possessing the common flagellar antigen  $b$  and others containing an antigen  $c$  specific for  $y$ . The bacilli that possessed the group antigen only were referred to as being in the *group*

chemical fractionation, namely that the treatment used may partly but fundamentally alter the substance we are looking for. The pneumococcal polysaccharide as originally isolated was not antigenic. By less chemically active fractionation, an acetylated form of the substance is produced, which is immunologically far more significant (Avery and Goebel 1933).

Lancefield (1926) separated antigenic components from hemolytic streptococci by chemical methods. She found that the component that confers type-specificity is acid soluble and contains some 14 per cent. of protein nitrogen. In its purified state it is hapten like, in that it gives specific precipitation in the test tube but fails to stimulate antibody production *in vivo*. In addition to this type-specific antigen there is a polysaccharide component that is shared by many types of hemolytic streptococci, but differentiates the species into large sub-groups (Lancefield 1933), and there is a nucleo-protein antigen that is shared by a wide variety of streptococci, hemolytic and non-hemolytic. Studies by Todd and Lancefield (1928) (see also Lancefield and Todd 1928) have shown that the change from the virulent matt to the avirulent glossy form is associated with the loss of the type-specific component, the polysaccharide component being retained.

The capsulated bacillus of Friedlander has given results entirely analogous to those obtained with the pneumococcus (Heidelberger, Goebel and Avery 1925; Julianelle 1925). The species may be divided into a number of sharply demarcated serological types. This type-specificity is conferred by a polysaccharide hapten present in the capsule. The body of the bacillus contains a nucleo-protein antigen that is shared by all types.

Polysaccharide haptens have also been isolated from the tubercle bacillus (Landlaw and Dudley 1925; Enders 1929), from *Bact. lactis aerogenes* (Tomczik and Kurotchkin 1928), from organisms of the *Salmonella* group (Furth and Landsteiner 1928, 1929), from cholera vibrios (Landsteiner and Levine 1927; Jermoljewa and Bujanowskaja 1930), from Shiga's dysentery bacillus (Meyer 1930; Morgan 1931), from the anthrax bacillus (Tomczik 1930; Schockaert 1928; Tomczik and Szongott 1932) and from yeasts (Mueller and Tomczik 1924; Stone and Garrod 1931; Duncan 1932a).

Recently the isolation of antigens more nearly in the native form has been facilitated by gentler and in some cases more specific methods of fractionation. Among these we may mention trichloroacetic acid (Borin and Mesrobian 1933) and diethylene glycol (Morgan 1937) for the extraction of the main antigenic complexes from salmonella and dysentery bacilli (see Chapters 29 and 30). Raistrick and Topley's (1934) method for the same type of substance by tryptic digestion of acetone-extracted bacteria, and Fuller's (1938) formamide method of extracting group-specific components from streptococci. Sonic vibrations apparently liberate antigens from bacteria in a more native form. Mudd and his colleagues (1937) succeeded by this means in isolating from *Str. pyogenes* a labile antigenic substance built up of smaller, serologically specific fractions extracted by Lancefield. By similar methods Sevag and his colleagues (1941) separated from the disintegrated streptococcal cells large numbers of antigenic "macromolecular" particles containing lipins, nucleic acid, protein, carbohydrate and a pigment.

Julianelle and Wieghard (1934) and Thompson and Khorazo (1937) isolated carbohydrate substances responsible for the type specificity of *Staph. aureus*. By milder procedures Verwey (1940) isolated an additional, type specific protein antigen. According to Menzel and Rake (1942) the type-specific substance of the Type II meningococcus owes its serological activity to a carbohydrate polypeptide complex differing from the type specific polysaccharide and type-specific proteins described for other species of cocci. (For the characterization of protein antigens in the gonococcus see Stokinger *et al.* 1944, and of various antigenic fractions in *C. diphteriae* see Wong and Tung 1939, 1940; Wong 1940 and Hoyle 1942.)

The case of the anthrax bacillus presents points of interest. The normal virulent form of this bacillus is capsulated and gives a rough colony on agar, the avirulent variant is non-capsulated and gives a smooth colony on agar (Preisz 1904; Eisenberg 1912). This, then, is one of the cases in which the normal relation between smoothness and virulence is reversed.

It was natural to suppose that the polysaccharide component isolated from *B. anthracis* was the capsular material but Tomesik and Szongott (1932-1933) report that this is not the case. They state that the polysaccharide is a somatic component common to the virulent capsulated rough colony forming type and to the avirulent non capsulated smooth-colony forming type. The capsular substance is a protein like substance. Ivánovics and his colleagues identified the capsular substance as a polypeptide which on hydrolysis yielded d( ) glutamic acid almost in the amount to be expected if the starting substance were built up of glutamic acid only. This antigenically distinct capsular polypeptide was also found in *B. mesentericus* and certain other members of the *Bacillus* group (Ivánovics and Erdos 1937, Ivánovics and Bruckner 1937a & b 1938).

There is another way in which chemical methods may be applied in the investigation of antigenic structure. Bacteria may be treated with various chemical solvents or other reagents with or without heat and the effect of this treatment on their immunological behaviour may be studied (see White 1927-1928-1929-1932-1933).

As an example of an indirect method of chemical analysis we may cite the comparison of the serological reactions of a natural antigen with that of a synthetic antigenal hapten is of a known chemical nature. Thus Goebel (1936) confirmed the importance of glucuronic acid in the molecular structure of the capsular polysaccharide of Types II, III and VIII pneumococci by demonstrating the cross reactions between the polysaccharides and antisera to glucuronic acid azo-proteins. The method is not entirely specific since a group of non identical haptens having certain chemical similarities may cross react with one another. Thus the precipitation of antibody to galactose by an azo protein by the polysaccharide of pneumococcus Type I confirms the importance of the galacturonic acid identified in the polysaccharide molecule but antisera to azo protein antigens containing a benzene carboxylic radicle (which has never been identified in Type I pneumococci) also precipitate with the polysaccharide (Goebel and Hotchkiss 1937). The cross reaction in this case appeared to depend on the reactivity of the antisera with acidic groups irrespective of the hapten radicle that bore them (see also Goebel 1941).

The first essential for the characterization of a bacterial product is the certainty that it has been synthesized by the bacterium and not derived from the culture media from which it was obtained. It will be obvious that blood and other animal proteins used in culture and adsorbed to the bacterium, may induce the formation of antibodies that are later confused with those induced by the bacteria (see, for example, Bliss 1938).

Bailey and Raffel (1941) record that even infusion broths will substantially modify the antigenicity of bacteria grown in them. In association with bacteria non antigenic substances like agar may be antigenic (Sordelli and Mayer 1931, Morgan 1936b, Sickles and Rice 1938). The mechanism of this association is unknown, but Partridge and Morgan (1942) have provided a model for it by making artificial antigenic complexes of agar and the conjugated protein component of the endotoxin of *St. alige*.

The development of culture media whose components are all known and all of relatively low molecular weight will eliminate this source of error (See for example Freeman *et al.* 1940).

It should be emphasized that only in a few cases has the chemical constitution of antigens been fully determined. The antigenic substances isolated from bacteria are usually either large molecules or large associations of molecules. The interpretation of their biological activity in terms of chemical structure must always be tentative in the absence of satisfactory evidence that the product is pure. Unfortunately the biologists and the chemists' conception of purity are not always coincident, and each tends to use criteria borrowed from the other, without full appreciation of their limitations. For instance the fact that a substance has

undergone several successive precipitations by the same reagent or successive crystallizations from the same solvent, is no guarantee of its purity, for loosely bound heterogeneous complexes will exhibit constant precipitability, and mixed and contaminated crystals are common in biochemistry. For further details on this important point, reference should be made to Pirie's (1910) review of the subject.

### The Sharing of Antigenic Components between Unrelated Bacteria, or between Bacteria and Other Cells

In the course of the numerous studies that have been made on antigenic structure, instances have come to light of the sharing of a particular antigenic component by bacteria that show no systematic relation to one another, or by bacteria on the one hand and plant and animal tissues on the other. It is possible in some cases that the reported immunological cross reactions may have been due to the presence of contaminating antigens discussed in the previous section. It will be noted that the various types of pneumococci figure largely in the examples given below. This predominance is not necessarily a peculiarity of the species, more probably it is a reflection of the extensive American work on the pneumococcal antigens.

Thus pneumococcus Type I and certain coliform bacilli are serologically related (Barnes and Wight 1935). The capsular polysaccharide of pneumococcus Type II is closely similar to that of the Type B pneumobacillus (Avery *et al.* 1925, Julianelle 1926), to polysaccharides isolated from a species of yeast (Sugg, Richardson and Neill 1929) and to a species of *Leuconostoc* (Sugg and Hehre 1942), and cross reactions occur between this type and *Past. leipsticka* (Dingle 1934).

Antisera to Types II and III pneumococci react with a polysaccharide obtained by partial hydrolysis of gum arabic (Heidelberger, Avery and Goebel 1929), and of gum acacia, cherry and other vegetable gums (Marrack and Carpenter 1938). Miller and Boor (1934) report cross reactions between Type III pneumococcus on the one hand, and meningococcus and gonococcus on the other, and Kauffmann and Langvad Nielsen (see Morch 1942) between Type XXXV and a *Salmonella* species. There are similar relationships between Type VI pneumococcus and *H. influenzae* Type a (Chapman and Osborne 1942, Vetter 1943, Zepp and Hodes 1943).

The polysaccharide of pneumococcus Type XIV is related to that which characterizes the antigen of the human Group A red blood cell (Goebel *et al.* 1939) and both are related to the somatic polysaccharide of the anthrax bacillus (Ivanovics 1940).

Many other examples could be cited of relationship between markedly differing species, particularly within the group of the Gram negative intestinal bacilli. Perhaps one of the most striking antigenic relationships is that between certain species of *Proteus vulgaris* and certain species of *Rickettsia* (see Chapter 39) where the antigenic varieties in one group are to some extent paralleled by antigenic varieties in the other.

An equally curious example of this sharing of antigenic components is afforded by the presence in a wide variety of bacteria of Forssman's heterophile antigen (see p. 1069) which is a constituent of the red blood corpuscles and tissue cells of certain animal species (see Rothacker 1913, Iijima 1923, Schmidt 1925, Meyer 1926, 1930, 1931, Meyer and Morgan 1935, Powell 1926, Yasui 1929, Combiesco *et al.* 1930, Eisler 1931a, b, Eisler and Howard 1931, 1932, Bailey and Shorb 1931, Buchbinder 1935, Morgan 1937, Goebel and Adams 1943, Goebel *et al.* 1943).

There is, of course, nothing very bizarre in the occurrence of identical, or closely similar, antigenic groupings in living cells that have no close systematic relationship. The specificity that antigen-antibody reactions detect is, as we have seen, a chemical one. The fact that it is also biological, in the systematic sense, depends on the

way in which the chemical substances or the complexes of enzyme systems that synthesize them are distributed in nature. It is very probable that as our search extends, we shall come on one instance after another in which antigenically similar substances are found in entirely dissimilar biological situations.

**The Localization of Antigenic Components in the Bacterial Cell**—The observations recorded above, and many others that will be referred to in subsequent chapters (see particularly the antigenic structure of the typhoid paratyphoid group Chapter 30) not only tell us that there are many different antigens in bacterial cells, and that these antigens have different chemical constitutions—they give us some indication of whereabouts in the bacterial cell these different antigens are placed. They indicate also that this antigenic anatomy is a factor of primary importance in determining that cell's immunological behaviour. Our conception with regard to the way in which such components are actually arranged can be set out most easily in diagrammatic form. But it must be emphasized that our diagram is not a picture. By placing an antigen at the cell surface we are implying that it behaves as though it were there. By placing it beneath the surface we mean that it seems in the normal form of the organism, to be overshadowed by some other bacterial component. How the components are really arranged we do not know except that we can certainly allocate some to flagella or capsules and are almost certainly right in supposing that changes in antigenic behaviour are associated with changes at the cell surface. It is quite likely that antigenic variation is associated with a change in the amount of a particular antigenic component as well as a change in its situation. A component, for instance, that has been unmasked by the loss of another component may be produced in greater amount when it assumes a dominant position on the active surface of the cell. Perhaps in some cases it is only represented in the normal cell in a rudimentary form.

Bearing these caveats in mind we may consider the diagrammatic arrangements of antigens set out in Fig. 44.

At A is represented a portion of a flagellated bacillus in its normal virulent smooth form, with one antigen at the surface of its flagella, a second and a third antigen at the cell surface, a fourth antigen situated more deeply and masked by antigens two and three, a fifth antigen situated more deeply still and a sixth antigen situated centrally in the cell body.

At B is a portion of a bacillus having the same antigens, but without flagella. We might regard it as an O variant derived from A. Alternatively we might alter the nature of the antigens, maintaining their arrangement, and regard it as the normal, smooth form of any non flagellated organism.

At C is a rough variant from A. It has kept its flagella and its normal flagellar antigen, but it has lost the antigenic components that determined the nature of the cell surface in the normal, smooth form. A deeper antigen has been unmasked. It will be noted that a few of the antigenic components that originally lay more deeply still are represented as having now found a place at the cell surface.

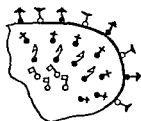
In some cases (see White 1933) antigenic variation by loss may proceed still further. The characteristic rough somatic antigen may disappear and the bacterial surface may be dominated by components that lay very deeply in the normal, smooth form.

At D is an organism, such as the pneumococcus, having a capsule that consists mainly of one kind of antigen (or haptén). Beneath it, in the cell body, is another antigen which is entirely covered by the capsule.



A

- Flagellar Antigen
- Predominant Surface Antigen
- More deeply situated Antigen
- Still more deeply situated Antigen
- Centrally situated Antigen



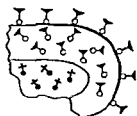
B

As above without Flagella



C

- Flagellar Antigen
- Uncovered Antigen now at surface
- More deeply situated Antigen partly at Surface
- Centrally situated Antigen



D

- Capsule Antigen
- Antigen of Cell Body



E

- Antigen of Cell Body now at Surface

## REFERENCES

- ALEXANDER, H. E. and HEIDELBERGER, M. (1940) *J exp Med*, 71, 1  
 ANDREWES, F. W. (1922) *J Path Bact.*, 25, 505 (1925) *Ibid.*, 28, 345  
 ARKRIGHT, J. (1920) *J Path Bact*, 33, 358, (1921) *Ibid.*, 34, 36, (1924) *Brit. J exp Path.*, 5, 23  
 AVERY, O. T. and GOEBEL, W. F. (1933) *J exp Med*, 58, 731  
 AVERY, O. T. and HEIDELBERGER, M. (1923) *J exp Med*, 38, 81, (1925) *Ibid.*, 42, 367  
 AVERY, O. T., HEIDELBERGER, M., and GOEBEL, W. F. (1925) *J exp Med*, 42, 709  
 AVERY, O. T. and MORGAN, H. J. (1925) *J exp Med*, 42, 347  
 AVERY, O. T. and NEILL, J. M. (1923) *J exp Med*, 42, 355  
 BAILEY, G. H. and RAFFEL, S. (1941) *Amer J Hyg.*, 33, 66  
 BAILEY, G. H. and SHORR, M. S. (1931) *Amer J Hyg.*, 13, 831  
 BARNES, L. A. and WIGHT, E. C. (1935) *J exp Med*, 62, 291  
 BEYER, H. G. and REICH, A. L. (1904) *J med Res.*, 12, 313  
 BIGGEE, J. W., BOLAND, C. R., and O'NEARA, R. (1927) *J Path Bact.*, 30, 261  
 BLISS, E. A. (1938) *J Immunol.*, 34, 337  
 BOVIN, A., ILARD, Y., and SARCHON, P. (1939) *C R Soc Biol.*, 131, 867, 870  
 BOVIN, A. and MESROBIANT, L. (1933) *C R Soc Biol*, 112, 76, (1937) *Rev Immunol.*, Paris 3, 319  
 BORDET, J. and RENAUX, E. (1930) *Ann Inst Pasteur*, 45, 1  
 BRAUN, H. and NODAKE, R. (1924) *Zbl Bakt.*, 92, 429  
 BUCHMÜLLER, L. (1935) *Arch Path.*, 19, 841  
 BURNET, F. M. (1934) *Brit J exp Path.*, 15, 354  
 CASTELLANI, A. (1902) *Z Hyg InfektKr.*, 40, 1  
 CHAPMAN, O. D. and OSBOURNE, W. (1919) *J Bact.*, 44, 620  
 COMBESCO, D., STATATESCO, S., NESTORESCO, N., and ADAM, C. (1930) *Arch roum Path. exp Microbiol.*, 3, 241  
 CRAIGIE, J. (1931) *J Immunol.*, 21, 417  
 DINGLE, J. H. (1934) *Amer J Hyg.*, 20, 143  
 DUNCAN, J. T. (1932a) *Brit J exp Path.*, 13, 499, (1932b) *Ibid.*, 13, 498.  
 DURHAM, H. E. (1901) *J exp Med*, 5, 353.  
 EISENBERG, P. (1912) *Zbl Bakt.*, 63, 305  
 EISLER, M. (1931) *Z ImmunForsch.*, 70, 49, (1931) *Ibid.*, 73, 37  
 EISLER, M. and HOWARD, A. F. (1931) *Z ImmunForsch.*, 71, 473, (1932) *Ibid.*, 78, 461  
 ENDERS, J. F. (1919) *J exp Med.*, 50, 777  
 FELIX, A. (1924) *J Immunol.*, 9, 115  
 FELIX, A. and ROBERTSON, M. (1928) *Brit J exp Path.*, 9, 6  
 FREEMAN, G. C., CHALLINOR, S. W., and WILSON, J. (1940) *Biochem J.*, 34, 307  
 FULLER, A. T. (1935) *Brit J exp Path.*, 19, 130  
 FURTH, J. and LANDSTEINER, K. (1928) *J exp Med*, 47, 171 (1929) *Ibid.*, 49, 727  
 GOEBEL, W. F. (1936) *J exp Med*, 64, 29, (1940) *Ibid.*, 72, 33.  
 GOEBEL, W. F. and ADAMS, M. H. (1943) *J exp Med*, 77, 435.  
 GOEBEL, W. F., BEESON, P. B., and HOAGLAND, C. L. (1939) *J biol Chem.*, 129, 455  
 GOEBEL, W. F. and HOTCHKISS, R. D. (1937) *J. exp Med.*, 66, 191  
 GOEBEL, W. F., SHEDLOVSKY, T., LAVIN, G. I., and ADAMS, M. H. (1943) *J biol Chem.*, 148, 1  
 GRIFFITH, F. (1923) *Rep publ Hlth. med Subj.*, Lond No 18.  
 GRUBER, M. and DURHAM, H. E. (1936) *Munch. med Wochr.*, 43, 285  
 HEIDELBERGER, M. and AVERY, O. T. (1923) *J exp Med.*, 38, 73 (1924) *Ibid.*, 40, 301  
 HEIDELBERGER, M., AVERY, O. T., and GOEBEL, W. F. (1929) *J exp Med.*, 49, 847  
 HEIDELBERGER, M., GOEBEL, W. F., and AVERY, O. T. (1925) *J exp Med.*, 42, 701  
 HEIDELBERGER, M. and KARAT, E. A. (1934) *J exp Med*, 60, 643, (1936) *Ibid.*, 63, 737.  
 (1937) *Ibid.*, 65, 885 (1938) *Ibid.*, 67, 545.  
 HEIDELBERGER, M. and HENDALL, F. E. (1934) *J exp Med.*, 59, 519  
 HENDERSON, D. W. (1939) *Brit J exp Path.*, 20, 11  
 HENRIKSEN, S. D. and HEIDELBERGER, M. (1941) *J exp Med.*, 74, 103.  
 HOFMEIER, K. (1927) *Z ImmunForsch.*, 50, 71  
 HOWIE, J. W. and CRICKSHANK, J. (1940) *J Path Bact.*, 50, 235  
 HOYLE, L. (1919) *J Hyg., Camb.*, 42, 416  
 IJIMA, T. (1923) *J Path Bact.*, 28, 519  
 IVÁNOVICS, G. (1940) *Z ImmunForsch.*, 98, 373  
 IVÁNOVICS, G. and BRUCKNER, V. (1937a) *Z ImmunForsch.*, 90, 304, (1937b) *Ibid.*, 91, 170, (1938) *Ibid.*, 93, 119  
 IVÁNOVICS, G. and ERDŐS, L. (1937) *Z ImmunForsch.*, 90, 5.  
 JERMOLJEW, Z. W. and BUJANOWSKAJA, I. S. (1930) *Z ImmunForsch.*, 68, 346  
 JULIANELLE, L. A. (1926) *J exp Med.*, 44, 113, 683, 735



- JULIANELLE, L. A. and WIEGHARD, C. W. (1934) *Proc Soc exp Biol & Med* 31, 947  
 KAUFFMAN, F. (1936) *Z Hyg Infekthkr*, 118, 549, 119, 103, (1939) *Acta path microbiol scand*, 16, 278  
 KAUFFMAN, F. and TESDAL, M. (1937) *Z Hyg Infekthkr*, 120, 168  
 KRUIP, P. H. DE (1921) *J exp Med*, 23, 773  
 LAIDLAW, P. P. and DUDLEY, H. W. (1926) *Brit J exp Path*, 6, 197  
 LAMARCA, C. (1940) *J infect Dis*, 67, 103, 205  
 LANCEFIELD, R. C. (1928) *J exp Med*, 47, 91, 469, 857, (1933) *Ibid*, 57, 571  
 LANCEFIELD, R. C. and TODD, E. W. (1928) *J exp Med*, 48, 769  
 LANDSTEINER, K. and LEVINE, P. (1927) *J exp Med*, 46, 213  
 MARRACK, J. and CARPENTER, B. R. (1938) *Brit J exp Path* 19, 53  
 MEVZEL, A. E. O. and PACE, G. (1942) *J exp Med* 75, 437  
 MEYER, K. (1926) *Z Immunforsch*, 45, 97, (1930) *Ibid*, 68, 98, (1931) *Ibid*, 71, 331  
 (1938) *C R Soc Biol* 129, 455 (1939) *Ann Inst Pasteur* 62, 282  
 MEYER, K. and MORGAN, W. T. J. (1935) *Brit J exp Path*, 16, 476  
 MILES, A. A. (1933) *Brit J exp Path*, 14, 43, (1939) *Brit J exp Path* 20, 63.  
 MILLER, C. P. and BOOR, A. K. (1934) *J exp Med*, 59, 75  
 MONCH, E. (1942) *J Immunol*, 43, 177  
 MORGAN, W. T. J. (1931) *Brit J exp Path*, 12, 62 (1936a) *Rep int Congr Microbiol*, 423 (1936b) *Biochem J*, 30, 909, (1937) *Ibid* 31, 2003  
 MUDD, S., CZARNETSKY, E. J., PETTIT, H., and LACKMAN, D. (1937) *Publ Hlth Rep Wash* 52, 434  
 MUELLER, J. H. and TOMCSIK, J. (1924) *J exp Med*, 40, 343  
 NETER, E. (1943) *J Immunol*, 46, 239  
 ORCUTT, M. L. (1924) *J exp Med*, 40, 43, 627  
 PARTRIDGE, S. M. and MORGAN, W. T. J. (1942) *Brit J exp Path* 23, 84  
 PIRIE, N. W. (1940) *Biol Rev, Camb* 15, 377  
 POWELL, H. M. (1926) *J Immunol*, 12, 1  
 PREISE, H. (1904) *Zbl Bakt*, 35, 280, 416, 537, 657  
 RAISTRICK, H. and TOPELEY, W. W. C. (1934) *Brit J exp Path* 15, 113  
 REIMANN, H. A. (1925) *J exp Med* 41, 587; (1927) *Ibid*, 45, 1, 807  
 ROTHACKER, A. (1913) *Z Immunforsch* 16, 491  
 SCHMIDT, H. (1925) *Z Immunforsch*, 43, 422  
 SCHOCKAERT, J. (1928) *C R. Soc. Biol* 99, 1242  
 SEVAG, M. G. SMOLENS, J. and STERN, L. G. (1941) *J Biol Chem*, 139, 925  
 SICKLES, G. M. and RICE, C. E. (1938) *J Immunol* 34, 235  
 SMITH, T. and REAGAN, A. L. (1903) *J med Res* 10, 89  
 SORDELLI, A. and MAYER, E. (1931) *C R Soc, Biol*, 107, 736  
 SPRINGUT, E. (1927) *Z Immunforsch*, 52, 25.  
 STOKINGER, H. E., ACKERMAN, H., and CARPENTER, C. M. (1944) *J Bact*, 47, 141  
 STONE, K. and GARROD, L. P. (1931) *J Path Bact*, 34, 429  
 SUGG, J. Y. and HEURE, L. J. (1942) *J Immunol*, 43, 119  
 SUGG, J. Y., RICHARDSON, L. V. and NEILL, J. M. (1929) *J exp Med* 50, 579  
 THOMPSON, R. and KHORAZO, D. (1937) *J Bact*, 34, 69  
 TODD, E. W. (1928a) *Brit J exp Path*, 9, 1, (1928b) *J exp Med* 48, 493  
 TODD, E. W. and LANCEFIELD, R. C. (1928) *J exp Med*, 48, 751  
 TOMCSIK, J. (1930) *Z Hyg Infekthkr*, 111, 119  
 TOMCSIK, J. and KURATSKIN, T. J. (1928) *J exp Med* 47, 379  
 TOMCSIK, J. and SZOGOTT, H. (1932) *Z Immunforsch*, 76, 214, (1933) *Ibid*, 78, 80  
 VERWEY, W. F. (1940) *J exp Med*, 71, 635  
 WEBSTER, L. T. (1926) *J exp Med* 41, 571  
 WEIL, E. and FELIX, A. (1917) *Wien Klin Wochr*, 30, 1509  
 WHITE, P. B. (1927) *J Path Bact*, 30, 113, (1928) *Ibid*, 31, 423, (1929) *Ibid*, 32, 83  
 (1932) *Ibid*, 35, 77, (1933) *Ibid* 36, 65  
 WILSON, G. S. (1930) *J Hyg Camb* 30, 40  
 WONG, S. C. (1940) *Proc Soc exp Biol & Med* 45, 850  
 WONG, S. C. and TUNG, T. (1939) *Proc Soc exp Biol & Med*, 44, 42, 821, (1940) *Ibid* 43, 749  
 YASUI, K. (1929) *Z Immunforsch* 63, 440  
 ZEPP, H. D. and HODES, H. L. (1943) *Proc Soc exp Biol & Med* 52, 315  
 ZINSSER, H. and PARKER, J. T. (1923) *J exp Med*, 37, 375

## CHAPTER 9

### BACTERIAL VARIATION

In the earlier bacteriological writings from the days of Pasteur and Koch onwards there will be found scattered references to certain bacterial strains that have deviated in one way or another from the modal form of the particular species concerned. In the earliest days of all when the doctrine of spontaneous generation was dying but not yet dead it was indeed the fixity rather than the variability of bacterial species that was in dispute. It was not however until the beginning of the present century that any serious attempt was made to study bacterial variation as a problem *sui generis* or to apply to bacteria the concepts that had proved so fruitful in the study of the higher plants and animals. During the first decade or so of the present century these attempts were sporadic but during the last twenty five years an immense impetus has been given to this line of inquiry by a series of converging studies and the relevant literature has expanded from a trickle to a flood.

The significance of many of the observations that have been recorded is at the moment exceedingly difficult to assess. Some of them serve to illustrate the wide range of variation that may occur within a single bacterial species but tell us little or nothing in regard to the relative frequency of the different variants described or the factors on which the variation depends. Others are concerned at least in part with problems that have been described in Chapter 2—the existence of a complex bacterial life-cycle of filtrable forms of bacteria of some form of sexual reproduction and so on. In the present chapter we shall discuss the general problem of bacterial variation, as illustrated by a series of observations the selection of which must of necessity be to some extent arbitrary. A more detailed account of certain variants will be found in the systematic descriptions of the different genera and species given in later chapters.

#### Terminology

The application to bacteria of terms that have been coined to express changes in form or function occurring in higher plants or animals is not without its dangers and it is possible that there is little real justification for the use of such a term as mutation in connection with the variations which bacteria may undergo. Some biologists would attach two implications to the use of this term—the suddenness of the change—the variation *per saltum*—and the permanency of the change, once it has occurred. Dobell (1913) regards as a mutation any permanent change which is transmitted to subsequent generations of bacteria without any implication in regard to the suddenness or gradualness of the change or the manner of its acquisition. We must at all events remember that most of our conceptions with regard to variation and heredity have been built up on data derived from observations and experiments on living things which pass through a sexual cycle.

and, so long as we regard bacteria as asexual organisms multiplying by simple binary fission, we must avoid the tendency to misapply concepts which have their essential basis in the segregation, and conjugation, of a special system of reproductive cells. Some of the concepts are applicable, as we noted in Chapter 2 the persistence of specific characters through a large number of generations of a bacterium implies an hereditary mechanism of some kind, and one that behaves as a unit within the bacterial cell. For the purposes of bacterial genetics, it is irrelevant whether a nuclear apparatus has been demonstrated morphologically or not. A logical deduction from the mathematical data of genetical experiments with other species leads to a chromosome and gene mechanism. We have as yet insufficient data on bacterial heredity to justify the assumption that here also there is a similar mechanism, but in the meantime, it is convenient and indeed sensible, to use those concepts of general genetics which are applicable to bacteria.

We may postulate a haploid or a polyploid nucleus, scanty morphological studies suggest that haploid and perhaps diploid nuclei are likely to be commonest in bacteria. The mutations observed in bacteria are characterized by absence of the intermediate forms, and a moderate readiness to reversion. If these are due to genic changes in the nucleus, the most likely of all known types of gene change is transposition (Landegeen 1935). It should be noted, however, that single-cell cultures, which are essential for the proper study of bacterial genetics do not necessarily represent a single cell in the genetical sense, for many single cells appear to contain more than one nuclear unit. In this respect at least a technique for the morphological demonstration of a nucleus is a necessary foundation for the formulation of a "mathematical" nucleus referred to above.

In one sense, bacteria offer a fruitful field for the study of mutations. Observable mutations occur with great rarity among more complex multicellular organisms. If bacterial mutations are of the same order of rarity, the bacteriologist will have ample opportunities for observing them, since the colony that grows from a single cell after a day's inoculation on an artificial medium may contain  $10^7$ - $10^8$  individual viable cells, representing the end result of an even greater number of divisions during which mutation could have occurred.

The term "bacterial dissociation" is frequently employed to denote a particular type of bacterial variation (see Hadley 1927, 1937, and Morton 1940, for detailed reviews). This term, in its generally accepted sense, denotes the appearance, in a bacterial culture, of forms which differ sharply, in one or more characters, from the "normal" forms of the parent strain, that is, the strain may be said to have undergone dissociation into two types, differentiated from each other in colony form in antigenic structure, or in some other way. The variation must be discontinuous in type, even though there is some overlapping, and the dissociated, or variant form, must be sufficiently stable to maintain its new characters over several generations, whether or not it eventually reverts, wholly or in part, to the normal form from which it was derived.

Those who uphold the view that bacteria pass through a complex life cycle have naturally sought to relate the phenomenon of dissociation to the phases of this cyclical development (Hadley 1927), but the term may be employed in a purely descriptive sense, without any reference to the possible existence of modes of reproduction other than simple binary fission. It is, however, doubtful whether "dissociation" has any advantage over "variation" as a descriptive term, and, since it is desirable, at the present stage, to avoid any implication in regard to

the underlying mechanisms involved, it seems wiser to adhere to the older name, which serves our purpose well because of its very vagueness.

### **Correlated Variations.**

An observed variation in any given bacterial character clearly gains in significance if it is found to be uniformly or frequently associated with a change in some other character, or characters. It is a fairly safe assumption that correlated variations of this kind indicate some relatively major change in genetic make-up. Whether the correlated character changes are different expressions of a single character factor, or are due to changes in two or more genetic factors that are themselves associated as a result of the reproductive mechanisms of the cell, we cannot tell, but in a few instances, as in the smooth  $\rightarrow$  rough variations to which we have already referred in Chapter 8, and which we shall shortly describe in more detail we can relate many of the associated character changes to a loss of the ability to synthesize and store a particular chemical component of the bacterial cell.

### **Impressed Variations**

By an impressed variation is meant a variation that occurs in response to a particular environmental stimulus, so that by applying the stimulus we can induce the variation at will. It should be noted that if this term is to be applied in its strict sense it is the genetic variation that must be impressed, not merely the character change by which this variation is recognized. We know, for instance, certain genetic variations in insects that lead to the appearance of well-defined character changes under particular environmental conditions. In the absence of these conditions the insects appear to conform to the normal type, though the genetic differences, which are themselves quite independent of these environmental factors, persist all the time. The criteria that justify the conclusion that a particular variation has been induced by a particular environmental stimulus are clearly that the application of the stimulus should regularly be followed by the appearance of the variation in question, and that the variant form should persist, over many generations at least after the stimulus has been withdrawn. The appearance of a variant form in response to a given stimulus, followed by immediate reversion to the normal form when the stimulus is no longer applied, should be regarded as a temporary adaptation to a changed environment, rather than as a variation in the sense in which that term is used here. It is often extremely difficult, in the light of our present knowledge to determine in which category a given change in bacterial form or function should be placed.

It will be convenient to discuss the variability of each of the more important bacterial characters in turn indicating where possible, whether the variation in question is correlated with others, and whether it occurs naturally or in response to any known change in environment.

### **Variations in Morphology**

There are innumerable accounts in the literature of changes in shape, size or structure of bacterial cells. Some of these such as the involution forms that appear in cultures of the plague bacillus when that organism is grown on agar with a high salt-content are clearly a direct response to an environmental stimulus, and are not inherited. In other instances it is very difficult to tell whether or not true variation has occurred. From among the many examples available, we may select the following as illustrating the kinds of variation that have significance from our present point of view.

Barber (1907), starting with a single strain of *Bact coli*, selected and subcultured, by means of a micromanipulator, individual bacterial cells that had grown to an unusual length. In this way he was able to isolate three strains that grew in the form of long rods throughout many successive generations, showing no tendency to revert to the modal short bacillary form of the original parent strain. This would appear to afford an instance of the perpetuation of a natural heritable variation by simple selection.

We have already, in Chapter 8, noted the occurrence of the  $H \rightarrow O$  variation—the loss by a flagellated organism of the capacity to produce flagella, associated of course with a loss of motility. When this variation occurs naturally, the variant  $O$  form usually shows no tendency towards reversion. The importance of this change from the point of view of antigenic analysis led to a search for methods by which it could be induced at will. It has been found that growth on agar containing 0.1 per cent phenol largely, or completely, suppresses the formation of flagella (Braun 1918), but the non flagellated cells so obtained give rise to the normal flagellated form when subcultured on ordinary media, so that we are here dealing not with an impressed variation but with a temporary adaptation to environment.

Another striking morphological variation is the occurrence of asporogenous variants of such spore bearing bacilli as *B anthracis* (Preisz 1904, Eisenberg 1912). Many of these naturally-occurring asporogenous variants have shown no tendency to revert to the normal spore bearing form. Pasteur (1881a, b) found that the growth of *B anthracis* at 42.5° C for about a month resulted in a great decrease in the frequency of spore formation, as well as in a decrease in virulence, and Roux (1890) obtained asporogenous strains of this organism by growing it in the presence of low concentrations of antiseptics. Whether the asporogenous strains obtained by these methods were examples of an impressed variation or of a temporary adaptation to environment is difficult to determine. Later experiments by Bordet and Renaux (1930), however, strongly suggest the occurrence of an impressed variation of the genetic type. They found that certain strains of *B anthracis*, yielding the normal, flat, filamentous colony when grown on solid media, gave rise on prolonged incubation to a characteristic type of papillary daughter colony consisting of asporogenous bacilli. By repeated subculture from these daughter colonies a completely asporogenous strain of *B anthracis* could be isolated. When the medium on which these strains of *B anthracis* were grown was deprived of its calcium by treatment with oxalate, spore formation was stimulated, and the papillary, asporogenous daughter colonies did not appear. When the calcium content of the medium was increased, by the addition of a little calcium chloride, the frequency of spore formation decreased and the papillary, asporogenous daughter colonies became very numerous. Repeated subcultures from these daughter colonies again produced a completely asporogenous strain (see also Bordet, P 1930).

There would seem to be an interesting difference between the asporogenous strains of *B anthracis* obtained by the method of Pasteur, and those obtained by the method of Bordet and Renaux. Although growth at 42.5° C for several days leads to a decrease in the average virulence of a culture and to the appearance of various abnormal bacillary forms, many of which are non spore bearing, Preisz (1911) was unable to find any correlation between the capacity to form spores and virulence as tested by animal inoculation. Spore bearing strains might be virulent

or avirulent, so might asporogenous strains. In Bordet and Renaux's experiments the actively sporogenous strains obtained by cultivation on oxalated media were highly virulent, while the asporogenous strains obtained by subculturing from the daughter colonies on media with a high calcium content were completely avirulent. It would seem, then, that loss of ability to form spores is sometimes, but not always, associated with loss of virulence. The most reasonable hypothesis would seem to be, either that asporogenous strains may be produced by different genetic mechanisms, or that the presence in a medium of excess of calcium stimulates some genetic change in addition to that on which the loss of spore-bearing capacity depends. We should not, it may be noted, on the basis of these divergent results, be justified in regarding this loss and the loss of virulence as correlated variations in the usually accepted sense. The use of precisely defined media will in all probability throw light on a number of morphological variations, though there are at present only a few isolated observations on this point. For example, in an amino-acid glucose salt medium, and sub-optimal amounts of an unidentified growth factor present in liver extract, gonococcal strains produced large, distorted, swollen, 'vacuolated,' and dumb-bell forms, in the presence of ample growth factor, the cells grew in characteristic diplococcal forms (Lankford, Scott, Cox and Cooke 1943). Badger (1944) records an interesting variation in a Type III pneumococcus for which choline was an essential nutrient. The choline could be replaced by ethanolamine, and in the presence of the latter, the pneumococcus grew in characteristic long chains. Again, Pappenheimer and Shaskan (1944) found that *Cl. welchii* grew as regular rods in a medium containing enough iron to ensure maximum growth, toxin production and breakdown of carbohydrate. With a reduction of iron content only, there was a depression of these activities, and the clostridia grew as curved, elongated and entirely atypical bacilli.

There are many other ways in which bacterial variants may depart from the normal morphological type of the species to which they belong. A capsulated organism, such as the pneumococcus, may, for instance, give rise to a non-capsulated variant. This particular variation is, however, associated with other important changes in behaviour, and it will be more convenient to consider it in the section dealing with antigenic variation.

#### Variations in Biochemical Reactions

Since the earliest days of bacteriology, differences in fermentation reactions have been extensively utilized in differentiating bacterial species or types that belong to the same genus or group, as judged by morphological or other criteria. To be of use from this point of view the fermentation reactions of any given species, or type, must, of course be constant. This has been found to be the case, to the extent that it is usually possible to select empirically a number of substrates that a given organism, in its normal form, will consistently alter or leave unaltered. It has, however, been found that for any given bacterial species there are other substrates that are sometimes fermented, sometimes not, from the systematic point of view we should say that those particular substrates in relation to that particular organism, had little differential value. Similarly, some bacterial species vary more than others in their fermentative abilities, and the differential value of fermentation tests therefore varies from one bacterial group to another.

In trying to assess the significance of the numberless recorded instances of variants that differ in their fermentation reactions from the parent strain from which they were derived, we must keep these facts constantly in mind. We may

regard any given bacterial species as equipped with an armoury of enzymic mechanisms of the kind discussed in Chapter 3. Some of these may perhaps readily be lost by disuse and as readily regained if called into activity by appropriate stimuli. Others may, perhaps, be easily adapted to deal with some substrate that does not differ too greatly from that which the enzyme naturally attacks. Others, again, will depend on more constant and fundamental cell mechanisms; these will be lost, regained or altered only as the result of some deep-seated variation in cell structure.

It is indeed impossible to draw any hard and fast line between those changes in fermentative ability that arise as temporary adaptations to environment and those that may be regarded as variations of a more permanent kind though certain instances can be assigned with some confidence to one category or the other.

**Adaptive and Constitutive Enzymes**—The enzyme response of a bacterium to a substrate may fall into one of two classes. The enzyme may be produced only in the presence of the substrate or it may be produced whether the substrate is there or not. Thus Wortmann (1882) described a bacterium that produced amylase in a starch medium, but none in a starch free medium, and contrasted it with a yeast, which produced invertase whether sucrose was present or no. For these two classes Karstrom (1930, 1937) proposed the names 'adaptive' and 'constitutive' enzymes. For example the production of molecular hydrogen from glucose or formic acid by *Bact. coli* was found to be due to two adaptive enzymes, one a glucose hydrogenlyase, the other a formic hydrogenlyase (Stephenson and Stuckland 1932, 1933; Yudkin 1939). Not only were the enzymes adaptive but they were apparently formed *in situ* as well as in reproducing cells. Some protoplasmic growth, however, appeared to be necessary since in the absence of nutrient broth no enzymes were formed (see also Stephenson and Yudkin 1936). The adaptation disappeared in the absence of the stimulating substrate and was therefore not heritable.

Enzyme formation may also be conditioned by the presence of materials which are unrelated to the substrate, presumably because they are needed for enzyme synthesis (see Jacoby 1916, 1917, 1918; Pasmore and Yudkin 1937). Quastel (1937) developed this concept by postulating that enzymes themselves were metabolites whose rates of formation and destruction follow the same physicochemical laws as those controlling the metabolism of other metabolites in the cells; the presence of substrate therefore might or might not be the particular condition necessary for the production of an enzyme. In his view the adaptive and constitutive enzymes represent the limits of variability of cellular enzymes, the constitutive having the least, the adaptive the greatest range under different environmental conditions.

Euler and Cramer (1913) were able to stimulate the production of invertase in a yeast by the addition of fructose or glucose and also by mannose. It appears therefore, that not only substrates but the products of hydrolysis of substrates and chemicals related to the substrate act as stimulants of adaptive enzymes. According to Yudkin (1938) the Mass Law serves to reconcile many of these phenomena of enzyme adaptation. If it is assumed that the enzyme and its precursor are in a state of equilibrium in the cell with the precursor predominating and the enzyme in undetectable amounts, a substrate, its hydrolytic products, or a related substance will by combining with the enzyme, shift the equilibrium so that more enzyme is formed from the precursor. The mere presence of the

insufficient; it must be utilized by the metabolizing cell. Thus, the production of the enzyme hydrolysing the specific polysaccharide of Type III pneumococcus by Dubos and Avery's (1931) bacillus was retarded when nutrients more readily assimilable than the polysaccharide were added to the culture (see also Dubos 1910).

It should be noted that the appearance of an enzyme does not necessarily imply its increased production, the effect may be due to activation of an enzyme already existing in considerable quantity. It may also be due to the removal of an inhibitor, rather than the addition of a substrate. For instance the tryptophanase system of *Bact. coli* is adaptive but when tryptophan is present glucose and phenylalanine will inhibit the production of the enzyme system by resting bacteria (Evans, Handley and Happold 1940). Dawson and Happold (1943) suggest that the phenylalanine may act by competing with tryptophan for a labile component common to two enzyme systems, one the tryptophanase the other concerned with carbohydrate storage in the cell.

Kocholaty and his colleagues (Kocholaty and Hoogerheide 1938, Kocholaty and Weil 1939) report enzyme adaptations to variations in pH. They produced a shift in the pH optima of, for example, the alanine and pyruvic dehydrogenases of *Cl. sporogenes* by varying the pH of the culture media and found that *Cl. histolyticum* grown in a casein medium produced proteinases with an optimum activity at pH 7.0 but when grown in a casein glucose medium at a lower pH produced proteinases with an optimum at pH 6.0. Cells from glucose-casein cultures when transplanted to the plain casein medium within 20 hours, had produced the proteinase with an optimum at pH 7.0. They were also able to train *Cl. histolyticum* to attack casein or gelatin alone though normally it attacks both equally well. This specificity was very labile however for when amino-acids not present in the homologous protein were added to the medium the resulting enzymes attacked both proteins. To explain these phenomena they sought to combine Yudkin's mass action theory with Quastel's theory of enzymes as metabolites by assuming that relatively few colloidal carriers are available for a number of enzymes and that under different conditions these combine with different active groupings to form enzymes of different activity or specificity. On this basis Yudkin's precursor may be a colloidal carrier in equilibrium with various prosthetic groups of enzymes. On the other hand this type of pH adaptation may depend on a multiplicity of enzymes. van Heyningen (1940) for example described two proteinases in *Cl. histolyticum*, one activated by cysteine, appearing in the first 12 hours of growth one appearing later which was inhibited by cysteine.

In other organisms a close relation is demonstrable between the pH optima of enzymes and the cultural conditions in which the enzymes come into play. Thus *Bact. coli*, *Str. faecalis* and certain clostridia produce amines from amino-acids by specific decarboxylases that act only between the limits pH 2.5 and 5.5. These decarboxylases are formed only when the organisms are grown in acid media (Gale 1940, 1941). Again at a pH in the vicinity of 5.0 *Bact. aerogenes* decomposes pyruvic acid with the formation of acetyl-methyl-carbinol, at a higher pH this activity is entirely suppressed and decomposition proceeds by breakdown into acetic and formic acids (Silverman and Werkman 1941). Extending these investigations, Gale and Epps (1942) showed that *Bact. coli* could grow in a casein digest at any pH between 4.5 and 9.0 and that though with changing growth pH the enzymic constitution of the bacteria changed there was no evidence of any shift in the pH optima of individual enzymes. The enzymes concerned fell into two groups. The formation of those in the first group increased as the growth pH deviated from the pH at which their action was maximum so that in each cell the drop in activity due to the pH change was compensated by the



increased production of enzyme. Catalase, urease and formic dehydrogenase were included in these enzymes, one of whose essential functions appeared to be removal of inhibitory products of metabolism; these were maintained at an efficient level over a large pH range. The enzymes of the second group displayed only a small degree of compensatory formation when the growth pH shifted from the pH of their optimal activity. They behaved like adaptive enzymes, but adaptive in respect of pH rather than of substrates. Nevertheless they contributed to the stability of cellular equilibrium. Thus in an acid medium amino acid decarboxylases and consequently amines were produced; on the other hand amino acid deaminases and consequently hydroxy acids were produced in an alkaline medium.

Adaptation occurs much more readily in physiologically young than in older cells (see for example Hegarty 1939) and we have already seen that adaptation takes place in resting bacteria. The independence of reproduction and some forms of adaptation is confirmed by the interesting study of Doudoroff (1940) on the adaptation of *Bact. coli* to sodium chloride.

When the cultural conditions such as air supply, concentration of nutrient and pH were standardized, a fairly constant fraction of a fresh water culture of *Bact. coli* was able to reproduce when inoculated into a medium of definite NaCl concentration. The acclimatization to salt was independent of reproduction, since bacteria in the stationary phase were far more readily acclimatized than those in the phase of logarithmic growth or the phase of decline. Only acclimatized cells were capable of reproduction in saline media, but if after acclimatization the non-dividing cells were returned to salt-free media they rapidly lost the capacity to grow in saline media. Once acclimatized, the cells could be propagated in saline media without further acclimatization, though the division rate was considerably lower than that in salt-free cultures.

Optimal acclimatization to growth even in high salt concentration was achieved by preliminary exposure to a single intermediate salt concentration. In other words the adaptive response of the bacteria consisted firstly of a reversible acclimatization independent of reproduction, and secondly of a selection of the cells with the widest range of potentialities for growth in various concentrations of saline.

The distinction between adaptive and constitutive enzymes is perhaps more a quantitative than a qualitative one. The production of constitutive enzymes varies according to the conditions of culture, and sometimes according to the presence or absence of the substrate. Moreover, adaptive enzymes are constitutive in the sense that they are a constant feature of the bacterial cell. As far as we are aware, there is no recorded instance of a bacterial strain losing the capacity to make a given adaptive enzyme response by continuous subculture in media which preserve the general characters of the strain. In other words, though by definition the adaptation is not heritable, the specific adaptability certainly is. We do not know whether the potentiality for adaptation is maintained as a precursor or as the enzyme itself, but in some cases it is possible that the enzyme is present as such when no substrate is present, though in undetectable amounts.

**Variation and Mutation in Enzyme Systems**—At the other end of the scale of variation we have those changes which are presumably due to the selection over a large number of bacterial generations of relatively rare mutations. Changes in environment of the kind that induce an adaptive response in bacteria may also serve to make mutations manifest. In a case of this kind the distinction between the two effects would lie mainly in the persistence of the mutation effect when

the original environment was restored. We must nevertheless guard against a too literal interpretation of this difference, for as we have pointed out above bacterial generation under experimental conditions is so rapid as to compensate for a very low mutation rate (the ratio of the number of mutations to the total number of cell divisions). It follows that if mutations in the reverse direction were as frequent as those in the direction of an observed variation, variation and reversion might be accomplished with such speed as to suggest a temporary adaptation.

Mutations are not equally rare in all bacterial strains. Indeed some produce variants so constantly and in such relatively large numbers that they are often designated as 'unstable' strains. Deskowitx (1937) concluded from a study of unstable colony variants of *Salmonella typhimurium* that unstable strains differed from stable only in having a mutation rate of the order of 1 per cent. or more. The existence of these "unstable" strains raises a practical point of some importance in distinguishing adaptation from true mutation.

We have seen in Chapter 3 that the typhoid bacillus though normally dependent for its growth on the presence of tryptophan can be trained to synthesize this essential metabolite from ammonium salts, and in Chapter 6 there are numerous examples of habituation of bacteria to growth in otherwise inhibitory concentrations of various antibacterial agents. These changes in most cases permanent are excellent examples of impressed variation in biochemical constitution. A few examples from recent work on the vitamin metabolism of bacteria and yeasts will serve to show the effects of such training and the degree of success which attends it.

Wood, Anderson and Werkman (1938) trained a strain of propionic acid bacteria to dispense with thiamin, and later showed (Silverman and Werkman 1939) that the trained cultures were synthesizing a substance that had the biological qualities of thiamin. Leonian and Lilly (1942-1943) induced several strains of the yeast *Saccharomyces cerevisiae* to grow without one or more of certain essential vitamins including thiamin, pyridoxin, inositol, pantothenic acid and in some cases biotin. At least two of the variants grew without all five vitamins and in some cases the growth in the absence of one vitamin induced the power to dispense with other vitamins as well. Some variants reverted easily but many of them reverted only after six months subcultivation on a medium containing all the vitamins.

Hoser and Wright (1943) trained four strains of dysentery bacilli to dispense with nicotinamide. The variants could be obtained either by serial subculture in a glucose amino-acid medium containing diminishing quantities of nicotinamide, or by incubation of a large inoculum in a nicotinamide free medium. Under the latter conditions the proportion of variants to the total of viable cells increased during incubation. The variants resembled the parent strains both antigenically and in their gross fermentation reactions but never developed as luxuriantly as in media containing optimal amounts of the vitamin. Nevertheless they had acquired the power to synthesize a substance physiologically equivalent to nicotinamide for filtrates of their cultures in vitamin free media supported the growth of strains of bacteria known to require nicotinamide.

Whereas the typhoid bacillus appears to acquire the ability to synthesize tryptophan with relative ease its normal inability to attack lactose seems to depend on a more fundamental peculiarity of cell organization, since it is extremely difficult to produce lactose fermenting strains of this organism, even by prolonged training in media containing this substrate as the main available source of carbon. Twort (1907) has succeeded in producing one such strain, and we may perhaps regard the extreme rarity with which this change has been induced as

evidence that it depends on a major variation in cell organization, though it must be noted that Penfold (1910a) found a lactose-fermenting strain of this organism to be very unstable, with a marked tendency to revert to the non lactose fermenting form

In this connection we may refer to an interesting type of variability in enzymic activity which was first demonstrated by Massini in 1907. This observer described a coliform bacillus which, on first isolation, failed to ferment lactose, and hence gave rise to colourless colonies on an agar medium, containing lactose and an indicator that gave a red colour in the presence of acid. From the third day of incubation onwards, small papillæ began to appear on these colonies, and took on a red tint, indicating that the bacilli composing them were breaking down the lactose with the formation of acid. Subcultures from these papillæ gave non papillated red colonies, showing that the power to ferment lactose had been transmitted to the descendants of the bacilli which had originally formed the red papillæ, and repeated subcultures showed that this power was not subsequently lost. Subcultures from the colourless parts of the original colonies however, gave rise to colourless colonies on which red papillæ appeared after about 3 days, just as in the case of the original culture, and repeated subcultures from the colourless portions of the colonies of successive generations gave similar results. Thus, the non lactose-fermenting form of this organism showed a constant tendency, when grown on a lactose containing medium, to give off lactose fermenting variants in which the new character appeared to be permanent. To this organism Massini gave the name of *Bacterium coli mutabile*. Such a bacterial strain may, as Dobell has pointed out, be likened to the ever sporting races of plants which have frequently been described. Massini's observations have been confirmed, in all essentials by many subsequent workers (Burk 1908, Benecke 1909, Burri 1910, Kowalenko 1910, Baerthlein 1912a, b), while Benecke, and Kowalenko, added greatly to the significance of their results by starting with a culture obtained from a single bacterial cell thus eliminating the possibility that the phenomena resulted from an original admixture of strains. In a careful quantitative study, Lewis (1934) showed that the mutation rate in this organism was such that when it was grown in a lactose free culture medium, one in every 100 000 viable cells produced a lactose fermenting colony. Lewis also observed that the proportion of variants was relatively constant. A relatively constant proportion of variant and normal cells has been recorded by other observers as characterizing certain mutating strains (see, for example, Solotorovsky and Buchbinder 1941).

Experiments along similar lines, employing other species belonging to the same bacterial group, have been carried out by Penfold (1910a and b, 1911a, b and c, 1912) and Muller (1908, 1911). The observations recorded by these observers have made it clear that the behaviour of Massini's *Bact. coli mutabile* is by no means a bacteriological curiosity, but that, given a substrate appropriate to the particular species under investigation, many members of the coli typhoid dysentery group will adapt themselves to a particular nutrient material which they do not immediately attack by giving rise to variants endowed with the power of breaking down this particular substrate.

Thus, *Salm typhi* usually fails to ferment dulcitol, but when grown on a medium containing that alcohol, it gives colonies which develop dulcitol fermenting papillæ (Penfold 1910a, b 1911), the same organism behaves similarly towards rhamnose (Muller 1911), while *Salm paratyphi B* behaves similarly towards

raffinose From Müller's account it would appear that the rhamnose fermenting variants of *Salm typhi*, and the raffinose-fermenting variants of *Salm paratyphi B*, are non-reverting modifications of the parent strain. Apparently, also, subcultures from the non-fermenting portions of the colonies in these two organisms, showed the same tendency to throw off fermenting variants in the form of papillæ, as was observed by Massini in the case of *Bact coli mutabile*. Penfold's observations do not, however, confirm the absence of a tendency towards reversion. In connection with the appearance of dulcitol fermenting forms of *Salm typhi*, in particular, he finds that subcultures from the dulcitol fermenting papillæ, or from fermenting cultures in dulcitol peptone water, show a marked tendency to revert to the non-fermenting parent form during the earlier generations. If the selective process is continued through a long series of successive generations the tendency to reversion becomes less and less, though it appears doubtful whether absolute permanency is ever attained. Penfold concludes from his results that the more rapidly a particular species acquires the ability to ferment a particular substrate, the less tendency is there for subsequent reversion, while the longer and more rigorous is the training required to bring about the appearance of fermenting variants, the longer must that training be afterwards continued to make a lasting impression on that particular strain.

Certain experiments carried out by Penfold (1911b, c) and by Revis (1911, 1912) have brought to light a different type of impressed variation in bacteria. Penfold found that, by growing certain strains of *Bact coli* on an agar medium containing sodium monochloroacetate, he was able to isolate strains which retained the power of producing acid in all the usual carbohydrate media, but which had lost the power of producing gas in many of them. Moreover, he found that while the power to produce gas from sugars, or from substances giving rise to sugars on hydrolysis, was usually suppressed, the power of forming gas from alcohols, such as mannitol or dulcitol, was usually unaffected, or but slightly diminished. The only exception noted was in the case of rhamnose, a methyl pentose, which was fermented with gas formation by the variant strains.

Goodman (1908) obtained variants showing differences in fermentative ability by an essentially different technique. Starting with a particular strain of *C diphtheria*, which produced a certain degree of acidity in dextrose broth, he inoculated 15 tubes of this medium from a single colony, and determined the degree of acidity attained after a few days. From the tube showing the highest acidity he inoculated 15 tubes of the same medium and a similar number from the tube showing the lowest acidity. This process he repeated through 36 successive subcultures. At the end of this series the high acid strain produced a titratable acidity more than twice as great as the parent strain while the low acid strain produced no acidity at all. As would be expected this strain had also lost its power to produce acid from maltose, but it is of interest to note that its power to produce acid from dextrin was almost unaffected. These observations would appear to afford an example of the separation of a bacterial strain into a fermenting and a non-fermenting variant, by a simple process of selection without any modification of the environmental conditions, for though it is true that the high acid strain was in fact subjected to a high concentration of hydrogen ions during its successive subcultures, while the low acid strain was not, it is difficult to understand how the latter condition could lead to the production of a variant which had lost its power to ferment dextrose and maltose, while retaining its power to ferment dextrin.

**Variations in Pigment Production**—It has long been recognized that different strains of a particular bacterial species which gives rise to a coloured growth on the ordinary laboratory media may vary widely in their power of pigment production, and that any particular strain may lose this power as the result of repeated subculture and may regain it for no apparent reason at some later period. In many cases it has been demonstrated that particular environmental conditions are favourable or unfavourable to pigment production but there are many cases on record in which we cannot reasonably attribute the loss of pigmentation to such external influences.

An interesting series of observations have been recorded by Pettger and Sherrick (1911) who studied a strain of the red *Citromobacterium prodigiosum* which had partially lost its pigment producing capacity. By successive subcultures from growths on solid media using in one series the most pigmented part of the growth and in another that part which showed least pigmentation they were able to separate a strain which produced an intensely coloured growth and a strain which gave almost colourless colonies. The segregation of these two types occurred early in the series of subcultures and there appeared to be some tendency for the highly pigmented variant to revert to the slightly pigmented type though the property of intense pigmentation was successfully maintained by selection through a long series of subcultures. There was, however, no apparent tendency for the non-pigmented variant to acquire the property of pigment production. Within recent years there have been numerous records of colourless variants of species that are normally pigmented. In some cases at least these variants differ sharply from the parent strain in the form of colony produced as well as in the absence of pigment and in such instances the variation appears to have much in common with the type discussed in the succeeding section.

#### Antigenic Variations and the Changes in Colonial and other Characters Associated with Them the Smooth-Rough Variation

The Smooth  $\rightarrow$  Rough (S  $\rightarrow$  R) type of variation has already been referred to in our discussion of antigenic structure (pp 277, 278) but it is of such fundamental importance in the general problem with which we are here concerned that it is necessary to consider it in considerably greater detail.

Arkwright (1920 1921 1924) described variants of bacteria belonging to the coli typhoid dysentery group which were characterized by the formation of rough or granular colonies on solid media (see Fig 45) by giving granular growths in broth or peptone water and in many cases by undergoing spontaneous agglutination in the presence of 0.85 per cent sodium chloride.

These properties—colonial roughness, granular growth in fluid media and instability in saline—are associated in some species at least with recognizable changes in morphology (Wilson 1930) and

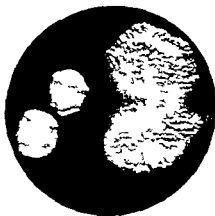


FIG 45—*Salmon typhi*  
Smooth and rough colonies, 24 hours  
growth on agar ( $\times 8$ )

with an alteration in the method of cell division (Nutt 1927). If normal smooth strains of *Salmonella typhi* are grown in a thin layer of agar between a slide and a cover-slip and watched under the microscope, it will be noted that each cell division is soon followed by separation of the daughter cells, which slip past each other and come to lie side by side. In rough variants derived from these smooth strains, the daughter cells tend to adhere, end to end, for some time after division has occurred. Short chains are formed and angular bends develop at the junctions between adjacent cells. The tendency of rough bacilli to agglutinate spontaneously in normal saline appears to be due in many cases to the presence, at the bacterial surface, of some lipoidal substance, since extraction with alcohol at a temperature of 50-60° C removes the salt-sensitiveness of many rough strains (White 1927). It seems probable that this alcohol-soluble constituent is present in the normal smooth form as well as in the rough variant, but in the former it does not determine the character of the cell surface.

In the organisms referred to above, and in many others, the S → R variation is associated with the loss of the polysaccharide antigen that characterizes the surface of the normal smooth form, and, though the rough variant has a polysaccharide constituent of its own, there seems little doubt that the change from smoothness to roughness is associated with a relative increase of lipid as compared with polysaccharide components at the cell surface, and a consequent change in colloidal behaviour from hydrophile to hydrophobe. The hydrophobe qualities of a suspension of rough variants is reflected by increased agglutinability by salt and other non specific agents, like the dye trypanflavine. There is not, however, any dependable connection between this degree of agglutinability and roughness. For example, trypanflavine agglutinates not only rough variants of bacteria in the coli typhoid group, but also Vi strains of *Salmonella typhi*, and flagellated salmonella bacilli in the group phase (Sertie and Boulgakov 1936, 1937, Hirsch 1937). There would also appear to be a laying bare of protein constituents; since rough variants of salmonella bacilli, and certain other organisms, usually give a positive Millon reaction while normal smooth strains do not (see White 1929b).

In the particular case of the pneumococcus the change from smoothness to roughness is associated with the loss of the characteristic capsule, and with it of the specific capsular polysaccharide that determines type-specificity. Among the antigenic components that are left are a nucleo-protein antigen and a minor polysaccharide component that is common to all pneumococcal types (see Avery and Heidelberger 1923, Griffith 1923, Reimann 1925, Tillett, Goebel and Avery 1930).

A similarly associated loss of capsule and virulence has been observed in *Pasteurella septica* (Priestley 1936) and *H. influenza* (Chandler, Fothergill and Dingle 1937), which, like the pneumococcus, had at the same time lost their capacity to produce specific soluble substances. In bacilli with no morphologically distinguishable capsule, there is nevertheless a demonstrable loss of antigen characterizing the virulent form. Thus, Borvin and Mesrobian (1936) could extract 8-9 per cent. of O antigen from smooth *Salmonella typhi* but none from rough forms, and Miles and Pirie (1939) found a progressive diminution in the yield of main antigen from *Br. melitensis* as cultures increased in agglutinability by heat, salt, and a specific antiserum prepared against rough strains.

It must not be supposed, however, that the mere power to synthesize large amounts of polysaccharide substances, whether they appear as a capsule with a defined edge, as a mucoid extra cellular substance, or as increased cellular content, necessarily characterizes the virulent smooth form. For example, Borvin, Mesrobian, Magheru and Magheru (1936) found a high polysaccharide content in all of five smooth strains of *Bact. coli*, but

it was also high in five of eight intermediate strains and in one of seven rough strains. Among the coli typhoid group, indeed the production of large amounts of polysaccharide mucoid substance is sometimes associated with no change, or with a fall in virulence. Mucoid variation is often favoured by growth at 25° C instead of 37° C (Birch Hirschfeld 1935, Morgan and Beckwith 1939 vor dem Esche 1940b). Vor dem Esche (1940a) noted the appearance of mucoid variants in the stool of a woman who had been treated by inoculations with an autogenous vaccine during convalescence from paratyphoid infection.

In many of these instances the change from smoothness to roughness is also associated with a complete or partial loss of virulence, a factor which is clearly of the first importance from the point of view of the medical bacteriologist.

Again (see Chapter II) the  $S \rightarrow R$  variation is usually associated with a change in sensitivity to the lytic action of various strains of bacteriophages, the filtrable viruses that propagate on, and at the expense of the bacterial cells.

Changes that are exactly similar have been described in *Pasteurella* (de Kruij 1921, Webster 1925), many other bacilli of the coli typhoid group (White 1926), *Staph aureus* (Bigger *et al* 1927), and in a large number of other species (see, for instance, Hadley 1927). We may, indeed, regard it as a variation to which most if not all bacteria are subject.

The  $S \rightarrow R$  variation affords an excellent example of a correlated variation. If we accepted the change in colony form as the essential criterion of the change from roughness to smoothness, we should note that the rough variant differed from the smooth form in the following characters:

- (1) Roughness or granularity of colonies.
- (2) Instability in saline.
- (3) Loss of the antigenic component characterizing the surface of the bacterial cell in the normal smooth form, whether this component is normally present in the form of a definite capsule or not.
- (4) Loss of virulence, partial or complete.
- (5) Altered sensitivity to various bacteriophages.

In accepting such a list of correlated characters we are, however, met with the difficulty that the variations observed in certain bacterial species fail to fall into line.

For instance, the normal, virulent, capsulated form of the anthrax bacillus gives a flat, uneven, filamentous colony, while the avirulent non-capsulated variant is raised, circular and smooth (Preis 1904, 1911, Eisenberg 1912). The hæmolytic streptococci appear also to form an exception to the general rule. The colony given by the normal virulent form is finely granular, not smooth, and the avirulent variant, which has lost its type specific antigen, is smoother, not rougher than the normal form. For this reason Todd (1923), who first gave a detailed description of these variants, used the term "matt" to describe the normal colony form, and "glossy" to describe that of the avirulent variant.

According to Dawson, Hobby and Olmstead (1933) the range of variation possible between highly virulent and non virulent forms of streptococci is much wider than that indicated by Todd's matt  $\rightarrow$  glossy. They described in a number of hæmolytic streptococci four types of variants—a capsulated M variant, forming large, watery, mucoid colonies; an MS variant, forming colonies corresponding to Todd's "matt" variants; a non capsulated S variant, corresponding to Todd's "glossy," and an R variant. With progressive loss of characters associated with virulence the variation proceeds as follows— $M \rightarrow MS \rightarrow S \rightarrow SR \rightarrow RS \rightarrow R$  where SR and RS represent variants intermediate between S and R forms (see also Marison 1940, Seastone 1943). A similar series of variants

occurs in other species. It has been described, for example, in *M. tetragenus* (Reimann 1937a) and in *Ham influenzae* (Chandler, Fothergill and Dingle 1939). In the influenza bacillus, the  $M \rightarrow S \rightarrow R$  variation occurred spontaneously, the reversion  $R \rightarrow S$ , but not  $S \rightarrow M$ , could be induced by serial passage through mice. There is also another colonial variant, found in a number of bacterial species, which does not fall into line with the  $S \rightarrow R$ , or the extended  $M \rightarrow S \rightarrow R$  scheme of variation (see Morton 1940). It is the D (dwarf colony) variant described by Hadley (1927, 1937), and must be distinguished from his G (gonidial) variants, which give minute colonies that in some cases are said to contain filtrable elements capable of growing into the normal, modal forms of the parent bacteria. The D variant is characterized mainly by the small size of the colony, in comparison with the S variant. The variations  $S \rightarrow D$  and  $D \rightarrow S$  have been observed in a number of species. In many cases, colony size is the only conspicuous feature distinguishing D from S variants. Thus, the D and S variants of a strain of *Salmonella typhi* (Morris, Sellers and Brown 1941) and of a Group C streptococcus (Morton and Sommer 1944) differed little in virulence, or in biochemical and serological reactions.

Nevertheless, in general, there is in each of the species we have described a variation of the same essential character as those listed above—loss of normal, type specific, surface antigen associated with loss of virulence. There are, however, variations within the S form, both natural, and induced by exposure to antisera or bacteriophage, which have no obvious connection with  $S \rightarrow R$  variation by loss. For example, Takita (1937) records a change from "specific" antigens to "group" antigens in the *Sh. flexneri* group, analogous to the diphasic flagellar variation in the salmonellae; and Kauffman (1941) records a variation by loss of one of the sub-types of XII somatic antigen in certain strains of *Salmonella*.

There are other reasons also for rejecting the hypothesis of a necessary connection between virulence and the cultural and antigenic characters of smoothness. A loss of, or a considerable change in, virulence may occur apart from the loss of the specific antigenic component characterizing the normal smooth form.

Wilson (1928) observed variants of *Salmonella typhi murium* which were both smooth and avirulent, and which must be regarded as true variants, as judged by their failure to revert easily to the virulent parent type. Borvin (1939) found that two strains of *Salmonella typhi murium*—one with a mouse M.L.D. of over 10 000 bacilli, the other with an M.L.D. of under 200, both yielded approximately the same amount of "smooth" lipopolysaccharide endotoxin, and that the two endotoxins were equally toxic and had equal immunizing power. Thus virulence may vary independently of the O antigen. Again, Shaffer, Enders and Wu (1936) described two strains of Type III pneumococcus, which were both fully capsulated, and antigenically identical. One was avirulent for rabbits, and had a greater tendency than the other, a virulent strain, to lose its capsule. The rough strains of both could be induced to revert to the smooth forms by growth in the presence of smooth killed cultures (see p. 305 below), and, whether the killed culture was of the original virulent or avirulent strain, both reverted to their original high or low virulence. The virulence, in fact, was not an expression of the degree of "smoothness" of the two strains, but dependent on some stable difference in physiological behaviour. Large changes in virulence can in fact be induced in a strain without altering its smooth characters. Thus, Hadley and Wetzel (1943), starting with a rough variant of an alpha hemolytic streptococcus, raised its virulence by serial passage through mice. The total increase in virulence during the transformation from rough to smooth was 140-fold; subsequent passage of the smooth form through mice raised its virulence 5 000-fold. It may be noted also that Bhatnagar (1940) and Jawetz and Meyer (1944) could detect no difference in the antigenic surface of virulent and avirulent strains of *Past. pestis*, though, as the latter authors suggest, in this case virulence may have been associated with an antigenic component that did not appear on the surface of the bacillus.



The exact nature of the change in colony form though it happened to give the conventional name to this particular kind of variation is clearly of quite secondary importance. We may note as a point of interest that the avirulent non specific variants of *B anthracis* and *Str pyogenes*, which happen to give smoother colonies than the normal virulent forms, have protein instead of polysaccharide components as their dominant surface antigens when in the normal virulent state.

We can, if we wish, use some term other than Smooth  $\rightarrow$  Rough to denote the particular variations but it seems undesirable to do so since they belong in all essentials to the same category. Alternatively we could abandon 'smooth' and 'rough' altogether as descriptive terms and select some new name to describe the loss of the normal surface antigen that is the essential factor concerned. It seems simpler, as we have suggested in Chapter 8 to use the initial letters of the terms 'smooth' and 'rough' to designate the variation of which the first examples observed happened to be associated with smooth and rough colony formation but to dissociate 'S' and 'R' from a designation of colony form. We should then define the 'P' variant as differing from the normal 'S' form in the following ways:

- (1) Loss of the antigenic component characterizing the surface of the bacterial cell in the normal smooth form whether this component is normally present in the form of a bacterial capsule or not
- (2) Loss of virulence partial or complete
- (3) Altered sensitivity to various bacteriophages
- (4) A change in colony form usually, but not always in the direction of increased granularity or roughness
- (5) A change in the hydrophobe or hydrophile properties of the cell usually but not always in the direction of a decreased affinity for water and a consequently increased sensitivity to the flocculating action of electrolytes

It must not be supposed that the S  $\rightarrow$  R variation represents the limit of the loss of particular antigenic components that bacterial variants may display. An excellent example of this progressive variation by loss is provided by the detailed studies which White has carried out on members of the typhoid paratyphoid group of bacteria (see White 1926, 1927, 1928, 1929a & 1931a & 1932, 1933). The polysaccharide components that characterize the surface of the bacterial cell in the normal smooth form are shared by certain types which are further differentiated from one another by the antigenic components contained in the flagella (see Table 47, p. 713). With these flagellar antigens we are not here concerned. When the normal smooth polysaccharide antigen is lost the surface of the cell is dominated by antigenic components that are shared by all members of the typhoid paratyphoid group and by some related bacteria. These include a polysaccharide component that differs from that characterizing the normal smooth form and another antigen or pair of antigens that are apparently protein in nature and have been named by White  $p_1$  and  $p_2$ . As the result of further variation the rough or R form may lose its particular polysaccharide component and then give rise to a form the antigenic behaviour of which is determined entirely by the components  $p_1$  and  $p_2$ . Situated still more deeply in the bacterial cell is another antigen which White has named  $\tau$ , but it seems doubtful whether this component is ever exposed at the cell surface as the result of loss-variation.

It may be noted that the S  $\rightarrow$  R variation, and still more the progressive loss variations referred to in the preceding paragraph, are relatively irreversible.

The  $S \rightarrow R$  variation occurs frequently under ordinary laboratory conditions of cultivation, and may be readily induced by the methods that we shall shortly describe. The reverse change ( $R \rightarrow S$ ) seldom if ever, occurs under the ordinary conditions of cultivation and it is very difficult, though not impossible to induce it by any specific stimulus, when the original  $S \rightarrow R$  change has been complete. The evidence indicates that this  $S \rightarrow R$  variation, at least as a quality of a culture as distinct from that of a component cell, is not a sudden "all-or none" process, but a gradual or step like change, so that intermediate SR forms appear between the typical S and the fully degraded R. In these partially degraded SR variants reversion to the normal S form may be more easily induced.

Prolonged growth of a normal smooth strain in any of the ordinary fluid media of the laboratory, followed by plating on ordinary agar, will usually result in the appearance of a proportion of rough, or partially rough, colonies. A bacteriophage that causes lysis of the normal smooth strain provides another, and very potent, method by which this change can be induced (see Chapter II). The contamination of a bacterial culture with a bacteriophage is not however, an entirely desirable procedure for this particular purpose, and the best method available is that introduced by Griffith (1923), who showed in the particular case of the pneumococcus, that rough variants could readily be produced by growing the normal smooth form in the presence of an antiserum acting on the type specific capsular polysaccharide. This method, the efficacy of which has been repeatedly confirmed by other workers, appears to be of quite general applicability.

The induction appears to act partly by selection. In a broth culture bacilli with a great deal of the antigen in question will be flocculated in a deposit by the antibody, and variants with none or less of it will tend to remain in suspension so that a sample from the upper part of the fluid will contain a relatively high proportion of the variants. A method that can be used with motile flagellated bacteria is growth in a semi-solid agar medium containing flagellar antibodies, those bacteria that by reason of their motility spread from the original inoculum through the semi-solid agar will tend to be those with little of the homologous flagellar antigen. By this second method Gnosspelius (1939) induced flagellated variants in diphasic salmonella bacilli possessing antigens different from the original characteristic "type" or "group" antigens. The variation was irreversible and though the new antigenic types retained their capacity for diphasic variation, it was found that only the phase which had undergone impressed variation had altered. Thus the new group phase alternated with the "original" type phase, and a "new" type phase alternated with the "original" group phase. Eriksson and Malmstrom (1939) induced a similar variation in *Salmonella newport* and found that the "new" flagellar variant had acquired an antigen of the  $\beta$  type, i.e., an  $\alpha \rightarrow \beta$  variation (see p. 716) had been induced. These organisms were originally diphasic. Starting with the monophasic *Salmonella paratyphi* A, Bruner and Edwards (1941) induced four variants: one in the original specific phase, one corresponding to the group phase of diphasic salmonellae, and two new phases with antigenic components unlike any described for the salmonellae.

It is clear that induced variations which in this case revealed not only a potentiality for diphasic variation but also two hitherto undiscovered antigens, offers a means of exploring hidden antigenic relationships within groups of bacteria.

As an example of a phage-induced variation we may cite the production from typhoid bacilli with Vi, O and R antigens of variants which have either Vi and no O antigen, or O antigen without Vi (see e.g., Hauffmann 1936 Craigie and Brandon 1936).

We are then in the change from S to R, dealing with a striking example of a variation, of a very definite type, that can be induced at will by certain specific

stimuli. It seems probable that bacteria afford particularly favourable material for this field of biological study.

Although the colonial changes associated with the  $S \rightarrow R$  variation have been described in particular detail it must not be supposed that they constitute the only variations in colony form to which bacteria are subject. Such is far from the case. We have already noted D variants and in the coli typhoid group of bacilli the occurrence of mucoid variants that clearly do not conform with the M forms in the  $M \rightarrow S \rightarrow R$  series observed in pneumococci, streptococci and the influenza bacillus. This type of variation is not infrequently stimulated when a non mucoid bacterium is submitted to the action of a bacteriophage to which it is sensitive. It seems likely that most variations associated with a change in colony form will be found to be associated also with a change in the antigenic components at the bacterial surface but not necessarily with that particular change on which the  $S \rightarrow R$  variation depends.

### The Transmutation of Antigenic Types

The loss of a specific antigenic component in the  $S \rightarrow R$  variation and its re appearance when as occasionally happens the rough variant again gives rise to the normal smooth form naturally raises the question as to whether it is possible for a rough strain to acquire the power of synthesizing not the specific antigen that characterized the smooth strain from which it was derived but some different antigen that is characteristic of another serological type belonging to the same species. Is it possible for instance to transmute a smooth Type I pneumococcus via the non capsulated rough variant into a smooth Type II or Type III pneumococcus? The problem is so important, in its biological interest and implications that the evidence must be considered in some detail.

The pioneer experiments in this field were those of Griffith (1928). He injected mice subcutaneously with living cultures of rough avirulent pneumococci mixed with large amounts of heat killed smooth pneumococci belonging to the same or another type. From the animals so inoculated smooth virulent pneumococci were frequently recovered not only was a rough strain induced to revert to the smooth type from which it was derived, but a rough variant from a Type II strain was changed to a smooth Type I strain a rough variant from a Type I strain to a smooth Type II strain rough variants of Type I or Type II strains to a smooth Type III strain, and so on.

These results were confirmed by Neufeld and Levinthal (1928) by Reimann (1929) and by Dawson (1930a, b). The study of this phenomenon was considerably advanced by the experiments of Dawson and Sia (1931) who were able to bring about a similar change *in vitro* by growing rough variants in a medium containing a heavy suspension of heat killed smooth pneumococci of the type it was desired to produce. The addition of an anti rough serum greatly assisted the transmutation but was not an essential factor. In further experiments (Sia and Dawson 1931) it was found that the transmutation could not be induced by growing rough pneumococci in the presence of purified pneumococcal polysaccharide and that heat killed smooth pneumococci obtained from old autolysed cultures or from suspensions that had been subjected to repeated freezing and thawing were unsuitable for this purpose. These results clearly demonstrated that the presence of the polysaccharide antigen belonging to a given type could not induce a rough variant to manufacture that particular antigenic component and pass on the capacity

to synthesize it to subsequent generations and they suggested that some enzyme readily liberated from the pneumococcal cells destroys some substance that is an essential stimulant of this change. Alloway (1939) was able to change rough variants of Type II pneumococci to smooth Type III or smooth Type I by growing the former on Berkefeld filtrates of extracts derived from the latter together with normal pig serum which contains anti R agglutinins. In further experiments (Alloway 1933) he substituted for the Berkefeld filtrate a preparation obtained by dissolving pneumococci of the required type in a solution of sodium desoxycholate, and precipitating the extract so obtained with alcohol. When rough pneumococci were grown in serum broth to which a saline extract of such a precipitate was added they gave rise to smooth strains of the type from which the extract was derived.

Recently Avery, MacLeod and McCarty (1944) in defining both the conditions for effecting the transformation and in making a presumptive identification of the substance inducing the transformation have made a noteworthy advance in the subject of bacterial variation. To effect the transformation a reactive variant must be selected from an irreversibly rough (R) strain of Type II pneumococcus. It is grown in broth free from as yet unidentified inhibitors, containing R anti serum free from enzymes that destroy the transforming principle, and the transforming substance itself. This substance isolated from a Type III pneumococcus was active in a dilution of  $1/6 \times 10^{11}$  appeared to be homogeneous with a particle weight of 500 000 and had the gross chemical constitution the exclusive susceptibility to a specific depolymerase and certain physicochemical properties, of a desoxyribonucleic acid.

It appears that this particular type of nucleic acid interacts with the R cell to give rise to a series of enzyme reactions culminating in the synthesis of Type III capsular polysaccharide. Once the transformation is established it is permanent, and the transformed cell continues to produce both polysaccharide and the specific desoxyribonucleate. We have in this phenomenon an outstanding example of an impressed genetic variation, i.e. a type-specific heritable mutation induced by a specific chemical agent. The mechanism of its action is unknown, but we may provisionally ascribe it to a direct effect on a bacterial gene.

#### Variations in Virulence or Toxicity

Variations in the characters on which bacteria depend for the production of disease in man and animals are clearly of particular importance to the medical bacteriologist. It would however be altogether impossible in the course of a general survey to give illustrative instances of the innumerable types of variation that are associated with some change in virulence or in toxicity. We may however note a few general principles leaving particular instances to be dealt with in the systematic description of the different bacterial species or in the chapters devoted to the diseases to which they give rise.

Taking *virulence* to mean the capacity for tissue invasion and *toxicity* to mean the power to produce a soluble toxin (see Chapter 44) we may note that these two characters depend on different factors so that where both are present in the same bacterial species they may vary independently. A hemolytic streptococcus for instance may lose its power of invading the tissues of a particular animal host without necessarily losing its power to produce a filtrable hemolysin.

We have seen that R  $\rightarrow$  S variation may be impressed on relatively rough strains of infective bacteria by passage through a susceptible animal. The varia-

tion is usually considered to be the result of selection of a few virulent organisms, which are either present in, or develop from, the injected culture

Zelle (1942), working with a strain of *Salmonella typhi* marium, has recently provided evidence of discontinuous variations of S and R forms, and their selection by the environment in the infected host. For example, an unstable S variant was shown, by a micro technique of separating cells as they divided *in vitro*, to throw stable R mutants. In one experiment, the division of an S cell, the unstable variant, into an S and an R daughter cell, was observed directly. The R mutation was observed twice in 296 divisions, a relatively high mutation rate which conforms to the hypothesis already noted, that instability is a manifestation of high mutation rates of the order of 1 per cent. Mixtures of small numbers of stable virulent variants with large numbers of less virulent stable variants were injected into mice, the organs of which after death yielded a culture with an increased proportion of the virulent variants. In other tests, virulent and avirulent variants were injected into inbred strains of susceptible and highly resistant mice. It was hoped to demonstrate a differential effect of resistant and susceptible host environments on the variants, such as a reduction in virulence of highly virulent strains propagated in the presumably less selective tissues of the susceptible mice. The only change observed, however, was an enhancement of virulence of certain variants, and this occurred both in susceptible and in resistant mice, showing that whatever the general susceptibility of the mice, any less virulent mutant, thrown by an injected strain of given virulence, was always more readily destroyed than the bacteria from which it arose.

Non toxigenic strains of *C. diphtheriae*, *C. tetani* and other normally toxigenic species have been frequently described. There is no reason to suppose that this loss of toxigenicity is in any way related to the S  $\rightarrow$  R variations, since there is no evidence that the production of a filtrable toxin is affected by the presence or absence of the smooth somatic antigen.

## REFERENCES

- ALLOWAY, J. L. (1932) *J. exp. Med.*, 55, 91. (1933) *Ibid.*, 57, 263.  
 ARKWRIGHT, J. A. (1920) *J. Path. Bact.*, 23, 358. (1921) *Ibid.*, 24, 36. (1924) *Brit. J. exp. Path.*, 5, 23.  
 AVERY, O. T. and HEIDELBERGER, M. (1923) *J. exp. Med.*, 38, 81.  
 AVERY, O. T., MACLEOD, C. M. and MCCARTY, M. (1944) *J. exp. Med.*, 79, 137.  
 BADOER, F. (1944) *J. Bact.*, 47, 509.  
 BAERTHELEIN (1912a) *Zbl. Bakt.*, 66, 21. (1912b) *Ibid.*, Ref. 54, 178.  
 BARBER (1907) *Kansas Univ. Sci. Bull.*, 4, No. 3, 1.  
 BENECKE W. (1909) *Z. indukt. Abstamm. u. Vererb. Lehre*, 2, 215.  
 BHATTACHAR, S. S. (1940) *Indian J. med. Res.*, 28, 1-17.  
 BIGGER, J. W., BOLAND, C. R., and O'MEARA, R. A. Q. (1927) *J. Path. Bact.*, 30, 261.  
 BIRCH HIRSCHFELD, L. (1935) *Z. Hyg. Infekt.*, 117, 626.  
 BOIVIN, A. (1930) *C. R. Soc. Biol.*, 132, 370.  
 BOIVIN, A. and MESROBEANU, L. (1936) *C. R. Acad. Sci.*, 203, 1402.  
 BOIVIN, A., MESROBEANU, L., MACHÉREU, G., and MACHÉREU, A. (1936) *C. R. Soc. Biol.*, 121, 169.  
 BORDET, J. and REYNAUX, E. (1930) *Ann. Inst. Pasteur*, 45, 1.  
 BORDET, P. (1930) *Ann. Inst. Pasteur*, 45, 26.  
 BRAUN, H. (1918) *Berl. Lin. Wochs.*, 55, 637.  
 BRUNER, D. W. and EDWARDS, P. R. (1941) *J. Bact.*, 42, 467.  
 BURK, A. (1908) *Arch. Hyg.*, 65, 325.  
 BURRI, R. (1910) *Zbl. Bakt.*, 11te Abt., 28, 321.  
 CHANDLER, C. A., FOTHERGILL, L. D., and DENOUE, J. H. (1937) *J. exp. Med.*, 66, 789.  
 (1939) *J. Bact.*, 37, 415.  
 CRAIGIE, J. and BRANDON, K. F. (1939) *J. Path. Bact.*, 43, 233, 249.  
 DAWSON, J. and HAPFOLD, F. C. (1943) *Biochem. J.*, 37, 389.  
 DAWSON, M. H. (1930a) *J. exp. Med.*, 51, 99. (1930b) *Ibid.*, 51, 123.  
 DAWSON, M. H. HOBBS, G. L., and OLIMSTAD, M. (1938) *J. infect. Dis.*, 62, 133.

- DAWSON, M. H. and SLA, R. H. P. (1931) *J. exp. Med.*, 54, 681  
 DESKOWITZ, M. W. (1937) *J. Bact.*, 33, 349  
 DOBELL, C. (1913) *J. Genetics*, 2, 325  
 DOLDOROFF, M. (1940) *J. gen. Hyg.*, 23, 580  
 DUBOS, R. (1940) *Bact. Rev.*, 4, 1  
 DUBOS, R. and AVERI, O. T. (1931) *J. exp. Med.*, 54, 51  
 EISENBERG, P. (1912) *Zbl. Bakt.*, 63, 300  
 FRIKSSON, E. and MALMSTRÖM, F. (1939) *Z. Hyg. Infektkr.*, 121, 629  
 ESCHÉ, P. VOR DEM (1940a) *J. Immunforsch.*, 88, 75, (1940b) *Ibid.*, 98, 333.  
 FULER, H. and CRAMER, H. (1913) *Hoppe Seyl. Z.*, 83, 430  
 EVANS, W. C., HANDLEY, W. R. C. and HARFOLD, F. C. (1940) *Biochem. J.*, 35, 207  
 GALE, E. F. (1940) *Biochem. J.*, 34, 392, 846, 853, (1941) *Ibid.*, 35, 66  
 GALE, E. F. and EPPS, H. M. R. (1942) *Biochem. J.*, 36, 600  
 GNOSPELUS, A. (1939) *Z. Hyg. Infektkr.*, 121, 529  
 GOODMAN, H. M. (1908) *J. infect. Dis.*, 5, 421  
 GRIFFITH, F. (1923) *Min. Hlth, Rep. publ. Hlth med. Subj.*, 18, (1928) *J. Hyg., Camb.*, 27, 113  
 HADLPA, P. (1927) *J. infect. Dis.*, 40, 1, (1937) *Ibid.*, 60, 129  
 HADLEY, P. and WETZEL, V. (1943) *J. Bact.*, 45, 529  
 HEGARTY, C. P. (1939) *J. Bact.*, 37, 145.  
 HEYKINGEN, W. E. VAN (1940) *Biochem. J.*, 34, 1540  
 HIRSCH, W. (1937) *J. Path. Bact.*, 44, 349  
 JACOBY, M. (1916) *Biochem. Z.*, 77, 124, 402, 405 (1917) *Ibid.*, 79, 30, 80, 307, 81, 332, 83, 74, 84, 308 (1918) *Ibid.*, 86, 329, 88, 33.  
 JAWETZ, E. and MEYER, H. F. (1943) *J. infect. Dis.*, 73, 124  
 KARSTRÖM, H. VON (1930) see STEPHENSON and STICKLAND (1932), (1937) *Ergebn. Enzymforsch.*, 7, 300  
 KAUFMANN, F. (1936) *Z. Hyg. Infektkr.*, 119, 103, (1941) *J. Bact.*, 41, 127  
 KOCHOLATY, W. and HOOGHEIJDE, J. C. (1938) *Biochem. J.*, 32, 477  
 KOCHOLATY, W. and WEIL, L. (1938) *Biochem. J.*, 32, 1690.  
 KOSER, S. A. and WRIGHT, M. H. (1943) *J. Bact.*, 46, 239  
 KOWALENKO, A. (1910) *Z. Hyg. Infektkr.*, 66, 277  
 KRUIJ, P. DE. (1921) *J. exp. Med.*, 33, 773  
 LANKFORD, C. E., SCOTT, V. COX, M. F., and COOKE, W. R. (1943) *J. Bact.*, 45, 321  
 LEONIAN, L. H. and LILL, V. G. (1942) *Science*, 95, 638, (1943) *J. Bact.*, 45, 329  
 LEWIS, I. M. (1934) *J. Bact.*, 28, 619  
 LINDEGREN, C. C. (1935) *Zbl. Bakt.*, II te Abt., 92, 40, 83, 113.  
 MASSINI, R. (1907) *Arch. Hyg.*, 61, 200  
 MILES, A. A. and PIRIE, N. W. (1939) *Brit. J. exp. Path.*, 20, 83  
 MORGAN, H. R. and BECKWITH, T. D. (1939) *J. infect. Dis.*, 65, 113.  
 MORISON, J. E. (1940) *J. Path. Bact.*, 51, 401  
 MORRIS, J. F., SELLERS, T. F. and BROWN, A. W. (1914) *J. infect. Dis.*, 68, 117  
 MORTON, H. E. (1940) *Bact. Rev.*, 4, 177  
 MORTON, H. E. and SOMMER, H. E. (1944) *J. Immunol.*, 48, 123.  
 MÜLLER, R. (1906) *Zbl. Bakt. Ref.*, 42, Beitr., 57, (1911) *Zbl. Bakt.*, 58, 97  
 NEUFELD, F. and LEVINTHAL, W. (1928) *Z. Immunforsch.*, 55, 324.  
 NUTT, M. M. (1927) *J. Hyg., Camb.*, 26, 44  
 PAPPEVHEIMER, A. M. and SHASKAN, E. (1944) *J. biol. Chem.*, 155, 260  
 PASSMORE, R. and VEDRIN, J. (1937) *Biochem. J.*, 31, 318.  
 PASTEUR, L. (1881a) *C. R. Acad. Sci.*, 92, 429, (1881b) *Ibid.*, 92, 666.  
 PENFOLD, W. J. (1910a) *Brit. med. J.*, ii 1672, (1910b) *J. Path. Bact.*, 14, 406, (1911a) *J. Hyg. Camb.*, 11, 30, (1911b) *Proc. roy. Soc. Med.*, 4 (Path. Sec.), 97, (1911c) *J. Hyg. Camb.*, 11, 487 (1912) *Ibid.*, 12, 195.  
 PREISE, H. (1904) *Zbl. Bakt.*, 35, 200, 416, 537, 607, (1911) *Ibid.*, 58, 510  
 PRIESTLEY, F. W. (1936) *J. comp. Path.*, 49, 349, *Brit. J. exp. Path.*, 17, 374  
 QCASTEL, J. H. (1937) *Enzymologia*, 2, 3  
 REIMANN, H. A. (1925) *J. exp. Med.*, 41, 587, (1929) *Ibid.*, 49, 237, (1937a) *J. Bact.*, 33, 499 (1937b) *Ibid.*, 33, 513  
 RETTOER, L. F. and SHERNICK, J. L. (1911) *J. med. Res.*, 24, 260  
 REYS, C. (1911) *Zbl. Bakt.*, II te Abt., 31, 1, (1912) *Proc. roy. Soc., B*, 85, 192  
 ROUX, E. (1890) *Ann. Inst. Pasteur*, 4, 25  
 SEASTONE, C. V. (1943) *J. exp. Med.*, 77, 21  
 SERTIC, V. and BOULGAROV, N. A. (1936) *C. P. Soc. Biol.*, 123, 901 (1937) *Ibid.*, 124, 217  
 SHAFER, M. F., ENDERS, J. F., and WIT, C.-J. (1936) *J. exp. Med.*, 64, 281  
 SIA, R. H. P. and DAWSON, M. H. (1931) *J. exp. Med.*, 54, 701  
 SILVERMAN, M. and WERKMAN, C. H. (1939) *J. Bact.*, 38, 25, (1941) *J. biol. Chem.*, 138, 35.  
 SOLOTOVSKY, M. and BUCHBINDER, L. (1941) *J. Immunol.*, 40, 243.

- STEPHENSON, M and STICKLAND, L. H. (1932) *Biochem J.*, **26**, 712, (1933) *Ibid.*, **27**, 1528
- STEPHENSON, M and LUDKIN, J. (1936) *Biochem J.*, **30**, 506
- TALITA, J. (1937) *J Hyg Camb.*, **37**, 271
- TILLET, W. S., GORBEL, W. F., and AVERY, O. T. (1930) *J exp Med.*, **52**, 835
- TODD, E. W. (1928) *Brit J exp Path.*, **9**, 1
- TWORT, F. W. (1907) *Proc roy Soc, B.*, **79**, 329
- WEBSTER, L. T. (1925) *J exp Med.*, **41**, 571
- WRITE, P. B. (1926) *Spec Rep Ser med Res Coun, Lond*, No 103, (1927) *J Path Bact*, **30**, 113, (1928) *Ibid.*, **31**, 423, (1929a) *Ibid.*, **32**, 85, (1929b) *Med Res Coun* "System of Bacteriology," **4**, 86 (1931a) *J Path Bact.*, **34**, 23, (1931b) *Ibid.*, **34**, 325, (1932) *Ibid.*, **35**, 77, (1933) *Ibid.*, **36**, 65
- WILSON, G. S. (1928) *J Hyg Camb.*, **28**, 295, (1930) *Ibid.*, **30**, 40
- WORTMANN, J. (1882) *Hoppe Seyl Z.*, **8**, 297
- WOOD, H. G., ANDERSEN, A. A., and WERKMAN, C. H. (1938) *J Bact.*, **36**, 201
- LUDKIN, J. (1937) *Biochem J.*, **26**, 1859, (1938) *Biol Rev.*, **13**, 93
- ZELLE, M. R. (1942) *J infect Dis.*, **71**, 131

## CHAPTER 10

### THE CLASSIFICATION OF BACTERIA

As has been indicated in preceding chapters, it is the behaviour rather than the nature of bacteria which has interested the bacteriologist. It is not surprising, therefore, to find that the field of systematic bacteriology has been very largely neglected. The study of bacteria has indeed never passed through that phase of detailed and accurate description, which has formed so important a part of the foundations of botany and zoology.

This neglect is not entirely attributable to lack of interest. In dealing with the morphology of bacteria, we have pointed out the difficulties which are inherent in any study of bacterial structure. As a result of these difficulties, the bacteriologist has come to rely very largely on physiological characters in the differentiation of bacterial groups, and the study of the antigen-antibody reactions has led to the elaboration of a technique which is peculiar to this field of biology. In addition to these methods of studying bacteria in artificial culture, the medical bacteriologist, who is primarily interested in the role of micro-organisms in disease, has naturally developed the habit of testing the pathogenicity of the strains he has isolated by the experimental infection of laboratory animals.

Employing a combination of these methods the bacteriologist has learned by experience to identify a large number of well-differentiated and stable bacterial types and to these he has given names. The criteria that have determined the classification and nomenclature of bacteria are not, therefore, such as would be accepted by the systematist in any other branch of biology, and the bacteriologist himself has not in general troubled overmuch as to the validity of a system which has developed rather as the result of luck than of cunning.

The inconvenience of a total absence of classification, reflected in a chaotic nomenclature, has, however, been so great, that various attempts have been made to introduce some sort of order into the bacteriological household. We cannot here enter into any historical description of the various systems which have been propounded except to note that a comparison of those suggested by Zopf (1885), Migula (1894), Kruse (1896), Lehmann and Neumann (1896) and Orla Jensen (1909) will reveal how widely the lines of cleavage may differ, when a large biological group is viewed from different angles. Those who desire more detailed information on this aspect of the question are referred to the two reports of the Committee of the Society of American Bacteriologists on characterization and classification of bacterial types (1917, 1920), the monograph by Buchanan (1925), the manual by Bergey and his colleagues (1939), and to a paper by Buchanan and others (1928), which sets out in diagrammatic form the classifications suggested by Migula, Orla Jensen, Buchanan, Castellani and Chalmers (1920), Lehmann



and Neumann, Bergey and his colleagues, and the earlier Committee of the Society of American Bacteriologists which reported in 1917 and 1920

As a result of the activities of the American Society, the whole question of bacteriological classification and nomenclature has been reopened during recent years. It cannot be said that the system propounded by the American Committee (1920) has met with the entire approval of bacteriologists in general, while systematists in other biological sciences would probably question the validity of the whole basis upon which the classification is founded. There does not however, appear to be any compelling reason for the bacteriologist to abandon for purposes of classification the criteria on which he has come to rely for purposes of identification, and few of us would be willing to admit that our systematic grouping must have a purely morphological basis simply because structural differences have been found to afford adequate classificatory criteria in the case of more highly differentiated plants and animals. While admitting that morphological differences must be given their full weight and accepting them as the natural basis for our primary subdivisions we might argue that our differential criteria depending as they do on differences in chemical structure rather than on the gross architecture of the cell, come nearer to the heart of the matter than do those adopted by botanists or zoologists.

We may note that our assessment of the significance of any particular differential criterion rests largely on a statistical basis. Our first concern is to determine the variability of a given character within a particular bacterial strain. If it is constant, it may be of value for purposes of classification. If it varies but in such a way that the variation is itself characteristic, it may still have classificatory value. If it varies in an entirely random and unpredictable fashion it cannot be used for purposes of identification or classification. Once it has been shown that a given character is of service in identifying a particular strain we can examine the distribution of this character among a sample of strains which possess other characters in common. In this way we gradually obtain a picture of the frequency distribution of many different characters among large samples of strains. The significance we attach to any particular character then depends in the main on its association with other characters. If we find that a particular group of strains resemble each other in several different characters and differ in these same characters from all other groups we feel justified in regarding the group as biologically valid and in attaching an added significance to each of the associated characters as a differential criterion within the larger group of which our homogeneous group forms a part. If on the other hand one particular character varies independently of all other characters, within a group which has many other characters in common we shall not in general attach the same significance to it, from the point of view of classification. In assessing characters in this way, we shall not of course accord all characters equal rank *a priori* and limit our consideration entirely to their frequency distribution and degree of association. Some characters will be given more weight than others and our arrangement of characters in descending order of importance will depend entirely on the function we envisage for our classification of bacteria. The classification may be predominantly utilitarian. For example as medical bacteriologists we might concentrate on the outstanding features of the medically important bacteria and ignore all other bacterial species, excepting those which are sufficiently like the medically important species to cause trouble in identification. This is, in fact the working procedure

in applied medical bacteriology, but it is not a sound basis for a classification of bacteria in general. A classification intended to accommodate the species familiar to animal, plant, industrial, biochemical and "pure" bacteriologists should depend mainly upon characters whose selection is securely based on agreed general principles. Its design should be consistent in that, should a species be discovered with a hitherto unknown combination of differential characters, it could be accommodated in the system without dislocating it. Clearly, any working classification will be a compromise between the utilitarian and the "logical" classification based on *a priori* conceptions of the relations of bacterial species.

The most promising *a priori* conception upon which to base a classification, and one which has proved fruitful in many other branches of biology, is the conception of species in a phylogenetic series. We have already discussed in Chapter 3 the relation of nutritional requirements to the possible evolution of bacterial species, and some of the dangers of a too-ready acceptance of the phylogenetic hypothesis. Our chief objection to the phylogenetic conception of the nutritional series was the impossibility of deciding which end of the series—organisms with the most complex or organisms with the least complex nutritional requirements—represented the starting point. With morphological characters there is perhaps less difficulty, for the spherical shape being the simplest and the most economical shape that could be taken by a unicellular organism (see Thompson 1942) may with some justification be taken as the primitive type. From this primitive coccus we can assume developments in the direction of aggregates of cocci, of bacteria, and of thread and mycelial forms, and in some members of each group postulate the acquisition of flagella, capsules, and other of the more striking morphological features of bacteria.

In their valuable review of bacterial classification, Kluver and van Niel (1936) in fact make this assumption the starting point of their proposed natural system of classification. Each of the main morphological groups springing from the cocci is subdivided according firstly to the main sources of energy of the bacteria, and secondly according to the most favoured substrates and their modes of dissimilation. Thus, there are the photosynthesizing autotrophs and heterotrophs, and the chemosynthesizing autotrophs and heterotrophs, four groups that are further divided according to their most favoured modes of dissimilation.

In choosing these modes, Kluver and van Niel point out that the type of attack is more important than the range of attack. Thus, the difference between an organism which splits glucose into lactic acid, and one which splits it into butyric and acetic acid,  $\text{CO}_2$  and hydrogen, is more fundamental than the difference between two organisms of the first type, one of which attacks maltose.

The principles have been further developed by Stanier and van Niel (1941), who propose the term *Monera* to cover all micro-organisms without true nuclei, plastids and sexual reproduction. Fig. 46 summarizes the proposed arrangement of *Monera*. The reader is referred to the original paper for details of their system, which may be compared with that set out on p. 319.

In our preoccupation with bacteria, parasitic on the larger animals, we are, however, concerned with distinctions that as yet do not fall into the province of the morphologist and biochemist, and while agreeing with the criteria governing the subdivisions that can be achieved in this province, we shall not find them particularly helpful with characters like pathogenicity or antigenic structure. Both are expressions of certain biochemical features in the organism, and may

ultimately fall into line with better studied features of bacterial economy, but until then we have no *a priori* grounds for evaluating the biological importance in classification of antigenic structure, or, say, the power of producing a characteristic toxin

- A Organisms photosynthetic with the evolution of oxygen and possessing the typical green plant chlorophylls phycocyanin and sometimes phycoerythrin, and colourless, non photosynthetic counterpart, clearly recognizable as such

#### Division I Myxophyta

- B Organisms not so characterized

- I Unicellular or mycelial organisms with rigid cell walls Motility when present by means of flagella Endospores, cysts, or conidia may be formed

##### Class I Eubacteria

- (a) Organisms photosynthetic, but not producing oxygen.

##### Order 1 Rhodobacterales

- (b) Non photosynthetic organisms.

##### 1 Unicellular

##### Order 2 Eubacterales

##### 2 Mycelial organisms

##### Order 3 Actinomycetales

- II Unicellular rod shaped organisms, without rigid cell walls Always creeping motility Microcysts and fruiting bodies may be formed

##### Class 2 Myxobacteria

##### One order Myxobacterales

- III Unicellular, spiral organisms without rigid cell walls Motility by means of an elastic axial filament or modified fibrillar membrane

##### Class 3 Spirochaetae

##### One order Spirochaetales

- IV Organisms not falling into the previous classes

#### FIG 46

Our main trouble is that we have no rules, and the few conventions which take their place are honoured as much in the breach as in the observance. It seems quite clear that nothing but some form of international agreement with regard to classification and nomenclature will put an end to the existing state of chaos. Whether it will be possible to adopt, in their entirety, the rules of botanical nomenclature, is a problem which only the future can decide. There are obvious advantages in adopting the Linnaean binomial nomenclature, which has served the purposes of zoologists and botanists in general, but it is doubtful whether the bacteriologist will not be forced to make frequent use of additional terms, designating races, varieties, or types. The frequent use of trinomial or quadriminomial names is, however, a cumbersome procedure, and it may be found necessary to regularize the use of letters or numbers, which is a current convention in bacteriological terminology. The Linnaean admonition, "*varietales levisimas non curat botanicus*," may serve the turn of the systematic botanist, and the bacteriologist would probably be well advised to bow to it in naming those groups

which he intends to regard as genera or species, but he cannot ignore small differences, and he needs a vocabulary which will allow him to talk or write about the bacterial types which interest him

This leads to the consideration of another difficulty, which has grown acute during recent years, and will clearly increase rather than diminish in the absence of some agreed international ruling. There is no agreement at all as to the end from which a bacteriological classification should start. Are we to begin by an intensive study of one or another relatively small group, seeking to differentiate within it all the identifiable and stable types, and giving names to these? Or are we first to differentiate the larger groups, and only when these have been adequately demarcated seek to divide them into their constituent species, varieties or types? As a method of mapping out the ground either approach will serve, but they lead, unfortunately, to quite incompatible nomenclatures. The method of the intensive study of a small or relatively small, bacterial group has been adopted by several groups of workers within recent years, and has resulted in such conspicuously successful systematic descriptions as those of the *Salmonella* group (see Chapter 30), or of the hæmolytic streptococci and the pneumococci (see Chapter 25). In each of these instances the final differentiation has depended, entirely or almost entirely, on an analysis of antigenic structure. The workers who have been engaged in the study of the *Salmonella* group have, however, adopted, at the extreme end of the scale, differentiable types listed as *Salmonella typhi*, *Salmonella dublin*, *Salmonella eastbourne*, and so on. Those who have studied the pneumococci and hæmolytic streptococci have, we think more wisely, labelled their recognizable types with numbers, or letters, or letters and numbers combined, though, except in the case of the pneumococci, there is as yet no general agreement as to the lettering or the numbering.

We do not, ourselves, think that the agreed definition of species and genera, which has still to be achieved, should be prejudged by the results obtained by antigenic analysis, particularly in the light of recent work on the sharing of antigens among species generally regarded as distinct (see Chapter 8). In the coli typhoid dysentery group, however, there are major distinctions, both antigenic and otherwise, which serve to divide the salmonella and the dysentery bacilli from the colon group of organisms in the genus *Bacterium*, and in the light of the growing number, we have accordingly assigned the enteric and food poisoning bacilli to the genus *Salmonella* and the dysentery to the genus *Shigella*.

While remaining convinced that, in naming genera and species, weight should be given to other criteria in addition to antigenic relationships, we should wish to record our entire agreement with those who, like White (1937), hold that antigenic analysis affords the best available method of differentiating the ultimate types or varieties into which bacteria are divided, that the antigenic similarities and differences provide a most valuable clue to the natural relationships of these types and the lines along which they have probably been evolved, and that each type or variety so differentiated should be given a distinctive label. We may perhaps add that this distinctive labelling is of particular importance to the medical bacteriologist, since it is the antigenic make-up of a bacterium that determines all its immunological reactions in the body as well as in the test-tube. We must, however, try to be consistent. We ought not to use letters and numbers for one set of labels, specific names for another, though current bacteriological usage compels us to do both.

Apart from the increasing importance of antigenic analysis in the classification of bacteria, it may be noted that there is a growing tendency to enlarge the range of criteria employed in the differentiation of types and species, and to rely less exclusively on the somewhat crude series of fermentation reactions that played so large a part in earlier systematic studies. The reaction of an organism to variations in the partial pressure of carbon dioxide, its resistance to various dyes its tolerance of a high concentration of hydrogen ions, all these and many other criteria are being increasingly employed in defining bacterial groups and in tracing the relation of one group to another.

There is one criterion commonly employed whose use we believe should be discontinued, from both the formal and the utilitarian standpoint namely the ecological. The relationship of an organism in its natural state to other forms of life in its environment is conditioned by many other factors besides those inherent in its own protoplasm, and in the absence of knowledge about those factors we cannot say what features of the habitat of an organism are necessarily connected with it. Consequently, habitat is on *a priori* grounds likely to be misleading as a differential character. It will be even more misleading on utilitarian grounds, since we classify bacteria in order to make precise bacteriological explorations of our environment. If, then the definition of a species includes habitat in a given type of environment, we may delay its recognition in another equally important environment. It is for example, only recently that the probable identity of *Phytomonas polycolor*, a tobacco plant pathogen and *Ps. pyocyanea* an organism infecting wounds in war, has been recognized (Elrod and Braun 1911).

One essential character of any systematic nomenclature is stability and those who have to read or write about bacteria at the present time are in a singularly unhappy position in this respect. When the same organism is masquerading as *Bacillus typhosus*, *Bacterium typhosum*, *Salmonella typhi* or *Eberthella typhi* while another answers with equal readiness to the names of *Micrococcus melitensis*, *Bacillus melitensis*, *Brucella melitensis* or *Alkaligenes melitensis* all printed in italics with a capital letter to the generic name, the student, or even the more practised reader of bacteriological literature, may be excused some degree of confusion.

It would, perhaps, be simplest to await some agreed solution of our difficulties and use the moribund nomenclature which was current before the first world war till some better system with authoritative support is offered in its stead. There are, however, real disadvantages in such a course. It is desirable, especially from the student's point of view, that a name should be as informative as possible. The scientific name of a living organism should tell us as much as possible about that organism itself, and about its relation to other organisms with different names. The latter problem is the particular concern of the systematist, and it may be many years before we know enough about the relationship of bacteria to evolve a system of classification in which those relationships can be adequately expressed. It is however, possible to allot names to bacterial groups which will give us a considerable amount of information with regard to the species, races, or types, of which they are constituted. In this respect the conventional bacteriological nomenclature of the past fifty years has been a conspicuous failure. Nothing could be less informative than the name *Bacillus*, when that name is applied to any rod shaped bacterium, and the student, who has memorized the

names *B. typhosus*, *B. pestis*, *B. anthracis*, and *B. tuberculosis*, has obtained very poor value for his effort. If adopting a more rational nomenclature, he memorizes the names *Salmonella typhi*, *Pasteurella pestis*, *Bacillus anthracis*, and *Mycobacterium tuberculosis*, he will, when he has studied the groups concerned, have a very useful picture of each of these organisms as typifying a separate genus, and the fact that some other organism is called *Salmonella enteritidis*, or *Pasteurella aviseptica*, or *Bacillus subtilis*, or *Mycobacterium phlei*, will convey to him some knowledge of its salient characteristics.

We are, ourselves, convinced that the correct approach to bacteriology, irrespective of the particular field in which the student intends ultimately to work, is to gain some knowledge of bacteria as living things, and such knowledge can most easily be obtained by grouping like forms together for the purposes of study, comparing them with other groups, and noting the differences and resemblances. For these groups we need names, even if they must, for the moment, be regarded as provisional.

This requirement can be fulfilled by adopting one of the several systems of classification and nomenclature that have been advocated within recent years. This is quite definitely a policy adopted *faute de mieux*. None of these systems has received any official or international sanction. As Buchanan, Breed and Rettger (1928) point out, neither of the systems drawn up by committees appointed by the Society of American Bacteriologists has been officially approved by that Society. The selection of one of the existing systems therefore remains a matter of personal choice, and, whichever system is selected, there is no reason to suppose that it, or the nomenclature based upon it, will receive international sanction without modification.

The terms "genus" and "species," as applied to bacteria, seem to us to defy definition, except as designations for two convenient groupings, of which the genus is the larger including group, and the species the smaller included group. For this reason, and because of the absence of any form of international agreement, we doubt the usefulness, at the present time, of naming orders, families, sub-families and tribes. Nor do we feel that the time is ripe for the creation of large numbers of genera or for the erection of an inelastic system into which all known varieties of bacteria are to be forced, each with its appropriate label.

It appears to us that the classification advocated in the final report of the first American Committee (1920) offers a carefully constructed scheme on which a useful nomenclature can be based, and that it has been designed on general lines which most bacteriologists would regard as sound, moreover, the Committee themselves make no claim to finality, and are at pains to indicate the tentative nature of some of the groupings they suggest.

We have therefore adopted the system of nomenclature set out in the final report of the first American Committee (Winslow *et al.* 1920), with a few minor modifications. We have merged the genus *Diplococcus* in the genus *Streptococcus*, since it appears to us that the pneumococcus should be included in the latter group. We have adopted the genus *Brucella*, which has already received an official recognition from many bacteriologists, since the group which contains the bacillus of Malta fever, and the bacillus of bovine abortion, appears to have as good a title to generic rank as the group which contains the plague bacillus and the bacilli causing hæmorrhagic septicæmia in animals, though we frankly admit

that the content of the *Brucella* group is extremely difficult to define. As mentioned already, we have separated the genera *Salmonella* and *Shigella* from the more inclusive genus *Bacterium*. We have combined the genera *Erythrobacillus* and *Chromobacterium* under the latter name, since it does not appear to us that the differences in morphology, physiological reactions or habitat between *Erythro prodigiosus* and such species as *Chr indicum ruber* merit separate generic rank. The final classification of these small saprophytic, chromogenic bacilli must await a more detailed study of the group, and it seems doubtful whether pigment formation should be accepted as a generic character. We have excluded the genus *Erwinia*, since the differentiation of coliform organisms of plant origin from those met with in animal tissues seems to rest on no satisfactory basis.

We append to this chapter a diagrammatic representation (Fig 47) of the classification given in the final report of the first American Committee (see Buchanan *et al* 1923) with the modifications referred to above, noting that we are concerned with the list of genera, rather than with the grouping of these genera into tribes, families or orders.

We also include in this chapter a summarized description of the characters of each genus, taken from the final report of the first American Committee (Winslow *et al* 1920), and emended in some cases in the light of more recent studies of the various groups.

In the remainder of this book, we shall employ these generic names, when referring to any species which appears to be clearly assignable to one of the listed genera. In general, bacteria which can be so assigned are already provided with a specific name, which is not in dispute. Such binomial names will be printed in italic, the generic name being given a capital letter, and used in an abbreviated form.

Other well recognized designations for various bacteria will, of course, be freely used, with the recognition that we are using the common name for a particular organism, instead of its scientific name—a practice universally followed in biological science. Such common names will be printed in ordinary type, and without a capital initial letter. For instance, the scientific name of the organism which causes tuberculosis will be written as *Mycobacterium tuberculosis*, or more shortly as *Myco tuberculosis*, but it will be generally referred to as the tubercle bacillus, similarly with *Corynebacterium diphtheriae*, *C diphtheriae*, or the diphtheria bacillus, *Pasteurella pestis*, *Past pestis* and the plague bacillus, *Brucella abortus*, *Br abortus* or the bacillus of bovine abortion, *Neisseria gonorrhoeae*, *N gonorrhoeae* or the gonococcus, *Streptococcus pneumoniae*, *Str pneumoniae* or the pneumococcus.

It is a common practice to refer to members of the *Streptococcus* and *Staphylococcus* groups as streptococci and staphylococci respectively. These terms are colloquial expressions, and we see no reason why a similar use of other generic names in the plural, or of a limited adjectival use of the generic name in the singular, should not be sanctioned. Thus, "the rickettsiae" refers to the members of the genus *Rickettsia*, "the brucellae" to the members of the genus *Brucella*, "the pasteurellae" to the members of the genus *Pasteurella*, and so forth. A salmonella antigen means an antigen met with in one or more members of the *Salmonella* genus, a proteus type of growth means a growth characteristic of members of the *Proteus* genus, and so on, but when the genus is specifically referred to, the adjective should preferably be printed with a capital and in italic, as, for instance, "members of the *Salmonella* group". The procedure is convenient, the terms

employed in most cases euphous, and the meaning with but two exceptions is unambiguous.

The two exceptions are the terms "bacillus" and "bacterium." The term bacillus means any rod-shaped organism, and the term bacterium means any organism whatever in the general class of *Schizomycetes*. Hence, neither of them is available for designating members of the two well-defined genera, *Bacillus* and *Bacterium*. In these cases we have retained the customary circumlocutions, "members of the *Bacillus* group" and "members of the *Bacterium* group."

There remain a considerable number of bacteria, which cannot yet be accorded a scientific name in some cases because the available descriptions are not sufficiently detailed to allow us to determine their systematic relationships, in others, because the characters, as described, do not seem to warrant the inclusion of the organism in any of the recognized genera. No useful purpose would be served by suggesting new generic names which would have no validity. As we have emphasized above, any system of nomenclature employed at the present time must be a temporary expedient, pending some form of international agreement, and it appears to us that our aim should be to use those names which seem most likely to be retained when such agreement is reached. When dealing with those organisms which, at present, defy classification, we have therefore frankly abandoned the use of a scientific name, with its conventional italic and capital letter, and have employed the most convenient designation available. It is unfortunate that many of these organisms have been given the generic name of *Bacillus*. As this name is reserved for the spore-bearing aerobes, it cannot be used, in the conventional form, for bacteria which do not in fact belong to that genus. We have, in general, adopted the expedient of referring to *Bacillus* x as "the x bacillus." In describing bacteria whose title to specific rank appears to us doubtful, though their generic position is not in doubt, we have in some cases employed a similar convention.

We would here add a protest against the habit, which is unfortunately frequent among medical bacteriologists, of coining new names for bacterial strains which they have isolated, without appending an adequate description of the organism or determining whether the organism in question corresponds with one that has already been described. This laxity has in the past led to much confusion, and it would be greatly to the advantage of bacteriology in general if editorial authority could be exerted to prevent the publication in medical or scientific journals of descriptions of newly named bacterial species, where these requirements are not properly fulfilled. The existence in this country, and in America, of adequate collections of type cultures provides the material for such comparisons as may be necessary.

Fig 47 provides a summary, in chart form, of the American classification, with the modifications referred to above.

The following are the genera included in this classification, given in the order in which they are listed in the chart.

**Actinobacillus.**—Gram negative non-acid fast rods sometimes occurring in long chains or in unjointed filaments. In lesions in the animal body no mycelium is formed but at the periphery finger-shaped cells or clubs may be visible.

Type species. *Actinobacillus lignieresii*.

**Leptotrichia.**—Thick, long, straight or curved threads, unbranched, frequently clubbed at one end and tapering to the other. Gram positive when young. Threads fragment



into short, thick rods. Anaerobic or facultative Non motile Filaments sometimes granular No aërial hyphæ or conidia Parasites or facultative parasites  
Type species. *Leptotrichia buccalis*

## SCHIZOMYCETES

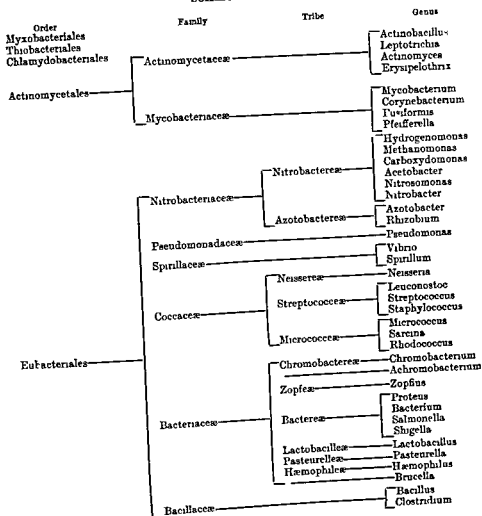


FIG 47

**Actinomyces**—Organisms growing in the form of a much branched mycelium, which may break up into segments or produce spores. Aërial mycelium often formed under suitable conditions. Mainly aerobic, but may be microaerophilic or even anaerobic. Usually saprophytic, but some species are parasitic on plants or animals, and may give rise to disease. In animal body organisms are frequently arranged in colonies composed of radiating threads with clubbed ends. Non motile. Some species are acid fast.

Type species. *Actinomyces bovis* Harz.

**Erysipelothrix**—Rod-shaped organisms with a tendency to the formation of long filaments, which may show branching. The filaments may also thicken and show

characteristic granules. No spores. Motility slight or absent. Gram positive. Slight fermentative activities. Microaerophilic. Usually parasitic.

Type species. *Erysipelothrix rhusopathiae*

*Mycobacterium*.—Slender rods which are stained with difficulty but which, when once stained, are acid fast. Cells are sometimes swollen clavate, cuneate or even branched. Non motile. Gram positive. No endospores. Growth on media slow. Aerobic. Several species are pathogenic to animals.

Type species. *Mycobacterium tuberculosis*.

*Corynebacterium*.—Gram positive rod like forms, arranged usually in a palisade. Not acid fast. Often with club-shaped swellings at the poles generally with irregularly staining segments or granules. Non motile non-sporing. Growing aerobically or under microaerophilic conditions, but often capable of anaerobic cultivation. Never forming gas in carbohydrate media, in which they may or may not produce acidity. They may or may not liquefy gelatin or serum. Some species produce a powerful exotoxin.

Type species. *Corynebacterium diphtheriae*.

*Fusiformis*.—Obligate parasites. Anaerobic or microaerophilic. Cells frequently elongate and fusiform staining somewhat unevenly. Filaments sometimes formed. Non branching, sometimes highly pleomorphic. Non motile. No spores. Reaction to Gram's stain variable, but mainly Gram negative. Growth in laboratory media feeble.

Type species. *Fusiformis termittidis*

*Pfeifferella*.—Small, slender, usually non motile, Gram negative rods, often staining irregularly, and sometimes forming threads or showing a tendency towards branching. Growth on all media is rather slow. Gelatin may be slowly liquefied, fermentation of carbohydrates is very weak. Characteristic brown honey like growth on potato.

Type species. *Pfeifferella mallei*.

*Hydrogenomonas*.—Monotrichate short rods capable of growing in the absence of organic matter, and securing growth energy by the oxidation of hydrogen (forming water).

Type species. *Hydrogenomonas pantotropha*.

*Methanomonas*.—Monotrichate short rods capable of growing in the absence of organic matter and securing growth energy by the oxidation of methane (forming carbon dioxide and water).

Type species. *Methanomonas methanica*.

*Carboxydomonas*.—Rod-shaped cells capable of securing growth energy by the oxidation of carbon monoxide (forming carbon dioxide).

Type species. *Carboxydomonas digoxycarboxiphila*.

*Acetobacter*.—Cells rod-shaped frequently in chains, non motile. Cells grow usually on the surface of alcoholic solutions as obligate aerobes, securing growth energy by the oxidation of alcohol to acetic acid. Also capable of utilizing certain other carbonaceous compounds, as sugar and acetic acid. Elongated, filamentous, club-shaped, swollen and even branched cells may occur as involution forms.

Type species. *Acetobacter aceti*.

*Nitrosomonas*.—Cells rod-shaped or spherical, motile or non-motile. Motile forms possess polar flagella. Capable of securing growth energy by the oxidation of ammonia to nitrites. Growth on media containing organic substances scanty or absent.

Type species. *Nitrosomonas europaea*.

*Nitrobacter*.—Cells rod-shaped, non-motile, not growing readily on organic media or in the presence of ammonia. Cells capable of securing growth energy by the oxidation of nitrites to nitrates.

Type species. *Nitrobacter winogradskyi*.

**Azotobacter**—Relatively large rods, or even cocci, sometimes almost yeast-like in appearance, dependent primarily for growth energy upon the oxidation of carbohydrates. Motile or non motile, motile forms possess a tuft of polar flagella. Obligate aerobes, usually growing in a film upon the surface of the culture medium. Capable of fixing atmospheric nitrogen when grown in solutions containing carbohydrates and deficient in combined nitrogen.

Type species. *Azotobacter chroococcum*.

**Rhizobium**—Minute rods, motile when young. Specialized forms abundant and characteristic when grown under suitable conditions. Obligate aerobes, capable of fixing atmospheric nitrogen when grown in the presence of carbohydrates and in the absence of compounds of nitrogen. Produce nodules upon the roots of leguminous plants.

Type species. *Rhizobium leguminosarum*

**Pseudomonas**—Rod shaped organisms, usually motile, by means of polar flagella. Generally Gram negative. Non-sporing. Aerobic, some species are facultative anaerobes. Frequently produce a water-soluble pigment, which is yellow, green blue purple or brown in colour, and which diffuses through the medium. Some species form a non diffusible yellow pigment, and some species are photogenic. Fermentation of carbohydrates as a rule not active. Frequently gelatin-liquefiers, and active ammonifiers. Common in soil and water. Many yellow species are plant parasites.

Type species. *Pseudomonas pyocyanea*

(On grounds of priority the American Committee recommend that this organism should be called *Ps. aeruginosa*.)

**Vibrio**—Short, curved, rigid rods, arranged singly or united into S-forms or spirals. Motile by a single polar flagellum, which is usually relatively short. (Some species may have two or three polar flagella.) Non-sporing. Usually Gram negative. Aerobic and facultatively anaerobic. Many species liquefy gelatin and are active ammonifiers. Commonly found in water. Most species are saprophytic, a few are pathogenic to man.

Type species. *Vibrio cholerae*

**Spirillum**—Rigid rods of spiral form, varying considerably in the number, length and breadth of the spirals. Usually motile by means of a tuft of polar flagella (5 to 20). The flagella occur at one or both poles, their number varies greatly, and is difficult to determine, since in stained preparations several are often united into a common strand. Generally Gram positive. Some species form a reddish yellow or greenish yellow pigment. Found in water or putrid infusions.

Type species. *Spirillum undula*

**Neisseria**.—Gram negative cocci, usually arranged in pairs. Strict parasites, often growing poorly on ordinary media, but growing well on serum media. Frequently pathogenic.

Type species. *Neisseria gonorrhoeae*

**Leuconostoc**.—Spherical or ovoid cells, arranged in pairs and chains, the cocci are surrounded by a gelatinous envelope, which unites them into zoogloeal masses. Usually Gram positive, but decolorize easily. Saprophytes usually growing in cane-sugar solutions.

Type species. *Leuconostoc mesenteroides*

**Streptococcus**.—Spherical or ovoid cells arranged in short or long chains, or in pairs. Non-sporing, usually non-motile. Most species Gram positive. Some species form capsules. Growth tends to be relatively slight on artificial media, and some species grow poorly in the absence of added native protein. Several species produce characteristic changes in media containing blood. Various carbohydrates are fermented with the production of acid. Most species fail to liquefy gelatin. Most species are aerobic and facultatively anaerobic, some are anaerobic. Many species are normally parasitic on man or animals. Some species are highly pathogenic, and some produce soluble toxins.

Type species. *Streptococcus pyogenes*

**Staphylococcus**—Spherical or ovoid non motile Gram positive cells arranged in grape-like clusters on solid media and in pairs, small groups or short chains in liquid media. On agar the growth is of a golden white or yellow colour. Great variation in biochemical activities, hæmolytic power, and pathogenicity. Actual or potential parasites.

Type species. *Staphylococcus aureus*

**Micrococcus**—Spherical or ovoid cells non motile arranged in pairs, tetrads or groups, but not in grape-like clusters or chains. Generally Gram positive. Grow freely on ordinary media. Sometimes produce a yellowish pigment. Gelatin liquefaction is not constant, and is usually slow. Fermentative activities weak. Usually non pathogenic to man or animals.

Type species. *Micrococcus luteus*

**Sarcina**.—Has same characters as *Micrococcus* except that cell division occurs under favourable conditions in three planes so that cubical packets are formed.

Type species. *Sarcina ventriculi*

**Rhodococcus**—Spherical or ovoid cells occurring in groups or regular packets. Usually Gram positive but are easily decolorized. Growth on agar abundant with formation of red pigment. Weak fermentative powers. Gelatin rarely liquefied. Nitrates generally reduced. Saprophytes.

Type species. *Rhodococcus rhodochrous*

**Chromobacterium**.—Small non-sporing aerobic rods usually motile and usually Gram negative, producing a yellow, red or violet pigment, which is generally insoluble in water. Saprophytic, commonly found in water or soil.

Type species. *Chromobacterium violaceum*

**Achromobacterium**.—Motile or non motile, Gram negative rods, usually small to medium in size, forming no pigment on agar, and varying in their fermentative ability. Optimum temperature for growth about 25° C, but often good growth at 37° C. Saprophytic, commonly found in water, soil, and milk.

**Zopfius**—Long rods, occurring in evenly curved chains. Gram positive. Motile. Spider-web growth on solid media. Facultative anaerobes. Carbohydrates and gelatin not attacked. Hydrogen sulphide not formed.

Type species. *Zopfius zopfi*

**Proteus**—Highly pleomorphic rods, filaments and curved cells being common in young cultures. Gram negative. Actively motile. Characteristic spreading growth on moist media. Often liquefy gelatin, and often produce vigorous decomposition of proteins. Ferment glucose and usually sucrose, but not mannitol or lactose, with production of acid and gas.

Type species. *Proteus vulgaris*

**Bacterium**—Gram negative, non-sporing rods often motile with peritrichate flagella. Some species capsulated. Easily cultivable on ordinary laboratory media. Aerobic and facultatively anaerobic. All species ferment dextrose with the formation of acid, or acid and gas. Many species are active fermenters of a wide range of carbohydrates and allied substrates. Typically intestinal parasites of man and animals, although some species may occur in other parts of the body, on plants, or in the soil. Many species are pathogenic.

Type species. *Bacterium coli*

**Shigella**—Gram negative, non motile rods, 2-3  $\mu$  long by 0.5-0.7  $\mu$  broad. Non capsulated, non-sporing. Ferment a variable number of carbohydrates with the production of acid. Lactose is not attacked except by some species, and then not for two days or more. Reduce nitrates to nitrites form ammonia but not hydrogen sulphide,

are Voges Proskauer negative, and fail to grow in Koser's citrate. Facultative anaerobes. Some species are antigenically related. One species produces a toxin. Most species are pathogenic to man, giving rise to dysentery or sometimes acute gastro enteritis. Found as a rule, in the intestinal tract of human dysentery patients and contacts.

Type species. *Shigella shiga*

**Salmonella**—Gram negative, non sporing rods, usually 1-3  $\mu$  long by 0.5-0.7  $\mu$  broad. Primarily intestinal parasites, widely distributed in man, mammals and birds. With few exceptions all species are motile, by peritrichate flagella. Easily cultivable on ordinary media. Aerobic and facultatively anaerobic. Apart from a few species that form acid only, acid and gas are produced from glucose, mannitol, dulcitol and sorbitol. Lactose, sucrose, and adonitol, and, except rarely, salicin are not fermented. Indole and acetyl methylcarbinol are not formed. Gelatin is seldom liquefied. H<sub>2</sub>S production is usual. The species are closely related to each other by somatic and flagellar antigens, and most species are diphasic. Pathogenic for man, animals, birds, or all three giving rise to typhoid poisoning, enteritis, or typhoid like infections.

**Lactobacillus**—Rods, often long and slender. Gram positive, non motile without endospores. Usually produce acid from carbohydrates, as a rule lactic. Some species grow best at 40° to 44° C., and some species are microaerophilic. Surface growth on media is poor.

Type species. *Lactobacillus caucasicus*

**Pasteurella**—Small, Gram negative, ovoid bacilli, showing bipolar staining. Aerobic and facultatively anaerobic. Powers of carbohydrate fermentation relatively slight, no gas produced. Gelatin not liquefied. Parasites in man and animals, producing characteristic infections.

Type species. *Pasteurella aviseptica*

**Haemophilus**—Minute rods, sometimes almost coccal, sometimes thread like, may be highly pleomorphic. Non motile, non sporing, Gram negative, not acid fast. Dependent for their growth on the presence of some factor, which is supplied by blood pigments, and by certain plant tissues. Some species require for their growth a second factor which is present in blood in most plant tissues in yeast, or in the cells of other bacterial species. All known species appear to be obligatory parasites, some are pathogenic.

Type species. *Haemophilus influenzae*

**Brucella**—Small, non sporing Gram negative coccobacilli. Non motile. Grow rather poorly on ordinary media or may require special media. Aerobic, no growth under strict anaerobic conditions. Growth often improved by CO<sub>2</sub>. Little or no fermentative action on carbohydrates. Usually tend to produce alkali in litmus milk, and a brown pigmentation on potato. Strict parasites, occurring in man and animals, and producing characteristic infections.

Type species. *Brucella melitensis*

**Bacillus**—Aerobic, spore-bearing rods, usually Gram positive. Often occur in long threads, and form rhizoid colonies. Form of rod not greatly changed at sporulation. Liquefy gelatin. Mostly saprophytes.

Type species. *Bacillus subtilis*

**Clostridium**—Anaerobic or microaerophilic rods, producing endospores, which are usually wider than the vegetative organisms in which they arise—so-called clostridium forms. Generally Gram positive. In young cultures often decompose protein media through the agency of enzymes, and often ferment carbohydrates. Many species are pathogenic.

Type species. *Clostridium butyricum*

## REFERENCES

- BERGEY, D. H., *et al* (1931) "Manual of Determinative Bacteriology" 5th Edit. Baillière, Tindall and Cox, London
- BUCHANAN, R. E. (1925) "General Systematic Bacteriology" Baltimore.
- BUCHANAN, R. E., BREED, P. S., and RETTOER, L. F. (1929) *J. Bact.*, 18, 387
- CASTELLANI, A. and CHALMERS, A. J. (1920) *Ann Inst Pasteur*, 34, 600
- ELROD, R. P. and BRACY, A. C. (1941) *Science* 94, 520
- KELLYVER, A. J. and NIEL, C. B. VAN (1936) *Zbl Bakt. Hte. Abt.*, 94, 369
- KRUSE, W. (1896) "Die Mikroorganismen." Flugge, Leipzig
- LEHMANN, and NEUMANN (1896) (See 1920) "Atlas u. Grundriss der Bakt. u. Lehrb., etc" 6th Edit. Munich
- MIGULA, W. (1894) (See 1900) "System der Bakterien," Bd. II, Jena.
- ORLA-JENSEN, S. (1909) *Zbl. Bakt.*, Hte. Abt., 22, 305
- Reports Comm. Soc Amer Bacteriol. WINSLOW, C. E. A., BROADHURST, J., BUCHANAN, R. E., Krumwiede C., ROGERS, L. A., and SMITH, G. H. (1917) *J. Bact.*, 2, 505 (1920) *Ibid.*, 5, 191
- STANIER, R. Y. and NIEL, C. B. VAN (1941) *J. Bact.*, 42, 437
- THOMPSON, D. A. W. (1942) "On Growth and Form" 2nd. ed., Camb. Univ. Press.
- WHITE, P. B. (1937) *Zbl. Bakt. Hte. Abt.*, 96, 145
- ZOFF, W. (1885) "Die Spaltpilze" 3 Aufl. Breslau.

## CHAPTER II

### THE BACTERIOPHAGE

IN 1915, Twort described a curious degenerative change that he had observed in cultures of a staphylococcus derived from calf lymph. He was able to transmit this change from one culture of the susceptible organism to another, by placing on the surface of an inoculated agar slope a drop of a highly diluted filtrate from a suspension of an earlier growth that had undergone the degenerative change. In 1917 d'Herelle recorded his first series of observations on the lytic properties of filtrates of mixed cultures obtained from the faeces of patients suffering from bacillary dysentery. In these preliminary studies, he was able to demonstrate the occurrence of rapid and generalized lysis in a growing broth culture of a dysentery bacillus to which some of the filtrate from the original mixed culture had been added, and the transmission of the lytic agent in a prolonged series of cultures of the susceptible bacterium, by the addition to each new culture of a filtrate obtained from the preceding one after lysis had occurred.

There can be no doubt, though d'Herelle has strenuously opposed this view, that these two descriptions afford different examples of the same essential process. With our present knowledge, we are indeed, able to identify earlier records of curious happenings and appearances in bacterial cultures as instances of this lytic change, but in none of these cases was the nature of the process studied in any detail, nor was its transmission by bacteria free filtrates demonstrated. Twort's original paper, on the other hand, contains a complete demonstration of all the essential features of this important and significant reaction, and it has hence come to be generally known as the Twort d'Herelle phenomenon."

D'Herelle's observations, which he has recorded in numerous papers from 1917 onwards and collected in three monographs (d'Herelle 1921, 1926, 1930), have, however, been far more detailed and extensive, and have played a major part in the development of our present conceptions. The name that he applied to the lytic agent, *Bacteriophage*, has come into general use, familiarly shortened to the diminutive *phage*, and the view, consistently maintained by him, that this agent is a filtrable virus, parasitic on bacterial cells, has won increasing support, particularly within recent years.

An extensive literature has grown around this subject, but we can here do no more than summarize certain of the more important observations, and the conclusions that have been drawn from them. The results obtained in recent studies have, indeed, deprived many of the earlier records of all save historical interest.

### The General Characters of Bacteriophage Lysis

The observations of d Herelle, of Twort, and of other early workers established the following facts in regard to the behaviour of the lytic agent

- (1) It will pass through filters that hold back all bacteria
- (2) It acts upon susceptible bacteria in such a way as to bring about their lysis during the phase of active bacterial growth. This lytic action may be demonstrated in several different ways

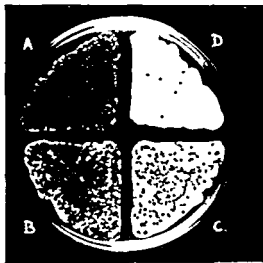


FIG 48

Effect of increasing dilutions of a phage on bacterial growth on an agar plate

few drops of a phage-containing filtrate may be spread over this inoculated surface before the plate is incubated. If the filtrate is very active and has not been diluted no growth may develop over the area on which it has been spread, or there may be a few resistant colonies (Fig 48A). With a moderate dilution of the filtrate there will result irregular, confluent areas of clearing (Fig 48B). With a still greater dilution there will be a number of well separated clear areas, each of them circular, or roughly circular, in outline (Fig 48C). These are the *taches vierges*, *plaques* or *plages*, of the French authors; the *Locher* of the German. As the phage filtrate is still further diluted these plaques become fewer and fewer (Fig 48D), a linear relation existing between the degree of dilution and the number of plaques at least over a considerable range.

Phage action may also be demonstrated on a solid medium by the appearance of "bitten," or "nubbled" colonies, of the type shown in Fig 49, or, in certain rather exceptional instances, in

(a) A phage-containing filtrate may be added to a broth, or peptone-water, culture of a susceptible bacterium, the addition being made either at the time the broth is inoculated, or during the early stages of bacterial growth. The addition of an active filtrate will result, after a variable period in a relatively sudden clearing of the turbid growing culture. This clearing may be partial, or apparently complete, but, even in the latter case, prolonged incubation usually results in renewed bacterial growth.

(b) The surface of an agar plate may be thickly inoculated with a susceptible bacterium, so as to give a confluent growth, and a



FIG 49

Colony of *Esch* coli on agar showing a "bitten" segment at the periphery due to the action of a bacteriophage ( $\times 8$ )



the occurrence of a vitreous, or granular, degeneration in colonies that have already attained a relatively advanced stage of growth

(3) The lytic agent may be propagated indefinitely in association with growing cultures of the bacteria on which it acts. A few drops of a filtrate from the first culture in which lysis has occurred may be added to a second young growing culture of the bacterium, this may be filtered after lysis and the filtrate added to a third freshly inoculated culture and so on in series. A phage filtrate may be active in very high dilution ( $1 \times 10^{-8}$  or higher), and in successive passages, carried out as above the original titre is often increased during the earlier transfers, and is then maintained. It is clear, therefore, that the lytic agent, whatever it may be is actively reproduced during the lytic process. There are few instances of its reproduction in the absence of bacterial cells or in the presence of dead bacterial cells (see Krueger and Baldwin 1937). Under certain conditions however it can be produced in the presence of bacterial cells that though living are not growing or dividing. There has been some controversy on this point (see Otto and Muntz 1923, Twort 1925, 1926, Gratia and Rhodes 1926, Gohs and Jacobsohn 1927, Bronfenbrenner and Muckenfuss 1927) but recently Krueger and his colleagues have defined conditions in which phage appears to be produced without growth of the bacterial cell (Scribner and Krueger 1937, Krueger and Fong 1937, Northrop 1939).

(4) Any given lytic agent will be found to be most active against one bacterial species or type, or against a few species or types that are known to be related to one another. Against unrelated species or types, there is usually no action. The phage is, then a highly specific agent.

(5) Phage lysis has been observed in a large number of bacterial species. It has also been reported in yeasts and actinomyces (Wiebols and Wieringa 1936).

(6) A secondary result of phage action is the appearance, in the bacterial culture that is undergoing lysis of variants that are resistant to the action of the particular phage concerned. These variants are usually susceptible to the action of other phages.

(7) A large number of bacteria are known to carry a lytic agent to which they are themselves resistant. These phage resistant carrier strains are called lysogenic. Lysogenic strains are indistinguishable from others except for the fact that they release an agent which is actively lytic for phage sensitive bacteria. A lysogenic strain, and the bacteria sensitive to the lytic agent it carries usually belong to the same species or group of micro organisms.

(8) One of the commonest natural habitats of the phage is the intestinal tract of man and animals, and filtrates active against one or more bacteria can almost always be obtained from any specimen of faeces, or from material that has been subjected to faecal pollution.

### The Nature of the Bacteriophage

As we have seen, d Herelle has consistently maintained the view that the phage is a filtrable virus, parasitic on bacteria. This hypothesis has always been in accord with many of the most striking features of phage behaviour (see Flu 1923, Reichert 1924, Schuurman 1925). There can be little doubt that it would have gained early and general acceptance but for the fact that certain recorded observations seemed almost irreconcilable with it.

Because of these difficulties in accepting the virus hypothesis various alternative theories have been propounded.

Katushima (1930) suggested that the phage was a catalyst, activating a pro-ferment present in the bacteria themselves. On this view the liberation of the ferment in an active form would of course have to be regarded as an essential consequence of the lysis of the bacterial cells, the process, when once set going, being self-reproducing.

Bordet (see Bordet and Ciucu 1920; Bordet 1923, 1925) sought to reconcile the conception of an inanimate phage with its reproduction in an unlimited series in a rather different way. He suggested that the phenomenon was a true autolysis, the active agent being produced exclusively from the bacteria themselves. The origin of the autolysis he traced to a disturbance of the normal equilibrium between the assimilative and metabolic activities of the bacterial cell, adding the supposition that the substances set free during the autolysis of the cells initially affected were able to act in some way upon susceptible but hitherto unaffected cells, and to initiate in them the same series of autolytic changes. Once started the process would thus be transmissible in series, provided that susceptible bacteria were present and that these bacteria were metabolically active.

Northrop (1939) emphasized the greater likeness of many of the characteristics of phage production and action to the production and action of enzymes, and in support of his thesis demonstrated the similarity in the production of an extracellular gelatinase and of phage from a lysogenic strain of *B. megatherium*.

Another hypothesis, advanced by Hadley (1937, 1938) and since put forward in a rather different form by Wollman (1925, 1937, 1938, 1939, 1934a & b, 1935) [see also Wollman and Wollman 1937 and Lwoff 1936], assumes the phenomenon to be purely bacterial in origin but relates it to the genetic not to the metabolic activities of the bacterial cell. The phage on this view would be regarded as analogous to some gene-carrying constituent of the bacterial cell, or some filtrable phase in a complex life-cycle. It must, of course, be assumed that the addition of this cellular component or bacterial phase to a young culture of susceptible organisms so alters their genetic behaviour that they undergo lysis during the process of multiplication, and, in so doing, reproduce the active agent in large amount. As further evidence for their view Wollman and Wollman (1936, 1938) maintain that each living cell of a phage-carrying strain of *B. megatherium* liberates one and only one particle of phage: if the phage were an externally infecting virus, it is argued, the number present in a carrier cell should vary from cell to cell. Gratus (1936) and Flu (1938a) working with the same strain of bacterium have both been unable to elicit the one-to-one ratio described by the Wollmans, both find the number of particles liberated can exceed the number of bacterial cells (see also Lewis and Worley 1936).

There are, we think, adequate reasons for accepting the virus hypothesis, at least as the most probable explanation of all the recorded facts. In setting out the evidence that seems to us to justify this conclusion it will be convenient to discuss, *seriatim*, certain aspects of the nature and behaviour of the phage: but before doing so one point may be made clear since it will be involved in our consideration of each other question in turn.

D. Herelle, for reasons that are not easy to appreciate, has upheld the view that the phage is a single living organism, *Protobius bacteriophagum*, which may adapt itself to live at the expense of a wide variety of different bacteria. The very extensive evidence that is now available is quite incompatible with this view. There is not one phage but an enormous number of different phages: and in order to obtain constant and reproducible results it is as necessary to work with pure strains of phage as with pure cultures of bacteria. Neglect of this precaution has rendered many recorded studies of very doubtful value.

Pure strains of phage may be obtained in two ways. Since a given strain of

bacterium will in many cases be susceptible to only one of the phages contained in a crude mixed filtrate, repeated transfer in growing cultures of this bacterium will often eliminate all phages except the one to which it is sensitive. This method is not, however, always reliable. A crude filtrate will often be found to contain more than one phage acting on the test bacterium selected. In such a case each of these phages may be propagated in successive transfers. It often happens that one or more are reproduced more rapidly than the others, and so have a better chance of transfer, but such phage cultures may remain mixed throughout a number of generations. An alternative, and better, method is to pick from isolated plaques on agar plates, just as we pick isolated colonies in attempting to purify a mixed bacterial culture. By a combination of these two methods it is usually possible to obtain phage strains of undoubted purity, but the procedure may be laborious, and contamination is very liable to occur. For these reasons strict attention to details of technique is required in any work of this kind (see Isheshov *et al* 1933a, Rakićen and Rakićen 1937).

The ways in which various strains of phage can be differentiated from one another will be considered in later sections of this chapter. At the moment we are concerned only with the point that such strains exist, and are identifiable.

#### The Physical State of the Phage. Is it Particulate or in Solution?

Given a bacteria free filtrate containing active phage we may determine whether it behaves as a homogeneous solution or whether the activity is distributed in a discontinuous fashion throughout the solution. The answer will depend entirely on the sensitivity of the method of examination. The question therefore "Is phage particulate?" must be understood in the restricted sense "Is the lytic activity in the solution continuous or discontinuous?" Should it prove to be continuous, the fact is not evidence that phage is not particulate in the general sense of the term but merely that the technique was too insensitive to detect the separate units in action.

As d'Herelle showed isolated plaques form on agar cultures over which a high dilution of phage filtrate has been spread. As further evidence that the phage activity was discontinuous d'Herelle noted that when a very high dilution of phage filtrate was distributed in equal volumes among a large number of tubes, each containing a young growing culture of a sensitive bacterium lysis occurred in some tubes but not in others indicating that the active agent was not uniformly dispersed in solution. This observation has been confirmed by a number of subsequent observers (Bronfenbrenner and Korb 1925b, McKinley and Holden 1926, Bronfenbrenner 1927) and both Feemster and Wells (1933) and Luria (1940) have shown that the proportion of tubes showing lysis among large numbers inoculated with varying amounts of phage filtrate is in accord with statistical expectations based on the assumption that the agent is dispersed in lytic particles.

A possible escape from the obvious implication of these observations was provided by the assumption that only a few of the bacteria in any culture were susceptible to phage action. This assumption had little inherent probability and could hardly account for the test-tube experiments referred to above but it was accepted by some observers, mainly on account of other observations that seemed to tell against a grossly particulate nature. Thus Bordet (1923) stated that it was impossible to concentrate a phage filtrate by centrifugation at high speeds. We now know that this can readily be done if the speed is adequate. Olsen and Yasaki (1923) stated that the phage was volatile. This would have finally disposed of the view that it was particulate, or a living agent of any nature,

but the results recorded were undoubtedly due to technical errors (Spät 1924, Borchardt 1924 Gildemeister and Herzberg 1924b Meissner 1924b, Gercke 1925, Bronfenbrenner and Korb 1925a)

The size of the lytic particle in a number of phage preparations has been determined and it has been found that different strains of phage are each characterized by a certain relatively narrow range of particle sizes.

The size has been estimated in various ways. Filtration through membranes of approximately known pore sizes indicated diameters of the order of 20-50  $m\mu$  (Stassano and de Beaufort 1925 Bechhold and Villa 1926, Zinsser and Tang 1927) Elford and Andrewes (1932) used carefully graded collodion membranes and showed that different pure strain phages had each a different particle size and that size was independent of the particular strain of bacterium upon which they were being propagated. One phage for instance, had a mean diameter of 8-12  $m\mu$ . The diameters of a group of phages lay between 50 and 75  $m\mu$  while other strains gave intermediate values. Yaot and Sato (1935) recorded similar results. Schlesinger (1932-33a 1933) Elford (1936), McIntosh and Selbie (1937) and Pale, Krassnoff, Haber, Reimé and Voet (1938) estimated the diameter of phage particles by measuring their rate of sedimentation in a high-speed centrifuge. The results were consistent with those observed by filtration and confirmed the existence of a wide range of particle-size. Wollman and Lacassagne (1940) estimated a similar range of sizes from the results of inactivation of phages by X radiation (see also Luria and Exner 1941)

Reference to Chapters 2 and 41 will show that the larger phages are big enough to be demonstrated by modern microscopical methods. Merling Eisenberg (1933) and Eisenberg Merling (1941) obtained photomicrographic images in visible light of staphylococcus and *Bact. coli* phages which on photometric measurement yielded estimates of size similar to those obtained by ultrafiltration and ultracentrifugation studies. Barnard using the technique of photography by monochromatic ultra violet light, has obtained photographs of a phage of this type which show it to be composed of uniformly sized particles with a diameter of about 50  $m\mu$  (see Burnet 1933c). These large phages produce in lysed broth cultures a turbidity that can be detected by its Tyndall effect, and the intensity of this effect has been found to provide an accurate measure of the concentration of the phage in the filtrate (Schlesinger 1932-33b, Schuurman and Schuurman ten Bokkel Huinck 1936)

Electron micrographs reveal not only the size of phage particles but also a structure. Ruska (1941) noted "sperm shaped" particles in his photographs, and more recently Luria Delbruck and Anderson (1943) have shown that three of four *Bact. coli* phages they studied had an opaque "head" consisting of a pattern of granules, about 80  $m\mu$  in diameter, and a less opaque "tail" about 120  $m\mu$  long (Figs. 50 51, 52)

Our conception of the lytic particle of a size that closely characterizes the various types of phage as the ultimate unit of phage, does not accord with recent descriptions of certain physical and serological properties of *Bact. coli* phage. By diffusion experiments with dilute solutions Kalmanson and Bronfenbrenner (1939) deduced a particle weight of 1,500 000 for a coli phage. The particle weight of the corresponding lytic unit was 27 times as great. Again, the maximum amount of antibody that combined with a lytic unit of phage was found to be far more than could be accommodated on the surface of a simple lytic unit. By postulating sub units of 1,500 000 particle weight, the quantitative serological data fall into line with those from reactions of other antigen antibody systems (Hershey, Kalmanson and Bronfenbrenner 1943). The lytic unit appears to be made up of about 30 independent diffusing units, bound by 25 per cent. of carrier

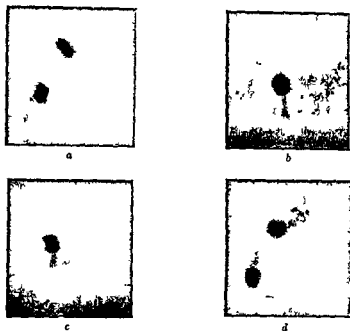


FIG. 50—Electron micrographs of a *Doct col* phage ( $\times 30,000$ )  
(From a photograph kindly supplied by Dr. S. E. Luria)

protein into an aggregate that in concentrated solutions behaves in lysis in the photographic field in the ultracentrifuge and in the ultrafilter as a unit particle. The titration of lytic activity by the plaque method thus reveals only one in thirty of the sub units. The lytic unit readily dissociates into diffusing particles also containing carrier protein which in turn are made up of ultimate particles having a particle weight of 50,000. If these facts are confirmed then the chemical descriptions of phage as a nucleo-protein (see Schlesinger 1934, Northrop 1938) may prove to be descriptions of carrier protein only. The identity of the protein isolated from phage preparations with phage itself has already been questioned on technical grounds (see Moriyama and Ohashi 1937, Flu 1938b).

The justifiable inferences from all these observations take us far beyond the conclusion that the phage is particulate. They accord well with the view that phages are filtrable viruses. They accord badly with any other hypothesis that has yet been propounded. A particulate lytic agent might conceivably consist of particles of bacterial protoplasm on to which some active principle had been adsorbed. If so the adsorbing material and the nature of the complex formed must be specific for each phage in a high degree or the particles resulting from disruption of a bacterium by one type of phage would be of varying sizes and would be unlikely to differ consistently in size from those produced by the action of another phage.

#### Other Characters of the Phages

Most phages within the pH range over which they remain active appear to carry a negative electric charge in this way resembling bacteria and filtrable viruses (Todd 1927, Krueger *et al.* 1929, Burnet and McKie 1930a, Natarajan and Hyde 1930).

The fact that phages will grow only in the presence of multiplying or living bacteria renders the study of their metabolic activities exceedingly difficult. Apart from the work of Schüler (see Editorial 1935) which suggests that phosphatase is the only hydrolytic enzyme possessed by the bacteriophage, such studies as have been recorded have, in fact, yielded negative or ambiguous results (Bronfenbrenner 1921-25, 1926, Gozony and Suranyi 1925, Kauffmann 1925-26, Bronfenbrenner and Reichert 1926-27, Schwartzman 1926-27, Bachmann and Wohlfeil 1927).

The resistance of various phages to heat and to other physical or chemical agents has been studied in some detail. Certain early statements ascribed to lytic filtrates a degree of resistance to heat, and to such chemical agents as chloroform, toluene, alcohol, acetone, etc., that seemed to place them in a category apart from other living things except the spore-bearing bacteria (see Kabeshima 1920). These statements played a not inconsiderable part in the controversy as to whether the phage was a living or non living agent, but they were not in accord with all the records existing at the time—Twort (1915) for instance, had stated that his lytic agent was inactivated by heating at 60° C for 1 hour—and they have since been shown to have no general validity. All the evidence suggests that different strains of phage like different species of bacteria differ from one another in their resistance to heat, to drying and freezing (see Knorr and Ruf 1931, Campbell Renton 1941) to shaking (Campbell Renton 1942) and to various other physical and chemical reagents, and that the range of sensitivity, taken as a whole is much the same in the two cases.

A few observations of general interest in regard to this particular problem may be briefly noted. Baker and Nanavutty (1929) found that the time relations of the inactivation of a Shiga phage by ultra violet light were closely similar to the killing of *Bact. coli* by the same agent. The time required to inactivate the Shiga phage or a staphylococcal phage did not differ greatly from that required to kill *Bact. coli*, but the time required to inactivate trypsin was 20-30 times as long, and for the inactivation of diastase about 120 times as long. In this respect therefore the phages studied behaved as living things, not as 'ferments' (see also Gates 1934). Wright and Kersten (1937) and Campbell Renton (1937) noted a wide variation in the sensitivity of different phages to ultra violet light.

Schultz and Krueger (1938) recorded the inactivation of a staphylococcus phage by methylene blue. Clifton (1931) found that this inactivation occurred when the mixture was exposed to sunlight but not in the dark, and concluded that it was due to the oxidation of the phage by the photodynamically activated dye. Perdrau and Todd (1933), using more exact methods record similar results. Different phages have been found to vary widely in their sensitiveness to this photodynamic action, though none is entirely resistant (Burnet 1933b, 1934). Staphylococcus phage may also be inactivated by safranine, the inactivation is partly photodynamic (Krueger and Baldwin 1935).

Phage is inactivated by aldehydes and the inactivation reversed by the addition of substances with free amino or imino groups which compete with the phage for the aldehyde (Hendall and Colwell 1938, Hügler and Oleinik 1943). The activity of the phage appears to depend in part upon certain of the free  $-NH_2$  or  $-NH$  groups on its surface. Levin and Lominski (1936) record reversible inactivation of phage by lecithin, which appears to prevent access of phage to sensitive bacteria. Williams, Sandholzer and Berry (1940) record the inactivation of a phage by cholesterol and cephalin, by bacterial and certain non bacterial phospholipins but not by lecithin, sphingomyelin or plasma phospholipins.

A well marked antagonistic action on the viability of phages, of Ca, Mg, Ba and Sr ions on the one hand, and of Na and K ions on the other can be demonstrated (Burnet and McKie 1930a, Sertic 1937a, Gratia 1940) suggesting that these ions are acting on a protein constituent at the surface of a living cell. Diminution in phage counts are also

observed after treatment of the phage at 50° C with various anions (Ohashi 1934). The inhibition of lysis by which inactivation is measured may be due to an action, not on the phage particle itself, but on the sensitive bacterium. For instance, Geat (1943) has shown that with high concentrations of univalent cations, like Na, and moderate concentrations (0.1 molar) of Mg ions, plate lysis may be completely masked by overgrowth of resistant bacteria, which are apparently produced in large numbers in the presence of the cations. Some of the larger phages are rapidly inactivated by a strong urea solution a reagent that acts similarly on many bacteria and viruses (Burnet 1933b, 1934).

It has been shown by several workers that some, but not all, phages are unable to produce lysis in bacterial cultures growing in a medium from which the calcium ions have been removed by previous treatment with citrate (Stassano and de Beaufort 1923, Bordet 1926, Asheshov 1926, Burnet 1933b). This fact allows a useful differentiation of phages into 'citrate sensitive' and 'citrate resistant' strains, but it should be noted that an initially citrate sensitive phage may often be trained to produce lysis in a citrate containing medium.

### The Mechanism of Phage Lysis

In considering the mechanism of phage lysis we may first deal with the phenomenon in a general sense, leaving the problem of specificity for later consideration.

d Herelle's conception of the lytic process, based of course on the view that the phage is a parasitic virus, is simple and straightforward. He believes that a phage particle enters a growing bacterial cell, multiplies within it, and causes its more or less explosive disintegration when the limit of distension has been reached. This limit appears to vary over a considerable range, but disruption usually occurs when the number of particles in the cell have reached some figure between 6 and 60. In support of this view, d Herelle states that the addition of very small amounts of phage to a growing culture is followed by a step like increase in phage titre during the earlier stages of growth, successive sudden increases occurring at intervals of 20 to 30 minutes. After a few such jumps in titre the phage concentration rises logarithmically until lysis occurs. This change from a discontinuous to a continuous rise would, of course, be expected, since the successive increases would soon get out of step, and their combined effect would give a steady rise in titre.

Burnet (1929c) has recorded observations that are in entire accord with d Herelle's view. To a number of small tubes each containing a young actively growing culture of a sensitive bacterium, he added a phage filtrate so highly diluted that, on the average, each tube received a single particle. At short intervals thereafter the whole contents of one of the tubes was spread on the surface of an agar plate, and the resulting plaques were counted. The results showed that there was no detectable increase during the first 20 minutes or so. After this time there was a sharp and sudden rise. Thus, a series of tubes plated at one minute intervals, over the appropriate time range, gave 1, 0, 1, 2, 0, 1, 80, 0, 1, 120, 1, 230, 0 and 100 plaques. The absence of values intermediate between 1 and 80, makes it clear that the free phage in the culture was not increasing by twofold steps, as would occur during the early generations of a single bacterium multiplying by binary fission, and the only obvious explanation of such findings is that the phage particles are dividing in, or on, a bacterium, and are suddenly liberated into the surrounding fluid when that bacterium disrupts. These observations have been fully confirmed by Ellis and Delbruck (1939) and Hershey and Bronfenbrenner (1943).

The process of phage lysis has also been studied quantitatively by observations carried out on a lysing culture as a whole (see, for instance, Lepper 1923). Krueger and Northrop (1931) determined the rates of growth of staphylococci and staphylococcal phage in a lytic mixture. After a short lag period the concentration of bacteria and phage both increased steadily up to the moment of visible lysis. The rate of increase of phage was consistently greater than that of the bacteria. Clifton and Morrow (1936) demonstrated

a similar relationship in a lytic mixture of *Bact. coli* and a coli phage. In both cases, massive lysis was initiated when the number of phage particles per bacterial cell reached a certain value. In the *Bact. coli* system it was 1/600. The steady rates of phage production as Burnet (1934) points out are not incompatible with the conception of liberation of phage in bursts from infected bacteria but may well represent the statistical average of large numbers of sudden bursts occurring at different times. Krueger and Fong (1937) later showed that the relationship they had observed between bacterial count and phage count was peculiar to their conditions of measurement, which happened to have been near the optimum both for phage production and bacterial growth, and, as we have already noted they were apparently able by altering these conditions to demonstrate increase of phage in the absence of bacterial multiplication. This important observation, that phage is liberated in large amounts from infected bacteria in the absence of bacterial multiplication has been repeated by Spizizen (1943a) but in view of the large number of previous negative reports, it may be wise to await its further confirmation.

We may distinguish three stages in the production of phage namely adsorption of phage on the bacteria (Fig. 51) a period of constant phage count in which phage grows in or on the bacteria and release of phage when the phage count rises (Fig. 52).

**Adsorption.**—The rate of adsorption depends on the concentration of both phage and bacteria. The adsorption capacity of a bacterial species for a phage to which it is sensitive varies with the strain (see for instance Hershey and Bronfenbrenner 1943) and the physiological state of the bacterial cell. Delbruck (1940) found that an actively growing culture of *Bact. coli* adsorbed 200 particles per cell while the cells of starved culture of the same strain adsorbed only 20 particles per cell. When a phage filtrate is adsorbed by excess of bacteria a certain amount of residual phage is left which was regarded by Krueger (1931) as evidence that adsorption was reversible. An alternative view (Schlesinger 1932-33a, b and Delbruck 1942) regards the residual phage as having lessened affinity for the bacterium.

**Constant Period.**—The constant period varies in the same way that division of bacteria varies with temperature (Ellis and Delbruck 1939).

**Release of Phage.**—At the end of the constant period the release of phage occurs steadily for a certain period during which all the infected bacteria one after the other are lysed, and then ceases a step being followed by a halt in the curve of phage production. The released phage is adsorbed to other bacteria, and in due time a second step release occurs, and so on each succeeding step being less clearly marked as the cycles of adsorption on growth and release in each infected bacterium overlap one another in time. Within certain limits the constant period and the size of the step release are independent of the number of phage particles adsorbed and it appears that if more than one particle is adsorbed, only one of them actively induces the production of more phage. Lysis of the growing culture as a whole occurs when the phage concentration reaches a threshold limit for the number of bacteria present. In his studies of *Bact. coli* phage systems Delbruck (1940) demonstrated in bacteria, at a given stage of growth a close similarity of the adsorption capacity of the bacterium the threshold limit for lysis, and the yield of phage at the end of the constant period and suggested that each bacterium might produce as many phage particles as it had receptors to which adsorption could take place. Using a similar system Hershey and Bronfenbrenner (1943) did not observe any simple relation between phage yield and adsorptive capacity.

It may be noted that phage particles, when present in adequate concentration, are able to kill bacteria without producing lysis. Andrewes and Elford (1932) have demonstrated this direct killing action by studies with a citrate sensitive coli phage. Mixtures of this phage with a sensitive *Bact. coli* when plated on citrate agar gave normal growth without lysis, so long as the phage concentration was below a certain limit. But when this limit was exceeded 90-99 per cent. of the *Bact. coli* were killed in the sense that they failed to develop on the citrate agar, although there was, of course still no lysis.





FIG 51

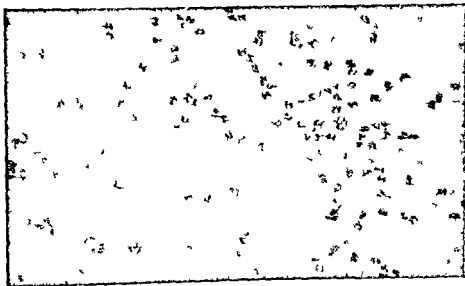


FIG 52

FIGS 51 and 52—Electron micrographs of phage action

FIG 51 Adsorption of phage to the surface of *Bact coli* ( $\times 54\,000$ )

FIG 52 The release of phage from a lysed *Bact coli*. The outline of the cell wall and the bacterial debris are seen at the left of the picture ( $\times 21\,000$ )

(From micrographs kindly supplied by Dr S. E. Luria.)

It should also be noted that although the multiplication of phage particles and the characteristic lysis associated with it demand the presence of living bacteria the phage is readily absorbed by dead bacterial cells (see Krueger 1931). As we shall see in a later section this absorption is specific.

Apart from these quantitative data, it is, of course, possible to observe the occurrence of phage lysis under the microscope, and this has in fact been done (see d'Herelle 1921, 1926, Twort 1922, Wollman 1925, Burnet 1925, v Preisz 1925, Manninger 1926, da Costa Cruz 1926, Bronfenbrenner *et al* 1927, Bronfenbrenner 1928, Bayne-Jones and Sandholzer 1933). Not all of these observations have been made under the best modern conditions of microscopy or photomicrography, so that their value, particularly in regard to matters of detail, varies considerably. They differ, also, in certain particulars of the actual happenings that are described, and there can be little doubt that phage lysis may, in fact, occur in several different ways, according to the nature of the phage and bacterium involved.

Swelling of the bacterial cells is usually seen in the early stages of the process, and is sometimes accompanied by the development of granules. The disruption of the swollen bacterial cell is usually very sudden. The cell may appear to explode, giving place to a cloud of granular debris, or it may lose its outline and disappear, without the liberation of the granular cloud.

Merling Eisenberg (1938) (see also Eisenberg Merling 1941) was able to make direct observations on the disruption of invaded bacteria, he found that 150-200 phage particles were liberated from cells of *Bact coli* and 2-4 from cells of *Staph aureus*. Delbrück (1940) distinguishes two forms of lysis, lysis from without and lysis from within. Lysis from without occurs almost instantly when phage is adsorbed at the threshold limit. The cells swell into spherical bodies. Lysis from within occurs in *Bact coli* after the adsorption of one or a few particles, but not until the phage has multiplied to the threshold limit. The cells appear to fade more or less quickly. Electron micrographic studies of *Bact coli* undergoing phage lysis (Luria, Delbrück and Anderson 1943) gave precise pictures of the adsorption of phage, lysis of the cell, and liberation of phage particles from the interior of the cell which were numerically in accord with the results obtained by concurrent phage counts of the infected cultures studied. They also revealed that in multiple infection of a cell the adsorbed particles did not seem to enter the cell, but remained attached to the cell wall.

The mechanism of the actual disintegration of lysed bacteria is not understood. In some cases it is doubtful whether such disintegration occurs, at any rate at the moment when the turbidity of the culture disappears. With some phages the absence of microscopically apparent disintegration at this moment suggests that the main change is an approximation of the optical character of the bacteria to that of the suspending medium.

The liberation by phage of a lytic enzyme at the time of lysis was postulated by d'Herelle (1926). Sertic (1929, 1937b, Sertic and Boulgakov 1939) observed a halo round each plaque formed by certain coli phages in an opaque growth of *Bact coli*. The halo was due to the induction of variants of the *Bact coli* used, by a "lysin" which would pass an ultrafilter, the variants grew as a transparent film. The addition of the phage free lysin to a mixture of *Bact coli* and a phage to which it was insensitive resulted in lysis. The lysin apparently destroyed certain antigenic constituents of the insensitive *Bact coli* thereby creating a variant of different sensitivity (see also Schuurman 1936). Evans (1940) was able to enhance the activity of streptococcal phages on resistant strains by adding a small quantity of a susceptible strain to the mixture, she explained the reaction in terms of nascent phage. It is probable that she was dealing with a lysin similar to that of Sertic. Campbell Renton (1937) observed a related phenomenon when phage filtrates inactivated by ultra violet light were tested on sensitive organisms: there was a diffuse lysis of the bacteria without the formation of plaques. In both these cases the lytic agent appeared to be derived from the bacteriophage. This view was also supported by Gratia (1937). On the other hand Bronfenbrenner and Muckenfuss (1927) obtained a lytic enzyme from both normal and phage lysed cultures of staphylococci. Wollman (1934a) upheld the bacterial origin of lytic enzymes. Phage action can be enhanced

lytic enzymes other than those derived from bacteria or phage. Lysozyme for instance, will enhance the action of cholera phage (White 1937).

Pirie (1939) observed an increase in reducing sugars and a decrease in precipitable carbohydrate during lysis by coli phage. No enzyme was detected in the phage itself but the coli bacillus yielded an enzyme that attacked the bacterial cellular polysaccharides. Though no direct connection exists between this phenomenon and lysis by the phage the association of lysis with the enzymic destruction of those cellular elements known to adsorb phage is at least suggestive. In a later paper (1940) Pirie records the release of adsorbed phage from *B. megatherium* by treatment with lysozyme and the destruction of the bacterium's capacity to absorb phage by preliminary incubation with lysozyme. The lysozyme hydrolysed a carbohydrate present in *B. megatherium* and as Pirie suggests it is possible that carbohydrate is one that determines the specific absorption of the phage.

### Phage Production and Phage Precursor

The rapid production of phage from bacteria following the adsorption of one or a few phage particles has suggested to some workers the possibility that the substances of bacterial origin from which the phage is derived exist as a precursor requiring only a few comparatively small transformations to convert it into phage. The analogy of the activation of a pepsinogen by a pepsin has been cited in this connection. It breaks down however in one important respect for with the activation of enzyme precursors by similar enzymes, it is the nature of the precursor which determines the enzyme produced. Thus chicken pepsin converts swine pepsinogen into swine pepsin and swine pepsin converts chicken pepsinogen into chicken pepsin (Herriott Bartz and Northrop 1938). A bacterium on the other hand, may be attacked by several distinct phages and each phage reproduces itself. If a single phage precursor is postulated it must then be sufficiently primitive in form to yield a variety of phages when suitably activated. Krueger and Baldwin (1937) made a cell free extract of staphylococci in the presence of which the titre of a phage filtrate doubled in 2 hours. In staphylococci this phage precursor was more easily destroyed than either the cells which produced it or the phage into which it was converted. Apparently protein in nature its activity was reduced by antistaphylococcal serum, by heating to 45° C for 20 minutes by iodoacetic acid, and by light in the presence of methylene blue. It was not liberated from cells disintegrated by sonic vibrations (see Krueger, Scribner and Mccracken 1940, Krueger, Brown and Scribner 1941).

Spizizen (1943b) has opened up a new field in the study of phage precursors by applying the methods elaborated by workers on bacterial nutrition to the study of phage production by *Bact. coli* in phosphate buffer. Several amino acids, certain phosphorylated compounds (including nucleic acid and co-enzyme I) 4-carbon dicarboxylic acids and ferric, ferrous magnesium and manganese ions stimulated phage production. Glycine and glycine anhydride were particularly effective in this respect, inducing a marked increase in phage in the absence of bacterial multiplication. The production of phage was even greater if the organisms were exposed to the glycine anhydride for several hours before the addition of the infecting dose of phage, suggesting that in the interval the cells either had adapted themselves to a more rapid glycine metabolism, or had built up reserves of a phage precursor. It appears that certain fundamental metabolic processes in the cell are required for phage multiplication but not necessarily all the processes that lead to cell multiplication. The specificity of some of these processes is illustrated by the fact that the action of glycine was specifically inhibited by the

addition of its sulphone derivatives, amino-methane sulphonc acid, and that the inhibiting action displayed by sulphanilamide was antagonized by *p*-aminobenzoic acid (see Chapter 6)

### The Factors that Determine Phage Specificity

It has been well established, from the earliest days of bacteriophage study, that any given strain of phage has a relatively limited range of activity. A coli phage, for instance, will not act on a staphylococcus, or on a diphtheria bacillus. It was soon found, however, that phage specificity, while varying widely in range with different strains of phage, might be much narrower than this. A coli phage would act on some strains of *Bact. coli*, but not on others, and so on. The obvious assumption was that different strains of the same bacterial species differed in "phage sensitivity," but no attempt was made by the earlier workers to determine on what factors this sensitivity depended. A very significant advance in our knowledge of this problem has been made by the studies of Burnet and his colleagues.

In a study of the activity of different strains of phage on the species and types included within the *Salmonella* group of bacteria, it was found (Burnet 1927) that there was a close relation between the sensitiveness of different bacterial species, or types, to a particular phage, and the distribution of particular surface somatic antigens. Thus, a particular phage was active against *Salm. typhi*, *Salm. enteritidis* and *Salm. pullorum*, all of which possess the somatic antigen IX, but was inactive against *Salm. paratyphi A* and *Salm. cholerae-suis* which do not possess this antigen. If, however, the normal smooth strains of these species were replaced by their rough variants in which the smooth somatic antigens are lost, their place being taken by a common rough somatic antigen, then a phage that attacked one rough variant would attack all the others. Some phages were found to attack only the normal smooth forms, their range of activity then being determined by the distribution of the various smooth somatic antigens. Other phages attacked only rough forms. A few phages were able to attack both rough and smooth forms (Burnet 1929a).

Further study revealed similar correspondence between antigenic structure and sensitivity to various phages. Schmidt (1931) noted that different species of salmonella bacilli could be differentiated by their sensitiveness to different selected phages. In some cases the correspondence is exact, in others less so. Lately it has proved possible to extend the range of phages specific for different antigenic varieties of certain bacterial species by adapting phages to antigenic types for which at first they show little affinity. For example, following the discovery that the action of certain phages on *Salm. typhi* depended on the presence of the Vi antigen of Felix and Pitt (1934) (see Sertie and Boulgakov 1936a, Scholtens 1936, 1937, Craigie and Brandon 1936a), Craigie and Yen (1938) were able to develop phages with selective action against 11 different varieties of *Salm. typhi*. In a similar manner Felix and Callow (1943) were able to adapt naturally occurring phages active against Vi forms of *Salm. paratyphi B*, and so extend the range of phages selective for different varieties of the bacilli. It will be obvious that once the correspondence between a bacterial and phage type is established, bacterial strains may be identified and assigned to a given phage type by tests with known strains of phage.

In many cases, a group of bacteria, homogeneous by serological tests, proves on phage testing to consist of a number of stable sub types. The recognition of these sub types, as Craigie and Yen first demonstrated, adds greatly to the precision of bacteriological surveys for epidemiological purposes. (See also Helmer, Kerr, Dolman and Ranta 1940, Lazarus 1940, 1941, Dolman, Kerr and Helmer 1941, Desranleau 1942, Hutchinson 1943, Felix 1943.)

There are a number of bacterial species in which the relationship of antigenic structure

and phage sensitiveness has been recorded, in the Flexner group of dysentery bacilli (Burnet and McKie 1930b, Claiberg and Marcuse 1932 Radojčić 1936 Miller 1937, Wheeler and Burgdorf 1941), among rough and smooth dysentery strains (Denys 1932) among variants of a *Bact coli* strain (Sertic and Boulgakov 1937), among rough and smooth strains of *V cholerae* (Asheshov *et al* 1930 1933b), in *Salm paratyphi C* (*cholerae suis*) (Levine and Frisch 1936 Frisch and Levine 1936), in Group A streptococci (Evans 1936 1940, 1942, Evans and Verder 1938 Evans and Sockrider 1942), among certain serological types of *C diphtheriae* (Keogh Simmons and Anderson 1938), in *Staph aureus* (Burnet and McKie 1929 Williams and Timmins 1938 Fisk 1942a b) and in mucoid strains of *Bact coli*, *Bact aerogenes* and *Bact friedlanderi* (Rakieten, Eggerth and Pakieten 1940).

It should be noted that these varieties of antigens and combinations of antigens associated with the specificity of phages are somatic antigens. Sertic and Boulgakov (1936b) however, record a phage which acted only on flagellated forms of *Salm typhi*.

The intimate relationship between phage sensitivity and the nature of the surface bacterial antigens has been confirmed by observing the effects on phage lysis of different bacterial extracts. Levine and Frisch (1934) tested the fractions obtained by alcoholic precipitation of saline extracts prepared from bacteria of the *Salmonella* and *Shigella* groups and found that they specifically inhibited the lysis of the homologous organisms by phage. Gough and Burnet (1934), working with more highly purified fractions, record similar results. The fractions obtained in this way were known to consist mainly of the specific polysaccharide somatic antigens. Still further confirmation has been obtained by the demonstration by Levine and his colleagues (Levine Frisch and Cohen 1934 Levine and Frisch 1935) that heat killed bacteria of the *Salmonella* group absorb phages specifically, in general accordance with their antigenic structure as revealed by agglutination tests.

Working with staphylococcal phage Rakieten, Rakieten and Doff (1936) demonstrated that autolysates of sensitive staphylococci inhibited phage action but those of resistant cocci did not. The same specificity was displayed by heat killed staphylococci. Freeman (1937) found that crude preparations of the specific polysaccharide of a *Staph aureus* strain inactivated a phage to which it was sensitive (see also Burnet and Freeman 1937). The rates of inactivation of a phage by culture filtrates are not compatible with the view that total inactivation follows simply from combinations of phage and inhibitory substances (Ellis and Spaizen 1941), there appear to be two competing processes, one producing inactivated the other partly inactivated phage and full inactivation is achieved only by circumstances favouring one process at the expense of the other (see below Phage Antiphage Reaction).

There can, then, be no doubt at all that the antigenic structure of the bacterial surface is one of the main factors in determining the accessibility of the bacterial cell to a given strain of phage, and we must suppose the phage particles to be so constituted that those of one strain can find attachment, or entry, only at an area of bacterial surface that is characterized by a particular antigenic structure, while those of another strain have wider potentialities and are able to attack a bacterium through more than one type of antigenic surface.

It should, however, be emphasized that this is not the only factor that determines phage sensitivity, or, at least, that there is not complete parallelism between sensitivity and antigenic structure as revealed by the available serological tests. Burnet (1929b) for instance notes that a phage resistant strain, appearing as the result of the lytic action of a particular phage on a particular sensitive bacterium may show exactly the same antigenic structure, as revealed by serological tests as the sensitive strain from which it was derived. He notes further that the resistant strain not only shows no lysis, but fails to adsorb the phage, so that some change

in surface structure would appear to be concerned, though it is not detectable by the ordinary serological methods.

### The Variations Induced in Bacteria in Response to Phage Lysis.

In an earlier section of this chapter, it was noted that a secondary bacterial growth usually appears in a culture that has undergone phage lysis. If this growth is further examined, it will be found to be composed of bacteria that are resistant to the strain of phage that caused the lysis, though they will usually be sensitive to other strains of phage.

Sometimes these resistant strains show a well marked antigenic difference from the original sensitive strain, for instance, the lysis of a smooth strain of a dysentery bacillus by an anti smooth phage is frequently followed by the appearance of characteristically rough resistant variants. But, as we have seen, resistant strains are not always of a recognizably different antigenic type from the original sensitive strain, and, in the great majority of cases, we have at present no evidence as to whether the change in phage sensitivity has, or has not, been accompanied by a change in antigenic structure. What we do know is that submission to the action of a phage is one of the most potent methods of inducing bacterial variation (see Arkwright 1924, Hadley 1927, 1928), that the variants so produced may be of many different kinds, that they share the character of resistance to the phage that induced the variation, and that some at least of them differ antigenically from the parent strain.

The kind of variation induced is not necessarily similar to variations which, like those noted in Chapter 9, occur in other circumstances. These latter variations may be indeed inhibited by the presence of phage. For instance, Lewis and Worley (1936) noted that spontaneous dissociation of *B. megatherium* occurred much less promptly in the presence of phage than in its absence.

It is clear that the existence of phage resistant strains of bacteria, and the possibility of producing them at will, afford a method of studying in greater detail the phages acting on single bacterial species.

Bail (1923) was the first to stress the importance of cross resistance tests between different phages and different bacterial strains as a method of phage differentiation, and many others have since employed this method in the study of phages acting on particular groups or species of bacteria such as the *Salmonella* group (Burnet 1926b), the *Shigella* group (Burnet and McKee 1930b, Morison 1932), or the cholera vibrios (Asheshov *et al* 1930, 1933b, Morison 1932). Many of these studies, however, were concerned with differences in phage sensitivity between different bacterial species or types, as well as between different variants derived by phage action from a single bacterial strain. It is with differences of the latter type, and their application as a method of separating and identifying different phages that are active against a single bacterial species or type, that we are here concerned. This method has been very extensively developed by Asheshov and his colleagues, and we may take an illustrative example of an experiment performed according to their technique (Asheshov *et al* 1933b).

Suppose that we have three strains of phage, all acting on the same strain of a given bacterium, but which we suspect, for one reason or another, to belong to different types. We allow each of the three phages, which we may label I, II and III, and a mixture of them (I II III), to act on four separate cultures of our sensitive bacterium growing in a fluid medium. We allow lysis to occur, and a secondary growth of resistant bacteria to follow it. We take a large agar plate, and mark it off into 16 squares ( $4 \times 4$ ). Over each square in the first column of 4 squares we make a thick seeding from the secondary growth from our Type I phage, over each square in the second column we seed the secondary

growth from our Type II phage, over the squares in the third column the secondary growth from our Type III phage, and over the squares in the fourth and last column the secondary growth from the culture containing all three phages. Then we take a filtrate containing Type I phage, and spread a small drop of it over a circular area in the middle of the top square of each column (i.e. of each square of the first row). In each square of the second row we make a similar inoculation of Type II phage, in each square of the third row an inoculation of Type III phage, and in each square of the fourth and last row an inoculation of phages I, II and III.

If the phages actually belong to different types, as judged by this test, the results will be as shown in Fig. 53, in which the dark circles indicate the occurrence of confluent lysis over the circular area inoculated with the phage.

It will be noted that Phage I acts on the secondary growth from Phages II and III but not on the secondary growth from Phage I and so on. None of the three phages acts on the secondary growth from the culture submitted to the action of all three phages. The mixture of three phages acts on the secondary growth from each of the three separate phages.

If we desire to test another phage, to see whether or not it is identical with any of our three Types I, II, or III, we include it in such a series of tests, adding the additional row and column to our squares. If it is identical with any of our three types it will behave as that type does. If it is a new type (say Type IV) then it will lyse the secondary growths from Types I, II and III and its own secondary growth will be lysed by each of these types.

This leads us to a consideration of other ways in which phages may be differentiated from one another. One of these is the method of antigenic analysis, which has been so extensively employed in the identification and classification of bacteria.

#### Phages as Antigens The Phage-Antiphage Reaction.

It was shown by Bordet and Cruca (1921) that the phage is antigenic and that an antiphage serum has the power of neutralizing the phage and so preventing its lytic action on bacteria. Numerous workers have since studied this phenomenon, and it has been clearly shown (a) that the inhibition of phage lysis is due to antibodies acting on the phage itself, and not to the antibacterial antibodies that are usually present in an antiphage serum, (b) that the antibodies are specific for particular types of phage, phages thus showing an antigenic specificity of exactly the same kind as that displayed by bacteria, but (c) that the antibodies have no action on the bacteria from which the phage arose, that is, phage and bacterial host are serologically distinct. Many of the papers already referred to contain

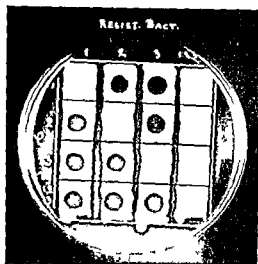


FIG. 53

Differentiation of phage types by their action on resistant bacteria

references to the antigenic behaviour of the phages studied, as well as to their other properties

The studies of Burnet and his colleagues clearly show the close analogy of the serological reactions of phage with those of bacteria. Bacteria which have been allowed to adsorb large amounts of a particular phage, will absorb the antibody from a homologous specific antiphage serum, from which all antibodies acting on the bacteria themselves have been removed. The treated bacteria are agglutinated by the antiphage serum (Burnet 1933c).

Filtrates from phage preparations contain soluble substances that inhibit the reaction of the phage with its antibody and will to some extent reverse a phage-antibody union that has already taken place (Burnet 1933d, Burnet *et al.* 1937). The inhibitory substance here is analogous with the specific soluble substance that characterizes many species and types of bacteria. Suspensions of large-particle phages free from bacteria and bacterial debris are agglutinable by antiphage sera (Burnet 1933c, Merrill 1936) and the agglutinating power of the serum for various related phages runs parallel with its power of phage-neutralization.

The quantitative relations between a phage and its homologous neutralizing serum were first studied in detail by Andrewes and Elford (1933a, b) who found that, over a considerable range of phage dilutions, a given amount of antiserum neutralized a constant percentage of the phage present irrespective of the number of phage particles exposed to its action (see also Clifton, Mueller and Rogers 1935). This certainly indicates that the particles are inactivated individually and not by aggregation, for if aggregation was effective in reducing the phage-count in the serum-phage mixture we should expect a greater percentage drop in the count as the concentration of phage and therefore the likelihood of collisions of sensitized phage particles, increased. The results suggest also that as we have seen in other antigen-antibody systems (Chapter 7) the phage or the antibody or both may be heterogeneous and that in any phage preparation a certain percentage of the particles either unite feebly with antibody, or are in some way insusceptible to inactivation after union (see also Halmanson and Bronfenbrenner 1942). The validity of these interpretations is questioned by Hershey and his colleagues (1943, 1944) on the grounds that there is little direct, independent evidence of heterogeneity of either antibody or phage. They have observed two types of antibody effect in a coli phage. Thus with a given amount of homologous antiserum the phage was sensitized, and its infectivity thereby increased, with double this amount the phage was neutralized and became non-infective. The variations in activity displayed by phage-antiserum mixtures may therefore be interpreted in terms of a homogeneous phage varying proportions of which are unaltered, increased in infectivity or decreased in infectivity, according to the degree of union with an essentially homogeneous antibody.

Though there are obvious parallels between inactivation of phage by adsorption to bacterial extracts, and by antiphage sera it does not appear that the two inactivators unite with the same part of the phage particle (Burnet and Freeman 1937, Burnet *et al.* 1937) for antigenically similar phages lyse antigenically unrelated bacteria, and phages for antigenically similar bacteria may themselves be antigenically unrelated. Moreover by adaptation the affinity of a phage for a bacterium may be profoundly modified without any alteration in antigenic quality. Nevertheless, the active sites of union of antibody and bacterial inactivator appear to be close together on the phage particle for union with antibody blocks union with bacterial inactivator, though in some cases the phage is only partly neutralized by the antibody producing lysis on the prepared plate after a delay which may represent the time required for a freeing or a redistribution of the antibody on the phage particle making the bacterial receptor again accessible. The variation observed in the degree of neutralization and in the rate of the phage-antibody reaction with temperature (Hershey 1941) suggests that the surface of the phage particle



varies in reactivity, so that not all collisions of antibody and phage in the neutralization mixtures are fruitful (Kalmanson, Hershey and Bronfenbrenner 1942). Even with fully neutralized phage, there is no evidence that union of antibody with phage alters it permanently, even after prolonged contact (Kalmanson, Hershey and Bronfenbrenner 1942). After 56 days' contact with antibody at 37° C., Kalmanson and Bronfenbrenner (1943) were able to reactivate a just neutralized phage by digesting the inactive phage with papain. The digestion apparently removed only a portion of the combined antibody molecules but in doing so made the bacterial receptors fully accessible.

The relations of phage inactivation by antisera and by bacterial antigens are summarized in Fig. 51, which is modified from a scheme devised by Delbrück (1942).

REACTION	RESULT
$B + P \rightarrow BP$	Adsorption of phage to bacteria
$b + P \rightarrow bP$	Adsorption of phage to free bacterial antigen
$BP + B$	No lysis or delayed lysis according to degree of phage inactivation
$B + \beta \rightarrow B\beta$	Antigen antibody union
$b + \beta \rightarrow b\beta$	Antigen antibody union
$P + \pi \rightarrow P\pi$	Antigen antibody union
$I\pi + B$	No lysis or delayed lysis according to degree of phage inactivation
$I\pi + b$	No adsorption of phage to free bacterial antigen
$BP + \pi \rightarrow BP\pi$	Antigen antibody union, agglutination of phage coated bacteria by antiphage serum

FIG. 51. The reactions of phage inactivation by bacterial antigens and specific antiphage sera.

$B$  = bacterium

$b$  = antigenic component of bacterium associated with specific adsorption of phage

$\beta$  = antibody to  $b$

$P$  = phage with affinity for  $B$

$\pi$  = antibody to  $P$

### Other Methods of Phage Differentiation, and the Identification and Classification of Phages

The phage plaques that have so frequently been referred to in preceding sections have characters that are of great value in the differentiation of one phage from another. For any one phage, acting on any one sensitive bacterium the plaques formed on an agar plate are usually closely similar, both in size and in form. But different phages acting on the same bacterium may give plaques that differ sharply and characteristically from one another.

To take first the character of plaque size the existence of large plaque forming phages and small plaque forming phages has long been recognized and used as a basis for differentiation (see Bail 1923). Elford and Andrewes (1932) were able to show that there is an inverse relation between plaque size and phage size. The small phages give big plaques and *vice versa*. It is an obvious assumption that the small phages can diffuse more readily through the surface bacterial growth, and so extend over a larger radius. The probability of this assumption is increased by the demonstration by Andrewes and Elford (1933b) that a large plaque forming phage incompletely neutralized by an antiphage serum and probably aggregated into small clumps, gives small instead of large plaques.

But size is not the only character that differentiates one phage plaque from another, there are often marked differences in the edge of the circular clearings

These may be entire or eroded sharp or bevelled. They may, or may not be surrounded by a halo in which the bacterial growth is altered in appearance, though not completely lysed. Phage plaques, indeed, present variations in morphology as distinctive as those shown by bacterial colonies, and have the same classificatory

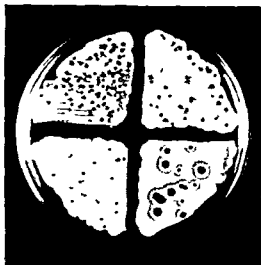


FIG. 55

Different types of plaque produced by different phages acting on the same bacterial culture

value. Asheshov (1924) (see also Asheshov *et al.* 1933b) has paid particular attention to these differences in plaque morphology, and Fig. 55 illustrates the differences in size and form, that may be observed among the plaques produced by different phages acting on the same strain of a sensitive bacterium.

If, then, we seek to classify any particular group of phage strains, we can make use of the following series of tests:

(A) The determination of the species of bacteria against which each phage is active. This will enable us to differentiate broad groups—coli phages, dysentery phages, salmonella phages, staphylococcal phages, cholera phages, and so on.

(B) By more detailed tests of cross resistance we can subdivide these broad groups, each into a number of different types—so many different types of dysentery phage, so many of cholera phage, and so on.

(C) We can apply to the same problem of the subdivision of our broad groups the method of antigenic analysis, using specific antiphage sera.

(D) We can study the size and morphology of the plaques formed, and divide our original groups of phages into large-plaque-forming strains, small plaque-forming strains, and so on.

(E) We can also apply certain other biological tests, such as relative resistance to heat, or to the photodynamic action of methylene blue, or to the presence of citrate in a medium.

When, in fact, we apply several of these tests, our confidence in each of them is increased by finding that the results they give are highly correlated. The classification derived from detailed resistance tests corresponds closely with that derived from neutralization tests with antiphage sera. The phages producing a particular type of plaque when acting on a particular sensitive bacterium, are usually found to fall into the same antigenic group. An admirable series of studies in which this correlation is brought out very clearly have been recorded by Burnet (1933a, b) and other evidence all pointing in the same direction, will be found in the studies of Asheshov and his colleagues (1930, 1933b), in papers by Sertic and Boulgakov (1935a, b) and in many of the other records referred to above. The correspondence is not of course, absolute. Just as some groups of bacterial strains would be divided into the same species or types, whether classified on the basis of

fermentation tests or antigenic structure, while other groups that are identical in regard to their fermentation reactions may be divided into different types on the basis of serological tests so one particular method of analysis may serve to separate strains of phage that would be grouped together when tested by another. But correlation of characters is as frequent among the phages as it is among the bacteria, and leaves no reasonable doubt that, by making full use of the methods now at our disposal, we can separate and identify phage types that have just as much claim to be regarded as biological entities as have the various species or types of bacteria.

#### Variation and Adaptation in Phages

The literature contains very numerous records of phage variation and phage adaptation. D. Herelle, as has been noted, regards the phage as a single virus that may become adapted to attack a wide variety of bacterial species. Many of the earlier accounts of adaptation must be discounted because the technique adopted did not ensure the purity of the original filtrates. There are on the other hand numerous observations which show quite clearly that adaptation occurs though its range is probably more limited than was at one time supposed. Here again the evidence suggests that the phage behaves in the same way as other known micro organisms.

We may note, for example, that d. Herelle and Rakietyen (1935) adapted a staphylococcus bacteriophage, initially sensitive to neutralization by antiphage serum, to lyse cocci in the presence of high serum concentrations. Burnet and Lush (1936) observed a mutant from a *Staph. albus* phage. The two varieties were similar, both serologically and by cross tests on resistant survivors (see preceding section), though one had less lytic power than the other. But in one case the resistant strain was lysogenic and in the other, not, and infection of a culture with the weak variant protected it against lysis by the strong variant.

#### The Ecology of Phages

The problem of phage ecology raises points of the greatest biological interest and importance. We have already noted that phages acting on one or other of the normal or pathogenic intestinal bacteria can almost always be isolated from faeces, from sewage or from polluted water supplies (see for instance, Sonnenschein 1927, Stewart and Ghosal 1931, Gildemeister and Watanabe 1931, Schlossmann 1932, von Vagedes and Gildemeister 1934, De and Paul 1940, de Assumpção and Leite e Silva 1942, Guelin 1943). This clearly suggests that wherever particular species of bacteria occur in nature, there also are likely to be found phages to which these bacteria are sensitive. But that is merely the first, and least interesting step in the problem.

Just as it was tacitly assumed, in the earliest days of medical bacteriology, that a pathogenic bacterium, when it became parasitic on its natural host would always, or almost always, cause the specific disease of which it was the causal agent, so it was assumed in the early days of bacteriophage studies that a phage, if it became attached to, and multiplied in association with, a particular bacterium would cause phage lysis. The conception of phage carriers among bacteria, or of a normal phage flora living symbiotically with certain bacterial species, came relatively late.

The demonstration that a phage active against a particular species of bacterium could sometimes be isolated from an old laboratory culture of that organism by repeated filtration and the addition of each successive filtrate to a fresh broth culture of the bacterium

in question (Gildemeister and Herzberg 1923, Kuttner 1923, Kline 1937, Hadley 1928, Fukuda 1928, Klieneberger 1932) was at first regarded as a powerful argument against the virus theory, and in favour of the view that the phage was a product, enzymic or other, of the bacteria themselves. It was indeed the failure of many workers to confirm these findings (Ogata 1924, Reichert 1924, Meissner 1924a, Arkwright 1924, Sonnenschein 1927) rather than any realization that they were quite compatible with the virus theory, that deprived them of much of their force. It is quite probable (see Burnet 1934) that some, at least, of the positive findings were due to technical errors. But in the light of our present knowledge it would be by no means surprising if many of them were correct.

Our present conception of the real nature of phage-bacterium parasitism has been determined largely by the proved existence of what are known as "lysogenic strains" of bacteria.

The classical examples of these lysogenic strains are the *Bact. coli* strain, which was first described by Lasbonne and Carrere (1922) and later studied by Bordet and Renaux (1928), and another *Bact. coli* strain which was studied by Gildemeister and Herzberg (1924a). Both these strains, although they themselves show no evidence of phage lysis, regularly yield filtrates which lyse *Sh. shiga*, i.e. they are permanent carriers of a phage to which they are themselves resistant, but to which *Sh. shiga* is sensitive.

Such strains were, for a time, regarded as bacteriological anomalies, but such a view is no longer tenable. Here, again, our present concepts have been largely influenced by the studies of Burnet (1932, 1934).

In a careful study of 34 strains of *Salm. enteritidis* he found that 27 were lysogenic, in the sense that they yielded a phage to which they were themselves resistant, but which produced transmissible lysis in other, specially susceptible bacterial strains. From these 27 lysogenic strains three different types of phage were obtained: A, B, and D, fourteen strains yielded phage B, seven phage D, two A and B, and one A and D. Phage A was also frequently isolated from lysogenic strains of *Salm. paratyphi A*, *Salm. paratyphi B* and *Salm. typhi murium* while phages B and D and another phage N were sometimes present in strains belonging to these species. Such results as these, as Burnet points out, are incompatible with the view that the lytic agent in these lysogenic strains is some component of the genetic apparatus of the bacterial cell.

In the case of *Salm. paratyphi C*, on the other hand, all strains examined, whether they had been isolated in Russia, South America, or the East Indies, were found to be carrying a single antigenic type of phage. A similar state of affairs may, as Burnet points out, exist in the case of *C. diphtheriae*, since Smith and Jordan (1931) found that every strain examined was lysogenic when tested against a single sensitive indicator strain of the same species. The occasional occurrence of 'muddled' colonies that have been noted by many workers in certain bacterial cultures represent, in Burnet's view, lysogenic strains in which the resistance of the carrying bacterium is slightly unstable.

We must, then, if we are to accept the virus hypothesis, to which all other evidence clearly points, also accept the view that symbiosis between phage and bacterium is an exceedingly common event—so common that it would at the moment be unwise to assert that any bacterial strain was certainly not carrying phage.

What place this symbiotic process will eventually take in our conceptions of bacterial structure, bacterial variation, antigenic behaviour, and other similar problems we cannot yet prophesy. It may be a relatively minor one, but it may not.

(For some of the methods of unmasking lysogenic strains, see Flu 1936b, Lominski 1938, Fisk 1942a)

We may note here one of the few instances of a geographical distribution of a phage. White (1937), examining a number of strains of *V. cholerae*, found a certain phage in all strains isolated in India, but in none of several strains of Chinese or Japanese origin.

In the carrier state represented by a lysogenic strain (see Burnet 1934) it is clear that the multiplication of phage and bacterium must be so co-ordinated that, when a bacterium divides, each daughter cell receives its quota of phage. It has in fact been shown (den Dooren de Jong 1931, Cowles 1931) that when a lysogenic strain is a spore bearer, the phage is present in the spore and in the new generations that arise from it. It is also noteworthy that in such strains the phage appears to share the heat resistance of the host, being relatively resistant in the spore, and susceptible when in the vegetative cell. Wollman and Wollman (1939) regard this fact as strong evidence for the bacterial origin of phage.

One other aspect of the ecology of phages must be mentioned, namely, the animal origin of phage. Certain workers concluded that the passage of various bacteria through the intestinal tract of warm-blooded animals results in the appearance of phages not previously present either in the animal's intestine or in the cultures administered. Neither Naito (1936), who worked with hens reared from the egg on sterile diets, nor Glaser (1938), who used aseptically reared flies, could find support for this notion. The recorded cases appear to have been due to insufficiently sensitive methods for detecting phage. Ohishi (1939) for instance, obtained phage more easily from the viscera than from the faeces or intestinal mucosa of mice.

The possible relation of phages to bacterial infections in man and animals is considered in Chapter 54.

## REFERENCES

- ANDREWES, C. H. and ELFORD, W. J. (1932) *Brit. J. exp. Path.*, **13**, 13. (1933a) *Ibid.*, **14**, 367. (1933b) *Ibid.*, **14**, 376.  
 ARKWRIGHT, J. A. (1924) *Brit. J. exp. Path.*, **5**, 23.  
 ASHESHOV, I. N. (1924) *J. infect. Dis.*, **34**, 536. (1926) *C. R. Soc. Biol.*, **94**, 687.  
 ASHESHOV, I. N., ASHESHOV, I. KHAN, S., and LAHIRI, M. N. (1930) *Indian J. med. Res.*, **17**, 971. (1933a) *Ibid.*, **20**, 1101.  
 ASHESHOV, I. N., ASHESHOV, I. KHAN, S., LAHIRI, M. N., and CHATTERJI, S. K. (1933b) *Indian J. med. Res.*, **20**, 1127.  
 ASSUMPTIÃO, L. DE and LEITE E SILVA. (1942) *Arg. Hig. Saude publ.*, **14**, 99.  
 BACHMANN, W. and WOHLFEIL, T. (1927) *Zbl. Bakt.*, **104**, 256.  
 BAIL, O. (1923) *Z. Immunforsch.*, **33**, 57.  
 BAKER, S. L. and NARAYAN, S. H. (1929) *Brit. J. exp. Path.*, **10**, 45.  
 BAYNE JONES, S. and SANDHOLZER, L. A. (1933) *J. exp. Med.*, **57**, 279.  
 BECHTOLD, H. and VILLA, L. (1926) *Z. Hyg. Infektkr.*, **105**, 601.  
 BORCHARDT, W. (1924) *Altn. Nachr.*, **3**, 278.  
 BORDET, J. (1923) *Brit. med. J.*, **1**, 175. (1925) *Ann. Inst. Pasteur*, **39**, 717. (1926) *C. R. Soc. Biol.*, **94**, 403.  
 BORDET, J. and CIUCA, M. (1920) *C. R. Soc. Biol.*, **83**, 1296. (1921) *Ibid.*, **84**, 280.  
 BORDET, J. and RENAUX, E. (1928) *Ann. Inst. Pasteur*, **42**, 1283.  
 BROUFEVRENNER, J. (1924-25) *Proc. Soc. exp. Biol.*, N.Y., **22**, 81. (1926) *Science*, **63**, 51. (1927) *J. exp. Med.*, **45**, 373. (1928) 'Filterable Viruses' (Rivers) Baillière, Tindall and Cox, London, p. 373.  
 BROUFEVRENNER, J. J. and KORE, O. (1925a) *J. exp. Med.*, **41**, 73. (1925b) *Ibid.*, **42**, 431.  
 BROUFEVRENNER, J. and MUCKENFUS, R. S. (1927) *J. exp. Med.*, **45**, 887.  
 BROUFEVRENNER, J., MUCKENFUS, R. S. and HETLER, D. M. (1927) *Amer. J. Path.*, **3**, 562.  
 BROUFEVRENNER, J. J. and REICHERT, P. (1926-7) *Proc. Soc. exp. Biol.*, N.Y., **24**, 17.

- BURNET, F. M. (1925) *J. Path. Bact.*, 28, 407. (1927) *Brit. J. exp. Path.*, 8, 121. (1929a) *J. Path. Bact.*, 32, 15. (1929b) *Ibid.*, 32, 349. (1929c) *Brit. J. exp. Path.*, 10, 109. (1932) *J. Path. Bact.*, 35, 851. (1933a) *Ibid.*, 36, 307. (1933b) *Ibid.*, 37, 179. (1933c) *Brit. J. exp. Path.*, 14, 93. (1933d) *Ibid.*, 14, 100. (1933e) *Ibid.*, 14, 302. (1934) *Brit. Rev.*, 9, 332.
- BURNET, F. M. and FREEMAN, M. (1937) *Aust. J. exp. Biol. med. Sci.*, 15, 49.
- BURNET, F. M., KEOGH, E. V., and LUSH, D. (1937) *Aust. J. exp. Biol. med. Sci.*, 15, 22.
- BURNET, F. M. and LUSH, D. (1936) *Aust. J. exp. Biol. med. Sci.*, 14, 27.
- BURNET, F. M. and MCHIE, M. (1929) *Aust. J. exp. Biol. med. Sci.*, 6, 21. (1930a) *Ibid.*, 7, 183, 199. (1930b) *J. Path. Bact.*, 33, 637. (1933) *Ibid.*, 36, 299.
- CAMPBELL-RENTON, M. L. (1937) *J. Path. Bact.*, 45, 237. (1941) *Ibid.*, 53, 371. (1942) *Ibid.*, 54, 235.
- CLAUBERG, H. W. and MARCUSE, K. (1932) *Zbl. Bakt.*, 124, 29.
- CLIFTON, C. E. (1931) *Proc. Soc. exp. Biol.*, N. Y., 28, 745.
- CLIFTON, C. E. and MORROW, G. (1936) *J. Bact.*, 31, 441.
- CLIFTON, C. E., MUELLER, E., and ROGERS, W. (1935) *J. Immunol.*, 29, 377.
- COSTA CRUZ, J. DA. (1926) *C. R. Soc. Biol.*, 95, 1501.
- COWLES, P. B. (1931) *J. Bact.*, 22, 119.
- CRAIGIE, J. and BRANDON, K. F. (1936a) *Canad. publ. Hlth. J.*, 27, 165. (1936b) *J. Path. Bact.*, 43, 233.
- CRAIGIE, J. and YEN, C. H. (1938) *Canad. publ. Hlth. J.*, 29, 449, 484.
- DE, S. P. and PAUL, S. (1940) *Calcutta med. J.*, 37, 499.
- DELBUECK, M. (1940) *J. gen. Physiol.*, 23, 631, 643. (1942) *Advances Enzymology* 2, 1.
- DENTS, P. (1932) *Ann. Inst. Pasteur*, 43, 349.
- DESRAVILLAC, J. M. (1942) *Canad. publ. Hlth. J.*, 33, 122.
- DOLMAN, C. E., KERR, D. E., and HELMER, D. E. (1941) *Canad. publ. Hlth. J.*, 32, 113.
- DOOREN DE JONG, L. E. DE V. (1931) *Zbl. Bakt.*, 120, 1.
- Editorial (1935) *Lancet*, 1, 818.
- EISENBERG, MERLING, H. B. (1941) *J. Path. Bact.*, 53, 365.
- ELFORD, W. J. (1936) *Brit. J. exp. Path.*, 17, 399.
- ELFORD, W. J. and ANDREWES, C. H. (1932) *Brit. J. exp. Path.*, 13, 416.
- ELLIS, E. L. and DELBUECK, M. (1939) *J. gen. Physiol.*, 22, 365.
- ELLIS, E. L. and SPIELEN, J. (1941) *J. gen. Physiol.*, 24, 437.
- EVANS, A. C. (1936) *J. Bact.*, 31, 423. (1940) *Ibid.*, 39, 597. (1941) *Ibid.*, 44, 207.
- EVANS, A. C. and SOCKRIDER, E. M. (1942) *J. Bact.*, 44, 211.
- EVANS, A. C. and VERDEN, E. (1938) *J. Bact.*, 38, 133.
- FERMSTER, R. F. and WELLS, W. F. (1933) *J. exp. Med.*, 58, 350.
- FELIX, A. (1943) *Brit. med. J.*, 1, 435.
- FELIX, A. and CALLOW, B. R. (1943) *Brit. med. J.*, 1, 127.
- FELIX, A. and PITT, R. M. (1934) *J. Path. Bact.*, 38, 409.
- FICK, R. T. (1942a) *J. infect. Dis.*, 71, 153. (1942b) *Ibid.*, 71, 161.
- FLC, P. C. (1923) *Zbl. Bakt.*, 90, 362. (1930a) *Ann. Inst. Pasteur*, 60, 610. (1930b) *Acta leidenensia (Schol. med. trop.)*, 12-13, 103, 113, 118.
- FREEMAN, M. (1937) *Aust. J. exp. Biol. med. Sci.*, 15, 221.
- FRISCH, A. W. and LEVINE, P. (1936) *J. Immunol.*, 30, 89.
- FUKUDA, Y. (1928) *Z. Immunforsch.*, 54, 369.
- GATES, F. L. (1934) *J. exp. Med.*, 60, 179.
- GERCKE, A. (1925) *Zbl. Bakt.*, 94, 387.
- GEST, H. (1943) *J. infect. Dis.*, 73, 158.
- GILDEMEISTER, E. and HERBERG, K. (1923) *Zbl. Bakt.*, 91, 12. (1924a) *Ibid.*, 93, 402. (1924b) *Klin. Wochs.*, 3, 186.
- GILDEMEISTER, E. and WATANABE, H. (1931) *Zbl. Bakt.*, 122, 556.
- GLASER, R. W. (1930) *Amer. J. Hyg.*, 27, 311.
- GORE, W. and JACOBSON, L. (1927) *Z. Immunforsch.*, 53, 12.
- GOUGH, G. A. C. and BURNET, F. M. (1934) *J. Path. Bact.*, 38, 201.
- GÖZONY, L. and SCHRIST, L. (1920) *Zbl. Bakt.*, 85, 333.
- GRATIA, A. (1936) *Ann. Inst. Pasteur*, 56, 307. (1937) *C. R. Soc. Biol.*, 126, 418. (1940) *Ibid.*, 133, 445, 702.
- GRATIA, A. and RHODES, B. (1926) *Lancet*, 1, 204.
- GUELYN, A. (1943) *Ann. Inst. Pasteur*, 69, 219.
- HADLEY, P. (1927) *J. infect. Dis.*, 40, 1. (1928) *Ibid.*, 42, 263.
- HELMER, D. E., KERR, D. E., DOLMAN, C. E., and RANTA, L. E. (1940) *Canad. publ. Hlth. J.*, 31, 433.
- D'HERELLE, F. (1917) *C. P. Acad. Sci.*, 165, 373. (1921) "Le Bactériophage son rôle dans l'immunité." Masson, Paris. Eng. transl., Baltimore and London, 1922. (1926) "Le Bactériophage et son Comportement." Masson, Paris. Eng. transl., Baltimore and London. (1930) "Bacteriophage and its clinical applications." Thomas, Springfield, Illinois.

- D HERELLE, P and RAKIETEN M L (1935) *J Immunol.* 23, 413  
 HERRIOTT P M, HARTZ, Q R., and NORTHOPE, J H (1938) *J gen Physiol.* 21, 575  
 HERSHEY, A D (1941) *J Immunol.* 41, 279  
 HERSHEY, A D and BRONFENBRENNER, J (1943) *J Bact.* 45, 211  
 HERSHEY, A D, KALMANSON, G., and BRONFENBRENNER, J (1943) *J Immunol.* 46, 267, 281, (1944) *Ibid.* 48, 221  
 HUTCHINSON, J R (1913) *Brit med J* ii 130  
 KADOSHIMA, T (1920) *C R Soc Biol.* 83, 219  
 KALMANSON, G and BRONFENBRENNER, J (1933) *J gen Physiol* 23, 203  
 KALMANSON, G M and BRONFENBRENNER, J (1942) *J Immunol.* 45, 13 (1943) *Ibid* 47, 387  
 KALMANSON, G M., HERSHEY, A D, and BRONFENBRENNER, J (1942) *J Immunol.* 45 1  
 KAUFFMANN, F (1925-26) *Z Hyg InfektAr.* 105, 594  
 KENDALL, A I and COLWELL, C. A (1934) *J infect Dis.* 63, 81  
 KEOGH, L. V., SIMMONS, R T., and ANDERSON, G (1938) *J Path Bact.* 46, 505  
 KLEVENBERGER, E. (1932) *Zbl Bakt.* 123, 318  
 KLIOTER, I J and OLKINIE, F (1913) *J Immunol.* 47, 325.  
 KLIVE, G M (1927) *J Lab clin Med.* 12, 1074  
 KNORR, M and ROY, H (1935) *Zbl Bakt.* 133, 280  
 KRUEGER, A P (1931) *J gen Physiol.* 14, 493  
 KRUEGER, A P and BALDWIN, D M (1935) *J infect Dis.* 57, 207, (1937) *Proc Soc exp Biol A* 37, 393  
 KRUEGER, A P, BROWN, B B., and SCHRIENER, E J (1941) *J gen Physiol.* 24, 631  
 KRUEGER, A. P and FONG J (1937) *J gen Physiol.* 21, 137  
 KRUEGER, A P and NORTHOPE, J H (1931) *J gen Physiol.* 14, 233  
 KRUEGER, A P, RITTER, R C., and SMITH, S P (1929) *J exp Med.* 50, 739  
 KRUEGER, A P, SCHRIENER, F J., and MCCRACKEN, T (1940) *J gen. Physiol.* 23, 705  
 KUTTZER, A G (1923) *J Bact.* 7, 49  
 LAZARUS, A S (1940) *Amer J publ Hlth.* 30, 1177; (1941) *Ibid.* 31, 60  
 LEFFER, F (1923) *Brit J exp Path.* 4, 204  
 LEVIN, B S. and LOMIVSKI, I (1936) *C R Soc Biol.* 122, 1286  
 LEVINE, P and FRISCH A W (1934) *J exp Med.* 59, 213, (1935) *J infect Dis* 57, 104 (1936) *J Immunol.* 30, 63  
 LEVINE, P., FRISCH, A. W., and CONEX L. V (1934) *J Immunol.* 28, 321  
 LEWIS, I M and WORLEY, G (1936) *J Bact.* 32, 195  
 LISBONKE, M and CARRELL, L. (1922) *C R Soc Biol.* 86, 569  
 LOMIVSKI, I (1934) *C. R Soc Biol.* 129, 264  
 LURIA, S (1940) *Ann Inst. Pasteur.* 64, 415.  
 LURIA, S F., DELBECQ, M and ANDERSON, T F (1943) *J Bact* 48, 57  
 LURIA, S F and LIXNER, F M (1941) *Proc nat Acad Sci Wash.* 27, 370  
 LOWRY, A (1936) *Ann Inst Pasteur.* 66, 165  
 McINTOSH, J and SELWICK F R (1937) *Brit J exp Path* 18, 162  
 MCKINLEY, F B and HOLDEN, M (1926) *J infect Dis.* 39, 451  
 MANVINGER, R. (1926) *Zbl. Bakt.* 99, 203  
 MEISSNER, G (1924a) *Zbl Bakt.* 91, 149, (1924b) *Ibid.* 92, 421  
 MERLING FISCHBERG, A B (1938) *Brit J exp Path.* 19, 338  
 MERRILL, V H (1936) *J Immunol.* 30, 169  
 MILLER, A A (1937) *Ann Inst. Pasteur.* 58, 709  
 MORISON, J (1932) "Bacteriophage in the treatment and prevention of cholera." Lewis & Co., London.  
 MORIYAMA, R and OHASHI S (1937) *J Shanghas Sci Inst.* Sect IV, 3, 155 181  
 NAITO, R. (1936) *Zbl Bakt.* 138, 34  
 NATARAJAN, C. V and HYDE, R R (1930) *Amer J Hyg.* 11, 652  
 NORTHOPE, J H (1938) *J gen Physiol.* 21, 335, (1939) *Ibid* 23, 59  
 OGATA N (1924) *Zbl Bakt.* 83, 329  
 OHASHI, S (1938) *J Shanghas Sci Inst.* Sect IV, 3, 279, (1939) *Ibid.* 5, 1  
 OLSEN O and YASAKI, Y (1923) *Klin Wochr.* 2, 1879  
 OTTO R and MÜNSTER, H (1923) *Z Hyg InfektAr* 100, 402  
 LAFC, M., KRASSNOFF, D., HABER, P., REINIE, L. and VOET, J (1938) *Ann Inst Pasteur.* 60, 227  
 PEDRAZ, J R and TODD C (1933) *Proc. roy Soc., B* 112, 277  
 PIRIE A (1939) *Brit J exp Path.* 20, 99, (1940) *Ibid.* 21, 125  
 PREISE, H VON (1925) "Die Bakteriophagie, etc." Fischer Jena.  
 RADOSCH, M M (1936) *Zbl Bakt.* 136, 326  
 RAKIETEN, M L., LOCKERTH A H and RAKIETEN, T L. (1940) *J Bact.* 40, 529  
 RAKIETEN, M. L. and RAKIETEN, T L. (1937) *Yale J Biol Med* 10, 191  
 RAKIETEN M L RAKIETEN, T L., and DOFF, S (1936) *J Bact.* 32, 505

- REICHERT, F (1924) *Zbl Bakt.*, 81, 235.  
 RUSKA, H (1941) *Naturwiss.*, 29, 367  
 SCHLESINGER, M (1932-33a) *Z Hyg Infektkr.*, 114, 161. (1932-33b) *Ibid.*, 114, 716.  
 (1933) *Biochem Z.*, 264, 6. (1934) *Biochem Z.*, 273, 306.  
 SCHLOSSMAN, K. (1932) *Z Hyg Infektkr.*, 114, 6a.  
 SCHMIDT, A. (1931) *Zbl Bakt.*, 123, 202, 207  
 SCHOLTENS, R T (1930) *J Hyg., Camb.*, 38, 452. (1937) *Ibid.*, 37, 315.  
 SCHULTZ, E. W and KRUEGER, A P (1928) *Proc Soc exp Biol.*, N Y., 28, 97  
 SCHUTTMAN, C J (1925) *Zbl Bakt.*, 95, 97. (1936) *Ibid.*, 137, 433.  
 SCHUTTMAN, C J and SCHUTTMAN TEN BOSKEL HUIJINK A. M (1936) *Zbl Bakt.*, 138, 264  
 SCHWARTZMAN, G (1926-27) *Zbl Bakt.*, 101, 62  
 SCRIBNER, E S. and KRUEGER, A P (1937) *J gen Physiol.*, 21, 1  
 SERTIC, V (1939) *C P Soc Biol.*, 100, 477. (1937a) *Ibid.*, 124, 14, 93. (1937b) *Ibid.*, 126, 1074  
 SERTIC, V and BOULGAKOV, N (1935a) *C R Soc Biol.*, 119, 983. (1935b) *Ibid.*, 119, 985.  
 SERTIC, V and BOULGAKOV, N A (1936a) *C R Soc Biol.*, 122, 30. (1936b) *Ibid.*, 123, 887. (1937) *Ibid.*, 126, 734. (1939) *Ibid.*, 132, 444  
 SMITH G F and JORDAN, E. O (1931) *J Bact.*, 21, 75  
 SONNENSCHEIN, C. (1927) *G Batt Immun.*, 2, 32.  
 SPAT, W (1924) *Med. Klin.*, 20, 184  
 SPIELER, J (1943a) *J infect Dis.*, 73, 212. (1943b) *Ibid.*, 73, 222  
 STASSANO, H. and DE BEAUFORT, A. C. (1925) *C R Soc Biol.*, 93, 1378.  
 STEWART, A. D and GHOSAL, S C. (1931) *Indian J med Res.*, 19, 137  
 TODD, C (1927) *Brit. J exp Path.*, 8, 369  
 TWOOT, F W (1915) *Lancet*, ii 1241. (1922) *Brit. med J.*, ii 293. (1925) *Lancet*, ii 642  
 (1926) *Ibid.*, i. 416  
 VAGIDES, K. VOY and GILDEMEISTER, E. (1934) *Zbl Bakt.*, 131, 414  
 WHEELER, K. M. and BURGDORF A L. (1941) *Amer J publ Hlth.*, 31, 325.  
 WHITE, P B (1937) *J Path. Bact.*, 44, 276.  
 WIEBOLS G L and WIERINGA, K T (1936) "Fonds Landbouw Export Bureau, 1916-18. No. 16.  
 WILLIAMS C H SANDHOLZER, L. A., and BERRY, G P (1940) *J Bact.*, 40, 517  
 WILLIAMS, S. and TIMMINS, C. (1938) *Med J Aust* ii, 657  
 WOLLMAN, E. (1925) *Ann. Inst. Pasteur*, 39, 789. (1927) *Ibid.*, 41, 883. (1928) *Bull Inst Pasteur*, 26, 1 (1929) *Ann. Inst. Pasteur*, 43, 309. (1934a) *C R Soc Biol.*, 115, 1616.  
 (1934b) *Bull Inst. Pasteur*, 32, 945 (1935) *Lancet*, ii 1312.  
 WOLLMAN, E. and LACASSAGNE, A. (1940) *Ann Inst. Pasteur*, 64, 5.  
 WOLLMAN E. and WOLLMAN E. (1932) *Ann Inst. Pasteur*, 49, 41. (1936) *Ibid.* 56, 137. (1935) *Ibid.*, 60, 13. (1939) *C P Soc Biol.*, 131, 442.  
 WRIGHT E. V and KERSTEN H (1937) *J Bact* 34, 639  
 YAOI H and SATO, K. (1935) *Jap J exp Med.*, 13, 56a.  
 ZIESSER, H and TANG F F (1937) *J exp Med.*, 46, 357



## PART II

### SYSTEMATIC BACTERIOLOGY

#### CHAPTER 12

#### THE METHODS OF OBTAINING PURE CULTURES, AND THE IDENTIFICATION OF BACTERIA

##### METHODS OF OBTAINING PURE CULTURES OF BACTERIA

ONE of the first essentials in the study of bacteriology is the preparation and maintenance of pure cultures of bacteria. Neglect of this leads inevitably to confusion. During the sixties and seventies of last century, micro organisms were perforce cultivated in liquid media, and as the preparation of pure cultures in such media is often difficult and sometimes impossible relatively little progress in the identification of particular species was made in these years. It was the introduction of solid media by Robert Koch in 1881 that rendered possible the easy separation of different organisms from one another in a mixed culture, and provided a means for distinguishing macroscopically between different species of bacteria. Koch found that on a suitable solid medium, such as potato or gelatin, most organisms formed characteristic colonies by which they could be readily identified, by inoculation of separate colonies into tubes of a liquid medium, pure cultures could be obtained. These could again be streaked on to a solid medium, and if the resulting colonies were all of the same appearance, it might be concluded that the liquid culture probably contained only one species of bacterium. This method—Koch's plating method—affords the simplest and most rapid means of separating one organism from another, we shall later describe it more fully.

We have now at our command numerous methods of purifying cultures. Some are of limited utility, or are suited solely to certain organisms, others are of wider applicability. Without discussing the technical details we shall give a brief description of the principles underlying the more important of these methods.

**A. Dilution Method.**—In point of time, this was the first method introduced for obtaining pure cultures of a bacterium, a method which we owe to Lister (1878). The mixed culture is diluted with sterile tap water, or other suitable fluid, till there is only about one organism in every two drops of the mixture. A series of tubes containing broth is then seeded, each with one drop of the diluted culture. If the dilution has been correctly gauged, there should be a growth in approximate

every alternate tube. This method affords one no certainty that the cultures obtained are pure, further study must be undertaken to ascertain this. The objections to the method are that too much guess-work is involved in judging the correct dilution, and that several tubes of medium are inevitably wasted. It is useful, however, in a modified form in conjunction with the plating method. That is to say, when the culture is thick, it is wise to dilute it considerably before plating out, in this way there is more likelihood of obtaining single colonies from single organisms.

**B. Koch's Plating Method.**—Originally the solid medium used was spread out in the melted state on microscopic slides and allowed to set, these were then streaked with a needle dipped in the culture, and incubated in a moist bell jar. Later, large glass plates were used, these had to be specially levelled by means of adjustable screw supports, and covered with a bell jar. The method now employed is to pour the melted medium into Petri dishes, each of these is provided with a cover, which protects the medium from contamination. The plating method may be employed in one of two ways. Either the culture material may be streaked on the surface of the solid medium, or it may be mixed with the medium in the melted state, poured out into Petri dishes, and allowed to set. The former method results in a surface growth, the latter in a growth throughout the whole thickness of the medium. As a rule the former method is the more useful. In streaking the surface of the medium, a drop of the fluid culture may be placed in the centre of the dish and spread out in all directions by means of a sterile glass or metal rod bent at a right angle. This results in an even distribution of organisms over the plate. If, however, single colonies are particularly desired—and this is usual—it is best to make a series of streaks across the plate with a platinum loop dipped in the culture, the streaks should be about 10 mm. apart, and may be crossed at right angles, the platinum loop should not be re-charged with culture during the process. If preliminary dilution has not been performed, it is often advisable to continue the streaking over a second or even a third plate without re-charging the loop. On the first plate the growth may be entirely confluent, but on the second and third single colonies will generally be obtained. These single colonies can then be examined with a hand lens, and picked off with a platinum needle into broth. Except with certain organisms, single colonies obtained in this way, especially if the culture has been previously diluted, are generally derived from single organisms, and are hence pure. The purity of colonies picked from over crowded plates is less certain. In using the surface streak method it is important that the plates should be fairly dry, if there is a film of moisture on the medium—resulting partly from condensation and partly from exsiccation—organisms, particularly if motile, are apt to form a confluent growth over the whole surface. This point must be borne in mind, especially when attempting to isolate anaerobic bacteria. These organisms instead of growing up from the medium frequently spread in a thin layer over it, the edges of the colonies are difficult to define, and if there is a film of moisture over the medium, they coalesce, thus rendering their isolation impossible. The pour plate method is, in general, of less value, the deep colonies are not usually as characteristic as the surface colonies, and to pick them off involves stabbing the medium with a platinum wire—a process that takes longer than the simple one of surface picking. In carrying out this method of plating, a tube containing 15 ml. of the solid medium is heated in water till the medium is melted, for gelatin a temperature of 30° C. will suffice, for agar the water must be boiled.

The gelatin tube can be inoculated directly with a drop of the diluted culture, the agar tube should be cooled to  $45^{\circ}\text{C}$  before inoculation. The culture is then thoroughly mixed by rotation of the tube, and the mixture is poured out into a Petri dish and allowed to set. Gelatin sets at about  $25^{\circ}\text{C}$ , agar at  $38^{\circ}\text{C}$ . In warm weather the gelatin plates should be set on ice.

**C Shake-tube Method.**—This is used chiefly for anaerobes. A test tube containing about 15 ml of solid medium, generally made up with a reducing agent such as glucose, is heated till the medium is melted, a drop of culture is delivered into the medium, which has been cooled to a suitable temperature, and thoroughly mixed by gentle shaking and rotation. The medium is then allowed to set in the tube in a vertical position. Incubation is carried out aerobically or anaerobically. The great value of this method is that it affords a simple means of grading the oxygen pressure in the medium, on the surface the pressure is atmospheric, at the bottom of the tube, particularly if a reducing agent has been added, the conditions are completely anaerobic. Thus the aerobic bacteria grow at or just below the surface, the anaerobic bacteria grow near the bottom, and the facultative aerobes and anaerobes are distributed throughout the medium. The single colonies, which appear, are often fairly characteristic. To pick them off, the test tube is cut round with a diamond at the middle of the column of medium, the two halves of the tube drawn apart, and the medium allowed to fall gently into a sterile Petri dish. A stout platinum wire is then used to fish the colonies, it is stabbed into the medium over the particular colony desired, taking care that no other colony is touched on the way, and as soon as it has come into contact with the colony, it is withdrawn and inoculated into broth. Sometimes it is advisable to cut the medium into pieces with a sterile scalpel before attempting to pick off the colonies. This method was used at one time for purifying anaerobes, but now that the technique of obtaining anaerobiosis has improved, it has been largely replaced by the more reliable surface plating method. In order to avoid breaking a test tube every time a colony has to be picked off, it is better to use a Veillon tube instead of the ordinary test-tube. This consists of a piece of glass tubing, about 1 cm in diameter, and 8 or 10 inches long. One end is fitted with a rubber cork, the other with a cotton wool plug. The medium is poured into the tube till it reaches about half or two thirds of the way up. Inoculation is performed in the usual way. When it is desired to pick off a colony, the rubber cork and the woollen plug are removed, and the whole column of medium expelled by a stout glass rod into a Petri dish. The tubes and corks can be used over and over again.



FIG 56—  
VEILLON  
TUBE

**D. Motility.**—Various methods have been devised for making use of motility to separate motile from non motile organisms. Rovida's (1925) tube, which is a modification of that invented by Carnot and Garnier (1902), may be used for this purpose. It consists of a large test-tube to the bottom of which is fused a glass tube of 7 mm internal diameter. This inner tube has a constriction near the lower end, below which are three small holes to afford a communication between the inside of the inner and the outer tubes. Above the constriction there is a

plug of glass wool, which supports a layer of sand. Broth is poured into both tubes till it reaches the same level. The mixed culture is seeded into the broth of the outer tube and the apparatus is put in the incubator. The organisms that are motile will pass through the holes into the inner tube, grow up through the wool and sand, and produce a turbidity in the broth of the inner tube, from this they may be recovered in pure culture. The non motile organisms remain confined to the broth of the outer tube. An alternative method described by Craigie (1931) is now extensively used for the separation of motile from non motile organisms of the *Salmonella* group. It consists of an ordinary test tube,  $6 \times \frac{1}{2}$  in., containing a piece of glass tubing 4 in. long, having the bottom cut off obliquely. About 6-8 ml. of melted semi-solid (0.25 per cent.) nutrient agar are poured into the outer tube. After autoclaving the agar is cooled to  $45^{\circ}\text{C}$ ., and the inner tube which projects above the level of the agar, is inoculated with the organism under test. If motile organisms are present, or appear as the result of incubation they grow down the inner tube and up the outer tube, from the top of which they may be subcultured in a day or two's time (see also Tulloch 1939).

**E. Optimum Temperature**—It is sometimes possible to make use of the optimum temperature of growth of an organism when it is desired to obtain it in pure culture, as for instance when one wants it to multiply freely in a mixed culture. The thermophilic bacteria may be separated from other organisms by incubating the medium at about  $60^{\circ}\text{C}$ ., none of the ordinary bacteria will grow at this temperature so a pure growth of the thermophilic organisms is obtained. Again, certain bacteria will not grow at  $22^{\circ}\text{C}$ ., whereas others will. If a mixture of the two is incubated at  $22^{\circ}\text{C}$ ., only one type will develop, this may then be picked off pure. In this way *A. catarrhalis* may be separated from the meningococcus. In water analysis, when it is desired to know the numbers of potentially pathogenic bacteria in a given sample, the cultures are incubated at  $37^{\circ}\text{C}$ ., many of the saprophytic forms fail to grow at this temperature, and the resulting growth consists largely of potential parasites.

**F. Aerobic and Anaerobic Incubation.**—This is a simple method of separating aerobes from anaerobes. Incubated aerobically, the strict anaerobes will not grow, incubated anaerobically, the strict aerobes will not grow. And as most facultative anaerobes seldom grow as well under anaerobic as under aerobic conditions anaerobic incubation favours the strict anaerobes more than the facultative ones.

**G. Heating**—Heating a mixed culture at  $80^{\circ}\text{C}$  for 10 minutes will destroy all the vegetative non-sporing bacteria while leaving the spores unaffected. This method is largely used in the preliminary purification of the anaerobes. In the body, most of the organisms with which the anaerobes are likely to be contaminated are non sporing cocci and bacilli, all of these are destroyed at  $80^{\circ}\text{C}$ ., and consequently the anaerobic organisms alone develop.

**H. Selective Bactericidal Substances**—Certain substances with a germicidal action are useful in destroying susceptible organisms, while leaving the more resistant unaffected. One of the best examples of this method is the isolation of the tubercle bacillus from sputum by the use of antiformin. Tubercle bacilli are very resistant to chemical disinfectants, even though they are easily killed by heat. If the sputum, which generally contains numerous other organisms, is treated with 15 per cent antiformin (equal parts of 15 per cent NaOH and Liq. sodæ chlorinatæ

B P), for a time varying from 5 to 60 minutes according to the thickness of the sputum, and inoculations are then made on to egg medium, the tubercle bacilli will develop in pure culture, without the antiformin they would be overgrown in 24 hours or less. Fifteen per cent sulphuric acid is often used for the same purpose. Subcultures should be made at intervals of from 5 to 20 minutes.

**I Agglutinating Serum**—If it is suspected that relatively few organisms of a particular species are present in a bacterial culture or suspension so that direct plating is unlikely to prove successful, it is sometimes possible to concentrate them by adding a specific high titre agglutinating serum, incubating for 2 hours, and centrifuging. The organisms, which are clumped together, are easily thrown down and are present almost exclusively in the deposit. Plates may then be streaked from this directly. This method is sometimes of value in isolating the typhoid bacillus from water. The same principle underlies the method of separating the phases of motile diphasic organisms of the *Salmonella* group. If, for example, a Craigie tube (see p. 351), containing semi solid agar to which a small quantity of agglutinating serum active against Phase 1 has been added is inoculated with the strain under test the Phase 1 organisms will be agglutinated and rendered non motile, whereas the Phase 2 organisms will grow down the inner and up the outer tube, and be recoverable from the surface of the agar (see Tulloch 1939).

**J Filtration**—This method may be used to separate the filtrable viruses from the ordinary bacteria, or if a fairly coarse candle is used, such as a Berkefeld N or V, it may be used to separate very small bacteria like *Bact. pneumosintes* from larger organisms. Many spirochaetes will also pass through Berkefeld filters. This property is made use of in separating them from the bacteria with which they are often contaminated.

**K. Selective and Enrichment Media**—For the isolation of special organisms from others that are likely to overgrow it in culture, it is common to add to the medium substances having either a stimulating effect on the organism it is desired to cultivate or an inhibitory effect on those it is desired to suppress. If the substance is added to a liquid medium the result is an absolute increase in the numbers of the special organism which becomes at the same time relatively more numerous to others than in the original material used for the inoculum. Such a liquid medium is known as an *enrichment medium*. If, on the other hand the substance is added to a solid medium it acts selectively, enabling a greater proportion of the special organisms to form colonies than would otherwise have been possible. Such a solid medium is known as a *selective medium*. It is common to use the two types of media in conjunction, inoculating the material first into a liquid enrichment medium so as to obtain a relative and absolute increase in the numbers of the special organism, and then plating the culture on to a solid selective medium so as to favour the development of colonies of the special organism at the expense of others. It is important to realize that colonies on a selective medium are not necessarily pure. At the base of the colony other organisms may be present which, though unable to develop on the selective medium itself, will nevertheless grow rapidly when transferred to a non selective medium. Thus fermentation results may be misleading if the sugar tubes are inoculated directly from colonies on a selective medium. It is therefore wise to re plate colonies from a selective medium on to a plain medium to ensure their purity before testing the biochemical antigenic or other properties of the organism under study.

Blood, serum, and ascitic or hydrocele fluid are substances that are frequently used to stimulate the growth of certain organisms, glucose and other sugars, extracts of vegetable and animal tissues, and certain salts such as potassium nitrate, are likewise used for the same purpose. On the other hand certain aniline dyes phenol, telluric acid, bile salts, and numerous other substances are used for inhibiting the growth of various organisms. Gentian violet, in a concentration of about 1-10 000, suppresses the growth of most Gram positive organisms, while allowing most Gram negative organisms to develop. Used at 1-500 000 it is of value in separating streptococci from staphylococci since the latter organisms are inhibited by this concentration. Brilliant green is often used for preventing the growth of lactose-fermenting organisms in cultures from the stools. It is added to the faecal suspension in broth in a concentration of about 1-150 000, the culture is incubated and plated out on a suitable medium after 18 to 24 hours. Many Gram negative bacteria are inhibited by potassium tellurite in a concentration of 1-80 000 or more, while penicillin exercises an inhibitory action mainly on Gram positive bacteria (Fleming 1932).

**L. Indicator Media.**—These are media that contain an indicator which changes colour when a certain organism or group of organisms develops. Thus, if it is known that the organism which it is desired to cultivate produces  $H_2S$ , lead acetate may be added to the medium, the colonies of the organism are coloured brown, owing to the production of lead sulphide, and can be readily picked off for identification. The diphtheria bacillus reduces sodium tellurite, whereas many of the organisms likely to be associated with it in a throat swab do not. When this substance is added to the medium, the colonies of the diphtheria and of the diphtheroid bacilli are coloured black, whereas those of the streptococci and numerous other organisms are colourless. Litmus and neutral red are two dyes that are frequently used to indicate the production of acid from some carbohydrate incorporated in the medium. Colonies of organisms that ferment the sugar are coloured red owing to the production of acid, whereas those that do not do so take on the alkaline colour of the dye—blue and yellow respectively. Blood is a very useful indicator. Some organisms produce no alteration in it others form from it a green pigment, while others lyse it completely. It is usually added to agar in a concentration of 5 per cent. The colonies of the first class leave the medium unchanged, those of the second class are surrounded by a greenish ring, those of the third class by a perfectly clear transparent ring. It is used particularly in the differentiation of the streptococci.

Selective and indicator media are frequently combined. Thus in MacConkey's medium bile salts are added to inhibit the growth of organisms other than those capable of multiplying freely in the intestine, and lactose and neutral red are added to distinguish the lactose-fermenting coliform organisms from the non lactose-fermenting group.

**M. Pathogenicity Methods.**—The introduction of the pathogenicity method of separating organisms from one another we owe to Koch (1880). By this means he succeeded in separating streptococci from *Erysipelothrix muriseptica*. When the mixed culture was injected into the ear of a house mouse, *Ery muriseptica* proliferated, invaded the blood stream, and could be obtained in pure culture from the heart's blood after death, the streptococcus proliferated locally but did not invade the blood stream. As it was mixed in the local lesion with *Ery muriseptica* it could not be obtained in pure culture. But Koch found that if the mixed culture

was injected into a field mouse the streptococci proliferated invaded the blood stream and caused death while *Ery muriseptica* did not grow at all, the streptococcus was therefore obtained in pure culture from the blood. This principle is of wide application. It is used particularly to isolate organisms that are pathogenic to a certain laboratory animal from other closely similar organisms that are not pathogenic. Thus *B anthracis* can easily be separated from *B subtilis* or *B megatherium* by the injection of a mouse or a guinea pig. It is also used to isolate pathogenic organisms which are not easy to grow in culture or which are readily overgrown by contaminating organisms. As examples we may quote the tubercle bacillus in pus or the pneumococcus in sputum. The contaminating organisms are rapidly killed in the animal body whereas the pathogenic organism multiplies and can be recovered in pure culture from the tissues.

**N Single Cell Methods**—The aim of these methods is to obtain a culture of a given organism from a single bacterial cell. If this can be carried out successfully then the resultant culture must obviously be pure. If the technique for single cell isolation was simple and flawless this would be the ideal method for the purification of cultures. In fact, however several of the methods advocated for this purpose suffer from optical or other defects which seriously detract from their value. In Barber's (1908) method the culture is diluted and a series of tiny droplets prepared. These are placed on the under surface of a cover slip forming the roof of a special chamber, and examined under the microscope. When a drop is found containing only one organism it is picked off with a special capillary pipette and transferred to a fluid medium. This method has been widely used but it suffers from the defect that in viewing a spherical droplet the optical conditions are such as to render accurate observation of particles at the water air interface very difficult or impossible. Hence there is no absolute certainty that a single cell has been picked. A method devised by Topley, Barnard and Wilson (1921) eliminates these particular optical defects. A loopful of a young gelatin culture at 37° C is placed on a slide and covered with a quartz cover glass. Under dark ground illumination a single organism is picked out which is well removed from any other organisms and is covered with a minute droplet of mercury. The preparation is exposed for a short time to ultra violet irradiation with the object of destroying all organisms except the single one that has been protected by the mercury droplet. After incubation overnight the preparation is again examined and if successful a colony will be observed at the site previously occupied by the protected organism. This can then be transferred to a liquid medium. Adequate controls are necessary to prove that the irradiation was sufficient to kill all non protected organisms. Several other methods have been described.

#### THE IDENTIFICATION OF MICRO ORGANISMS

Having once obtained a pure culture of a particular organism it is necessary to establish its identity by an appropriate series of tests. This may require a few weeks or it may take several months to complete. Many of the reactions may have to be tested three or four times to make sure of their consistency. It is often desirable to prepare photographs recording the morphology and colonial appearances

on the most important media, these will be found of great value for future comparison. For studying the properties of an organism the following scheme is suggested.

**A Morphology**—Under this heading we include the shape and size of the organism, its arrangement, motility, the number and distribution of flagella, the shape and situation of spores, and capsule formation. It is impossible to study all the properties on a single medium, motility for example should be looked for in a young rapidly growing broth culture, preferably not more than 6 to 8 hours old, flagella are sought for on a young agar culture, spores in a culture that has been growing for some days, capsules in a pathological exudate, and so on. The shape and size of the organisms are subject to considerable variation, and it is important to gain some idea of the extent of this variation. With a few exceptions, such as the corynebacteria, most organisms are larger in a young than in an old culture (Henrici 1926). Measurements, for example, of *Salmonella typhimurium* in a 4-hours culture on agar showed that the average size was  $2.30 \mu \times 0.79 \mu$ , in the same culture after 26 hours the size was only  $1.13 \mu \times 0.49 \mu$ . In volume the organisms from the young culture were over five times that from the old. On further incubation the average size decreased still more (Wilson 1926). When taking measurements of a given strain it is therefore important to record the age of the culture from which they were taken. Even in one and the same preparation the individual organisms may vary considerably in size and shape, this may be so marked as to justify the term "pleomorphic". Thus coccoid, bacillary, and filamentous forms may all be present together, or besides the usual rods there may be club forms, navicular forms, granular forms, large bloated forms, shadow forms, and so on. Moreover, the appearance of the organisms is often considerably influenced by the type of medium on which they are grown. Chain formation, for example, is more evident in liquid media than on solid. The typical morphological appearance of the diphtheria bacillus is seen best on Loeffler's serum, on agar the organisms tend to be more solid and less granular. The nature of the medium often influences the production of spores and of capsules. Some organisms, such as *B. anthracis*, form spores readily in artificial culture, but never do so in the animal body. On the other hand, capsules are quite frequently found in the body, but less often in artificial culture. The arrangement of the organisms should be carefully studied. If they are cocci, they may be arranged singly, in pairs, tetrads, packets, clusters or chains, if bacilli, they may be arranged singly, in pairs end-to-end, in bundles, chains, clusters, or in Chinese-letter forms in which the individual bacilli lie more or less at right angles to each other. If vibrios they may be arranged singly, in S-forms, semicircles, in wavy chains composed of S-forms strung end-to-end, or they may present the fish-in-stream appearance. Though most organisms show two or three types of arrangement, it is usual for one of these to be predominant. This comes to be regarded as the typical arrangement. It cannot be emphasized too strongly that the morphology of bacteria is subject to variation depending on the age of the culture, the nature of the medium, the particular strain used, the temperature of incubation, and a number of other factors, the extent of this variation can be learnt only by experience.

**B Staining Reactions**—The morphology of bacteria may be studied in a hanging-drop preparation, by dark ground illumination, or in stained films. By each of these methods different information may be gained. Staining methods, in addition to revealing the morphology of the organism, may render evident



differences in the chemical constitution of different organisms, or of different parts of the same organism. For studying the morphology, it is advisable to use a weak stain, otherwise so much dye may be absorbed as to alter the appearance of the organism. Gram's stain is of great value in that it serves to divide all bacteria into one or other of two classes—the Gram positive and the Gram negative. The Ziehl Neelsen method of staining is likewise of value, since it serves to distinguish the acid fast from the non acid fast bacilli. Numerous other stains are used for special purposes, such as the demonstration of flagella, capsules, spores, and meta chromatic granules.

By a study of the morphology and the staining reactions, it is generally possible to identify the group to which a given organism belongs. In certain instances when the origin of the organism is known it is possible to make a presumptive diagnosis of its actual identity, though it should be clearly understood that in medical bacteriology, a provisional identification of this kind is valid only if in consonance with a clinical diagnosis established on other grounds. Thus acid fast bacilli in the cerebro-spinal fluid of a patient with clinical symptoms of meningitis may provisionally be identified as tubercle bacilli, Gram negative diplococci in the pus of an infant with ophthalmia neonatorum are probably gonococci, and Gram negative bipolar staining ovoid bacilli in the gland juice from a patient with an inguinal bubo, in an area where plague is prevalent, may provisionally be regarded as plague bacilli. As a rule, however, it is impossible to identify an organism by morphology and staining alone.

**C. Cultural Reactions**—Under this heading must be included a study of the surface, and often of the deep, colonies formed on solid media and of the type of growth in fluid media. Nutrient agar is the usual medium on which colony formation is studied but if the organism fails to grow on agar, then some other medium must be chosen. The colonies are best examined after 24 hours incubation at 37° C and again at intervals for a week. In describing them, particular attention should be paid to their shape, size, elevation, structure, colour, transparency, surface, edge, consistency, and emulsifiability, differentiation into central and peripheral areas should also be noted. The type of growth following a streak inoculation on an agar slope should be studied, attention being paid particularly to the profuseness of growth, to the elevation, colour, surface, and edge and to any change in the medium itself. The type of growth in a gelatin stab culture should also be studied, and notes made of the degree and extent of the growth, the presence of a surface growth, the presence or absence of liquefaction and if liquefaction occurs of the particular type which it assumes (see Chapter 13). In any systematic examination, the growth should be studied on certain special media such as Loeffler's serum, glycerine potato, and coagulated egg.

The cultural reactions of the different groups of bacteria are fairly distinctive and even within a given group there may be differences between the members. Some organisms moreover have a characteristic form of growth, which enables them to be distinguished from morphologically similar organisms. As a rule however, a study of the cultural reactions merely indicates the group to which a given organism belongs, it does not distinguish between the different members. It serves to confirm the conclusions reached from the examination of the morphology and staining reactions.

**D. Resistance**—Organisms vary considerably in their resistance to inimical agencies. Roughly speaking, three classes may be distinguished.

(1) The bacteria that are susceptible to low degrees of heat, and low concentrations of chemical disinfectants, this class includes the non sporing bacteria and the vegetative forms of the spore-bearing bacteria. They are destroyed by moist heat at  $60^{\circ}\text{C}$  in half an hour and by 1 per cent phenol within an hour.

(2) The bacteria that are susceptible to low degrees of heat but are resistant to low concentrations of disinfectants, this class includes the acid fast bacteria, which are killed at  $60^{\circ}\text{C}$  in half an hour, but resist destruction by chemical agents in the cold often for several hours.

(3) The bacteria that are resistant both to low degrees of heat and low concentrations of disinfectants, this class includes the sporing forms of the spore-bearing bacteria. To kill them with certainty, steam under pressure at a temperature of  $120^{\circ}\text{C}$  for half an hour should be employed or high concentrations of disinfectants, for example 5 per cent phenol, maintained for several hours.

A study of the resistance of a given bacterium will, as a rule, merely serve to confirm the conclusions already reached by the three previous methods of examination, but occasionally it is in itself of some diagnostic importance. Thus certain of the non sporing vegetative bacteria for example the enterococcus are not destroyed at  $60^{\circ}\text{C}$  in half an hour, they require a temperature of  $60^{\circ}\text{C}$ , this abnormal heat resistance is of value in differentiating this species of streptococcus from other species, which are readily killed at the lower temperature. Whenever an organism is suspected of forming spores, the heat resistance must be tested, and not till the suspected spores have definitely been found to be resistant to heat should the conclusion be reached that they really are spores. Many forms have been interpreted in the past as being true spores, which on subsequent examination have been found to be devoid of the characteristic property of heat resistance.

**E Metabolism.**—Under this heading is included a study of the oxygen pressure required for growth, the optimum temperature for growth, pigment formation, hæmolytic production, and the effect on growth of adding different substances to the medium. It is usual to divide bacteria into 3 classes according to their oxygen requirements. (1) Strict aerobes these organisms will grow only in the presence of free oxygen. (2) Strict anaerobes these will grow only in the absence of free oxygen. It must be noted however that growth will occur in the presence of molecular oxygen provided the medium contains a reducing system capable of bringing about a sufficiently low O-R potential. (See Chapters 3 and 36.) (3) Facultative anaerobes these grow best under aerobic conditions, but are able to grow under anaerobic conditions. To these may be added a fourth class the microaerophiles comprising those organisms that grow best under a pressure of oxygen lower than that of the atmosphere. According to their temperature requirements bacteria may be divided into (1) the mesophilic, which have an optimum temperature between  $20^{\circ}\text{C}$  and  $40^{\circ}\text{C}$ , and (2) the thermophilic which have an optimum temperature between  $60^{\circ}\text{C}$  and  $70^{\circ}\text{C}$ . In medical bacteriology, the important distinction lies between those organisms that will grow at room temperature as well as at  $37^{\circ}\text{C}$ , and those that will grow at  $37^{\circ}\text{C}$  but not at room temperature. The latter class includes many of the highly parasitic organisms. The power to hæmolyse may be studied by growing the organism on blood agar plates or by mixing varying dilutions of a broth culture with a suspension of washed red cells. This property is of considerable import

ance and is employed as a primary criterion for differentiating between the members of the streptococcal group. The effect on growth of adding blood serum glucose nitrates and bile salts to the medium is important since it is often of differential value. The formation of pigment should be studied on various media and at different temperatures as a rule it will be found that pigment is best formed on the surface of a solid medium at a temperature of 20-30° C. In liquid media or in the depth of solid media and at temperatures above 35° C. pigment is formed less abundantly under strict anaerobic conditions it is formed only by exceptional organisms such as *F. velutigenus*.

A study of the salient metabolic functions of an organism as a rule adds considerably to the information derived from the previous methods of examination. Oxygen and temperature requirements and pigment formation especially are of great classificatory value being frequently used for the differentiation of species.

**F Fermentation Reactions and Other Biochemical Properties**—Under this heading we include a study of the fermentative action on certain carbohydrates and alcohols (colloquially spoken of as sugars) of the proteolytic powers especially the digestion of gelatin egg and serum of the fat splitting powers of the power to reduce certain dyes such as methylene blue and litmus or certain salts such as nitrate and tellurite of the production of catalase the production of indole from peptone the formation of  $\text{NH}_3$  and  $\text{H}_2\text{S}$  the final hydrogen ion concentration in glucose broth and the power to utilize certain salts such as tartrates and citrates.

As a rule the fermentation of sugars is observed qualitatively the formation of acid being rendered evident by the inclusion in the medium of an indicator and the liberation of gas by an inverted Durham tube or by a special fermentation tube. For the testing of the other biochemical properties certain fairly stereotyped methods have been evolved which it is unnecessary to describe here.

In bacterial differentiation the biochemical reactions are often of the greatest importance in many groups of organisms the classification is made on the basis of sugar fermentation of proteolytic power or of both tests taken together. The sugar tests especially afford a means of bringing out the finer distinctions between closely allied organisms. The oxidation of tartrates and citrates is employed in the differentiation of the coliform group of bacteria.

**G Antigenic Structure**—For identifying bacteria the serological reactions most frequently employed are agglutination and complement fixation. Provided that adequate controls are used these reactions—particularly agglutination—afford the most rapid and reliable method of identifying a given bacterium. For the identification to be complete the organism should be agglutinated to titre by a serum prepared against the organism which it is supposed to resemble and should absorb all the agglutinins from that serum moreover the type organism should be agglutinated to titre by a serum prepared against the unknown organism and should likewise remove all agglutinins from it. That is to say there should be complete cross agglutination and cross absorption between the two sera and the two organisms. In certain groups of bacteria the serological method is found to be much the quickest and most satisfactory way of distinguishing between the different members and it is therefore extensively used for rapid identification. In other groups the agglutination method is not of much help either because there are numerous varieties within the species or because the organisms are auto-agglutinable or for some other reason. Apart from affording a rapid means of

and if it is desired to preserve or to publish a description of the new species or type it is essential to make a full and careful record of all its characters and reactions so that they will be available for comparison with organisms isolated at a future date or with organisms isolated by other observers. Such a record should always contain careful comparisons with those species or types which most nearly resemble the newly isolated organism.

## REFERENCES

- BARBER, M. A. (1908) *J. infect. Dis.* 5, 379.  
 CARNOT, P. and GARNIER, M. (1907) *C. R. Soc. Biol.* 54, 748.  
 CRAIGIE, J. (1931) *J. Immunol.* 21, 417.  
 FLEMING, A. (1937) *J. Path. Bact.* 35, 831.  
 HENRICI, A. T. (1926) *J. infect. Dis.* 38, 54.  
 KOCH, R. (1880) Investigations into the Etiology of the Traumatic Infective Diseases. New Sydenham Soc. Lond. (1881) *Mit Reichsgesundh.Amt* 1, 1.  
 LISTER, J. (1878) *Quart. J. med. Sc.* 18, 177.  
 ROVIDA, G. (1925) *Sperimentale* 79, 1003.  
 TOPLEY, W. W. C., BARNARD, J. E. and WILSON, G. S. (1921) *J. Hyg. Camb.* 20, 221.  
 TULLOCH, W. J. (1939) *J. Hyg. Camb.* 39, 324.  
 WILSON, G. S. (1926) *J. Hyg. Camb.* 25, 150.

identification of certain bacteria the serological method is as a rule the most delicate method available for bringing out the finer distinctions between closely allied organisms. In this respect it is more valuable even than the biochemical tests. It is often the only method available for differentiating between the sub-species or varieties of a given species of organism. Of recent years increasing attention has been paid to the precipitin test which is particularly useful when homogeneous suspensions of bacteria cannot be obtained for agglutination or when, as with the haemolytic streptococci a particular antigen can be extracted from the organisms and recognized quickly by a simple precipitin technique.

**H. Pathogenicity**—The pathogenicity of bacteria is usually tested on laboratory animals especially the guinea pig rabbit rat and mouse. It may be advisable to introduce the organism directly into the tissues by inoculation subcutaneously intramuscularly intraperitoneally, or intravenously or it may be given by the mouth or in the form of a spray, which the animal is made to inhale. Pathogenicity tests are open to numerous errors, but provided these are adequately guarded against they often afford very important information. This is limited however to certain groups of organisms. In the study of the purely saprophytic bacteria and of certain bacteria that are harmless to laboratory animals, the pathogenicity test is of no value except to establish the absence of virulence. It is used chiefly in distinguishing virulent from avirulent members of the same genus or species. But it is also used to distinguish between closely allied organisms both of which are virulent to the same animal but which produce in it lesions of varying extent or localization or which differ in their virulence to different species of animal.

For the complete identification of a pathogenic species, it may be necessary to determine whether or not it forms a soluble exotoxin that is to say, whether sterile filtrates of cultures grown for a suitable time in suitable fluid media produce death with characteristic lesions. In species which produce such exotoxins neutralization with a specific antitoxin may play an important part in identification. In the description of any newly isolated pathogenic species a record of the toxicity or non toxicity of filtrates should always be included.

It will be realized that the complete identification of an unknown organism is often a lengthy proceeding. As a rule it is easy to refer it to its proper genus this can be done by simple examination of the morphological and staining reactions, aided at times by the cultural reactions. But its more exact denomination requires the use of the most delicate tests at our disposal namely the biochemical, serological, and pathogenicity tests. It is advisable never to place too much weight on any one test errors of technique or of interpretation are always liable to occur. If a large series of tests is carried out and the organism is studied by several different methods, then the chances of being misled are very greatly reduced.

The complete identification of a given organism with any known type is not, of course, always possible, within the type species there are often varieties differing in minor respects from the type organism. It is very important to realize this in any large collection of organisms of apparently the same species, there will almost invariably be found a number that differ from the rest in one or more of their properties sometimes these differences are so numerous, or a single one of them may be so important that it is necessary to revise one's classification.

If the characters of the organism differ from those of any described species,

and if it is desired to preserve or to publish a description of the new species or type it is essential to make a full and careful record of all its characters and reactions so that they will be available for comparison with organisms isolated at a future date or with organisms isolated by other observers. Such a record should always contain careful comparisons with those species or types which most nearly resemble the newly isolated organism.

## REFERENCES

- BARBER M A (1908) *J infect Dis* 5 379  
 CARNOT P and GARNIER M (1902) *C R Soc Biol* 54 718  
 CRAIGIE J (1931) *J Immunol* 21 417  
 FLEMING A (1931) *J Path Bact* 35 831  
 HENRICI A T (1926) *J infect Dis* 38 54  
 KOCH R (1880) Investigations into the Etiology of the Traumatic Infective Diseases  
 New Sydenham Soc Lond (1881) *Mitt ReichsgesundhAmt* 1 1  
 LISTER J (1878) *Quart J micr Sci*, 18 177  
 ROVIDA G (1925) *Sperimentale* 79 1053  
 TOPLEY W W C, BARNARD J E and WILSON G S (1921) *J Hyg Camb* 20 224  
 TULLOCH W J (1939) *J Hyg., Camb.*, 39 324  
 WILSON G S (1926) *J Hyg Camb* 25 150

## CHAPTER 13

### DESCRIPTION OF THE METHODS USED IN THE SYSTEMATIC EXAMINATION OF BACTERIA, AND A GLOSSARY OF THE TERMS EMPLOYED

WE have already dealt in Chapter 12 with the methods of isolating pure cultures of bacteria, and with the various criteria that are employed in their identification. In the present chapter we describe a routine which may be used in examining the various *morphological*, *cultural*, and *biochemical* properties of bacteria, and define the terms which we shall employ in the description of these properties.

**Morphological Appearance of Bacteria.**—The chief points to be noted are the following

*Shape*—Spheres, short rods, long rods, filaments, commas, or spirals

*Axis*—Straight or curved.

*Size*—Length and breadth

*Sides*—Parallel, bulging, concave, or irregular

*Ends*—Rounded, truncate, concave, or pointed

*Arrangement*—Singly, in pairs, in chains, in fours, in groups, in grape-like clusters in cubical packets, in bundles, or in Chinese letters.

*Irregular Forms*—Variations in shape and size, club, filamentous, branched, navicular, citron, fusiform, giant swollen forms and shadow forms.

*Motility*—Motile or non motile

*Flagella*—Monotrichate, amphitrichate, lophotrichate, peritrichate (Fig 13, p 31)

*Endospores*—Spherical, oval, or ellipsoidal, equatorial, subterminal, or terminal, single or multiple, causing bulging of bacillus or not (Fig 6, p 24)

*Capsules*—Present or absent

*Staining*—Even, irregular, unipolar, bipolar, beaded, barred, and variations in depth between different organisms. Presence of metachromatic granules, reaction to Gram and to Ziehl-Neelsen stains

#### Surface Colonies on Solid Media.

*Shape*—Circular, irregular, radiate, rhizoid.

*Size*—In millimetres

*Elevation*—Effuse, raised, low convex, convex or dome-shaped, umbonate, umbilicate, with or without bevelled margin

*Structure*—Amorphous, fine, medium, or coarsely granular, filamentous, curled.

*Surface*—Smooth, contoured, beaten-copper, rough, fine, medium, or coarsely granular, ringed, papillate, dull or glistening

*Edge*—Entire, undulate, lobate, crenated, erose, fimbriate, curled, effuse

- Colour*—Colour by reflected and transmitted light, fluorescent, indescent opalescent self-luminous
- Opacity*—Transparent, translucent, or opaque
- Consistency*—Butyrous viscid, friable membranous
- Emulsifiability*—Easy or difficult, forms homogeneous granular, or membranous suspension when rubbed up in a drop of water with a platinum loop
- Differentiation*—Differentiated into a central and a peripheral portion (Fig 57).

#### Growth on Stroke Culture

- Degree*—None, scanty, moderate abundant profuse, discrete or confluent
- Form*—Filiform, spreading, rhizoid
- Elevation*—Effuse or raised
- Surface*—Smooth, contoured, beaten-copper, rough, finely, moderately, or coarsely granular, papillate, heaped up, dry or moist
- Edge*—Entire undulate, lobate, crenated, erose, fimbriate, curled effuse
- Colour, Opacity, Consistency and Emulsifiability*—As for colonies.
- Odour*—Absent, decided resembling—
- Medium*—Coloured, digested, crystal formation (Fig 57)

#### Growth in Stab Culture

- Degree*—As for stroke culture Also position of optimal growth
- Form*—Filiform, beaded with or without branching
- Extent*—Depth in tube to which growth occurs
- Surface*—Surface growth present or absent, if present, diameter surface and edge
- Colour and Opacity*—As for stroke culture
- Liquefaction*—Present or absent, if present crateriform napiform infundibuliform, saccate, or stratiform
- Medium*—As for stroke culture (Fig 57)

#### Growth in Shake Culture

- Position*—Uniform growth throughout tube, or position of optimal growth
- Surface*—Surface growth present or absent
- Colonies*—Size, shape, colour, opacity, outgrowths from periphery if any
- Gas*—Present or absent, medium disrupted
- Medium*—Coloured, digested, or rendered turbid

#### Growth in Fluid Medium.

- Degree*—None, scanty, moderate abundant or profuse
- Turbidity*—Present or absent, if present slight moderate or dense uniform granular, or flocculent
- Deposit*—Present or absent, if present slight moderate, or abundant, powdery granular, flocculent membranous or viscid, disintegrating completely or incompletely on shaking
- Surface Growth*—Present or absent, if present ring growth around wall of tube or surface pellicle, which is thin or thick with a smooth granular or rough surface, and which disintegrates completely or incompletely on shaking
- Odour*—Absent, decided, resembling—



## Growth in Blood Agar

Colonies—Description of surface colonies

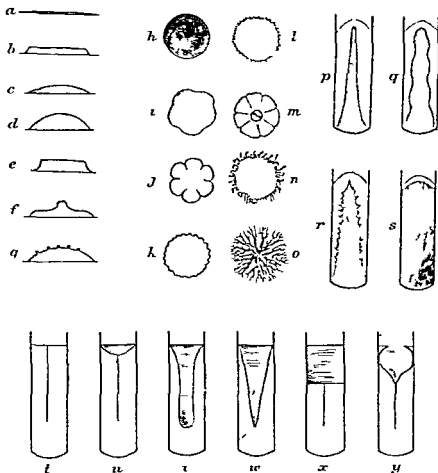
Hæmolysis—Present or absent, if present of  $\alpha$  or  $\beta$  type

FIG 5

a-g Elevation of colonies. a Flat or effuse b Raised c Low convex. d Convex or dome shaped. e Raised with concave bevelled edge f Umbonate g Convex with papillate surface h-o Edge of colonies h Entire i Undulate j Lobate k Crenated. l Erode or dentate m Radially striated periphery with lobate edge n Fimbriate o Rhizoid or arborescent. p-s Growth on agar streak culture p Filiform. q Slightly spreading with undulate edge. r Slightly spreading with erose edge. s Spreading t-y Growth in gelatin stab culture. t Filiform growth without liquefaction. u Crateriform liquefaction. v Saccate liquefaction. w Infundibuliform liquefaction. x Stratiform liquefaction. y Vapiform liquefaction.

Resistance—Tested usually by placing a 24-hours' broth culture, containing 5 ml. of medium in a  $\frac{1}{8}$ -inch test tube, in a water bath at such temperatures as 55° C for 1 hour, 60° C for half an hour, and 80° C. for ten minutes, and sub-

culturing into a favourable medium. This is of course, merely a rough differential test. For accurate purposes, the column of fluid culture should be enclosed in a capillary tube thin enough to ensure rapid heating of the organisms to the temperature of the water in the bath.

### Metabolic Properties

*Oxygen Pressure required for Growth*—Aerobic, facultatively anaerobic, obligatory anaerobic, microaerophilic

*Increased Carbon Dioxide pressure required for Growth*—The growth of many organisms is favoured by a partial pressure of CO<sub>2</sub> higher than that—0.03 per cent—in the atmosphere, and some entirely fail to grow in the absence of a raised pressure.

*Effect of Temperature on Growth*—Limits between which growth occurs. Optimal temperature for growth.

*Pigment Formation*—Tested usually on an agar slope incubated at 22° C, or left at room temperature in the light after preliminary incubation at 37° C.

*Effect of Modifying the Constitution of the Medium*—Effect on growth of adding to the medium blood, serum, ascitic fluid, glucose, glycerine, potassium nitrate, bile salts, or other substances.

### Biochemical Reactions

*Fermentation of Sugars*—Tested in 1 per cent peptone water containing 1 per cent of the sugar and Andrade's indicator. A Durham's tube is included. For certain groups of organisms, which do not grow well in this medium, 5 per cent of serum is added. Horse serum may be used as a rule but in testing the fermentation of maltose it is better replaced by human or rabbit serum, since it contains an enzyme maltase, which may lead to a false reaction (Hendry 1938). Acid or acid and gas production is noted.

*Litmus Milk*—No change, acid or alkali, clot, clot disrupted by gas, peptonization, saponification. The term "clot" is unfortunately used for both an acid clot and a rennet clot. An acid clot results from the precipitation of the caseinogen, it is soft gelatinous, does not retract and can be completely dissolved in alkali. A rennet clot is due to the coagulation of the caseinogen under the influence of bacterial enzymes. A few hours after its formation it retracts with the expression of a clear greyish-coloured fluid called whey, the clot itself is firm and cannot be dissolved by alkali. Calcium caseinogenate is soluble in water. When acid is produced from the lactose of the milk, the calcium combines with it and the caseinogen, which is insoluble, is precipitated. This is the mechanism of formation of the acid clot. In coagulation by rennet the soluble calcium caseinogenate is converted into insoluble calcium caseinate, which forms the curd.

*Indole*—Tested in 1 per cent peptone water after 5 days growth using Bohme's reagents. One ml of ether is added to the culture, which is shaken thoroughly, and then allowed to stand till the ether collects on the surface. 1 ml Solution A is run down the side of the tube, if no colour appears within a minute, 1 ml. of Solution B is added. A positive reaction is characterized by a colour varying from a faint pink to a deep magenta. According to Happold and Hoyle (1934) xylene is better than ether.

## Solution A

<i>para</i> -dimethylaminobenzaldehyde	4 gm.
96 per cent. Alcohol	380 "
Concentrated HCl	80 "

## Solution B

Saturated watery solution of potassium persulphate.

An alternative method depending on the volatility of indole at 37° C, is recommended by Holman and Gonzales (1923). It consists in placing a strip of filter paper soaked in a saturated watery solution of oxalic acid and subsequently dried between the cotton wool plug and the tube. The paper should be carefully folded so as to present the maximum surface to the volatilizing indole, which turns it a pink colour.

Some organisms form indole, but break it down more rapidly than they produce it, and hence may give a false negative reaction (Reed 1942).

**Methyl Red Test (M.R.)**—Tested by adding 5 drops of an 0.04 per cent. solution of methyl red to a culture in glucose phosphate medium (peptone 0.5 gm.,  $K_2HPO_4$  0.5 gm., glucose 0.5 gm., water 100 ml., pH 7.5). Culture grown for 5 days at 30° C or 3 days at 37° C.

Red colour = positive

Yellow colour = negative

**Voges Proskauer Test (V.P.)**—Tested by adding 1 ml. of a 10 per cent. solution of KOH to a glucose phosphate culture grown for 5 days at 30° C or 2 days at 37° C. The colour develops slowly, and the test should be read after 18 to 24 hours.

Pink fluorescence = positive

No coloration = negative

A higher proportion of positive reactions is obtained by the use of O'Meara's (1931) modification. A knife point of creatine is added to the culture, followed by 5 ml. of 40 per cent. sodium hydroxide. The tube is shaken thoroughly for 2 to 5 minutes. A positive reaction is characterized by the appearance of a pink colour within about 2 minutes, unaccompanied by fluorescence. The development of the colour may, however, be delayed for an hour or longer.

An even more sensitive test for acetylmethylcarbinol is that described by Barnett (1936). It consists in adding 0.5 ml. of a 5 per cent. alcoholic solution of  $\alpha$ -naphthol and 0.2 ml. of 40 per cent. KOH solution to 1 ml. of culture. In a positive reaction a pink colour appears in 2-5 minutes, deepening to magenta or crimson in half an hour. In a negative reaction the mixture remains colourless for an hour or so, when it may become copper-coloured owing to the action of KOH on the  $\alpha$ -naphthol. Traces of pink coloration are best neglected.

**Nitrate Reduction.**—Tested on a broth culture containing 0.1 per cent.  $NaNO_3$ , grown for 5 days at 37° C by the Griess-Nosva method.

## Solution A

$\alpha$ -naphthylamine	1 gm.
Water	22 ml.

Dissolve filter, and then add 180 ml. of dilute acetic acid (sp. gr. 1.04).

**Solution B**

Sulphanilic acid 0.5 gm

Dilute acetic acid 150 ml

Add 1 ml of Solution A followed by 1 ml of Solution B

Pink red or maroon colour = positive

No coloration = negative

A negative reaction may sometimes be due to the reduction of the nitrite to gaseous nitrogen almost as rapidly as it is formed or to the production of hydroxylamine. The first possibility may be examined by growth in a gas fermentation tube, or by chemical estimation of the nitrate the second by testing for nitrite in the way just described after preliminary oxidation of the hydroxylamine with iodine (see Landsey and Rhines 1932 Conn 1936 Reed 1912). A control tube should always be tested.

**Ammonia**—Tested on a peptone water culture grown for 5 days at 37° C by adding Nessler's reagent

Brown colour = positive

Faint yellow colour = negative

**Hydrogen Sulphide**—Tested on lead acetate medium (heart extract broth containing 4 per cent peptone and 2.5 per cent agar. Sterilize and add an equal quantity of a sterile 0.1 per cent solution of basic lead acetate.)

Brown or black coloration = positive

No coloration = negative

The lead acetate may be replaced by 0.05 per cent ferric ammonium citrate or 0.03 per cent ferrous acetate (Zobell and Feltham 1934). A higher proportion of positive reactions is obtained with some organisms by incubating at 30° C instead of 37° C (Tittler 1931). The most delicate method is to grow the organisms in a slope tube of liver extract agar and to include between the cotton wool plug and the tube a slip of filter paper soaked in 10 per cent lead acetate solution and subsequently dried. The amount of browning or blackening of the paper is measured in millimetres. A fresh slip may be inserted daily.

**Methylene Blue Reduction**—Tested on a 24 hours broth culture at 37° C. Add 1 drop of 1 per cent aqueous methylene blue and incubate at 37° C.

Complete decolorization = strong positive

Green coloration = weak positive

No decolorization = negative

**Catalase**—Tested on a 24 hours agar slope culture at 37° C. One ml of H<sub>2</sub>O<sub>2</sub> (10 vols) is poured over the growth and the tube is set in an inclined position.

Gas bubbles produced = positive

No gas produced = negative

For an account of the methods of examining the antigenic structure and the pathogenicity of bacteria reference must be made to the chapters dealing with the particular organism under consideration.

**GLOSSARY OF DESCRIPTIVE TERMS**

**Aerobic** growing in the presence of free oxygen *strictly aerobic* growing only in the presence of free oxygen

- Amorphous* (colonies) without visible differentiation in structure.
- Amphitrichate* having a single flagellum at each pole
- Anaerobic* growing in the absence of free oxygen, *strictly anaerobic*, growing only in the absence of free oxygen, *facultatively anaerobic*, growing both in the presence of and in the absence of oxygen. It must be noted, however, that growth of even strict anaerobes will occur in the presence of molecular oxygen, provided the medium contains a reducing system capable of bringing about a sufficiently low O R potential (see Chapters 3 and 36)
- Banded* (stained bacteria) deeply staining granules arranged at regular intervals along the course of the rod (In stab or stroke culture) disjointed or semi confluent colonies along the line of inoculation
- Beaten copper* multiple small crateriform depressions on the surface of a growth, resembling beaten copper
- Bipolar* at both ends or poles of the bacterial cell
- Butyrous* growth of butter like consistency
- Chains* four or more bacterial cells attached end to end
- Chromogenesis* the production of colour
- Citron* shaped like a lemon, having a small knob at each end
- Clavate* club shaped
- Coagulation* formation of a firm clot in milk with the subsequent separation of the casein from the whey
- Contoured* an irregular, smoothly undulating surface
- Convex* the segment of a sphere of short radius, *Low convex* the segment of a sphere of long radius
- Crateriform* a saucer shaped liquefaction of the medium.
- Crenated* small shallow indentations of the edge, which has a scalloped appearance
- Cuneate* wedge shaped
- Curled* composed of parallel chains in wavy strands, as in anthrax colonies
- Effuse* growth thin, hardly raised at all from the medium
- Endospores* thick walled spores formed within the bacterial cell
- Entire* with an even margin.
- Equatorial* situated about equidistant from each end
- Erose* border showing fine, pointed, tooth like projections
- Filaments* applied to morphology of bacteria, refers to thread like forms, generally unsegmented, if segmented, to be distinguished from chains (*qv*) by the absence of constrictions between the segments
- Filamentous* growth composed of long, often interwoven threads
- Filiform* in stroke or stab cultures, a uniform growth confined to the line of inoculation
- Fimbriate* fine, sometimes recurved, processes projecting from the edge of the colony or growth
- Flocculent* containing small adherent masses of bacteria of various shapes floating in the culture fluid, or deposited at the bottom
- Fluorescent* having one colour by transmitted light and another by reflected light
- Friable* growth dry and brittle, when touched with a platinum needle
- Granular* composed of granules, fine, medium or coarse

- Hæmolysis** on blood agar plate  $\alpha$  hæmolysis ' colonies surrounded by a greenish ring  $\beta$  hæmolysis ' colonies surrounded by an area of clearing, which is transparent (see Chapter 24)
- Heaped up** irregular, coarse processes projecting considerably above the level of the rest of the growth
- Infundibuliform** in form of a funnel or inverted cone
- Iridescent** exhibiting changing rainbow colours in reflected light
- Lenticular** surface colony, which is convex and translucent, and which acts like a plano convex lens, giving an inverted image of an object viewed through it Deep colony, which is shaped like a lentil
- Lobate** having the margin deeply undulate, producing lobes (see *Undulate*)
- Lophotrichate** having a tuft of flagella at one or both poles
- Luminous** glowing in the dark, phosphorescent
- Maximum Temperature** temperature above which growth does not take place
- Membranous** growth thin, coherent, like a membrane
- Microaerophilic** growing best under a lowered oxygen pressure
- Minimum Temperature** temperature below which growth does not take place
- Mirror like** having a smooth glistening surface, in which reflections of surrounding objects, *e.g.* window bars, can be seen
- Monotrichate** having a single flagellum at one pole
- Napiform** liquefaction in form of a turnip
- Navicular** shaped like a boat.
- Opalescent** coarsely iridescent, like an opal
- Opaque** objects, *e.g.* window bars cannot be seen through growth
- Optimum Temperature** temperature at which growth is most rapid
- Papillate** growth beset with small nipple like processes
- Pellicle** bacterial growth forming either a continuous or an interrupted sheet over the culture fluid
- Peptonization** rendering curdled milk soluble by the action of peptonizing enzymes
- Peritrichate** having flagella disposed around the organism
- Punctiform** very small but visible to naked eye, under 1 mm in diameter
- Radiate** showing fissures or ridges arranged in a radial manner
- Raised** growth thick, with a comparatively flat surface, and with abrupt or terraced edges
- Rhizoid** growth of an irregular branched or root like character, as in *B mycorides*
- Ring** growth at the upper margin of a liquid culture, adhering to the glass
- Ringed** having one or more circular depressions or elevations on the surface, sometimes giving a draughtsman like appearance
- Rough** general term for an irregular surface the irregularity being of a coarsely granular type, or resembling morocco-leather or a relief map
- Saccate** liquefaction in form of an elongated sac, tubular, cylindrical
- Spreading** growth extending much beyond the line of inoculation, *i.e.* several millimetres or more, sometimes over an entire tube or plate
- Stratiform** liquefying to the walls of the tube at the top and then proceeding downwards horizontally
- Subterminal** situated towards the end
- Terminal** situated at the extreme end.

*Translucent*: objects, e.g. window bars, are visible through growth, but growth is not water-clear.

*Transparent*: growth is water-clear.

*Truncate*: ends abrupt, square.

*Turbid*: cloudy; may be a uniform, flocculent, or granular turbidity.

*Umbonate*: having a button-like, raised centre.

*Undulate*: border wavy, with shallow sinuses.

*Unipolar*: at one end only of the bacterial cell.

*Viscid*: sticky, semi-fluid; on withdrawal of the needle, the growth follows it in the form of a thread; sediment on shaking rises as a coherent swirl.

#### REFERENCES

- BABBITT, M. M. (1936) *J. Path. Bact.*, **42**, 441.  
CONN, H. J. (1936) *J. Bact.*, **31**, 225.  
HAPFOLD, F. C. and HOYLE, L. (1934) *Biochem. J.*, **28**, 1171.  
HENDRY, C. B. (1938) *J. Path. Bact.*, **46**, 383.  
HOLMAN, W. H. and GONZALES, F. L. (1923) *J. Bact.*, **8**, 577.  
LINDSEY, G. A. and RHINES, C. M. (1932) *J. Bact.*, **24**, 489.  
O'MEARA, R. A. Q. (1931) *J. Path. Bact.*, **34**, 401.  
REED, R. W. (1942) *J. Bact.*, **44**, 425.  
TITSLER, R. P. (1931) *J. Bact.*, **21**, 111.  
ZOBELL, C. E. and FELTHAM, C. B. (1934) *J. Bact.*, **28**, 169.

## CHAPTER 14

### ACTINOMYCES AND ACTINOBACILLUS

#### ACTINOMYCES

**DEFINITION** *Actinomyces*, Harz 1877

Organisms growing in the form of a much branched mycelium which may break up into segments or produce 'spores'. Aerial mycelium often formed under suitable conditions. Mainly aerobic, but may be microaerophilic or anaerobic. Usually saprophytic, but some species are parasitic on plants or animals and may give rise to disease. In animal body organisms are frequently arranged in colonies composed of radiating threads with clubbed ends. Non motile. Some species are acid fast. The type species is *Actinomyces bovis* Harz.

The term *Actinomyces bovis* was originally given by Harz to a mould like organism which was found by Bollinger (1877) in the lesions of cattle suffering from a peculiar disease of the tongue and jaw, now known as Actinomycosis. This organism was first cultivated by Wolff and Israel in 1891 under anaerobic conditions. An aerobic organism, which is occasionally present in actinomycotic lesions, but which is probably not aetiologicaly related to the disease, was isolated in the same year by Bostroem (1891). In order to avoid confusion with the anaerobic pathogenic type, we have suggested that Bostroem's organism should be called *Actinomyces graminis*. Since then a number of similar organisms have been isolated from a variety of diseases in man and animals, and from such situations as soil, grains, and grasses.

These organisms appear morphologically as jointed or unjointed filaments, which frequently show true branching. In culture, rod forms are not uncommon. In the animal body, many of the pathogenic species are characterized by the formation of granules of varying size, which are found to consist of a filamentous mycelium surrounded by radiating clubs—a picture which is responsible for the term "ray fungus" (Botanically the term 'ray' refers to the marginal portion of a composite flower, consisting of ligulate florets arranged radially). In their staining reactions some species are acid fast, though the majority are non acid fast. It is evident that these organisms bear some resemblance to those of *Mycobacterium*, and in the classification furnished by the American Committee of Bacteriologists in 1917 (Report 1917), *Actinomyces* and *Mycobacterium* were included in a single family, known as the *Mycobacteriaceæ*. In the 1920 report (Report 1920), however, it was decided to create a separate family of *Actinomycetaceæ*, which should contain the genera *Actinobacillus*, *Leptothrix*, *Actinomyces*, and *Erysipelothrix*. For descriptive purposes it is convenient to consider these genera separately, and in the present chapter we shall confine ourselves to a description of *Actinomyces* and *Actinobacillus*.



Other terms such as *Streptothrix* or *Nocardia* have been applied to organisms of the *Actinomyces* group. Since the term *Streptothrix* was given by Corda in 1839 to a genus of fungi belonging to the *Hyphomycetes*, quite different from the group that we are considering, this name is obviously inapplicable. The term *Nocardia* was coined later than *Actinomyces*, and is likewise inapplicable.

**Habitat**—Many members of this group lead a saprophytic existence on grains and grasses, and in water. As these substances are widely used as foods it is not unnatural that by their means *Actinomyces* often gains access to the alimentary and respiratory tracts of man and other animals. They have been isolated chiefly from man and cattle, and also from pigs, chickens, rabbits, dogs, elephants, lizards and oysters (Foulerton 1910). Most members live in the soil, where they play an important part in the biological processes that are occurring there.

There is at least one species that appears to be a strict parasite in man and animals. Numerous members are pathogenic for plants causing such diseases as potato scab.

**Morphology**—On culture media the morphology is variable. In ordinary film preparations the anaerobic Wolff Israel type occurs chiefly as rods 3-4  $\mu$  long by 0.6  $\mu$  broad which from their arrangement, their clubbed ends, and their irregular staining bear a resemblance to certain members of the corynebacteria, careful search, however, will generally reveal a few definite filaments, some of

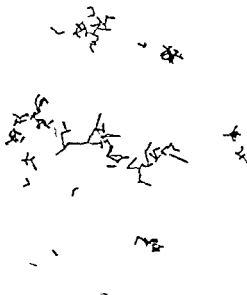


FIG 58—*Actinomyces botis*  $\times 1000$

From an agar slope culture 14 days 37° C. anaerobically

which may show true branching (Fig 58). The aerobic type in young cultures occurs chiefly as long unsegmented straight or wavy filaments which show simple or dichotomous branching (Fig 59) and which not infrequently grow upwards from the surface as aerial hyphae. Later these filaments undergo segmentation and break up into rod forms of varying length and oval coccoid bodies which are usually referred to as spores. In some strains of the aerobic type segmentation is visible within 24 hours, in others it may not occur for 3 weeks or more. On solid media the filaments are arranged in loose groups or in a tangled mycelium, but in broth definite colonies occur consisting of a densely matted central core of filaments and a peripheral zone in which the filaments are more loosely disposed (Fig 63). These colonies in liquid media are common to both the aerobic and the anaerobic types. The rods and filaments may stain evenly, but as a rule granular staining is evident. After growth for some time in liquid media both the aerobic and the anaerobic types may show involution

forms, consisting mainly of spherical or club shaped swellings on the ends of the filaments

If, instead of making film preparations, the growth of the organisms is followed by Orskov's (1923) agar block technique, it will be seen that in some species the aerial mycelium gives rise, without preliminary segmentation, to circular or oval "spores" There is evidence that these "spores" are rather more resistant to inimical agencies generally than the plum mycelium They may, for example, resist moist heat at 65° C for as long as 3 hours The segmented mycelium, observed in many species, is no more resistant, however, than the unicellular mycelium

All members are non motile and all, with a few possible exceptions, are Gram positive The anaerobic types are uniformly non acid fast The aerobic types may be differentiated into

(1) acid fast, these resist decolorization with 1 per cent sulphuric acid for 5 minutes, but are usually decolorized by the application of 25 per cent  $H_2SO_4$  for a similar length of time, there is, however, a marked variation in the acid fastness of different species (2) Non acid fast

In the animal body the morphology is often different from that on culture media The anaerobic Wolff Israel type grows in the form of definite colonies, which appear in the pus or in sections of the tissues as granules or "Drusen"

When crushed and examined microscopically these granules are seen to consist of a central filamentous Gram positive mycelium surrounded by a peripheral zone of large, Gram negative clubs In old colonies the mycelium is replaced by a mass of short Gram positive rods and coccoid bodies which appear to have resulted from the disintegration of the filaments The clubs vary in size, but may be as long as 10  $\mu$  and as broad as 5  $\mu$  Their mode of origin has given rise to much discussion On the whole it seems probable that there are two entirely different types of club, one observed in artificial culture and derived from the organism itself, the other observed in the animal body and derived from the host The first type, or "culture club" as Orskov (1923) calls it, represents the swollen end of the mycelial filament The second type or "tissue club," as we may call it, appears to be due to the deposition around the end of the filament of some material, probably rich in lipid, by the tissues of the host Tissue clubs are observed, not only in ray fungus infections, but also in lesions caused by other

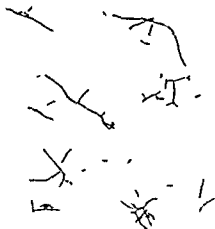


FIG 59—*Actinomyces graminis*  $\times 1000$   
From a broth culture 24 hours 37° C aerobically

organisms such as tubercle bacilli and staphylococci. Even dead tubercle bacilli are said to stimulate their production.

In sections of tissues the filaments may be differentiated from the clubs by a modified Ziehl Neelsen stain. If a section is stained with carbol fuchsin, decolorized for 20 to 30 seconds with 1 per cent  $H_2SO_4$  and counterstained with methylene blue, the clubs appear red and the filaments blue.

The aerobic types, when growing in the animal body, generally form a tangled mycelium without evidence of ray or of club formation, but exceptions do occur, as with *Actinomyces madurae*, which forms definite granules similar to those of the Wolff Israel type.

The most striking feature of the *Actinomyces* is their pleomorphism. All forms may be seen—filaments, rods, cocci and even spirilla. In the anaerobic type rod forms predominate in culture, in *Actinomyces madurae* filaments. But in most of the aerobic types all forms are seen, coexisting in a single culture. For a detailed

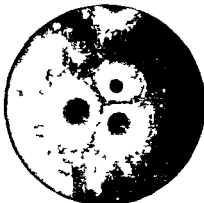


FIG 60—*Actinomyces graminis*.  
Colonies on agar plate 7 days, 37° C. aerobically  $\times 8$

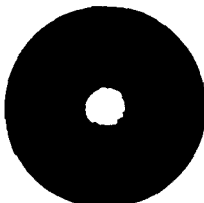


FIG 61—*Actinomyces bovis*.  
Colony on agar plate 14 days, 37° C. anaerobically  $\times 8$

description of their morphology, the reader is referred to a monograph by Laeske (1921).

**Cultural Reactions**—In general, growth on artificial media is readily obtained. The usual media suffice, but the addition of glucose or glycerol is beneficial. The aerobic types, with a few exceptions, multiply rapidly, so that in 24 hours definite evidence of growth is visible on agar. The anaerobic types, on the other hand, grow more slowly, taking 3 or 4 days to form macroscopic colonies. Great diversity of cultural appearance is noticeable, particularly in the aerobic species. The descriptions that follow refer only to some of the commoner types.

On an agar plate the aerobic types form round, low convex opaque, finely granular colonies, which later undergo differentiation into a raised, knob-like, sometimes radially striated centre and an effuse, ground-glass like periphery. The surface is finely granular and often has a "chalk powder" covering due to the formation of aerial spores, the edge is rhizoid, indented, or feathery (Fig 60). Most strains form pigment—yellowish, pink, or orange in colour—which becomes apparent after a few days' incubation, and which may show progressive alterations in tint. This is especially noticeable in cultures that have been incubated at 37° C.,

and subsequently left in the dark at room temperature under suitable conditions, aerial hyphæ may develop, giving rise to a characteristic bloom on the surface of the colony—the chalk powder appearance just described.

The anaerobic Wolff Israel type forms smaller colonies, not apparent for 3 or 4 days, they are more compact, greyish or porcelain white in colour, and have a nodular surface (Fig 61)

On glycerol or glucose agar the aerobic types give a luxuriant, confluent heaped up, worm cast pigmented growth, adherent to the medium, of tough consistency, and difficult to emulsify (Fig 62). The anaerobic type grows in the form of discrete colonies, which are only slightly adherent to the medium, and are much easier to emulsify.

In a glucose agar shake culture the aerobic types give a thick pigmented growth confined entirely or almost entirely to the surface. The anaerobic type gives a characteristic band like growth situated about 0.5 to 1 cm below the surface with a few larger discrete colonies scattered throughout the medium below. No growth at all occurs in the upper few millimetres.

In broth the aerobic types often form a thick, dry, dull, scaly or nodular, pigmented surface pellicle, which may extend for some distance up the sides of the tube. A ropy or membranous, sometimes pigmented sediment

After a variable time



FIG 62 —  
*Actinomyces graminis*  
Culture 14 days 37° C  
glycerine agar slope  
aerobically

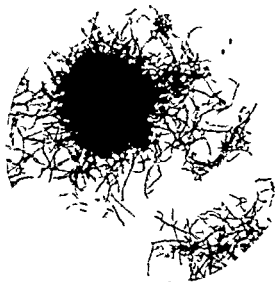


FIG 63 — *Actinomyces graminis* × 1000  
A small granule composed of radiating filaments, from a  
broth culture 24 hours 37° C aerobically

forms augmented by frequent deposits from the surface membrane. The broth remains clear, or at most shows a finely granular turbidity. Aerial hyphæ may sprout from the surface pellicle. Sometimes growth commences at the bottom and characteristic fluff balls, resembling the head of a seeding dandelion, develop. The anaerobic type grows in the form of compact whitish granules with a nodular surface deposited at the bottom of the tube, there is no turbidity and no surface growth. Nitrate broth is more favourable for growth than ordinary broth.

The cultural characteristics on other media can

be ascertained from the descriptions of the individual species

**Resistance**—The members of this group show no special resistance to heat or disinfectants. Most are killed in 15 minutes when exposed to moist heat at 60° C. Some attempts have been made to find whether filaments containing so-called spores are specially resistant. Vincent (1894) working with *Actinomyces madura* stated that the spores were killed at 80° C in 3 minutes whereas the non sporing forms were killed at 60° C in 3 to 5 minutes. Ørskov (1923) has likewise found that the spores are more resistant than the plain mycelium; they may survive exposure to moist heat for 3 hours at 65° C. Goyal (1936) however was unable to show that the spores were any more resistant than the filaments; both were killed by a time temperature combination varying with different strains from 30 minutes at 60° C to more than one hour at 72° C. There is of course a great difference between the resistance of the *Actinomyces* spores and that of true bacterial spores.

Cultures of the aerobic type if kept at room temperature remain viable for months; cultures of the anaerobic type usually die out in about 6 to 8 weeks. If kept in the incubator they die very much more quickly.

**Growth Requirements**—There is a fairly sharp division between the aerobic and the anaerobic types. The aerobic types are unable to grow under strictly anaerobic conditions while the anaerobic types are unable to grow at any rate on solid media and when first isolated in the presence of air. In liquid media especially in nitrate broth and in the depths of solid media such as glucose agar the anaerobic types may flourish when kept under aerobic conditions showing that they are not strict anaerobes but rather organisms having a preference for a low oxygen pressure—micro-aerophiles. Intermediate types are met with (Bruns 1899; Lignieres and Spitz 1903) usually having a preference for anaerobic conditions (Naeslund 1925). The development of both aerobic and anaerobic types is favoured by the addition of 10 per cent CO<sub>2</sub> to the atmosphere. The range of temperature over which the aerobic species are able to grow is very wide. Many of the water and soil strains multiply even at 3–6° C while nearly all strains grow between 6° and 30° C. About 30° C some strains fail to grow but the majority have their optimum temperature round about 37° C. Thermophilic species are encountered with an optimum temperature between 40° and 70° C. The anaerobic strains are more fastidious in their requirements and fail to grow if the temperature varies more than a few degrees from the optimum of 37° C (Lieske 1921). Growth is improved as a rule by the addition of glucose or glycerol sometimes by blood or serum.

**Biochemical Characteristics**—The anaerobic type produces acid but no gas in glucose, maltose, lactose and sucrose; strains of human origin (see p. 39) ferment also mannitol and salicin (Erikson 1940). Fermentation is often very slow. The aerobic types have as a rule, no action on these substances. Many of the aerobic types turn litmus milk slightly alkaline and peptonize it slowly; the anaerobic type turns it acid (Negroni and Bonfiglioli 1937–38; Slack 1942). A few members are proteolytic, digesting gelatin and serum but the majority have no proteolytic power. The anaerobic type cannot grow in the presence of bile salts; some of the aerobic types are able to do so.

None of the strains that we have tested was hæmolytic but Waksman (1918) has noted hæmolysis in certain strains and has correlated this property with the power to digest proteins.

Pigment formation is characteristic of many of the aerobic species, the usual colour being some shade of pink or brown

**Antigenic Structure**—Colebrook (1921) working with three strains of the Wolff Israel type, found that their agglutination reactions differed. Aoki (1936*a, b*), using agglutination and complement fixation reactions, found that 6 anaerobic strains all fell into one group, but that 19 aerobic strains fell into eight different groups. The careful observations of Erikson (1940) have shown that by agglutination, aided when necessary by absorption of agglutinins, the anaerobic species can be divided into (1) a group containing strains of human origin which are antigenically homogeneous though differing in their degree of agglutinability, and (2) a group containing strains of bovine origin, likewise homogeneous, but with much less power of giving rise to antibodies in rabbits. It must be noted however, that all of Erikson's bovine strains were of Australian origin, whether European strains behave similarly is not known.

**Pathogenicity**—The anaerobic Wolff Israel type appears to be responsible for actinomycosis in man and cattle. The aerobic types are mostly saprophytic, but occasionally they are able to give rise to chronic granulomatous lesions in man and other animals.

For laboratory animals both types have a low pathogenicity. Inoculated in a large dose subcutaneously into rabbits they give rise to a circumscribed abscess which may persist for weeks or months, in the pus *Drusen* may often be found (Hasegawa *et al.* 1938). Intraperitoneal injection of the Wolff Israel type into guinea pigs and rabbits may give rise to small nodules containing typical granular pus, but the lesions are neither extensive nor fatal. By the repeated inoculation of massive doses intravenously Slack (1942) was able to kill rabbits in 6–10 weeks, and to demonstrate post mortem the presence of macroscopic or microscopic abscesses and focal necroses in the lungs and liver, sometimes containing granules with hyalinized clubs. The acid fast members of the aerobic group, such as *Actinomyces asteroides* are more pathogenic. Injection of this organism by the subcutaneous intravenous, or intraperitoneal route leads to a progressive fatal infection of guinea pigs and rabbits in 5 days to 4 weeks. Post mortem small tubercles are found scattered throughout the organs, being especially numerous in the lungs, liver, and spleen (Eppinger 1891, MacCallum 1902). Microscopically these nodules contain tangled filaments, sometimes arranged in the typical ray form.

**Variation in *Actinomyces bovis***—Not all the anaerobic strains isolated from actinomycotic lesions conform to the description that we have given. Lentze (1938) pointed out that in certain cases an anaerobic bacillus of more diphtheroid appearance might be found differing in its morphological, cultural biochemical and antigenic characters from the classical Wolff Israel type. He designated the new form as the S and the old as the R form. Erikson (1940) however, rightly objects to this terminology, and since the majority of the new strains are of animal origin, refers to Lentze's S and R forms by the terms bovine and human respectively. Whether these different forms are stable varieties analogous to the bovine and human types of the tubercle bacillus, must await further observation.

The chief differential characters of strains of bovine origin are as follows—The mycelium undergoes fragmentation very rapidly so that rod forms are predominant, extensive ramification is rare, growth is scanty, the colonies are smoother, softer, have an entire edge, and are not adherent to the medium, aerial hyphae are not formed, as they may occasionally be by the classical form (Erikson

1940), in liquid media there is sometimes a slight turbidity, and a wispy or light flocculent deposit, mannitol and salicin are seldom fermented, and antigenically they constitute a separate group

### CLASSIFICATION

Lieske (1921), who made an extensive study of the *Actinomyces* from the point of view of a botanist, found so much variation in the behaviour of individual strains that he was unable to arrive at any satisfactory classification. Subsequent workers, however, have felt that Lieske overestimated the difficulties of this task. Attempts at classification have been made along both morphological and physiological lines. The most prominent exponent of the former group during recent years is Orskov (1923) who devised the following scheme

**Group I** "Spores" give rise to a unicellular branching mycelium. This affords the substratum for an aerial mycelium, which consists of rather thicker branched filaments. 'Spores' are formed from the aerial hyphae without any previous segmentation of the cytoplasm. "Spore" formation commences at the tip of the thread and proceeds towards the base. "Spores" are more resistant to heat than the plain mycelia. Minor points are that the primary mycelium is of cartilaginous consistency, and often sends roots into the agar. The aerial mycelium may arise centrally or peripherally, and may not appear for a long time. Condensation of the cytoplasm occurs at regular intervals. Whole appearance of organism is fairly uniform. Gelatin is often liquefied. This group seems to include most of the non acid fast aerobic species.

**Group IIA** Both the primary and the aerial mycelium are of the same diameter, and both undergo early segmentation into irregular fragments. The aerial mycelium arises very early in the culture. It starts at the centre of the primary mycelium and spreads concentrically towards the periphery. No "spore" formation occurs, and the segmented mycelial fragments are no more resistant to heat than the unicellular mycelium. The whole appearance is very pleomorphic, coccoid, bacillary, filamentous, clubbed, and irregular shaped forms are common. The growth is generally soft in consistency, and does not adhere to the medium. A reddish insoluble pigment is frequently formed. Gelatin is not usually liquefied. This group includes many of the aerobic acid fast species, such as *Actinomyces asteroides* and *Actinomyces farcinicus*.

**Group IIB** Differs from Group IIA in forming no aerial mycelium. [Erikson (1930), however denies this, and would do away with the distinction between the A and B groups.] Besides segmentation, the so-called angular division is common in this sub-group and is responsible for the diphtheroid appearance of these organisms in film preparations. Group IIB comprises mainly the anaerobic species, of which the most important is the pathogenic organism described by Wolff and Israel.

**Group III** The characteristic feature of this group is the formation of oval 'spores' at the extreme tips of the mycelial branches. No aerial mycelium is produced. Only one species has so far been recognized—*Actinomyces clalcea*.

Nae-lund (1925) put forward a classification based primarily on physiological characteristics. We have modified it very slightly, and present it in the following form, paying attention mainly to the organisms that are parasites or potential parasites of animals.

*Actinomyces*

- A Predominantly anaerobic types *Actinomyces bovis* Harz
- B Predominantly aerobic types
  - (1) Non acid fast *Actinomyces graminis* Bostroem  
*Actinomyces caprae*  
*Actinomyces madurae*  
*Actinomyces somaliensis*
  - (2) Acid fast *Actinomyces farcinicus*  
*Actinomyces asteroides*  
*Actinomyces gypsoides*
- C Facultative aerobic types *Actinomyces muris*

The creation of a special subdivision for the facultative aerobic types is dictated, partly by convenience, and partly by the differences of the main species *Actinomyces muris* from the anaerobic Wolff Israel type

There seems little doubt that the organism *Streptobacillus moniliformis*, to which so much attention has been called in recent years by Levaditi, Nicolau and Poincloux (1925), Parker and Hudson (1926), Levaditi Selbie and Schoen (1932), Strangeways (1933), and Mackie, van Rooyen, and Gilroy (1933) is the same as the organism isolated by Schottmüller (1914), Blake (1916) and Tileston (1916), from one type of rat bite fever in human beings, and called *Streptothrix muris rattis*. Since the original name for this organism claims priority over *Streptobacillus moniliformis*, we propose to adopt it. Modification, however is necessary, partly to suit the binomial nomenclature, and partly because the term *Streptothrix* is not valid. In its place we suggest the name *Actinomyces muris*.

The inclusion, however, of this organism in the *Actinomyces* group is admittedly tentative, and may well have to be revised in the light of future work. The observations of Klenberger, of Dienes, of Hedman and others (see pp. 939-945) have revealed the existence of a group of small pleomorphic bodies resembling in many ways organisms of the pleuropneumonia group. The first of these bodies was isolated from a culture of *Streptobacillus moniliformis*. Whether it was a variant form of this organism or a symbiont remains doubtful, but since the other pleuropneumonia like bodies that have been described do not appear to have been associated with *Streptobacillus moniliformis*, it seems premature to transfer this organism to the pleuropneumonia group and call it as Hedman (1941) suggests, *Asterococcus muris*. We propose therefore, to include it temporarily at least in the *Actinomyces* genus, but to describe the pleuropneumonia like bodies associated with it in the chapter on the pleuropneumonia organism and associated foras. Van Rooyen (1936) would exclude *Streptobacillus moniliformis* from the *Actinomyces* group, because he was unable to demonstrate the presence of true branching in this organism, but his observations are contrary to those of most other observers, and the reasons he advances for assigning it to the *Hamophilus* group are not convincing. There is some ground for believing (see Dienes and Edsall 1937) that the organism isolated by Theobald Smith from the pneumonic lungs of calves and called by him *B. actinoides* is closely related to, if not identical with, *Streptobacillus moniliformis*. Since there is still some doubt about this, we shall for the moment describe it in the *Actinobacillus* group (see p. 389) to which its normal morphological appearance would naturally assign it.

Erkson (1935), who has recently studied a number of new parasitic species of *Actinomyces*, has suggested a scheme of classification containing both morphological and physiological characteristics. She accepts Orskov's grouping, but does away with the distinction between his Groups IIA and IIB. She objects to the use of oxygen requirements as a basis of classification on the ground that the distinction



between the aerobic and anaerobic species is not sufficiently sharp. Instead, she places reliance on pigment formation, and proteolytic action.

Waksman and Henrici (1943) suggest a classification based primarily on the fragmentation or not of the mycelium. Organisms in which the mycelium breaks up into bacillary or coccoid elements they would place in the family *Actinomycetaceae*, this would comprise two genera—*Actinomyces* for the anaerobic and *Nocardia* for the aerobic species. Organisms in which the mycelium does not fragment they would place in a new family *Streptomycetaceae*, this would comprise two genera—*Streptomyces* for those species in which multiplication occurs by conidia in chains from aerial hyphae, and *Micromonospora* for those species in which multiplication occurs by single terminal spores or short sporophores. It will be realized that agreement on the classification of members of the *Actinomyces* group is still far from being reached.

A detailed description of some of the more important members is appended, followed by notes on others that are of less importance. These descriptions are based in part on our own observations of relatively few strains. For a differential table see p. 392 (Table 24), and for a general description of different types see Lieske (1921), Naeslund (1925), Setti (1929), and Rosebury (1944).

#### *Actinomyces bovis* Harz

**Isolation.**—Described by Bollinger in 1877, named *Actinomyces lotis* by Harz in 1877, and first isolated by Wolff and Israel in 1891.

**Habitat.**—Strict parasite found in lesions of actinomycosis in man and cattle. Frequent in human mouth and in salivary calculi.

**Morphology.**—*Glycerol agar*, 7 days at 37° C. Long and short rods predominate, long continuous or segmented threads with a straight or curved axis, showing simple or dichotomous branching, S-shaped or spiral organisms, coccoid forms. The rods resemble, and are arranged like, certain members of the corynebacteria, sides parallel or irregular, ends rounded, clubbed or tapered, axis straight or curved, great variation in appearance, irregular staining is usual, granular and beaded forms are not uncommon. Non-motile. Non-sporing. Gram positive. Non-acid fast.

**Agar Plate.**—7 days at 37° C. anaerobically. Poor growth of round, 0.5–1.0 mm. in diameter, convex, opaque, amorphous colonies with smooth dull surface and entire edge, greyish-white by transmitted, porcelain white by reflected light, butyrous or friable consistency, emulsifiability not difficult as a rule. 21 days, rather larger, 1–1.5 mm. in diameter, umbonate, with slightly irregular nodular surface and lobate edge, differentiated into a glistening raised centre and a dull shelving periphery resembling a rosette. Colonies may grow into medium.

**Agar Slope.**—7 days at 37° C. anaerobically. Moderate growth of discrete colonies similar to those described. Numerous greyish-white floccular masses of coarsely granular structure in water of condensation, they are irregular in shape, have an irregular edge, and are opaque. In the condensation water there is also a finely granular turbidity.

**Gelatin Slab.**—No growth at 23° C. After 12 days at 37° C., the culture shows, when cooled, a band of growth 4 mm. deep with its upper margin 1 mm. below the surface. Growth consists of very fine greyish-white interlacing filaments, looking like cotton wool. No liquefaction.

**Broth.**—5 days at 37° C. anaerobically. Poor to moderate growth, deposit of compact, white, mulberry like granules with nodular surface, often adherent to each other, not disintegrated on shaking. No turbidity, no surface growth, no odour.

**Glucose Agar Shake.**—5 days at 37° C No growth for 1 cm below surface. Then comes a turbid band, about 0.8 mm deep, consisting of large numbers of tiny colonies. Throughout the rest of the medium are scattered discrete, irregularly round, opaque, greyish white colonies, about 0.1–1.0 mm in diameter, with smooth or slightly knobby surface.

**Loeffler's Serum.**—7 days at 37° C anaerobically Moderate, partly confluent, raised, shiny growth of low convex, rounded colonies about 0.5 mm. in diameter No liquefaction.

**Glycerol Egg**—14 days at 37° C anaerobically Poor, slightly raised, confluent growth with finely granular surface due to imperfect fusion of colonies No liquefaction.

**MacConkey**—No growth in either solid or liquid medium.

**Potato**—14 days at 37° C anaerobically Very poor growth of discrete, round, 1 mm in diameter, whitish low convex colonies with smooth glistening surface and entire edge.

**Resistance.**—At 37° C cultures live for about 1 to 4 weeks, sometimes longer Dried on glass and kept in the dark organisms may live for 7 weeks or more. Killed by moist heat at 60° C. in 15 minutes.

**Metabolism**—Anaerobe of the microaerophilic type Will not grow on surface culture exposed to the air Optimum temperature for growth 37° C, growth below 30° C is either very slight or absent Optimum pH 7.3–7.6 No hæmolytic of horse red cells No pigment formation. Growth is improved by nitrates, glycerol, blood an increased partial pressure of CO<sub>2</sub> and sometimes by glucose.

**Biochemical**—Acid, no gas, in glucose, maltose, mannitol, lactose, sucrose, and salicin within 21 days under anaerobic conditions LM acid Indole —, MR —, VP —, Nitrate reduction + +, H<sub>2</sub>S —, NH<sub>3</sub> +, MB reduction —, Catalase —.

**Antigenic structure**—Two groups distinguishable by agglutination, one containing strains of human, the other of bovine origin.

**Pathogenicity**—Responsible for actinomycosis in man and cattle Very slight pathogenicity for laboratory animals Intrapertoneal inoculation of a broth culture into a rabbit or guinea pig may be followed by appearance of small nodules chiefly in the great omentum, containing the typical clubbed colonies of *Actinomyces*. The animals live indefinitely.

#### *Actinomyces maduræ*

**Isolation.**—Isolated from pale variety of Madura foot by Vincent in 1894 Called by him *Streptothrix maduræ*.

**Habitat.**—Found in pale variety of Madura foot Saprophytic existence probable, but not demonstrated.

**Morphology**—Glycerol agar, 14 days at 37° C Long, non segmented filaments 0.4–0.6  $\mu$  thick, showing true and false branching, sides parallel, ends often tapering. Arranged in a mycelium sometimes aggregated into dense masses. Stain evenly. Later fragmentation may occur with production of ovoid bodies. Non motile. Gram positive. Non acid fast.

**Agar Plate.**—5 days at 37° C Small, round, convex colonies about 0.5 mm in diameter 14 days, larger, 1–3 mm. in diameter, greyish yellow, opaque, irregularly heaped up, nodular, umbonate colonies, resembling worm casts which have a smooth glistening surface. Very adherent to medium; consistency horny, very difficult to emulsify. Whole colony looks like a rosette.

**Agar Slope.**—7 days at 37° C Poor growth of discrete, dull, greyish white opaque irregularly heaped up colonies with nodular surface.

**Gelatin Slab**—14 days at 20° C Moderate, filiform growth consisting mostly of small, discrete, greyish white colonies, having a darker centre and a lighter feathery

periphery, growth extends to bottom of tube. Slightly raised surface growth about 3 mm. in diameter. Slight liquefaction after 6 weeks.

*Broth*—7 days at 37° C. Poor growth. Deposit of little greyish white puff balls, looking like colonies of moulds, and having a dense centre and a lighter periphery, often cohering in groups of two or three. No turbidity, no surface growth; no odour. Later a white efflorescent surface growth may appear.

*Loeffler's Serum*.—7 days at 37° C. Poor growth of isolated colonies. 21 days, moderate growth of heaped up nodular colonies. No liquefaction.

*Glucose Agar Slope*.—7 days at 37° C. Discrete colonies raised, heaped up with worm cast surface, moist and glistening.

*Glycerol Agar Slope*.—12 days at 37° C. Luxuriant raised, confluent, greyish white worm cast growth, very tough, adherent to medium, and difficult to emulsify.

*Glycerol Egg*.—7 days at 37° C. Mostly confluent growth of rounded, dome-like colonies with dull nodular surface. Very tough adherent to medium, and difficult to emulsify. No liquefaction.

*Potato*.—7 days at 37° C. Discrete heaped up, nodular, yellowish brown colonies. 18 days heaped up dry, chalky white and greyish brown, worm-cast colonies. Later may take on a rose-red colour.

*MacConkey*.—No growth on solid or liquid media.

*Resistance*.—Destroyed by moist heat at 60° C. in 5 minutes.

*Metabolism*.—Aerobic, very slight growth on glycerol agar anaerobically. Optimum temperature 37° C. grows at 20° C. Forms sometimes a rose-red pigment on potato. Growth improved by glycerol and glucose.

*Biochemical*.—Ferments no sugars. L.M. turned slightly alkaline, may be peptonized. Indole—M.R.—, V.P.—, Nitrate reduction +, H<sub>2</sub>S—, NH<sub>3</sub> sl. +, Catalase v. sl. +, M.B. reduction—.

*Pathogenicity*.—Subcutaneous inoculation into rabbits, guinea pigs, mice, and cats causes a local nodule, which increases in size for a month and then retrogresses. Responsible for pale or ochroid variety of Madura disease in man.

#### *Actinomyces graminis* Bostroem

*Isolation*.—Isolated by Bostroem in 1891 from human actinomycosis. Subsequently isolated by numerous workers from different lesions in man and other animals.

*Habitat*.—Saprophyte living on grains and grasses. Often gains access to mouth and respiratory passages of man and other animals.

*Morphology*.—Agar 24 hours at 37° C. Long filaments, 0.6  $\mu$  wide, showing true and false branching, long and short rods, and coccoid bodies. Sides parallel, ends rounded or tapering. Filaments are straight, wavy, or spirillar, and may or may not be segmented. Arranged in small loose groups, or in a mycelium. In broth ray forms are frequent, with compact centre and radiating filaments. Staining of rods and filaments is often irregular. Non-motile. Gram positive. Non acid fast.

*Agar Plate*.—24 hours at 37° C. Round, greyish white, low convex, dull, opaque colonies with finely granular surface and erose edge. 7 days, rounded, up to 3 mm. in diameter, umbonate, granular colonies, with raised, opaque, primrose-yellow, radially striated centre, and effuse, greyish ground-glass periphery. Surface granular, edge feathery or rhizoid. Generally tough and adherent to agar, and difficult to emulsify. Colonies may be folded on surface and coral-like or they may be undifferentiated with a nodular surface and lobate edge. Colour may be chalky white, yellow or brown.

*Agar Slope*.—24 hours at 37° C. Abundant, slightly raised, greyish-white, faintly translucent or opaque growth with dull, finely granular or mealy surface and entire or erose edge. 7 days, surface is whitish and moderately granular, growth may be heaped up in places.

- Gelatin Slab**—7 days at 20° C Moderate filiform growth of confluent, greyish white, feathery colonies, extending to bottom of tube Slightly raised surface growth, 3 mm in diameter After 7 weeks the growth near the surface is orange pink Liquefaction unusual.
- Broth**—24 hours at 37° C Moderate growth, ropy or membranous sediment, not disintegrating on shaking, ring growth and finely granular almost invisible surface pellicle, turbidity absent, or slight and finely granular 7 days, thick surface pellicle, extending up sides of tube, dry, dull, and scaly with pinkish yellow or orange nodules in places, heavy floccular deposit, pinkish in colour No odour
- Glucose Agar Shake**—8 days at 37° C Good growth confined to surface except for a few tiny colonies in upper 5 mm. of medium Surface growth is thick, raised, confluent, dull, greyish white with several secondary colonies developing on it Sometimes surface growth is heaped up, yellowish brown, and of worm-cast type with no colonies below surface 21 days, growth is brick red or yellowish-orange in colour
- Loeffler's Serum**—24 hours at 37° C Good, raised, moist, glistening confluent growth with nodular surface and lobate edge 24 days, no liquefaction.
- Dorset Egg**—10 days at 37° C Good, confluent, raised, yellowish growth with nodular surface and edge formed of single colonies. 21 days no liquefaction
- MacConley's Agar**—6 days at 37° C Growth of small 0.1 mm in diameter pinkish opaque colonies; growth very poor compared to that on agar In liquid medium there is good growth with a heavy granular deposit
- Potato**—24 hours at 37° C Poor, slightly raised chalky white growth with powdery surface. Later, growth may turn yellowish-orange or ochre brown
- Persistence**—Cultures remain viable for months.
- Metabolism**—Aerobic. No growth under strict anaerobic conditions. Optimum temperature 37° C, grows at 20° C. No haemolysin for horse red cells. Yellowish orange or pink pigment formed, particularly in old cultures stood at room temperature Growth is improved by glucose, sometimes by serum
- Biochemical**—No fermentation of sugars. Litmus milk turned slightly alkaline in 6 days, may be slowly peptonized Indole—MR—, VP— Nitrate reduction +, NH<sub>3</sub> +, H<sub>2</sub>S very slight +, Catalase + MB reduction—
- Pathogenicity**—Appears to be non pathogenic to laboratory animals

### Actinomyces muris

**Synonyms**—*Streptothrix muris ratti* Schottmüller, *Streptobacillus moniliformis* Levaditi.

**Isolation**—Isolated by Schottmüller (1914) from human patients bitten by rats

**Habitat**—Natural parasite inhabiting the nasopharynx of rats (Strange ways 1933)

**Morphology**—*Loeffler's serum* at 37° C Slender branching filaments, 0.4–0.6  $\mu$  wide, growing in interwoven masses After 18–24 hours fragmentation of the filaments sets in, and many of the filaments are replaced by chains of bacillary or coccoid bodies Very marked pleomorphism Occasional filaments show spherical, oval, fusiform



FIG 64—*Actinomyces muris* From a Loeffler slope, 2 days 37° C aerobically ( $\times 1000$ )

form or club shaped swellings occurring terminally sub-terminally, or in some other situation—hence the term *moniliformis*. These swellings may be 2-5 times the diameter of the filament and may project from one side only. According to Kluenerger (1912) the moniliform appearance and certain other appearances are due to the pleuropneumonia or LI organism which is a constant companion to *Actinomyces muris*. In the animal body the morphology is more regular and lacy. Great irregularity in depth of staining. Non motile. Usually described as Gram negative but may be Gram positive in young cultures. Non-acid fast. (See Fig 64)

*Nutrient Agar at 37° C*—No growth

*Glucose Agar at 37° C*—No growth

*Serum Agar Plate*—2 days at 37° C Circular greyish yellow, low convex almost watery, amorphous colonies 0.2-0.3 mm in diameter with smooth glistening surface and entire edge, butyrous in consistency and easily emulsifiable. No differentiation. Little or no increase in size on further incubation

*Gelatin Stab*—7 days at 20° C No growth

*Nutrient Broth at 37° C* No growth

*Serum Broth*—2 days at 37° C No turbidity. Abundant greyish white coarsely granular sediment, looking like fluffy bread crumbs, miniature cotton balls, or tiny snow flakes not disintegrating completely on shaking. No surface growth. No odour

*Glucose Agar Shake*—7 days at 37° C No growth.

*Loeffler's Serum*—2 days at 37° C Discrete circular, low convex colonies, similar to those on serum agar but rather larger—0.5-0.7 mm in diameter. 7 days some colonies may show a differentiation into a slightly raised umbonate centre with a flatter periphery having an irregular or crenated edge, surface appears finely granular and rather dull. Growth may be confluent from the start, and appear slightly raised colourless with a glistening beaten copper surface and a more or less entire edge. No liquefaction even after 3 weeks.

*Dorsal Egg*—2 days at 37° C Similar to colonies on Loeffler's serum but perhaps slightly smaller—0.3-0.6 mm in diameter. No liquefaction even after 3 weeks.

*Horse Blood Agar*—2 days 37° C Colonies resemble those on serum agar. No haemolysis.

*Potato*—7 days at 37° C No growth

*MacConkey's Agar*—7 days at 37° C No growth

*Growth in Developing Egg*—Inoculated on to the chorio-allantoic membrane of the developing chick embryo, *Actino. muris* invades the embryo and becomes localized almost exclusively in the synovial lining of the joints, where it appears to grow mainly as an intracellular parasite, the embryo dies within 4 days (Buddingh 1944).

*Persistence*—Destroyed in serum broth by heating to 55° C. for 30 minutes. Dies out in culture very readily. Serum broth cultures may remain viable at 37° C. for a week.

*Metabolism*.—Grows aerobically, but grows equally well or better under anaerobic conditions. Growth said to be improved by 10 per cent. CO<sub>2</sub>. Optimum temperature 37° C. Little or no growth at 22° C. No haemolysin for horse red cells. No pigment formation. No growth on ordinary media, but growth occurs in presence of serum, ascitic fluid or blood, not improved by glucose or glycerol.

*Biochemical*—Not thoroughly studied. In serum sugar media acid is produced within 3 days in glucose and salicin, sometimes in maltose and lactose. Litmus milk unchanged. Indole—, M.R.—, V.P.— Nitrate reduction—, H<sub>2</sub>S— catalase—, M.B. reduction—

—Different strains appear to be antigenically homogeneous.

able in man for one type of rat bite fever—sometimes described as Haverhill fever. May give rise to an epizootic disease by oedematous swelling of the feet and legs, arthritis, oophadenitis. Intraperitoneal inoculation of mice with 0.5 ml. ure is usually fatal in 1-2 days, no characteristic post mortem

appearances visible. Subcutaneous inoculation into one of the hind feet often leads to a more or less perfect reproduction of the natural disease. Comparatively avirulent for rats, guinea pigs and rabbits though intravenous inoculation of culture into rabbits may sometimes lead to arthritis.

*Variation*—Phuro-neumonia like bodies are constantly associated with this organism (see p. 346), it is still doubtful whether they are variant forms or symbionts.

*Note*—Dick and Tunnichiff (1918) isolated a similar organism—*Actinomyces putorii*—from a boy bitten by a weasel.

## A Other Anaerobic Types

Tunnichiff (1926) describes a weakly Gram positive motile anaerobic organism which she isolated from a tonsillar granule. Smear preparations of the granule showed thick bacilli with rounded ends, filaments, tightly waved spirilla and cocci. Sections stained with Giemsa showed bundles of filaments with the bacillary forms radiating from them. In anaerobic culture on ascitic fluid tissue medium and sheep-blood agar rosettes and test tube-brush like forms appeared, similar to those in the original material. It is very doubtful whether this organism should be included in the *Actinomyces* group (see also Tunnichiff and Jackson 1930).

## B Other Aerobic Types

### (1) NON ACID-FAST TYPES

*Actinomyces capre*—Described by Silberschmidt (1899) in 1897. It was isolated from the lung of a goat supposed to be suffering from tuberculosis. Consists of very thin wavy filaments, showing a varying degree of branching. Filaments segment into rod and coccoid forms. On agar dry colonies, flattened in the centre with an irregular warty folded surface. In both surface growth of dry discoid colonies and a rough deposit. No change in litmus milk. Abundant growth on potato of rose-red colonies, later becoming chalky white. No liquefaction of gelatin. Aerobic. Subcutaneous injection into rabbits produces an abscess. Intravenous injection sometimes causes tubercles in various organs. Guinea pigs are rather more susceptible than rabbits (see also Galli Valerio 1912).

*Actinomyces smallensis*—Described by Brumpt in 1906 (see Brumpt 1927). Was first isolated by Bouffard in French Somaliland from patients affected with mycetoma. Consists of long branching filaments with truncate or sometimes tapering ends. Gram positive. Grows on agar, but better on blood agar. On this medium colonies are at first small, circular, convex and translucent but after a few days they become irregularly heaped up, nodular, worm cast or crateriform, they are opaque, vary in colour from white through yellowish-orange, to brown or black, often show radial segmentation which gives them a stellate appearance, are extremely tough in consistency and adherent to the medium and have a peculiar odour. In broth no turbidity or surface growth but a deposit of little greyish white puff balls. On potato a white folded layer of growth which in 5 to 6 days becomes yellow. Peptonizes milk. Ferments no sugars. Gives rise in man to mycetoma of the hand or foot. Lesions contain hard smooth yellowish red granules 1 mm in diameter not dissociated by caustic potash.

### (2) ACID FAST TYPES

*Actinomyces farcinieus*—Isolated by Nocard in 1888 from cattle suffering from farcy. Branching filaments growing in a mycelium. Gram positive, feebly acid fast (see p. 376). On agar it forms small irregular, raised opaque yellowish white colonies with a dull mammillated powdery surface. Dry, scaly, pale yellow plaques on potato. Irregular whitish masses in broth, some of which remain at the surface and others fall to the bottom. No liquefaction of gelatin. No change in litmus milk. No growth anaerobically. In cultures, the organism forms filamentous felted masses and diphtheroid like bacilli. Cultures remain viable for 4 months at 37° C, killed by heat at 70° C in 10 minutes. Intra-

peritoneal injection is fatal to guinea pigs in 9 to 20 days, post mortem, milary nodules over peritoneum, containing a little pus, in the pus are masses of bacilli. Intravenous injection of guinea pigs causes the formation of generalized milary nodules, particularly abundant in the lungs, liver, and spleen. Milary nodules follow intravenous injection of cows and sheep. Rabbits, dogs, cats, horses, and asses are resistant to intravenous or intraperitoneal injection. Subcutaneous injection causes a slowly progressive abscess, which ulcerates and heals.

*Actinomyces asteroides*.—Isolated by Eppinger in 1891 from a brain abscess in a glass-grinder. Consists of threads showing true and false branching, threads may be long, or short and segmented, tiny rod forms also seen. In the body it forms long granular, interlacing filaments with no ray or club formation. Gram positive. Acid fast, though not so strongly as the tubercle bacillus. Aerobic, no growth anaerobically. Destroyed by heat at 70° C. in 5 minutes. On agar—at first whitish, later ochre-coloured, umbonate colonies, having a raised, dry, wrinkled, opaque centre and a moist glistening more translucent periphery with a myceloid edge, whole colony star-shaped—hence the name *asteroides*—may be a central depressed crater, later, colour deepens to orange, and the wrinkling of the surface becomes more marked. Agar slope—raised ochre growth with a mealy surface and entire edge. Gelatin—very slow growth without liquefaction. Potato—red raised growth with a granular, and later wrinkled, surface, a chalk white bloom may develop due to a velvet like upgrowth of fine filaments into the air. These uplifted filaments have terminal chains of coccoid bodies or spores, which, when transferred to broth, sprout and give rise to long filaments or star like clusters (MacCallum 1902). Broth—white surface pellicle, which falls to the bottom, and is renewed several times, no turbidity. After subcutaneous, intraperitoneal, or intravenous injection, rabbits and guinea pigs die in 5 days to 4 weeks. Post mortem, the viscera, especially lungs, liver, and spleen, are studded with small white nodules. Abscesses may develop in the muscles, kidneys, and other organs—these abscesses contain branching test tube brush forms with laterally radiating clubs. An organism called *Actinomyces variabilis* with similar pathogenicity but different cultural reactions to Eppinger's strain was isolated by Cohn (1913) from the bladder of a man with pyuria.

*Actinomyces gypsoides*.—Isolated by Henrici and Gardner (1921) from the sputum of a woman. Acid fast branching filaments were found in the sputum, sometimes in mycelial form. Agar slope—thin greyish veil, soon becoming thick, opaque, and chalky white, surface dry and wrinkled, growth finely adherent to the medium and very brittle. Potato—growth similar to that on agar. Gelatin stab—surface growth only, liquefaction stratiform and complete in a week. Broth—small white flakes coalescing to form a thick, wrinkled, snow white surface pellicle extending up sides of tube. Litmus milk—yellowish surface pellicle, milk is turned alkaline and curdled, litmus reduced, later digestion. Media containing peptone are darkened (tyrosinase). Growth improved by dextrose, maltose, and glycerol. No carbohydrates fermented. Intravenous injection into rabbits is fatal in 2 days post mortem, minute abscesses in viscera, especially kidneys, which are studded with yellowish white nodules. Intraperitoneal injection into guinea pigs is fatal in 4 to 6 days, post mortem, small tubercle-like nodules over peritoneum, omentum shrunken and studded with nodules.

An acid fast strain described by Birt and Leishman (1902) gave a snow white growth on solid media, peptonized milk, but did not liquefy gelatin. Another acid fast strain described by Berestnew (see Feistmantel 1902) gave a grey to whitish growth, liquefied gelatin, but was non pathogenic to laboratory animals. More recently, Goldsworthy (1937) has given a good description of an acid fast strain isolated from a patient with pulmonary actinomycosis.

### C Other Facultative aerobic Types

Described by Naeslund (1925), who isolated organisms of two different types from the human mouth.

Type I consists morphologically of branching and often very sinuous relatively short threads usually arranged in a radiating fashion, rods and granules are also present. In old cultures there is marked pleomorphism. No aerial spores. Chiefly Gram positive, non acid fast. Culturally, growth in dextrose broth occurs in the form of round oval or ovoid colonies, white to yellowish grey in colour which appear at the bottom in about a week. On saliva dextrose agar a slightly shiny colourless film appears becoming granular after a few days, some of the granules may develop into small greyish white nodular adherent colonies, having a narrow translucent finely striated margin. Little or no growth on gelatin, potato or milk. Optimum temperature for growth  $37^{\circ}\text{C}$ , no definite growth at  $20^{\circ}\text{C}$ .

Type II consists of very long threads of fairly even thickness showing typical but infrequent branching. No definite aerial spores formed. Gram positive but the greater part of the mycelium consists of Gram negative elements with occasional Gram positive segments. Non acid fast. Culturally, in saliva dextrose broth round greyish white more or less translucent colonies develop at the bottom in about a couple of weeks. On saliva glucose agar growth at first occurs in a thin film but in 1 to 2 weeks isolated pinhead colonies appear hard or soft in consistence and surrounded by a narrow translucent border. No definite growth on gelatin, potato or in milk.

### ACTINOBACILLUS

Lignières and Spitz (1902) isolated a non motile non branching Gram negative bacillus from the lesions of cattle suffering from a disease which in many respects resembled actinomycosis. They called the organism the *actinobacillus*, and the disease to which it gave rise actinobacillosis. Two other organisms have since been described, having some points of similarity with this bacillus and it is therefore convenient to consider them as forming a group to which the generic name *Actinobacillus* may be applied.

**DEFINITION** *Actinobacillus* Brumpt (Emended from the American Committee's Report)

Gram negative, non acid fast rods, sometimes occurring in long chains or in unjointed filaments. In lesions in the animal body no mycelium is formed, but at the periphery finger-shaped cells or clubs may be visible.

Type species is *Actinobacillus lignieresii*, Brumpt

The classification we suggest is as follows

#### *Actinobacillus*

A Aerobic and facultatively anaerobic *Actinobacillus lignieresii*  
*Actinobacillus actinomycetem-*  
*comitans*

B Preferring raised  $\text{CO}_2$  pressure *Actinobacillus actinoides*

A description follows of the separate organisms

#### *Actinobacillus lignieresii*

For isolation see above. Appears to be a strict parasite

**Synonym**—Probably the same as *Bact. purifaciens* (see Tunncliffe 1941)

**Morphology**—In young cultures it is a small rod-shaped organism, in older cultures it is coccobacillary, and various involution forms appear. In serum broth long streptobacillary forms are common. In glucose agar shake cultures long tangled unbranched filaments may be formed accompanied by smaller bacilli and coccoid bodies (Griffith 1916). Dimensions of the bacilli are given by Lignières and Spitz (1902) as  $1.15-1.25\ \mu$  long by  $0.4\ \mu$  broad. Non motile non sporing non acid fast. Stains readily especially with carbol fuchsin and is Gram negative frequently shows bipolar staining.



In lesions in the animal body small granules are found which consist of tufts of radially disposed clubs similar to those in actinomycosis. An important point of difference is that the centre of the granule is occupied not by a Gram positive filamentous mycelium such as is formed by *Actinomyces bovis* but by minute Gram negative bacilli which may quite readily be overlooked. Though both the bacilli and the clubs formed by *Actinobacillus lignieresii* are Gram negative it is possible to differentiate between them by a modified Ziehl-Neelsen stain, as was pointed out by Bosworth (1923). If a section of affected tissue is stained with carbol fuchsin decolorized for 90 to 30 seconds with 1 per cent  $H_2SO_4$  and counterstained with methylene blue the clubs appear red and the bacilli blue. For pus one of the best stains is glycerine picro-carmin which stains the clubs yellow and the pus cells pink.

**Cultivation**—Cultures are best obtained by grinding up infective pus in a mortar and seeding on to agar. Growth occurs readily under aerobic conditions and less readily under anaerobic conditions. The optimum temperature for growth is  $37^\circ C$ . very slight growth occurs at  $20^\circ C$ .

On agar in 24 hours at  $37^\circ C$  small circular bluish grey translucent colonies with a smooth surface and an entire edge up to 1.5 mm in diameter, are formed. Further incubation results in a considerable increase in size—up to 4 mm.—due to peripheral extension of the colony. On an agar slope the growth of freshly isolated strains is poor consisting of small, discrete translucent bluish colonies or of a thin, dry confluent layer of growth adherent to the medium. After cultivation for some time in the laboratory the organism grows more readily giving a confluent, filiform viscous growth with a thickened edge.

In stab agar there is a whitish opaque spot at the surface, no growth occurs down the stab. In gelatin stab growth is very poor and is not visible for some days. A small opaque spot appears at the surface, no growth occurs down the stab and there is no liquefaction.

FIG. 65—*Actinobacillus lignieresii*. From a liver agar slope 2 days  $37^\circ C$  aerobically ( $\times 1000$ ).

Coagulated serum is not a very favourable medium only a thin whitish growth is formed.

On acid potato there is no growth. On alkaline potato a slight glistening greyish yellow growth appears.

In peptone broth there is a slight uniform turbidity. In old cultures a surface film may develop and an abundant deposit. Growth in broth is improved by serum.

**Resistance** The organism is killed by heating to  $60^\circ C$  in 10 minutes. It rapidly succumbs to drying. Cultures do not live long and should be transplanted every few days. Infected pus preserved in sealed tubes may remain virulent for a month or two.

**Biochemical Reactions**—The fermentative ability of this organism is a little doubtful. Glucose, maltose, mannitol and sucrose are generally rendered acid though not markedly so in a day while lactose may be fermented later. Acid is produced in litmus milk, but no clot. Indole is formed apparently in small quantity.

**Antigenic Structure** Little known. Tunnichoff (1941) states that bovine and ovine strains appear to be similar but that strains of either type may fail to agglutinate with a type serum.

**Pathogenicity**—No exotoxin is formed. The organism is responsible for Actinobacillosis in cattle. The virulence of different strains seems to vary considerably and while cattle inoculation experiments are successful with some strains they are completely negative with others (Magnusson 1928). Most workers including ourselves have been unable to produce any specific lesions in laboratory animals. The following statements therefore which are taken from Lignières and Spitz (1902) must be accepted with considerable reserve. Subcutaneous inoculation of pure cultures into cattle produces an abscess identical with those occurring spontaneously in the pus granules are found consisting of bacilli surrounded by clubs. Intra-peritoneal inoculation of a whole agar culture is fatal to a guinea pig in 12 to 24 hours. Post mortem there is an abundant turbid peritoneal exudate rich in polymorphonuclear cells; the organisms can be cultivated from the exudate but rarely from the blood. Intrapertoneal injection of  $\frac{1}{2}$  an agar culture into male guinea pigs produces a typical Straus reaction. In 2 days the testicles are markedly inflamed and the two layers of the tunica vaginalis are adherent; the scrotum is red, swollen and tender. The animal loses weight and dies in 5 to 7 days. Post mortem small purulent granules the size of a hemp seed formed of a very thin membrane containing white or yellowish thick homogeneous pus and scattered over the peritoneal serosa—particularly on the inferior surface of the diaphragm, the liver, spleen and omentum. Around the testis there is a thick purulent exudate gumming the two layers of the tunica vaginalis together. In the pus of these lesions tufts of clubs are found though not in large numbers; they are rather smaller than those seen in cattle. Subcutaneous inoculation of guinea pigs causes a local abscess which may resolve; ulceration rarely occurs; clubs are not usually demonstrable in the pus.

Rabbits, cats, and dogs are considerably more resistant than guinea pigs but succumb to intravenous inoculation. Small lesions may develop in mice or rats after subcutaneous inoculation. Pigeons and fowls are resistant.

**Actinobacillus actinomycetem comitans**—Described by Hinger in 1912 under the name *Bact. actinomycetem comitans*. Found in lesions caused by *Actinomyces bovis* as densely packed Gram negative coccobacilli (Colebrook 1930). In culture the rod forms are 1.0–1.5  $\mu$  long; the coccoid forms are 0.6–0.8  $\mu$  in diameter. Intermediate forms are frequent. The organism is non-motile. In broth or liquid gelatin at 37°C it forms isolated, translucent granules 0.5–1.0 mm. in diameter along the sides of the tube; most numerous near the surface; several hundreds of these colonies may develop. After some days they fuse into a greyish white mass forming a ring round the tube and a pellicle over the surface. The granules can be picked off the wall of the test tube with a loop but are very difficult to break up. Later they may become opaque and greyish white. On agar it gives rise to small tough colonies not unlike those of streptococci, adherent to the medium. The organism flourishes under both aerobic and anaerobic conditions. There is no growth at room temperature. Cultures live for 4 weeks. It is toxic on injection into rabbits but does not set up a true infection.

**Actinobacillus actinoides**—This organism was isolated by Smith in 1918 from the lungs of calves suffering from epizootic pneumonia and called by him *B. actinoides*. In the animal body it appears as a minute Gram negative bacillus arranged in groups. In the condensation water of coagulated serum it forms minute whitish flocculi which consist of a central mass of radiating non-branching filaments ending peripherally in clubs. In tissue agar cultures the organism grows as aggregations of rounded ring-like bodies 2  $\mu$  in diameter having a minute refringent speck on the periphery or near the centre. There are thus three distinct forms in which this bacillus occurs. Most strains are capsulated (Smith 1921b) but the capsule does not stain with the usual dyes. Growth occurs only under a raised pressure of CO<sub>2</sub>—as in sealed tubes. On coagulated serum whitish flocculi appear in the condensation water in 3 days at 37°C and after several

weeks very tiny, elevated, pointed like colonies may appear on the slant. Growth may be obtained on agar to which a piece of guinea pig's spleen has been added. No growth on ordinary media or on ascitic fluid. Non pathogenic for laboratory animals on experimental injection. Subcutaneous injection into calves causes a large necrotic swelling with caseous contents, ulceration occurs in 4 weeks. Intratracheal injection into calves causes small necrotic foci in the lungs, identical with those observed in the natural disease (Smith 1921a, see also Jones 1922). The fact that it has been isolated from the lungs of white rats suffering from pneumonia (Jones 1922) suggests that it may be a natural parasite of these animals. For further references to this organism see Smith (1921a b). A similar organism, differing only in minor particulars has been isolated from the middle ear of white rats, in which it was causing suppurative (Nelson 1930, 1931). The resemblance of *Actinobacillus actinoides* to *Streptobacillus moniliformis* (see p. 381) has been noted by Dienes and Fildall (1937), but in view of its reported lack of pathogenicity for laboratory animals and its bacillary morphology in the lungs of calves, it is probably wiser to treat it separately for the moment.

## REFERENCES

- AOKI, M. (1936a) *Z. Immunforsch.* **87**, 100. (1936b) *Ibid.* **87**, 200.  
 BIRT, C. and LEITCHMAN, W. B. (1902) *J. Hyg., Camb.* **2**, 120.  
 BLAKE, F. G. (1916) *J. exp. Med.* **23**, 39.  
 BOLLINGER, O. (1877) *Zbl. med. Wiss.* **15**, 481.  
 BOSTROEM, F. (1891) *Beitr. path. Anat.* **9**, 1.  
 BOWWORTH, T. J. (1923) *J. comp. Path.* **36**, 1.  
 BRUMPT, E. (1927) "Précis de Parasitologie" Masson et Cie, Paris p. 1201.  
 BRUNS, H. (1899) *Zbl. Bakt.* **26**, 11.  
 BUDDINGH, G. J. (1944) *J. exp. Med.* **80**, 59.  
 COHN, T. (1913) *Zbl. Bakt.* **70**, 290.  
 COLEBROOK, L. (1920) *Brit. J. exp. Path.* **1**, 197. (1921) *Lancet* **i**, 893.  
 DICK, G. F. and TUNNICLIFFE, R. (1918) *J. infect. Dis.* **23**, 183.  
 DIENES, L. and EDWALL, G. (1937) *Proc. Soc. exp. Biol., N.Y.* **38**, 740.  
 EFFINGER, H. (1891) *Beitr. path. Anat.* **9**, 287.  
 ERIKSON, D. (1935) *Spec. Rep. Ser. med. Res. Coun., Lond.* No. 203. (1940) *Ibid.* No. 240.  
 FEISTMANTEL, C. (1902) *Zbl. Bakt.* **31**, 433.  
 FOULETTON, A. G. R. (1910) *Lancet*, **i**, 551, 626, 769.  
 GALLI VALERIO, B. (1912) *Zbl. Bakt.* **63**, 555.  
 GOLDSWORTHY, N. L. (1937) *J. Path. Bact.* **45**, 17.  
 GOYAL, R. K. (1930) "Contribution à l'étude des Streptothricées" Jouve & Cie Paris.  
 GRIFFITH, F. (1916) *J. Hyg., Camb.* **15**, 190.  
 HASSEGAWA, S., NAKAMOTO, T., MIYASAKI, Y., ARIMITSU, T., and AKIYOSHI, M. (1938) *Jap. J. med. Sci.* **V**, 3, 27.  
 HEILMAN, F. R. (1941) *J. infect. Dis.* **69**, 32.  
 HENRICI, A. T. and GARDNER, E. L. (1921) *J. infect. Dis.* **28**, 232.  
 JONES, F. S. (1922) *J. exp. Med.* **35**, 361.  
 KLIFFEBERGER, EMMY (1942) *J. Hyg., Camb.* **42**, 485.  
 KLINGER, R. (1912) *Zbl. Bakt.* **62**, 101.  
 LENTZE, F. A. (1938) *Zbl. Bakt.* **141**, 21.  
 LEVADITI, C., NICOLAU, S., and POINCELOUX, P. (1925) *C. R. Acad. Sci.* **180**, 1188.  
 LEVADITI, C., SELBIE, R. F., and SCHÖNEN, R. (1932) *Ann. Inst. Pasteur* **48**, 308.  
 LIESKE, R. (1921) "Morphologie und Biologie der Strahlenpilze" (Actinomyceten) Gebrüder Borntraeger, Leipzig.  
 LIGNIÈRES, J. and SEITZ, G. (1902) *Bull. Soc. cent. Med. Vét.* **20**, 487, 546. (1903) *Arch. Parasit.* **Paris** **7**, 428.  
 MACCALLUM, W. G. (1902) *Zbl. Bakt.* **31**, 520.  
 MACKIE, T. J., ROOYEN, C. F. VAN, and GILROY, E. (1933) *Brit. J. exp. Path.* **14**, 132.  
 MAGNUSSON, H. (1928) *Acta path. Microbiol. scand.* **5**, 170.  
 NÄSLUND, C. (1925) *Acta path. Microbiol. scand.* **2**, 110.  
 NEGROVI, P. and BONFIGLIOLI, H. (1937-38) *Folia biol.* No. 82 p. 351.  
 NELSON, J. B. (1930) *J. infect. Dis.* **46**, 61. (1931) *J. Bact.* **21**, 187.  
 NOCARD, C. (1888) *Ann. Inst. Pasteur* **2**, 293.

- ØRSKOV, J (1923) "Investigations into the Morphology of the Ray Fungi." Levin and Munksgaard, Copenhagen
- PARKER, F and HUDSON \ P (1926) *Amer J Path.*, 2, 357
- Report (1917) *J Bact.*, 2, 505, (1920) *Ibid.*, 5, 191
- ROOYEN, C E. VAN (1936) *J Path Bact.*, 43, 455
- ROSEBURY, T (1944) *Bact Rev.*, 8, 189
- SCHOTTMULLER, H. (1914) *Derm. Wochs.*, 58, Supp., p. 77
- SETTI, C (1929) *G Batt Immun.*, 4, 585
- SILBERSCHMIDT (1899) *Ann Inst Pasteur*, 8, 841
- SLACK, J (1942) *J Bact.*, 43, 193.
- SMITH, T (1918) *J exp Med.*, 28, 333, (1921a) *Ibid.*, 33, 441, (1921b) *Ibid.*, 34, 593
- STRANGEWAYS, W I (1933) *J Path. Bact.* 37, 45
- TILESTON, W (1916) *J Amer med Ass.*, 66, 995
- TUNNICLIFF E A. (1941) *J infect. Dis* 69, 52
- TUNNICLIFF, R (1926) *J infect. Dis*, 38, 366
- TUNNICLIFF R and JACKSON, L. (1930) *J infect. Dis.*, 46, 12
- VINCENT H (1894) *Ann. Inst. Pasteur*, 8, 129
- WAKSMAN, S A. (1918) *J infect Dis*, 23, 547
- WAKSMAN, S A and HENRICI A T (1943) *J Bact.*, 46, 337
- WOLFF, M and ISRAEL, J (1891) *Archives Arch.*, 128, 11

## CHAPTER 15

### ERYSIPELOTHRIX AND LISTERELLA

#### DEFINITION

Rod-shaped organisms with a tendency to the formation of long filaments which may show branching. The filaments may also thicken and show characteristic granules. No spores. Motility slight or absent. Gram positive. Slight fermentative activities. Microaerophilic. Usually parasitic.

The type species is *Erysipelothrix rhusiopathiae*, the causative organism of swine erysipelas.

The first member of this group to be described was the bacillus of mouse septicæmia *Erysipelothrix muriseptica*. It was found by Koch in 1880 in the blood of mice that had been injected subcutaneously with putrefying blood. In 1882 Loeffler (1886) observed a similar bacillus in the blood vessels of the skin of a pig that had died of swine erysipelas. [It is possible that the bacillus observed four months previously by Thuillier (Pasteur and Thuillier 1883) in pigs dying of *rouget* was the same organism as that described by Loeffler but this is not absolutely clear.] Another organism closely allied to *Ery. rhusiopathiae* was found by Rosenbach in cases of human erysipeloid. Subsequent workers have recorded the presence of *Erysipelothrix* in outbreaks of polyarthritides in sheep and joint ill in lambs and in occasional infections of cattle, horses, turkeys, peacocks and man (see Beaudette and Hudson 1936, Paterson and Heatley 1938, Greener 1939). There is also reason to believe that it is a not uncommon parasite of fish though conclusive evidence of this is still lacking (Klauder 1926, 1932, Schoop (1936) for example who recorded its isolation from fish used the mouse inoculation method so that it is impossible to be certain whether the organisms came from the fish or from latently infected mice).

In 1926 Murray, Webb and Swann at Cambridge described a disease of rabbits characterized by a large mononuclear leucocytosis, and caused by a small Gram positive non sporing bacillus which they termed *Bact. monocytogenes*. The same organism has since been isolated by a number of workers from various diseases in animals and man characterized most often by a generalized infection tending to localize in the liver, myocardium or central nervous system (see p. 1287). For this organism Pirie (1927) suggested the generic name of *Listerella*. Without for the moment discussing the appositeness or the validity of this name we may point out that *Erysipelothrix rhusiopathiae* and *Listerella monocytogenes* resemble each other in so many respects that it is convenient to describe them together.

**Habitat**—Enough has already been said to indicate the wide range of animals infected by these organisms. Though both of them appear to be mainly parasites *Ery. rhusiopathiae* is said to be present in the slime surrounding the body of various fish (Klauder *et al.* 1926, Klauder 1932, Schoop 1936) and in sewage derived from abattoirs (Hettche 1937).

**Morphology.**—The work of Spryszak and Szymanowski (1929), Meyn (1931) Redlich (1932) and Barber (1939) has made it clear that both *Erysipelothrix* and *Listerella* occur in a smooth and a rough form, each characterized by closely associated morphological and colonial appearances. In the smooth form they appear as small, straight or slightly curved, Gram positive rods with rounded ends, about 0.8–2.5  $\mu$  long and 0.3–0.6  $\mu$  broad arranged singly, in small packets or groups or in short chains. In the rough form long filaments, up to 60  $\mu$  or more, predominate some of which are seen breaking down to form chains of bacilli. *Erysipelothrix* is more slender than *Listerella* and is non motile. The motility of *Listerella* however, which is said by some workers to be due to a single polar flagellum and by others to peritrichate flagella is very sluggish. According to Seastone (1935) it is demonstrated best in a 4 hour glucose broth culture. Flagella are said to be developed better at 25° C than at 37° C (see Paterson 1939 Griffin and Robbins 1944).

**Cultural Characters.**—In the smooth form the colonies after 24 hours' incubation at 37° C are very small, circular, convex, amorphous, and water-clear, with a smooth glistening surface and entire edge. On further incubation *Erysipelothrix* colonies show little or no increase in size, those of *Listerella* become larger and less transparent. In the rough form the colonies are rather larger and flatter, their matt surface, curled structure, and fimbriate edge render them not unlike miniature anthrax colonies. On the whole *Listerella* colonies tend to be larger and less transparent than those of *Erysipelothrix*. In gelatin stab culture the growth of *Erysipelothrix* on first isolation is often of the lamp-brush type, *Listerella* forms a filiform growth with no lateral outgrowths. *Erysipelothrix* fails to grow on MacConkey's medium, *Listerella*, according to Paterson (1937) and our own observations forms small colonies on this medium, but Barber (1939) was unable to confirm this. *Listerella* forms a soluble haemolysin, *Erysipelothrix* does not (Barber 1939) though haemolysis may occur around colonies in blood agar.

**Resistance.**—Most strains of *Erysipelothrix* are killed by exposure to moist heat for 15 minutes at 55° C, *Listerella* survives this temperature for 30 minutes but is killed within 60 minutes (Barber 1939). *Erysipelothrix* is resistant to salting, pickling and smoking and may remain alive in putrefying carcasses for months. Hettche (1937) found that it survived for 4 to 5 days in drinking water, and for 12 to 14 days in sewage and aquarium water. *Listerella* is said to be fairly resistant to penicillin (Foley *et al* 1944).

**Growth Requirements and Metabolism.**—The growth of both organisms is favoured by glucose and to a less extent by blood and serum. According to Colella (1936) growth of *Erysipelothrix* occurs best in 0.1 per cent glucose broth and 0.5 per cent glucose agar, larger quantities of sugar were found to be inhibitory. A liver digest medium favours both organisms (Murray, Webb, and Swann 1926 Vawter 1937). On first isolation *Listerella* like *Erysipelothrix*, may give a band growth just below the surface of a shake agar culture (Gibson 1935), whether this is due to a preference for CO<sub>2</sub> or for a lowered partial pressure of oxygen is not clear. Both organisms are facultative anaerobes. Huttner (1942) found that the growth requirements of *Listerella* were simpler than those of *Erysipelothrix*. *Listerella* could grow in peptone water without serum or thioglycollate, whereas *Erysipelothrix* could not. Growth of *Listerella* is said to occur in an acid hydrolysed "vitamin free" medium containing glucose and inorganic salts, provided riboflavin,

biotin, and hæmin are added (Porter and Pelczar 1941) Growth of both organisms occurs between about 15° and 44° C, but is best at 30° to 37° C

**Biochemical Characters**—The fermentative activity of *Erysipelothrix* is less than that of *Listerella* Neither organism forms gas *Erysipelothrix* produces acid in glucose and lactose, *Listerella* produces acid in glucose, maltose, salicin, mannose, rhamnose, and dextrin, and slowly or in only small amount in lactose, sucrose and glycerol (see Deem and Williams 1936, Barber 1939, Julianelle 1941 Harvey and Faber 1941) According to Barber (1939) and Julianelle (1941) *Listerella* gives a positive and *Erysipelothrix* a negative Voges Proskauer reaction though Harvey and Faber (1941) record a negative reaction for the strains of *Listerella* that they studied

**Antigenic Structure**—There is general agreement that *Erysipelothrix* and *Listerella* are antigenically distinct Watts (1940) who studied 43 strains of *Erysipelothrix*, found that 38 appeared to belong to one antigenic type and 5 to another Each group possessed a heat stable specific antigen, and in addition two heat labile antigens, which were present in different proportions in the two groups and were responsible for cross agglutination Seastone (1935), Webb and Barber (1937), Schultz, Terry, Brice, and Gebhardt (1938), Paterson (1939 1940a) and Julianelle and Pons (1939) have brought evidence to show the existence of some antigenic diversity among *Listerella* strains. Paterson recognizes four types the division being made primarily on the H and secondarily on the O antigens

**Pathogenicity**—Both organisms have a wide range of pathogenicity for animals under natural conditions, and both may occasionally infect man (pp 1285 7) The diseases to which they give rise will be described in Chapter 58 here we shall concern ourselves with their effect on laboratory animals Three striking characteristics are possessed, though in differing degree by *Erysipelothrix* and *Listerella* Both produce a considerable monocytosis in rabbits pin point focal necroses in the liver of mice, and conjunctivitis in rabbits and mice After intra venous inoculation into rabbits each of the organisms gives rise to a considerable monocytosis, which reaches its maximum in 3 to 7 days (Webb and Barber 1937, Barber 1939) In mice inoculated subcutaneously or intraperitoneally focal necroses of the liver, up to pinhead in size are nearly always found post mortem in *Listerella* infections, but are much less numerous and striking in infections with *Erysipelothrix* Conjunctivitis, on the other hand, is commoner in *Erysipelothrix* than in *Listerella* infections of mice (Barber 1939) If however, the organisms are instilled into the conjunctival sac of the rabbit, or rubbed gently on the everted lid *Listerella* gives rise to a severe conjunctivitis and keratitis whereas *Erysipelothrix* causes a milder conjunctivitis without keratitis (Anton 1934, Graham, Hester, and Levine 1940, Julianelle 1941) According to Barber (1939) *Listerella* kills guinea pigs but not pigeons, *Erysipelothrix* kills pigeons but not guinea pigs In assessing these differences it must be remembered that the virulence of both organisms is subject to variation, and that unless freshly isolated strains in the smooth phase are used the results of inoculation may be equivocal In general it may be said that both organisms give rise to a septicæmic infection, and that *Erysipelothrix* has a tendency to localize in the skin, endocardium, and joints, and *Listerella* in the liver, myocardium, and central nervous system Both *Erysipelothrix* and *Listerella* appear to be more virulent in the smooth than in the rough form (Schoening *et al* 1938, Barber 1939)

Inoculation of *Erysipelothrix* into Animals

**SWINE.**—Loeffler (1886) who first isolated the swine erysipelas bacillus failed to reproduce the disease in swine with pure cultures, but Schutz (1886) later succeeded in doing so. Broth cultures injected subcutaneously proved fatal to two pigs, one animal dying in 3 the other in 4 days, there were typical findings at the necropsy and the bacilli were recovered in pure culture from the blood and spleen, and from the pleural and peritoneal exudates. Artificial cultures rapidly lose their virulence for swine. Collins and Goldie (1940) produced polyarthritis by repeated intravenous inoculation of cultures. In addition there was a focal inflammatory polyarthritis, focal necrosis of the liver and myocardium, lymphadenopathy, a monocytosis, and endocarditis but skin lesions were never found.

The bacillus is pathogenic for mice, pigeons and rabbits, but not for guinea pigs.

**MICE.**—0.001–0.1 ml. of a 24 hours broth culture injected subcutaneously or intraperitoneally is usually fatal in 2 to 3 days. During life the mice develop conjunctivitis and their lids become glued together with a muco-purulent secretion. Arching of the back is very common, and constipation is usual. Post mortem the vessels of the skin and subcutaneous tissue are congested, the spleen is enlarged and the lungs are bright red and oedematous. Bacilli are usually abundant in the blood and viscera; they are found particularly within the phagocytic cells, in which they appear to multiply (Tenbroeck 1920).

**PIGEONS.**—0.001–0.1 ml. of a 24 hours broth culture inoculated intramuscularly proves fatal in 3 or 4 days as a rule. Death is often preceded by paralysis of the legs, dyspnoea, and convulsions. Post mortem there is a black hæmorrhagic mass in the muscle at the site of inoculation, the spleen is enlarged, there are often punctiform hæmorrhages in the mucosa and viscera, and there is almost constantly a clear lemon yellow exudate in the pericardium (Crump 1914). The bacilli are fairly numerous in the blood and organs.

**RABBITS.**—0.5 ml. of a 24-hours broth culture inoculated intravenously sometimes proves fatal in 2 to 3 days. A marked oedematous swelling or erysipelatous rash develops in the injected ear and there is a rise in temperature and a loss in weight. Post mortem besides the rosy skin lesion there is congestion of the viscera, and often a clear lemon yellow pericardial exudate, there may be large hæmorrhages into the lungs. The bacilli are scarce. If the disease is not acutely fatal, a monocytosis occurs, reaching its maximum in 3 to 7 days. In animals dying about this time occasional tiny focal necroses may be found in the liver and areas of mononuclear cell reaction may be seen in sections of the spleen. Inoculation of the conjunctiva gives rise to conjunctivitis which often proves fatal. After subcutaneous inoculation death seldom occurs.

Inoculation of *Listerella* into Animals

**FISH.**—Hettebe (1937) and Brunner (1938) have shown that both freshwater and sea fish can be readily infected by feeding or by intraperitoneal inoculation with the bacilli. The organisms are widely distributed in the tissues and may be recovered after several weeks. They are particularly abundant in the kidneys and are excreted in the urine. The infection appears to be of the covert type, the fish showing no evidence of illness.

**MICE.**—Subcutaneous or intraperitoneal inoculation of 100 million living organisms of a highly virulent culture causes death in about 1 to 4 days. During life conjunctivitis sometimes occurs. At post mortem multiple tiny focal necroses are found scattered throughout the liver. The organisms can be readily recovered from the spleen and heart blood.

**RABBITS.**—Intravenous inoculation proves fatal in 24 hours or not for several days according to the dose and virulence of the strain. Animals surviving for some time develop a monocytosis which reaches its maximum in about 3 to 7 days. Post mortem examination of animals dying after a few days reveals the presence of multiple focal necroses in the liver and rarely in the spleen and myocardium. Necrosis of the supra-



imals is common. Occasionally abscesses in the myocardium and inflammation of the meninges may be met with (see Burn 1935). After intraperitoneal inoculation much the same lesions are found, but in addition there is a sero-fibrinous peritonitis, with abscesses containing thick white pus in the rolled-up omentum. The organisms can rarely be recovered from the blood stream. Instillation of a pure culture into the conjunctiva or swabbing of the everted lid gives rise to a severe conjunctivitis within 24 hours followed by keratitis, the animal itself rarely dies.

**GUINEA PIGS.**—These animals die after inoculation with large doses. The lesions at necropsy are similar to those in mice. The organisms may be recovered from the spleen and sometimes from the heart blood.

**CHICK EMBRYOS.**—Paterson (1940b) has shown that *Listerella* gives rise to focal lesions on the chorio-allantoic membrane of chick embryos.

**Classification.**—In the *Erysipelothrix* group three species were differentiated by Rosenbach (1909)—*muriseptica*, *porci* and *erysipeloides*. Rickmann (1909) however, pointed out that the morphological and cultural distinctions on which Rosenbach relied for differentiation were insufficient to serve as a means of classification, and since he found that all three organisms agglutinated to the same titre with immune sera, and exhibited the same pathogenicity to animals he concluded that they should be regarded as belonging to a single species. To this the name *Erysipelothrix rhusiopathiae* is now commonly applied.

For the organism isolated by Murray, Webb, and Swann (1926) and called non-committally by them *Bacterium monocytogenes* Pirie (1927) suggested the generic name *Listerella* in honour of Lord Lister. Apart from the inappropriateness of this name—Lord Lister having neither discovered nor worked with this organism—it was too premature, since at that time no thorough comparison had been made between *Bact. monocytogenes* and *Erysipelothrix*, which it resembles in many respects (Topley and Wilson 1936).

Barber (1939) and Julianelle (1941) have now carried out a comparative study of these two organisms, and their observations seem to show that the relationship between them is even closer than was at first suspected. Thus both organisms are small, Gram positive rods, showing an extraordinary similarity in the morphological and cultural appearances of their smooth and rough forms. Both have much the same growth requirements and much the same degree of resistance. Both have a wide range of pathogenicity for animals, and both occasionally give rise to disease in man. In rabbits, experimental inoculation of either organism results in the development of a generalized infection accompanied by the appearance of conjunctivitis and, if the disease is not acutely fatal, of a considerable monocytosis, at post mortem focal necroses are found in the liver. Against these similarities, it may be said that *Listerella* is faster and is motile, attacks rather more sugars, is antigenically distinct, and is fatal experimentally to guinea pigs but not to pigeons, in contrast to *Erysipelothrix* which is fatal to pigeons but not to guinea pigs.

No one who is familiar with systematic bacteriology and who has observed several strains of each of these organisms side by side can deny that the differences between them are less than those existing between different species of many other organisms classified in the same genus, as, for example, between members of the *Corynebacterium*, *Pasteurella*, *Bacillus*, or *Clostridium* groups. The differences between *Erysipelothrix* and *Listerella* are those characteristic of specific, not of generic, differences, and it seems to us to be doing violence to the principles

of bacterial taxonomy to classify them in separate genera. Our proposal, therefore, is to include them both in the *Erysipelothrix* genus, calling them *Ery rhusiopathiae* and *Ery monocytogenes* respectively. It may be pointed out that in any case the name *Listerella* is invalid, since it was given to a mycetozoan by Jahn in 1906 (see Pirie 1940).

### *Erysipelothrix rhusiopathiae*

*Synonyms*—*B. rhusiopathiae* *sensu* Kitt, *Erysipelothrix porci* Rosenbach; *B. erysipelas sensu*

*Isolation*.—Observed independently by Thuillier (Pasteur and Thuillier 1883) and Loeffler (1886) in 1882.

*Habitat*.—Found on the mucosae and tonsils of swine.

*Morphology*.—*Smooth form* Small, slender, straight or slightly curved rods, 0.8–2.0  $\mu$  long and 0.3–0.4  $\mu$  broad. Arranged singly, in small packets, in small groups, or in short chains. *Rough form* Long chains of bacilli and interlaced filaments of variable length. Staining is fairly regular, but sometimes deeply stained granules may be seen. Non motile. Non sporing. Gram positive.

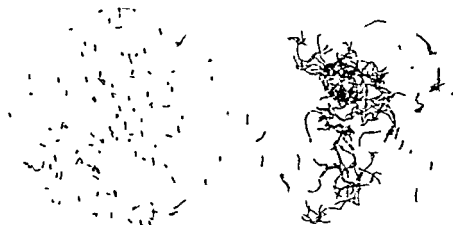


FIG. 66.—*Erysipelothrix rhusiopathiae*.

Left Smooth form. Right Rough form. From a surface agar culture, 3 days 37° C. ( $\times 1000$ )

*Agar Plate*—24 hours, 37° C. *Smooth form* Round, convex, tiny, amorphous, water clear colonies, 0.1 mm. in diameter, with smooth glistening surface and entire edge, butyrous and easily emulsifiable. No increase in size on further incubation. *Rough form* Rather larger and flatter, 0.2–0.4 mm. in diameter, with a granular curled appearance and fimbriate edge, resemble miniature anthrax colonies.

*Agar Slope*.—24 hours, 37° C. Very poor, partly confluent, slightly raised, colourless, transparent growth with an irregular surface due to imperfect fusion of individual colonies, and an edge which is very finely dentate or made up of single colonies. Practically no change on further incubation, growth may become slightly viscous.

*Gelatin Plates*.—Deep colonies in 3 or 4 days resemble snow-flakes, they are very small, but when magnified 50 times they are seen to consist of a granular centre with branching threads radiating outwards.

*Gelatin Slab*.—Growth occurs slowly and is subject to considerable variation, apparently depending to some extent on the reaction of the medium. May be a simple

filiform growth extending to the bottom of the tube. Usually lateral outgrowths occur, these may be ill-defined looking like snow flakes nebulae or the conventional bursting bomb or they may be definite branches producing a lamp-brush appearance. The outgrowths may extend for a distance of only 2 or 3 mm from the stab or they may reach the sides of the tube. The smooth form tends to remain restricted to the line of inoculation while the rough form grows out laterally. No liquefaction. The lamp-brush form may not be obtained till after 2 or 3 subcultures in gelatin.

*Broth*—24 hours 37° C. Smooth form Slight uniform turbidity with very slight powdery deposit disintegrating on shaking. After a few days the broth may clear and a viscous deposit become evident. Rough form Little or no turbidity. Flocculi of varying size or tangled hair like masses of growth appear and settle on the sides or bottom of the tube, they are difficult to disintegrate by shaking.

*Loeffler's Serum*.—7 days, 37° C. Very poor, confluent slightly raised colourless growth slightly better than on agar.

*MacConkey's Agar*.—No growth.

*Potato*.—No visible growth.

*Glucose Agar Shake*.—Very tiny colonies throughout medium on first isolation there may be a band growth just below surface.

*Resistance*.—In broth cultures the bacilli are killed by moist heat at 55° C. in 15 minutes. In meat they are highly resistant to salting pickling and smoking surviving for 1 to 3 months, they are likewise resistant to putrefaction, remaining alive and virulent for months in putrefying buried cadavers. Apparently succumb readily to drying, provided this is complete.

*Metabolism*.—Microaerophilic but will grow under both aerobic and anaerobic conditions. Optimum temperature for growth is 30° C., grows between about 15° and 44° C. Growth favoured by glucose and slightly by blood. Hemolysis occurs round deep colonies in 10 per cent horse blood agar plates.

*Biochemical*.—Sugar reactions are variable. Usually forms acid in dextrose and lactose, not in maltose mannose rhamnose mannitol sucrose dextrin, or salicin. L.M. no change or very slight acid. Indole negative. MR negative. VP negative. Nitrates slight reduction. Catalase  $\pm$ . MB reduction negative.  $\text{NH}_3$  negative.  $\text{H}_2\text{S}$   $\pm$ .

*Antigenic Structure*.—Evidence of two antigenic types each possessing a heat-stable specific antigen, and two heat-labile antigens that are present in different proportions in the two groups and are responsible for cross-agglutination.

*Pathogenicity*.—Causes swine erysipelas in swine and erysipeloid in man. Experimentally it is pathogenic for mice pigeons and rats but not for guinea pigs. A non fatal dose inoculated into rabbits gives rise to a circulating monocytosis. Virulence for swine is said to fall on artificial cultivation. No exotoxin is produced.



FIG. 6.—*Erysipelothrix rhusiopathiae*.

Smooth form Surface colonies on agar 24 hours 37° C. ( $\times 8$ )

### *Erysipelothrix monocytogenes*

*Synonyms*.—*Bact. monocytogenes* Murray Webb and Swann, *Listerella monocytogenes* Fine.

*Isolation*.—By Murray, Webb and Swann (1926) from rabbits suffering from a natural disease.

*Habitat*.—Widespread animal parasite.

**Morphology**—*Smooth form* Small, straight or slightly curved rods 0.8–2  $\mu$  long and 0.5–0.6  $\mu$  broad. Arranged singly, in pairs side by side or in V form, in small packets, or in short chains. *Rough form* Long interlacing filaments some of which show segmentation into rods. Filaments are particularly abundant in cultures at room temperature. Sluggishly motile by polar, or according to some workers peritrichate flagella. Stains fairly uniformly, but sometimes shows beading. Non-sporing. Gram positive.

**Agar Plate**—24 hours, 37° C Poor to moderate growth, having slightly acid smell reminiscent of the acne bacillus. *Smooth form* Circular, low convex or convex amorphous colonies, 0.2–0.8 mm in diameter, with smooth mirror like surface and entire edge, almost transparent by transmitted, greyish yellow by reflected light, butyrous in consistency and easily emulsifiable. 48 hours Slightly larger, 0.8–1.0 mm in diameter, with tendency to crinkling of edge. Older colonies are up to 1.5 mm in diameter, and have a brownish slightly granular centre and a translucent effuse periphery with a finely crenated margin, when magnified, the colony has a poached-egg appearance. *Rough form* 48 hours Rather larger than the smooth form, 1.5–2.0 mm in diameter flatter, with a matt surface umbonate centre, and a finely fimbriate edge. After 5 days colony may reach 5 mm. in diameter, and show a small, smooth, darker centre with a wide, more effuse, translucent finely granular peripheral extension having a fimbriate edge.

**Agar Slope**—24 hours, 37° C Thin, raised, confluent translucent growth with beaten copper surface and finely crenated edge. Growth becomes more abundant on further incubation. Pough form is similar, but shows a slightly spreading edge.

**Gelatin Slab**—Slow, filiform growth, not liquefying the gelatin, narrow surface growth.

**Broth**—24 hours 37° C *Smooth form* Slight to moderate turbidity, with slight powdery deposit disintegrating on shaking, no surface growth. *Pough form* Little or no turbidity, with thread like masses of deposit that are difficult to disintegrate.

**Horse Blood Agar Plate**—24 hours, 37° C Circular, low convex colonies, 0.7–1.2 mm in diameter, with smooth mirror like surface and entire or very finely crinkled edge, milky white by reflected light. Colonies are surrounded by a narrow zone of more or less complete  $\beta$  haemolysis.

**Loeffler's Serum**—2 days 37° C Thin, raised growth, like that on agar, with slightly beaten-copper surface and very finely crenated edge, no liquefaction, even after 3 weeks.

**MacConkey's Agar**—24 hours, 37° C Circular, convex, colourless colonies, 0.1–0.4 mm in diameter, with smooth surface and entire edge. 5 days No increase in size or change in colour, but surface tends to be slightly granular and edge slightly irregular.

**Potato**—2 days, 37° C Slightly raised, greyish white growth, with finely granular surface and very finely crenated edge.

**Glucose Agar Shale**—2 days, 37° C Tiny colonies throughout medium. May be a band growth, on first isolation, just below surface.

**Resistance**—Survives moist heat for 30 minutes at 55° C, but is killed within 60 minutes. Cultures remain viable for several months.

**Metabolism**—Aerobic and facultatively anaerobic. Optimum temperature for growth 30°–37° C, growth occurs between 20° and 44° C. Growth favoured by glucose and liver extract and to a less extent by blood and serum.  $\beta$  haemolysis occurs around colonies on horse blood agar, weak soluble haemolysin formed in broth cultures.

**Biochemical**—Sugar reactions are variable. Usually produces acid in glucose, maltose, salicin, mannose, rhamnose, and dextrin, and slowly or in only small amount in lactose, sucrose, and glycerol. L.M. slight acid. Indole negative. M.R. + V.P.  $\pm$ . Nitrate reduction negative. M.B. reduction weakly positive. H<sub>2</sub>S negative. NH<sub>3</sub> in serum peptone water weakly positive. Catalase  $\pm$ .

- Antigenic Structure.**—Probably 4 types, distinguished primarily on flagellar and secondarily on somatic antigens. Types 1 and 2 have the same H but different O antigens. Types 3 and 4 have each specific H and specific O antigens.
- Pathogenicity.**—Causes disease in a wide variety of animals, including occasionally man. Experimentally it is pathogenic for mice, rabbits, guinea pigs, but not pigeons. Non-lethal doses set up a monocytosis in rabbits. Virulence often falls on artificial cultivation. No exotoxin is produced.

## REFERENCES

- ANTON, W. (1934) *Zbl Bakt.*, 131, 89.  
 BARBER, MARY (1939) *J Path Bact.*, 48, 11.  
 BRAUDETTE F R and HUDSON, C B. (1936) *J Amer vet med Ass.*, 88, 475.  
 BRUNTER, GERTH (1938) *Zbl Bakt. Abt.*, 97, 477.  
 BURN, C G. (1935) *J Bact.*, 30, 573.  
 COLELLA, C. (1930) *Nuova Vet.*, 14, 6.  
 COLLINS, D H and GOLDIE, W. (1940) *J Path Bact.*, 50, 323.  
 CRIMP, P. (1914) *Ann. Stat. Mal. Best. Napoli*, 2, 107.  
 DEEM, A W and WILLIAMS, C L. (1936) *J Bact.*, 32, 303.  
 FOLEY, E J, EPSTEIN, J A, and LEE, S W. (1941) *J Bact.*, 47, 110.  
 GIBSON, H J. (1935) *J Path Bact.*, 41, 239.  
 GRAHAM, R, HESTER, H R, and LEVINE N D. (1940) *J infect Dis.*, 68, 91.  
 GREENER, AERIL W. (1939) *Brit J Derm Syph.*, 51, 372.  
 GRIFFIN, A M and ROBBINS M L. (1944) *J Bact.*, 48, 114.  
 HARVEY, P C and FARER, J E. (1941) *J Bact.*, 42, 677.  
 HETTCHER H O. (1937) *Arch Hyg.*, 119, 178.  
 HUTNER, S H. (1942) *J Bact.*, 43, 629.  
 JULIANELLE, L A. (1941) *J Bact.*, 42, 367, 385.  
 JULIANELLE L A and PONS, C A. (1939) *Proc Soc exp Biol N Y.*, 40, 364.  
 KLAUDER, J V. (1926) *J Amer med Ass.*, 86, 536, (1932) *J industr Hyg.*, 14, 222.  
 KLAUDER, J V, RIGTER, L L, and HARRIS M J. (1926) *Arch Derm Syph N Y.*, 14, 662.  
 KOCH, R. (1880) 'Investigations into the Etiology of Traumatic Infective Diseases'. New Sydenham Soc London.  
 LOEFFLER (1886) *Arch Reichsgesundh.Amt.*, 1, 46.  
 MEY, A. (1931) *Zbl Bakt.*, 122, 507.  
 MURRAY, E G D, WEBB, R A, and SWANN, M B R. (1926) *J Path Bact.*, 29, 407.  
 PASTEUR and THUILLIER. (1883) *C R Acad Sci.*, 97, 1163.  
 PATERSON, J S. (1937) *vet Rec.*, 49, 1533, (1939) *J Path Bact.*, 48, 25 (1940a) *Ibid*, 51, 427, (1940b) *Ibid*, 51, 437.  
 PATERSON, J S and HEATLEY, T G. (1938) *vet J.*, 94, 33.  
 PIRIE, J H H. (1927) *Publ S Afr Inst med Res.*, 2, 163 (1940) *Nature*, 145, 264.  
 PORTER, J R and PELCZAR M J. (1941) *J Bact.*, 42, 141.  
 REDLICH, E. (1932) *Z Infekth. Haustiere.*, 42, 300.  
 RICKMAN, (1909) *Z Hyg InfektKr.*, 64, 362.  
 ROSENBAUM, F J. (1909) *Z Hyg InfektKr.*, 58, 343.  
 SCHOKING, H W, GOCHENOUR, W S, and GRIFFIN, C G. (1933) *J Amer vet med Ass.*, 92, 61.  
 SCHOOFF, G. (1936) *Dtsch. tierarztl. Wochn.*, 44, 371.  
 SCHULTZ, E W, TERRY, M C, BRICE, A T, and GERHARDT, L P. (1938) *Proc Soc exp Biol.*, N Y, 38, 605.  
 SCHÜTZ (1886) *Arch Reichsgesundh.Amt.*, 1, 56.  
 SEASTONE, C V. (1936) *J exp Med.*, 62, 203.  
 SPETZLAK, A and SZYMANOWSKI Z. (1929) *C R Soc Biol.*, 100, 1151.  
 TENBROECK, C. (1920) *J exp Med.*, 32, 331.  
 TOPLEY, W W C and WILSON, G S. (1936) 'The Principles of Bacteriology and Immunity', 2nd ed., p. 709.  
 VAWTER L R. (1937) *J Amer vet med Ass.*, 90, 635.  
 WATTS, P S. (1940) *J Path Bact.*, 50, 365.  
 WEBB, R A and BARBER, M. (1937) *J Path Bact.*, 45, 523.

## CHAPTER 16

### MYCOBACTERIUM

#### DEFINITION *Mycobacterium*

Slender rods which are stained with difficulty but which when once stained are acid fast. Cells are sometimes swollen, clavate, cuneate, or even branched. Non motile. Gram positive. No endospores. Growth on media slow. Aerobic. Several species are pathogenic to animals.

Type species is *Mycobacterium tuberculosis*

**The Acid fast Bacteria.**—The acid fast bacteria are so called because of their ability, when once stained to resist subsequent decolorization by mineral acids. The degree of acid fastness varies with different members of the group, and in any single member is liable to alteration with changed environmental conditions, these differences are never so distinct or so constant as to serve as a reliable means of differentiating between the members of the group.

The first member to be discovered was the *leprosy* bacillus in 1868 (see Hansen 1874). In 1882 came the discovery by Koch of the *mammalian tubercle* bacilli. The work of Smith (1888), Vagedes (1893), Ravenel (1901), Kossel, Weber and Heuss (1904-1905), the English Royal Commission (Report 1911), and Park and Krumwiede (1910) during the years 1898-1910, showed that these mammalian bacilli could be divided into two types—the human and the bovine.

The discovery of the *avian* type of tubercle bacillus was due largely to the work of Rivolta (1889), Maffucci (1890-1892), Cadot, Gilbert and Roger (1890), Sibley (1890), and Straus and Gamaleia (1891) during the years 1889 to 1891. The work of Sibley (1889), Bataillon, Dubard and Terre (1897), Ledoux Lebard (1893-1900), Friedmann (1903), and Kuster (1905) from 1889 to 1905 served to differentiate a fourth type of tubercle bacillus—the *cold-blooded* type. In 1895 Johnes and Frothingham described the organism which is now known as *Johnes's* bacillus, and which is responsible for a chronic enteritis in cattle. The rat leprosy bacillus was discovered by Stefansky (1903) in 1901 at Odessa. During the years 1885 to 1906 a number of workers demonstrated the existence of the *saprophytic acid fast* bacilli—a group which though able, under experimental conditions, to give rise in mammals to lesions closely simulating those of tuberculosis, does not appear capable of causing a definitely progressive disease. Amongst these bacilli the most important are (1) the butter bacillus *Myco butyricum*—isolated by Rabnowitsch in 1897 from 23 out of 80 specimens of market butter examined in Germany, and since then by numerous other workers (Petri 1898, Korn 1899, 1900, Tobler 1901, Beck 1905, Pellegrino 1906), (2) Moeller's Grass bacilli 1 and 2. Moeller isolated his first bacillus in 1898 from timothy grass (*Phleum pratense*), hence it is generally referred to as *Mycobacterium phlei* 1 or more familiarly as the

timothy grass bacillus, his second bacillus was isolated in 1899, from the dust of some plant material used as fodder, (iii) the Mist bacillus or *Myco stercoris* this was isolated by Moeller in 1901 from a dung heap and later from the faeces of cows, donkeys, and other herbivora, it owes its name to the German term for manure (*Mist*); (iv) the Smegma bacillus, *Myco smegmatis* this was first described by Alvarez and Tavel in 1880, but was not obtained in pure culture till 1897, when Laser (1897) and Czaplewski (1897) cultivated it independently. This organism is present in varying numbers in smegma of both males and females it has been found in the smegma of dogs (Pellegrino 1906)

Finally, in 1937 Wells isolated an acid fast bacillus from voles (*Microtus agrestis*) suffering from natural tuberculosis. This organism differs in certain respects from the other mammalian types, and is probably best referred to as the murine type of tubercle bacillus or *Myco muris*

We shall describe the tubercle and saprophytic acid fast bacilli together in the body of this chapter, but shall reserve the leprosy, rat leprosy and Johnes bacillus for a separate description at the end

**Habitat**—The tubercle bacilli are essentially pathogenic, so far as we know they do not multiply naturally outside the animal body. The human bovine and murine bacilli give rise to mammalian tuberculosis (see Chapter 59). The avian type is found chiefly in birds, though it often infects pigs and occasionally cattle. It is sometimes present in hens' eggs (see Gloyne 1933). The cold blooded type is responsible for disease in cold blooded animals and fish. The saprophytic acid fast bacilli are found in such diverse surroundings as butter milk smegma, grass, manure, and faeces, they are also widely distributed in dust and water. The presence of metal seems to favour their growth and they can almost invariably be found in scrapings from metal cold water taps (Brem 1909, Beitzke 1910) and metal wind instruments (Jacobitz and Kayser 1910). They have been reported in cultures made from a gangrenous lung (Rabinowitsch 1900) from human faeces (Mironescu 1901), from the tonsils (Marzinowsky 1900 Beck 1905) from the nasal secretion (Karlinski 1901, Marchoux and Halphen 1912) from the intestinal contents of insects (Pellegrino 1906), from cow's milk (Albiston 1930) from a pleural exudate (Beaven and Bayne Jones 1931), from pus (Bruynoghe and Adant 1933) from sputum (Cummins and Williams 1933), and from blood (Tiedemann 1931 Schwabacher 1933a). In view, however, of the frequency of acid fast bacilli in dust, it seems probable that some of these organisms gained access to the cultures by air contamination and were not necessarily present in the material from which they were apparently derived. The leprosy bacillus is a specific parasite of man, and the rat leprosy bacillus of rats. Johnes's bacillus infects cattle, and to a less extent sheep, in both of which it causes a chronic enteritis.

**Morphology and Staining**—The acid fast bacilli are rod shaped 'organisms straight or slightly curved, with more or less parallel sides and rounded ends, they are arranged either singly, in small groups or bundles, or in groups of three or four with the individual bacilli lying at acute angles to each other, resembling diphtheria bacilli. Their size varies considerably according to the medium on which they are grown. In the animal body they are generally longer and thinner than in culture. Their length is 1-4  $\mu$ , but occasional forms as long as 8  $\mu$  are seen, in breadth they vary from about 0.3-0.6  $\mu$ . Long filamentous acid fast bacilli have been described, but it is probable that these belong to the *Actinomyces* group (see Chapter 14). Numerous authors, however have stated that

the tubercle bacilli are capable of producing filaments—particularly in liquid media—but these forms are not encountered under ordinary conditions. Clubbed forms, resembling the typical clubs of the diphtheria bacillus are not uncommon in culture. Branched forms have been described by some authors but are probably infrequent except in the avian bacilli. Staining is either uniform or granular. In the latter type the granules may be restricted to the poles or they may be evenly distributed throughout the length of the bacillus—the so-called beaded form. In bacilli that appear to be undergoing degeneration the staining is often irregular both in depth and in situation. In young cultures it is common to find a certain proportion of non acid fast forms. The morphology of the developing organisms may be studied by Pryce's (1941) slide-cell technique of micro-culture.

On the average human bacilli tend to be long thin and curved and to show granular staining while bovine bacilli tend to be short straight and thick and to show uniform staining. Their morphology however is so variable and is so dependent upon environmental factors that no weight can be attached to these



FIG 68—*Mycobacterium tuberculosis*

Glycerine egg-culture 4 weeks, 3° C., showing short, straight forms of bacilli (× 1000)



FIG 69—*Mycobacterium tuberculosis*

Glycerine agar culture 4 weeks, at 3° C., showing some short straight forms, and some longer curved forms (× 1000)

criteria in the identification of individual strains. Murine bacilli, according to Griffith (1949) are slender and often longer than human bacilli. Curved forms of the shepherd's crook, sickle spiral and S-type are abundant and characteristic. Some organisms show fine granulation and vacuolation along their whole length. The acid fast bacilli are Gram positive. Staining is not always easy but with a 5 per cent solution of gentian violet in alcohol and aniline oil, aided by gentle warming it is usually possible to obtain satisfactory preparations. According to Kretschmer (1934) the Gram positiveness is independent of treatment with iodine and is closely bound up with the property of acid fastness.

The organisms are resistant to simple solutions of the aniline dyes. To overcome this difficulty several methods of staining have been devised. Koch (1889) first stained the tubercle bacillus by immersion for 24 hours in an alkaline solution of methylene blue. Ehrlich (1882) improved on this by using aniline oil basic fuchsin or aniline oil methyl violet. By this means the bacilli were stained in 15 to 30 minutes and subsequently resisted decolorization with 33 per cent  $\text{HNO}_3$  for a few



seconds. He believed that the reason why the bacilli were resistant to ordinary stains was that they were surrounded by a capsule which was permeable only to alkalies. Ziehl (1882) showed that this conception was wrong, the bacilli could be stained quite satisfactorily by a dye of acid reaction. The stain he advocated was a 2 per cent alcoholic methyl violet solution in carbolic acid water. Later Ziehl employed carbol fuchsin. Neelsen increased the strength of phenol in the stain and the *Ziehl Neelsen method* is the one that is now usually employed.

It consists in covering the film with carbol fuchsin (basic fuchsin 1 part absolute alcohol 10 parts, and 5 per cent phenol in water 100 parts) and gently heating till the steam rises, the heating is continued for 5 to 15 minutes the water lost by evaporation being replaced by fresh stain. The film is then washed thoroughly in water and treated with a 15-20 per cent solution of a mineral acid. If the film is from a pure culture the effect of the acid will be merely to dissolve the excess stain, but if a film of tuberculous pus or a section of tuberculous tissue is being treated the acid will turn the preparation yellow, indicating that the stain has been removed from the tissue cells or from other organisms that may be present. The treatment with acid is continued for 5 to 10 minutes as a rule, till subsequent washing with water causes no more than a faint pink tinge to reappear. The film is then thoroughly washed in running water to remove all the acid. It is counterstained with a 1 per cent aqueous solution of methylene blue for 5 minutes after which it is washed and dried in the usual way. By this method the acid fast bacilli are coloured red while the tissue cells and all other organisms are coloured blue. Some workers prefer a yellow counterstain—usually 1 per cent picric acid. The success of this method depends partly upon the heat employed which renders the waxy material in the tubercle bacillus more permeable to aqueous dyes and partly on the phenol which acts as a mordant.

Numerous other methods of staining have been described (Spengler 1907, Herman 1908, Mori 1911, Bozzelli 1914, Schulte-Tigges 1920, Kieffer 1921, Shoub 1923, Pottenger 1912).

The property of acid fastness appears to be due to the presence in the bacilli of unsaponifiable wax (Anderson 1932). This substance was apparently first isolated by Aronson in 1898. It was referred to as one of the higher alcohols by Bulloch and Macleod (1904), and was termed "mykol" by Tamura (1913). The larger the amount of chloroform soluble wax the greater is the resistance of the bacilli to decolorization (Darzine 1932). The tubercle bacilli contain more of this substance than the saprophytic acid fast bacilli (Table 25), and are therefore usually more strongly acid fast. The degree of acid fastness however, is dependent on a number of factors, and no reliance should be placed on it in determining the particular type of organism under investigation. Not all workers are agreed on the simple chemical explanation of acid fastness just given. Sordelli and Arena (1934) for example, state that it is a property of intact bacilli, and believe that it depends on the existence of a semi permeable membrane around the organisms which allows fuchsin to diffuse in but does not allow acid fuchsin to diffuse out.

In formal fixed tissue sections the bacilli often stain very poorly with Ziehl Neelsen. According to Fielding (1934) this is due to the development of an acid reaction following autolysis of the tissues and can be overcome by fixing in a weakly alkaline solution of formal or by staining with alkaline fuchsin.

*Much's granules*—Much (1907) brought evidence to show that under certain conditions the bacilli might be present in the tissues in the form of non acid fast granules. Starting from the observation that in the *Perlsucht* nodules of cattle, and in cold abscesses of man

it was often impossible to find acid fast bacilli in films even though the presence of tubercle bacilli could be shown by culture and by pathogenicity experiments, he devised a number of different staining methods to determine whether bacilli of any sort could be demonstrated microscopically. The method that he found most successful was to stain for 24 to 48 hours in aniline gentian violet or carbol methyl violet at a temperature of 37° C., to treat with Lucol's iodine solution and to decolorize by a mixture of absolute alcohol and clove oil, or by a dilute mineral acid and a mixture of alcohol and acetone. Examining someiliary tubercles of a calf that had been injected with virulent bovine bacilli, he failed to find any bacilli in preparations stained with Ziehl-Neelsen, but in preparations stained by his own method he found large numbers of fine rods in the tubercles, often accompanied by small rounded granules arranged singly in pairs, or in short chains resembling beaded bacilli. The rods preponderated in the necrotic portions of the tubercles, the granules in the peripheral zones—both were coloured violet and both were numerous within the cells. Small pieces of the lung were seeded on to the serum slopes, and incubated at 37° C.—smears were examined daily. In smears stained with Ziehl-Neelsen no bacilli were found for 6 days, when acid fast rods appeared—but in smears stained by Much's method fine granules and rods were visible after 3 days. Small pieces of the lung injected subcutaneously into guinea pigs gave rise to generalized fatal tuberculosis in 8 weeks—acid fast bacilli were found in the tissues of the dead animals. Much obtained similar results with other tuberculous material. He concluded that—(i) There is a form of tubercle bacillus that is not stainable by Ziehl, but is stainable by Much's method—it is granular. (ii) In tuberculous organs this granular form may be the only stainable form of bacillus present. (iii) The granular form may be accompanied by fine rods, which likewise do not stain with Ziehl. (iv) The granular forms are virulent. (v) There are transition forms between the Gram + granules, the fine Gram + rods, and the acid fast rods and granules.

For a long time comparatively little attention was paid to Much's work—but of late years a number of observers have studied the growth of acid fast bacilli in suitable culture preparations, and have demonstrated the presence of granular forms similar to those described by Much. The interpretation of these forms, however, has given rise to controversy. While Swaney (1923) and Kahn (1930) hold that they represent a stage in the life-cycle of the bacilli, Oerskov (1939) believes that they are products of degeneration. The bundles of extremely fine rods that Kahn described as forming part of the life-cycle are interpreted by Oerskov as crystals, formed partly from the medium and partly from bacillary products. According to Legian and Porter (1944) many of the non-acid fast forms are frank artefacts resulting from trauma to the organisms during the preparation of the film. These observers find that destruction of the integrity of the cell is accompanied by a loss of the acid fast staining property. Young organisms are more readily destroyed than old—and the greater the effort spent in spreading the growth with a spatula on the slide—the more numerous are the non-acid fast rods and granules seen microscopically. The micro-motion pictures obtained by Wyckoff (1934) and Wyckoff and Smithburn (1933) show that the young bacilli increase in size before dividing—but that, as the culture ages, division continues without previous enlargement. The resulting organisms, therefore, become shorter and shorter till true coccoid forms staining intensely acid fast appear. Transplanted into a fresh medium, these short forms again give rise to typical bacilli. The sequence of events is so similar in general outline to the behaviour of non-acid fast bacteria—that there seems no justification for postulating the existence of any special cycle of development.

**Filtrable Forms of the Tubercle Bacillus**—Closely connected with the presence of Much granules is the existence of the so-called filtrable forms of the tubercle bacillus. Since Fontes (1910) original observation, numerous workers have claimed to demonstrate under appropriate conditions the presence of filter passing forms possessing a low degree of pathogenicity for guinea pigs and constituting a special stage in the life-cycle of the organism. This subject was reviewed in the second edition of Topley and Wilson (pp

291-3), and the conclusion reached that no satisfactory evidence had been brought forward to prove the existence of such forms. The more recent findings of Soltys and Taylor (1944) support this conclusion, and there seems no justification for considering at length the significance of observations that were almost certainly the result of faulty technique.

**Chemical Structure of Acid-fast Bacilli.**—Largely owing to the work of Anderson and his colleagues (1927 *et seq.*) at Yale University, valuable information has been obtained in recent years on the chemical structure of the mycobacteria. Large quantities of bacilli of different types, grown on Long's (1926) synthetic medium, were extracted with a mixture of alcohol and ether, and the resulting extract was treated with chloroform and with acetone. In this way the lipid material was separated into three fractions, consisting of glycerides, phosphatides, and wax. The alcohol-ether extract also contained a considerable amount of polysaccharide and some basic compounds that could be precipitated by  $\text{HgCl}_2$  and by phosphotungstic acid. The results of the fractionations are given in Table 25.

TABLE 25

PERCENTAGE FRACTIONS OF LIPID AND OTHER MATERIAL ISOLATED FROM ALCOHOL-ETHER AND CHLOROFORM EXTRACTS OF ACID-FAST BACILLI (Chargaff, Pangborn and Anderson 1931)

	Type of Organism			
	Human tubercle bacillus H 37	Avian tubercle bacillus	Bovine tubercle bacillus	Timothy Grass bacillus
Phosphatide	6.54	2.20	1.53	0.59
Acetone-soluble fat	6.20	2.19	3.34	2.75
Chloroform-soluble wax	11.03	10.79	8.62	4.98
Total lipins	23.78	15.26	13.40	8.37
Polysaccharide	0.87	1.02	1.09	3.90
Dried bacterial residue	75.01	83.71	85.50	87.70

It will be observed that the total lipin content was highest in the human type of bacillus and lowest in the saprophytic acid fast bacillus. The polysaccharide content, on the other hand, was arranged in the reverse order. Further analysis showed that the phospholipins contained saturated and unsaturated fatty acids and glycerophosphoric acid, moreover, on hydrolysis they yielded large amounts of water-soluble carbohydrates, of which mannose and inositol seemed to be the two most important. Besides palmitic, linoleic, and linolenic acids, there were two fatty acids of special interest. One, which was optically active and isomeric with cerotic acid, was termed phthioic acid, the other, which was optically inactive and isomeric with stearic acid, was termed tuberculostearic acid. Of the waxy material, one portion was purified and found to be a white powder melting at  $200^{\circ}\text{--}205^{\circ}\text{C}$ , the remainder formed a yellowish salve-like mass which was called "soft wax." The purified wax yielded on hydrolysis about 56 per cent of unsaponifiable wax, this corresponded to the higher alcohols of previous workers, and proved to be acid fast. The purified wax also contained polysaccharides which on hydrolysis yielded a number of sugars including mannose, *d*-arabinose and galactose (see also Gough 1932). The "soft wax" appeared to be a complex glyceride. From the acetone-soluble fat of the human tubercle bacillus a yellow pigment was isolated. This pigment, to which the name phthiocol has been given, is one of the hydroxynaphthaquinones, and is the oxidant of a reversible oxidation.

reduction system whose  $E'_0$  is among the lowest reported for systems of biological origin (Ball 1934). The polysaccharides in the ether extract were apparently different from those in the phosphatide or wax.

A nitrogen free and phosphorus free polysaccharide was prepared by Hooper Renfrew and Johnson (1934) by alkaline hydrolysis of the acetol product of the crude carbohydrate from a protein free ultrafiltrate. The molecular weight of the pure polysaccharide has been estimated by Seibert Pedersen and Tiselius (1938) at 9 000.

Besides the lipid material and the polysaccharides acid fast bacilli contain proteins that are soluble in water. From cultures of tubercle bacilli on synthetic media Long and Seibert and their colleagues (1926-1928) isolated various proteins of which one appears to be the active principle of tuberculin. In this country Gough (1933) obtained evidence of the presence in cultures of human tubercle bacilli of two proteins having different chemical and immunological characters. The later work of Menzel and Heidelberger (1933) who fractionated the residue of frozen and dried bacilli after extraction in the cold with acetone and ether, revealed the great complexity of the proteins in the acid fast bacilli. Differences were noted between the proteins of saprophytic acid fast bacilli, avian bacilli and mammalian bacilli. At least three antigenic components were found among the proteins separated from bacilli of the human type. The protein fraction PPD b3 which appears to be responsible for the tuberculin reaction was found by Seibert Pedersen and Tiselius (1938) to have a molecular weight of 16 000 and to contain 4.1 per cent. of polysaccharide and 3 per cent. of nucleic acid. Further purification by Seibert and Glenn (1941) resulted in the reduction of the nucleic acid to 1.2 per cent. and of the molecular weight to 10 500.

The cellular reactions to the various fractions extracted from tubercle bacilli have been studied for many years by Sabin and her colleagues (for references see Sabin 1938 and Sabin and Joyner 1938). They may be summarized briefly as follows:

Four types of reaction have been noted: (a) exudation of neutrophils from the blood vessels to the tissues; (b) stimulation of the phagocytic mononuclear cells of the tissues; (c) multiplication of fibroblasts; and (d) local increase in lymphocytes. The polysaccharide when introduced into the tissues of suitable animals called forth neutrophils from the blood vessels but had no further action. The lipins, in addition to evoking this reaction, stimulated the phagocytic mononuclear cells of the tissues. After injection of the phosphatide fraction tubercles developed consisting of epithelioid cells and of their multinucleated derivative the Langhans giant cell. Caseation sometimes followed. The waxes, higher alcohols, and hydroxy-acids led to a multiplication of monocytes and their fusion into giant cells of the foreign body type. The reaction to the proteins was more complex. They gave rise not to one type of the phagocytic mononuclear cell, but to every type. They induced the formation of monocytes, of stimulated monocytes of macrophages of epithelioid cells singly and in tubercles, and of giant cells, both of the Langhans and the foreign body type. The degree of complexity of the reaction was least with the soluble proteins and greatest with the insoluble. A consistent increase in fibroblasts was found only after the injection of a wax like fraction isolated from a saprophytic acid fast organism. The new formation of lymphocytes was observed irregularly after injection of the phosphatide and protein fractions. Reactions in general were greater in tuberculous than in normal animals.

The polysaccharide is non-toxic on intravenous injection into rabbits (Sabin *et al.* 1931) but like the protein it may play a part in the phenomenon of allergy. It reacts

with a precipitating serum though it is incapable of calling forth the production of antibodies (Laidlaw and Dudley 1925 Mueller 1926). The purest protein fractions appear to be practically non antigenic, the fractions with larger molecular weight give rise on injection to antibodies (for reviews see Wells and Long 1932 Anderson 1932, Sabin 1932 Calmette 1936 Seibert 1941 1944).

### Cultural Reactions

The acid fast bacilli vary in the ease with which they grow under artificial conditions. At one end of the scale are the saprophytic acid fast bacilli and the cold blooded tubercle bacilli, which grow well in 2 or 3 days on ordinary media. At the other end are the mammalian tubercle bacilli, which grow only on special media, and which may take 2 weeks or more to form a layer of growth visible to the naked eye, in between come the avian bacilli, which grow poorly or not at all on ordinary media, but which give a good growth in a few days on glycerinated media. The most satisfactory media for cultivation of the tubercle bacilli are inspissated serum, coagulated egg and potato. The addition of 5 per cent glycerine to these media is of great value enhancing the growth of all acid fast organisms with the exception of the murine and partial exception of the bovine tubercle bacillus. Its incorporation in nutrient agar or in broth renders these media suitable for the growth of human and bovine bacilli and, except for the murine type, greatly improves the growth of other members of the group.

One of the striking characteristics of the acid fast group is the friable tenacious consistency of the growth, and its adhesiveness to the medium. This is evident not only in the process of subculturing the organisms when a stout platinum loop has to be used but particularly in endeavouring to form a suspension of the organisms in saline or other fluid. Instead of giving rise to a uniform turbidity they settle in granules to the bottom and leave the supernatant fluid clear, to produce a homogeneous suspension they must be ground up thoroughly in an agate mortar—a process that may take anything from half an hour to several days to complete. Similarly, it is difficult to make a uniform film of a culture for microscopic examination, even with a stout platinum loop it is impossible to break up the growth completely, and the film remains granular. As a rule the most difficult growths to emulsify are those of the human and bovine types. Growth of the other types usually present less difficulty. Sometimes a creamy almost butyrous, growth is formed by members of the avian and cold blooded types, which can be emulsified rapidly, but even with these suspensions the disintegration of the bacillary clumps is rarely complete, against an illuminated dark ground a fine granularity is visible to the naked eye.

Another feature that is common to all the tubercle bacilli, though not to the saprophytic acid fast bacilli, is the rather pleasant, sweet fruity odour of their cultures. This odour, so far as we know, is peculiar to the tubercle bacilli, it is developed on all the usual media in which growth occurs both solid and liquid, and is most readily detected in tubes that have been corked during incubation.

In studying the cultural reactions of the acid fast bacilli it is important to realize that, on first isolation the reactions of a given organism may seem to be anomalous. It is often not till two or three generations, and sometimes more, have been spent in artificial media that the characteristic reactions of the type develop. Thus a cold blooded bacillus when first isolated may grow very poorly—almost like a human strain—but in 3 or 6 months in the laboratory it will become accustomed to its new surroundings and instead of giving a poor discrete growth

on glycerine egg in 4 weeks it will give a profuse confluent growth in 5 or 6 days

**Cultural Differentiation between the Human and Bovine Tubercle Bacilli.**—For differentiating between the human and bovine types of tubercle bacilli it is desirable to seed a number of different media—serum glycerine serum, egg glycerine egg glycerine agar glycerine potato and glycerine broth—with the organisms to be tested preferably as soon after their isolation as possible. To obtain the maximum differentiation the organisms used for seeding should never have been grown on a medium containing glycerine. It is advisable to inoculate two or three tubes of each medium and to seal the tubes so as to prevent evaporation. For examining the growth in glycerine broth flasks are most suitable so that a large surface of

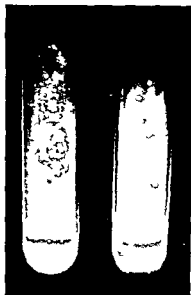


FIG 0—*Myco tuberculosis*

Human type Left Glycerine serum, 14 days, 3 °C Right Plain serum 14 days, 3 °C showing beneficial effect on growth of addition of glycerine.

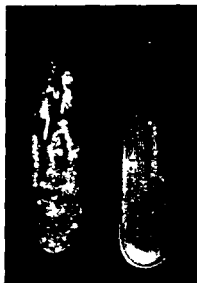


FIG 71—*Myco tuberculosis*

Left Humantype Glycerine agar 3 weeks, 3 °C. Right Bovine type Glycerine serum, 3 weeks, 3 °C.

medium is exposed to the air. Rapid methods of differentiating tubercle bacilli are to be regarded with caution. The method for example described by Jensen (1932) which claims to yield a result within 4 weeks is open to several objections. Though possibly giving a high proportion of correct answers in experienced hands, it will not commend itself to those who realize how much bacteriology has suffered in the past from the use of faulty or inadequate technique.

On all media the growth of the human type is greater in amount than that of the bovine type for this reason the human type is called eugonic the bovine type dysgonic. On media containing no glycerine such as coagulated serum or egg the superior growth of the human bacillus is barely evident but on media containing glycerine it is most striking. The effect of glycerine on growth is one of the most important of the differential characteristics, glycerine favours the growth

of the human type, but has little or no effect on that of the bovine type (Figs 70 and 71). Thus on glycerine agar the human bacillus gives a thick confluent wrinkled growth, while the bovine bacillus gives only a poor, effuse ground glass growth or may fail to grow at all. On glycerine potato again the human bacillus gives a raised confluent, wrinkled or warty growth, while the bovine bacillus gives a poor, often discrete, effuse growth, and similarly with the other media.

Another point of distinction is that the human type often forms a yellowish or orange pigment, whereas the bovine type never does. The pigment is noticeable only on batches of serum that have a rich yellow colour. For this reason the serum should come from an old cow, pale serum from a young cow, or from another species of animal, is useless. On a golden yellow serum the human type often produces a rich yellow or orange yellow growth, the bovine type gives a non-pigmented growth. On a serum coloured only slightly yellow the growth of the human type is cream or light yellow in colour. The differentiation of the human and bovine types may take months to complete, often when the organisms are first isolated they grow very poorly, and it is not till they have become accustomed to saprophytic conditions that they give the best growths of which they are capable.

**Cultural Differentiation of the Murine from the Human and Bovine Bacilli.**—The murine type as exemplified by the vole bacillus, grows much more slowly than either of the other two mammalian types, taking four weeks or more to form colonies visible to the naked eye on egg medium. Little or no growth occurs in primary culture on media containing glycerine, and even in subculture growth is poor. Unlike the other two types, the murine bacillus grows in plain trypticized broth producing film-like colonies on the surface and a fine deposit. It grows likewise on potato without glycerine.

**Cultural Differentiation between the Human and Avian Tubercle Bacilli.**—When first isolated from lesions in birds, the avian bacillus may closely resemble the human bacillus but as a rule after a few generations in the laboratory it takes on a more rapid and luxuriant growth, so that differentiation on cultural grounds alone is rendered possible. There are certain avian strains, however, that remain



FIG 72.—*Myco tuberculosis*

Human type. Glycerine broth 6 weeks 37°C showing thick wrinkled surface growth spreading up the sides of the flask.

permanently like the human type, on solid media these strains are indistinguishable from human strains, but differentiation is generally possible by the growth in glycerine broth, in this medium the human bacillus forms a thick wrinkled surface pellicle (Fig 72), whilst the avian bacillus grows at the bottom of the flask forming a granular deposit, or sometimes spreading out in a veil like manner over the bottom, and part way up the sides of the flask. Occasionally the avian bacilli give rise to a diffuse turbidity in broth. Why it is that some bacilli grow as a pellicle on the surface, others form a veil over the bottom, while still others grow diffusely, is not known. It may depend on the oxygen pressure most suitable for growth, or, as some workers think, on the surface tension of the medium. By lowering the surface tension to below 42 dynes, Larson (1926) states that it is possible to induce organisms, which usually form a surface pellicle, to grow diffusely or at the bottom of the flask.

Most avian strains grow more rapidly and more profusely than human strains, and tend on solid media to give a more creamy, homogeneous, and less granular growth than that of the human bacillus (Fig 73). Though growth is favoured by glycerine, it is possible to get the avian bacillus to grow on simple media like nutrient agar or broth without the addition of any glycerine, this is more difficult with the human bacillus. Another point of difference is that the human strains do not grow at a temperature above 40° C, whereas the avian strains will grow up to 43° or 45° C. A point of not much importance is that avian cultures generally live longer than human ones, they may be found viable after 1 or even 2 years, human cultures are often dead in a couple of months, though occasionally they may survive for much longer. Another point of small importance is that a few avian strains when grown on glycerine egg medium give a faint pink-coloured growth, human bacilli never give a pink coloration.

**Cultural Differentiation between the Avian and Cold-blooded Tubercle Bacilli.**—In culture the avian and the cold blooded bacilli resemble each other closely. On first isolation they can easily be distinguished by the difference in their optimum temperatures of growth, the avian bacillus growing best at about 40° C, the cold blooded at 25° C. After prolonged subculture in the laboratory this difference is partly obliterated, the cold blooded bacillus comes to grow quite well at 37° C., though the avian bacillus will rarely grow below 30° C. Practical differentiation can therefore be made by growing the organisms at 25° C, if growth is as good at this temperature as at 37° C, the strain is a cold blooded one, if growth occurs freely at 37° C, but fails to occur at 25° C, the strain is an avian one. Apart from the differences in optimum temperature there are no definite cultural characteristics distinguishing the two types, except the pink coloration of certain avian strains when grown on glycerine egg. Minor differences such as the worm-cast growth of the cold blooded type on glycerine potato, and the denser nature of the growth on the bottom of flasks of glycerine broth are not sufficiently constant to be reliable. As a rule the cold blooded bacillus grows more rapidly than the avian bacillus, cold blooded strains often give a definite layer of growth in 2 or 3 days, growth of avian strains is often not visible for 4 or 5 days.

**Cultural Characteristics of the Saprophytic Acid-fast Bacilli.**—These organisms grow rapidly at room temperature giving rise in 2 or 3 days to a profuse confluent growth. Different strains vary in their cultural reactions, but on the whole their appearance on solid media is very characteristic. On glycerine agar there is a



luminant, raised dry growth yellow, pink or brick red in colour, with a coarsely granular surface, resembling dry bread crumbs (Figs 74 and 75). This growth is unlike that of any of the tubercle bacilli. A few strains however, are non pigmented. In glycerine broth some strains give a turbidity, but most of them do not, there is generally a surface growth which may be thick and wrinkled like the human tubercle bacillus thick and coarsely granular or more thin and delicate a deposit of granular material is usual. Growth occurs in gelatin, but no liquefaction is produced. The optimum temperature of growth varies, but is generally in the neighbourhood of  $37^{\circ}\text{C}$ . All strains grow freely at room temperature several grow at  $45^{\circ}\text{C}$ , while a few multiply even at  $55^{\circ}\text{C}$  (see Schwabacher 1933b).

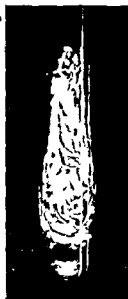


FIG 73 — *Myco tuberculosis*  
Avian type Glycerine  
agar 3 weeks  $37^{\circ}\text{C}$ ,  
showing abundant growth  
of butter cream type



FIG 74 — *Myco stercorea*  
(Mist bacillus)  
Glycerine agar 3 weeks  
 $37^{\circ}\text{C}$  showing abundant  
heaped up growth

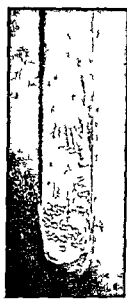


FIG 75 — *Myco phlei* I  
Glycerine agar 3 weeks  
 $37^{\circ}\text{C}$  showing abundant  
growth with wrinkled  
surface

**Variation**—The phenomenon of variation in acid fast bacilli has occupied the attention of large numbers of workers in recent years. The field is a wide one but so far the results of exploring it have not been fruitful. Many contributions to its study have been purely descriptive, and have been made without any apparent realization of its biological significance. The terms used by different workers to denote the variants they have observed have differed so widely that the nomenclature is in a state of the utmost confusion. Changes in virulence have been postulated on inadequate grounds and attempts to measure small differences in virulence have often been confidently performed with numbers of animals quite insufficient to provide a definite answer.

One of the chief difficulties in work of this type lies in distinguishing between fixed and environmental variants. Our own experience suggests that a great many of the so called variants are nothing more than adaptations to

changed conditions. If, for example, a suspension of tubercle bacilli is inoculated simultaneously in equal quantities on to a number of different media, quite a variety of colonial types will develop (Fig 76). If the different types are picked off and inoculated all on to the same medium, complete similarity of colonial appearance will often result, indicating that no fundamental biological change has been effected. What degree of fixity variant types may have it is very difficult to ascertain, and we believe that before further progress is possible, a very careful study of the limits of transient environmental variation will have to be made. Without expressing any opinion on the degree of fixity of types or of giving a critical review of the extensive bibliography that has grown up on the subject of variation, we shall confine ourselves to describing two main types possessing correlated morphological and cultural appearances.



FIG 76—*Myco tuberculosis*—bovine type.

Surface colonies on three different media, inoculated at the same time from the same suspension and incubated under identical conditions. 37 days, 37° C. The figure illustrates the dependence of the colonial form on the nature of the medium.

Left: Modified Dorset egg medium. Middle: Egg yolk medium. Right: Egg yolk agar medium.

On Dorset or 5 per cent glycerol egg medium many cultures of tubercle and saprophytic acid fast bacilli produce colonies that are either smooth or rough. In stroke cultures the smooth forms give rise to a moist butyrous growth with a smooth glistening surface, while the rough forms produce a dry, rather friable growth, with a rough dull, and often heaped up surface, looking not unlike dry bread crumbs (Fig 77). Rubbed up in water the smooth growth yields a fairly homogeneous suspension, the rough growth a granular suspension. Single colonies of the smooth form tend to be circular, convex, with a generally smooth surface and an entire edge, rough colonies are irregular in outline, are often

heaped up and convoluted have a roughish granular surface and tend to be surrounded by a spreading veil like peripheral extension (Fig 78). In glycerol broth the smooth forms may grow diffusely throughout the medium they may form a thin smooth surface pellicle or they may cover the bottom of the flask with a reticulated veil like growth the rough forms usually grow as a thick wrinkled surface pellicle unaccompanied by turbidity. Morphologically smooth strains consist of fairly long curved slender sometimes beaded bacilli lying more or less parallel to one another and occurring in bundles (Fig 79), less often they are rather short stout or ovoid bacilli staining evenly and arranged singly or in groups (Fig 80). Rough strains on the other hand consist of rather short sometimes ovoid bacilli or coccobacilli arranged in Chinese letter forms and in dense masses (Fig 81). This morphological difference in the arrangement of the smooth and rough types is similar to that described by Nutt (1927) and Wilson (1930) for *Salm typhimurium* and by Soule (1928) for *B anthracis* and *B subtilis*. It has been well pictured by Schwabacher (1933b) and Wyckoff (1931) for saprophytic acid fast and cold blooded tubercle bacilli respectively. It depends essentially on the mode of division. The bacilli of the smooth type separate completely after division and slip past each other so as to come to lie in parallel. The bacilli of the rough type exhibit an angular division the organisms not separating completely but coming to lie at an obtuse angle to each other resembling a green stick fracture or forming long tangled masses which become heaped up and convoluted as growth continues. It is hardly necessary to add that many intermediate forms occur partaking of some smooth and some rough characteristics.

Whether the smooth and rough types differ metabolically antigenically and in virulence it is impossible to say definitely at present. Many workers assert that the smooth type of tubercle bacillus is more virulent than the rough but further evidence is desirable on this point.

(References: Petroff 1927 Petroff Branch and Steenken 1927a b Petroff and Steenken 1930 1935 Kraus and Gerlach 1929 Uhlenhuth and Seiffert 1930 Begbie 1930 1931 Dreyer and Vollum 1931 Reed and Rice 1931a b Rice and Reed 1931 Rice 1931 Toda 1931 Seiffert 1932 Schwabacher 1933b Seibert Long and Morley 1933 Wyckoff 1934 Meisner and Prausnitz 1934 Steenken *et al* 1934 Birkhaug 1935 Denys 1935 Shaffer 1935 Smithburn 1935.)

**Resistance**—Acid fast bacilli possess much the same degree of susceptibility to heat as other non sporing bacteria but a rather higher degree of resistance to chemical disinfectants. This behaviour is probably related to their content of



FIG 77—Saprophytic acid fast bacilli.  
Left smooth strain showing raised confluent butyrous growth, with smooth glistening surface.  
Right rough strain, showing raised confluent friable heaped up growth with dry dull rough surface.  
Glycerol egg 2 months 37° C.

waxy substances, which are less permeable to cold than to warm aqueous solutions. There is evidence that their resistance is more or less proportional to the amount

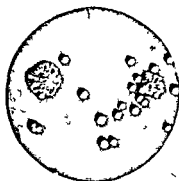


FIG 78 —*Myco tuberculosis*—avian type

Surface colonies of smooth and rough forms. Petraghani medium, 7 weeks, 37° C.



FIG 79 —Glycerine egg 1 month, 37° C, showing long curved bacilli arranged in bundles. Smooth type of morphology

of waxy material present (Shen 1934). Use is made of this differential susceptibility in isolating tubercle bacilli from contaminated material. Treatment of sputum, pus, or other products, with 15 per cent antiformin or 15 per cent sulphuric acid



FIG 80 —Glycerine egg 1 month 37° C, showing fat ovoid forms arranged individually and in groups. Smooth type of morphology



FIG 81 —Glycerine egg 1 month, 37° C, showing mostly short irregularly stained rods and coccobacilli arranged singly, in Chinese letter forms and in dense clumps. Rough type of morphology

for 5 to 20 minutes is often sufficient to destroy contaminating non-sporing bacteria without killing the tubercle bacilli, which can then be cultivated on suitable media. For the effect of relatively concentrated solutions of disinfectants reference may

be made to Calmette (1936) With regard to weaker solutions Douglas and Hartley (1934) found that a saline suspension containing 1 mgm of moist bovine bacilli per ml was sterilized within 24 hours by exposure at room temperature to 0.5 per cent phenol or 0.02 per cent merthiolate Phenol in 0.25 per cent concentration failed to sterilize the suspension in 14 days, while 0.1 per cent formal apparently just failed to do so In our own experience, using the cultural instead of the animal inoculation method, 0.5 per cent phenol cannot be relied on to destroy all tubercle bacilli in 24 hours In milk tubercle bacilli are killed in 20 minutes at 60° C, provided it is contained in a closed vessel (for references, see Wilson 1942) In an open vessel a pellicle forms on the surface which protects the organisms to some extent, so that a few bacilli may escape destruction for an hour (Smith 1899 Oerskov 1925, Meanwell 1927) In polluted water kept in the dark at room temperature tubercle bacilli may remain alive for at least 3 months (Rhines 1935) while in soil or cow dung exposed on pasture land during the summer and autumn in this country they may remain alive and virulent for 2 to 6 months (Williams and Hoy 1930, Maddock 1933) The organisms are comparatively resistant to drying, and provided they are protected from sunlight they may survive for months under suitable conditions In books contaminated with the sputum of tuberculous patients, they may remain viable from 2 weeks to 3½ months (see Smith, C R, 1942b) Numerous experiments have been performed to test the action of light on tubercle bacilli (Rochaix and Cohn 1911, Mayer 1921 Caldwell 1925, Eidinow 1927, Mayer and Dworski 1932, Smith, C R, 1942a) Most workers have found that they are rapidly destroyed, if spread in a thin layer by bright sunlight, or by ultra violet rays from a mercury vapour lamp The most effective appear to be the short ultra violet rays Thus bacilli suspended in saline when exposed in quartz flasks to rays of 2300-7620 Å U, were killed in 10 minutes those exposed through window glass to rays of 3300-5720 Å U were not completely killed even in 1 hour Probably the rays shorter than 3300 Å U are the most lethal (Eidinow 1927) Blood, serum, and other proteins protect the bacilli against ultra violet light

Growth of tubercle bacilli in glycerine broth is inhibited by the addition of very small quantities of certain aniline dyes, thus a concentration of 0.0004 per cent thioflavine or 0.0002 per cent methylene blue entirely prevents growth, smaller quantities have little or no effect Thymol is active in a concentration of 0.004 per cent, most metals in colloidal suspension, however, are inactive even in a concentration of 10 per cent (Karwacki and Biernacki 1925)

**Metabolism**—The optimum hydrogen ion concentration for growth in 4 per cent glycerine broth is said by Ishimori (1924) to lie between the following points

Human type of tubercle bacillus	pH 7.4-8.0
Bovine " " "	pH 5.8-6.9
Cold blooded " " "	pH 6.2-7.7
Rabinowitsch's butter bacillus	pH 5.7-8.5
Timothy grass bacillus	pH 7.5-9.1
Moeller's grass bacillus	pH 7.4-7.7

Dernby and Näslund (1922) found that growth of human and bovine tubercle bacilli in 3 per cent glycerine veal broth occurred between pH 4.5 and 8.0, the optimum being pH 6.0-6.5, their figures, it will be seen, do not agree with those

of Ishimori, who found that the human preferred a more alkaline reaction than the bovine bacillus.

The optimum temperature for growth of the human, bovine, murine, and, generally the saprophytic acid fast bacilli, is  $37^{\circ}\text{C}$ ; for the avian  $40^{\circ}\text{C}$ , and for the cold blooded bacillus  $25^{\circ}\text{C}$ . The human, bovine, and avian types do not grow below  $30^{\circ}\text{C}$ , the cold blooded and saprophytic acid fast types grow freely at  $20^{\circ}\text{C}$ . Many saprophytic acid fast bacilli grow at  $45^{\circ}\text{C}$ , and a few at  $55^{\circ}\text{C}$ .

The tubercle bacillus is an aerobe, it will not grow under strictly anaerobic conditions. The optimum partial pressure of oxygen is said to be 40-50 per cent for the human type (Novy and Soule 1925) and 60-70 per cent for the avian and saprophytic acid fast types (Uga 1935). Novy and Soule in their very careful study of the respiration of the tubercle bacillus found that  $\text{CO}_2$  had little effect on growth, and no inhibition occurred till a partial pressure of 60 per cent was reached. Subsequent observations, however, by Rockwell and Highberger (1926) and others have shown that  $\text{CO}_2$  is beneficial for growth. For optimal development it is desirable to incubate cultures in a high partial pressure of oxygen and about 10 per cent  $\text{CO}_2$ . A few measurements have been made on the oxygen uptake of the bacilli (Dieckmann and Menzel 1932), and on the oxidation reduction potential established in phosphate buffer solutions (Aksmanzew 1933).

Moisture is an essential requirement of the tubercle bacillus *in vitro*. For good growth to occur, plenty of condensation water—supplied best by passing steam into the tube before inoculation—and an abundance of air are essential. Growth does occur in sealed tubes provided plenty of moisture is present, but it ceases after 3 or 4 weeks.

The effect of glycerine on the growth of the tubercle bacillus has been the subject of much controversy. Nocard and Roux in 1887, working apparently with an avian strain, were the first to notice the beneficial action of this substance. Since then it has been found that the addition of glycerine, generally in a concentration of 5 per cent, greatly increases the growth of all acid fast bacilli with the exception of the murine, and to some extent the bovine tubercle bacillus. On this organism glycerine is not without effect, for it will enable it to grow—though very poorly—on agar, potato, or broth, on which no growth otherwise occurs. But the favourable action of glycerine on the bovine bacillus is not to be compared with that on other types, the addition of glycerine, for example, to serum or egg medium, makes no difference to the growth of the bovine bacillus, whereas it greatly increases the growth of all the other acid fast bacilli.

The difference in the effect of glycerine on growth was made use of by Smith (1904-05) in the elaboration of a test for distinguishing between the human and bovine types. He found that if bovine tubercle bacilli were grown in glycerine broth, which had an acid reaction to phenol phthalein, the acidity gradually decreased, till after full growth had occurred the reaction was about neutral, in cultures of human bacilli, on the other hand, after an initial production of alkali, the reaction gradually became acid again. That is to say, the final reaction of a bovine culture was about neutral of a human culture decidedly acid. This difference in the reaction curve of the two types of mammalian bacilli has been investigated by numerous workers, some of whom have confirmed, and some of whom have not confirmed, Smith's observations. Undoubtedly one of the reasons for this discrepancy is that most of the work was done before the days of accurate determination of acidity by measurement of the  $\text{H}^+$  ion concentration was possible. Smith's view, put briefly, was that the human bacillus was able to utilize glycerine with the con-

sequent production of acid whereas the bovine bacillus was unable to use it and so caused an increased hydrolysis of protein and an alkaline reaction. Whether Smith's view is correct or not, it is still impossible to say. The reaction curve of broth cultures is determined by so many factors, such as the production of  $\text{NH}_3$ ,  $\text{CO}_2$ , amino-acids and other substances, some of which are volatile, and some of which are not, that it is almost impossible to determine whether the production of acid is due to disintegration of the glycerine by the bacilli, or to hydrolysis of the protein material. Harden (1913) who made a special investigation of this subject, failed to obtain evidence of any definite distinction in biochemical behaviour between the human and the bovine types. Kendall, Day and Walker (1920), who have examined the question more recently, conclude that glycerine probably exerts a sparing action on the protein constituents of the medium in cultures of the human, but not in cultures of the bovine type, but as noted elsewhere (p. 57) the existence of this protein sparing effect is very doubtful. It is not possible to discuss this problem more fully (for a full discussion of the work up to 1917 see Cobbett 1917) but we may conclude that as a practical means of differentiating the human from the bovine bacillus determination of the glycerine broth reaction curve is too complex for routine use.

Glucose seems to act in much the same way as glycerol (see Kauffmann 1932). The growth of tubercle bacilli is likewise improved by the addition of extracts of acid fast bacilli to the medium, certain organisms, such as *Johnes bacillus* will not grow at all unless such an extract is added (see p. 441). The tubercle bacillus has been grown in a number of synthetic media (see Wells and Long 1932) its growth is said to be increased by the addition of substances containing vitamin B such as yeast, and of orange tomato or cabbage juice (Uyei 1927) (but see p. 66). Schmidt (1925) found 0.01 per cent of iron chloride to favour growth but Davies (1937) had the opposite experience. The effect of blood seems to be complex: tubercle bacilli grow well in liquid blood particularly if lysed by saponin (Pryce 1941), but are generally inhibited by blood on the surface of a solid medium (Evans and Hanks 1939). Old potatoes that blacken on standing are more favourable for growth than new ones that do not (McCarter and Tatum 1937). Saprophytic acid fast bacilli are said to grow in a simple solution of inorganic salts containing neither carbon nor nitrogen (Bruner 1931; Gordon and Hagan 1937). There is evidence that tubercle bacilli can oxidize certain fats, such as those of olive oil and butter (Sedyeh and Selber 1927).

*Pigment production is very variable.* On deeply coloured batches of ox serum the human bacillus often forms a rich yellow or orange yellow growth. A pink coloration on glycerol egg medium is not uncommon with avian strains and according to Blacklock (1932) is sometimes given by rapidly growing bovine strains. Cold blooded bacilli generally give colourless growths. On the other hand, the growth of nearly all the saprophytic acid fast bacilli is accompanied by the production of a yellow, pink, orange, or brick red pigment. On Sauton's medium a green coloration is not infrequently observed with cultures of mycobacteria: its intensity varies from strain to strain (Lange 1932, de Grohier 1933, Kraus and Koref 1933). Pigment formation seems to depend on a number of different environmental factors particularly the composition of the medium and the oxygen supply. Reed and Rice (1929) found that the addition of iron favoured its formation. On a glycerol agar medium containing 0.02 per cent of ferric citrate human, bovine, and avian types of tubercle bacilli, and saprophytic acid fast bacilli all formed pigment of varying depth, whereas on the same medium without iron many strains formed no pigment at all. Pigment production is not well developed at  $37^\circ \text{C}$ , it is best seen in

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The optimum temperature for growth of the human, bovine, murine, and, generally the saprophytic acid fast bacilli, is 37° C, for the avian 40° C, and for the cold blooded bacillus 25° C. The human, bovine, and avian types do not grow below 30° C, the cold blooded and saprophytic acid fast types grow freely at 20° C. Many saprophytic acid fast bacilli grow at 45° C, and a few at 55° C.

The tubercle bacillus is an aerobe, it will not grow under strictly anaerobic conditions. The optimum partial pressure of oxygen is said to be 40–50 per cent for the human type (Novy and Soule 1925) and 60–70 per cent for the avian and saprophytic acid fast types (Uga 1935). Novy and Soule in their very careful study of the respiration of the tubercle bacillus found that CO<sub>2</sub> had little effect on growth, and no inhibition occurred till a partial pressure of 60 per cent was reached. Subsequent observations, however, by Rockwell and Highberger (1926) and others have shown that CO<sub>2</sub> is beneficial for growth. For optimal development it is desirable to incubate cultures in a high partial pressure of oxygen and about 10 per cent CO<sub>2</sub>. A few measurements have been made on the oxygen uptake of the bacilli (Dieckmann and Menzel 1932), and on the oxidation reduction potential established in phosphate buffer solutions (Aksianzew 1933).

Moisture is an essential requirement of the tubercle bacillus *in vitro*. For good growth to occur plenty of condensation water—supplied best by passing steam into the tube before inoculation—and an abundance of air are essential. Growth does occur in sealed tubes provided plenty of moisture is present, but it ceases after 3 or 4 weeks.

The effect of glycerine on the growth of the tubercle bacillus has been the subject of much controversy. Nocard and Roux in 1887, working apparently with an avian strain, were the first to notice the beneficial action of this substance. Since then it has been found that the addition of glycerine, generally in a concentration of 5 per cent, greatly increases the growth of all acid fast bacilli with the exception of the murine and to some extent the bovine tubercle bacillus. On this organism glycerine is not without effect, for it will enable it to grow—though very poorly—on agar, potato or broth on which no growth otherwise occurs. But the favourable action of glycerine on the bovine bacillus is not to be compared with that on other types, the addition of glycerine, for example, to serum or egg medium, makes no difference to the growth of the bovine bacillus, whereas it greatly increases the growth of all the other acid fast bacilli.

The difference in the effect of glycerine on growth was made use of by Smith (1904–05) in the elaboration of a test for distinguishing between the human and bovine types. He found that if bovine tubercle bacilli were grown in glycerine broth which had an acid reaction to phenol phthalein the acidity gradually decreased till after full growth had occurred the reaction was about neutral, in cultures of human bacilli on the other hand after an initial production of alkali, the reaction gradually became acid again. That is to say the final reaction of a bovine culture was about neutral of a human culture decidedly acid. This difference in the reaction curve of the two types of mammalian bacilli has been investigated by numerous workers, some of whom have confirmed and some of whom have not confirmed Smith's observations. Undoubtedly one of the reasons for this discrepancy is that most of the work was done before the days of accurate determination of acidity by measurement of the H<sup>+</sup> ion concentration was possible. Smith's view, put briefly, was that the human bacillus was able to utilize glycerine with the con-



sequent product on of acid whereas the bovine bacillus was unable to use it and so caused an increased hydrolysis of protein and an alkaline reaction. Whether Smith's view is correct or not it is still impossible to say. The reaction curve of broth cultures is determined by so many factors, such as the production of  $\text{NH}_3$ ,  $\text{CO}_2$ , amino-acids and other substances some of which are volatile and some of which are not that it is almost impossible to determine whether the production of acid is due to disintegration of the glycerine by the bacilli or to hydrolysis of the protein material. Harden (1913) who made a special investigation of this subject, failed to obtain evidence of any definite distinction in biochemical behaviour between the human and the bovine types. Kendall Day and Walker (1930) who have examined the question more recently conclude that glycerine probably exerts a sparing action on the protein constituents of the medium in cultures of the human but not in cultures of the bovine type but as noted elsewhere (p. 57) the existence of this protein sparing effect is very doubtful. It is not possible to discuss this problem more fully (for a full discussion of the work up to 1917 see Cobbett 1917) but we may conclude that as a practical means of differentiating the human from the bovine bacillus determination of the glycerine broth reaction curve is too complex for routine use.

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cultures that have been incubated for some time at 34° C. and then transferred to a dark cupboard at room temperature. In this respect the acid fast bacilli resemble most other chromogenic bacteria.

The tubercle bacilli do not secrete a true exotoxin but endotoxins are liberated on autolysis of the bacilli in broth cultures. These appear to be of complex constitution and are more fully considered in the section on tuberculin (see Chapter 59). There is evidence that the endotoxins of all the acid fast bacilli are closely alike. Positive reactions may be obtained in animals infected with the mammalian type by injection of tuberculin made from the human, bovine, avian or cold blooded type of bacillus (Ledoux Lelard 1900 Wolbach and Ernst 1904). Intravenous injection of killed cultures of the mammalian, and especially of the avian tubercle bacilli is often fatal to rabbits. Twort and Craig (1913) state that the smegma, mist and timothy-grass bacilli are also toxic for rabbits, whereas the cold blooded tubercle and the butter bacilli are relatively non toxic.

**Biochemical Reactions.**—Very little work has been done on the biochemical reactions of the acid fast bacilli. Froun and Guillaume (1923a, b) brought evidence to show that human and avian bacilli could break down glucose, maltose, glycerol, and trehalose with the production of acid. Merrill (1930) who worked with one human, one avian, three cold blooded and several saprophytic acid fast strains and who made quantitative estimations of the sugar content of the medium before and after growth found that glucose was utilized by all the strains, arabinose by all but one, sucrose by some and lactose by none. The reaction however did not become acid nor did any acid cleavage products accumulate. Instead the medium became more and more alkaline during growth owing to the production of  $\text{NH}_3$ , though the rate at which it did so was less than in broth free from sugar. The conclusion was reached that the organisms oxidized the carbohydrates completely to  $\text{CO}_2$  and water.

Our own observations based on an incomplete study of 7 cold blooded and 36 saprophytic acid fast strains, showed that there was no detectable acid production in any of the usual sugar media. Litmus milk was turned alkaline by the saprophytic acid fast but usually acid by the cold blooded strains. In peptone water ammonia was produced in considerable quantity. Indole production was uniformly negative though Rabinowitch (1897) stated that it was sometimes positive. Catalase production was always positive.  $\text{H}_2\text{S}$  was formed in variable amount. The reduction of methylene blue in broth was weak or absent. The methyl red and Voges-Proskauer reactions were consistently negative.

In milk tubercle bacilli are able to grow but they produce in it no visible change.

**Antigenic Structure.** By agglutination, absorption of agglutinins and complement fixation the acid fast bacilli fall into four serological types—mammalian, avian, cold blooded, and saprophytic acid fast—the human and bovine types being indistinguishable (Tulloch *et al.* 1924 Cumming 1925 Wilson 1925 Griffith 1925 Furth 1926 Klopstock 1931 Kauffmann 1932). Direct agglutination is not sufficient to differentiate between the different types. Absorption of agglutinins is essential. By this means it appears that there is an antigen common to the human, bovine and avian types but that the avian type also possesses an antigen which is not present in the mammalian types (Wilson 1925). Grünberg (1935) has reached similar conclusions on the basis of the precipitin test. Wulff (1925) concluded that by the absorption of agglutinins technique human and bovine strains could be separated but his results still await confirmation. According to Wells (1944)

the murine type is serologically indistinguishable from the human and bovine types. Observations suggest that group specificity is determined by the polysaccharides, type specificity by the proteins (Seibert 1941). There is evidence that the avian type may be divided serologically into a small number of subtypes (Wulff 1925, Furth 1926, Schaefer 1937, Harpoth 1938). It is possible that the cold blooded and saprophytic acid fast types are heterogeneous but insufficient strains have as yet been examined to affirm this definitely.

It is noteworthy that serological methods are less valuable in classifying the tubercle bacilli than cultural methods: the bovine and human bacilli can be easily separated by their cultural reactions, yet serologically they form a homogeneous group. This affords an exception to the rule that serological examination is a far more delicate method of differentiation than cultural examination.

**Pathogenicity and Experimental Infection of Animals**—Under natural conditions the human tubercle bacillus gives rise to disease mainly in man, monkey, pigs and occasionally in dogs and parrots, the bovine bacillus to disease in cattle, pigs, horses, man, and occasionally dogs, cats and sheep, the murine bacillus to disease in voles and possibly other members of the family of *Muridae*, the avian bacillus to disease in birds, and occasionally in pigs, sheep and cattle, and the cold blooded bacillus to disease in cold blooded animals and fish. Saprophytic acid fast bacilli, though occasionally isolated from the tissues, rarely seem able to give rise to progressive disease (see also Chapter 59).

The virulence of tubercle bacilli is subject to variation. Though usually virulent on isolation, they not infrequently become more or less avirulent during subculture in the laboratory. Very little exact information, however, based on an adequate number of animal tests, is available about the difference in virulence of freshly isolated strains of the same type, or about the factors that are responsible for changes in virulence occurring *in vitro* or *in vivo*.

The experiments of Villemin, recorded in 1868, furnished convincing proof of the transmissibility of human tuberculosis to animals. Material taken from different types of tuberculosis in man—catarrhal pneumonic forms, caseous pulmonary and chronic pulmonary tuberculosis, disseminated tuberculosis, scrofula, sputum and the blood removed from a tuberculous patient after death—and introduced beneath the skin of rabbits, was successful in setting up the disease in 17 out of 21 animals, 3 of the 4 animals that did not contract tuberculosis died within a week of erysipelas. Villemin also transmitted bovine tuberculosis to rabbits, and made the important observation that material from bovine tuberculosis set up a more rapid and more generalized disease than material from human tuberculosis, when inoculated into these animals. Convincing though Villemin's experiments were, the final proof of the transmissibility of the disease was not furnished till Koch in 1882 succeeded in producing tuberculosis in animals by injection of pure cultures of the tubercle bacillus. In 1898 Smith brought evidence to show that bovine tubercle bacilli were more virulent when injected into rabbits and calves than human bacilli. This difference was substantiated by other workers, in particular by Vagedes (1898), Ravenel (1901), the English Royal Commission on Tuberculosis (Report 1911), Kossel, Weber and Henss (1904, 1905), and Park and Krumwiede (1910).

**CATTLE**—The subcutaneous injection of 50 mgm. of bovine bacilli from young serum cultures into calves sets up an acute, rapidly generalizing disease, proving fatal in about 6 weeks to 3 months. At necropsy the main features are a local lesion, focal glandular swelling and abscess formation, generalized glandular lesions, and lesions in the viscera.

most extensive in the lungs and spleen and less extensive in the liver and kidneys. Smaller doses give less constant results. The intravenous injection of even 1 mgm. of bovine bacilli produces a severe and fatal tuberculosis. Cattle may likewise be infected with bovine bacilli by inhalation and feeding.

The subcutaneous injection of calves with *human* bacilli in no matter what dosage never gives rise to a progressive disease. At most there is a local abscess and swelling of the focal glands when the animals are killed after 4 or 5 months, the lesions are minimal and are often calcified. The intravenous injection of human bacilli may cause death by toxæmia with acute infiltration and oedema of the lungs but there is no true infection. The same toxæmia may be produced by the intravenous injection of avian bacilli and even of the non pathogenic saprophytic acid fast bacilli.

The intramuscular inoculation of a calf with 80 mgm. of *murine* bacilli gave rise to a local abscess and to inconspicuous lesions in the regional lymphatic glands. Even 10 mgm. intravenously excited no more than a mild chronic retrogressive tuberculous reaction (Griffith and Dalling 1940).

The subcutaneous injection of calves with *avian* bacilli in a dose of 500 mgm. produces no more than a local lesion with perhaps a caseous nodule in the nearest lymphatic gland. Intravenous inoculation however of even 5 mgm. may prove fatal in 2 to 3 weeks. Post mortem there are milary tubercles in the lungs the spleen though showing no macroscopic tubercles is enlarged and contains great numbers of tubercle bacilli visible microscopically.

**GOATS**—*Bovine* bacilli inoculated subcutaneously in a dose of 1 mgm. give rise to a fatal generalized disease. *Human* bacilli, even in a dose of 100 mgm. cause no more than small retrogressive lesions. *Avian* bacilli in a dose of 100 mgm. rarely give rise to progressive disease on the other hand the animals may remain chronically infected and excrete the bacilli in the milk for a long time (Griffith 1931).

**SHEEP** resemble goats in being highly susceptible to infection with *bovine* bacilli moderately susceptible to infection with *avian* bacilli, and resistant to infection with *human* bacilli.

**PIGS**—Injected subcutaneously the *bovine* type in a dose of 10 to 50 mgm. produces rapidly fatal milary tuberculosis. The *human* type in a dose of 50 mgm. produces a local lesion with slight dissemination. *Avian* bacilli injected subcutaneously do not give rise to progressive disease but they may multiply and be disseminated through the internal organs, where they remain alive for some considerable time. Young pigs fed with bovine bacilli develop widespread disease and generally die when fed with human bacilli they may develop extensive glandular and pulmonary disease. Though the human bacilli cause less severe and less extensive tuberculosis than bacilli of the bovine type they are more pathogenic for pigs than they are for cattle.

**RABBITS**—*Bovine* bacilli injected subcutaneously in a dose of 0.1-1.0 mgm. produce a generalized disease fatal in 2 to 3 months. At necropsy a local caseous lesion caseous focal glands, innumerable little grey tubercles with caseating centres or larger irregular nodules in the lungs numerous projecting hemispherical nodules in the cortex of the kidneys, and tubercles in the spleen and liver are found. Given intraperitoneally even minute doses such as 0.000 000 001 mgm. of bovine bacilli are said to prove fatal in 2 to 3 months (Cobbett 1917). Intravenous injection of 0.01-0.1 mgm. of bovine bacilli proves fatal in 3 to 6 weeks.

The subcutaneous injection of *human* bacilli in a dose of 1-100 mgm. never causes fatal tuberculosis. Human bacilli may however give rise to lesions in the lungs and kidneys. Usually the lungs contain a few small grey tubercles or caseous nodules, and the kidneys show a few milary tubercles in the cortex. But the acute fatal milary tuberculosis seen after injection of bovine bacilli never results from the subcutaneous injection of human bacilli. Intravenous and intraperitoneal injections are less reliable for purposes of differentiation. 0.01 mgm. of human bacilli given intravenously rarely if ever proves fatal, but larger doses may give rise to a progressive fatal disease. Doses of 10-50 mgm. given

intraperitoneally may likewise prove fatal on occasion. Extensive lesions in the lungs and kidneys are often seen both after intravenous and intraperitoneal injection (see Cobbett 1932). The rabbit in fact possesses a considerable resistance to the human type of bacillus but not so great as the ox or the goat. Rabbits are also fairly resistant to infection with murine bacilli. Intravenous inoculation of 0.1 to 1.0 mgm may cause death from acute miliary tuberculosis. Smaller doses 0.01 mgm intravenously, or larger doses subcutaneously, produce trivial lesions and the bacilli gradually die out: an abscess in the lung however may provide a nidus in which the bacilli may live for a long time. The avian bacillus is less virulent for rabbits than the bovine but more virulent than the human bacillus. Subcutaneous injection gives rise to a chronic disease; intraperitoneal injection of large doses to a rapidly fatal peritonitis; and intravenous injection to an acutely fatal infection. The macroscopic lesions caused by the avian bacillus are much less obvious than those caused by the bovine bacillus, sometimes no tubercles are visible. After intravenous inoculation Yersin (1898) found the spleen greatly enlarged and the liver enlarged to a less extent: no macroscopic tubercles were seen but microscopically there were numerous small tuberculous nodules in the liver and spleen containing large numbers of tubercle bacilli, the kidneys and lungs appeared practically normal. Thus proliferation of the bacilli in the body without macroscopic tubercle formation is known as the Yersin type of disease, it is seen both in rabbits and guinea pigs injected with avian bacilli, and in rats injected with both mammalian and avian types. Chronic avian infections are generally characterized by infection of the joints (Griffith 1941c).

After injection of the bovine or human type of bacillus no matter what route is chosen the lesions in rabbits are most evident in the lungs and kidneys: the spleen and liver suffer less, the lymphatic glands, with the exception of the regional glands, hardly at all. Not infrequently the joints, mammary glands and testes show lesions. For differentiating between bovine and human bacilli in the rabbit the best doses to employ are 10 mgm subcutaneously, 1 mgm intraperitoneally and 0.01 mgm intravenously (Cobbett 1917).

Feeding the rabbit with 1-10 mgm of bovine bacilli sets up a disease that proves fatal in 2 to 3 months.

**GUINEA PIGS.**—The guinea pig is highly susceptible to experimental infection with bacilli of the human and bovine type. Indeed it has been stated that the subcutaneous injection of a single tubercle bacillus may succeed in producing disease—though only of a slow type showing little tendency to generalization (Wamoscher and Stocklin 1937; Doerr and Gold 1932). Less than 10 living bacilli even of virulent strains cannot be relied upon to cause disease in every animal (Schwabacher and Wilson 1937). With between 10 and 100 bacilli intramuscular inoculation into the thigh usually sets up a slowly progressive disease often characterized in the later stages by healing of the initial lesions. The bovine bacillus is rather more virulent than the human bacillus, this can be demonstrated however only by a series of comparative experiments in which carefully measured doses are introduced. Table 26 from Griffith quoted by Cobbett (1917) who gives an excellent review of the pathogenicity of tubercle bacilli to animals illustrates this.

TABLE 26  
SHOWING GREATER VIRULENCE OF BOVINE THAN HUMAN BACILLI TO GUINEA PIGS

Method of Injection	Dose of Bacilli	Average Duration of Life in Days	
		Bovine Type	Human Type
Intraperitoneal	1.0 mgm	11.9 (15 animals)	29.5 (17 animals)
	0.1 mgm.	16.4 (58 animals)	27.9 (40 animals)
Subcutaneous	1.0 mgm	49.6 (15 animals)	80.9 (30 animals)
	0.1 mgm	45.6 (58 animals)	65.0 (40 animals)

The subcutaneous injection—usually made in the left thigh—of either human or bovine bacilli, even in minute doses, is followed by death from generalized tuberculosis in about 6 to 15 weeks. At necropsy there is a caseous local lesion—the superficial inguinal, and often the femoral, glands are much enlarged and caseous—the sublumbar, portal, mediastinal, and bronchial glands are enlarged and generally caseous—the spleen is greatly enlarged, and is beset with irregular necrotic areas, varying considerably in size, of a yellowish white waxy appearance—the liver is enlarged and contains smaller necrotic areas of a yellow or greenish colour, the lungs contain small numbers of rounded gelatinous tubercles. The most striking appearance is afforded by the necrotic areas in the spleen and liver—these are peculiar to the guinea pig. They are more marked after injection with bovine than with human bacilli. True tubercles are not often seen except in the very early stages of the disease—and in the lungs. It will be noted that the distribution of lesions is quite different in the guinea pig from that in the rabbit. In the rabbit they are most marked in, and often confined to, the lungs and kidneys—in the guinea-pig the lymphatic glands, spleen, and liver bear the brunt of the disease—the lungs are but slightly affected, and the kidneys practically never.

Intraperitoneal injection of human or bovine bacilli is followed by death in 2 or 3 weeks. Post mortem, there is a local caseous abscess in the abdominal wall—the superficial inguinal glands are enlarged and caseous—the omentum is rolled up, thickened, and caseous—the portal and often the mediastinal glands are enlarged and caseous—and the spleen and liver may show small foci of necrosis. The effect of intravenous inoculation of minimal doses has been described by Fust (1933).

There is a disease that occurs naturally in guinea pigs known as pseudotuberculosis—it is caused by *Pasteurella pseudotuberculosis* (*B. pseudotuberculosis rodentium*). The lesions to which it gives rise include white rounded nodules in the spleen, liver and mesenteric glands. These are unlike the necrotic lesions caused by the true tubercle bacillus, but may confuse those who are not well acquainted with the disease—especially since when inoculated subcutaneously into normal guinea pigs, the pseudotubercle bacillus gives rise to a local lesion and caseation of the focal lymphatic glands. The differential diagnosis can be made by microscopical and cultural examination. Microscopically short non-acid fast, Gram negative bipolar stained bacilli are seen, though often only in small numbers—culturally these organisms grow readily on ordinary media within 24 hours (see Chapter 32).

Murine bacilli are much less pathogenic for the guinea pig than the other two mammalian types. Relatively large doses, 0.1 and 1.0 mgm., intraperitoneally may cause death from acute or chronic atypical generalized tuberculosis—but smaller doses intraperitoneally and subcutaneous doses up to 5 mgm., give rise to no more than retrogressive lesions remaining more or less localized to the site of inoculation.

Arian bacilli are very much less virulent for the guinea pig than the human and bovine types. Subcutaneous injection is followed by a local abscess and swelling of the regional glands—death is unusual unless a large dose is given. Intraperitoneal injection of large doses is followed by death in a few weeks. Post mortem, the omentum is thickened, the spleen is large and red, there are minute grey points in the liver and there may be fluid in the pleural cavities. Though neither after subcutaneous nor intraperitoneal injection are macroscopic tubercles visible—smears or cultures made from the spleen and liver will generally reveal the presence of tubercle bacilli—Yersin type of disease.

**RATS AND MICE.**—In the past these animals have been regarded as comparatively resistant to tuberculosis. More recent work, however particularly with mice seems to show that their resistance has been rather overestimated. Subcutaneous inoculation, except in large doses, seems to have little effect. Intraperitoneal inoculation of mice with 1 mgm. of human or bovine bacilli sets up a disease usually proving fatal in 3 to 4 weeks, while intravenous inoculation with 0.2 mgm. is often fatal in 2 to 3 weeks. Post mortem, the lungs are studded with translucent, gelatinous, milky tubercles—similar tubercles are sometimes present on the pericardium—the spleen shows a variable degree of enlarge-

ment but no macroscopic tubercle formation. Microscopically, the lungs contain enormous numbers of bacilli, while in smears of the spleen, liver, and kidneys the organisms are usually plentiful but not abundant. Smaller doses give rise to a less rapidly fatal disease, in which there is time for the pulmonary tubercles to become partly aggregated into irregular, hard, caseous masses. Small doses cannot be relied upon to cause progressive infection when injected intraperitoneally, but intravenous inoculation with as few as 10-100 living bacilli has, in our experience, often been followed by chronic disease. If the animals are killed within 3 months after such small doses, no tubercles may be visible in the lungs but quantitative bacteriological examination leaves no doubt that the bacilli are actively proliferating (Schwalmacher and Wilson 1937). Left to themselves the animals may live for several months or even a year, at post mortem some of them show miliary tuberculosis of the lungs. Mice may also be infected with very small numbers of bovine tubercle bacilli if these are inhaled in the form of an aerosol mist (Glover 1944). The virulence of the human and bovine types for mice seems to be very much the same (Lange 1922; Stamatin and Stamatin 1939). The avian type was found by Gunn and his colleagues (1934) never to cause fatal disease after intravenous inoculation of 0.25 mgm. but our experience does not bear this out. We do, however, find that the avian bacillus is less virulent than the mammalian types. According to Lange (1922) tuberculous mice appear to be relatively non-allergic. Field mice (*Arvicola arvalis*) were found by Koch (1886) to be more susceptible than house mice, after subcutaneous inoculation they died in 4 to 6 weeks with extensive tuberculosis of the lungs, liver and spleen.

**VOLES**—Wells (1938) found that the vole (*Microtus agrestis*) was useful for distinguishing between the human and the bovine types of tubercle bacilli. In his experience bovine bacilli in a dose of 0.001 mgm. intraperitoneally produced extensive and progressive disease, whereas human bacilli, even in a dose of 1 mgm., produced no more than an insignificant lesion in the mesentery. Griffith (1939a, 1941a, c) states that the vole is susceptible to infection with all three mammalian types and with the avian type. The bovine type is the most virulent giving rise when inoculated parenterally or introduced by feeding to generalized progressive tuberculosis which runs a rapid course and is characterized by widespread caseation of the lymphatic glands and great multiplication of bacilli in the lesions. The human and the avian bacillus can multiply in the tissues of the vole, but have little tendency to produce macroscopic lesions. The murine type gives rise to generalized tuberculosis which runs a more chronic course than that caused by the bovine type, and is distinguished by the occurrence in the areolar tissues of masses of necrotic or caseous material composed largely of acid fast bacilli.

**OTHER ANIMALS**—The dog is relatively resistant to experimental tuberculosis but young animals may be infected by intraperitoneal inoculation with the mammalian types. Horses and asses are also resistant, subcutaneous injection sets up no more than a local lesion. Cats are highly susceptible to bovine bacilli, slightly susceptible to avian and resistant to human bacilli. Subcutaneous injection of 0.1 mgm. of bovine bacilli sets up invariably a rapidly fatal generalized tuberculosis. Monkeys and anthropoid apes are easily infected either by subcutaneous inoculation or by feeding with the human and bovine types but are comparatively resistant to the avian type. The golden hamster (*Cricetus auratus*) is very susceptible to experimental infection with the bovine and slightly less so to the human type, both of which produce progressive generalized tuberculosis. The murine type causes generalized tuberculosis of slow development with the production of lesions that do not undergo necrosis or caseation. The avian type is the least virulent and rarely gives rise to macroscopic lesions (Griffith 1939b, 1941b). Ferrets in captivity brought up on raw milk sometimes contract tuberculosis. Experimentally they are susceptible to the bovine but only very slightly to the human type of bacillus (Dunkin Laidlaw and Griffith 1929; Dalling and O'Brien 1935).

**BIRDS**—With the exception of the parrot, the cockatoo, the canary, and possibly certain birds of prey, birds are resistant to infection with the mammalian bacilli. Sub-

cutaneous intraperitoneal or intravenous injection of avian bacilli into fowls or pigeons gives rise to fatal tuberculosis. At necropsy the main lesions are found in the liver and spleen which contain rather hard caseous tubercles well differentiated from the surrounding tissue. After feeding experiments tubercles are often seen projecting through the peritoneal covering of the intestine.

**COLD BLOODED ANIMALS**—The cold blooded type of tubercle bacillus is unable to set up progressive disease in mammals or probably birds after subcutaneous injection of large doses a local lesion may result, but the disease does not spread. Frogs, turtles, lizards, snakes fish and other cold blooded animals are susceptible to experimental inoculation with the cold blooded type and are generally regarded as insusceptible to the mammalian and avian types. Griffith (1941c) however states that the water or grass snake (*Tropidonotus natrix*) is highly susceptible to experimental infection with the avian type. He (1941d) has also found that mammalian and avian bacilli, if inoculated into the dorsal sac of a toad multiply slowly in the liver and may be recovered 1, 2 or 3 years later. Subcutaneous or intraperitoneal inoculation with cold blooded bacilli gives rise to lesions that are often widely distributed throughout the viscera. Their nature depends on the site of inoculation and the time of survival. Soft, nodular lesions in the liver or lungs, filled with creamy or caseous material are not uncommon. Sometimes the liver is studded with little greyish white granules almost confluent (see Ledoux Lebard 1900, Friedmann 1903, Kuster 1903). Whether cold blooded bacilli are pathogenic for birds is not clear. Aronson (1936) states that he isolated a cold blooded bacillus from certain salt water fish—a sergeant major (*Abudefduf mauritii*) three croakers (*Micropogon undulatus*) and two sea bass (*Centropristis striatus*)—which proved pathogenic for pigeons.

**Developing Chick embryo**—A few observations have been made on the growth of tubercle bacilli on the chorio-allantoic membrane of the developing chick embryo. According to Fite and Olson (1944) the lesions produced under these conditions by the human type of bacillus differ from those caused by the bovine and avian types. Though the method may be of differential value it does not appear to be suited for estimating the virulence of strains to their natural animal host.

Table 27 modified from Cobbett (1932) summarizes the reaction of different animals to the four main types of tubercle bacilli.

The *saprophytic acid fast bacilli* are unable to set up a progressive infection in mammals or birds. Nevertheless when injected intraperitoneally in fairly large doses especially together with some fatty protective substance such as butter they may give rise to extensive lesions closely simulating those of true tuberculosis (see Rabinowitsch 1897, Grassberger 1899, Hagan and Levine 1932). This appears to result from the dissemination of the bacilli in the tissues by the lymph stream and leucocytes and the subsequent focal reaction of the tissues around them. True nodule formation and caseation may occur and acid fast bacilli are found microscopically in the lesions. After subcutaneous or intramuscular inoculation the lesions are usually confined to the local site and the regional lymphatic glands. The disease may be distinguished however from tuberculosis by culture and by further inoculation. Cultures made from the lesions on to glycerine agar will reveal a growth of saprophytic acid fast bacilli in 2 or 3 days while further inoculation into a guinea pig using a saline suspension of a small portion of one of the lesions will prove innocuous. Histologically the lesions caused by the saprophytic acid fast bacilli show more exudation than proliferation, there is less tendency to caseation and more to suppuration, polymorphonuclear are commoner than epithelioid cells, and typical giant cells with peripheral nuclei are rare (Rabinowitsch 1897). Saprophytic acid fast bacilli injected intravenously



TABLE 27

SHOWING THE SUSCEPTIBILITY OF CERTAIN ANIMALS TO EXPERIMENTAL INFECTION WITH THE FOUR MAIN TYPES OF TUBERCLE BACILLI

Susceptible to	Species of Animal	Severity of Disease caused by Types of Tubercle Bacilli			
		Human	Bovine.	Murine.	Avian
General infection by bovine type	Ox goat cat	±	+++	± (calf)	±
	Pig	±	+++	?	±
	Horse (a)	0	+	?	0
	Rabbit (d)	±	+++	±	+ Y
	Vole (d)	±	+++	++	±!
Infection by both bovine and human types	Guinea pig (b)	+++	+++	±	+ Y
	Dog (a)	±	±	?	0
	Anthropoid apes and other monkeys	++	++	?	0
	Other wild animals in captivity	++	++	?	?
	Rat (c) mouse (c)	+++ Y	+++ Y	+++ Y	+ Y
	Golden hamster	+++	++	+	+ Y
	Parrot, cockatoo and canaries (f)	++	++	?	++
Insusceptible to mammalian types	Fowls and other domestic birds	0	0	0	++

± A local retrogressive lesion

± Localized tuberculosis with sometimes slight dissemination

+, ++, +++ Different degrees of progressive tuberculosis.

+ Y Tuberculosis of the Yersin type

(a) Spontaneous tuberculosis relatively uncommon difficult to infect experimentally

(b) Extremely easy to infect experimentally but seldom contracts tuberculosis naturally

(c) Spontaneous tuberculosis is rare, after intravenous inoculation more die with pulmonary tuberculosis of the lungs and an enlarged spleen containing tubercle bacilli in great numbers

(d) Spontaneous infection with the murine type appears to be common The natural disease and the experimental disease caused by the murine type are characterized by tuberculosis of the lungs and lymphatic system and the occurrence of necrotic or caseous material in the subcutaneous areolar tissue reminiscent of rat leprosy Experimental infection with the bovine type gives rise to acute generalized tuberculosis

(e) Produces slowly progressing generalized tuberculosis without necrosis or caseation of the lesions

(f) Canaries are said to be less susceptible to infection with the human and bovine types than parrots and cockatoos (Tulloch 1936)

into laboratory animals or even into larger animals such as calves, may give rise to severe illness, sometimes followed by death (Kossel *et al* 1904, Twort and Craig 1913) The bacilli do not multiply in the tissues, and therefore no true infection is set up, the symptoms appear to be due to toxæmia following the liberation of endotoxins from the bacilli in contact with the tissues. Similar results can be obtained with dead bacilli. This toxæmia following the intravenous injection of living or dead acid fast bacilli whether of the saprophytic acid fast or the true tubercle type, into susceptible animals, is a phenomenon that may lead to confusion if differentiation of the various types is attempted by the intravenous route. For this reason the subcutaneous method of injection is generally to be preferred.

**Type Differentiation by Pathogenicity**—At the risk of repetition we give for the sake of clearness the methods by which the three main types of tubercle bacilli may be differentiated

The *human* and the *bovine* types of tubercle bacilli are able, when injected subcutaneously in minute doses, to give rise to a progressive and fatal disease in guinea pigs. But whereas the bovine type is able likewise to produce a progressive, generalized and fatal disease in rabbits, cats, voles, goats, and calves, the human bacillus cannot do so. This fundamental distinction can be elicited in practice only by strict regard to certain factors, such as dosage and route of inoculation. The experience of the Royal Commission showed that the best differentiating dose for calves was 50 mgm. subcutaneously, and for rabbits 10 mgm. subcutaneously. The bacilli are taken from a 1 to 3-weeks old culture on inspissated serum and weighed moist. Under these conditions the bovine bacillus sets up a generalized fatal disease, while the human bacillus causes at most a localized and retrogressive disease confined to the site of inoculation, and sometimes to one or more of the internal organs. Often no lesions are visible at all. Animals vary so much in their susceptibility to experimental inoculation that it is advisable to inject two or three simultaneously, and to repeat the test on a fresh series of animals if the results are not precise. The time taken to produce death is probably not of such importance as the extent of the lesions at necropsy (Park and Krumwiede 1910) but generally speaking calves inoculated with a bovine strain may be expected to die in about 6 to 8 weeks, and rabbits in 6 to 12 weeks. A more rapid method of differentiation consists in the intravenous inoculation of rabbits with 0.001 mgm. or even less. In such a dose the human bacillus has little effect. The bovine kills the animal in 4 to 6 weeks, and at post mortem milary tuberculosis is found affecting particularly the lungs and kidneys. According to Wells (1944) an equally satisfactory and perhaps even more rapid method consists in the subcutaneous inoculation of 1 mgm. into the Orkney vole (*Microtus orcadensis*). The human bacillus is practically non pathogenic to this animal, giving rise to no more than a local lesion with perhaps minimal lesions in the focal lymphatic nodes and occasionally a few scattered tubercles in the lungs, whereas the bovine bacillus causes generalized glandular and visceral tuberculosis, proving fatal as a rule in 3 to 5 weeks.

The separation of the avian from the mammalian bacilli is generally quite straightforward. The avian bacillus is pathogenic for fowls and pigeons, but not for the guinea pig. The mammalian bacilli on the contrary, are pathogenic for the guinea pig but not for fowls and pigeons. Cobbett (1917) recommends for differentiating purposes the injection of 10 mgm. of culture subcutaneously into guinea pigs, and 10 mgm. intraperitoneally into fowls or pigeons. The mammalian bacilli prove fatal to guinea pigs in about 4 to 8 weeks, and at necropsy there is generalized tuberculosis with extensive caseation of the glands and large necrotic areas in the spleen and liver. The avian bacilli often give rise to no lesions at all, or only to a slight local collection of caseous material, but sometimes, if the animals are killed after a few weeks, the focal glands will be found to be hyperemic, and the spleen enlarged and congested, smears of the spleen, liver, and lungs from these cases may reveal a few acid fast bacilli (Straus and Gamaleia 1891). After injection with avian bacilli, fowls and pigeons die in a variable time, often considerably emaciated, if they are still alive at the end of 3 months they should be killed. Disease is most evident in the spleen, liver, and kidneys, which show numerous tubercles or hard caseous areas, bacilli are abundant in smears of these organs.

The mammalian bacilli generally produce no recognizable lesions in fowls or pigeons, but occasionally a local abscess forms, which contains a few bacilli. Avian bacilli are pathogenic to rabbits if inoculated intravenously, giving rise to the Yersin type of disease (see p 423). Griffith used to inject 0.01 mgm intravenously into rabbits and 10 mgm intramuscularly into hens. Sometimes, however, avian strains are encountered, particularly from pigs, that are pathogenic to fowls but not to rabbits.

**Lesions caused by the Injection of Dead Tubercle Bacilli.**—The injection into animals of dead tubercle bacilli, preferably killed by heat, gives rise to lesions of varying degrees of severity. Subcutaneous injection provokes merely a local abscess, but after intravenous injection the dead bacilli are deposited in the viscera, where they give rise to small follicular lesions closely resembling true tubercles, and sometimes undergoing caseation. This disease is known as *Necro tuberculosis* (Miller 1905). Apparently the dead bacilli are attacked and broken down by the cells of the body and their intracellular toxins liberated, these toxins then act on the tissues, and give rise to the characteristic lesions of tuberculosis. For the production of necro tuberculosis, virulent bacilli are not essential, we have produced lesions in rabbits by the intravenous injection of dead avirulent bacilli, such as Calmette's BCG strain.

**Classification**—We have already indicated that the acid fast bacilli may be divided into six types—human, bovine, murine, avian, and cold blooded tubercle bacilli, and the saprophytic acid fast bacilli. The distinction between the types rests chiefly on cultural appearances and pathogenicity, aided when necessary by serological reactions. These types are remarkably constant so far as we are aware, no clear evidence has ever been produced to prove that one type may change into another. Numerous experiments have been made by different methods to bring this about, but no one has yet succeeded in doing so.

Nevertheless, the division of the mammalian tubercle bacilli is not always clearly defined. From time to time aberrant strains are encountered, which depart from the standard type either in cultural behaviour, in pathogenicity, or in both. The most fruitful source of these aberrant types is lupus, thus from 140 cases of lupus examined by A. S. Griffith (1924), no fewer than 99 yielded strains of aberrant type. Apparently, when growing in the skin, tubercle bacilli are liable to undergo a modification in virulence, so that both human and bovine types become less pathogenic for experimental animals. Aberrant types are encountered in other lesions, though much less frequently, thus Griffith (1916) found 4 aberrant strains amongst a total of 212 strains isolated from sputum, and 5 amongst 141 strains isolated from bone and joint disease (1916–17), Eastwood and F. Griffith (1916) likewise isolated 10 aberrant out of a total of 261 strains from cases of bone and joint disease. Aberrant strains have occasionally been found in animals, especially horses (A. S. Griffith 1924, Stableforth 1929).

The mammalian bacilli may be subdivided as follows (Table 28, p 432).

Of these various types the attenuated eugonic human and the attenuated dysgonic bovine are the ones usually found in lupus, the attenuated dysgonic human and attenuated eugonic bovine are rare, and have been met with in lupus only. The dysgonic human type was the type isolated from sputum and from bone and joint disease, in the cases already referred to. On serum and glycerine serum it grows luxuriantly with the production of yellow pigment. But on glycerine agar and glycerine potato growth is very slow, an effuse grey glazed layer of growth

TABLE 28 (Modified from A. S. Griffith 1924)

SHOWING THE DIFFERENTIAL CHARACTERISTICS OF MAMMALIAN TUBERCLE BACILLI.

Human.	Classification.	Cultural Characteristics.	Virulence
1	Typical human	Eugonic on all glycerine media. Pigmented growth on bovine serum	High for guinea pig, low for rabbit (standard human virulence)
2	Dysgonic human	Dysgonic on glycerine agar potato, and broth. Pigmented growth on bovine serum	Like (1)
3	Attenuated dysgonic human	Like (2)	Very low for rabbit and lower for guinea pigs and monkeys than standard human strains.
4	Attenuated eugonic human	Like (1)	Like (3)
Bovine.			
1	Typical bovine	Dysgonic on all glycerine media no pigment formation on bovine serum	High for guinea-pig and rabbit (standard bovine virulence).
2	Eugonic bovine	Growth better than typical bovine strains, but not equal to human	Like (1)
3	Attenuated eugonic bovine	Like (2)	Less than standard bovine virulence for all animals, but higher for calf and rabbit than standard human.
4	Attenuated dysgonic bovine	Like (1)	Like (3).
Murine	Typical murine	Grows very slowly and dysgonic on glycerine media.	High for voles, but very low for guinea pigs and rabbits.

occurs, on which a few large isolated warty and pigmented colonies may eventually develop. Similarly on glycerine broth there is only a thin layer of growth with an occasional raised island of irregular appearance. In virulence this variety resembles the standard human type, being high for the guinea pig and low for the rabbit. These dysgonic human strains are most likely to be confused with the attenuated bovine strains, differentiation can be effected by making a sufficiently wide and prolonged series of tests. The dysgonic bovine strains do not give a yellow pigmented growth on bovine serum, and their virulence is lowered not differentially but uniformly, that is to say, their virulence is lowered not only for the more resistant calf and rabbit, but also for the highly susceptible guinea pig and monkey.

Whenever an aberrant strain is encountered, the possibility of the cultures being impure must always be considered. Thus the Royal Commission (Report 1911) examined 2 cultures that were eugonic on glycerine media and were highly virulent to calves and rabbits. By plating out these cultures they were able to separate off a dysgonic virulent strain from a eugonic strain of slight virulence for the calf and rabbit, in other words the original cultures contained a mixture of bovine and human types.

Considerable attention has been paid to these atypical strains because they have been regarded by some workers as evidence of instability of type. Griffith who has had the greatest experience of these atypical strains is, however,

firmly of the opinion that they represent no more than modifications of the fixed types, they do not represent transitional forms between the types. By passage through suitable animals it is often possible to restore the normal properties of their type to these atypical strains, but it has never yet proved possible to modify them in such a way that they change into a different type. Thus an attenuated dysgonic bovine strain, if passed through a series of rabbits, will often regain the standard virulence of the bovine type, it will not take on a eugonic growth and come to resemble the human type (Griffith 1924). The extensive observations of Jensen and Frimodt Møller (1936) and of Frimodt Møller (1939) are in general accord with Griffith's findings.

We may conclude therefore that the types of tubercle bacilli are fixed, that occasionally modifications due to residence in a particular environment may result but that no alteration of environment or any other factor has yet been successful under experimental conditions of changing an organism of one type into that of another.

Classification of the saprophytic acid fast strains is very unsatisfactory. Numerous types have been described, and have been differentiated on morphological and cultural appearances, the degree of acid fastness, the formation of pigment and the optimum temperature of growth (see Haag 1927, Pinner 1932, Thomson 1932, Schwabacher 1933b, Gordon and Hagan 1938).

A large amount of detailed information, much of which has been overlooked by subsequent workers on the various properties of the tubercle bacilli, is contained in the Reports (1907, 1909, 1911, 1913) of the Royal Commission on Tuberculosis. Those anxious to learn more about these organisms are advised to study these reports carefully, as well as the numerous subsequent publications of Dr A. Stanley Griffith, many of which have appeared in the *Journal of Pathology and Bacteriology* and the *Journal of Hygiene* during the past thirty years.

#### *Mycobacterium tuberculosis, Human type*

*Isolation*—By Koch in 1882 from human tuberculous lesions.

*Habitat*—Strict parasite causing tuberculosis in man, pigs, monkeys, dogs, and parrots.

*Morphology*—Rod shaped organisms 1-4  $\mu$  long and 0.2-0.8  $\mu$  broad, straight or slightly curved, with parallel or irregular sides, and rounded ends, arranged singly or in small clumps, non motile, non sporing, and non capsulated. Fail to stain with simple dyes except after prolonged exposure. Stain best with hot carbol fuchsin, resist decolorization with 25 per cent  $H_2SO_4$  and with absolute alcohol for 10 minutes. Gram positive, staining may be even or granular, beaded forms are common. In the animal body the bacilli are larger than in culture. Non acid fast and clubbed forms are not uncommon in culture.

*Coagulated Ox serum*—4 weeks, 37° C. Thin effuse, confluent, greyish yellow growth, with a finely granular surface, looking like ground glass. Colour may be golden yellow, consistency friable, emulsifies with great difficulty.

*5 per cent Glycerine Ox serum*—4 weeks, 37° C. More luxuriant, thicker, raised, confluent, yellow or golden yellow growth, with a coarsely granular surface, irregularly heaped up in places, often a granular film over the water of condensation. Consistency friable, emulsifies with great difficulty.

*Dorset Egg*—4 weeks, 37° C. Rather poor, discrete or confluent, slightly raised, yellow growth, with a finely granular surface.

*5 per cent Glycerine Egg*—4 weeks, 37° C. More luxuriant, raised, confluent, greyish growth with a coarsely granular surface, growth irregularly nodular in places, heaped up in places.

*5 per cent. Glycerine Agar*—4 weeks, 37° C Thick, raised, confluent, cream-coloured growth with a nodular or wrinkled surface.

*5 per cent. Glycerine Potato*—4 weeks, 37° C Thick, raised, confluent growth, creamy or yellow in colour, with a wrinkled, nodular, or warty surface

*5 per cent. Glycerine Broth*—4 weeks, 37° C Greyish white surface pellicle often irregularly thickened in places. On further incubation the pellicle increases in thickness, develops a deeply wrinkled surface, and spreads for about  $\frac{1}{2}$  inch up the sides of the flask. No turbidity, but often a slight granular deposit.

*Plain Agar or Broth*—No growth, as a rule.

*Resistance*.—Cultures live for 4 to 8 weeks as a rule but may remain viable for a year. Bacilli are killed by moist heat at 60° C. in 15 to 20 minutes. In excised tissues, kept at 37° C., they die in about a week. In dried sputum most of the bacilli die in a few days. Are fairly susceptible to sunlight and ultra-violet light. Moderately resistant to chemical disinfectants, in sputum may survive exposure to 5 per cent. phenol or antiformin for 24 hours.

*Metabolism*.—Growth occurs between pH 4.5 and 8.0, optimum pH is 7.0-7.6. Optimum temperature 37° C., very little growth, if any, below 30° C. Growth occurs best in an atmosphere of 40-50 per cent. O<sub>2</sub>, no growth under strictly anaerobic conditions. Growth is improved by addition of glycerine and of dead acid fast bacilli to the medium, and is said to be improved by substances rich in Vitamin B and by very small quantities of iron salts. Golden yellow pigment produced on glycerinated ox serum.

*Biochemical*.—Very little known about biochemical properties. Evidence that the bacilli can produce acid from glucose, maltose, lactose, sucrose, glycerol and trehalose

*Antigenic Structure*.—By agglutination, absorption of agglutinins, and complement fixation the human bacilli are shown to form a homogeneous group indistinguishable from bovine tubercle bacilli but easily distinguishable from avian cold blooded and saprophytic acid fast bacilli.

*Pathogenicity*.—Produces tuberculosis in man, pigs, monkeys, dogs and parrots. Experimentally, it is highly pathogenic for the guinea pig but not for the rabbit, cat, goat or ox. A minute quantity injected subcutaneously into a guinea pig at high causes death in 6 to 12 weeks. P.M. local caseous abscess, enlargement and caseation of the inguinal, sublumbar, portal, axillary, and bronchial glands, enlargement of the spleen with production of irregular yellowish areas of necrosis, enlargement of the liver with production of smaller, irregular, greenish yellow areas of necrosis, few rounded tubercles in the lungs. Tubercle bacilli are numerous in the local lesion and the inguinal glands, more scanty elsewhere.

The characters of the different types of tubercle bacilli, including the saprophytic acid fast bacilli, are summarized in the table on pp 435-7 (Table 29)

### The Leprosy Bacillus

The causative organism of leprosy, which was one of the first micro-organisms shown to be pathogenic to man, was discovered by Hansen in 1868 though his published account did not appear till 1874. He observed it in the tissues of lepers, where it occurs in large numbers in the granulation tissue cells. The bacilli vary in size and shape, they may be straight or slightly curved, 1-8  $\mu$  in length, with parallel sides and rounded ends, arranged chiefly in clumps or bundles and staining evenly, or they may resemble diphtheroids and show granular staining confined to the poles or distributed throughout their length. They are not easily stained without a mordant, they are Gram positive and strongly acid fast. They are non motile and non sporing.

Though the leprosy bacillus was one of the first of the pathogenic organisms to

TABLE 29

THE GROWTH OF THE VARIOUS TYPES OF TUBERCLE BACILLI ON DIFFERENT MEDIA

	Human	Bovine	Murine	Avian.	Cold blooded	Saprophytic Acid fast
<i>Nutrient agar 4 weeks at 37° C</i>	No growth	No growth	No growth	Poor, partly confluent, effuse, trans- lucent, ground glass growth, with finely granu- lar surface	7 d 22° C Moderate, semi conflu- ent raised irregularly heaped up growth with smooth or granular sur- face	7 d 22° C or 37° C Abun- dant, conflu- ent, raised, irregularly heaped up growth with smooth granular or worm cast surface
<i>Broth 4 weeks at 37° C</i>	No growth	No growth	Usually no growth	No turbidity or surface growth, but a moderate viscous mem- brano granu- lar deposit, partly dis- integrating on shaking	7 d 22° C Similar to avian type	7 d 22° C or 37° C No turbidity, dull dry, scaly or granular, often pig- mented, sur- face pellicle and a viscous membrano granular deposit, partly disin- tegrating on shaking
<i>Coagulated Ox serum 4 weeks at 37° C</i>	Thin, effuse, confluent, greyish yellow growth with finely granu- lar surface like ground glass	Similar to the human type, but often poorer		Similar to the human type	7 d 22° C Moderate, confluent, slightly raised growth with finely nodular sur- face	7 d 22° C or 37° C Abun- dant, raised, confluent, often pig- mented growth, irregularly heaped up in places, and with smooth or nodular surface
<i>Glycerine Ox serum 4 weeks at 37° C</i>	More luxuri- ant thicker, raised, con- fluent, yellow or golden yellow growth, with coarsely granular surface, irregularly heaped up in places	Similar to growth on plain ox serum	Similar to the bovine type, but colourless	Luxuriant, raised, con- fluent, yellow or golden yellow growth with a smooth creamy sur- face	Similar to growth on plain serum, but often more abundant	Similar to growth on plain serum, but often more abundant

TABLE 29 (continued).

	Human.	Bovine.	Marine.	Avian.	Cold blooded.	Saprophytic Add fast.
<i>Dorset egg 4 weeks at 37° C</i>	Rather poor, discrete or confluent, slightly raised, greyish yellow growth, with a finely granular surface	Similar to human type, but often poorer	Very slow Just visible in 4 weeks as pearly white hemi- spherical colonies, sometimes with narrow frilled mar- gin, or occa- sionally with wide- spread outgrowth having fern like markings on surface. Colonies may be conical and granular, blackberry like, or umbilicated.	Similar to human type, but surface is more smooth and less granular	7 d. 22° C Moderate, slightly raised, confluent growth with moist, glistening, finely granu- lar surface	7 d. 22° C or 37° C Abun- dant, con- fluent, often pigmented, raised growth with dull, dry, finely granular sur- face. growth may be irregularly heaped up
<i>5° Glycerine egg 4 weeks at 37° C</i>	More luxuri- ant, raised, confluent, greyish yellow growth, with coarsely granular surface, growth irregularly nodular and heaped up in places	Similar to growth on Dorset egg	Sometimes no growth If colonies do develop they are slightly larger than on egg, ivory white conical, with smooth or rough surface and frilled margin	Sometimes similar to human type, but usually more luxuri- ant, of a creamy yellow or slightly pinkish colour, with a smoother surface, growth often resembles butter cream	7 d. 22° C Good, raised, confluent growth, with relatively smooth sur- face, often resembles butter cream	7 d. 22° C or 37° C Luxuriant growth, raised, dry, yellow, pink, or brick red in colour, with a coarsely granular sur- face resem- bling dry bread crumbs.
<i>5° Glycerine agar 4 weeks at 37° C</i>	Good, some- times lux- uriant, gen- erally con- fluent, raised creamy white growth with a finely granular, scaly or wrinkled surface	Poor growth of small, discrete, granular colonies, sometimes a thin, effuse, con- fluent film	Extremely thin trans- lucent grey layer, con- sisting of minute discrete, flat, glistening colonies	Sometimes similar to human type, but usually more luxuri- ant, creamy white, with a paint-like surface	Luxuriant growth in 7 days at 22° C., creamy white and paint like	Similar to growth on glycerine egg



be described very little more is known about it now than at the time of its original discovery. Numerous attempts have been made to cultivate it, and many different organisms have actually been isolated from the tissues of lepers but it is by no means certain that the real causative organism of the disease has yet been grown in pure culture.

In 1901 Kedrowski cultivated apparently from 4 cases of leprosy, a non acid fast diphtheroid bacillus showing true branching. In very young cultures, 10 to 14 hours old most of the bacilli withstood decolorization with 5 per cent.  $H_2SO_4$  but in older cultures the organisms were non-acid fast except for the metachromatic granules. He stated that when injected into rabbits, the organisms became acid fast after a residence of several weeks in the tissues. He thought that the bacillus belonged to the *Actinomyces* group and that the acid fast rods seen in human leprosy represented only one stage in the developmental cycle of a single pleomorphic species. In 1905 Emile Weil obtained a growth of Gram positive acid fast bacilli on a medium consisting of a mixture of glycerine glucose peptone agar and egg yolk. Single colonies appeared about the 5th day, and increased in size till the 15th or 20th day. Cultures were likewise successful in hens eggs, the leprous juice being inoculated directly into the yolk of the whole egg. Subcultures were never obtained. In 1900 Clegg cultivated from 8 out of 10 lepers a weakly acid fast chromogenic bacillus. The primary cultures were obtained in symbiosis with amoebae and cholera vibrios, by heating these cultures to  $60^\circ C$  for 30 minutes he obtained a pure culture of the acid fast bacillus. The organisms, both morphologically and culturally, resembled the ordinary saprophytic acid fast bacilli, they resisted decolorization with alcohol for 3 minutes, but were largely decolorized by 5 per cent HCl in 2 minutes. Cultures on agar were of a bright orange colour. Local lesions resulted from injection of the bacilli into guinea pigs, similar apparently to those following injection of such organisms as *Myc. phlei* and the smegma bacillus.

In 1910 Dural cultivated from 4 cases of leprosy a non-chromogenic acid fast bacillus, cultures were made on an agar or banana medium enriched with a 1 per cent. solution of cysteine or tryptophan. Glistening white colonies 1.2 mm in diameter appeared after 1 to 2 months at  $32-33^\circ C$ , but not at  $37^\circ C$ . The organisms were nearly as acid fast as the tubercle bacillus, but were decolorized by 30 per cent.  $HNO_3$ , followed by 90 per cent alcohol, they failed to grow on ordinary media. When injected, even in small numbers, subcutaneously or intraperitoneally into Japanese dancing mice, they gave rise to glistening white nodules, resembling milary tubercles, these were disseminated throughout the body but were especially numerous in the spleen and lymph nodes. These nodules resembled the human lesions very closely, they contained acid fast bacilli in enormous numbers, chiefly intracellular in position. Injection into guinea-pigs, rabbits, rats, and mice was without effect. In the same year Twort (1910) isolated an acid fast bacillus from the nasal discharge of a leper by growth on an egg medium containing a glycerine extract of dead tubercle bacilli. Growth was not visible for 6 weeks. In 1911 Rost cultivated an acid fast chromogenic, highly pleomorphic bacillus from 3 cases of nodular leprosy at Rangoon. Primary isolation was obtained on a milk broth medium containing volatile alkaloids from rotten fish. Growth occurred in 3 days, and was yellow, pink or orange red in colour. A monkey injected repeatedly by different routes developed small nodules under the skin containing acid fast bacilli. Using a medium similar to Post's, but in which the rotten fish distillate was replaced by distilled water, Williams in 1911 cultivated two organisms from 5 cases of leprosy in Persia and Bombay, one was a non acid fast streptothrix the other an acid fast bacillus like Rost's. In old cultures of the latter organism in liquid media, a non acid fast diphtheroid appeared.

In 1912 Bayon (1911-12) isolated a non acid fast diphtheroid bacillus, sometimes showing branching from a case of nodular leprosy in London, it was morphologically identical with Kedrowski's bacillus. Intraperitoneal injection into a rat caused a lump at the site of inoculation. 3 months later the lump was punctured, and the fluid that

not yet been cultivated. Agglutination tests carried out with the serum of lepers do not help in determining the aetiological significance of these organisms, for it has been found that tubercle bacilli and saprophytic acid fast bacilli are agglutinated as well as the bacilli cultivated from leprosy (Duval and Wellman 1912). And since it is impossible to reproduce typical leprosy in laboratory animals by inoculation even with ground up leprosy material, it is clear that animal inoculation tests are likewise useless in deciding this question.

There is reason to believe that the non-chromogenic acid fast bacilli cultivated by various workers were in fact the real leprosy bacillus. Examination, however, of the records reveals the fact that, though colonial development was obtainable in the first two or three subcultures, attempts to carry on the bacilli indefinitely in subculture almost invariably failed. Even in Soule and McKinley's work, which incidentally Schlossmann (1933), Duval and Holt (1934a), Holt (1934a, b), and Dharmendra and Lowe (1935) have failed to confirm, more and more difficulty was experienced in obtaining growth with each successive subculture. Duval (1910) many years ago brought evidence to show that leprosy bacilli were able to grow in artificial media only so long as some of the original human lepromatous tissue persisted in the culture, and that the bacilli derived their nutriment almost entirely from the autolytic products of this human protein material (see also Duval 1934). These findings are supported by the more recent work of Schlossmann (1933). If they are correct, they seem to afford an explanation of many of the observed facts. Whether it will be possible, as Duval and Holt (1934b) anticipate, to replace the natural autolytic tissue products by protein split substances derived from other sources, is still doubtful. We may conclude tentatively that the leprosy bacillus has been grown in culture, but that its indefinite subcultivation in artificial media has never been unequivocally demonstrated.

In these circumstances it is regrettable that a number of stock cultures in various collections are labelled *Mycobacterium leprae* and that so much work has been expended in studying the various properties of these so-called leprosy bacilli. Many of the strains are undoubtedly ordinary saprophytic acid fast bacilli, and their description under a false name can do nothing but cause confusion in the literature. (For attempts at experimental reproduction of leprosy, see Chapter 60.)

### The Rat Leprosy Bacillus

This organism was first described by Stefansky (1903) in 1901 at Odessa, where it was giving rise to a leprosy like disease in rats. The organisms which have never yet been definitely cultivated, are 3-5  $\mu$  long, are often slightly curved and have rounded ends. They are Gram positive and strongly acid fast, withstanding 5 per cent  $H_2SO_4$  and 95 per cent alcohol for at least 5 minutes. Staining is often granular. According to Marchoux (1933) the rat leprosy bacillus is killed by exposure to moist heat at 60° C for 15 minutes. It cannot withstand drying but it remains viable for 2 years or more in infected organs preserved in 40 per cent glycerol at 0-6° C. From the fact that human leprosy cannot be conveyed to rats, it is probable that the leprosy and the rat leprosy bacilli are different. There is reason to believe that some cases of human leprosy may be caused by the rat leprosy bacillus (see Chapter 60). (For experimental reproduction of the disease in rats, see Chapter 60.)

## John's Bacillus

This organism was found by John and Frothingham (1895) in a chronic disease of cattle characterized by massive infiltration of the intestinal tract. It was believed at first to be an avian type of tubercle bacillus, but subsequent work has rendered it clear that it is a different species of organism.

*Morphologically* it is a short, thick rod, 1-2  $\mu$  long, generally straight, with rounded ends and parallel or slightly bulging sides, it is non motile. Gram positive, and strongly acid fast. Staining is generally uniform but may occasionally be granular. It was first isolated in pure culture by Twort in 1910 on a glycerine egg medium containing dead tubercle bacilli (Twort and Ingram 1912, 1913). Subsequently it was found that the saprophytic acid fast bacilli could replace the tubercle bacilli, and that a glycerine extract of the organisms, or even liquid tuberculin, could be used (McFadyean *et al* 1912). Twort and Ingram advise the following medium. Cultures of *Mycophiles* are killed by steaming, the growth is scraped off and dried, the bacilli are then ground in a mortar, and added in 0.5-1.0 per cent concentration to Dorset's egg medium containing 4 per cent of glycerine. On this medium primary cultures of John's bacillus consist of tiny, dull white colonies, rarely visible to the naked eye in less than 4 weeks, they are more or less circular in shape, and may be discrete or confluent. As they grow older, they increase in size, become more elevated and turn a dull yellowish white colour, the edges remain thin, and from them numerous irregular striations rise towards the central peak. In subcultures the growth is more copious, more confluent, and may be slightly wrinkled. If the saline in Dorset's egg medium is replaced by peptone beef broth, the growth is still better, and if sheep's brain broth or horse liver extract is used, the growth cannot be distinguished from a Dorset's egg culture of human tubercle bacilli. On glycerine agar containing *philes* extract the growth is slower and less vigorous. In glycerine broth with *philes* extract added growth occurs in the form of a thin surface film which may later show irregular areas of thickening. After several subcultures growth is more profuse and may even occur in plain glycerine broth without the addition of *philes* extract. Growth occurs between 23° C and 43° C, the optimum temperature being 39° C. In culture the bacilli are very short, often only about 1  $\mu$  in length, there is no branching but occasional club forms are seen. Primary cultures are best made by treating the washed intestinal mucosa with 20 per cent antiformin for about 30 minutes, and seeding on to a glycerine egg medium containing dead *philes* or tubercle bacilli (see Minett 1942). In what way the added organisms serve to promote the growth of John's bacillus is not known, they probably provide some nutrient substance, or enzyme, necessary for its metabolism, but that this substance is not specific to acid fast bacilli is shown by the fact that alcoholic extracts of such diverse substances as currant grapes, figs, oats linseed, and the fungus *Cantharellus aurantiacus*, are all able to replace the acid fast bacilli (Twort and Ingram 1914).

Whether the bacilli giving rise to John's disease in sheep are identical with those in cattle is not clear. Apparently they are more difficult to grow, and in primary culture may take 6 or 7 months to develop (Dunkin and Balfour Jones 1935). On the other hand McEwen (1939) claims to have infected calves by feeding them with material from diseased sheep, so that the difference between the bovine and ovine bacilli may be less than Dunkin (1936) suggests.

**Reproduction of the Disease in Animals**—The injection intravenously intra peritoneally or subcutaneously of pure cultures of *Johnes bacillus* into bovine animals frequently gives rise to the typical disease from the lesions of which pure cultures of the bacillus can be recovered (Twort and Ingram 1913). Feeding may also be successful. Sometimes goats and sheep can be infected by inoculation of pure cultures. Lambs are said to be more susceptible than adult sheep (McEwen 1939). Reproduction of the disease in laboratory animals has not so far been successful. Johnes and Frothingham (1895) and Twort and Ingram (1913) found that guinea pigs, rats, and mice were refractory. Later Twort and Craig (1913) found that the intraperitoneal inoculation of 100–150 mgm. of bacilli into rabbits gave rise in the abdominal cavity to a few nodules which were slightly caseous; the animals remained perfectly well and showed no signs of toxic disturbances. The same lesions were produced by *Mycophila*. Boquet (1925) found that the intraperitoneal injection of 5–10 mgm. of culture into white rats gave rise to pinhead greyish nodules on the surface of the peritoneum and omentum, these nodules contained pus very rich in bacilli. The mesenteric glands were enlarged; the tracheo-bronchial glands were enlarged, hard and sclerotic and contained enormous numbers of bacilli. Even more marked lesions were obtained when the injection was repeated in 15 to 20 days with a dose of 10–30 mgm. White mice developed similar but less chronic lesions. It is doubtful whether these changes can be considered specific for *Johnes bacillus*, similar lesions can often be produced by the saprophytic acid fast bacilli. It is certain that no one has yet reproduced in laboratory animals the typical enteritis of the natural disease.

## REFERENCES

- AKSIANKIEW M. I. (1933) *Z. Tuberk.*, 68, 249.  
 ALBISTON H. E. (1930) *Aust. vet. J.*, 6, 123.  
 ALVAREZ and TAVEL. (1885) *Arch. Phys. norm. Path.*, 6, 303.  
 ANDERSON R. J. (1932) *Physiol. Rev.*, 12, 166.  
 ANDERSON R. J. and Colleagues. (1937 et seq.) 34 papers, mainly in *J. Biol. Chem.*, 1937–33 and 3 papers in *Z. physiol. Chem.* (1930) 191, 157–166, 12.  
 ARONSON H. (1898) *Berl. Hin. Wochr.*, 35, 484.  
 ARONSON J. D. (1906) *J. infect. Dis.* 39, 315.  
 BALL E. G. (1934) *J. Biol. Chem.*, 106, 515.  
 BATAILLON E., DEBARD and TERRE, L. (1897) *C. P. Soc. Biol.*, 49, 446.  
 BAYON H. (1911) *Trans. R. Soc. trop. Med. Hyg.*, 5, 153.  
 BEAVER P. W. and BAYNE-JONES S. (1931) *J. infect. Dis.*, 49, 390.  
 BECK, M. (1905) *Tuberk. Arb.* 3, 145.  
 BEGHE, R. S. (1930) *Edinb. med. J.* 37, 187. (1931) *Ibid.*, 38, 173.  
 BEITZKE, H. (1910) *Berl. Hin. Wochr.*, 47, 1451.  
 BIRKHAUG K. E. (1935) *Ann. Inst. Pasteur* 54, 19.  
 BLACKLOCK, J. W. S. (193) *Spec. Rep. Ser. med. Res. Coun., Lond.*, No. 12.  
 BOQUET A. (1925) *C. P. Soc. Biol.*, 93, 219.  
 BOZELLE, R. (1914) *Ann. Stat., Mal. Best. Napoli* 2, 1.  
 BREX W. V. (1909) *J. Amer. med. Ass.* 53, 609.  
 BRUNER, D. W. (1934) *J. infect. Dis.* 55, 26.  
 BRUYNOGHE, R. and ADAM M. (1933) *C. R. Soc. Biol.*, 111, 1031.  
 BULLOCH W. and MACLEOD J. J. R. (1904) *J. Hyg. Camb.* 4, 1.  
 CADOT P. J., GILBERT A., and FOGER, H. (1900) *C. P. Soc. Biol.*, 42, 93.  
 CALDWELL, M. E. (1925) *J. infect. Dis.*, 37, 465.  
 CALMETTE, A. (1930) "L'infection bacillaire et la tuberculose chez l'homme et chez les animaux. 4th ed. Masson et Cie Paris.  
 CALMETTE, A., VALTIS, J., NEGRE, L., and BOQUET A. (1925) *C. R. Acad. Sci.*, 181, 491.  
 CHARGAFF E., FANGBORN M. C. and ANDERSON R. J. (1931) *J. Biol. Chem.*, 90, 45.  
 CLEGG M. T. (1909) *Philipp J. Sci. B* 4, 403.

- COBBETT, L. (1917) "The Causes of Tuberculosis" Cambridge, (1932) *J Path Bact.*, 35, 681
- CUMMING, W M (1925) *Tubercle, Lond*, 7, 10a
- CUMMINS, S L and WILLIAMS, E M (1933) *Tubercle, Lond*, 15, 49
- CURRIE, D H, CLEGG, M T, and HOLLMANN, H T (1912) *Publ Hlth Bull, Wash.*, No 47
- CZAPLEWSKI, E (1897) *Munch med. Wschr.*, 44, 1192
- DALLING, T and O BRIEN, R A (1935) *Brit med J.*, i 897.
- DARLIVE, E (1932) *Ann Inst Pasteur*, 49, 743
- DAVIES, R (1937) *J Path Bact.*, 45, 773
- DENYS, P (1935) "Contribution à l'étude de la variabilité du virus tuberculeux" Imprimerie Saint Alphonse, Louvain
- DERNBY, K G and NILSSON C (1922) *Biochem Z.*, 132, 393
- DHARMENDRA and LOWE, J (1934) *Indian J med Res*, 25, 83.
- DIECKMANN, H and MENZEL, G (1932) *Z Hyg Infekth.*, 113, 709
- DOERR, R and GOLD, E (1932) *Z Immunforsch.*, 74, 7
- DOUGLAS, S R and HARTLEY, P (1934) *Tubercle, Lond*, 16, 97
- DREYER, G and VOLLUM, R L (1931) *Lancet*, i 9
- DUNKIN, G W (1936) *Proc roy Soc Med.*, 30, 83
- DUNKIN, G W and BALFOUR JONES S F B (1935) *J comp Path.*, 48, 236
- DUNKIN, G W, LAIDLAW, P P, and GRIFFITH A S (1930) *J comp Path*, 42, 46
- DUVAL, C W (1910) *J exp Med.*, 12, 649, (1934) *Proc Soc exp Biol*, 1, 32, 494
- DUVAL, C W and HOLT, R A (1934a) *Proc Soc exp Biol N Y*, 31, 453, (1934b) *Ibid*, 31, 828
- DUVAL, C W and WELLMAN, C (1912) *J infect Dis.*, 11, 116
- EASTWOOD, A and GRIFFITH F (1916) *J Hyg, Camb.*, 15, 257
- EHRICH (1892) *Deutsch med Wschr.*, 8, 269
- EIDINOW, A (1927) *Brit med J.*, ii 160
- ÉMILE WEIL, P (1905) *Ann Inst Pasteur*, 19, 793
- EVANS, B and HANES J H (1939) *Proc Soc exp Biol, N Y*, 40, 112
- FIELDING, J W (1934) *Aust J exp Biol.*, 12, 1
- FITE, G L and OLSON, B J (1944) *Publ Hlth Rep, Wash.*, 59, 1423
- FONTÈS, A (1910) *Mem Inst Osw Cruz.*, 2, 141
- FRIEDMANN, F F (1903) *Zbl Bakt.*, 34, 647 793
- FRIMODT MÖLLER, J (1939) "Dissociation of Tubercle Bacilli: Investigations on the mammalian types including BCG" H. K. Lewis and Co., Ltd, London
- FROUIN, A and GUILLAUMIE (1923a) *C R Soc Biol*, 88, 1002, (1923b) *Ibid*, 88, 1095
- FURTH, J (1926) *J Immunol.*, 12, 273
- FUST, B (1938) *Z Hyg Infekth.*, 120, 547
- GLOVER, R E (1944) *Brit J exp Path.*, 25, 141
- GLOYNE, S R (1933) *Bull Hyg, Lond.*, 8, 39
- GORDON, R E and HAGAN, W A (1937) *Amer Rev Tuberc.*, 36, 349 (1938) *J Bact.*, 36, 39
- GOUGH, G A C (1932) *Biochem J.*, 26, 248, (1933) *Ibid J.*, 27, 1049
- GRASSBERGER, R (1899) *Munch med Wschr.*, 46, 341, 382.
- GRIFFITH, A S (1916) *Lancet*, i 721, (1916-17) *J Path Bact.*, 21, 54 (1924) *Tubercle, Lond*, 5, 569, (1925) *Tubercle, Lond*, 6, 417, (1931) *J comp Path.*, 44, 144 (1939a) *J Hyg Camb.*, 39, 244 (1939b) *Ibid*, 39, 154 (1941a) *Ibid*, 41, 250 (1941b) *Ibid*, 41, 250 (1941c) *Ibid*, 41, 284, (1941d) *Ibid*, 41, 259, (1941e) *Tubercle, Lond*, 22, 33, (1942) *J Hyg, Camb.*, 42, 527
- GRIFFITH, A S and DALLING, T (1940) *J Hyg, Camb.*, 40, 673
- GROLIER, A DE (1933) *C R Soc Biol.*, 113, 1500
- GRÜNBERG, B (1935) *Z Tuberk.*, 73, 197
- GUNN, F D, NUNESTER, W J, and HOUEN, E T (1934) *Proc Soc exp Biol, N Y.*, 31, 527
- HAAO, F E (1927) *Zbl Bakt.*, Hte Abt., 71, 1
- HAGAN, W A and LEVINE, P (1932) *J Amer vet med Ass.*, 81, 723
- HANSEN, G H A (1874) *Norsk Mag Lægevidensk.*
- HARDEN, A. (1913) *Rep roy Comm Tuberc, Lond.*, 6, part II Appendix
- HARPOTH H (1938) *Z Tuberk.*, 79, 140
- HERMAN, M (1908) *Ann Inst Pasteur*, 22, 92
- HOLT, R A (1934a) *Proc Soc exp Biol, N Y*, 31, 567, (1934b) *Ibid*, 31, 643
- HOOPER, F E, RENFREW, A G, and JOHNSON, T B (1934) *Amer Rev Tuberc.*, 29, 66
- ISHIMORI, K (1924) *Z Hyg Infekth.*, 102, 329
- JACOBITE and KAYSER, H (1910) *Munch med Wschr.*, 57, 1175
- JENSEN, K A (1932) *Zbl Bakt.*, 125, 222
- JENSEN, K A and FRIMODT MÖLLER, J (1936) *Acta tuberc Scand.*, 10, 83 217
- JOHNE, H. A and FROTHINGHAM, L. (1895) *Deutsch Z Tiermed.*, 21, 439

- KARV, M. C. (1930) *Tubercle, Lond.*, 11, 202.
- KARLINSKI, J. (1901) *Zbl Bakt.*, 29, 521.
- KARWACKI, L. and BIERNACKI, S. (1925) *Ann Inst Pasteur*, 39, 476.
- KATTFMANN, F. (1932) *Z Hyg InfektKr.*, 114, 121.
- KEDROWSKI, W. J. (1901) *Z Hyg InfektKr.*, 37, 52.
- KENDALL, A., DAY, A., and WALKER, A. (1930) *J infect Dis.*, 28, 45.
- KIEFFER, J. (1924) *Amer Rev Tuberc.*, 5, 662.
- KLOPFSTOCK, F. (1931) *Klin Wochr.*, 10, 967.
- KOCH, R. (1882) *Berl Hin Wochr.*, 19, 221, (1886) 'Microparasites in Disease.' New Sydenham Soc., London.
- KORN, O. (1899) *Zbl Bakt.*, 25, 532, (1900) *Ibid.*, 27, 481.
- KOSSEL, H., WEBER, A., and HEUSS (1904) *TuberkArb.*, 1, 1 (1905) *Ibid.*, 3, 1.
- KRAUS, R. and GERLACH, F. (1929) *Z ImmunForsch.*, 62, 339.
- KRAUS, R. and KOREF, O. (1933) *Z Tuberk.*, 67, 42.
- KRETSCHMER, O. S. (1934) *J Lab clin Med.*, 19, 350.
- KUETEL, E. (1905) *Munch med Wochr.*, 52, 57.
- LAIDLAW, P. F. and DUDLEY, H. W. (1925) *Brit J exp Path.*, 6, 197.
- LANG, B. (1932) *Z Hyg InfektKr.*, 98, 229.
- LANG, L. (1932) See KOLLE, W. (1932) *Dtsch med Wochr.*, 58, 304.
- LARSON, W. P. (1936) *Lancet* ii 1231.
- LASER, H. (1897) *Munch med Wochr.*, 44, 1191.
- LEDoux LEBARD (1895) *C R Soc Biol.*, 50, 610, (1900) *Ann Inst Pasteur*, 14, 535.
- LONG, E. R. (1926) *Amer Rev Tuberc.*, 13, 393.
- LONG, E. R. and SEIBERT, F. B. (1926, 1928) See series of 10 papers in *Amer Rev Tuberc.*, 13, 17.
- LOWE, J. and DHARMENDRA (1937) *Indian J med Res.*, 25, 329.
- MCCARTER, J. R. and TAYLOR, E. L. (1937) *Ann Rep Director 1935-36, Agric. exp Sta Calif Wisconsin Bull.*, No. 438.
- MCCOY, G. W. (1914) *Publ Hlth Bull.*, Wash., No. 61, p. 27.
- MCEWEN, A. D. (1939) *J comp Path.*, 52, 69.
- M FADYEAN, J., SHEATHER, A. L., and EDWARDS, J. T. (1912) *J comp Path.*, 25, 217.
- MCKINLEY, E. B. (1934) *Medicine, Baltimore*, 13, 377.
- MCKINLEY, E. B. and LEON, W. DE (1937) *Int J Leprosy*, 5, 259.
- MCKINLEY, E. B. and VERDER, E. (1933) *Proc Soc exp Biol.*, N Y., 30, 659.
- MADDOCK, E. C. G. (1933) *J Hyg, Camb.*, 23, 103.
- MAFFUCCI, A. (1890) *Zbl allg Path path Anat.*, 1, 409, (1892) *Z Hyg InfektKr.*, 11, 445.
- MARCHOUX, E. (1933) *Per fran Derm Venereol.*, 9, 323.
- MARCHOUX, E. and HALPHEV, E. (1912) *C R Soc Biol.*, 73, 249.
- MARZINOWSKY, E. J. (1900) *Zbl Bakt.*, 28, 39.
- MAYER, E. (1921) *Amer Rev Tuberc.*, 5, 75.
- MAYER, E. and DWORSKI, M. (1932) *Amer Rev Tuberc.*, 28, 105.
- MEANWELL, L. J. (1937) *J Hyg, Camb.*, 28, 392.
- MEISNER, I. and PRAUSNITZ, C. (1934) *Zbl Bakt.*, 132, 23.
- MENZEL, A. E. O. and HEIDELBERGER, M. (1938) *J biol Chem.*, 124, 89, 301.
- MERRILL, M. H. (1930) *J Bact.*, 20, 235.
- MILLER, J. (1905) *J Path Bact.*, 10, 1.
- MINETT, F. C. (1942) *J Path Bact.*, 54, 209.
- MIRONOVICH, T. (1901) *Z Hyg InfektKr.*, 37, 497.
- MOELLER, A. (1898) *Dtsch med Wochr.*, 24, 376, (1899) *Zbl Bakt.*, 25, 369, (1901) *Ibid.*, 30, 513.
- MORI, N. (1911) *Ann Sta Mal Best. Napoli*, 1, 327.
- MUCH, H. (1907) *Beitr Klin. Tuberk.*, 8, 85.
- MUELLER, J. H. (1926) *J exp Med.*, 43, 9.
- MUGARD and POY (1887) *Ann Inst Pasteur*, 1, 19.
- NOVY, F. G. and SOULE, M. H. (1925) *J infect Dis.*, 36, 165.
- NOTT, M. M. (1927) *J Hyg, Camb.*, 26, 44.
- ORSKOV, J. (1932) *Zbl Bakt.*, 123, 271.
- ORSKOV, J. L. (1932) *C P Soc. Biol.*, 92, 400.
- PARK, W. H. and KREMWIEDE, C. (1910) *J med. Res.*, 23, 205.
- PELLEGRINO, P. L. (1906) *Ann Igene (ser.)*, 18, 163.
- PETRI, R. J. (1893) *Arch Psichogerundh. dmt.*, 14, 1.
- PETROFF, S. A. (1927) *Proc Soc exp Biol.*, N Y., 24, 632, 956.
- PETROFF, S. A., BRANCH, A., and STEENKEN, W. (1927a) *Proc Soc exp Biol.*, N Y., 25, 14 (1927b) *Amer Rev Tuberc.*, 19, 9.
- PETROFF, S. A. and STEENKEN, W. (1930) *J exp Med.*, 51, 831; (1935) *J infect Dis.*, 56, 277.
- PINVER, M. (1932) *Proc Soc exp Biol.*, N Y., 30, 214.
- POTTINGER, J. E. (1942) *Amer Rev Tuberc.*, 45, 549, 558.

- PRYCE, D. M. (1941) *J Path Bact*, 53, 327
- RABINOWITZ, L. (1897) *Z Hyg InfektKr*, 26, 90. (1900) *Dtsch med Wschr*, 26, 257
- RAVENEL. (1901) *Trans Brit Congr Tuberc*, 3, 553
- REED, G. B. and RICE, C. E. (1929) *J Bact*, 17, 407; (1931a) *Canad J Res*, 4, 349. (1931b) *Ibid*, 5, 111
- Report (1907) Royal Commission on Tuberculosis, 2nd interim Rep., H.M. Stat. Off., London. (1909) *Ibid*, 3rd interim Rep. (1911) *Ibid*, Final Rep. (1913) *Ibid*, Final Rep., Appendix
- RHINES, C. (1935) *Amer Rev Tuberc*, 31, 493
- RICE, C. E. (1931) *Canad J Res*, 5, 375
- RICE, C. E. and REED, G. B. (1931) *Canad J Res*, 5, 122
- RIVOLTA (1889) *G anat Fisiol*, 1, 122
- ROCHAIX, A. and COLIN, G. (1911) *C R Acad Sci*, 153, 1253
- ROCKWELL, G. E. and HIGBERGER, J. H. (1926) *J infect Dis*, 38, 92
- ROST, C. R. (1911) *Sci Mem med Sanit Dep, India*, No 42, p. 7
- SABIN, I. R. (1932) *Phynol Rev*, 12, 141. (1938) *J exp Med*, 68, 837
- SABIN, F. R., DOAN, C. A., and FORENER, C. E. (1930) *J exp Med* 52, Suppl No 3
- SABIN, F. R., MILLER, F. R., DOAN, C. A., and WISEMAN, B. H. (1931) *J exp Med* 53, 51
- SABIN, F. R. and JOYNER, A. L. (1938) *J exp Med*, 68, 853
- SALLE, A. J. and MOSER, J. R. (1937) *Int J Leprosy*, 5, 163 2-3
- SCHAEFER, W. (1935) *C R Soc Biol*, 120, 1185
- SCHAEFER, W. (1937) *Ann. Inst Pasteur*, 58, 388
- SCHLOSSMAN, K. (1930) *Zbl Bakt*, 115, 474. (1933) *Ibid*, 128, 369
- SCHMIDT, H. (1925) *Zbl Bakt*, 94, 94
- SCHULTE TIGGES, H. (1920) *Dtsch med Wschr*, 46, 1225
- SCHWABACHER, H. (1933a) *Spec Rep Ser med Res Coun Lond* No 182 p 104 (1936b) *Ibid*, p 124
- SCHWABACHER, H. and WILSON, G. S. (1937) *Tubercle, Lond* 18, 442
- SEIDYK, A. and SELIBER, G. (1927) *C R Soc Biol*, 97, 57
- SEIBERT, F. B. (1941) *Bact Rev*, 5, 60. (1944) *Chem Rev* 34, 107
- SEIBERT, F. B. and GLENN, J. T. (1941) *Amer Rev Tuberc* 44, 9
- SEIBERT, F. B., LONG, E. R., and MORLEY, N. (1933) *J infect Dis*, 53, 175
- SEIBERT, F. B., PEDERSEN, K. O., and TISELIUS, A. (1938) *Amer Rev Tuberc* 38, 399
- SEIFFERT, W. (1932) *Z Immunforsch*, 74, 116
- SHAFFER, M. F. (1935) *J Path Bact*, 40, 107
- SHEV, T. H. (1934) *J Shanghai Sci Inst*, Sect iv 1, 157
- SHIGA, K. (1929) *Zbl Bakt*, 114, 511
- SHOUB, H. L. (1923) *J Bact*, 8, 121
- SIBLEY, W. K. (1889) *Zbl Bakt*, 5, 831. (1890) *Lancet* 1, 804
- SMITH, C. R. (1942a) *Amer Rev Tuberc*, 45, 334. (1942b) *Ibid*, 46, 549
- SMITH, T. (1898) *J exp Med*, 3, 451. (1899) *Ibid*, 4, 217 (1901 5) *J med Res* 8, 233
- SMITHBURN, K. O. (1935) *J exp Med*, 62, 645
- SOLTYS, A. and TAYLOR, A. W. (1944) *J Path Bact* 56, 173
- SONNENSCHNIG, C. (1930) *Zbl Bakt* 117, 284
- SORDELLI, A. and ARENA, A. (1931) *C R Soc Biol*, 117, 63
- SOULE, M. H. (1928) *J infect Dis*, 42, 93. (1934) *Proc Soc exp Biol* 11, 31, 1197
- SOULE, M. H. and MCHINLEY, F. B. (1932) *Amer J trop Med*, 12, 1, 441 (1938) *Amer Ass Advancement Sci, Symposium Ser* 1, 87 Science Press, N. Y.
- SPENGLER, C. (1907) *Dtsch med Wschr*, 33, 337.
- STABLEFORTH, A. W. (1929) *J comp Path*, 42, 91
- STAMATIN, N. and STAMATIN, L. (1939) *Ann. Inst Pasteur*, 63, 209
- STEENLUND, W., OATWAY, W. H., and PETROFF, S. A. (1934) *J exp Med* 60, 515
- STEFANSKI, W. K. (1903) *Zbl Bakt*, 33, 481
- STRATS J. and GAMALIELA, N. (1891) *Arch Med. exp.*, 3, 457
- SWEATY, H. C. (1928) *Amer Per Tuberc*, 17, 53
- TAMURA, S. (1913) *Z phynol Chem*, 87, 85
- THOMSON, H. M. (1932) *Amer Rev Tuberc*, 26, 162
- TIEDENMANN, H. J. (1931) *Zbl Bakt*, 122, 483
- TORLER, M. (1901) *Z Hyg InfektKr*, 36, 120
- TODA, T. (1931) *Z Hyg InfektKr*, 112, 463
- TULLOCH, W. J. (1936) *Edinb med J*, 43, 141
- TULLOCH, W. J., MUNRO, W. T., ROSS, G. R., and CUMMING, W. M. (1921) *Tubercle, Lond* 6, 18, 57, 105
- TWORT, C. C. and CRAIG, T. (1913) *Zbl Bakt*, 68, 435
- TWORT, F. W. (1901) *Proc roy Soc., B*, 83, 156.

- TWOOT, F W and INGRAM, G L Y (1912) *Proc roy Soc., B* 84, 517, (1913) "A Monograph on Johnes's Disease" London (1914) *Zbl Bakt* 73, 277
- UGA T (1930) *Jap J exp Med*, 13, 167
- UHLENHUTH P and SEIFFERT, W (1930) *Z Immunforsch*, 69, 187
- UYET, N (1927) *J infect Dis*, 40, 433
- VAGGEDES, (1898) *Z Hyg Infekthr*, 28, 276
- VILLEMIN J A (1868) "Études sur la Tuberculose" Paris.
- WÄMOSCHER, L. and STOECKLIN, H (1927) *Zbl Bakt*, 104, Beiheft, p 86
- WELLS, A Q (1937) *Lancet*, i, 1221, (1938) *Brit J exp Path.*, 19, 324, (1944) *Pers comm*
- WELLS, H G and LONG, F R (1932) 'The Chemistry of Tuberculosis. Baillière, Tindall & Cox, London
- WHERRY, W B (1930) *J infect Dis*, 46, 263
- WILLIAMS, R S and HOY, W A. (1930) *J Hyg, Camb*, 30, 413.
- WILLIAMS, T S B (1911) *Sci Mem med Sanit Dep, India* No 42 p 15
- WILSON G S (1925) *J Path Bact*, 28, 69, (1930) *J Hyg, Camb.*, 30, 40. (1947) 'The Pasteurization of Milk.' Ed Arnold & Co, London.
- WOLBACH S B and ERNST, H C (1904) *J med Res*, 12, 295
- WULFF F (1925) *Acta path microbiol scand.*, 2, 149
- WYCKOFF, R W G (1934) *Amer Rev Tuberc.*, 29, 339
- WYCKOFF, R W G and SMITHBURN H C (1933) *J infect Dis*, 53, 201
- YEOMAN, D and PORTER, K. R (1944) *J Bact* 48, 83
- YERSIN M A (1888) *Ann Inst Pasteur*, 2, 240
- ZIEHL, F (1882) *Dtsch med Wochr* 8, 401



## CHAPTER 17

### CORYNEBACTERIUM

#### DEFINITION *Corynebacterium*

Gram positive rod like forms, arranged usually in a palisade. Not acid fast. Often with club shaped swellings at the poles, generally with irregularly staining segments or granules. Non motile, non sporing. Growing aerotically or under microaerophilic conditions, but often capable of anaerobic cultivation. Never forming gas in carbohydrate media in which they may or may not produce acidity. They may or may not liquefy gelatin or serum. Some species produce a powerful exotoxin.

Type species, *Corynebacterium diphtheriae*

The generic name *Corynebacterium* was allotted by Lehmann and Neumann in 1896 to the group of bacteria containing the diphtheria bacillus and other species resembling it in morphology. By its derivation the name emphasizes the tendency to the formation of club-like forms that is characteristic of the type species and of several other species within the generic group. This name was accepted by the Committee appointed by the Society of American Bacteriologists (Winslow *et al* 1920), and was adopted as the valid generic name in the monograph on diphtheria issued under the aegis of the Medical Research Council in 1923 (Andrewes *et al*). It is gaining increasing currency in the literature and is very unlikely to be superseded. The summary of the generic characters, as recorded by the American Committee, was emended by the Bacteriological Committee of the Medical Research Council by the omission of aerobiosis as a generic character, the addition of a reference to the fermentative activities of the group, and the omission of any reference to toxin production. There appear to us to be advantages in calling attention to a striking biological character possessed by the type species and perhaps shared by some related species. With the restoration of this reference to toxigenicity we should adopt the summary of generic characters as given by the Committee of the Medical Research Council.

Although *C. diphtheriae* is universally accepted as the type species, it was not in fact the first to be described. Reymond and his colleagues in 1881, and again in 1883, described the isolation from the conjunctival sac of a bacillus which is now recognized, under the name of *C. xerosis*, as belonging to this genus. This organism was described more fully by Kuschbert and Neisser in the latter year, which also witnessed Klebs' description of the diphtheria bacillus in the diphtheritic false membrane. It was not until 1884, the year following, that Loeffler published his classical paper on diphtheria, and provided a description of the causative organism which afforded a standard of reference for all subsequent studies on this bacterial group. Any claims that might have accrued to *C. xerosis* on account of priority would in any case have been vitiated by the fact that it is quite impossible at the present time

to be sure that Raymond, and Kuschbert and Neisser, were dealing with a single bacterial species. It is moreover, equally impossible to identify with certainty any of the strains now labelled *C. xerosis* with those described in the early 'eighties. The stage of technical development that bacteriology had then reached did not permit each new organism that was isolated to be fully studied and described. The importance of *C. diphtheriae* as a human pathogen focussed attention on the differentiation of the species from others with which it might be confused. Our picture of the type species thus became more and more complete as time went on but to the related diphtheroids little attention was paid beyond that necessary to determine their probable relation to the diphtheria bacillus itself. As Andrewes and his colleagues point out it is extremely difficult to determine which of the strains, to which specific names have from time to time been assigned, represent well-differentiated species. Some of those who have reviewed the group as a whole have been liberal in the distribution of titles (Graham Smith 1908, Mellon 1917, Eberson 1918, Chalmers and Macdonald 1920). Here, as elsewhere,

we propose to adopt a conservative view, and to list as species only those organisms which have been adequately described and appear from this description to be reasonably well-differentiated. In discussing this question of classification and nomenclature in greater detail we may consider *C. diphtheriae* as a species since it has in fact been so regarded in most of the observations to which we shall refer. Recent work has, however, shown that it is divisible into three well-differentiated types and that this differentiation is of considerable importance from the medical point of view. The discussion of this particular problem may, however, conveniently be dealt with in a separate section.



FIG. 82.—*C. diphtheriae*

From 24 hours culture on Loeffler's serum  
( $\times 1000$ )

#### Habitat.

Though attention has in the past been concentrated almost entirely on the parasitic members of the *Corynebacterium* group, which live mainly on the skin and mucous surfaces of their animal host, there is reason to believe that some members are adapted to the saprophytic life. Diphtheroid bacilli are common in heated milk products (Mattick 1944) and appear to occur also in the soil. Many of the parasitic species are pathogenic, others form an important constituent of the normal bacterial flora of various hosts.

#### Morphology

The club-form from which the name is derived is only one of many shapes which may be assumed by the individual cells of the type species, *C. diphtheriae*. This organism is indeed characteristically pleomorphic. One of the most typical forms (see Fig. 82), in films prepared from a 24-hour culture on Loeffler's serum is that of a

long, rather slender bacillus, often slightly curved, with rounded, somewhat swollen ends and sometimes with localized swellings elsewhere, and staining unevenly with such dyes as methylene blue but in the same, or in other cultures, there will also be found much shorter forms, cells which stain solidly and evenly, cells in which the irregular staining takes the form of a series of transverse bars, and cells in which the combination of uneven staining and localized swellings gives to a single bacillus the appearance of a short chain of streptococci. This diversity of structure has led to attempts to classify the various forms of *C. diphtheriae* into definite types, indicated by names, numbers or letters, and to a description of individual strains in accordance with the predominant morphological form. It is quite true that different strains of this organism differ very markedly in morphology and that a notable frequency of one particular type of cell may characterize a particular strain throughout repeated subcultures. It seems, however, very doubtful whether it is wise to assign labels to different strains of this organism on the basis of morphological characters alone. Moreover, as Goldsworthy and Wilson (1912) have shown the morphology of diphtheria bacilli varies greatly on different batches of Loeffler's serum, depending on its mode of preparation. A strain which, on one batch, develops into typical long, curved, granular forms may, on another batch, appear as short rods devoid of granules, closely resembling an ordinary diphtheroid bacillus.

The interest shown by the older workers in the morphology of the diphtheria bacillus as a criterion of differentiation from other members of the group was due mainly to the extensive use of 'stroke' cultures on Loeffler's serum. Since the introduction of blood tellurite medium reliance has been placed to an increasing extent on the appearance of individual colonies, and morphology has been degraded to a secondary position. Furthermore, each of the three types of diphtheria bacillus now recognized, *gravis*, *intermedius*, and *mitis*, has been found to possess its own more or less characteristic morphology, with the result that microscopical examination is used more to distinguish between these three types than between the diphtheria bacillus and diphtheroid bacilli.

Another feature that characterizes *C. diphtheriae* as a species, and serves to distinguish it from some, but by no means all, of the related 'diphtheroids' is the presence of the metachromatic granules described by Babes (1886) and by Ernst (1888, 1889). These granules are coloured a reddish purple when a film preparation is stained with a suitable sample of methylene blue. They may be demonstrated more clearly by the differential stain devised by Neisser, or by one of its many modifications. A single cell may contain one or more of these granules, seldom more than half a dozen, usually two or three. When only one or two are present they show a definite tendency to be situated at one or both poles.

The arrangement of the bacilli in film preparations is at least as characteristic as the form of the individual cells. Adjacent cells tend to lie at any angle to one another, forming a V or an L according to the degree of angular displacement, and groups of such pairs form characteristic clusters, resembling Chinese letters, or cuneiform writing. It would appear, from the observations of Hill (1893-1902), that this particular arrangement results from incomplete separation at the moment of division, the daughter cells remaining attached at one point, and bending on this attachment as on a hinge as growth proceeds.

Finally, it may be noted that *C. diphtheriae* provided the first instance in which true branching was demonstrated in a bacillary species. The observations of Hill showed that this appearance was not an artefact, but could be observed to take

place during the growth of the living cell. This character, among others, decided the American Committee to separate this genus from the *Eubacteriales*, and include it with some others in a family of the order *Actinomycetales*. It has, however, been noted in Chapter 2 that rudimentary branching has occasionally been observed in such typically bacillary forms as those of the genus *Bacterium*.

The morphology of other species of corynebacteria departs, to a greater or less extent, from that of *C. diphtheriae*. Sometimes, as with *C. oris* and *C. murium*, the resemblance may be so close that an experienced observer would be unable to distinguish either of these species from the true diphtheria bacillus on morphological appearances alone. Sometimes, as with *C. hofmanni*, the differences are so clear cut that little difficulty arises.

It is characteristic of many species of diphtheroids that their morphology, as displayed in films from young cultures, is far less variable than that of *C. diphtheriae*. A film of *C. xerosis*, for example, may show a marked resemblance to some of the average long forms of *C. diphtheriae*, but while pleomorphism will usually be minimal in the former it will be marked in the latter. Similarly the so-called *C. coryzae segmentosum* may be very similar to the barred form which is sometimes assumed by *C. diphtheriae*, but again pleomorphism is slight or absent.

With regard to the staining reactions of this group the bare statement that the corynebacteria are Gram positive bacilli needs some qualification. The type species retains the stain to a sufficient degree to differentiate it quite clearly from the frankly Gram negative bacteria, but it is decolorized by alcohol more easily than are many Gram positive species. The metachromatic granules, on the other hand, retain the stain tenaciously, so that moderate overdecolorization, followed by the use of a red or brown counterstain, may give a picture very similar to that afforded by the use of Neisser's stain. Of the other species within this genus, *C. hofmanni* is very resistant to decolorization, so that the application of Gram's stain, followed by a prolonged exposure to alcohol (15 mins or so), affords a useful differential criterion between these two species. Of the rest, some species behave as *C. hofmanni*, others as *C. diphtheriae*. Absence of flagella and lack of motility are characteristic of the genus, as also is the absence of capsulation.

#### Cultural Characters

With the exception of *C. acnes*, the members of this group grow on ordinary nutrient agar though to a variable degree. Many of the diphtheroids, including *C. hofmanni* develop freely, but the growth of all members is improved by the presence of natural animal protein. For several decades Loeffler's serum constituted the medium of choice. In stroke cultures on this medium there is a fairly abundant growth within 24 hours, having a moist, slightly creamy and sometimes faintly pigmented appearance. The degree of development, however, varies, and all transitions are seen between a slightly raised colourless film and a profuse succulent pigmented growth.

Since the introduction of blood tellurite medium attention has been concentrated mainly on the appearance of individual colonies, particularly in relation to the diphtheria bacillus. In 1931 McLeod and his colleagues at Leeds (Anderson, Happold, McLeod and Thomson 1931) recorded the differentiation of two types of this organism. One, which was prevalent in severe cases, they called *gravis*, the other, which was isolated from milder cases, they called *mitis*. In a subsequent report (Anderson, Cooper, Happold and McLeod 1933) they extended these observa-

tions, and concluded that certain strains which they had found previously to correspond to neither the *gravis* nor the *mitis* type constituted a third type—now generally known as *intermedius*. Robinson and Marshall (1934) at Manchester, and numerous subsequent workers, have confirmed the accuracy of these observations. The three colonial types with their associated morphological, biochemical and pathogenic characters are now generally recognized as stable varieties of the diphtheria bacillus (see Siemens 1938).

A general description of the colonial appearance of the *gravis*, *intermedius* and *mitis* types is rendered peculiarly difficult because of the absence of a standard medium. McLeod and his colleagues originally used a heated rabbit blood tellurite agar, on which the differentiation of the three variants was excellent. Later, however, it was found that some *mitis* and occasional *gravis* strains failed to develop on this medium. Numerous modifications have therefore been introduced. Rabbit and guinea pig blood in 5–10 per cent concentration give the best type differentiation but are difficult to obtain in quantity. Sheep, horse and ox blood are less satisfactory, but they are improved either by heating the medium in which they are contained or by lysis of the red cells (Neill 1937, Hoyle 1941). Against the first method is the objection, noted above, that some strains particularly of the *mitis* type, are inhibited by heated blood (Glass 1937, 1939a), against lysis there is the objection that the ability of the different variants to cause hæmolytic on blood agar plates is obscured. On the whole, therefore, we favour a medium made with unheated and unlysed blood containing sufficient potassium tellurite to prevent or restrain the growth of some of the organisms that are likely to be present in the nose or throat. A 10 per cent sheep blood 0.5 per cent glucose agar medium containing a final concentration of 0.03–0.04 per cent potassium tellurite is satisfactory in practice, though the type differentiation is not so good as on McLeod's medium.

The colonial appearances of the three types will be summarized more conveniently, along with their morphological, biochemical and other characters in a later section of this chapter (p. 462). Suffice it to say here that diphtheroid bacilli also develop on a tellurite medium, forming colonies which are usually low convex, undifferentiated, about 1–2 mm in diameter varying in colour from pearl grey to jet black, with an entire edge, and a smooth, finely granular, or liquorice type of surface. Most of them can be easily recognized, but some strains form colonies so closely resembling the *gravis*, *intermedius* or *mitis* types of the diphtheria bacillus that they can be distinguished from them only after careful study of their other characters.

Growth occurs readily in broth but is seldom abundant. The degree of turbidity, pellicle formation, and the amount and nature of the deposit vary with different members of the group, with some species, as for instance the three types of diphtheria bacillus, they are of value in identification.

Growth in gelatin at room temperature is generally poor to moderate. A few members of the group, like *C. ovis*, *C. pyogenes*, and certain diphtheroids liquefy the medium, but most members, including the diphtheria bacillus do not. Wright (Report 1942) has drawn attention to *gravis* like strains of a toxigenic diphtheroid bacillus that liquefy gelatin in slope but not in stab culture.

Resistance—*C. diphtheriae* is readily killed by heat, suspensions of the bacilli failing to survive 10 minutes' heating at 58° C. It is also easily destroyed by most of the usual antiseptics. It would appear to be relatively resistant to drying,

though the evidence on this point is somewhat conflicting. Concerning the resistance of the various species of diphtheroid bacilli we have far less precise information, such data as are available suggest that they behave, in this respect, in much the same way as the type species. An exception must, however, be made for *C. equi*, which is reported by Karlson, Moses and Feldman (1940) to be unusually resistant to certain chemical disinfectants like oxalic acid that are used in the cultivation of tubercle bacilli from contaminated material. Most vegetative organisms are destroyed within 15 minutes by exposure to 15 per cent. oxalic acid, *C. equi* is said to resist even a 5 per cent. solution for an hour.

### Metabolism and Growth Requirements

The oxygen requirements of different species within this genus vary widely. Though all species are apparently able to grow in the absence of gaseous oxygen, some species, including *C. diphtheriae* itself, develop far more freely under aerobic conditions and display this preference by growing as a film or veil over the surface of a liquid medium. Other species such as *C. acnes* and *C. typhi*, are microaerophilic.

The temperature range over which most members grow in artificial media extends from about 15° to 40° C., with an optimum at about 37° C.

It has already been pointed out that the growth of all species is improved by the addition of a natural animal protein to the medium. Both blood and serum favour growth, but some members, particularly the *mitis* type of *C. diphtheriae* are inhibited by heated blood as well as by blood treated by acid or alkali. According to Glass (1939a, b) this effect is apparent only in aerobic culture, and is abolished by the addition of sodium hydro-sulphite or by anaerobic cultivation.

So far as growth on synthetic media is concerned our information is as yet relatively scanty in spite of a considerable mass of experimental data (see Uschinsky 1893, 1897, Hadley 1907, Hadley and Gorham 1907, Koser and Rettger 1919, Davis and Ferry 1919, von Groer 1923, Hosoya and Kurova 1923, Robertson 1924, Braun and Hofmeier 1927, Braun and Mundel 1927, 1929, Braun, Hofmeier and Mundel 1929, Maver 1930, Landemann 1932, Ehrismann 1932-33, 1933, Hottinger and Hottinger 1933, Nitsch 1933, Schmidt 1933-34, Wadsworth and Wheeler 1934, Knight 1936). It may be summarized by noting (1) that *C. diphtheriae* appears to be incapable of growth with ammonia as the sole source of nitrogen, carbon being supplied in an organic form, (2) that growth, and toxin production often occur when amino-acids are added to such a synthetic medium, and that among these amino-acids, cystine, aspartic acid and perhaps tryptophan appear to be indispensable, and (3) that it is only certain 'non-exacting' strains that are capable of growing in such a medium, and even with these strains toxin production is usually much less abundant than with a more adequate food supply. Knight (1936) emphasizes the difficulty of assessing the real significance of many of the recorded findings in view of the great difficulty of obtaining most amino-acids in a state of chemical purity.

Attempts have been made (Bunker 1919, Hosoya *et al.* 1933, Mueller *et al.* 1933, Mueller 1935a, b, c) to identify the additional substances required for active growth and toxin production by fractionating peptone, meat extract and other types of complex protein extracts or hydrolysates in which *C. diphtheriae* is known to grow abundantly. Considerable progress has been made along these lines (see Chapter 3).

### Biochemical Reactions

The carbohydrates most frequently employed as test substrates for the differentiation of species within this genus are dextrose, maltose and saccharose. The type species, *C. diphtheriae*, produces acid but no gas in dextrose and maltose, but does not attack saccharose. Other species, such as *C. zosteris* produce acid in all three

sugars, others again, such as *C. hofmanni*, attack none of them. No species produces gas. Within the species *C. diphtheriae* the *gravis* type ferments starch and glycogen, the *mitis* and *intermedius* types do not. In practice great care is necessary in the preparation of the medium if reliable results are to be obtained. A medium containing serum is almost indispensable, but it has certain drawbacks. The serum, for example, may contain sufficient fermentable carbohydrate to give a false reaction, this can be overcome by suitable buffering. Unheated horse serum may hydrolyse starch due to the presence of a natural diastase (Hendry 1938) this may be destroyed by heating. Not all samples of soluble starch are satisfactory, each sample should be tested before use.

A suitable medium may be prepared by dissolving 0.5 per cent peptone and 0.1 per cent  $\text{Na}_2\text{HPO}_4$  in 1,400 ml. of distilled water, steaming for 15 minutes, filtering, adjusting to pH 7.4, adding 250 ml. of horse serum, steaming for 20 minutes, adding 11 ml. of Andrade's indicator, adjusting to pH 7.6-7.8, tubing in 3 ml. quantities, autoclaving at 10 lb. for 10 minutes and adding to each tube separately sufficient of a sterile solution of the sugar in distilled water to give a final concentration of 0.4 per cent starch or 1 per cent of other sugars (Robinson 1940).

A more detailed and extended table of the fermentation reactions of ten named species within this genus, and of eleven unnamed diphtheroids examined by Barratt (see Andrewes *et al.* 1923) is appended to this chapter, but experience has revealed the existence of several more fermentative types. The fermentation of certain other substrates by different varieties of the type species is considered below.

We may note here that certain species and types within this group, for instance *C. ovis*, *C. pyogenes* and certain unnamed diphtheroids liquefy gelatin, while *C. diphtheriae* and most diphtheroid organisms do not.

Many strains of *C. diphtheriae*, but not all, produce areas of hæmolysis on blood agar plates, and lyse red cells when these are added to broth cultures (Schwoner 1904, Costa *et al.* 1918, Goldie 1933). The red cells of the guinea pig are the most sensitive, then those of the rabbit, horse, man, pig, mouse and sheep in this order. There is a conflict of evidence in regard to the production of a soluble hæmolyisin. The earlier observers stated that culture filtrates did not cause hæmolysis, but Goldie (1933) reports that cell free filtrates are hæmolytic, and that their activity runs roughly parallel to their toxin content, though the hæmolyisin is not neutralized by antitoxin. There is a similar conflict in regard to the heat resistance of the hæmolytic agent, whether it be intracellular or extracellular. It was originally recorded as being inactivated by heating to 58° C. for half an hour. Goldie states that it is not inactivated by boiling. Lysis around colonies on blood agar is of some value in distinguishing between the three types of the diphtheria bacillus. Most *mitis* strains are hæmolytic, *intermedius* strains are not, and *gravis* strains vary in their activity.

Two species of diphtheroids that are pathogenic for animals, *C. ovis* and *C. pyogenes*, also exert a hæmolytic action on the red blood corpuscles of various species including the rabbit and horse. The other species and types within this genus that have been examined from this point of view appear to be non hæmolytic.

Some diphtheroid strains form phosphatase, the true diphtheria bacillus never does. The production of this enzyme may be tested for on a medium containing phenolphthalein phosphate (see Bray 1944).

antigenic relationship between these species, or between either of them and any serological type of *C. diphtheriae*. Similarly, Bailey (1925) studied two strains of toxigenic *C. diphtheriae*, two of *C. hofmanni* and three of *C. xerosis*, together with seventeen non-toxicogenic strains of *C. diphtheriae*. He was unable to detect any significant antigenic relationship between these two species of diphtheroids and the diphtheria bacillus.

The fact that agglutination tests fail to reveal any antigenic relationship between *C. diphtheriae* and the various diphtheroid organisms, does not, of course, mean that there is no sharing of antigenic components between them. It means only that, if such sharing exists, the shared antigens, either because of their position in the bacterium or for some other reason, are not concerned in the agglutination of the intact bacterial cells. It is not, therefore, surprising to find that certain antigenic components are distributed widely among the corynebacteria, and even among related genera. Thus Krah and Witebsky (1930) record that alcoholic extracts of the diphtheria bacillus, of certain diphtheroids and of the tubercle bacillus all fix complement in the presence of an antiserum prepared against any one of these organisms. Again, a strain of *C. diphtheriae* that has undergone antigenic variation and lost its type specific surface antigen may agglutinate with antisera prepared against diphtheria bacilli of other serological types or against various diphtheroids. Neill and his colleagues (1931) have described such a strain.

### Toxin Production and Pathogenicity.

The type species, *C. diphtheriae*, is an important human pathogen giving rise to a characteristic and often fatal disease, the lesions of which are, in the main, produced by the action of a powerful exotoxin. This diffuses throughout the body from the primary focus of infection, which is most frequently situated in the tonsillar region. The pathogenesis of diphtheria in man, and its diagnosis, prevention and treatment so far as these depend on bacteriological methods, are dealt with in Chapter 61. The characters of the specific toxin, its effects on certain laboratory animals, and its immunological relationships with toxins produced by other species and types within this genus, are, however, so important as differential criteria, that it is necessary to discuss them here.

The production of diphtheria toxin in artificial culture has been the subject of a large series of empirical observations. From these it is clear that optimal toxin production demands conditions which are not necessary for optimal growth.

Davis and Ferry (1919) tested the value of media prepared from beef infusion, peptone, and meat extract in various combinations with each other. The presence of beef infusion was found to be essential. This was later disproved by Wadsworth and Wheeler (1934) who devised a synthetic medium containing 2 per cent of peptone on which a toxin of high potency can be prepared (Eaton 1936, Pappenheimer and Robinson 1937). Hartley and Hartley (1922) tested various specimens of peptone, and found that while each brand of peptone gave a characteristic curve of toxin production, it was impossible to predict the value of any one brand by a preliminary chemical analysis. Hartley (1922) pointed out the superiority of a tryptic digest of horse muscle as a medium for the production of toxin, and his findings have been confirmed and extended by Watson and Langstaff (1927) who also confirm his observation that the value of such a medium is markedly influenced by the method of sterilization. In addition to growth promoting substances, there are apparently toxin inducing substances present, which are very labile to heat when the pH is at neutrality or on its alkaline side. Autoclaving at pH 8.0 or over may completely destroy the value of a medium for toxin production. This may be avoided by filtration through a Seitz press followed by a short steaming. The addition to the medium of maltose, or certain other energy sources considerably increases the toxin yield, particularly if sodium acetate or lactate is also added (Pope 1932, Ramon and Berthelot 1932, Pope and Smith 1932, Pope and Healey 1933a, Pope and Langgood 1939). The



iron content of the medium has to be carefully adjusted (Pappenheimer and Johnson 1936). The initial reaction of the medium is of great importance, a fact which has been emphasized by many workers (Bunker 1919 Hartley 1922 Andrewes *et al* 1923 Watson and Langstaff 1927). The most favourable starting reaction is at or just below pH 8.0 and it is important that during the 7 to 10 days which elapse between sowing the culture and harvesting the toxin the reaction should not swing far towards neutrality. The range over which growth of *C. diphtheriae* takes place extends from about pH 5.7 to pH 8.7 but the zone over which toxin production occurs appears to be limited to pH 7.5-8.2. The growth of the organism during its initial stages is associated with a slight production of acid probably derived in part from nitrogenous constituents of the medium. Later there is a reversion in the alkaline direction due to the splitting up of these organic acids with the formation of carbonates. The balance between these metabolic activities is in part determined by the oxygen pressure to which the culture is submitted. Partly for this reason partly perhaps for others the shape of the flask in which the medium is contained the thickness of the layer of medium itself and the type of plug used for closing the mouth of the flask all exert an influence on the grade of toxin produced. It is also important to eliminate any movement which will prematurely break up the veil of growth which forms at the surface of the medium. In practice it is found that the best results are obtained by growing the organism in a shallow layer of medium in a cylindrical bottle which is kept lying on its side and is plugged loosely with gauze or cotton wool (Bunker 1919 Hartley and Hartley 1922 Andrewes *et al* 1923 Watson and Langstaff 1927 Pope and Healey 1933b). The cultures are incubated at 37° C for 7 to 10 days at the end of which time phenol, or preferably toluol, is added in sufficient strength to ensure sterilization the flasks are allowed to stand for 24 hours and the contents are filtered. The crude toxin so obtained has now to be freed from the various non-toxic substances such as toxoids and substances derived from the bacterial cells and the medium with which the pure toxin is contaminated. Progress has been made in the attempt to isolate diphtheria toxin in a pure form but complete success has not yet been achieved. This is in part due to the fact that the toxin is very labile, and is readily inactivated or destroyed by heat or strong chemical reagents. It is probable that the specific material of a toxic filtrate—toxin and toxoid—constitutes a very small fraction of the total constituents of the crude filtrate (about 1 per cent according to Glenny 1923a). A considerable concentration can be achieved by precipitation with weak acids, or ammonium sulphate or acetone or by dialysis, or by fractional filtration through graded collodion membranes, or by adsorption on to aluminium hydroxide followed by elution, or by a combination of these methods (see Glenny and Walpole 1915 Watson and Langstaff 1926 Locke and Main 1928 Lenher *et al* 1931 Bunney *et al* 1931 Schmidt 1931 Schmidt *et al* 1931 Tasman and Pondman 1931 Brandwijk and Tasman 1937 1933 Zaydel 1937 Tasman and van Waasbergen 1932 Wadsworth *et al* 1937 Leonard and Holm 1933 Wadsworth and Quigley 1934 Eaton and Bayne-Jones 1934 Goldie 1934) Eaton (1936) however has shown that the toxin is denatured by acid, and therefore recommends a method of purification which does not necessitate the use of this reagent.

The evidence so far obtained suggests that diphtheria toxin is a heat-coagulable protein. The purest preparations contain about 16 per cent nitrogen 0.75 per cent sulphur 9 per cent tyrosine and 1.4 per cent tryptophan. The isoelectric point is pH 4.1. The toxin is extremely sensitive to denaturation by solutions more acid than pH 6 and by moderate heat. The amount of nitrogen per flocculating (Lf) unit is 0.00045 mgm. and the MLD for guinea pigs is about 0.0001 mgm. (Eaton 1936 Pappenheimer 1937 Pappenheimer and Robinson 1937). It is actively antitoxic and when injected into animals, stimulates the production of a powerful antitoxin. It can be converted into toxoid by suitable treatment with formalin, in this state it will cause a precipitate when mixed with specific anti-

serum and will give rise to antibody production in animals, but is no longer toxic. It appears to be produced within the bodies of the bacilli since toxin is liberated in bacterial suspensions submitted to treatment with sonic vibration (Morton and Gonzalez 1912).

The classical paper in which Loeffler (1884) first described the isolation and characters of the diphtheria bacillus, and the report by Roux and Yersin (1888) of the separation of the filtrable toxin, contain descriptions of the lesions produced by the living organism, or by its separated toxin in a variety of laboratory animals. These original observations have since been extended by a host of experimental studies. It will suffice to note here that, among laboratory animals, the guinea pig and the rabbit are the most susceptible, while rats and mice are extremely resistant. Dogs, cats, pigeons and other birds appear to occupy an intermediate position (See Loeffler 1884, 1890, Roux and Yersin 1888, Wernicke 1893, Goodman 1907, Coca *et al.* 1931, Glenny and Allen 1922, Andrewes *et al.* 1923). It may be noted that the bacillus appears to have little power of tissue invasion, whether the inoculum consists of a living culture, or of a toxic filtrate, death occurs as the result of a toxæmia in the strict sense. This general statement may require minor modification in regard to certain varieties of the diphtheria bacillus (see below). For our immediate purpose it will suffice to note the sequence of events that follow the injection of a living culture, or of a toxic filtrate, into the guinea pig.

If a guinea pig is inoculated subcutaneously into the flank with a dose of a virulent culture or of a toxic filtrate of a size which will produce death within a few days, a soft cedematous swelling usually appears at the site of inoculation within 12 to 18 hours and gradually extends. About the time the swelling appears or shortly thereafter the animal becomes obviously ill, developing a staring coat and sitting crouched in its cage. Death usually occurs between 18 and 96 hours according to the size of the dose of culture or filtrate inoculated. With very large doses the time to death may be even shorter but is never less than 10 to 14 hours (see Glenny 1925b). Animals that survive beyond the 4th day may develop cachexia and paralysis and die at some later period, but the pathogenesis of this condition appears to be essentially different from that of the acutely fatal toxæmia and it is with the latter that we are here concerned. When a guinea pig that has died within 4 days after a subcutaneous inoculation is examined post mortem, the typical findings are as follows.

At the site of inoculation is found an extensive area of gelatinous hæmorrhagic oedema extending to the skin superficially, and deeply to the muscles or to the parietal peritoneal membrane. If the animal has survived for several days the tissues in the more central parts of the cedematous area may be obviously necrotic. The regional lymph glands are usually swollen and congested. The peritoneum may contain a varying amount of fluid, which may be clear, cloudy or blood stained. The abdominal viscera as a whole are congested, but the most striking lesion is the marked swelling and congestion of the adrenal glands. On macroscopical section there are seen to be scattered hæmorrhages situated in the medulla in the cortex or in both. Sometimes all naked-eye distinction between cortex and medulla is lost. On opening the thorax a serous exudate will often be found in the pleural cavities, usually clear sometimes cloudy or blood stained. A pericardial effusion may or may not be present. Films prepared from such effusions reveal a marked preponderance of mononuclear cells.

It is of some interest to note the relative frequency of the more important lesions associated with acute diphtheritic toxæmia in the guinea pig. Wright (1894) records the findings in 160 necropsies: a local lesion was present in 90 per cent. of the animals, congestion of the adrenals in 81.2 per cent., and a pleural effusion in 42.5 per cent. Barratt (1923) records the post mortem findings in 50 guinea pigs which died within 72 hours after

sometimes it has been apparently healthy. It is exceedingly difficult, from the published reports, to determine whether these organisms have, or have not, played any pathogenic role, and in most instances they have not been studied in sufficient detail to allow of any systematic identification or classification. We shall therefore confine ourselves here to a brief description of the lesions produced by the named species referred to above.

*C ovis*, often referred to as the Preisz Nocard bacillus, (see Nocard 1889, Preisz 1894) causes caseous lymphadenitis in sheep and ulcerative lymphangitis in horses. It differs sharply from *C diphtheriae* in that it is a pyogenic organism, and invades the tissues. It resembles *C diphtheriae* in producing a filtrable toxin.

Nicolle, Lousseau and Forgeot (1912) have carefully recorded the lesions met with in guinea pigs, which have died as the result of inoculating either living cultures of *C ovis* or bacteria free filtrates subcutaneously. In the former case, and where the dose of living culture has been so adjusted that the animal dies about the 25th day, subcutaneous abscesses develop in various situations during life. At necropsy, in addition to these superficial lesions, small granulomatous masses are found in the liver, spleen and lungs and beneath the parietal peritoneum. In the male guinea-pig similar lesions are found in the tunica of the testis and epididymis. Some of these lesions may have developed into large caseous or caseo-purulent masses.

When a guinea pig is injected subcutaneously with a fatal dose of a toxic broth filtrate death occurs within a few days, often in less than 24 hours, from an acute toxæmia. The necropsy findings in such cases are entirely different from those described above. There is a local, subcutaneous, inflammatory, gelatinous oedema at the site of inoculation, often hæmorrhagic in character. The abdominal viscera are congested, and often show small hæmorrhages, particularly in the stomach, large intestine, and kidneys. The latter may be almost black in colour. There is, however, no congestion of the adrenals and no exudation into the pleura. Hall and Stone (1916) give a very similar picture.

Again, Petrie and McClean (1934) state that the effect produced by the injection of *C ovis* toxin into the skin of a guinea pig differs from that produced by the injection of diphtheria toxin. The former gives rise to a definitely papular lesion, and if the dose of toxin injected is large the lesions become pustular. According to Carne (1940) who defines the optimal conditions for toxin production, guinea pigs, rabbits, sheep, goats, pigs, horses, oxen, dogs and cats are all sensitive to its action.

It has been shown by Bull and Dickinson (1935) that the pyogenic substance is largely contained in the bacterial cells, as is suggested by the observations of Nicolle and his colleagues, and that it is relatively thermostable. Suspensions of *C ovis*, killed by heating at 60° C for 1 hour, no longer produce toxic death in susceptible animals, but they give rise to sterile abscesses when injected in adequate dosage.

It may be noted that the exotoxin of *C ovis* is different from that of *C diphtheriae*. Nicolle and his colleagues found that an antitoxin prepared against the former gave specific protection against *C ovis* toxin, whereas none was afforded by diphtheria antitoxin. Dassonville (1907), Hall and Stone (1916), Minett (1922a, b), and Barratt (1933) recorded some degree of protection by diphtheria antitoxin against the toxin of *C ovis*, but the detailed study of Petrie and McClean (1934) leaves little doubt that these effects were due to the fact that the sera of normal horses may contain varying amounts of *C ovis* antitoxin (*C ovis* is a natural pathogen of the horse), and that the two toxins are immunologically quite distinct from one another.

Petrie and McClean have, however, found evidence of the existence of varieties of diphtheroid bacilli that are, in respect of certain characters, intermediate between *C diphtheriae* and *C ovis*. These diphtheroids, all isolated by various observers from the

the injection of 2 ml. of a virulent culture, oedema, of varying degree, was present at the site of inoculation in 94 per cent. of the animals; the adrenals were abnormal in all and are noted as pink in 4 per cent., red in 22 per cent., and deep red in 74 per cent.; a pleural exudate was present in 44 per cent.

For details of the histological changes associated with these lesions, reference may be made to the monograph published by the Medical Research Council. We may, however, note a few points which have a direct bearing on diphtheria as it occurs in man. Mollard and Regaud (1895) recorded the occurrence of degenerative changes in the myocardium in experimental diphtheria, and Flexner (1897) noted that fatty degeneration of the cardiac muscle was almost constantly present in animals which died within a short time after inoculation. There has been some discussion as to whether these changes are primary, or are a sequel to an initial reaction in the interstitial tissue, or to a primary lesion of afferent nerve fibres. The careful and detailed studies of Dodgeon (1906) gave a clear answer to this question, and afforded strong experimental support to the suggestion of Bolton (1905), that the direct action of diphtheria toxin on the cardiac muscle is the most important cause of acute cardiac failure in human diphtheria. Examining a large series of guinea-pigs, killed or dying in various stages of acute diphtheritic toxæmia, Dodgeon demonstrated the occurrence of fatty degeneration of the diaphragmatic muscle within 4 hours after inoculation, and of the cardiac muscle within 16 hours. Similar results have since been recorded by Jaffe (1920).

We may note also, since this method is now frequently employed in the identification of a toxigenic strain of *C. diphtheria*, that the intradermal injection of toxin, or of living bacilli, leads to a localized erythematous lesion, followed by necrosis (Römer 1909). This effect, as also the lethal action of the subcutaneous injection of larger doses, can, of course, be specifically neutralized by an antitoxic serum.

Although the production of this filtrable toxin, with its characteristic action in the guinea pig and its property of being specifically neutralized by the homologous antitoxin, is one of the most important characters by which *C. diphtheria* is identified, there exist strains of bacilli that, while conforming in all other respects with the diphtheria bacillus, fail to form this filtrable toxin. These strains are commonly classed as non toxigenic, or avirulent, diphtheria bacilli. Whether they should in all cases be assigned to this species is perhaps doubtful, but there can be no reasonable doubt that many of them, at least, are actually non toxigenic variants of *C. diphtheria*. We have noted above that certain avirulent strains can be shown to be antigenically related to typical toxigenic strains, and the actual emergence of an avirulent variant from a virulent organism, under laboratory conditions, has been recorded by several observers.

Thus Crowell (1926), starting from a single-cell culture of a fully virulent strain, derived from this parent culture a series of daughter strains, one of which was entirely avirulent. All attempts to raise the virulence of this variant were without result. Cowan (1927) records the derivation of avirulent variants from 2 strains of virulent *C. diphtheria*, one of them the classical "Park 8" which has yielded toxin to most laboratories in the world. These variants were "rough," in the sense that they formed small, raised, dense and granular colonies, and gave increased deposit in broth, with an absence of pellicle formation.

The diphtheria bacillus is not the only species of *Corynebacterium* that is pathogenic under natural conditions. *C. oris*, *C. murium*, *C. pyogenes*, *C. equi* and *C. renale* certainly fall into this category. So probably does *C. acnes*.

A great variety of diphtheroid organisms have, at one time or another, been isolated from various tissues in man and animals. Sometimes the tissue from which the diphtheroid bacillus was isolated has been the site of some obvious lesion,

sometimes it has been apparently healthy. It is exceedingly difficult, from the published reports, to determine whether these organisms have, or have not, played any pathogenic rôle, and in most instances they have not been studied in sufficient detail to allow of any systematic identification or classification. We shall therefore confine ourselves here to a brief description of the lesions produced by the named species referred to above.

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Petrie and McClean have however found evidence of the existence of varieties of diphtheroid bacilli that are, in respect of certain characters, intermediate between *C. diphtheriae* and *C. ovis*. These diphtheroids, all isolated by various observers from the

human throat were originally studied by Barratt (1933) who found them to have the power of liquefying gelatin a property possessed by *C. ovis* but not by *C. diphtheriae*. Petrie and McClean found that one of these strains produced both diphtheria toxin and a toxin that was immunologically related to that of *C. ovis*. The remaining strains produced only a toxin related to that of *C. ovis*. Two strains, similar to those of Barratt have been described more recently by Wright at Liverpool (Report 1942). Both strains resembled the *gravis* type of diphtheria bacillus in their colonial appearance, fermentation of starch and virulence to the guinea pig but differed from it in liquefying gelatin in slope culture at 22° C though not in stab culture at 22° C or 37° C in being inagglutinable with *gravis* type antiserum and in being apparently unaffected by diphtheria antitoxin even 10 000 units of antitoxin failed to protect guinea pigs against the usual test dose of culture and the organisms were easily recoverable from the heart blood. One strain killed a rat injected intraperitoneally. Whether the toxin formed by these strains was neutralized by *C. ovis* antitoxin is not recorded.

*C. pyogenes* was first described by Lucet (1893) who isolated it from suppurative lesions in cattle. Similar organisms were isolated from similar lesions in cattle and swine by Grips (1895), Kunnemann (1903) and Glage (1903). Since then they have been recorded by many workers (see Holth 1908, Poels 1912, Ward 1917). The lesions to which they most frequently give rise under natural conditions appear to be suppurative pneumonia, suppurative arthritis and other suppurative lesions including mastitis, the animals affected being cattle, pigs, sheep and goats (see Merchant 1935, Magnusson 1938).

Among laboratory animals the rabbit appears to be most susceptible, guinea pigs less so and mice relatively resistant (see Holth 1908, Ward 1917, Brown and Orentl 1920).

The injection of living cultures of *C. pyogenes* into the rabbit is followed by the development of localized abscesses if the injections are given subcutaneously. If they are given intravenously or if generalization occurs after a subcutaneous inoculation abscesses develop in the bones and joints less frequently in other organs.

*C. ovis* and *C. pyogenes* produce suppurative lesions in animals, both liquefy gelatin and both produce hæmolytic. There is however a general consensus of opinion among those who have worked with them that they are different species. They tend to affect different animal hosts and among laboratory animals the guinea pig is very susceptible to infection with *C. ovis* but relatively resistant to *C. pyogenes*. There are also cultural differences. *C. pyogenes* liquefies gelatin rapidly and constantly, *C. ovis* slowly and irregularly. On a medium containing blood serum *C. pyogenes* gives small dewdrop colonies which slowly enlarge and become granular in the centre. *C. ovis* gives colonies that are circular, umbonate and opaque with a tendency to develop a yellowish pigment. Both organisms form a filtrable toxin. The toxin of *C. pyogenes* reaches its maximum concentration in culture after 48 hours at 37° C (Lovell 1937, 1944) that of *C. ovis* in about 5 days (Carne 1940). Lovell finds that the toxin of *C. pyogenes* is apparently identical with the hæmolytic but the toxin of *C. ovis* according to Carne (1939) is unrelated to the hæmolytic. It would appear that the toxins of the two organisms are distinct, the results of cross protection tests with antitoxin are however still awaited.

An organism that has sometimes been confused with *C. pyogenes* is *C. equi* which was isolated by Magnusson (1923) in Sweden from foals affected with pyæmia.

The differential characters of the two organisms are enumerated on p. 469 under *C. equi*.

Enderlen (1890-91) described a diphtheroid bacillus that he had isolated from the pus from a cow suffering from pyelitis. This organism has been named *C. renale*. A similar organism was isolated by Ernst (1905, 1906), and Jones and Little (1923) record an outbreak of infective cystitis and pyelitis in three dairy herds associated with the constant presence of a diphtheroid bacillus. In this last instance, at least, there would seem to be no reasonable doubt that the bacillus was aetiologicaly related to the disease, though the same organism may often be isolated from the genital tract of healthy calves (Jones and Little 1930). There are as far as we know no records of the lesions produced by this organism in laboratory animals though Jones and Little have reproduced the disease in cattle. Ernst, indeed states that the bacillus isolated by him produced no lesions in any animal and he therefore regarded it as devoid of pathogenic significance. No attempt has apparently been made to ascertain whether it produces a filtrable toxin. Its claim to rank as a separate species must rest in part on its predilection for the urinary tract in cattle but mainly on its behaviour in the laboratory. It is recorded as not producing a haemolysin and not liquefying gelatin in both of which characters it differs sharply from *C. pyogenes* (Merchant 1935).

*C. murium*, which was first described by Kutscher (1894) gives rise to a natural disease in mice, and its pathogenic activity is apparently confined to this small laboratory animal. It has been injected without effect into the guinea pig, rabbit, cat, dog, pigeon, hen, rat, goat, calf, sheep, cow and horse (Kutscher 1894, Bongert 1901), but Gundel, Gyorgy, and Pagel (1932) have recorded spontaneous infections in rats on a vitamin deficient diet. We may give here a brief description of the natural disease as well as of the results of experimental inoculation.

The natural disease has been described by Kutscher and by Bongert and has been observed on many occasions by the present authors during necropsies on mice though it is certainly relatively infrequent. The most characteristic lesion in the naturally occurring disease is the presence of large firm caseous areas in the lung. In sections or films from these lesions the bacilli are usually abundant. Caseous nodules may be found in the liver though they are less frequent. When present they project from the surface in contrast to the necrotic areas seen in mouse typhoid. The lymphatic glands of the axilla, neck, mediastinum and mesentery may be enlarged and caseous but the pulmonary lesions are frequently the only obvious sign of disease. Occasionally the bacillus may be isolated from a single caseous gland, found at necropsy without any other detectable lesion.

The disease may readily be reproduced by inoculating mice with pure cultures of *C. murium*, or by administration *per os*. The findings at necropsy depend largely on the route of administration. After feeding lesions develop in the mesenteric glands and in the liver. After intraperitoneal inoculation, which usually leads to death within a week the peritoneum is found to be studded with minute tubercles and there is a spreading granulomatosis of very varying extent involving the regional lymphatic glands, the liver, and less frequently the spleen. In our experience pulmonary lesions are much less frequent in the experimental than in the natural disease though they occasionally occur. Bongert (1901) has called attention to the trivial lesions which may sometimes be found post mortem after experimental infection. In animals dying after subcutaneous inoculation the only detectable lesion may be a small caseous abscess at the site of inoculation. Seeking an explanation for this fact he inoculated mice with filtrates of broth cultures or with cultures killed by heat, and found that death resulted in every case after about 10-14 days. No obvious lesions of any kind were found at necropsy. Mice fed with filtrates died in about the same time and with the same absence of lesions. We can in part confirm these findings

as regards the inoculation of filtrates, or of killed cultures, though our own results were far less uniform than those recorded by Bongert. It would appear that this organism produces an exotoxin which is fatal for mice. According to Bongert this toxin is relatively heat-stable since it withstands heating for 9 hours at 50° C. or for a few minutes at 74° C. but the particulars given are not sufficiently precise to allow of any definite conclusion with regard to the time or temperature required for inactivation.

The toxin of this organism has not, so far as we are aware, been compared with that of *C. diphtheriae* or of *C. ovis* but it seems exceedingly unlikely that there is any relationship, since the mouse is conspicuously resistant to diphtheria toxin, and the guinea pig and rabbit, which are very susceptible to *C. ovis* are resistant to *C. murium*.

It may be noted that Fischl, Koech and Kussat (1931) described an organism under the name of *Corynebacterium arthritis muris* which they isolated from the swollen ankle joint of a white mouse. On inoculation into the joints of normal mice and rats it gave rise to a similar arthritis. Little is known about this organism, but it appears to differ in some respects from *C. murium*.

There remains *C. acnes* a diphtheroid organism described in the lesions of cutaneous acne by Unna (1896). It was first isolated by Sabouraud (1897) and has since then been studied by Gilchrist (1900-1903) by Fleming (1909) and by Salmersen and Thompson (1909-10). Its claim to pathogenicity must rest in the main on its constant association with the disease in man. Salmersen and Thompson state that the two strains examined by them were pathogenic for the mouse but not for the guinea pig but their description of the lesions in the latter animal is extremely scanty. *C. acnes* is clearly marked as a distinct species from those described above by its peculiar growth requirements (see p. 471) and particularly by the fact that it is microaerophilic.

*C. typhi* another microaerophilic diphtheroid, was isolated by Plotz (1914) from the blood in typhus fever. It is now generally admitted to be an example of a parasitic diphtheroid with no established pathological role (Olitsky 1921).

### The Gravis, Mitis and Intermedius Types of *C. diphtheriae*

Attention has already been drawn on p. 450 to the recognition by McLeod and his colleagues at Leeds (Anderson, Happold, McLeod and Thomson 1931) of three stable varieties of the diphtheria bacillus—the *gravis*, *intermedius* and *mitis* types. The characters of these three types as recorded by Robinson (1934-1940) and Cooper, Happold, McLeod and Woodcock (1936) are summarized in Table 30.

This summary can do no more than serve as a guide to the recognition of the three types. On the whole the characters of the *intermedius* type are the most constant. In the identification of *gravis* strains starch fermentation is of particular value since though it may be delayed beyond 24 hours, it is seldom absent.

Sometimes it is difficult to distinguish not only between the three types, but between diphtheria bacilli and diphtheroids. Each of the types has one or more species of diphtheroid bacilli that closely resemble it. Space does not permit a detailed description of the differential characters of these organisms. In general, it will be found that the diphtheroid bacilli tend morphologically to be more regular in shape, size, depth of staining, distribution of bars or granules, and arrangement. Diphtheroids lie often in palisades, whereas true diphtheria bacilli show the Chinese letter type of distribution, or if they are in bundles, they have seldom the same regularity of arrangement as that of diphtheroids. The colonial differences are often very slight and, as they vary from one medium to another they can be learned only by close observation and long experience.



TABLE 30

CHARACTERS OF THE TYPES OF *DIPHTHERIA BACILLUS*

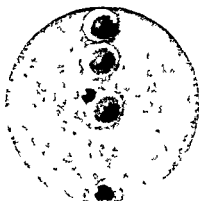
	Gravis	Intermedius	Mitis
Morphology on tellurite blood agar	Usually short rods, resembling irregular Hofmann. Staining fairly uniform, few or no granules and often a narrow unstained bar dividing the rod unequally. Some degree of pleomorphism with irregularly barred snow shoe and tear drop forms. May be coccoid on first isolation. Occasional strains resemble <i>intermedius</i> or <i>mitis</i> type.	Usually long irregularly barred rods often with terminal clubbing. Granulation generally poor. Pleomorphism always present. Distinguished from diphtheroids which may closely resemble them by irregularity of size, shape, barring and arrangement. Some strains indistinguishable from <i>mitis</i> .	Usually long, curved pleomorphic rods with prominent metachromatic granules. Except for some shadow areas protoplasm stains evenly. Some strains show barring with or without granules. Occasional strains are coccoid and others yeast like.
Colonial appearance on tellurite blood agar	An 18 hr colony is 1-2 mm. q., circular, low convex, pearly grey or with greyish black centre and paler semi-translucent periphery, with a smooth matt or rarely liquorice type of surface, and a commencing crenation of the edge. The colony is coherent, tending when touched, to move as a whole on the surface of the medium, has a consistency of cold margarine, and breaks up radially into small masses that are not easily emulsified. Slight hæmolytic strains. In 2-3 days colony reaches 3-5 mm in diameter, is flattened with a slightly raised centre, is slaty grey or greyish black in colour, often darker at the centre than at the periphery, has a frosted surface and a crenated edge and shows radial striation, especially towards the margin. When striation and differentiation are well developed, the term 'daisy head' colony is applicable.	Colonies are uniform, small, discrete, delicate, almost misty in appearance and undergo little increase in size between 24 and 48 hours. At 18 hours the colony is less than 1 mm q. is slightly raised, with or without umbonation or is of sugar loaf appearance, centre is greyish black and generally darker than periphery. Surface smooth or very finely granular, and edge entire or slightly spiky. Consistency is intermediate between the brittle <i>gravis</i> and the butyrous <i>mitis</i> type. Hæmolytic strains never seen. At 48 hours colony is not much larger, has a dull granular centre and a smoother more glutinous periphery and is dark in colour except for a lighter ring near the edge—frog's egg appearance. Edge may be entire or finely crenated. On further incubation colony may enlarge and come to resemble a daisy head colony of the <i>gravis</i> type.	Very variable in size, usually ranging between <i>intermedius</i> and <i>gravis</i> . At 18 hours colonies may be less than 1 mm q. up to 1.5 mm q. They are circular, convex usually of a mushroom grey colour darker than that of <i>gravis</i> though varying considerably with a smooth glistening surface and entire edge. Consistency is of soft butter and emulsifiability is easy. Small ring of hæmolytic strains is usual. At 48 hours colony is 2-4 mm q. undifferentiated and dark greyish black with sometimes a narrow paler margin. On further incubation colony may become flatter with a central elevation—poached egg appearance—and the surface may become granular and contoured or develop concentric rings or papular excrescences.

TABLE 30—continued

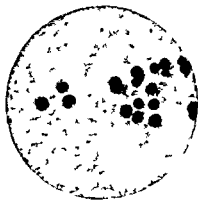
	Gravis	Intermedius	Mitis.
Growth in broth	Appearance variable Usually surface pel- licle and coarse granu- lar deposit with little or no turbidity	Appearance very con- stant. In 24 hours there is slight turbid- ity with little or no de- posit. In 48 hours broth has cleared and there is a very finely granular sediment which can be easily dispersed on shaking	Appearance variable Usually diffuse even turbidity denser than that of <i>intermedius</i> and moderate non- granular deposit. Soft pellicle may form on further incubation
Fermentation of starch and glycogen	+	—	—
Antigenic structure	At least 5 types recog- nizable by direct ag- glutination. Types I and II form the clas- sical daisy head col- onies. Types III, IV and V approach nearer to the <i>mitis</i> type.	Little known but prob- ably at least 2 types with specific antigens	Little known but at least a distinct types
Animal pathogenicity	Almost invariably viru- lent to guinea pigs.	Almost invariably viru- lent to guinea pigs.	Usually virulent to guinea pigs but strains from carriers are often avirulent

Note:  $q$  = diameter

Fermentation reactions are often of help since many diphtheroids either ferment sucrose or have no action at all on sugars. Some strains cannot be distinguished except by virulence tests and even this method leaves the true nature of an avirulent *mitis*-like organism doubtful. Whether in identifying the individual types of

FIG. 83—*C. diphtheriae*

Three colonies of *gravis* type and two of *mitis* type on blood tellurite agar ( $\times 8$ )

FIG. 84—*C. diphtheriae*

Colonies of *intermedius* type on blood tellurite-agar ( $\times 8$ )

*diphtheria* bacilli or in distinguishing between diphtheria bacilli and diphtheroid bacilli that closely resemble them too much attention should never be paid to any one character. Often it is only by observing all the characters of a particular

strain and assigning to each character a weight which experience alone can provide that a sound conclusion on its identity can be reached. Valuable help will be obtained by a careful study of McLeod's (1913) review.

### The Classification of Corynebacteria

It will be obvious from the foregoing discussion that the time has not yet arrived to attempt any general systematic classification of the large number of different types of corynebacteria that have been described. Certain organisms have however, been studied in sufficient detail to make it clear that they deserve specific rank. Among these the type species, *C. diphtheriæ*, is of course pre-eminent. Among the non-pathogenic species parasitic to man *C. hofmanni* is a well recognized species, and *C. xerosis* would probably fall into the same category. We follow Andrewes and his colleagues (1923) in excluding the so called *C. coryæ segmentosum* from the list of identifiable species. Including the various species and types, of human or animal origin that have been dealt with above, this leaves us with the following named species within the genus *C. diphtheriæ* (types, *gravis*, *mitis* and *intermedius*) *C. hofmanni*, *C. xerosis*, *C. ovis*, *C. pyogenes*, *C. equi*, *C. renale*, *C. murium*, *C. acnes* and *C. typhi*. For the rest, we are in entire agreement with Andrewes and his colleagues in believing that specific names should be withheld from the numerous diphtheroids that have been described until they have been examined in greater detail and their identity more fully established.

We append a summarized description of the named species and a tabular description of the fermentation reactions of the eleven types of diphtheroid bacilli differentiated by the Committee of the Medical Research Council (see Andrewes *et al.* 1923). Reference may also be made to a paper by Brooks and Hucker (1944) who divided 79 strains of animal diphtheroids into 3 groups on the basis of growth and biochemical reactions.

### *C. diphtheriæ*

Observed by Klebs (1883), isolated and described by Loeffler (1884).

The morphology and staining reactions of this species have been described above and the absence of motility and capsulation, common to the genus, has been noted.

**TYPE OF GROWTH.**—On Loeffler's serum, the colonies after 24 hours incubation at 37° C., are about 1 mm. in diameter, circular, convex, with a slightly raised centre, a smooth or finely granular surface and an entire edge granular in structure when viewed by transmitted light butyrous in consistency pale or deeper cream in colour moderately opaque and easily emulsifiable in water or saline. After 48 to 72 hours incubation the colony shows a varying degree of enlargement the centre becomes more raised, more opaque, and deepens in colour, while the periphery remains flat extends outwards and appears more transparent than the centre, giving the so-called poached-egg appearance.

On agar, growth is much less abundant and the individual colonies are for the most part smaller often having a diameter of 0.25 mm. or less after 24 hours incubation at 37° C. These small colonies which are greyish white in colour, convex with a raised central portion and usually with an entire margin are frequently mingled with a few larger, whiter colonies.

On gelatin, the growth is very similar to that on agar but develops much more slowly owing to the lower temperature of incubation. In stab culture growth develops along the whole length of the needle-track without lateral outgrowths and with a slight surface growth consisting of a raised central portion and a flatter periphery, sometimes showing an irregular margin. The medium is never liquefied.

On potato, growth is usually very scanty and often invisible to the naked eye.

On tellurite blood agar plates the colonial differences that characterize the *gravis*, *mitis* and *intermedius* types (see Table 30) can most readily be observed. These colonial differences are, however also observable on certain other media, such as tryptic serum agar (Dudley *et al.* 1934)

*Broth* See Table 30

*C. diphtheriae* is aerobic and facultatively anaerobic.

The optimum temperature for growth is in the near neighbourhood of 37° C. with a range from about 15° to 40° C. over which growth occurs.

Heat resistance is slight a temperature of 55° C. for 10 minutes sufficing to sterilize a suspension or broth culture.

**BIOCHEMICAL REACTIONS.**—*C. diphtheriae* ferments glucose, galactose, and maltose, with the production of acid but no gas. It has no action on saccharose, lactose or mannitol. Litmus milk is unchanged. Indole is not formed but, according to the results obtained by Fneber (1921), *C. diphtheriae* gives a colour reaction with sulphuric acid and potassium nitrite as a result of the formation of indole-acetic acid from tryptophan. This substance does not however give the colour reaction with paradimethylamido benzaldehyde which is characteristic of indole itself. Nitrates are reduced. Gelatin is not liquefied.

The *gravis* type, in addition to the carbohydrate substrates referred to above, ferments dextrin, starch and glycogen. The *mitis* and *intermedius* types give irregular results with dextrin and do not ferment starch or glycogen.

The *mitis* type is usually hæmolytic, the *gravis* type is usually non hæmolytic, and the *intermedius* type is consistently non hæmolytic.

**ANTIGENIC STRUCTURE.**—*C. diphtheriae* is divisible into a number of different antigenic types. The *gravis mitis* and *intermedius* types differ antigenically from each other and each is divisible into a number of antigenic sub-types.

**TOXIN PRODUCTION AND PATHOGENICITY.**—*C. diphtheriae* is pathogenic for man and for certain laboratory animals. It produces a powerful exotoxin with a characteristic action on the animal tissues (see above). Non virulent strains of the *mitis* type are not uncommon.

#### C. hofmanni

Von Hofmann, in 1888 isolated from the throats of normal persons a diphtheroid bacillus which was probably identical with the species which now bears his name. The incompleteness of the earlier descriptions does not allow us to identify with any certainty the various strains which were, about this time described under the general head of "pseudodiphtheria bacilli." The description which follows refers to a particular type of diphtheroid bacillus to which the name of *C. hofmanni* has been allotted by common consent. There are other forms of non fermenting diphtheroid bacilli which possess quite a different morphology. These must for the moment be left unnamed finding a temporary home in the appropriate groups of the fermentative types differentiated by the Committee of the Medical Research Council.

**MORPHOLOGY.**—Short rods, 1.5 to 2  $\mu$  in length, with parallel sides, rounded or slightly pointed ends with a straight axis, and a single unstained central septum. Metachromatic granules are usually absent if present they are few in number small and inconspicuous. There is little or no tendency to pleomorphism. The bacilli are arranged in parallel rows, or in irregular groups, with the usual angular displacement of adjacent cells (see



FIG. 30.—*C. diphtheriae*  
24 hours culture on  
Loeffler's serum

Fig 86) The bacilli are more tenacious of the Gram stain than *C diphtheria*, or than many other diphtheroids.

**GROWTH**—On *Loeffler's serum*, after 24 hours at 37° C., *C hofmanni* produces colonies which are larger than those of *C diphtheria*, and whiter in colour. They vary in diameter from 1 to 1.5 mm, and are circular, convex, smooth, and opaque, with an entire edge. They are homogeneous in structure, butyrous in consistency, and emulsify readily. After 48 hours they increase in size to a diameter of 2 mm or so, and the edge may become slightly erose. On *agar* this species, unlike *C diphtheria*, grows readily and abundantly, forming colonies very similar to those produced on serum. In contrast to *C diphtheria*, a confluent growth often occurs in primary culture, or in early subculture. In *broth* a moderate turbidity is produced, the growth gradually settling to the bottom as a powdery deposit. No pellicle is formed. In *agar* or in *gelatin stab* *C hofmanni* produces little growth along the needle track, but a profuse surface growth. *C hofmanni* is aerobic and facultatively anaerobic, the optimal temperature for growth is in the neighbourhood of 37° C. The resistance of this species to heat, or to antiseptics, has not been systematically examined, but there is no evidence that it differs in these respects from the type species.

**BIOCHEMICAL REACTIONS**—*C hofmanni* ferments none of the carbohydrates against which it has been tested, and these include all those referred to in the case of *C diphtheria*. It produces no change in litmus milk, does not liquefy gelatin, and does not produce indole. Nitrates are reduced. It does not produce hæmolytic.

**ANTIGENIC STRUCTURE**—All that is known on this point is that the antigenic make-up of such strains of *C hofmanni* as have been examined differs entirely from that of *C diphtheria* on the one hand, and from *C xerosis* on the other.

**TOXIN PRODUCTION AND PATHOGENICITY**—*C hofmanni* produces no toxin and is not pathogenic.

#### C xerosis

An organism which possessed certain of the characters which we ascribe to *C xerosis* was isolated from the conjunctiva by Reymond in 1881, and described somewhat more fully in 1883. The first detailed description of this organism was provided by Kuschbert and Neisser in 1883, if we may assume that these three records in fact refer to the same bacterial species. The original view, that *C xerosis* was ætiologically related to a particular conjunctival lesion, has been generally abandoned. Griffith (1901) states that this organism is the commonest bacterial inhabitant of the normal conjunctival sac. Andrewes and his colleagues record their doubt as to whether *C xerosis* is even now sufficiently well characterized to deserve specific rank, and their scepticism appears fully justified. The description which follows must be regarded as summarizing the characters of those strains to which most observers would allot this particular name.

**MORPHOLOGY**—The form assumed by *C xerosis*, in films from a culture grown on

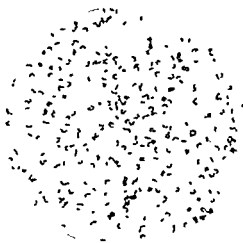


FIG 86—*C hofmanni*

From 24 hours culture on Loeffler's serum ( $\times 1000$ )

Loeffler's serum for 24 hours at 3° C. is not unlike the bacillary form of *C. diphtheriae* the differences consist in a preponderance of barred or segmented forms over the granular form the infrequency of club forms, the relative infrequency and inconspicuousness of metachromatic granules, though these are often present in small numbers, and the relatively slight pleomorphism. The cells of this species retain the Gram stain more tenaciously than those of *C. diphtheriae*.

**GROWTH.**—On *Loeffler's medium*, or on *agar* *C. xerosis* forms colonies which are smaller than those of *C. diphtheriae* or of *C. Hofmanni*; the margins may become irregular after 48 to 72 hours and the colonies tend to adhere firmly to the medium during the later stages of growth. *Broth* remains clear or shows a slight turbidity while a granular deposit forms at the bottom of the tube. No pellicle is formed. The general conditions of growth, as regards temperature, oxygen pressure etc., do not differ from those of the type species.

**BIOCHEMICAL REACTIONS.**—*C. xerosis* produces acid in glucose, maltose and saccharose but not in dextrin or mannitol. It does not acidify litmus milk, does not produce indole, and does not liquefy gelatin. Nitrates are reduced. It apparently forms no hæmolyisin.

**ANTIGENIC STRUCTURE.**—The antigenic constituents of those strains of *C. xerosis* which have been examined differ from those of *C. diphtheriae* on the one hand, and from *C. Hofmanni* on the other.

**PATHOGENICITY.**—There is no adequate evidence that *C. xerosis* is pathogenic.

### *C. oris*

(Synonym. *C. pseudotuberculosis oris*)

The Preisz Nocard bacillus (see Nocard 1889, Preisz 1894) was originally isolated from pseudotuberculous lesions in sheep. Similar organisms have been isolated from ulcerative lymphangitis of horses by many workers. It resembles the type species very closely in its general morphology and behaviour.

**MORPHOLOGY.**—Films from young cultures on *Loeffler's medium* show slender clubbed bacillary forms granular or segmented often with numerous metachromatic granules and exhibit a considerable degree of pleomorphism. The appearances are in fact indistinguishable from those presented by certain strains or cultures of *C. diphtheriae*.

**GROWTH.**—The organism grows well on *Loeffler's medium* giving colonies which are circular umbonate and opaque with a tendency towards the development of a distinct yellowish pigment. As they enlarge they often develop a series of concentric rings round the raised centre. The growth is described as peculiarly friable the colony breaking apart when touched by the needle (Hall and Stone 1916). The growth on *agar* is recorded as poor. Growth in *broth* is scanty but there is definite pellicle formation. On blood agar especially under anaerobic conditions, colonies are surrounded by a zone of hæmolysis. Moderate granular growth in gelatin with slight saccate liquefaction. Aerobic and facultative anaerobe growing best at about 3° C.

**BIOCHEMICAL REACTIONS.**—Forms acid from dextrose, maltose and glycerol. According to most workers (Hall and Stone 1916, Minett 1922a, b, Andrewes *et al.* 1923) mannitol, lactose and sucrose are not fermented, but Carne (1939) in Australia reports that about half of his 133 strains from sheep fermented these sugars. Dextrin may be weakly acidified. Carne (1939) gives the following additional reactions for his strains: M.R. weakly positive, V.P. negative, H<sub>2</sub>S formed by about half the strains. Gelatin is liquefied slowly and irregularly, nitrates reduced by some but not by all strains, indole not formed. *C. oris* produces a hæmolyisin, active against sheep, horse and rabbit corpuscles; according to Carne (1939) the hæmolyisin is thermolabile, non-antigenic, unrelated to the exotoxin, and is linked to the cells so that it cannot be obtained free in a filtrate.

**ANTIGENIC STRUCTURE.**—At present uninvestigated.

**PATHOGENICITY**—*C. ovis* is a natural pathogen of horses, sheep and perhaps cattle, and is pathogenic for rabbits and guinea pigs, but not for pigeons or fowls. It produces a soluble toxin which differs from that of *C. diphtheriae* (see above).

### *C. pyogenes*

First described by Lucet in 1893

**MORPHOLOGY**—The organism is a small, Gram positive, pleomorphic, diphtheroid bacillus, frequently assuming an almost coccoid form, staining irregularly with methylene blue, but apparently without metachromatic granules.

**GROWTH**—Scanty on plain media, but improved by addition of blood or serum. Grows aerobically and anaerobically. Optimum temperature 37° C. little or no growth at room temperature. Optimum pH 7.5. On *Loeffler's serum* *C. pyogenes* forms minute colonies in 24 hours at 37° C., which slowly enlarge, if incubation is continued until they may reach a diameter of 2-3 mm., the centre becomes granular and the medium is slowly liquefied, the liquefaction beginning as a small pit beneath each colony. On *blood agar* colonies rarely exceeding 1 mm. in diameter, are visible in 48 hours surrounded by a zone of  $\beta$  hemolysis (Brown and Orcutt 1920, Lovell 1937). In *serum broth* there is a granular growth without pellicle formation. On *gelatin* growth is slight but the medium is slowly liquefied. No growth on *MacConkey agar* or on *potato*.

**BIOCHEMICAL REACTIONS**—Acid in glucose, maltose, and later in lactose, occasional strains ferment mannitol and sucrose, dextrin and glycerol are also said to be fermented (Magnusson 1938). Litmus milk acidified and clotted within 3 days, clot is later digested. Gelatin coagulated serum, and coagulated egg albumin are gradually liquefied. Indole negative. Nitrates reduced to nitrites. Filtrable haemolysin produced which is most active on horse and rabbit corpuscles and which is destroyed at 56° C. in 30 minutes, appears to be identical with the toxin, reaches its maximum concentration in culture in 48 hours at 37° C. (Lovell 1937, 1941).

**RESISTANCE**—Rapidly killed at 57° C., and very sensitive to disinfectants (Brown and Orcutt 1920).

**ANTIGENIC STRUCTURE**—Brown and Orcutt studied 12 strains without finding any sharp difference in antigenic behaviour between them. Lovell (1937) studied 33 strains and except for 5 strains which had been subcultured for some years and which contained a major and a minor antigen, found that, irrespective of animal origin they appeared to be antigenically homogeneous.

**PATHOGENICITY**—Under natural conditions produces suppurative lesions in cattle, pigs, sheep, and goats but not in horses, is also pathogenic for the rabbit, producing suppurative lesions including arthritis. Experimentally, mice inoculated intraperitoneally with 100-1 000 million organisms die in a week with abscess formation in the omentum and liver. Rabbits inoculated subcutaneously develop localized abscesses. Inoculated intravenously they develop abscesses, particularly in the bones and joints. *C. pyogenes* forms a weak toxin which inoculated intravenously into rabbits in a dose of 1 to 5 ml., produces convulsions and death within 30 minutes (Lovell 1937).

### *C. equi*

This organism was isolated by Magnusson (1923) from foals affected with pyæmia. It has been confused with *C. pyogenes* but differs from it in several respects notably in its abundant growth on ordinary media, its pigment formation, its failure to liquefy coagulated serum, to lyse blood or to ferment carbohydrates and its pathogenicity for horses. The following description is taken from Magnusson (1938) and Karlson, Moses and Feldman (1940).

**MORPHOLOGY**—Fairly large, pleomorphic, Gram positive bacillus showing metachromatic granules, in pure and surface colonies may appear coccoid. Reported by some workers to be partly acid fast.

**GROWTH.**—Grows freely on ordinary media, forming large succulent colonies of irregular shape and pale pink colour. On potato the growth is moist, thick, and pale pink, later becoming deep reddish yellow. Slate to black colonies on tellurite blood agar. Grows well between 18° and 3° C.

**PERSISTENCE.**—Said by Karlson, Moses and Feldman (1940) to be unusually resistant to oxalic acid which is sometimes used for the destruction of non-acid fast bacilli.

**BIOCHEMICAL REACTIONS.**—No fermentation of sugars. Poor growth in litmus milk with no obvious change. No hemolysis. No liquefaction of coagulated serum or gelatin. Indole negative. Nitrates reduced to nitrites.  $\text{S}_{\text{H}}\text{H}_2\text{S}$  production. Catalase formed.

**ANTIGENIC STRUCTURE.**—Strain specificity said to be marked, so that the agglutination reaction is of little help in identification. By complement fixation, however a species-specific antigen can be demonstrated (Bruner, Dimock, and Edwards 1939).

**PATHOGENICITY.**—Gives rise under natural conditions to pyæmia in foal, characterized by a suppurative bronchopneumonia with intense purulent infiltration of the adjoining lymph nodes, and sometimes to intestinal ulceration and abscess formation in the mesenteric lymph nodes. Probably non pathogenic for swine though frequently present in the submaxillary lymph nodes. Experimentally the natural disease can be reproduced by intratracheal inoculation of foals. Subcutaneous inoculation produces local abscess formation with involvement of the focal lymph nodes in horses, pigs and goats. Gives rise to peritonitis not always fatal, when inoculated intraperitoneally into guinea pigs.

#### *C. renale*

This organism was first described by Enderlen (1890-91) but it is very doubtful whether all the strains of diphtheroid bacilli that have since been isolated from pyælis in cattle were identical with the bacillus isolated by him and it is by no means certain that the organism described by more recent workers is entitled to specific rank. It seems quite clear however that this organism differs in several ways from *C. pyogenes* with which it has often been confused. The incomplete description that follows is taken mainly from the papers of Jones and Little (1923, 1930) and Merchant (1933) and is given with considerable reserve.

**MORPHOLOGY.**—*C. renale* is a typical Gram positive barred diphtheroid, showing numerous metachromatic granules, and considerable pleomorphism.

**GROWTH.**—On serum agar gives moist, raised colonies showing a pigmentation that varies from cream to yellow. Later the growth becomes drier.

**BIOCHEMICAL REACTIONS.**—Most strains that have been examined have fermented dextrose alone with the production of acid. According to Merchant (1933) levulose and mannose may also be fermented. Gelatin is not liquefied. No hemolysin is produced.

**ANTIGENIC STRUCTURE.**—The few data available (see Merchant 1933) are insufficient to allow any adequate description.

**PATHOGENICITY.**—The organism has been isolated by several observers from cattle suffering from pyælis and Jones and Little (1923, 1930) have reproduced the disease experimentally in these animals by the injection of pure cultures. No data are available with regard to its pathogenicity for laboratory animals.

#### *C. murium*

(Synonym. *C. pseudotuberculosis murium*.)

Isolated from a mouse by Kutscher in 1894 and by Bongert in 1901. It has since been isolated by several observers (Andrews *et al.* 1923).

**MORPHOLOGY.**—In films from cultures on Loeffler's medium the appearances are very similar to those presented by *C. ovis* or by some strains of *C. diphtheriae* (see Fig. 87).



**GROWTH**—The type of growth on serum, agar, and gelatin appears to be very like that of *C. diphtheriae*. Pellicle formation in broth has not been recorded, but suitable conditions have perhaps never been secured.

**BIOCHEMICAL REACTIONS**.—Acid is produced from dextrose, maltose, and saccharose, galactose, dextrin, lactose, and mannitol are unchanged. Litmus milk is unchanged, or shows a slight and transitory acidity. Indole is not formed. Gelatin is not liquefied. Nitrates are reduced.

**ANTIGENIC STRUCTURE**—At present uninvestigated.

**TOXIN PRODUCTION AND PATHOGENICITY**—*C. murium* is a natural pathogen of mice, and appears to be pathogenic for no other species, with the possible exception of rats. There is evidence that some of its effects are due to the action of a soluble toxin (see above).

### C acnes

A diphtheroid organism was described in the ordinary lesions of cutaneous acne by Unna in 1898, and was isolated in culture by Sabouraud (1897).

**MORPHOLOGY**—This has been generally reported as very variable, according to the medium employed. degree of acidity, oxygen pressure, etc. Craddock (1942) distinguishes two types on blood agar: in large colonies the organisms are short, thick, irregular in shape, and often clubbed; in small colonies the bacilli are longer, thinner and curved. Often weakly Gram positive.

**GROWTH**—*C. acnes* will grow aerobically if the medium contains serum or blood, and is acidified by the addition of lactic, or of hydrochloric acid, so as to fall within the pH range 6.2-6.8. Growth occurs better under anaerobic conditions, and is favoured by glucose, glycerol, blood, boiled blood, Fildes' extract of blood and serum. On plain agar growth is poor or absent.

On a glucose agar plate anaerobically after 4 days at 37° C the colonies are circular, 0.2-0.4 mm in diameter, convex, amorphous, greyish white, with a smooth glistening surface and an entire edge; they are butyrous in consistency, and emulsify easily. The growth has a sour smell. After 6 weeks, colonies may be coloured pink. On blood agar anaerobically Craddock (1942) describes two types. Type I forms a large capped up colony, yellowish buff in colour with a wide zone of hæmolytic. Type II forms a small flat colony. In a glucose agar shake medium no growth occurs for about 10 mm below the surface; there is then often a band growth for 10-20 mm below the surface, with discrete colonies to the bottom of the tube, the medium becomes milky and opaque. Loeffler's serum, no liquefaction. In glucose broth there is a slight turbidity after 3 days anaerobically, and a slight, finely granular sediment, after a week or so there is a heavy loose floccular deposit, which occupies the lower centimetres of the tube and disintegrates on shaking to give a moderate turbidity. Slight to moderate turbidity in cooked meat medium. No growth in gelatin stab culture at 22° C.

**BIOCHEMICAL REACTIONS**—According to Sudmersen and Thompson (1909-10) *C. acnes* produces acid from glucose, galactose, maltose, glycerol, and mannitol, but does not

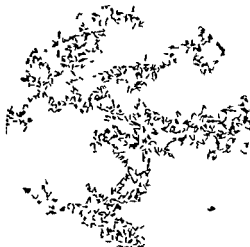


FIG. 87.—*C. murium*

From 24 hours culture on Loeffler's serum ( $\times 1000$ )

ferment lactose. Of 2 strains examined by Sudmersen and Thompson, one actively fermented saccharose, the other gave a late and slight acidity. In our experience acid is generally formed in glucose, maltose, and sucrose, sometimes in lactose but not in mannitol. Slight acid in litmus milk. Indole negative. Nitrates reduced. Gelatin not liquefied.

**ANTIGENIC STRUCTURE.**—According to Craddock (1942) two types can be distinguished by agglutination with specific rabbit sera.

**PATHOGENICITY AND TOXIN PRODUCTION.**—It is generally believed that *C. acnes* is aetologically related to the lesions from which it has been isolated. Experimentally it shows some degree of pathogenicity for the mouse.

### *C. typhi*

Isolated by Plotz (1914) from the blood of patients suffering from typhus fever. Originally regarded by Plotz and his colleagues as aetologically related to the disease (Plotz, Olitsky and Baehr 1915), but now generally admitted to be an example of a parasitic microaerophilic diphtheroid, without any established pathological significance (Olitsky 1921).

**MORPHOLOGY** is that of a small pleomorphic diphtheroid, with few elongated cells but numerous coccid forms. The rod forms which occur may be straight or curved, with rounded or pointed ends. Metachromatic granules are present.

**GROWTH.**—*C. typhi* appears to demand more strictly anaerobic conditions for growth than *C. acnes* but there appears to be no record of the result of acidification of the medium. It is recorded as giving a creamy white growth on Loeffler's serum, glucose serum agar, or potato, the growth takes on a light brown colour in its later stages on the latter medium.

**BIOCHEMICAL REACTIONS.**—Tested on ascitic agar containing 2 per cent. of the test carbohydrate, and incubated in Buchner tubes *C. typhi* is stated to produce acid from glucose, maltose and galactose, but not from mannitol, dextrin, lactose or saccharose.

**ANTIGENIC STRUCTURE** unknown.

**PATHOGENICITY**—Probably slight or absent.

We append, in tabular form (Table 31), the chief differential characteristics of the ten named species described above, and the fermentation reactions of the eleven groups of diphtheroid bacilli differentiated by the Committee of the Medical Research Council (Table 32). In both tables the + sign signifies the formation of acid. Since the fermentation reactions of this group are habitually tested in Hiss's serum water medium, the formation of acid will usually be followed by the formation of a clot, after a longer or shorter period, though some strains which produce definite acidity in the presence of a particular carbohydrate fail to clot the medium.

With regard to the eleven groups of unnamed diphtheroids, the Committee note that there is no correlation between the source from which any strain was derived and its biochemical reactions. Thus, of the 15 strains which fall into Group I, ten came from the nose, one from the ear, two from the eye, one from an infected wound, and one from a specimen of pus. As already noted, there was no consistent relation between pigment production and fermentative activity. Thus one strain of Groups IV, VII and XI produced pigment, as did the single representative of Group IX. The four strains of Group VI, on the other hand, and the three strains of Group VIII all formed pigment, so that these two groups would appear to be characterized by a marked tendency to pigment production. It may be noted that none of the strains examined produced the pink or red pigments which have

TABLE 31

Species	Production of Acid from						Liquefaction of Gelatin	Production of Hemolysin	Oxygen Requirements	Pathogenicity	Toxin Production
	Dextrose	Maltose	Saccharose	Dextrin	Lactose	Mannitol					
<i>C diphtheria</i> *	+	+	-	+	-	-	-	+	A & F An	+	+
<i>C diphtheria</i> (avirulent)	+	+	-	+	-	-	-	+		-	-
<i>C hofmanni</i>	-	-	-	-	-	-	-	-		-	-
<i>C xerosis</i>	+	+	+	-	-	-	-	-		-	-
<i>C pyogenes</i>	+	+	+	-	+	-	+	+		+	+
<i>C equi</i>	-	-	-	-	-	-	-	-		+	-
<i>C renale</i>	+	-	-	-	-	-	-	-		+	-
<i>C ovis</i>	+	+	-	±	-	-	+	+		+	+
<i>C murium</i>	+	+	+	-	-	-	-	-		+	+
<i>C acnes</i>	+	+	±	-	-	+	-	-	microaerophilic	+	-
<i>C typhi</i>	+	+	-	-	-	-	-	-		-	-

\* See Table 30 for differentiation between *gravis* *mitis* and *intermedia*

been so frequently noted by some observers and especially by those who have examined numerous strains from lymphatic glands and other tissues (Hoag 1907 Harris and Wade 1915). The Committee note that the strains selected have been tested on many occasions and at long intervals with scarcely varying results.

TABLE 32

FERMENTATION REACTIONS OF 19 UNSELECTED STRAINS OF DIPHTHEROIDS EXAMINED FOR COMMITTEE OF MEDICAL RESEARCH COUNCIL (see Andrewes *et al* 1923)

Group	Dextrose	Maltose	Galactose	Saccharose	Lactose	Dextrin	Mannitol	No of strains in Group
I	+	-	+	+	-	-	-	15
II	+	-	-	+	-	-	-	11
III	+	+	-	-	-	-	-	11
IV	+	±	-	-	-	-	-	6
V	+	+	+	-	-	-	-	4
VI	+	+	+	+	+	-	-	4
VII	+	+	-	+	-	-	-	5
VIII	+	+	+	+	-	-	-	3
IX	+	+	+	+	+	+	-	1
X	+	-	+	+	-	-	+	1
XI	-	-	-	-	-	-	-	18

## REFERENCES

- ANDERSON, J S, COOPER, H. E., HAPFOLD, F C, and McLEOD, J W (1933) *J Path Bact.*, 36, 169
- ANDERSON, J S, HAPFOLD, F C, McLEOD, J W, and THOMSON, J G (1931) *J Path Bact.*, 34, 667
- ANDREWS, F W *et al.* (1923) *Med. Res. Coun.*, "Monograph on Diphtheria." London.
- BABES, V (1896) *Bull. Soc. Anat., Paris*, 61, 72
- BAILEY, G H. (1925) *J Immunol.*, 10, 791
- BARR, M., GLENNY, A. T., POPE, C G., and LINGGOOD, F V (1941) *Lancet*, ii, 301
- BARRATT, M. M. (1923) see Andrews *et al.*, p. 174, (1933) *J Path Bact.*, 36, 369
- BOLTON, C (1903) *Lancet*, i, 278
- BOGERT (1901) *Z Hyg InfektKr.*, 37, 449
- BRANDWILK, A. C. and TASMAN, A. (1932) *Z Immunforsch.*, 77, 390, (1933) *Ibid.*, 78, 540
- BRAUN, H. and HOFMEIER, K. (1927) *Klin. Wochr.*, 6, 690
- BRAUN, H., HOFMEIER, K., and MÜNDEL, F (1929) *Zbl. Bakt.*, 113, 530
- BRAUN, H. and MÜNDEL, F (1927) *Zbl. Bakt.*, 103, 182, (1929) *Ibid.*, 112, 347
- BRAY, J (1944) *J Path Bact.*, 56, 497
- BROOKS, R F and HUCKER, G J (1944) *J Bact.*, 48, 295
- BROWN, J H. and ORCUTT, M L. (1930) *J exp. Med.*, 22, 219
- BRUNER, D W., DIMOCK, W W. and EDWARDS, P R. (1939) *J infect. Dis.*, 65, 92
- BULL, L. B. and DICKINSON, C. G. (1935) *Aust. vet. J.*, 11, 126.
- BUNKER, J W (1919) *J Bact.*, 4, 217
- BUNNY, W E., CIANCIARULO, J., and KAMILL, M. (1931) *J Immunol.*, 20, 417
- CARVE, H R. (1939) *J Path Bact.*, 49, 313, (1940) *Ibid.*, 51, 199
- CHALMERS, A. J. and MACDONALD, N. (1930) *J trop. Med. Hyg.*, 23, 83.
- COCA, A. E., RUSSELL, E. F., and BAUGHMAN, W H. (1921) *J Immunol.*, 6, 887
- COOPER, H. E., HAPFOLD, F C., McLEOD, J W., and WOODCOCK, H. E. DE C. (1936) *Proc. R. Soc. Med.*, 29, 1029
- COSTA, S. TROISIER, J., and DAUVERGNE, J (1918) *C. R. Soc. Biol.*, 81, 89
- COWAN, M. L. (1927) *Brit. J. exp. Path.*, 8, 6
- CRADDOCK, S (1942) *Lancet*, i, 553.
- CROWELL, M. J (1926) *J Bact.*, 11, 65
- DASOVILLE (1907) *Bull. Soc. cent. Méd. vet.*, 61, 576
- DAVIS, L. and FERRY, N S. (1919) *J Bact.*, 4, 217
- DUDGEON, L. S. (1906) *Brain*, 23, 227
- DUPLEY, S F., MAY, P M., and O'FLYNN, J A. (1934) *Spec. Rep. Ser. med. Res. Coun., Lond.*, No 195
- DURAND, P (1918) *C. R. Soc. Biol.*, 81, 1011 (1920) *Ibid.*, 83, 613.
- EAGLETON, A. J. and BAXTER, E. M. (1923) *J Hyg., Camb.*, 22, 107
- EATON, M. D (1936) *J Bact.*, 31, 367
- EATON, M. D. and BAYNE-JONES, S (1934) *J Bact.*, 29, 56
- EBERSON, F (1918) *J infect. Dis.*, 23, 1
- ERISMANN, O (1932-33) *Zbl. Bakt.*, 127, 111, (1933) *Z Hyg InfektKr.*, 115, 273
- ENDBLEIN, E. (1890-91) *Dtsch. Z. Tierheilk.*, 17, 325
- ERNST, W (1889) *Z Hyg InfektKr.*, 4, 25, (1889) *Ibid.*, 5, 428, (1900) *Zbl. Bakt.*, 39, 549-660 (1906) *Ibid.*, 40, 79
- EWING, J O (1933) *J Path. Bact.*, 37, 345
- FISCHL, V., KOZICH, M., and KUSSAT, E (1931) *Z Hyg InfektKr.*, 112, 421
- FLEMING, A (1909) *Lancet*, i, 1035
- FLEXNER, S (1897) *Rep. Johns Hopk. Hosp.*, 6, 209
- FRIEDER, W (1921) *Zbl. Bakt.*, 87, 204
- GILCHRIST, T C (1900) *Bull. Johns Hopk. Hosp.*, 9, 409, (1903) *J cutan. Dis.*, 21, 107
- GLACE, F (1903) *Z Fleisch u. Milchhyg.*, 13, 166
- GLASS, V (1937) *J Path Bact.*, 44, 233, (1939a) *Ibid.*, 48, 507, (1939b) *Ibid.*, 49, 549
- GLENNY, A T (1923a) *J Hyg., Camb.*, 24, 301, (1923b) *J Path Bact.*, 28, 201
- GLENNY, A T. and ALLEN, K. (1922) *J Path. Bact.*, 24, 61
- GLENNY, A. T. and WALPOLE, G S. (1915) *Biochem. J.*, 9, 298.
- GOLDIE, H. (1933) *C. R. Soc. Biol.*, 112, 1210 (1934) *Ibid.*, 116, 17
- GOLDSWORTHY, N E. and WILSON, H. (1942) *J Path Bact.*, 54, 183
- GOODMAN, H M. (1907) *J infect. Dis.*, 4, 509
- GRAHAM-SMITH, G S (1908) *Diphtheria*. Nuttall and Graham Smith Cambridge
- GRIFFITH, A. S. (1901) *Rep. Thomps. Yates Lab. Univ. Lpool.*, 4, 99
- GRIPS (1893) *Z Fleisch u. Milchhyg.*, 8, 166
- GROER, F VON (1923) *Biochem. Z.*, 133, 13
- GUNDEL, M., GÖRBOY, P., and PAGEL, W (1932) *Z Hyg InfektKr.*, 113, 629
- HADLEY, P B (1907) *J infect. Dis.*, 3, Suppl. No 3, 93

- HADLEY, P. B and GORHAM, F P (1907) *Zbl Bakt, Ref*, 40, 392  
 HALL, I C and STONE, R V (1916) *J infect Dis*, 18, 195  
 HARRIS, W H and WADE, H W (1915) *J exp Med*, 21, 493  
 HARTLEY, P (1922) *J Path. Bact*, 25, 479  
 HARTLEY, P and HARTLEY, O (1922) *J Path Bact* 25, 458  
 HAVENS, L C (1920) *J infect Dis*, 26, 393  
 HENDRY, CATHERINE B (1938) *J Path Bact* 46, 383  
 HILL, H. W. (1898) *J Boston Soc med Sci*, 3, 86, (1899) *Ibid*, 4, 78 (1902a) *J med Res*, 7, 115, (1902b) *Ibid*, 7, 202  
 HOAG, L (1907) *Boston med surg J*, 157, 10  
 HOFMANN, G VON (1888) *Wien med Wochr*, 38, 65, 103  
 HOLTH, H (1908) *Z Infekth. Hausiere*, 3, 155  
 HOSOYA, S and KUROYA, M (1923) *Sci Rep Inst infect Dis Tokyo Univ*, 2, 233  
 HOSOYA, S, OZAWA, E, and TANAKA, T (1933) *Jap J exp Med*, 11, 463  
 HOTTINGER, A and HOTTINGER, C (1933) *Z Kinderheilk*, 54, 440  
 HOYLE, L (1941) *Lancet* i, 175, (1942) *J Hyg, Camb*, 42, 416  
 JAFFE, R (1920) *Arb Inst exp Ther Georg Speyer Haus Frank am M*, 11, 5  
 JONES, F S and LITTLE R B (1925) *J exp Med*, 42, 593, (1930) *Ibid*, 51, 909  
 KARLSON, A G, MOSES H E, and FELDMAN, W H. (1940) *J infect Dis* 67, 243  
 KLEBS, E (1883) *Verh Cong inn Med, Wiesbaden*, 139  
 KNIGHT, B C J G (1936) *Spec Rep Ser med Res Coun, Lond*, No 210  
 KOSEK S A and RETTGER L F (1919) *J infect Dis*, 24, 301  
 KRAH, E and WITEBSKY, E (1930) *Z Immunforsch*, 66, 59  
 KÜNNEMANN, O (1903) *Arch wiss prakt Tierheilk*, 29, 128  
 KUSCHKE and NEISSER (1883) *Jber schles Ges Vaterl Kult*, 60, 50  
 KUTSCHER. (1894) *Z Hyg Infekth.*, 18, 327  
 LANGER, H (1916) *Zbl Bakt*, 78, 117.  
 LEHMANN, K. B and NEUMANN, R O (1896) "Atlas u Grundriss d Bakt u Lehrb d spez bakt Diagnostik 6th Ed, Munich  
 LEONARD, G F and HOLM, A (1933) *J infect Dis*, 53, 376  
 LEULIAR, A, SÉDALLIAN, P and CLAYEL (1931) *C R Soc Biol*, 107, 1136  
 LINDENBAUM, H (1932) *Z Hyg Infekth* 113, 288  
 LOCKE, A and MAIN E R (1928) *J infect Dis*, 43, 41  
 LOEFLER, F (1884) *Mitt ReichsgesundhAmt*, 2, 421 (1890) *Zbl Bakt* 7, 528  
 LOVELL, R (1937) *J Path Bact*, 45, 339, (1941) *Ibid*, 52, 295, (1944) *Ibid* 56, 525  
 LUCET, A (1893) *Ann Inst Pasteur*, 7, 325  
 MCLEOD, J W (1943) *Bact Rev*, 7, 1  
 MAGNUSOV, H (1923) *Arch wiss prakt Tierheilk*, 50, 22, (1938) *Vet Rec*, 50, 1459  
 MATTHEW, A T R (1944) *Pers comm*  
 MAYER M E (1930) *J infect Dis* 47, 384  
 MELLON, R R (1917) *J Bact*, 2, 269  
 MERCHANT I H (1935) *J Bact*, 30, 95  
 MINETT, F C (1922a) *J comp Path*, 35, 71 (1922b) *Ibid*, 35, 291  
 MOLLARD, J and REGAUD, C (1895) *C R Soc Biol*, 2, 828  
 MORTON, H. E and GONZALEZ L M (1942) *J Immunol* 45, 63  
 MUELLER J H (1935a) *Science*, 81, 50 (1935b) *J Bact*, 29, 383, (1935c) *Ibid*, 29, 515  
 MUELLER J H, KLISE K. S, PORTER E F and GRAYBIRL, A. (1933) *J Bact*, 25, 509  
 NEILL, G A W (1937) *J Hyg, Camb*, 37, 552  
 NEILL, J M, RICHARDSON, L V, FLEMING, W L, SUGG, J Y, and GASPARI E L. (1931) *Amer J Hyg*, 13, 499  
 NICOLLE M, LOISEAU G, and FORGOT P (1912) *Ann Inst Pasteur*, 26, 83  
 NITSCH J (1933) *Z Kinderheilk*, 54, 470  
 NOCARD, E (1889) *C R Soc Biol*, 1, 608  
 OLITSKY, P K (1921) *J exp Med*, 34, 593  
 PAPPENHEIMER A M (1937) *J Biol Chem* 120, 543  
 PAPPENHEIMER A M and JOHNSON S J (1936) *Brit J exp Path*, 17, 335  
 PAPPENHEIMER A M and ROBINSON E S (1937) *J Immunol*, 32, 291  
 PETRIE C F and McCLEAN, D (1934) *J Path Bact*, 39, 635.  
 PLOTZ, H (1914) *J Amer med Ass*, 62, 1506  
 PLOTZ H, OLITSKY, P K, and BAEHR G (1915) *J infect Dis* 17, 1  
 POELS (1912) *Tijdschr Leeuartsnijk*, 39, 90  
 POPE C G (1932) *Brit J exp Path*, 13, 207  
 POPE C G and HEALEY M (1933a) *Brit J exp Path*, 14, 77 (1933b) *Ibid*, 14, 87  
 POPE C G and LINGGOOD F V (1939) *Brit J exp Path*, 20, 297  
 POPE C G and SMITH M L (1932) *J Path Bact*, 35, 573  
 PREISZ, H (1894) *Ann Inst Pasteur* 8, 231  
 RAMON, G and BERTHELOT A (1932) *C R Soc Biol*, 110, 530  
 Report (1942) *Mon Bull, Emerg publ. Hlth Lab Serv*, Feb., p. 12

- REYMOND COLONNATI, and PERROCCITO (1881) *Cong period int ophthalm C R* 1880  
Milano, Annexes, 48 (1883) *G Accad. Med., Torino* 31, 519
- ROBERTSON R. C. (1904) *J infect Dis* 35 311
- ROBINSON D T (1934) *J Path. Bact* 38 501 (1946) *Pers comm*
- ROBINSON D T and MARSHALL, F N (1934) *J Path. Bact.* 38, 73 (1935) *Lancet* ii 441
- ROBINSON D T and PREEVEY A. L. P. (1936) *J Path. Bact.*, 43, 403
- RÖMER, P H (1909) *Z ImmunForsch* 3, 208
- ROUX, E. and YERSIN A. (1888) *Ann. Inst. Pasteur* 2, 609
- SABOURAUD P (1897) *Ann Inst Pasteur* 11, 134
- SCHMIDT H. (1933-34) *Zbl Bakt* 130 391
- SCHMIDT S. (1931) *Z ImmunForsch.*, 71, 101
- SCHMIDT S., HANSEN A., and KHAER, K. A. (1931) *Ann Inst Pasteur* 46 209
- SCHWOWER, J (1904) *Zbl. Bakt* 35 608
- SCOTT W M. (1903) *Rep publ Hlth med Subj Lond* No 22
- SIEMENS B W L. (1933) *Bydrage tot de Kennis der Typen van het Corynebacterium Diphtherie.* Van Gorcum and Co Assen.
- SMITH J (1923) *J Hyg Camb* 22, 1
- SUDMERSE H J and THOMPSON E. T. (1909-10) *J Path. Bact.* 14, 2-4
- TASMAN A and PONDMAN A. B F A. (1931) *Z ImmunForsch.*, 72, 240
- TASMAN A and WAASBERGEN J P VAN (1932) *Z ImmunForsch.*, 75 164
- UNNA P G (1896) *The Histopathology of Diseases of the Skin.* Eng Transl by Walker Edin
- UCHINSKY N (1893) *Zbl Bakt.*, 14 316 (1897) *Ibid* 21, 146.
- WADSWORTH, A. and QUIGLEY J J (1934) *Amer J Hyg.*, 20 220
- WADSWORTH A QUIGLEY J J and SICKLES, G R (1935) *J exp Med* 55 810
- WADSWORTH A and WHEELER, M W (1934) *J infect Dis.*, 55 123.
- WADSWORTH A and WHEELER, M W (1934) *J infect Dis* 55 123
- WARD A R (1917) *J Bact* 2, 619
- WATSON A. F and LANGSTAFF E. (1906) *Biochem J.*, 20, 763, (1927) *J Path. Bact* 30 383
- WERNICKE (1893) *Arch. Hyg.*, 18 190.
- WENSLow C. E. A. BROADHURST J BUCHANAN R. E., BRUNWIEDE, G., ROGERS L. A and SMITH G H (1900) *J Bact.*, 5 191
- WRIGHT J H (1894) *Boston med surg J.*, 131, 309
- ZAJDEL, R. (1930) *C R Soc Biol* 111, 109

## CHAPTER 18

### FUSIFORMIS

#### DEFINITION *Fusiformis*

Obligate parasites Anaerobic or microaerophilic Cells frequently elongated and fusiform, staining somewhat unevenly Filaments sometimes formed, non branching, sometimes highly pleomorphic Non motile No spores Reaction to Gram variable, but mainly Gram negative Growth in laboratory media feeble

A number of organisms, anaerobic or microaerophilic in their oxygen requirements, have been isolated by different workers from necrotic lesions in man and animals. The first of these was the so called *B. necrophorus*, which was observed by Loeffler (1884) in calf diphtheria. In 1896 Vincent described a fusiform bacillus, frequently associated with a spirochaete, *Trep. vincenti* (see Chapter 79), in necrotic and ulcerative lesions of the throat and other tissues in human beings. Veillon and Zuber (1898), studying the bacterial flora of appendicitis and other suppurative lesions, described a number of non sporing anaerobic bacilli, to which they gave the names of *B. ramosus*, *B. serpens*, *B. fragilis*, *B. furcosus*, and *B. fusiformis*, this last organism was apparently identical with the fusiform bacillus described by Vincent. Later work has revealed the frequent presence of organisms of this group in the mouth and on the teeth of man and certain animals (Tunnick 1906, Ellermann 1907, Varney 1927, Pratt 1927, Slinnetz and Rettger 1933, Bachman and Gregor 1936, Pesch and Schmitz 1936, Spaulding and Rettger 1937, Hine and Berry 1937, Kelly 1944) and in the healthy alimentary canal of man (Eggerth and Gagnon 1933, Weiss and Rettger 1937, Misra 1938, Lewis and Rettger 1940, Dragstedt, Dack and Kirsner 1941). With the more general use of improved anaerobic methods they have been found in association with various human infections, especially in France and the United States (see Chapter 79).

Relatively little attention has been paid to the systematic study of these organisms, and their classification presents considerable difficulties. Castellani and Chalmers (1920) proposed a genus *Bacteroides* to contain obligatory anaerobic bacilli that did not form spores, and the genus has since been adopted in Bergey's manual, where it covers a variety of morphologically different organisms. The American Committee of Bacteriologists (see Report 1920) have described a genus *Fusiformis*, for the inclusion of organisms with certain characteristics resembling in some respects those possessed by the *Corynebacterium* and the *Pfeifferella* groups. As the definition of this genus (see above) seems to cover the main characteristics of the organisms we have mentioned, it seems permissible, at least for the moment, to include these organisms within it.

**Group Characteristics**—The organisms of this group are typically rod shaped, but their size is subject to considerable variation. They may be very short, or they

may grow out into long filaments which are generally curved. Their width is also variable, not infrequently it is greater near the middle of the bacillus than at the end, giving the organism a fusiform appearance, the fusiform swellings may be as much as  $4-5\ \mu$  in diameter. Occasionally the greatest diameter of the bacillus is at one end, so that the organism appears clubbed. Pleomorphism is a marked characteristic of some species. The organisms are arranged singly, in pairs end-to-end, or in chains, pseudo-filaments showing evidence of subdivision are common. Some species show false branching with the result that V or Y forms are seen, but true branching does not occur. Some species are said to be motile, but the observations of recent workers render this doubtful. Spores are never formed. The organisms stain with the usual aniline dyes, but staining is often irregular. The reaction to Gram varies with different species. None of the organisms is acid fast.

Culturally, growth occurs under anaerobic or microaerophilic conditions and is said to be favoured by the presence of 2 per cent  $\text{CO}_2$ . Little is known of their respiratory mechanism, but it seems probable that in fluid media they do not produce such low oxidation reduction potential, as do many of the spore-bearing anaerobes (Dack and Burrows 1935). Some species grow readily in ordinary media, while others require the addition of natural animal protein. According to Svanetz and Rettger (1933) growth is stimulated by aqueous extracts of various vegetables and a potato extract gelatin medium is recommended for the preservation of stock cultures. The optimum temperature for growth is about  $31^\circ\text{C}$ ., some species will develop at  $23^\circ$  others not below  $30^\circ\text{C}$ . In solid media colonies frequently do not become visible for 3 or 4 days, and usually remain small. For the isolation of the intestinal non-sporing anaerobic bacteria Lewis, Bedell and Rettger (1940) recommend a glucose-cysteine agar medium at a pH between 6.3 and 7.0, enriched with yeast-extract and tomato juice, and state that growth is greatly improved by 10 per cent  $\text{CO}_2$  in the anaerobic atmosphere. Little is known of the precise growth requirements of these organisms though both pantothenic acid and pyruvic acid appear to be essential nutrients for a number of strains (West, Lewis and Wiltzer 1942).

The organisms are not particularly resistant, they are destroyed by exposure to moist heat at  $55^\circ\text{C}$  within an hour. The colonies of some species are highly oxygen-sensitive, dying within an hour of exposure to air (Hine and Berry 1937). The fermentation reactions have been incompletely studied, but some species produce acid and gas in certain carbohydrate media.

The organisms of this group appear to be obligatory parasites and may be cultivated from certain inflammatory processes, particularly those accompanied by necrosis and ulceration, as well as from the normal mouth, teeth, and faeces. There seems to be little doubt that some species are primarily responsible for the lesions from which they are isolated, but the aetiological role of others is probably more of a secondary nature. Many of the species are pathogenic to laboratory animals, producing necrotic lesions and death. Whether a true exotoxin is formed is not yet known.

Antigenically, little exact information is yet available about these organisms but there is evidence that differences in antigenic structure do exist between different types.

The recorded investigations of these organisms are of two kinds firstly, of organisms found in association with disease processes, and secondly, of organisms



isolated from the normal flora of the mouth and intestines. Both Varney (1927) and Slanetz and Rettger (1933) divided fusiform bacilli of the normal mouth into four types I to IV on the basis of morphological and serological characteristics but the divisions did not coincide. Agglutination tests revealed a high degree of strain specificity and a lesser degree of group specificity. Weiss and Mercado (1938) extracted immunologically type specific protein like substances from organisms of Slanetz and Rettger's Types I, II and III and also found some evidence of a group specific carbohydrate in the organisms. Spaulding and Rettger (1937) later divided 84 strains of fusobacteria from many sources into two groups: Group I containing their previously described Types I, II and some of Type III; Group II containing the remainder of Type III and Type IV. In a limited cultural study of 193 human mouth strains Pesch and Schmitz (1936) distinguished 6 types but concluded that these were varieties of a single *Bacterium fusiforme*. Hine and Berry (1937) studied 104 mouth strains dividing them into three species: *F. nucleatus*, *F. polymorphus* and *F. dentium*. The first two corresponded only very approximately to Spaulding and Rettger's Group I consisting of short or long fusiform rods of limited saccharolytic activity and *F. dentium* to Group II consisting of long relatively thick fusiform rods with marked saccharolytic activity.

Besides these fusiform bacteria of the mouth there is a large group of Gram negative non sporing intestinal bacteria in the contents of the alimentary canal. It is clear from the work of Eggerth and Gagnon (1933), Weiss and Rettger (1937), Misra (1938) and Lewis and Rettger (1940) that these anaerobes are the predominant organisms in the human lower intestine sometimes outnumbering *Bact. coli* by one hundredfold or more. They are biochemically and antigenically heterogeneous though some association between morphological, biochemical and antigenic characters has been noted.

However the relation of the normal flora of the alimentary canal to the organism responsible for infective lesions and particularly to those species which were described in the early years of this century is not yet clear. The intestinal species in man bear a morphological resemblance to some of the pathogenic species like *F. funduliformis* and certain fusiform bacilli but differ in biochemical reactions (Lewis and Rettger 1940). Until the heterogeneity of both the pathogenic and the normal strains has been reduced by further study speculation on cross relationships within the group is unprofitable. (See Prevot 1938 for a taxonomic review of this difficult group).

We append descriptions of some of the named species that have been found in association with infective processes. In the first description we have assumed the substantial identity of *F. necrophorus* and *Bacillus funduliformis* and retained the name *F. necrophorus* for the two (Dack et al 1938, Kirchheimer 1940, see also Lemierre, Grumbach and Reilly 1936).

**F. necrophorus**—Synonyms: Schmorl's bacillus, *B. diphtheriae vitulorum*, *Streptothrix cuniculi*, *Actinomyces necrophorus*, Bang's necrosis bacillus, *B. funduliformis*, *Bacteroides funduliformis*. First observed by Loeffler in 1884 in calf diphtheria. He succeeded in producing necrotic lesions in mice by subcutaneous inoculation of the diphtheritic membrane and in obtaining a primary culture of the organism from mice on calf serum but he failed to subculture it. Schmorl in 1891 encountered apparently the same bacillus in a spontaneous epidemic amongst his laboratory rabbits characterized by spreading necrosis of the lower lip. He inoculated mice with material from the rabbits and obtained

cultures of the organism from the mice. He classed it with the *Streptothrix* group under the *Leptothrix* or *Cladothrix* genera, and called it *Streptothrix cuniculi*. The organism has been described by a few other workers. (For a detailed description of it, see Orcutt 1930, Beveridge 1934.) Hallé (1898) named a bacillus associated with genital infections in man *B. funduliformis*, and later Teissier and his colleagues (Teissier *et al.* 1929, 1931) found the same organism in four cases of septicæmia in man. It probably corresponds to the species C described by Veillon and Zuber (1899) and its role in putrid infections of man has been confirmed by a number of workers (see Chapter 79). The human strains do not differ in any consistent way from the animal strains of *F. necrophorus* except that they are usually less pathogenic for laboratory animals (Dack, Dragstedt and Heinz 1937) and tend to be pleomorphic.

**MORPHOLOGY**—In the diphtheritic membrane of calves the bacilli appear as long threads—5 to 6 times as long as they are broad—arranged either in thick heaps or in long wavy rows. In the pleural or pericardial exudate of rabbits dying of lateral necrosis they usually appear as Gram negative, highly pleomorphic bacilli, varying in shape from cocci to bacilli and long threads. In the thread forms there are often oval unstained portions arranged at regular intervals looking like spores, but as they do not give the differential spore stain and are not particularly resistant to heat, they cannot be regarded as spores. The filaments may reach 80–100  $\mu$  in length, they are 0.75–1.5  $\mu$  thick, one end is often narrow and pointed, the other thicker or almost fusiform. They may be surrounded by a capsular material. Branching does not occur. In culture the organisms appear as straight or curved rods, or as filaments. The ends are rounded, and the sides are generally parallel, though fusiform enlargements including ‘ball’ forms up to 5  $\mu$  in diameter are not uncommon. Except in young cultures, staining is irregular, and beaded forms are common.

**CULTIVATION**—Schmorl cultivated the organism in deep inspissated sheep serum. It grows only under anaerobic conditions. Deep colonies in serum agar plates are round, pinhead in size, matt-white, with an entire edge, under a low power they have a thicker centre and an irregularly radiate periphery, under a high power the centre consists of a mesh work of threads, and the periphery of streaming threads, which pass often for some distance into the surrounding medium. According to Shaw (1933), deep colonies in a semi solid glucose serum agar medium have a grey, cotton like, fluffy appearance, while in stiffer agar they are brown, circular or biconvex, with an entire edge. Surface colonies, after 2 days on a special serum agar medium (V.F.), made up with a peptic digest of ox muscle and liver, are circular convex, about 1 mm. in diameter, almost water clear, and glistening, after 7 days they are differentiated into a convex centre and a narrow flat periphery with a slightly dentate edge, they are butyrous in consistency and easily emulsified (Beveridge 1934). In V.F. broth there is a dense turbidity, with a small dirty white deposit, which increases as the medium clears on further incubation. Stab growth in serum begins in 24 to 40 hours near the bottom of the tube, and spreads upwards to within  $\frac{1}{4}$ –1 cm. of the surface, the growth is filiform with radiate branches, the serum is partly clouded, but not liquefied. There is no growth on potato. In glucose agar, Schmorl obtained growth only in the presence of a coccus. He quotes this as an example of *metabiosis*—or the ability of one organism to grow in a particular medium only when another organism has prepared the medium for it, and rendered it suitable. V.F. gelatin (see above) is not liquefied. Turbidity and vigorous gas production occur in cooked heart medium and brain medium. A zone of hæmolytic about 0.5 mm. wide, is said to surround deep colonies in blood agar plates. Broth containing 10 per cent. of serum and a reducing agent like cysteine (0.1 per cent.) may support growth (Dack *et al.* 1938). Cultures, especially in liquid media, have a foul odour. Growth occurs between 30° and 40° C., with an optimum at 37° C.

**BIOCHEMICAL AND METABOLIC**—Gas, and a variable amount of acid, are produced in glucose, maltose and levulose. Fermentation of glycerol, galactose and sucrose is

variable Mannitol and lactose are not fermented A soft clot is formed in 4-14 days in litmus milk Indole +  $H_2S$  + Nitrate reduction — MR — VP — Methyl ene blue reduction + Catalase weakly positive

**ANTIGENIC STRUCTURE** —Antigenic relationships exist within the group and in some surveys the strains examined have fallen into a relatively few groups As a whole however the species is antigenically heterogeneous (see Orcutt 1930 Beveridge 1934 Dack Dragstedt and Heinz 1937, Henthorne Thompson and Beaver 1936 Walker and Dack 1939)

**PATHOGENICITY** —*F. necrophorus* appears to be responsible for several necrotic and gangrenous lesions in animals such as calf diphtheria labial necrosis of rabbits and foot rot of sheep and occasionally for necrotic lesions in man Organisms identical with or closely simulating *F. necrophorus* have been described by Harris and Brown (1927) in puerperal fever, by Dack Dragstedt and Heinz (1936) in chronic ulcerative colitis by Thompson and Beaver (1931) and Cohen (1932) in human lung abscesses by Beaver, Henthorne and Macy (1934) in hepatic abscesses, and by Lemierre (1936) as a cause of pyæmia and septicæmia (see also *Necrobacillosis* Chapter 79)

Experimentally, the organism is pathogenic for rabbits and mice It is non pathogenic for guinea-pigs dogs cats pigeons and hens, though McCullough (1938) succeeded in infecting guinea pigs deficient in vitamin C with human strains of *F. necrophorus* Inoculated subcutaneously into the lower lip of rabbits, it gives rise to the typical necrotic disease (see p 1788) and causes death in 4-12 days Injected intraperitoneally into rabbits, it proves fatal in about the same time, p.m. the peritoneum is covered with large white masses sticking the two layers of serosa together, similar masses of caseous material occur between the intestinal loops Cultures can be obtained from the peritoneum. Intravenous inoculation into rabbits is fatal in about 8 days, post mortem there is a fibrino purulent pleurisy, a caseous bronchitis and disseminated lobular pneumonia, the bacilli can be cultured from the affected organs Histologically, the organisms give rise to intense inflammation, followed by a rapid necrosis not only of the fixed tissue cells but also of the cells of the exudate no liquefaction of the necrotic material occurs

Mice inoculated subcutaneously at the root of the tail die after about 12 days in an emaciated condition. Two days after the inoculation, the local site is covered with a dry brownish crust, later a yellowish grey discoloration of the borders of the inoculation wound becomes visible and gradually spreads till at the time of death the whole of the lower third of the back is involved. A purulent conjunctivitis develops and the lids are stuck together Post mortem, the subcutaneous tissue around the wound and over the lower part of the back is converted into a tough, dryish yellowish white necrotic mass, the caseation involves the back muscles as well. The whole area is surrounded by œdema. Numerous organisms are found in the caseous material. The internal organs appear normal.

In both rabbits and mice the organism is said to be found only in the affected parts, never in the blood or in those internal organs that are free from lesions, but on this question there is some doubt. The ability of the organism to produce lesions seems to be largely due to its production of a necrotizing endotoxin (Beveridge 1934) There

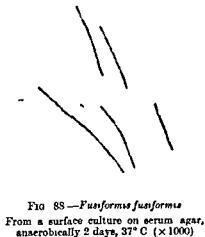


FIG 88.—*Fusiformis fusiformis*  
From a surface culture on serum agar,  
anaerobically 2 days, 37° C (× 1000)

is little evidence of the presence of a soluble substance of the exotoxin type (Scrivner and Lee 1934)

**F fusiformis**—This organism was first described by Vincent in 1896 who found it frequently associated with *Trep vincenti* (see Chapter 38) in necrotic and ulcerative lesions of the throat and other tissues. A similar and possibly identical organism was observed by Veillon and Zuber (1898) in appendicitis and other necrotic lesions of man and named by them *B fusiformis*. Smith (1933) has recorded its presence in tropical ulcer. In necrotic tissue the organisms appear as long rods 5–12  $\mu$  in length thickened in the middle and with tapering or pointed ends. The axis is straight or curved. They are arranged singly or in pairs end to end and are non motile. They stain readily with the usual aniline dyes, and are Gram positive but are decolorized if the treatment with alcohol is prolonged. In cultures the bacilli are pleomorphic (Leukowicz 1906) they vary greatly in size and do not always show the typical fusiform shape. Long filamentous forms are common. The various morphological types of fusiform bacilli (see above) cultivated from the normal mouth of man and animals vary from thin slightly pointed bacilli  $0.4 \times 3-5 \mu$  to long and thick slightly pointed bacilli  $0.7 \times 6-9 \mu$ . Tunnichiff (1903, 1933) considered the spirillar forms found in association with fusiform bacilli in lesions of the throat to be a phase of the fusiform type and found that spiral forms were relatively common in rough variants of fusiform bacilli. Hine (1937) considers that there is no good evidence of a genetic relationship between fusiform bacilli and spirochaetes.

Fig 89—*Fusiformis fusiformis*  
Blood broth culture an-  
aerobically 3 days  
37° C showing floccu-  
lar growth.

ated with stomatitis and to short diphtheroid bacilli producing no odour in culture found in healthy mouths.

The organism can be cultivated under anaerobic conditions at 37° C but not at room temperature in serum agar or serum broth. The deep colonies appear in 2 days and are 1 1/2 mm in diameter. The smallest colonies have a felt-like branched appearance, the larger ones are round and the largest of all are prismatic with angled projections and are of a yellowish colour. On the surface of serum agar the colonies are small and resemble those of streptococci, but on further incubation they may show a considerable peripheral extension and come to resemble colonies of *Cl sporogenes* (see p 876). In serum broth large white flocculi form in 24 hours and later sink to the bottom (Ellermann 1904) (see Fig 89). On plain agar there may be no growth at all or a thin very delicate whitish layer limited to the line of inoculation may develop after 48 hours (Weaver and Tunnichiff 1905). Loeffler's serum and Dorset's egg medium are both suitable. Neither is liquefied. Growth does not occur in media containing sugar unless blood or ascitic fluid is added. Peters (1911) found that in the presence of rabbit blood acid was produced in glucose maltose mannitol and lactose but not in sucrose. There was no indole formation. Group I organisms of Spaulding and Rettger's

The serological grouping of fusiform bacilli made by Rettger and his colleagues has been noted above. Bachmann and Gregor (1936) found two groups in mouth strains, corresponding respectively to filamentous odour producing bacilli associ-



FIG 90—*Fusiformis fusiformis*

Surface colony on serum agar anaerobically 7 days 37° C ( $\times 8$ ). Resembles Slanetz and Rettger's Type IV.

fusiform organisms fermented only glucose and sucrose were indole +, and  $H_2S$  + in the absence of cysteine. Group II organisms fermented glucose, sucrose maltose trehalose lactose and raffinose, were indole negative and  $H_2S$  + only in the presence of cysteine. The place of *F. juniformis* in these groups is not clear but it is more nearly related to Group II and to the Type IV of Slanetz and Pettger. Hine and Berry (1937) could find no such clear-cut association between fermentation reactions and cultural characters. The organisms do not form spores. They are killed in 15 minutes by moist heat at 55° C. but resist 1 per cent antiformin for 5 minutes. In cultures they may remain alive for 6 to 8 weeks. Injections subcutaneously or intramuscularly into rabbits and guinea-pigs may have little or no effect, at most an abscess is produced (Ellermann 1905).

*F. ramosus*—Said by Veillon and Zuber (1898) to be one of the commonest organisms in appendicitis, has also been reported in human gas gangrene and bacteraemia (Lemierre, Reilly and Bloch-Michel 1937). Gram positive non motile, non sporing rod slightly larger than *Erysipelothrix rhusopathiae*, arranged in pairs or short chains, in pus it is rather short but in culture it may form pseudo-filaments, which are really made up of numerous short rods. V forms and Y forms are common. No growth occurs at room temperature, and at 37° C. colonies do not become visible for 3 or 4 days. Surface colonies on agar are very fine effuse greyish white and translucent later they become granular and slightly cloudy. Deep colonies in agar are very small greyish points which under low magnification are seen to be ovoid and granular with hatched borders, as the colony grows, it becomes greyish yellow, and the edges become more defined. In broth it produces a uniform turbidity in 3 or 4 days with a greyish white muddy deposit. Cultures have a characteristic oetid odour but very little gas is formed. Strict anaerobe. Acid is formed in glucose, galactose and mannitol and sometimes in lactose saccharose and maltose. Lactose fermenting strains clot litmus milk. Indole — Gelatin not liquefied (Weinberg *et al.* 1937). Cultures live for over a month. Inoculated intravenously into rabbits it causes death in some days by intoxication and cachexia. Subcutaneous inoculation into rabbits causes abscess formation with death in 8 to 10 days.

*F. serpens*—Isolated by Veillon and Zuber (1898) from the pus of a mastoiditis in a child who died of pulmonary gangrene. Large, slightly motile Gram negative rod with rounded ends arranged in pairs end-to-end, or in pseudo-filaments. Strict anaerobe. Grows best at 37° C., but develops at 20° C. Surface colonies on agar after 48 hours are only just visible, later they form small cloudy translucent masses of greyish colour. Deep colonies in agar after 24 hours at 37° C. are small, clear rounded greyish, granular masses with hatched edges, and sometimes a bouquet of filaments at one point on the periphery later they increase in opacity and the edges become better defined. In gelatin stab there is a filiform growth, liquefaction occurs down the whole length of the stab the liquefied gelatin remains clear, and flocculi of growth fall slowly to the bottom forming a white deposit. Deep colonies in gelatin are round greyish, and liquefy the gelatin slowly. In broth there is a rapid turbidity, followed by a clearing of the medium with the formation of a white deposit. Acid and gas are formed from glucose levulose maltose, galactose and lactose. Indole — No haemolysin. Cultures have a foetid odour, but only a small amount of gas is evolved. Cultures remain viable for about 20 to 25 days. Pathogenic for mice, guinea pigs and especially rabbits but not so pathogenic as *F. ramosus*.

*F. fragilis*—Said by Veillon and Zuber (1898) to be the commonest organism in appendicitis. Non motile, Gram negative, rod with rounded ends, slightly smaller than the diphtheria bacillus, axis straight or slightly curved, arranged singly or in pairs end-to-end. Strict anaerobe. Difficult to isolate. Surface colonies on agar are extremely fine greyish, and translucent and tend to undergo autolysis in a few days. Deep colonies in agar do not become visible for 3 or 4 days, they are very small, round, irregularly round, or ovoid brownish yellow, opaque, with an entire edge. Deep colonies in gelatin appear in 8 to 10 days, they are panthiform, yellow, granular, with an entire edge no

liquefaction. Uniform turbidity in broth with a whitish deposit. Cultures have a foetid odour, but evolve little gas. Cultures in agar remain viable for less than a week. Subcutaneous inoculation into guinea-pigs produces abscess formation. The organism is more pathogenic for the rabbit, producing on subcutaneous inoculation an extensive phlegmon with sloughing of the skin, and death in 6 to 7 days. Intravenous inoculation of the rabbit is followed by death, but no organisms can be found in the tissues at necropsy probably death is due to toxæmia, for the same result is brought about by killed cultures.

*F. furcosus*.—Not so common in appendicitis as *F. fragilis* and *F. savourus*. In pus it is a very small rod with one of its ends forming two little branches like a Y. In culture it forms rods many of these increase in length, and divide at one of their extremities into two branches, each terminated by a swelling, others form branches, which themselves divide. The bacilli and their branches are never very long the swellings are rounded or pyriform, and are numerous. The whole organism is slightly larger than the tubercle bacillus. Non motile and Gram negative. Strict anaerobe. No growth at room temperature, at 37° C. growth is not visible for 3 or 4 days. Surface colonies on agar are very fine grey points, scarcely raised at all from the medium under low magnification they appear yellowish, very finely granular, and have transparent borders. Deep colonies in agar are so small and transparent that they are difficult to see when magnified, they are round and yellowish, with delicate regular borders. A fine precipitate is formed in broth. Cultures have a slightly foetid, sour odour, but evolve very little gas. Cultures remain viable for 15 to 20 days. Subcutaneous inoculation into a guinea pig produces an abscess, which generally heals, sometimes death occurs after several weeks from cachexia.

*F. melaninogenicus*.—Isolated by Oliver and Wherry (1921) from the mouth, infected wounds, urine and faeces of the human subject. On haemoglobin-containing media it forms a melanin like pigment. According to Burdon (1928) it is a small Gram negative non-sporing diplococcobacillus growing feebly on blood media unless mixed with other organisms, when its growth is more profuse. The pigment develops slowly after 1-2 days the colonies may be stained a pale brown in 4-5 days they are a deep black in colour. Cultures have a foul odour. By the time pigmentation is fully developed, the culture is often dead, which may account for the infrequency with which this organism has been isolated and its peculiar chromogenic character recognized. Weiss (1943) reported local inflammatory oedema, followed by necrosis, after intradermal injection of young cultures in rabbits, but no effect after intraperitoneal injection in mice.

Its pathogenicity is probably low. In man, it has been isolated from purulent lesions, usually along with other pathogenic bacteria, and sometimes from the blood.

## REFERENCES

- BACHMANN W and GREGOR, H. (1936) *Z. Immunforsch.*, 87, 250.  
 BEAVER, D. C., HENTHORPE, J. C., and MACT J. W. (1934) *Arch. Path.*, 17, 493.  
 BERENSON, W. I. R. (1924) *J. Path. Bact.*, 28, 45.  
 BURDON K. L. (1928) *J. infect. Dis.*, 42, 161.  
 CASTELLANI, A and CHAIKINS, A. J. (1920) "Manual of Tropical Medicine" 3rd Ed., London.  
 COHEN J. (1932) *Arch. Surg.*, 24, 171.  
 DACK, G. M. and BURROWS, W. (1935) *Proc. Soc. exp. Biol.*, N. Y., 22, 1441.  
 DACK, G. M., DRAGSTEDT, L. R., and HEIST, T. E. (1936) *J. Amer. med. Ass.*, 106, " (1937) *J. infect. Dis.*, 40, 330.  
 DACK, G. M., DRAGSTEDT, L. R., JOHNSON, R., and McCULLOUGH, N. B. (1938) *J. infect. Dis.*, 62, 169.  
 DRAGSTEDT, L. R., DACK, G. M., and KIESER, J. B. (1941) *Ann. Surg.*, 114, 653.  
 EGGERT, A. H. and GAGNON, R. H. (1933) *J. Bact.*, 25, 389.  
 FLEHMANN V. (1904) *Zbl. Bakt.*, 37, 229 (1905) *Ibid.*, 33, 353, (1907) *Z. Hyg. Infectkr.*, 58, 453.

- HALLÉ, J (1898) "Recherches sur la bacteriologie du canal génital de la femme" Thèse de Paris
- HARRIS, J W and BROWN, J H (1927) *Bull Johns Hopk Hosp*, 40, 203
- HENTHORN, J C, THOMPSON, L, and BEAVER, D C (1936) *J Bact*, 31, 255
- HINE, M K (1937) *J infect Dis*, 61, 198
- HINE, M K and BERRY, G P (1937) *J Bact*, 34, 517
- KELLY, F C (1944) *J infect Dis*, 74, 93
- KIRCHHEIMER, E (1940) *Ann Inst Pasteur*, 64, 238
- LEMIERRE, A (1936) *Lancet*, 1, 701
- LEMIERRE, A., GRUMBACH, A., and REILLY, J (1936) *Bull Acad Med, Paris*, 115, 945
- LEMIERRE, A., REILLY, J., and BLOCH MICHEL, H (1937) *Bull Acad Med, Paris*, 117, 322
- LEUKOWICZ, X (1906) *Zbl Bakt*, 41, 163
- LEWIS, K. H., BEDELL, M and RETTGER, L F (1940) *J Bact*, 40, 309
- LEWIS, K. H and RETTGER, L F (1940) *J Bact*, 40, 287
- LOEFFLER, F (1884) *Mitt ReichsgesundhAmt*, 2, 421
- MCCULLOUGH, N B (1938) *J infect Dis*, 63, 34
- MISRA, S S (1933) *J Path Bact*, 46, 204
- OLIVER, W W and WHEBBY, W B (1921) *J infect Dis*, 28, 341
- ORCUTT, M. L (1930) *J Bact*, 20, 343
- PESCH, K L and SCHMITZ, L (1936) *Zbl Bakt*, 136, 476
- PETERS, W H (1911) *J infect Dis*, 8, 455
- PRATT, J S (1927) *J infect Dis*, 41, 461
- PRÉVOT, A R (1938) *Ann Inst Pasteur*, 60, 235
- Report (1920) *J Bact*, 5, 191
- SCHMORL, G (1891) *Dtsch Z Thiermed*, 17, 375
- SCHWYNER, L H and LEE, A M (1934) *J Amer vet med Ass*, 85, 360
- SHAW, F W (1933) *Zbl Bakt*, 129, 132
- SLANETZ, L W and RETTGER, L F (1933) *J Bact*, 26, 599
- SMITH, E C (1933) *J Hyg, Camb*, 33, 95
- SPAULDING, E H and RETTGER, L F (1937) *J Bact*, 34, 535 549
- TEISSIER, P., REILLY, J., RIVALIER, E., and LAYANI, F (1929) *Paris med*, 71, 297
- TEISSIER, P., REILLY, J., RIVALIER, E., and STÉFANESCO, V (1931) *Ann Méd*, 30, 97
- THOMPSON, L. and BEAVER, D C (1931) *Proc Mayo Clin*, 6, 372
- TUNNICLIFF, R (1906) *J infect Dis*, 3, 148, (1923) *Ibid*, 33, 147, (1933) *Ibid*, 53, 280
- VABEY, P L (1927) *J Bact*, 13, 275
- VEILLON, A and ZUBER, A. (1898) *Arch Med exp*, 10, 517
- VINCENT, H (1896) *Ann Inst Pasteur*, 10, 488
- WALKER, P H and DACK, G M (1939) *J infect Dis*, 65, 285
- WEAVER, G H and TUNNICLIFF, R (1905) *J infect Dis*, 2, 446
- WEINBERG, M., NATIVELLE, R., and PRÉVOT, A R (1937) *Les Microbes Anaérobies*, Masson et Cie, Paris
- WEISS, C (1943) *Surg*, 13, 683
- WEISS, C and MERCADO, D G (1938) *J exper Med*, 67, 49
- WEISS, J E and RETTGER, L F (1937) *J Bact*, 33, 423
- WEST, R A., LEWIS, K. H and MILITZER, W E (1942) *J Bact*, 43, 155

## CHAPTER 19

### PFEIFFERELLA, AND CERTAIN ALLIED ORGANISMS

DEFINITION (emended from the American Committee's Report).

Small, slender, usually non motile, Gram negative rods, often staining irregularly and sometimes forming threads or showing a tendency towards branching. Growth on all media is rather slow, gelatin may be slowly liquefied, fermentation of carbohydrates is very weak, characteristic brown honey like growth on potato.

Type species is *Pfeifferella mallei*.

HISTORY.—The glanders bacillus was isolated by Loeffler and Schutz in 1882 (see Loeffler 1886) from a horse dying of acute glanders. An organism in many respects resembling *Pf mallei* was isolated by Whitmore and Krishnaswami (1912) from a glanders like disease of human beings in Rangoon. It was designated *B pseudomallei* by Whitmore (1913). Subsequently Stanton and Fletcher (1921-1925) gave the name of *Melioidosis* to this disease and *Bacillus whitmorei* to the causative organism.

The genus *Pfeifferella* was tentatively created by the American Committee as one of the genera intermediate in position between *Actinomyces* on the one hand and *Mycobacterium* on the other. The type species and the only listed member of the group, is the glanders bacillus or *Pf mallei*. The classification of this organism presents many difficulties and it is by no means certain that the genus *Pfeifferella* will gain permanent recognition.

As regards the fermentative powers of *Pf mallei*, the American Committee's definition states that carbohydrates are not fermented. According to our observations and those of Stanton and Fletcher (1932), acid is formed in glucose in 2 to 3 weeks—a small amount of acid sufficient however to give a pink colour with Andrade's indicator, is produced in salicin. Moreover there is a slow formation of acid in litmus milk, becoming apparent in about 5 days, and followed in 2 to 3 weeks by definite clotting.

In certain respects Whitmore's bacillus resembles *Pf mallei* but it differs from it in many others. For the moment we assign it to the *Pfeifferella* group entering a caveat that should that genus attain a permanent place in systematic bacteriology it is by no means certain that Whitmore's bacillus will be placed within it.

#### Group Characteristics

Morphologically the two organisms are fairly similar, *Pf whitmorei* however is rather smaller frequently shows bipolar staining and is motile. In films of the smooth form the organisms are arranged in long parallel bundles embedded in an interstitial substance presenting a very characteristic appearance, with Loeffler's methylene blue, the interstitial substance stains blue, the bacilli bluish



red, films of the rough forms show no interstitial substance, and resemble *Pf mallei* more closely

Culturally, the growth of *Pf whitmori* is more rapid and more profuse than that of *Pf mallei*. On glycerol agar it may give a smooth mucoid or a corrugated and wrinkled growth. Stanton and Fletcher (1927) have also described an ultra rugose variant, with an extremely corrugated surface, and a consistency so tenacious that if the colony is touched with a needle, it adheres to it and peels bodily off the medium. Growth in gelatin at 20° C is abundant, and liquefaction of the stratiform type is apparent in 4 or 5 days, *Pf mallei*, on the other hand, grows very poorly in gelatin at 20° C and never liquefies the medium, though according to Stanton and Fletcher (1925), if it is incubated at 37° C, permanent liquefaction occurs in 4 to 6 weeks. In broth the growth of the smooth form of *Pf whitmori* resembles that of *Pf mallei*, the rough form, however gives rise to a wrinkled surface pellicle. On potato the smooth form gives a case au lait growth resem-



FIG 91 —*Pfeifferella mallei*

From an agar culture 48 hours 37° C ( $\times 1000$ )

bling that of *Pf mallei*, whereas the rough form gives a much lighter growth of creamy or creamy yellow colour. On MacConkey's agar, *Pf whitmori* grows freely, forming red colonies, while *Pf mallei* entirely fails to grow.

Biochemically, *Pf whitmori* is an active fermenter on first isolation, producing acid in glucose, mannitol, lactose, sucrose and dulcitol, and decolorizing Andrade's indicator, but after long subcultivation in the laboratory it loses its power of fermenting any sugars except glucose. In litmus milk acid is produced in 3 days, later the casein is precipitated, and may be digested, the litmus is partly decolorized. *Pf mallei*,



FIG 92 —*Pfeifferella whitmori*

From an agar culture, 24 hours, 37° C ( $\times 1000$ )

on the other hand, though it may produce acid and clot, never peptonizes the medium. Freshly isolated strains of *Pf whitmori* liquefy blood serum and coagulated egg, but this property is often lost during cultivation in the laboratory.

*Pf mallei* has no digestive action on serum. Both organisms may produce a small amount of  $H_2S$  when tested by Huddleson's method on liver agar (For the preparation of mallein see Chapter 62)

Antigenically the two organisms appear to resemble each other closely Stanton and Fletcher (1925) who examined 5 strains of *Pf mallei* and 14 strains of *Pf ichimori* by agglutination absorption, and complement fixation found that the strains of *Pf ichimori* formed a homogeneous group but that the 5 strains of *Pf mallei* fell into two groups. One of these groups was very closely related to if not identical with the *Pf ichimori* strains. It is important to note that the 2 strains of *Pf mallei* which resembled *Pf ichimori* were isolated from ponies in Java and India respectively since they differed in certain respects from the other strains of *Pf mallei* which were isolated in England and Egypt it is just possible that they were variant strains of *Pf ichimori*. According to de Moor and his colleagues (1930) strains of *mallei* and *ichimori* show cross-agglutination to t tre and complete cross-absorption with specific sera. Cross allergic reactions are also said to be obtainable in infected animals with mallein and the corresponding product prepared from cultures of *Pf ichimori*. Verge and Pairemaure (1928) have reported a positive complement fixation reaction with the serum of a glandered horse in the presence of a *ichimori* antigen. Further work however is obviously desirable on the antigenic structure and relationship of these two organisms.



FIG 93 —  
*Pfeifferella*  
*ichimori*

Gelatin stab culture, 10 days, 20° C showing stratiform liquefaction.

Pathogenically *Pf mallei* gives rise to disease in equines, while *Pf ichimori* is a natural parasite of rodents. Both organisms can cause disease in man both are infective for laboratory animals and both are able to call forth the Straus reaction on intraperitoneal inoculation of male guinea pigs.

**Experimental Reproduction of Glanders in Animals.**—Experimentally glanders may be reproduced in horses asses and mules by feeding with cultures of *Pf mallei* and by subcutaneous inoculation. Sheep and goats are easily infected but cows and pigs are said to be resistant. Of laboratory animals the guinea pig and the field mouse (*Arvicola arvalis*) are the most susceptible to experimental inoculation rabbits and dogs are less so rats

birds and perhaps white mice are comparatively resistant.

**GUINEA PIGS** Loeffler (1886) made 85 experiments on guinea-pigs, and every animal developed the disease. Not all observers, however are agreed on the uniform susceptibility of the guinea-pig young animals seem to be more resistant than older ones.

After subcutaneous inoculation of a small amount of a pure culture an abscess develops, which ulcerates in 5 days the regional glands break down and discharge pus. At this stage the disease may become stationary or retrogress but usually it advances. In the second week hard nodular foci are palpable in the testicles and epididymis inflammation of the tunica vaginalis occurs and the testicle becomes fixed to the overlying skin finally ulceration takes place with the discharge of purulent material. In female guinea-pigs the mammae and labia may be inflamed. Swelling and inflammation of one or more joints is very common, sometimes leading to abscess formation and ulceration. Nodules often appear in the muscles, face back, or beneath the periosteum of the bones. In about a third of the animals nodules appear on the nasal mucosa, and crusts collect around the

external nares. Death follows in 1 to 8 weeks, generally in the 3rd or 4th week. At post mortem there is a local ulcer or scar, the regional glands are swollen, and contain greyish white purulent masses. Abscesses are found in the skin, subcutaneous tissues and around the joints. The lungs contain nodules, greyish yellow and easily emulsifiable, particularly under the pleura. In the spleen there are numerous submiliary nodules of a greyish yellow colour, projecting slightly above the surface. There are fewer nodules in the liver, these are more greyish white in colour. The kidneys are free, but nodules may be seen in the suprarenals. Small, greyish yellow nodules are found in the testicles, or larger foci, relatively firm with a caseous centre. The nasal mucosa is red and swollen, and may be covered with friable caseous masses. In recent lesions the bacilli can always be found, but in the older ones they are scanty.

After intraperitoneal injection the testicles swell in 2 to 3 days, by the 10th day they are greatly enlarged, death occurs in a fortnight as a rule. The testicular lesion—*Straus reaction* (Straus 1889a)—commences in the tunica vaginalis. The two layers of the serosa are covered with confluent yellowish white granules of pinhead size. On the 3rd or 4th day the layers are united by a thick, purulent exudate rich in bacilli. The scrotal skin becomes inflamed and adherent to the underlying tissues, later ulceration occurs (see Panisset 1910). After intraperitoneal injection of organisms of lowered virulence the Straus' reaction may occur without death ensuing.

Guinea pigs have been infected by insufflation with powdered cultures (Babes 1891).

**FIELD MICE**—After subcutaneous inoculation these animals die in 3 to 4 days. Post mortem, there is a greyish green infiltration at the site of injection. The lymph vessels leading to the enlarged glands are studded with little greyish white nodules. The spleen is greatly enlarged, and contains numerous yellowish white nodules projecting slightly above the surface. There are several tiny nodules in the liver.

**WOOD MICE**—The wood mouse—*Mus sylvaticus*—is less susceptible than the field mouse—*Arvicola arvalis*. After subcutaneous inoculation it develops a chronic disease not proving fatal for 3 to 6 weeks. Post mortem, the spleen is enlarged, and contains numerous pin head, greyish spots or nodules, sometimes there is a fibrino purulent exudate in the pleura, and enlargement of the lymphatic glands (Kitt 1887).

**RABBITS**—Subcutaneous inoculation causes a local ulcerating lesion and swelling of the neighbouring lymphatic glands. If the animal is killed after a month, as well as the glandular swelling there may be a few greyish nodules in the lungs, and ulcers on the nasal mucosa. After intravenous inoculation, numerous nodules develop in the spleen and liver, but death may be delayed for some weeks.

**WHITE MICE AND WHITE RATS**—These animals react to subcutaneous injection with a rapidly retrogressive local abscess. Occasionally an animal dies after about 7 weeks, and at post mortem shows caseous nodules in the spleen. Leo (1889) states that white mice can be rendered susceptible to glanders by being fed on a diet containing phloridzin. According to Sabototny (1926), white mice are more susceptible than has generally been believed. In his experiments, after subcutaneous inoculations with a pure culture, the mice often died of an acute infection in 30 to 72 hours, no macroscopic lesions were present at post mortem, but the bacillus could be recovered from the organs. After subcutaneous injection with glanders pus, they all died in 5 to 6 weeks of a chronic infection. Post mortem, the spleen was much enlarged and was riddled with nodules of varying sizes, there were also a few nodules in the liver and lungs.

**DOGS AND CATS**—After subcutaneous inoculation in dogs a local abscess occurs, followed by ulceration, the disease apparently remains localized (Galtier 1881). Acute fatal glanders may follow intravenous injection of large doses of bacilli, numerous small subcutaneous nodules develop, and at post mortem lesions are found in the liver and spleen (Straus 1889b).

Cats are more susceptible, after subcutaneous inoculation a local lesion occurs followed by death in 15 to 30 days; at post mortem nodules are found in the internal organs.

cultures there may be great variation in length and to a less extent in thickness, long filaments showing true or false branching, club pear, flask, and other irregular forms have been described. Staining is usually uneven, deeply stained alternating with poorly stained or unstained areas, bipolar staining is common. Non motile, non-capsulated, non sporing, Gram negative, non-acid fast

*Agar Plate*—2 days at 37° C Round, convex, amorphous, translucent, greyish yellow colonies 0.5–1.0 mm in diameter, with smooth glistening surface and entire edge, butyrous in consistency and easily emulsifiable 9 days, colonies are 1–2 mm in diameter, more opaque, and may have a very finely granular surface, sometimes the centre is coloured slightly brown.

*Agar Slope*—2 days at 37° C Moderate, confluent, raised greyish yellow translucent growth with beaten copper surface and edge undulate or formed of single colonies Medium unchanged Not so profuse as growth of *Bact coli*

*Gelatin Stab*—7 days at 20° C Poor to moderate, filiform growth extending nearly to the bottom of the tube and consisting of small discrete colonies, raised surface growth about 3 mm in diameter No liquefaction After 6 weeks the growth is often brownish in colour If incubated at 37° C gelatin is said to be permanently liquefied in 4 to 6 weeks

*Broth*—2 days at 37° C Moderate growth with moderate uniform turbidity and a slight powdery sediment disintegrating completely No surface growth 14 days, ring growth is present and there is a moderate, viscous deposit, very difficult to disintegrate

*Horse Blood Agar Plate*—3 days at 37° C Good growth of round, low convex, greyish green, opaque colonies 1 mm in diameter No hæmolytic, but plate is browned

*Loeffler's Serum*—3 days at 37° C Scanty growth, barely visible, of flat discrete colonies, 0.3 mm. in diameter No liquefaction

*Potato*—2 days at 37° C Moderate, raised greyish yellow growth After 4 days the colour deepens to café-au-lait, and in 10 days to chocolate

*MacConkey's Agar*—9 days at 37° C No growth.

*Resistance*—Killed by moist heat in 10 minutes at 55° C Cultures dried on threads may live for 3 or 4 weeks, but infected pus or discharge when dried, usually becomes sterile in a few days Killed by 2 per cent phenol in 1 hour, by 1/1000 HgCl<sub>2</sub> in 15 minutes, and by calcium hypochlorite with 2 parts of free chlorine per million in 30 minutes In culture the organism rarely survives longer than a month or 6 weeks

*Metabolism*—Aerobe, either no growth at all or only very slight growth after 14 days, under strict anaerobic conditions Optimum temperature 37°, little or no growth below 20° C. On culture media it grows slowly, growth is often not apparent for 2 days Tendency to formation of brownish pigment in cultures, especially on potato No hæmolytic is produced

*Biochemical*—Acid, no gas in glucose and sometimes very slight acid in saccharin, no other sugars fermented L.M. slight acid, after 2 to 3 weeks a clot forms, and the litmus is partly decolorized at the bottom. Indole —, M.R. —, V.P. —, nitrates reduced, H<sub>2</sub>S slight +, NH<sub>3</sub> +, M.B. reduction —, catalase slight +

*Antigenic Structure*—By agglutination, absorption and complement fixation there appear to be at least two types of *Pf mallei*, of which one is related to *Pf whitmorei* Saka moto (1930) has isolated from culture filtrates of *Pf mallei* a relatively non specific nucleo protein substance, and a soluble specific polysaccharide which gives a precipitation reaction with immune sera

*Pathogenicity*—Causes glanders or farcy in horses mules asses, and man Experimentally the disease can be reproduced in equines, goats, cats and guinea pigs sheep rabbits, dogs, rats and mice are less susceptible, pigs and cattle are resistant Intrapertoneal injection of the male guinea pig is followed in 2 days by swelling of the testicles, after 3 or 4 days the two layers of the tunica vaginalis are gummed

together by a thick, purulent, yellowish exudate. The scrotal skin becomes adherent, and ulcerates. Death in 14 days. P.M., besides the testicular lesions, there may be submiliary nodules in the spleen projecting above the surface, greyish white nodules in the lungs and liver, and subcutaneous or periarticular abscesses. Bacilli are present in the fresh lesions.

### *Pfeifferella whitmorei*

*Isolation*—Isolated by Whitmore and Krishnaswami (1912) from human patients with melioidosis

*Synonyms*—*B. pseudomallei*, *B. whitmorei*

*Habitat*.—Parasite of rodents and man

*Morphology*—Small, slender rods, 1-2  $\mu$  long and 0.4-0.5  $\mu$  broad, sides parallel, ends rounded, axis straight, arranged singly, in pairs end-to-end, or sometimes in long parallel bundles, the bacilli being embedded in an interstitial substance. Variations occur in depth of staining, bipolar staining common, especially in films from infected tissues. Motile. Gram negative, acid fast granules have been described in freshly isolated strains (Finlayson 1944). The short, oval, bipolar-stained rods are characteristic of the rough form, the longer, narrower rods, arranged in palisades with irregular staining and shadow forms, are characteristic of the smooth variant.

*Agar Plate*.—24 hours at 37° C Round, amorphous, low convex, translucent, greyish yellow colonies, 1-2 mm. in diameter, smooth, glistening surface and entire edge, consistency mucoid, emulsifiability easy. 14 days, colonies are opaque, often coloured yellow, brown, or pinkish, and may have a wrinkled centre.

*Agar Slope*.—24 hours at 37° C Abundant, confluent, raised, greyish-yellow, mucoid, spreading growth, with glistening, beaten-copper surface, and edge undulate or made up of single colonies. Growth more profuse than that of *Bact. coli*. Cultures have a mouldy, earthy smell.

*Gelatin Stab*.—10 days at 20° C Abundant, filiform growth, mostly of discrete colonies extending to bottom of tube. Stratiform liquefaction; between the liquefied and unliquefied portions of the gelatin there is a thick nodular pellicle of growth.

*Broth*.—24 hours at 37° C Good growth with moderate turbidity, and slight powdery deposit disintegrating completely, may be slight pellicle formation. 10 days, luxuriant growth with dense turbidity and a heavy viscous deposit disintegrating with difficulty. The rough form gives rise to a slight turbidity and a wrinkled surface pellicle. According to Nicholls (1930), the rough form produces about 0.2 per cent oxalate (calculated as calcium oxalate) in 4 days, the smooth form produces not more than about 0.01 per cent., but renders the medium alkaline—pH 8.4.

*Glycerol Agar*.—3 days at 37° C May be (1) profuse mucoid growth with smooth glistening surface, or (2) profuse growth with dull, wrinkled, corrugated, or honeycombed surface.

*Horse Blood Agar Plates*.—3 days at 37° C Abundant growth of round, low convex, greyish green colonies, 1-2 mm. in diameter. No haemolysis, except, perhaps, with freshly isolated strains.

*Loeffler's Serum*.—3 days at 37° C Good, confluent, slightly raised, creamy growth with smooth surface and undulate edge. Liquefaction by freshly isolated strains after a variable number of days. Sometimes the growth itself appears to become liquefied, and runs down the slope.

*Potato*.—24 hours at 37° C S type good growth of a creamy or lemon yellow colour not very easy to see, later the colour deepens to café-au-lait or chocolate. R. type forms dry, dull, dirty white growth.

*MacConkey's Agar*.—3 days at 37° C Abundant growth of red, opaque, low convex or umbonate colonies, 2-3 mm. in diameter.

**Resistance**—Destroyed by moist heat at 56° C within 10 minutes. Killed within about 10 minutes by 1 per cent phenol and 0.5 per cent formol. Organisms may survive for a month or more in water, feces and dried soil, and for a week in putrefying carcasses.

**Metabolism**—Aerobe, very slight growth on agar and in broth after 14 days under strict anaerobic conditions. Optimum temperature for growth 37° C. In culture it grows more rapidly and more abundantly than *Pf mallei*. No hæmolytic of sheep or horse blood, except perhaps with freshly isolated strains. Tendency to formation of brownish pigment in cultures, especially on potato. Oxalates formed in broth by rough form.

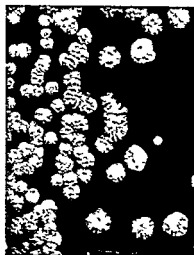


FIG 94—*Pf whitmori*

Surface colonies on glycerol agar plate  
Rough form  
(After Stanton and Fletcher)



FIG 95—*Pf whitmori*

Growth on glycerol agar. Left smooth form. Right rough form.  
(After Stanton and Fletcher)

**Biochemical**—On first isolation it forms acid, no gas in glucose, maltose, mannitol, lactose, dextrose, dextrin and sucrose, after long cultivation in the laboratory it may attack glucose only. Andrade indicator decolorized in 4–10 days. LM may be slightly acid in 3 days, casein is coagulated and may be digested, partial decolorization of litmus. Indole —, MR —, VP —, nitrates reduced,  $\text{NH}_3$  +,  $\text{H}_2\text{S}$  ± when tested by Huddleson's method on liver agar, MB reduction +, catalase +.

**Antigenic Structure**—By agglutination, absorption, and complement fixation cultures of *Pf whitmori* form a homogeneous group, which is said to be closely allied to one group of *Pf mallei*.

**Pathogenicity**—Causes melioidosis in man, rats, cats, dogs, guinea pigs, rabbits and occasionally horses. Experimentally rodents can be infected by feeding, injection into the tissues or by rubbing on the scarified skin. Produces abscess at site of inoculation, generalized adenitis, and nodular abscesses of the spleen and lungs. After ip injection of a small quantity of culture into a male guinea pig swelling of the testicles occurs in 2 days with caseous exudate between the two layers of the tunica vaginalis. If a large dose is used, death may occur in 24 hours from septicæmia, before there is time for the Straus reaction to develop or for nodular lesions to appear in the viscera. Small doses prove fatal in about a week. Guinea pigs are so susceptible that even minute numbers of bacilli brought into contact

with the scarified skin may give rise to a fatal infection. Rabbits are as susceptible as guinea-pigs, but rats are more resistant. The subcutaneous inoculation of 0.1 ml. of a 24 hour broth culture into a rabbit usually causes death in 3 weeks. Post mortem there is local necrosis and necrotic or purulent lesions are found in the lungs, spleen, liver, joints, bones or testicles, purulent arthritis is common in the second week. Cats, monkeys and goats can all be infected experimentally. The disease in the monkey is usually chronic, lasting 2 to 4 months, and is accompanied by severe emaciation. Horses are resistant (see Stanton and Fletcher 1937 de Moor *et al* 1932 Nicholls 1934). According to Nicholls (1934) the rough type is the normal virulent form while the smooth type is relatively avirulent if obtained completely free from rough organisms.

### Perez's Bacillus, or *Cocco-bacillus foetidus ozænae*

In 1899 Perez described under the name of *Cocco-bacillus foetidus ozænae* a small organism that he had isolated from the nose of patients suffering from ozæna. His work was confirmed by numerous observers (Hofer 1913a, b, Ward 1916, 1917, Busson 1923, Olinescu and Atineu 1925a, b) but there is insufficient evidence to show that this organism is causally related to ozæna. Perez (1901, 1913) found his bacillus also in the saliva and nasal mucus of apparently healthy dogs.

To avoid confusion it may be stated here that Perez's bacillus is different from the ozæna bacillus described by Loewenberg (1894) and by Abel (1896) which undoubtedly belongs to the Friedländer group (see Chapter 28).

The classification of Perez's bacillus is very doubtful, it is impossible to say to what group this organism will ultimately be assigned. Our reasons for including it in this chapter are to draw attention to certain points of resemblance that it shows to *Pf. whitmorei* and *H. bronchisepticus*.

Morphologically Perez's bacillus is a small, Gram negative cocco-bacillus which was originally described as being non motile but which is said by Ferry and Noble (1918) to be sluggishly motile and by Ward (1917) occasionally to acquire motility in culture. In fluid media and in old cultures on solid media long irregular deeply staining filamentous forms may appear. In broth, and on agar if the tubes are sealed, cultures have a heavy sweet unpleasant or even nauseating odour which is said by Ward to be due to volatile sulphur compounds. According to our own observations, agar cultures tend to wither after a few days suggesting the occurrence of autolysis. In gelatin stab cultures of Perez's bacillus fine lateral filaments often appear in 2 or 3 weeks radiating from the central stab. On potato a creamy white growth is formed. Biochemically the organism produces acid and gas in glucose only, in litmus milk there may be a slight acidity, indole is produced. Antigenically all strains are said by Ward (1917) to be homogeneous but according to Olinescu and Atineu (1925a) this is doubtful. An anti-serum to *H. bronchisepticus* is without action on Perez's bacillus and vice versa (Ferry and Noble 1918). Both Perez's bacillus and *H. bronchisepticus* are parasites of the respiratory tract and both seem to be secondary rather than primary agents in the causation of disease. It is probable that in the past they have been confused with each other. The following table (Table 33) modified from Ferry and Noble enumerates the main differences between them.

An organism similar to Perez's bacillus has been isolated by Shiga (1922) from the nose of ozæna patients. The chief differences between the two are that Shiga's organism often ferments maltose and sucrose as well as glucose it sometimes peptonizes milk it liquefies gelatin and it is not agglutinated by a serum prepared against Perez's bacillus. Blanco and Pangalos (1925) have isolated from the nose of ozæna patients a short actively

TABLE 33

DIFFERENTIATION OF *H. bronchisepticus* FROM PEREZ'S BACILLUS

	<i>H. bronchisepticus</i>	Perez's bacillus
<i>Morphology</i>	Small narrow bacillus	Small coccoid bacillus
<i>Gelatin Stab</i>	No lateral filaments	Lateral filaments after 2 to 3 weeks
<i>Potato</i>	Yellowish brown growth	Creamy white growth
<i>Lysmus Milk</i>	Alkaline	Slightly acid.
<i>Biochemical</i>	No fermentation	Acid and gas in glucose
<i>Indole</i>	—	+

motile Gram negative bacillus which they call *B. ozogenes*. It coagulates milk ferments glucose maltose mannitol lactose and sucrose with the production of acid and gas gives a variable indole reaction and forms  $H_2S$ .

**Cocco-bacillus fætidus-ozænæ** Perez

*Synonym*—Perez's bacillus

*Isolation*—By Perez in 1890 from nose of ozæna patients

*Habitat*—Nose of ozæna patients saliva and nasal mucus of normal dogs Probably strict parasite.

*Morphology*—Small coccoid-bacillus 1.2  $\mu$  long by 0.5  $\mu$  broad some strains are coccoid some are bacillary Parallel or bulging sides rounded ends axes straight arranged singly in bundles of two or three members and occasionally in pairs end-to-end. Long sinuous filaments and long straight or curved bacilli may be seen especially in old cultures. Staining is fairly even. Non-motile or sluggishly motile. Non-capsulated. Gram negative.

*Agar Plate*—24 hours at 37° C Round amorphous low convex greyish white translucent colonies up to 1 mm in diameter smooth glistening surface and entire edge consistency butyrous or viscous emulsifiably easy 7 days colonies are larger 1.2 mm. in diameter but owing to autolysis are flattened almost colourless less transparent and adherent to the medium surface is dull and very finely granular

*Agar Slope*—24 hours at 37° C Moderate confluent slightly raised greyish yellow slightly translucent growth with glistening beaten-copper surface and entire edge condensate on water consists of greyish white mucoid material. 7 days whole growth is flattened, and withered surface dry and covered with secondary colonies.

*Gelatin Stab*—7 days at 22° C Moderate mostly confluent filiform growth greyish white in colour extending to bottom of tube Surface growth 2–4 mm in diameter slightly raised or flattened. No liquefaction even after 3 weeks After 2 to 3 weeks delicate feathery outgrowths from the stab appear in the upper half of the medium.

*Broth*—24 hours at 37° C Moderate growth with moderate uniform turbidity and a slight powdery deposit disintegrating completely heavy unpleasant sweetish odour No surface growth.

*Horse Blood Agar Plate*—24 hours at 37° C Abundant greyish white growth No hæmolysis but plate is browned.

*Potato*—7 days at 37° C Poor confluent slightly raised creamy white growth with smooth surface potato browned.

*MacConkey's Agar*—24 hours at 37° C Good growth of small round low convex yellowish colonies not unlike those of *Salmonella typhi*.



*Resistance*.—Not known.

*Metabolism*.—Aerobe, facultative anaerobe. Optimum temperature for growth 37° C., but growth occurs slowly at 20° C. No hæmolysis of sheep's or horse's red corpuscles. Old cultures have a tendency to become brownish in colour. Fætid odour, especially noticeable in broth or serum broth cultures, and in corked agar cultures.

*Biochemical*.—Acid and gas in glucose only. L.M. slight acid. Indole +, M.R. —, V.P. —, nitrates reduced, H<sub>2</sub>S +; NH<sub>3</sub> ++; M.B. reduced, catalase ++.

*Antigenic Structure*.—Appears to be antigenically homogeneous, as tested by agglutination and complement fixation, but not many strains have been tested.

*Pathogenicity*.—Pathogenic for guinea pigs, rabbits, and mice. After intravenous injection of rabbits a mucopurulent, sometimes hæmorrhagic, nasal discharge develops, containing large numbers of the bacilli. Chronic rhinitis may develop, with atrophy of the turbinate bones. Death occurs in from 24 hours to several weeks, according to the dose injected. Intraperitoneal injection of 0.25 ml. of a saline suspension off agar into guinea pigs is fatal in 12 hours, post mortem, there is peritonitis with abundant hæmorrhagic exudate. A similar dose injected subcutaneously into mice is fatal in 48 hours, post mortem, there is gelatinous oedema at local site, and enlargement of the spleen. Neither in guinea pigs nor in mice can the bacillus be recovered from the blood.

## REFERENCES

- ABEL, R. (1896) *Z. Hyg. Infektkr.*, 21, 89.  
 BABES, V. (1891) *Arch. Méd. exp.*, 3, 619.  
 BLANC, G. and PANGALOS, G. (1925) *C. R. Soc. Biol.*, 93, 1267, 1268.  
 BUSSON, B. (1923) *Munch. med. Wochr.*, 70, 426.  
 FERRY, N. S. and NOBLE, A. (1918) *J. Bact.*, 3, 499.  
 FINLAYSON, M. H. (1914) *S. Afr. med. J.*, 18, 113.  
 GALLI VALENZIO, B. (1900) *Zbl. Bakt.*, 28, 303.  
 GALTIER, V. (1881) *C. P. Acad. Sci.*, 92, 303.  
 HOYER, G. (1913a) *Berl. klin. Wochr.*, 50, 2413; (1913b) *Wien. klin. Wochr.*, 26, 1011.  
 KITT, T. (1887) *Zbl. Bakt.*, 2, 241.  
 LEGROUX, R. and DUEHL, K. (1931) *C. R. Acad. Sci.*, 193, 1117.  
 LEGROUX, R. and GENEVREY, J. (1933) *Ann. Inst. Pasteur*, 51, 249.  
 LEO, H. (1889) *Z. Hyg. Infektkr.*, 7, 503.  
 LOEFFLER, (1886) *Arch. Psychiatr. u. Nervenheilk.*, 1, 141.  
 LOEWENBERG, (1894) *Ann. Inst. Pasteur*, 8, 292.  
 MAYER, G. (1900) *Zbl. Bakt.*, 28, 673.  
 MOOR, C. E. DE, SOKKARVEN, and WALLÉ, V. V. D. (1932) *Genetisk Tydschr. Ned. Ind.*, 24, 1618.  
 NICHOLLS, L. (1930) *Brit. J. exp. Path.*, 11, 393; (1934) *Ceylon J. Sci., Sect. D med. Sci.*, 3, 183.  
 OLIVESCIO, R. and ATINCE, A. (1925a) *C. R. Soc. Biol.*, 93, 740; (1925b) *Ibid.*, 93, 741.  
 PANISSET, L. (1910) *Rev. gen. Méd. vet.*, 15, 561.  
 PEREZ, F. (1899) *Ann. Inst. Pasteur*, 13, 937; (1901) *Ibid.*, 15, 409; (1913) *Berl. klin. Wochr.*, 50, 2411.  
 SABOLOTNY, S. S. (1926) *Zbl. Bakt.*, 93, 37.  
 SAKAMOTO, K. (1930) *J. Immunol.*, 18, 331.  
 SHIGA, M. (1922) *Zbl. Bakt.*, 88, 521.  
 STANTON, A. T. and FLETCHER, W. (1921) *Proc. 4th. Congr. Far Eastern Ass. trop. Med. Hyg.*, 2, 196; (1925) *J. Hyg., Camb.*, 23, 347; (1927) *Ibid.*, 26, 31; (1932) *Studies Inst. med. Res., F.M.S.*, No. 21.  
 STRAUS, I. (1889a) *Arch. Méd. exp.*, 1, 460; (1889b) *Ibid.*, 1, 489.  
 THOMPSON, L. (1933) *J. Bact.*, 26, 221.  
 VERGE, J. and PATREMAUKE, O. (1928) *C. P. Soc. Biol.*, 99, 18.  
 WARD, H. C. (1916) *J. infect. Dis.*, 19, 153; (1917) *Ibid.*, 21, 338.  
 WHITMORE, A. (1913) *J. Hyg., Camb.*, 13, 1.  
 WHITMORE, A. and KRISHNASWAMI, C. S. (1912) *Indian med. Gaz.*, 47, 262.  
 WILSON, G. S. (1934) *J. Hyg., Camb.*, 34, 361.

## CHAPTER 20

### AZOTOBACTER, RHIZOBIUM, NITROSOMONAS, NITROBACTER, HYDROGENOMONAS, MITHANOMONAS, CARBOXYDOMONAS, AND ACETOBACTER

In this chapter we group together a number of organisms playing an important part in the nitrogen metabolism of the soil.

#### THE AZOTIFYING OR NITROGEN FIXING BACTERIA

##### DEFINITION — *Azotobacter*

Relatively large rods, or even cocci, sometimes almost yeast like in appearance, dependent primarily for growth energy upon the oxidation of carbohydrates. Motile or non motile, motile forms possess a tuft of polar flagella. Obligate aerobes, usually growing in a film upon the surface of the culture medium. Capable of fixing atmospheric nitrogen when grown in solutions containing carbohydrates and deficient in combined nitrogen.

Type species — *Azotobacter chroococcum*, Beijerinck.

Isolated by Beijerinck in 1901. He described two species, *Az. chroococcum*, so called from the brown pigment to which it gives rise, and *Az. agilis*. The former is widespread in garden earth and in fruitful soil of all kinds, the latter was found in canal water in Holland. Since then four other species have been described—*Az. vinelandii*, *Az. beyerinckii*, *Az. woodstockii*, and *Az. vitreus*, all of them inhabit the soil.

All species of *Azotobacter* are highly pleomorphic, the cells may be short, thick and rod like, ellipsoidal, pyriform, or spindle-shaped, when occurring in pairs they often look like giant diplococci. Their size varies from 4–7  $\mu$  in length and 1.5–4  $\mu$  in breadth. Giant involution forms, looking like amebae or yeasts, are not uncommon or possibly peritrichate (Hofar 1914).

The organism is strictly aerobic.



FIG. 96 — *Azotobacter chroococcum*  
From an agar culture, 3 days 30° C ( $\times 1000$ )

They are motile by one or more polar flagella and are Gram negative.

It grows best in tap water containing 2 per

cent mannitol and 0.02 per cent  $K_2HPO_4$ . If the medium is spread out in a thin layer in a wide-bottomed flask, *Azotobacter chroococcum* forms a surface pellicle in 2 or 3 days, which gradually becomes brown, and may later even turn black. Potassium, calcium, or sodium propionate in 0.5 per cent solution may be substituted for the mannitol.

Of solid media one of the best has the following composition

Mannitol	2 gm
$K_2HPO_4$	0.02 "
Washed Agar	2 "
Aq Dist	100 "

*Azotobacter* grows best in solutions containing little or no combined nitrogen, according to Beijerinck (1901) ammonium salts are not easily assimilated and peptone can be used only to a slight extent.

The peculiar property of this group of organisms is to fix atmospheric nitrogen and to convert it into ammonia, nitrites and nitrates (Beijerinck and van Delden 1902). To do this they must be supplied with a source of energy in the form of a suitable carbohydrate, such as mannitol. The energy gained from the oxidation of this substance is utilized in the fixation of nitrogen. Garney (1918) found that in a synthetic mannitol medium inoculated with soil, about 8 mgm. of nitrogen were fixed for 50 ml of medium. Not all strains of *Azotobacter* are capable of utilizing mannitol (Smith 1930). *Azotobacter* is more susceptible to acid than most soil organisms. Its growth limits are about pH 6.5-8.6 in pure culture (Fred and Davenport 1918) but in soil it is probable that growth can occur down to about pH 6.0. Pigment formation is variable in *Az. chroococcum*, some variants form a brown pigment, others are achromogenic, intermediate forms occur (Omelianskv and Sewerowa 1911). Pigment is formed only in the presence of a free supply of oxygen. We give a description of the type species *Az. chroococcum* followed by notes on the lesser known species.

#### *Azotobacter chroococcum*

*Isolation*—By Beijerinck in 1901.

*Habitat*—Soil.

*Morphology*—Short thick rods with rounded ends, large ovoid forms, forms like giant diplococci, pear-shaped rods, and other forms.  $4-5 \mu \times 1.5-2 \mu$ . Slowly motile by polar flagella. Arranged singly, in pairs end-to-end, and in old cultures in packets. Cells often vacuolated, when grown on mannitol, cells may contain fat droplets. In surface membranes on liquid media cells are surrounded by a thick mucoid capsule. Large involution forms not uncommon. Non-sporing. Gram-negative. Non acid fast.

*Agar Plate*.—3 days at  $30^\circ C$ . Round colonies 1 mm. in diameter, convex, with smooth, moist glistening surface, edge entire, structure amorphous, consistency butyrous, easily emulsifiable, often differentiated into an opaque brown centre and a translucent lighter periphery. After 6 days the colonies measure up to 3 mm. in diameter, some remain homogeneous undifferentiated and opaque, others show an opaque brownish centre and a clear slightly radiate periphery.

*Agar Slope*.—2 days at  $30^\circ C$ . Moderate confluent or partly confluent slightly raised, translucent greyish-yellow growth with glistening beaten-copper surface, edge formed of single colonies.

*Gelatin Stab*.—6 days at  $20^\circ C$ . Poor to moderate, greyish white, filiform growth, consisting chiefly of discrete colonies, extends  $\frac{2}{3}$  way down tube, surface growth, 3-4 mm. in diameter with a lobate edge and irregular surface. No liquefaction.

*Broth*—2 days at 30° C Moderate growth, with moderate finely granular turbidity, and slight, finely granular sediment, not disintegrating on shaking After 6 days, surface ring growth.

*Loeffler's Blood Serum*—6 days at 30° C Moderate, confluent, creamy growth, with slight yellowish colour, smooth, mirror like surface, no liquefaction

*Potato*—10 days at 30° C Abundant, raised, opaque brown growth, with wrinkled honeycombed surface During first few days, growth is yellowish, but later it becomes brown. Potato unchanged.

*Resistance*—Killed by 55° C. in 30 minutes

*Metabolism*—Aerobic. No growth anaerobically Opt temp 28 30° C Limits of pH 6.5–8.6 in pure culture Pigment, grown in mannitol and other suitable media, a surface scum is formed, which becomes brown, and later black Pigment is insoluble in water, alcohol, CHCl<sub>3</sub>, ether, CS<sub>2</sub>, but slightly soluble in alkalis which destroy it

Nutritional. Grows best in 2–10 per cent mannitol solutions and in 0.5 per cent solutions of K, Ca, or Na propionate Growth on agar improved by 1 per cent mannitol

*Biochemical*—Fixes atmospheric nitrogen, converting it into NH<sub>3</sub>, nitrite and nitrate Capable of growing in media free from combined nitrogen but containing carbohydrates Can utilize nitrates for its nitrogen supply, NH<sub>3</sub> and peptone can be used only slightly

Utilizes dextrose, maltose, mannitol, lactose, dextrin, starch, glycerol, alcohol propionate, acetate, butyrate, citrate, lactate, malate, and succinate, gives off CO<sub>2</sub>, Indole— M/R — V/P — Nitrates— M/R reduction— Catalase + NH<sub>3</sub>, very slight L.M. partly decolorized and rendered clearer and more fluid

*Pathogenicity*—Nil

*Az. agilis* is a large, oval organism containing granules and vacuoles and provided with a bundle of polar flagella The other four species of *Azotobacter* that have been described differ in minor respects from the type species *Az. chroococcum*.

Numerous non sporing rod shaped organisms, capable of fixing nitrogen, have been found in horse and cow dung by Fulmer and Fred (1917) The chief of these, which they call *B. azophila*, is a rod shaped organism, 1.6 × 0.8 μ, motile and Gram positive It gives a light orange, wrinkled growth on agar, a heavy membranous surface growth in broth, and a brownish growth on potato Gelatin is liquefied, milk is peptonized Nitrates are reduced to nitrites and to gaseous nitrogen Indole + Strict aerobe Grows well in mannitol solution in which it fixes nitrogen in considerable quantities.

## RHIZOBIUM

### DEFINITION—*Rhizobium*

Minute rods, motile when young Specialized forms abundant and characteristic when grown under suitable conditions Obligate aerobes<sup>1</sup> capable of fixing atmospheric nitrogen when grown in the presence of carbohydrates and in the absence of compounds of nitrogen. Produce nodules on the roots of leguminous plants

Type species *Rhizobium leguminosarum*, Frank.

The first member of this group was isolated by Beijerinck in 1888, who named it *Bacillus radicicola* He found this organism in the root nodules of leguminous plants, and in numerous specimens of soil and water of different origin, he noted its variable morphology, he described its cultural and biochemical reactions, and he showed how the bacilli in the "swarmer" stage penetrated

<sup>1</sup> This is not absolutely certain we have observed growth of one strain in broth, though not on agar, under strictly anaerobic conditions

the pore-spaces in the roots of certain plants. Though suspecting it of being capable of fixing atmospheric nitrogen, he was unable to demonstrate this conclusively. Numerous strains of *Plazobium* have since been described, but as it is doubtful whether they are separate species, we shall confine ourselves to a description of *Plaz. leguminosarum*.

In pure cultures on nutrient agar this organism occurs in the form of rod,  $1.5-3 \mu \times 0.5 \mu$ , but by varying the constituents of the medium it can be made to pass through a cycle of changes described by Bewley and Hutchinson (1930) as follows: (1) Pre-swarmers stage, non motile. When a pure culture is inoculated into a neutral soil solution, the organisms assume this form in 4 to 5 days. Diameter about  $0.4 \mu$ . (2) Pre-swarmers stage, larger, non motile, cocci,  $0.8 \mu$  in diameter. Appear in mannitol agar. (3) Swarmers stage, very actively motile. Cells are ellipsoidal,  $0.9 \mu \times 0.18 \mu$  (Benjaminck). They are so small as to be



FIG. 9.—*Rhizobium leguminosarum*.

1. Pre-swarmers first stage. 2. Pre-swarmers second stage. 3. Swarmer. 4. Motile rod.  
5. Highly vacuolated rods.  
(After Bewley and Hutchinson.)

able to pass through a Chamberland filter. Appear in carbohydrate media. (4) Pod stage motile. Appearance favoured by carbohydrates, as long as these are abundant, the organism remains in this stage. Dimensions  $3.4 \mu \times 1 \mu$ . (5) Stage of high vacuolation. In a neutral soil extract, or in a medium in which the carbohydrate has been exhausted, the organisms become highly vacuolated, the chromatin divides into a number of bands. Later these bands become rounded off and escape from the rod as the coccoid pre-swarmers (Fig. 9.) The formation of the pre-swarmers may also be induced by the addition of calcium and magnesium carbonate to the medium, or by incubating the culture anaerobically. As Lewis (1933) points out, there is no reason to regard this succession of morphological changes as essentially different from that seen in other groups of bacteria, or as indicating the occurrence of a specialized form of reproduction, the vacuolated state appears to depend on the presence of fat globules.

Calcium phosphate causes a change from pre-swarmers to rods. Acid soils favour the production of highly vacuolated cells, and eventually kill the organisms.

Slightly alkaline soils support vigorous growth without altering the morphology. Relatively high temperatures, 30–37° C, prevent or postpone the change into the pre swarming stage.

Inside the root nodules the organisms assume curious Y shaped, pyriform, and racket-like forms, known as bacteroids. Filaments may likewise be formed. In culture the bacteroids develop into rods (Fremlin 1898).

The properties of *Rhiz leguminosarum* vary in accordance with the species of plant from which they are derived. Fred and Davenport (1918) who studied 21 strains from different *Leguminosae*, found a variation in their resistance to acids. Thus the strains from alfalfa and sweet clover had a limiting pH for growth of 4.9, for the garden pea and vetch of 4.7, for red clover and common beans of 4.2, for soya beans and velvet beans of 3.3, and for lupins of 3.15. All strains had much the same resistance to alkali. Incidentally their alkali tolerance was much greater than their acid tolerance.

Whether the strains from different plants should be regarded as varieties of one species is doubtful. Klummer and Krüger (1914), who examined a number of strains from eighteen different species of *Leguminosae* by means of the agglutination, complement-fixation, and precipitation tests, were able to classify them into 9 different groups. Bushnell and Sarles (1939) likewise found a large number of serological types. In their experience little relationship was noted between the ability of strains from different plants to cross agglutinate and to cross infect (see also Fremlin 1898, Kleczkowski and Thornton 1944).

*Rhiz leguminosarum* is an obligate aerobe, capable of fixing atmospheric nitrogen when grown in a medium free from combined nitrogen, but containing a fermentable carbohydrate. Such a medium is composed of

Manntol	10.0 gm
K <sub>2</sub> HPO <sub>4</sub>	0.2 "
NaCl	0.2 "
MgSO <sub>4</sub> ·7H <sub>2</sub> O	0.2 "
CaSO <sub>4</sub> ·2H <sub>2</sub> O	0.1 "
Distilled Water	1000 "

Inoculated into the root of a leguminous plant it gives rise to a nodule, from which it can be recovered in pure culture. Each variety is apparently specialized to attack its own or closely related species of plants, thus a strain isolated from a nodule on a pea (*Pisum sativum*) will produce nodules also on *vicia*, *Lathyrus* and *Lens*, but not on other legumes (Russell 1923).

We append a detailed description of *Rhiz leguminosarum*.

### *Rhizobium leguminosarum* Frank

*Synonyms*—*Bacillus radicola*, *Ps radicola*, nodule organism.

*Isolation*—By Beijerinck in 1888 from root nodules of leguminous plants.

*Ecology*—Soil, found also in water.

*Morphology*—Has a life-cycle with gross changes in morphology. Shows non motile coccoid forms, very small, highly motile ellipsoidal form (swarmer), 0.9  $\mu$   $\times$  0.2  $\mu$ , motile rods, 2–3  $\mu$   $\times$  0.5  $\mu$ , large "vacuolated" rod and Y forms. In root nodules shows filamentous forms, and Y forms (bacteroids). Swarmer forms motile by single, long, polar flagellum. Vacuolated forms show bands of chromatin. Non sporing, may be capsulated (see below). Gram negative. Non-acid fast.

*Agar Plate*—2 days at 25° C Round, 1 mm. in diameter low convex, greyish-yellow transparent colonies with smooth, glistening surface and lobate edge. Structure amorphous, consistency butyrous, easily emulsifiable. Differentiated into a smooth, raised, darker centre and a thin, effuse, radiate periphery. Some strains are said to form mucoid colonies, consisting of bacilli embedded in a mucous material which appears to consist of a carbohydrate material, yielding glucose on inversion (Kramar 1921).

*Agar Slope*—2 days at 25° C Moderate partly confluent slightly raised, greyish-yellow translucent growth, with glistening beaten-copper surface, and finely lobate edge.

*Gelatin Slab*—5 days at 20° C Moderate filiform growth extending to bottom of tube, slight surface growth 1 mm. in diameter, no liquefaction except in very old cultures.

*Broth*—2 days at 25° C Poor growth with slight turbidity, and slight powdery deposit disintegrating on shaking. 5 days, moderate turbidity, slight surface ring growth, and moderate viscous deposit giving rise on shaking to dense turbidity.

*Loeffler's Elood Serum*—6 days at 25° C Abundant, mostly confluent, raised, chrome-yellow growth, slightly tinted with pink, smooth glistening surface. Medium not coloured, no digestion.

*Potato*—6 days at 25° C Poor effuse, confluent whitish-pink growth, with smooth, moist glistening surface, medium not coloured, or coloured slightly grey.

*Resistance*—Killed at 60° C. in 1 hour. Resists drying and freezing.

*Metabolic*—Aerobic. No growth anaerobically on agar but moderate growth in broth.

Opt. temp. 25° C. Pigment yellowish-pink, not marked, on certain media only.

*Biochemical*—No acid or gas in glucose, malose, mannitol, lactose sucrose or salicin.

Indole—M.R.—V.P.—Nitrate reduction—Catalase+ M.R. reduction— $\text{NH}_3$ +  $\text{H}_2\text{S}$  slight+ L.M. acid and clot in 21 days. Capable of fixing atmospheric nitrogen in the presence of carbohydrates and in the absence of combined nitrogen.

*Antigenic Structure*—Can be divided by agglutination into numerous different groups.

*Pathogenicity*—Nil to man or animals. Produces nodules upon the roots of the *Leguminosae*.

## NITROSOMONAS

**DEFINITION**—*Nitrosomonas*.

Cells rod-shaped or spherical, motile or non-motile, motile forms possess polar flagella. Capable of securing growth energy by the oxidation of ammonia to nitrites. Growth on media containing organic substances is scanty or absent.

Type species *Nitrosomonas europaea*, Winogradsky

Winogradsky (1890a b c) finding that nitrification did not occur in a medium containing organic matter, seeded with soil prepared the following solution

Ammonium sulphate	1 gm.
Potassium phosphate	1 "
Pure water	1000 "

To each 100 ml. was added 0.5–1.0 gm. of basic magnesium carbonate, when this medium was inoculated with soil, nitrification occurred satisfactorily, the ammonia being oxidized to nitrite. Five different types of organisms were found, one of them, which grew around the particles of carbonate at the bottom of the flask, failed to grow in gelatin on transplantation, but could be cultivated in a purely inorganic medium. This organism was obtained in pure culture, and was found to be responsible for nitrification. Morphologically it consists

of ellipsoidal cells, intermediate between cocci and bacilli, its dimensions are  $1.1-1.8 \mu \times 0.9-1.0 \mu$ . It is motile by a single long flagellum. It is arranged singly, or aggregated into a zooglial mass by some slightly viscous substance, chains even of 3 or 4 members are rare.

Winogradsky found that this organism could grow in a medium devoid of all traces of organic matter. It must therefore obtain its carbon from the magnesium carbonate added to the medium, that is to say, it can assimilate the carbon of carbonic acid. It obtains its nitrogen from the ammonium sulphate, and oxidizes it to nitrite. From purely inorganic substances it can therefore synthesize organic matter—a process rarely accomplished independently of chlorophyll and sunlight.

Experiments showed that the amount of ammonia oxidized and the amount of carbon assimilated ran strictly parallel. For every 96 mgm of nitrous acid formed it assimilated only 1 mgm of carbon. This disproportion between the rapid oxidizing and the slow assimilating action of the organism explains why its growth is so slow.

Winogradsky and Omeliansky (1899) showed that *Nitrosomonas europaea* was very susceptible to the presence in the medium of organic nitrogenous substances such as peptone or asparagin. In fact the more nutritious a medium was for ordinary bacteria, the less suitable was it for the nitrite organism. It is possible however, for it to grow in the presence of organic matter (Boullanger and Massol 1904, Fremlin 1914, Bonazzi 1919), but in artificial culture the results are not satisfactory. On the ordinary laboratory media, for example, growth is scanty or entirely absent. In the soil it is probable that its susceptibility to the presence of organic matter is less.

For the study of single colonies the best medium is a silicic acid gel poured into plates (Winogradsky 1891). After 3 or 4 weeks' incubation small, compact, sharply-defined colonies appear, of a brownish colour. Growth can be hastened by pouring a solution of ammonia over the plate.

*Nitrosomonas europaea* is most active at a temperature of  $25-30^{\circ}\text{C}$ , in a well aerated medium contained in large flat bottomed flasks, which are slowly agitated. The presence of scoria (cellular lava) in the medium is beneficial, apparently by increasing the surface exposed to the air (Boullanger and Massol 1903). Under such conditions in an inorganic solution it may oxidize as much as 169 mgm of ammonia nitrogen to nitrite per litre of medium in 14 days (Bonazzi 1919). The accumulation of nitrite arrests the reaction. The type species is known as *Nitrosomonas europaea*, Winogradsky. Another similar organism, but of spherical shape, is called *Nitrosococcus americanus*, it is found in the New World.

*Nitrosomonas* is not by any means the only organism capable of forming nitrite from ammonia. Cutler and Mukerji (1931) isolated a number of organisms from soil that were able to perform this oxidation though not so actively as *Nitrosomonas*. A full description of these organisms has not yet been published, but most of them were Gram positive, non sporing, non motile rods,  $1.4-1.9 \mu$  long and  $0.70-0.85 \mu$  broad, which grew on ordinary agar, which oxidized various ammonium salts to nitrite, both in culture medium and in soil, which were unable to oxidize nitrite to nitrate, and which failed to grow in the absence of oxygen. Some strains liquefied gelatin and some did not. Nitrite formation was stimulated by the presence of 0.1 per cent sucrose, but no growth occurred in pure sugar solutions. Unlike *Nitrosomonas*, which requires a distinctly alkaline medium, these organisms were able



to form nitrite within a range of pH 4.8-7.3. Similar organisms have been described by Cutler and Crump (1933) who found that no fewer than 104 out of 229 strains of bacteria isolated from beet sugar effluent were able to produce nitrite from ammonium salts. Fremlin (1903, 1914, 1929-30) has worked for a long time on a very active nitroso-bacterium that grows in association with other organisms. S. Winogradsky and H. Winogradsky (1933) have described two further genera of nitrifying organisms—*Nitrosocystis* and *Nitrospira*. Their article should be consulted not only for an account of these organisms but also for much useful information on nitrifying bacteria in general.

## NITROBACTER

### DEFINITION—*Nitrobacter*

Cells rod-shaped, non motile, not growing readily on organic media or in the presence of ammonia. Cells capable of securing growth energy by the oxidation of nitrites to nitrates.

Type species *Nitrobacter winogradskyi*

*Nitrobacter winogradskyi* was isolated by Winogradsky in 1891—the year after his discovery of *Nitrosomonas europaea*.

It is a small rod shaped or pyriform organism, sometimes with one end drawn out or recurved. Size  $0.5 \mu \times 0.25-0.3 \mu$ . Non motile. Arranged in more or less dense masses.

Winogradsky (1896) cultivated it in a medium of the following composition:

NaNO <sub>2</sub>	1.0 gm
Pot. phosphate	0.5 "
MgSO <sub>4</sub>	0.3 "
Na <sub>2</sub> CO <sub>3</sub> (anhydrous)	0.5 "
NaClO <sub>4</sub>	0.5 "
Re-distilled water	1000 "

The medium is placed in a shallow layer in wide-bottomed flasks. Growth occurs in the form of a just perceptible gelatinous film at the bottom of the flask, no turbidity appears. The nitrite is oxidized to nitrate. No organic nitrogen nor carbon is required. Indeed the presence of organic matter hinders its development, though not so markedly as that of the nitrite organism. On a washed agar medium made up with sodium nitrite, sodium carbonate, and potassium phosphate (Omehansky 1899) single colonies are formed, light brown in colour and of irregular shape. Similar colonies, but smaller and more compact, appear on silicic acid gel plates. No growth occurs in broth.

In artificial culture the nitrate bacillus is very susceptible to the presence of ammonia, in soil it is less so. The accumulation of nitrate to the extent of 25 gm. per litre arrests the reaction (Boullanger and Massol 1903).

As well as the genera that we have described, there are numerous other organisms playing an important part in soil metabolism. We shall confine ourselves here to giving a definition of the genera *Hydrogenomonas*, *Methanomonas*, *Carboxydomonas*, and *Acetobacter*.

DEFINITION — *Hydrogenomonas*

Monotrichate short rod, capable of growing in the absence of organic matter and securing growth energy by the oxidation of hydrogen (forming water) Kaserer (1906) who first described the organism states that his species will also grow well on a variety of organic substances

Type species is *Hydrogenomonas pantotropha* Niklewski (1908) described two additional species *Hydrogenomonas* and *Hydrogenomonas*

DEFINITION — *Methanomonas*

Monotrichate short rods capable of growing in the absence of organic matter and securing growth energy by the oxidation of methane (forming carbon dioxide and water)

Type species is *Methanomonas methanica* (Söhngen 1906)

DEFINITION — *Carboxydomonas*

Rod-shaped cells capable of securing growth energy by the oxidation of carbon monoxide (forming carbon dioxide)

The type species *Carboxydomonas degarabopula* (Bejerinck and van Delden 1903) is described as non motile

DEFINITION — *Acetobacter*

Cells rod-shaped frequently in chains non motile Cells grow usually on the surface of alcoholic solutions as obligate aerobes securing growth energy by the oxidation of alcohol to acetic acid Also capable of utilizing certain other carbonaceous compounds as sugar and acetic acid Elongated filamentous club shaped swollen and even branched cells may occur as involutary forms

Type species is *Acetobacter aceti* (Thomson 1857)

## REFERENCES

- BEJERINCK, M W (1883) *Bot Ztg* 46 744 740 756 780 796 (1901) *Zbl Bakt II* 10 561  
 BEJERINCK, M W and DELDEN, A VAN (1903) *Zbl Bakt* 9 3 (1903) *Ibid* 10 33  
 BEWLEY, W F and HUTCHINSON, H B (1900) *J agric S* 10 144  
 BONAZZI, A (1919) *J Bact* 4 43  
 BOULLANGER, E and MASSOL, L (1903) *Ann Inst Pasteur* 17 49 (1904) *Ibid* 18 181  
 BUSHNELL, O A and SABLES, W B (1939) *J Bact* 33 401  
 CUTLER, D W and CRUMP, I W (1933) *Ann appl Biol* 20 791  
 CUTLER, D W and MCKENZIE, B K (1931) *Proc roy Soc B* 108 384  
 FRED, E B and DAVENPORT, A (1918) *J agric Res* 14 317  
 FREMLIN, H S (1898) *J Path. Bact* 5 389 (1903) *J Hyg Camb* 3 364 (1914) *Ibid* 14 149 (1929-30) *Ibid* 29 736  
 FULMER, H L and FRED, F B (1917) *J Bact* 2, 473  
 GAINES, P I (1918) *J agric Res* 14, 765  
 HOFER, A W (1914) *J Bact* 48 697  
 KASERER, H (1906) *Zbl Bakt II* 10 681  
 KLECZKOWSKI, A and TORMON, H G (1914) *J Bact* 48 661  
 KLIMMER, M and KRÜGER, R (1914) *Zbl Bakt II* 40 256  
 KRAMER, F (1911) *Zbl Bakt* 87 401  
 LEWIS, I M (1938) *J Bact* 35 573  
 NIKLEWSKI, B (1908) *Zbl Bakt II* 10 469  
 OMELIANSKY, V (1899) *Zbl Bakt II* 10 537  
 OMELIANSKY, W L and SZEWEROWA, O P (1911) *Zbl Bakt II* 10 29 643  
 RUSSELL, E J (1923) *The Microorganisms of the Soil* London  
 SMITH, N R (1935) *J Bact* 30 33  
 SÖHNGEN, N L (1906) *Zbl Bakt II* 10 513  
 THOMSON, R D (1857) *Leb gs Ann* 83 89  
 WINOGRADSKY, S (1890a) *Ann Inst Pasteur* 4 713 (1890b) *Ibid* 4 257 (1890c) *Ibid* 4 760 (1891) *Ibid* 5 97 577 (1896) *Zbl Bakt II* 10 329 377 429  
 WINOGRADSKY, S and O ELIANSKY, W (1899) *Zbl Bakt II* 10 379 377 479  
 WINOGRADSKY, S and WINOGRADSKY, H (1933) *Ann Inst. Pasteur* 50 350

## CHAPTER 21

### PSEUDOMONAS

#### DEFINITION — *Pseudomonas*

Rod-shaped organisms, usually motile by means of polar flagella. Generally Gram negative. Non-sporing. Aerobic, some species are facultative anaerobes. Frequently produce a water-soluble pigment, which is yellow, green, blue, purple, or brown in colour, and which diffuses through the medium. Some species form a non-diffusible yellow pigment, and some species are photogenic. Fermentation of carbohydrates as a rule not active. Frequently gelatin-liquefiers, and active ammonifiers. Common in soil and water. Many yellow species are plant parasites.

Type species *Pseudomonas pyocyanea*. (On grounds of priority the American Committee recommend that this organism should be called *Ps. aeruginosa*.)

*Ps. pyocyanea* was first isolated by Gessard in 1882 from 'blue pus.' *Ps. fluorescens* was described originally by Flüge (1896) under the name of *Bacillus fluorescens liquefaciens*. This organism appears to be closely related to *Ps. pyocyanea* the possible differences between them will be discussed later.



FIG 98 — *Pseudomonas fluorescens*

From an agar culture 24 hours, 37° C. (× 1000)

**Morphology** — The organisms of this group are rod shaped and rather slender. Their length is subject to considerable variation, even in a single strain some organisms may be very short, while others are long or actually filamentous. The sides are parallel and the ends rounded. They are arranged singly, in small bundles, or in short chains. They are motile by one or more polar flagella, they are non-sporing, they stain readily with the ordinary aniline dyes and are usually Gram negative. They are non-acid fast.

**Cultural Appearances** — Growth occurs readily on the usual media. Many species form a water-soluble pigment which diffuses through

the medium. On potato *Ps. pyocyanea* and *Ps. fluorescens* give a pigmented growth, which frequently assumes a café-au-lait colour, not unlike that given by organisms of the *Brucella* and *Pfeifferella* groups, and *V. cholera*.

**Resistance**—None of the members of the group forms spores and none is particularly resistant to heat or chemical disinfectants. They all succumb on exposure in a water bath to a temperature of 55° C in 1 hour. The greenish yellow fluorescent bacilli—*Ps. fluorescens* and *Ps. pyocyanea*—are said to be more resistant than other vegetative organisms to ultra violet light (Burge and Neill 1915). It is suggested that they are able to convert the short wave lengths into longer waves and hence dispose of the energy of the absorbed waves which would otherwise be spent in coagulating them (see Chapter 5).



FIG 93.—*Pseudomonas pyocyanea*  
Surface colony on agar 24 hours  
37° C ( $\times 8$ )

**Metabolic and Biochemical Characters**—Many of the organisms are obligatory aerobes others may give a very slight growth under anaerobic conditions. Growth of *Ps. pyocyanea* in broth is accompanied by a fall in oxidation reduction potential which reaches a limiting value of between Eh -0.100 and -0.200 volt. The pigment *pyocyanin* constitutes a reversible oxidation reduction system and



FIG 100.—*Pseudomonas fluorescens*  
Agar slope culture 48  
hours 22° C

acts as a respiratory catalyst but according to Reed and Boyd (1933) the changes of potential in culture are not dependent on the presence of this substance. The limiting temperatures for growth are about 0° C and 42° C. Generally speaking *Ps. pyocyanea* grows between 5° C and 42° C and has an optimum about 37° C whereas *Ps. fluorescens* grows between 0° C and 37° C and has an optimum about 25° C. The fermentative power is usually weak, acid but no gas is formed by some species in certain sugars. According to Moltke (1927) 4 strains of *Ps. pyocyanea* which he examined failed to ferment any of the usual sugars. In our experience acid is generally formed from glucose but from no other sugar. This is confirmed by Sandiford (1937) who examined 60 strains and found that glucose was the only sugar fermented. The formation of indole has been recorded by various observers but as Sandiford points out a false reaction may result from the action of the acid in Böhm's reagent on the pigment produced by the organisms. If the oxalic acid method is used for testing no indole formation can be demonstrated. Practically all strains liquefy gelatin and peptonize milk. The usual nitrate reduction test at 5 days is negative with most strains but this is often because the nitrate itself is reduced resulting in the production of gaseous nitrogen or one of its compounds. If the test is carried out after one day nitrites can be demonstrated (Ferramola and Monteverde 1939). All strains are said by Ferramola and Monteverde to utilize citrate as the sole source of carbon but to be incapable of producing  $H_2S$ . Ammonia is produced by all strains (Seleen and Stark 1943).

**Pathogenicity**—With the exception of *Ps. pyocyanea*, members of the group

are non pathogenic to man. *Ps. pyocyanea* itself gives rise occasionally to suppurative processes and less often to generalized infection. Among the commonest manifestations are middle ear suppuration in children, destructive lesions of the skin sometimes described as ecthyma gangrenosum, in children and adults, and necrotic and ulcerative lesions of the alimentary mucosa. The respiratory tract, the eye, the joints and the kidneys are sometimes affected. Wounds are often infected. There is also reason to believe that the organism plays a part in some cases of infantile diarrhoea. Infection may be primary or secondary, and is often acute and rapidly fatal. (For review of human infections see Waite 1908, Fraenkel 1917, and Lode 1929.)

Animals are rarely infected unless given large doses intravenously, when they may die of intoxication. *Ps. pyocyanea* however often produces fever and a local abscess after subcutaneous injection into rabbits, and, if highly virulent, it may prove fatal in 24 hours.

**Group forming Bluish-green Pigment**—*Ps. pyocyanea*, first isolated in pure culture by Gessard (1882), is widely distributed in nature, being found in water, sewage and sometimes on the normal skin, particularly of the axilla and perineum (Růžicka 1898). It is not infrequently found in wounds where it gives rise to blue pus. It may invade the nasal fossæ, the middle ear, the meninges, the bronchi and other organs and set up suppuration. In infants and young children it causes intestinal disturbances and diarrhoea, sometimes it enters the blood stream, and gives rise to a general infection (Williams and Cameron 1896). According to Pons (1927), *Ps. pyocyanea* is especially pathogenic in the tropics where it is not infrequently responsible for typhoid like infections and abscesses of the liver.

Injected subcutaneously or intravenously into guinea pigs or rabbits in a dose of 0.5–1.0 ml. of a 24 hours' broth culture it may cause death in 24 to 48 hours, post mortem there is a hæmorrhagic œdema at the site of injection (after subcutaneous inoculation), small punctate hæmorrhages are seen in the stomach and intestine, and sometimes nephritis. The bacilli can be recovered from the blood, viscera, and urine. As much the same appearances result from the injection of dead cultures it is probable that an endotoxin is responsible. Different strains vary in virulence, some do not kill for weeks others not at all.

Little is known of the antigenic structure of this organism. Boivin and Mesrobianu (1937) using their trichloroacetic acid technique, isolated a glyco-lipoid complex or endotoxin containing a polysaccharide hapten. By inoculation of rabbits with the whole complex they obtained antisera which precipitated the homologous endotoxin and the polysaccharide hapten, and which agglutinated the homologous bacteria. The endotoxin, as a whole, killed mice, but the polysaccharide hapten was non toxic on intraperitoneal inoculation, even in doses of 5 mgm. The authors conclude that the identity of the endotoxin with the somatic antigen of *Ps. pyocyanea* is similar to that in members of the *Salmonella* group.

In culture *Ps. pyocyanea* forms a bluish green pigment. Gessard (1890, 1891, 1892) found that this pigment consists of two different substances. One, pyocyanin is bluish green, non fluorescent, is formed in peptone media and is soluble in both chloroform and water. The other, fluorescin, is greenish yellow, fluorescent, is formed only in the presence of a phosphate, and is insoluble in chloroform, but soluble in water. By cultivation in different media he was able to obtain varieties

that produced pyocyanin or fluorescin only, and some that were completely achromogenic. Fordos (1860) obtained pyocyanin in long blue crystals from a solution in chloroform. Wasserzug (1887) showed that its formation was prevented by several substances, such as 5 per cent  $\text{KNO}_3$ , 8 per cent  $\text{KClO}_3$ , 0.5 per cent ammonium tartrate, 5 per cent  $\text{NaCl}$ , and by many disinfectants not strong enough to inhibit the growth of the organism. Jordan (1899) studied 7 strains of *Ps. pyocyanea*, 1 strain produced pyocyanin only, 5 both pyocyanin and fluorescin, and 1 fluorescin only. He found that the fluorescent pigment required for its formation both phosphate and sulphate, while neither of these substances was necessary for the production of pyocyanin. Both pigments are formed in suitable synthetic media. In old cultures a black pigment sometimes appears, this appears to be an oxidation product of pyocyanin. A yellowish brown pigment, which may also be found in cultures, appears to be an oxidation product of the fluorescent substance. Boland (1899), who worked with a solution of pyocyanin in chloroform, found that it became yellow if exposed to sunlight. Apparently the chloroform was broken up, and chlorine set free, which oxidized the pyocyanin to pyoxanthose. He showed that pyocyanin was largely dissolved by  $\text{HCl}$ , which turned it red, and pyoxanthose by 33 per cent  $\text{H}_2\text{SO}_4$ , which turned it reddish yellow. Turfitt (1936) found that the production of both pigments was favoured by 1 per cent glycerol of pyocyanin by 5 per cent glycerol, and of fluorescin by asparagin.

More recently Wrede (1930) has determined the constitution of pyocyanin, and shown that it can be synthesized by the organism from lactic acid and salts. Chemically he regards it as an entirely new type of dye, containing two pentavalent nitrogen atoms, but this requires confirmation (Michaelis 1935). It affords, moreover, the first instance in which phenazine derivatives have been found in nature. Its empirical formula is  $\text{C}_{22}\text{H}_{20}\text{N}_4\text{O}_2$ . Wrede states that it dissolves poorly in cold, but readily in warm, water, as well as in chloroform, nitrobenzol, pyridine, and phenol. It is fairly resistant to acids and forms with them red-coloured salts. To alkali, on the other hand, in the presence of oxygen, it is much less resistant and is rapidly broken down.

Turfitt (1937) obtained a preparation of fluorescin by growth of *Pseudomonas* in synthetic liquid media and adsorption of the pigment on to "suma-carb," followed by electro-dialysis. The final product, which had an ash value of less than 0.4 per cent, was an amorphous greenish brown powder, readily soluble in water, phenol, and acetic acid, but not in other organic solvents. A dilute aqueous alkaline solution showed a green fluorescence, becoming colourless and non-fluorescent on acidification. More concentrated alkaline solutions had a red colour and exhibited an intense green fluorescence. In alkaline solution the pigment showed a well-defined absorption band with its maximum at  $410\mu$ , in acid solution the band was less evident and was nearer the shorter wave-lengths. The empirical formula was found to be  $\text{C}_9\text{H}_7\text{O}_2\text{N}$ .

Summarizing, we may say that *Ps. pyocyanea* forms two pigments: (1) Pyocyanin, a bluish green pigment, soluble in both chloroform and water, from which it can be obtained in long blue crystals. For its production neither phosphate nor sulphate is required. (2) Fluorescin, a greenish yellow fluorescent pigment, soluble in water but not in chloroform. For its production both phosphate and sulphate are required. In old cultures it may be oxidized to a yellowish-brown pigment. *Ps. pyocyanea* forms pyocyanin and fluorescin, *Ps. fluorescens* forms only fluorescin. Both pigments are themselves oxidation products of colourless

substances. (For a careful study of the factors controlling the production of pyocyanin and other pigments, see Sullivan 1905-06)

From Hadley's observations (1927) it appears fairly certain that the power of forming pyocyanin is subject to discontinuous variation. Under artificial conditions of cultivation, many strains of *Ps. pyocyanea* tend to lose their ability to produce a bluish green coloured growth on agar. If such strains are inoculated into broth, and plated out, it will be found that only a proportion of the colonies are coloured bluish green, the remainder have merely the yellowish colour due to the presence of the fluorescent pigment. With many strains prolonged subcultivation is followed by the complete disappearance of the bluish green variants, and their replacement by the yellowish variants. Once a strain has lost its power of producing pyocyanin, it is unable to recover it. According to Cattell (1935), cultivation in broth containing 2 per cent zinc oxide or 1 per cent. zinc sulphate results after a few passages in suppression of pigment formation. The organisms, however, do not undergo dissociation, and when transferred back to normal media again give rise to pigment.

Emmerich and Low (1899) and Emmerich, Löw and Korschun (1902) found that old cultures of *Ps. pyocyanea* were highly bactericidal to many organisms. They ascribed this action to an enzyme "pyocyanase". More recently Schoental (1941) has brought evidence to show that the antibacterial and lytic action is due not to enzymes but to pigments. Of these, the most active is a yellow pigment,  $\alpha$ -oxyphenazine, which can be isolated in crystalline form from old cultures. It has an antibacterial action similar to that of the flavines and is less toxic to tissue cells than pyocyanin. In a 1/20,000 concentration in serum broth it inhibits growth of Gram positive cocci for 24 hours, and in a 1/10,000 concentration organisms of the *Neisseria*, *Corynebacterium*, *Proteus*, *Bacterium*, *Brucella* and *Clostridium* groups. An oily substance, which she isolated from cultures of *Ps. pyocyanea* 2 to 3 months old, proved very active against *V. cholera*, causing lysis in a 1/10,000 dilution.

An interesting feature of *Ps. pyocyanea* is its ability both in culture and in the animal body to form hydrocyanic acid (Patty 1921).

We append a detailed description of this organism.

### *Pseudomonas pyocyanea*

*Synonyms*—*Bacterium aeruginosum* (Schroter) Migula. *B. pyocyaneus*, Gessard.

*Habitat*—Intestinal canal, water, sewage, pus, sinuses, human skin, sometimes pathogenic to man.

*Morphology*—Rods, 1.5-3.0  $\mu \times$  0.5  $\mu$ , axis straight, ends rounded, sides parallel, arranged singly, or in pairs and short chains, motile by 1-3 flagella at one pole. Non sporing, non-capsulated. Gram negative. Non acid fast.

*Agar Plate*—2 days at 25° C. Round colonies, 1-2 mm in diameter, low convex, with smooth, moist, glistening surface, edge entire or undulate, structure amorphous; butyrous consistency, easily emulsifiable, fluorescent yellowish green colour, translucent. After 5 days it is differentiated into a smooth, convex, translucent centre and a radially striated, effuse, transparent periphery, with an undulate, lobate, or villous edge. Medium coloured green.

*Agar Slope*—2 days at 25° C. Good growth raised slightly, with beaten-copper surface, irregular edge, and sometimes clear phage-like areas, greenish yellow, translucent. Medium green. 5 days growth becomes effuse and scarcely visible.

*Gelatin Stab*—Moderate filiform growth to bottom of stab, slow crateriform liquefaction. After 14 days the upper 1-3 cm. are digested in stratiform manner, and

the fluid is turbid, yellowish green, and sometimes granular, there may be saccate liquefaction around the filiform growth as well

**Broth**—2 days at 25° C Abundant growth, with dense turbidity, yellowish green colour, thick white ring growth and thin surface pellicle, slight powdery sediment, disintegrating on shaking After 5 days there is an abundant, visco-floccular deposit, only partly disintegrating Mawkish odour, like trimethylamine

**Blood Serum**—7 days at 25° C Good, confluent, slightly raised growth, of greenish-yellow colour, medium is slightly green, medium is partly digested in 14 days

**Potato**—6 days at 25° C Abundant, slightly raised, confluent, greenish brown growth with moist, glistening contoured surface Potato coloured green Later, both the growth and the potato take on a brownish colour

**Resistance**—Destroyed by 55° C in 1 hour

**Metabolic**—Aerobic, no growth anaerobically Opt temp 30–37° C, limits 5–42° C Forms a green pigment soluble in chloroform and in water, called pyocyanin, forms a greenish yellow fluorescent pigment, soluble in water but not in chloroform Nutritional grows well on ordinary media, in synthetic media both phosphate and sulphate are essential for production of fluorescent pigment

**Biochemical**—Acid, no gas, in glucose Indole —, a false reaction may be given by Bohme's reagent, the acid of which turns pyocyanin red MR — VP — Nitrate reduction — H<sub>2</sub>S — NH<sub>3</sub> production + Catalase + MB reduced Starch diastase — LM peptonization and decolorization complete in 5 days at 30° C, may be slight preliminary clot, milk often turned green Growth in citrate

**Pathogenicity**—Low pathogenicity to man giving rise to diarrhoea and general infections in infants and to suppuration Cause of blue pus Gives rise to fever and local abscess after subcutaneous injection into rabbits

***Ps fluorescens***—This organism is found in water, hail (Bell 1902), sewage, and has been isolated from lemonade (Thom 1911) It is motile by one or more polar flagella Many authors consider it a variety of *Ps pyocyanea*, which has become adapted to a purely saprophytic existence (Tanner 1918, Růžicka 1898) The differential characters may be given as follows

<i>Ps pyocyanea</i>	<i>Ps fluorescens</i>
(1) Opt temp 37° C, grows at 42° C	Opt temp 25° C, no growth at 42° C
(2) Pyocyanin and fluorescent pigment formed	Fluorescent pigment only formed
(3) Liquefaction in gelatin stratiform and saccate	Liquefaction in gelatin not always present when present stratiform only
(4) Pathogenic to rabbits and guinea pigs	Non pathogenic to rabbits and guinea pigs

These differences are by no means constant Further, since varieties of *Ps pyocyanea* may occur that fail to produce pyocyanin, and since achromogenic varieties of both organisms are not uncommon, differentiation of the two organisms is often impossible

Jordan (1899) studied 58 strains of *Ps fluorescens* from water He found that 33 of them liquefied gelatin, and produced acid, clot, and peptonization in milk, 25 did not liquefy gelatin, and produced alkali in milk without coagulation The variations in reaction that may occur can be judged from the fact that Tanner divided 42 strains, which he studied, into no fewer than 27 different groups

*Ps fluorescens* is generally non pathogenic to animals, but it may give rise to a local abscess in rabbits and guinea pigs On the other hand, it is stated to be frequently pathogenic to plants, especially cultivated vegetables, such as carrots, cauliflowers and tomatoes, in which it causes areas of moist necrosis (Griffon 1909)



*Ps. cyanogena* is a motile bacillus possessing polar flagella. It forms two pigments one fluorescent the other varying in colour from blue to brown or black. In milk with an acid reaction it gives rise to a bright blue colour. It is the cause of epidemics of blue milk.

*Pseudomonas denitrificans*—This organism was originally described by Christensen (1903-04) under the name of *Bacillus denitrificans fluorescens*. Two varieties were recognized A and B. variety A was isolated from garden earth and was able to reduce nitrates to gaseous nitrogen. variety B was isolated from horse dung and was able to reduce nitrites but not nitrates to gaseous nitrogen.

*Variety A* is a small bacillus  $0.5-1.25 \mu \times 0.5-0.7 \mu$ . It is surrounded by a large capsule, measuring  $2-5 \mu$  in diameter. Appears to be slightly motile. Often shows bipolar staining. Is Gram negative. Grows freely at  $25^{\circ}\text{C}$ . Colonies on agar are 2-3 mm in diameter after 3 days, and are circular with an entire edge. They have an opalescent sheen. In an agar stroke culture a whitish glistening growth is formed and the agar is coloured bright green. In gelatin stab there is a filiform growth and a whitish surface growth with a lobate edge. The gelatin is not liquefied. In a gelatin stroke culture there is a dirty white layer of growth which fluoresces brilliantly in transmitted light. The gelatin is coloured bright-green. In broth there is a dense turbidity and a very thick wrinkled surface pellicle which climbs up the walls of the tube. In 0.2 per cent. nitrate broth a dense turbidity is produced and a foam due to the liberation of gaseous nitrogen, is seen reaching its maximum in 40 hours. The nitrate is not completely destroyed, even in 3 weeks.

*Variety B* is a larger bacillus  $1-3 \mu$  long by  $0.5-1.25 \mu$  broad, and is surrounded by a capsule. Motility doubtful. Gram negative. It is unable to reduce nitrates to nitrites, but is able to reduce nitrites to gaseous nitrogen. Colonies on agar are 2.5-3.5 mm. in diameter after 2 to 3 days and are flat, whitish, and so fluid that they may flow over the agar if the plate is stood on edge. In agar stroke culture there is a thinish filiform grey growth, having an effuse indescendent peripheral extension. After 10 days the agar is slightly coffee-coloured. In gelatin stroke culture there is a greyish or slightly brownish layer of growth, which fluoresces strongly in transmitted light. The gelatin is coloured brown. In broth there is a dense turbidity and a thin indescendent surface pellicle.

*Pseudomonas caviae*—This name was suggested by Scherago (1937) for a capsulated organism that he isolated from the blood and viscera of young guinea pigs dying from a rapid bacteræmic infection. Its general properties are as follows. Gram negative rods,  $1-5 \mu$  long by  $0.6-0.75 \mu$  broad occurring singly and in pairs end-to-end. Actively motile by 1 to 3 polar flagella. Encapsulated when isolated from the animal body. Surface colonies on agar are 1-3 mm. in diameter, convex, smooth, indescendent and translucent, with a finely granular structure. In agar slope cultures the medium becomes greenish yellow in a week, and later turns brownish yellow. Uniform turbidity in broth with a surface ring and pellicle and a heavy granular deposit, light yellow in colour. Infundibuliform liquefaction of gelatin, complete in 2 days. Scanty glistening light yellowish orange growth on potato. Aerobic, facultatively anaerobic. Grows well at  $22^{\circ}\text{C}$  and  $37^{\circ}\text{C}$ . Acid in dextrose, maltose, mannitol, lactose, sucrose and salicin. Litmus milk acidified, coagulated, and partly peptonized. Indole and ammonia formed but not  $\text{H}_2\text{S}$  or catalase. Nitrates reduced to nitrites. M.P.  $\pm$  V.P.  $-$  M.B. reduced. Reproduces the natural disease when inoculated parenterally into young guinea-pigs. Produces rapidly fatal septicæmia in mice inoculated intraperitoneally. Apparently non pathogenic to rabbits.

## REFERENCES

- BELLI, C M (1902) *Zbl Bakt.*, IIte Abt., 8, 445  
 BOVIN, A and MESROBEANU, L. (1937) *C R Soc Biol*, 125, 273  
 BOLAND, G W (1899) *Zbl Bakt*, 25, 897  
 BURGE, W E and NEILL, A J (1915) *Amer J Physiol*, 33, 399  
 CATALIOTTI, F (1935) *G Batt Immun.*, 15, 161  
 CHRISTENSEN, H. R. (1903-4) *Zbl Bakt.*, IIte Abt., 11, 190  
 EMMERICH, R and LÖW, O (1899) *Z Hyg InfektKr* 31, 1  
 EMMERICH, R., LÖW, O., and KORSCHUN, A (1902) *Zbl Bakt* 31, 1  
 FERRANOLA, R and MONTEVERDE J J (1939) *Bol Obras san Nacion Buenos Aires* 3, 272  
 FLÜGGE C G F W (1896) 'Die Mikroorganismen,' 3te Auflage 2te Theil p 292 Leipzig  
 FORDOS, M J (1860) *C R Acad Sci*, 51, 215 396  
 FRAENKEL, E. (1917) *Z Hyg InfektKr*, 84, 369  
 GESSARD, C (1852) *C R Acad Sci*, 94, 563, (1890) *Ann Inst Pasteur* 4, 88 (1891) *Ibid*, 5, 65, (1892) *Ibid*, 6, 801  
 GRIFFON, E (1909) *C R Acad Sci* 149, 50  
 HADLEY, P (1927) *J infect Dis*, 40, 74  
 JORDAN, E O (1899) *J exp Med*, 4, 627  
 LÖDE, A (1929) *Kollo & Wassermann Handb path Mikroorg* 6, 149 Gustav Fischer Jena  
 MICHAELIS L. (1935) *Chem Rev* 16, 243  
 MOLTKE O (1927) 'B proteus vulgaris' Copenhagen  
 PATTY, F A (1921) *J infect Dis*, 29, 73  
 PONS, R (1927) *Ann Inst Pasteur*, 41, 1338  
 REED, E M and BOYD, G B (1933) *Canad J Res*, 8, 173  
 RČIČKA, S (1898) *Zbl Bakt*, 24, 11  
 SANDIFORD, B R (1937) *J Path Bact*, 44, 567  
 SCHERAGO, M. (1937) *J infect Dis* 60, 245  
 SCHOENTAL, R (1941) *Brit J exp Path*, 22, 137  
 SELEEV, W A. and STARR, C N (1943) *J Bact*, 46, 491  
 SULLIVAN, M A (1905-6) *J med Res*, 14, 109  
 TANNER, F W (1918) *J Bact*, 3, 63  
 THÖNI, J (1911) *Zbl Bakt* IIte Abt 29, 616  
 TURPITT G E (1936) *Biochem J*, 30, 1323, (1937) *Ibid*, 31, 212  
 WAITE, H H (1908) *J infect Dis*, 5, 542  
 WASSERZUG E (1887) *Ann Inst Pasteur*, 1, 581  
 WILLIAMS E P and CAMERON, K (1896) *J Path Bact.*, 3, 344  
 WREDE, F (1930) *Z Hyg InfektKr*, 111, 90

## CHAPTER 22

### VIBRIO AND SPIRILLUM

#### VIBRIO

##### DEFINITION—*Vibrio*

Short curved rigid rods, arranged singly or united into  $\alpha$  forms or spirals. Motile by a single polar flagellum which is usually relatively short. (Some species may have two or three polar flagella.) Non sporing. Usually Gram negative. Aerobic and facultatively anaerobic. Many species liquefy gelatin and are active ammonifiers. Commonly found in water. Most species are saprophytic, a few are pathogenic to man.

Type species. *Vibrio cholerae*

The first member of this group to be described was *V. cholerae* which was found by Koch (1886) in the dejects of cholera patients. In 1888 Gamaleia (1858-1908) isolated a vibrio from the blood and intestinal contents of chickens dying from a cholera like disease at Odessa. To this organism he gave the name of *V. metchnikovi*. During the next 10 years a large number of other vibrios more or less resembling the cholera vibrio were isolated from different sources such as well river and sea water, the faeces of man and animals, cheese and intestinal abscesses of pigs (Dunbar 1893, Smith 1894, Dieudonne 1894, Kutscher 1895, Gotchlich 1895, 1906, Ruffer 1907, Crendiropoulo 1912, Craster 1913). The differentiation of many of these organisms from *V. cholerae* proved impossible until the introduction of the Pfeiffer test in 1894 (see Chapter 63). Even with the help of this test it was not always possible to decide whether they were different species or were merely variants of the main species. Since the chief interest of the vibrios—at least to the medical bacteriologist—has been their relationship to *V. cholerae* it follows that a careful systematic study of the saprophytic species has not yet been made. For this reason it is premature to attempt a classification of the members of this group.

**Morphology and Staining**—The vibrios are short curved rods looking like commas. In size they vary considerably from about 1 to 5  $\mu$  in length and about 0.3 to 0.6  $\mu$  in breadth. The commas may appear long thin and delicate or short, stunted and thick. They are arranged singly in  $\alpha$  shaped or occasionally semicircular pairs or in short chains. In fluid media spirals are often formed and in old cultures a variety of forms may be seen. Most of these are very small looking like granules and staining poorly but there are larger swollen shadow forms resembling bottles or clubs. When freshly isolated the resemblance to a comma is most striking but after long subculture in the laboratory the vibrios frequently lose their curved shape and are then not unlike coliform bacilli. The organisms are

very actively motile by a single polar flagellum. They stain best with dilute carbol fuchsin. They are Gram negative.

**Growth Requirements**—Growth occurs readily on the usual media. One of the most characteristic properties is the rapidity of growth in peptone water (1 per cent peptone, 0.5 per cent NaCl). Multiplication occurs chiefly at the surface, where, after 6 to 9 hours, a delicate membrane is formed. There is very little turbidity as a rule; the deposit that forms appears to be derived from the surface pellicle.

The vibrios are markedly aerobic; they grow best in the presence of abundant oxygen. Under strictly anaerobic conditions some of the members fail to grow altogether, the majority give rise to a very slight growth on agar or in broth in about a week. The optimum temperature is 30–40° C, no growth occurs macroscopically under 16° C.

For growth and survival a H ion concentration of pH 7.6–8.0 is most suitable. The organisms have a high alkali but a very low acid tolerance. Cultures containing a fermentable sugar are sterile in a day or two (Nobechi 1925).

A number of selective media have been devised for facilitating the isolation of *V. cholerae* from the faeces.



FIG 109—*Vibrio cholerae*  
Surface colonies on agar 24 hours  
37° C (× 8)



FIG 101—*Vibrio cholerae*  
From an agar culture 24 hours 37° C (× 1000)

One of the best known of these—Dieudonné's medium (1909)—is prepared by adding normal KOH solution to an equal quantity of defibrinated ox blood and heating to 100° C for half an hour. Thirty parts of this mixture are added to 70 parts of nutrient agar rendered neutral to litmus. According to Vedder and van Dam (1932) the medium should be allowed a day or two to ripen. During this time CO<sub>2</sub> is taken up from the air and NH<sub>3</sub> is given off. The medium when ready for use should have a pH of 9.0–9.6. At a lower pH coliform and other organisms grow and the medium is no longer selective; at a higher pH the growth of the cholera vibrio itself is inhibited. Ottolenghi's medium (see Bocchius 1911) consists of ox bile to which 3 per cent of a 10 per cent solution of crystalline sodium carbonate has been added. Steri-

lization is effected in the autoclave. Bandia's medium suitable for cultivation of the cholera vibrio from water is a peptone water solution containing dilute anticholera agglutinating serum; the vibrios multiply and fall to the bottom in clumps. Yen (1932–33) recommends a phenolphthalein starch medium for the isolation of the cholera vibrio. It depends on the unusual property possessed by this organism of rapidly fermenting starch in an alkaline solution (see Gordon 1900).

**Cultural Characters.**—On agar the colonies are not distinctive, they may be either clear and amorphous or finely granular. Small knob-like secondary colonies sometimes form in about a week on the surface of the parent colony. An effuse, transparent peripheral extension is not unusual. Crystals may form in the agar. Balteanu (1926) has described three colonial variants in cultures of cholera and cholera-like vibrios. Variant (1) was rugose, (2) had a more opaque centre and a transparent periphery, and (3) was opaque. Variants 1 and 2 reverted to type when subcultured on agar, variant 3 reverted slowly in broth but remained constant for a long time on agar. Variant 3 consisted of



FIG. 103.—*Vibrio cholera*.

Gelatin stab culture 5 days, 22° C. showing infundibuliform liquefaction.

non motile bacilli which had a mucoid envelope, the organisms contained a heat-stable antigen only and were apparently of the pure O form (see antigenic structure). Though the rugose form has been regarded by some workers as an extreme rough form, the observations of White (1935, 1940) show that it is a peculiar variant characterized by the secretion of a diffuse intercellular gelatinous substance or of actual capsules. It is unstable and is constantly tending to revert to the S or R form from which it is derived.

In gelatin stab many species produce liquefaction. On potato some of the members—including *V. cholera*—give a raised growth of case-au-lait colour resembling that of the *Brucella* group. Pfeiffer, Mallet, and Pelyocyanca. On MacConkey's medium growth is often poor, *V. cholera* flourishes well on it, but the non-pathogenic members grow poorly. The colonies are colourless when young but soon assume a pinkish red appearance; the medium changes simultaneously to a darker red. The rate at which the colour alters depends on the organism observed, the colonies of *V. cholera* may remain yellow for a week, those of the Asiatic vibrio are bright red in 3 days.

On Loeffler's serum growth is plentiful, and is sometimes accompanied by slow liquefaction.

**Resistance.**—None of the vibrios forms spores. Their resistance to heat and disinfectants is low and they are easily destroyed by drying (see Chapter 63). They are killed by heat at 55° C. in 15 minutes or less (Kitasato 1889) and by 0.5 per cent. phenol in a few minutes. Dried on cover slips they perish in about 3 hours. Gastric juice containing more than 25 degrees of acidity (degrees equivalent to number of ml. of N/10 NaOH necessary to neutralize 100 ml. of gastric juice) is said to kill cholera vibrios at once, in the absence of free acid the vibrios may survive for over 24 hours (Napier and Gupta 1942).

**Biochemical Characters.**—Acid, without gas, is generally formed within a day or two in glucose, maltose, mannitol and sucrose. Lactose is sometimes fermented after 10 or 14 days, and occasionally salicin. The Asiatic vibrio ferments glucose only—at least in liquid media. Litmus milk may be unchanged, more of it is acidified and sometimes it is clotted. Many species form indole, reduce nitrates to nitrites, and give the cholera red reaction. This reaction is performed

V.P. test (see p 355) According to van Loghem (1930) the El Tor vibrio gives usually a positive V.P. reaction

**Hæmolytic Formation.**—Many members of this group form a hæmolytic acting on sheep, horse, or rabbit cells. This can be demonstrated either on plates or in a broth culture. In general the cholera vibrio does not produce a soluble hæmolytic, while the El Tor and many non-cholera vibrios are able to do so. For diagnostic purposes a standard technique is essential. Much confusion has in the past been due to differences in method of studying hæmolytic. Zimmermann (1932, 1933) working with 5 per cent defibrinated sheep blood broth cultures, obtained conflicting results but when he used a medium made up with peptone, asparagin and ammonium lactate containing 4 per cent. sheep blood, and read his results after 48 hours incubation at 37° C., a clear differentiation between the non hæmolytic cholera and the hæmolytic El Tor and non-cholera vibrios was apparent. Van Loghem (1930) states that sheep or goat's blood should be used, guinea pig and rabbit red cells are too sensitive. He further points out that the cholera vibrio though not forming a true soluble hæmolytic, does digest blood pigment, this property which has been shown by Bernard, Guillemin and Gallut (1937) to be due to a ferment present in cultures but not in the bodies of the cholera vibrio, is responsible for the greenish discoloration around individual colonies on blood agar and for the complete clarification of the medium that occurs on further incubation. The El Tor vibrio digests blood likewise, but in addition it produces a soluble hæmolytic, the extraction of which has been recorded by Bernard, Guillemin and Gallut (1939). The observations of these workers suggest that the cholera vibrio forms a hæmolytic as well as a digesting ferment, but that it is mainly intracellular and, unlike that of the El Tor vibrio, does not diffuse out to any considerable extent into the medium. The identity of these two hæmolysins is suggested by the finding of Vassiliadis (1937) that the injection of non hæmolytic cholera vibrios into animal gives rise to anti hæmolysins for the El Tor vibrio as active as those prepared by injection of the El Tor vibrio itself.

Doorenbos (1936) finds that hæmolytic is very much more active in 8-hour than in 24-hour cultures. Many strains of true cholera vibrios that were non-hæmolytic after 24 hours produced hæmolytic of sheep or goat cells after 8 hours. With so many factors influencing hæmolytic production it would clearly be dangerous to place too much weight on this characteristic as a means of differentiating between the vibrios.

The common routine method for testing hæmolytic activity is to grow the organism in broth for 3 days at 37° C., to add 1 ml. of the culture to 1 ml. of a 5 per cent. suspension of washed goat or sheep red cells, to incubate for 2 hours at 37° C., and to read the results after the tubes have been left in the cold overnight.

**Toxin Production.**—Nicas and Rietsch (1934) injected dogs intravenously with the filtrate of a broth culture of *V. cholera* a week or more old. In their first series of experiments there were vomiting, defecation, and general depression, with recovery in an hour. In their second series there were dyspnoea, vomiting and paresis of the extremities, followed by recovery or death in 12 hours. At necropsy in the fatal cases, ecchymoses were found in the duodenum and larger hæmorrhages in the stomach. Filtrates of young cultures were innocuous.

Pfeiffer (1932) likewise experimented with filtrates. He found that even 4 ml. of a 20-days glycerine broth filtrate injected intraperitoneally into guinea pigs, had no

more than a slight toxic effect. Dead vibrios however are very toxic. Pfeiffer (1892-1895) found that the lethal dose of living vibrios on intraperitoneal injection into guinea pigs was 1.5 mgm. of an 18 hours' agar slope culture. When the vibrios were killed by chloroform or thymol the lethal dose was 3-4.5 mgm., when they were killed by drying it was 6 mgm., and when they were killed by heat at 55° C. for an hour it was 10-20 mgm. While immune serum was able to protect a guinea pig against several fatal doses of living vibrios it possessed no more protective power than normal serum against dead vibrios. From these experiments he concluded that in young cultures of *V. cholerae* there was a specific toxic substance bound to the bacterial bodies, and that the immune substances in the antiserum were not antitoxic but bactericidal in their action.

Von Dungern (1895) working with one highly virulent strain of cholera and another of very low virulence found that the lethal dose of heat killed organisms was the same in each instance. The toxicity of the cultures therefore bore no relation to their virulence.

Manwaring, Boyd, and Olami (1923) perfused the mammalian heart with 2-7 days culture filtrates of *V. cholerae* added in 5-10 per cent. concentration to Locke's solution. Though non-toxic for the conducting and contractile tissues the filtrates had a destructive effect on the capillary endothelium, as was evident from the oedema of the muscle and the hemorrhages that occurred beneath the endocardium and pericardium.

We may conclude that the cholera vibrio does not secrete a true soluble exotoxin but that it contains endotoxins which are liberated on the autolysis of the bacilli in culture or on the active disintegration of the bacilli by the cells of the animal body. The analogy that it presents with the meningococcus—another organism that readily undergoes autolysis—is very close though the cholera vibrio is far more toxic.

Hahn and Hirsch (1929) working with El Tor and other hæmolytic vibrios found that a soluble toxin was produced in peptone water cultures to which small quantities of glucose were added during growth, the reaction of the medium being kept alkaline by similar additions of NaOH. The toxin which passed through a Seitz filter became demonstrable in 6-10 hours and reached its maximum in 1-4 days. Bacterial counts indicated that the increase in toxicity of the culture coincided with the death of the organisms. The heat resistance of the toxin seemed to vary with different batches: sometimes it was destroyed in 2 minutes, at others not for 30 minutes when exposed to a temperature of 100° C. Injected intraperitoneally into guinea pigs in a dose of 0.25 ml. the toxin had a marked effect on the temperature, which often fell to 30° C. within 2-3 hours. The animals became progressively weaker, paralysis developed in their hind legs and they died in 6-10 hours. Post mortem the findings consisted of a large exudate in the peritoneal cavity, fibrinous purulent deposits on the liver and sometimes hyperæmia of the intestine. Injection of horses with increasing doses of toxin led to the appearance in the serum of antibodies capable of neutralizing to some extent the lethal action of the toxin for guinea pigs. It is to be noted that true non-hæmolytic cholera vibrios were almost completely devoid of toxin-producing power under the cultural conditions described (see also Andu and van Niekerk 1929). Takita (1939) found that the El Tor vibrio produced a true thermolabile exotoxin neutralizable by an antiserum according to the law of multiple proportions. Mice inoculated intravenously with 0.01 ml. died within 24 hours. The exotoxin appeared to be different from the hæmolysin.

**Antigenic Structure**—The antigenic structure of the vibrios has of late years received considerable attention. Kabeshima (1918) working with *V. cholera* discovered the occurrence of serological variants. Balternu (1926) found a heat labile H and a heat stable O receptor in the cholera vibrio. Immune serum prepared against organisms heated to 100° C. for 2 hours contained only O agglutinins. This finding was confirmed and extended by Shousha (1931), Abdoosh (1932) and Gol. (1932). These observations and a particularly careful study by Gardner in

Venkatraman (1935) have done much to clarify the confusion resulting from the work carried out before the importance of flagellar and somatic antigens had been appreciated. The analysis is, however, by no means complete, and the scheme reproduced here must be regarded only as a working hypothesis, certain in the future to require considerable modification.

Attention has been concentrated mainly on the cholera and cholera like vibrios, which we refer to for convenience as Group A. This group comprises organisms, most of which produce acid without gas in glucose, maltose, mannitol and sucrose, but not in dulcitol and which give the cholera red reaction. All organisms of Group A possess a common H antigen. The major O antigens on the other hand, of which six have already been differentiated, are much more specific and are used as a basis for the differentiation of Group A into sub-groups. The true cholera vibrios all appear to fall into sub-group I, which also contains most of the El Tor strains. Sub-groups II to VI contain organisms referred to as paracholera and cholera like that have been isolated from cases of choleraic diarrhoea or from water. Thus, according to Gardner and Venkatraman, the true cholera vibrio is a non-haemolytic organism containing the specific O antigen of sub-group I except by haemolysin production it is indistinguishable from El Tor vibrios containing the same O antigen. A non-specific O antigen shared to a variable extent by all the members of Group A has also been described by these workers.

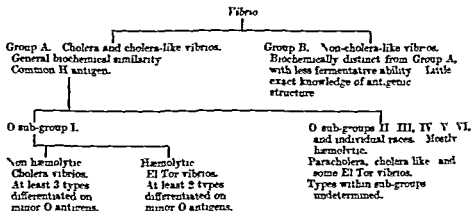


FIG. 104.

**Chemical Analysis.**—The chemical analysis of the *Vibrio* group has been intensively studied of late years by Linton and his colleagues in India (for early references see Linton, Shrivastava and Mitra 1935) who followed up the work of Landsteiner and Levine (1932). They find that the vibrios can be classified into six groups on the basis of two protein and three polysaccharide constituents (Table 34).

Proteins I and II show wide structural differences, but whether they are qualitatively distinct or whether each is a mixture of several proteins, is not yet decided (Mitra 1935). The polysaccharides exist in the cell as acetyl compounds (Linton and Mitra 1936). On hydrolysis polysaccharide I yields galactose and aldobionic acid, polysaccharide II arabinose and aldobionic acid, whereas the polysaccharide complex III yields glucose only. It is doubtful how far the polysaccharides can be regarded as distinct compounds. Linton, Shrivastava and Seal (1935) for example have found big differences in the physical, chemical and antigenic properties of preparations of a polysaccharide formed by a given vibrio grown on different media. Metabolic studies by Linton, Mitra and Mullick (1936a, b)



TABLE 34  
CHEMICAL CLASSIFICATION OF THE *VIBRIO* GROUP, ACCORDING TO LINTON AND HIS COLLABORATORS

	Protein	Polysaccharide	Source of Origin of Vibrio
Group I	I	I	Majority of strains from cholera cases
Group II	I	II	Some strains from cholera cases and some water vibrios
Group III	II	II	Mainly inagglutinable water strains
Group IV	II	I	El Tor strains and some agglutinable cholera strains found in India
Group V	II	III	Atypical cholera strains.
Group VI	I	III	Atypical cholera strains including certain old laboratory strains of true cholera vibrios

have shown that strains belonging to chemical Groups I and VI have a high rate of respiration and of aerobic glycolysis, strains of Group II are moderately active and those of Group III relatively inactive, strains of Groups IV and V have a high respiration but a low aerobic glycolysis rate. In phosphate buffered peptone water the final Eh of strains belonging to chemical Groups I, II, and VI is higher than of those belonging to Groups III, IV and V (Seal and Mitra 1939).

It is as yet too early to explain the antigenic behaviour of the *Vibrio* group in terms of chemical structure. As will be seen from Table 34, true cholera vibrios fall into chemical Groups I, II, IV and VI. In addition it has been found by Linton, Mitra and Seal (1933-39) that the transition of a true cholera vibrio from the smooth to the rough or  $\rho$  variant state may be accompanied by a change in its chemical grouping such as from Group I to Group IV. Chemical and antigenic differences have also been observed in cholera strains isolated from different stages of a given epidemic (Linton, Shrivastava, Seal and Mookerji (1938-39)).

In this country Bruce White has studied various fractions from members of the *Vibrio* group in particular relation to their antigenic structure. White (1935a) finds that, just as in the *Salmonella* group, the transformation of the smooth to the rough phase is accompanied by a loss of specific O antigens and the unmasking of a common rough antigen. In consequence, many organisms that are antigenically diverse in the smooth state show a close similarity in the rough. White has established four rough antigenic groupings. Rough Group A contains strains derived from Gardner and Venkatraman's smooth O sub group I, rough Group B from smooth O sub group II, rough Group C from smooth O sub groups III and IV, and rough Group D from unclassified smooth O sub groups.

Variants in a further stage of degradation known as  $\rho$  variants have been described by White (1934b, 1935a). Organisms of this type have lost their dominant rough O antigen and appear antigenically similar owing to the unmasking of a still deeper common  $\rho$  antigen. Little is yet known of the chemical structure of this  $\rho$  antigen, but it is very resistant to proteolytic digestion and includes a polysaccharide hapten referred to by White (1940c) as C3.

A study of the different types of cholera vibrios belonging to Gardner and Venkatraman's O sub group I led Scholtens (1933) and Heiberg (1936b) to postulate the existence of two qualitatively distinct somatic antigens A and B. Some strains appeared to contain A only, and some both A and B. White (1937b) on the other hand following up his work of 1936, disagrees with this interpretation. On the basis of agglutination and

absorption of agglutinins tests he believes that the Inaba (original), Hikojima (middle) and Ogawa (variant) strains of *V. cholerae*, which have been extensively used for serological work in the East, behave as if their heat-stable O antigens had the general structure AX, ABX, and BX. By means of precipitin tests carried out with sera prepared by the inoculation of rabbits with polysaccharide extracts, he finds that there are at least four O receptor groups on the smooth polysaccharide of the cholera vibrio. Both the Inaba and the Ogawa strains possess these four groups—two of which are type specific and two of which are common to both types. In each type one of the receptors is alkali labile and the other alkali stable.

White (1934a, 1935b) has described the occurrence of alcohol-soluble protein antigens which he refers to as Q antigens. There is reason to believe that these substances play a part in the non-specific O agglutination of boiled vibrios, described by Gardner and Venkatraman.

White's studies were interrupted by the war, but it may be worth while summarizing his findings to date. From vibrio bodies he has isolated (1) a heat labile protein antigen (White 1940b), (2) a heat-stable protein antigen possibly associated with a hapten Cy2 (White 1940d), (3) an alcohol-soluble Q protein fraction, (4) the differential S R and  $\rho$  antigens with their respective polysaccharide haptens Cz, C $\beta$  and C $\delta$ , (5) another hapten Cy1 is probably also of somatic origin, and (6) another, the "rugose" hapten (White 1940c) has been derived from the intercellular secretion of rugose variant strains. Antibodies for all these components may occur in the sera of rabbits immunized with living cultures of *V. cholerae*.

From this brief summary it will be apparent that the findings of Bruce White and of Linton and his colleagues are by no means easy to interpret, and that much further work will have to be done before the antigenic structure of even the cholera vibrio can be expressed in chemical terms. For a review of the chemistry and antigenic structure of the vibrios, see Linton (1940).

**Pathogenicity.**—The cholera vibrio causes Asiatic cholera in man. Metchnikoff's vibrio apparently is responsible for a choleraic disease in chickens (Gamaleia 1888a)—not for true chicken cholera, which is due to a member of the *Pasteurella* group. It is possible that *V. phosphorescens* may cause acute gastro-enteritis in man, but this has not been proved conclusively (Jermoljewa 1926).

A disease simulating cholera may be reproduced in guinea pigs and new born rabbits by certain experimental procedures (see Chapter 63). The cholera vibrio when given by mouth, or injected *per rectum*, is harmless to mice, rabbits, guinea pigs, and monkeys. Intrapentoneal injection into guinea pigs is fatal within 24 hours. If a small dose of vibrios— $\frac{1}{2}$  loopful of an 18-hours' agar culture—is given the animal dies of toxæmia and at necropsy the pentoneal cavity is sterile. If a larger dose is given,  $\frac{1}{2}$  loopful, cultures from the pentoneal cavity may be positive, and if a still larger dose is given, 1 or more loopfuls, the vibrios may be recovered also from the heart blood. Intrapentoneal injection of mice is fatal in 24 to 48 hours. Intravenous injection of young rabbits is fatal in 1 to 5 days. According to Botman (see van Loghem 1933) the rat is comparatively unaffected by the intrapentoneal inoculation of killed cholera vibrios, whereas it is susceptible to El Tor vibrios.

Metchnikoff's vibrio is more invasive than the cholera vibrio. Even after a small dose given intrapentoneally to guinea pigs, the vibrios can be recovered from the heart's blood. It is fatal to guinea pigs even when given subcutaneously, under these conditions the cholera vibrio gives rise merely to a local abscess. Both guinea pigs and chickens can be infected by feeding with *V. metchnikovi*. Moreover this organism is pathogenic to pigeons, on intramuscular injection, while the cholera

vibrio is not, except occasionally in large doses (Wherry 1905) Pigeons injected intramuscularly with  $\frac{1}{2}$  agar culture of *V. metchnikovi* die in about 8 hours with general septicæmia (Metchnikoff 1893) Intratracheal injection appears to be even more fatal, since not only guinea pigs, pigeons, and fowls, but also rabbits may be infected by this route (Gamaleia 1888b) Deneke's *Vibrio tyrogenus* is pathogenic for the guinea pig and the pigeon Half an agar culture injected intraperitoneally into a guinea pig was fatal in 6 hours, and a whole agar culture injected intramuscularly into a pigeon was fatal in 7 hours (Metchnikoff 1893) Finkler Prior's *Vibrio proteus* resembles *V. tyrogenus*, but is slightly less virulent

*V. phosphorescens* is pathogenic for guinea pigs rabbits and pigeons About 500 million organisms injected intraperitoneally into guinea pigs, intravenously into rabbits, or intramuscularly into pigeons proved fatal in 24 hours vibrios were isolated post mortem from the heart's blood of the pigeons (Jermoljewa 1926) If a guinea pig that has died after intraperitoneal injection is opened up and placed in the dark, the viscera are seen to exhibit a marked phosphorescence (Kutscher 1893)

Most other members of the group are non pathogenic The virulence of *V. cholera* is variable Freshly isolated strains are more virulent than those kept in the laboratory Moreover, even on isolation, the virulence of different strains to laboratory animals appears to vary Haffkine (1892) stated that it was possible to raise the virulence by passing the organisms through the peritoneal cavity of guinea pigs, between each injection the peritoneal exudate was exposed to the air for some time at room temperature By growing the vibrio in broth in a constantly aerated atmosphere and subculturing every 2 or 3 days, the virulence was said to diminish Gotschlich and Weigand (1895) also stated that the virulence might be raised by intraperitoneal passage through guinea pigs

**Variation**—The occurrence of smooth, rough and rugose forms of the cholera vibrio has already been referred to in the sections on cultural and antigenic characters Confusion has arisen from paying too much attention to colonial variation without a full study of antigenic and other properties As White (1938) points out, the essential feature of rough variants is their absence of the specific smooth polysaccharide

There is a widespread belief that cholera vibrios under unfavourable conditions such as in water, may lose their specific characters and be transformed into some other type of vibrio The alleged transmutation of vibrios in the laboratory by Lanton (1935), Lanton, Shrivastava and Mitra (1934-35) Lanton Seal and Mitra (1938) and Taylor and Ahuja (1935-36) has tended to strengthen this belief White (1937a) who has studied some of the strains before and after their alleged change, can find no evidence to support the conception of vibriotic transmutability Until further studies have been made it is probably wiser to adopt a strictly conservative attitude toward the limits of variation within the different species of this group

We append a summarized description of *V. cholera*, and brief notes on the characters of other species which have been described and named (For general classification of members of this group see Heiberg 1935)

### *Vibrio cholerae*

*Synonym*—Comma bacillus

*Isolation*—Koch in 1884 (1886)

*Habitat*—Intestinal contents of cholera patients and carriers

**Morphology**—Slightly curved bacillus, often resembling a comma. Varies considerably in size,  $1.5-4 \mu \times 0.2-0.4 \mu$ . One end often blunter than the other ends rounded. Axis generally curved. Sides converging or parallel. Arranged singly or in s-shaped pairs, sometimes short chains are found, and sometimes spirals. In the intestinal contents, arranged like fish in a stream. In old cultures the bacilli are very small, resembling granules, and stain poorly. Involution forms numerous. Actively motile by a single polar flagellum. Gram negative. Non sporing. Non-acid fast.

**Agar Plate**.—24 hours at  $37^{\circ} \text{C}$  Round, 1-2 mm. in diameter low convex translucent, greyish yellow colonies with smooth, or finely granular glistening surface and entire edge and of amorphous or finely granular structure consistency butyrus, easily emulsifiable. 7 days slightly larger edge entire or undulate surface sometimes studded with small, knob-like secondary colonies colony is sometimes surrounded by a narrow effuse, transparent peripheral extension. Crystals often formed in the medium.

**Agar Slope**.—24 hours at  $37^{\circ} \text{C}$  Good, raised, translucent greyish yellow layer of growth, with smooth, glistening surface and edge formed of single colonies. 7 days surface sometimes studded with small, knob-like secondary colonies. Crystals often formed in the medium.

**Gelatin Plate**.—2 days at  $23^{\circ} \text{C}$  Round 0.5 mm. in diameter amorphous, raised or low convex, greyish white opaque colonies, with smooth or slightly granular surface and entire or crenated edge. Zone of liquefaction around colony small flocculi of growth in liquefied gelatin.

**Gelatin Stab**.—3 days at  $23^{\circ} \text{C}$  Good filiform growth, confluent at top, discrete below extending to bottom of tube. Infundibuliform or napiform liquefaction thick, yellowish brown pellicle on surface of liquid gelatin, and coarsely granular turbidity.

**Broth**.—24 hours at  $37^{\circ} \text{C}$  Abundant growth with moderate turbidity a slight powdery deposit, and a thick surface pellicle breaking up on shaking into coarse membranous and granular pieces.

**Loeffler's Serum**.—10 days at  $37^{\circ} \text{C}$  Good growth with partial liquefaction.

**Horse Blood Agar Plates**.—24 hours at  $37^{\circ} \text{C}$  Abundant growth colonies are surrounded for 2 mm. by a zone of  $\alpha$  or  $\beta$ -hemolysis.

**Potato**.—7 days at  $37^{\circ} \text{C}$  Good, confluent cake-like growth with smooth glistening surface.

**MacConkey Plate**.—24 hours at  $37^{\circ} \text{C}$  Good growth of clear colourless colonies smaller than those on agar. After 7 to 9 days the colonies take on a reddish colour.

**Resistance**.—Not specially resistant. Easily killed by drying. Destroyed by heat at  $55^{\circ} \text{C}$  in 15 minutes. Dried on linen or threads they survive 1 to 3 days. Killed by 0.5 per cent. phenol in a few minutes. Survive in clean tap water up to 30 days, but perish in 24 hours in cesspool water.

**Metabolism**.—Strongly aerobic very slight growth noticeable on agar and in broth after a week under strictly anaerobic conditions. Optimum temperature  $37^{\circ} \text{C}$ . limits  $16-42^{\circ} \text{C}$ . Optimum pH 7.0-8.0 Limits for growth pH 6.4-9.6 Growth favoured slightly by blood. Grows well and rapidly in peptone water. No soluble haemolysin formed for sheep or goat cells. Proteolytic and diastatic ferments secreted.

**Biochemical**.—Acid, no gas, in glucose, maltose, mannitol, and sucrose in 1 to 3 days after 14 days there may be slight acid in lactose. L.M. acid, or acid and clot in 14 days. Indole + Cholera red reaction + M.R. — V.P. — Nitrates reduced.  $\text{NH}_3$  —  $\text{H}_2\text{S}$  — in 14 days. Catalase — M.B. reduction —

**Antigenic Structure**.—All strains have a common O antigen, but 3 sub-types are distinguishable. Immune sera prepared by injection of rabbits, goats, or horses with living vibrios contain specific bactericins, demonstrable by Pfeiffer's test.

**Pathogenicity**.—Causes Asiatic cholera in human beings. A similar disease may be repro-

duced experimentally in new born rabbits by feeding and in young guinea pigs by Koch's procedure. Pathogenic on *ip* or *iv* inoculation into guinea-pigs, rabbits, and mice, but not as a rule into pigeons. One loopful of an 18 hours agar culture of a virulent strain injected *ip* into a young guinea pig is fatal within 24 hours. P.M. congestion of peritoneal and pleural cavities with some sero sanguineous fluid. Small gut congested, may be fibrin over the abdominal viscera. Vibrios may or may not be cultivated from the peritoneal cavity. If a large dose is given the vibrios can be recovered from the peritoneal fluid and the heart blood. *Iv* injection of five loopfuls of an 18 hour agar culture of a virulent strain into rabbits is fatal in 48 hours or less. P.M. small gut congested and contains thin fluid. Vibrios generally recoverable from the blood. No true exotoxin formed but disintegrated bodies of bacilli are very toxic to animals. Virulence rapidly falls on artificial cultivation.

**Finkler-Prior's *Vibrio proteus***—Isolated by Finkler and Prior (1884) from the old putrid excreta of a patient suffering from gastro-enteritis. Morphologically and culturally it resembles the cholera vibrio but it can be differentiated by serological reactions and by its failure to give the cholera red reaction. Has frequently been found in water.

**Deneke's *Vibrio tyrogenus***—Found by Deneke (1885) in cheese. Resembles the cholera vibrio but liquefies gelatin more rapidly, grows poorly or not at all on potato, and does not give the cholera red reaction.

***Vibrio metchnikovi***—Isolated by Gamaleia (1885a) from the blood and intestinal contents of chickens dying from a cholera like disease at Odessa. Resembles the cholera vibrio very closely, gelatin is liquefied more rapidly, growth on MacConkey is poorer. Cholera red and other biochemical reactions are identical with those of *V. cholerae*. It is much more invasive when injected into animals, killing guinea pigs injected subcutaneously and pigeons injected intramuscularly (see Pathogenicity). Can be separated from *V. cholerae* by agglutination and Pfeiffer's reactions. Has been isolated from water.

***Vibrio phosphorescens***—Isolated by Dunbar (1893) from the Elbe in 1893. Shown by Kutscher (1893) to exhibit phosphorescence in the dark. This occurs on ordinary media at 22° C., reaching its maximum in 24 to 48 hours in gelatin broth or peptone water, and disappearing rapidly. It is a function of the living bacilli. Phosphorescence is not visible in cultures incubated anaerobically. *V. phosphorescens* grows in and liquefies gelatin, more rapidly than *V. cholerae*. Grows very poorly or not at all on potato. Haemolytic and diastatic, produces indole, gives acid in glucose, mannitol, lactose, and later maltose (Jermoljewa 1926). It has been isolated from human faeces (Jermoljewa 1926).

**El Tor *Vibrio***—Isolated by Gotschlich (1906) in 1905 from six pilgrims who had died of dysentery or gangrene of the colon at the Tor quarantine station on the Sinai Peninsula. Forms soluble haemolysin for sheep and goat cells. Gives atypical Pfeiffer reaction (Neufeld and Haendel 1907). Usually gives positive Voges Proskauer reaction (van Loghem 1938). Killed vibrios injected intraperitoneally are said to be more toxic for the rat than killed cholera vibrios (see van Loghem 1938). According to Takita (1933) the El Tor vibrio produces a true thermolabile exotoxin, distinct from the haemolysin and proving fatal to mice on intraperitoneal inoculation. Relation to *V. cholerae* still doubtful (see Fig. 104).

***Vibrio berolinensis***—Isolated by Neisser (1893) from water to which cholera vibrios had intentionally been added. Resembles the cholera vibrio closely, gelatin colonies are smaller and animal pathogenicity is low. But probably it is merely a variant of the true cholera vibrio. Similarly the "*Vibrio Ivanoff*" (Ivanoff 1893) which was cultured from the faeces of a typhoid patient to which cholera vibrios had been added, and which differed in unimportant particulars from the cholera vibrio, is also a variant of the true cholera vibrio (Dieudonné 1894).

*Vibrio danubicus*—Isolated by Heider (1893) from the Danube canal. Resembles the cholera vibrio closely but can be differentiated by serological reactions.

*Vibrio heliogenes*—Isolated by Fischer (1893) from the diarrhoeal faeces of a woman. Some of the mice inoculated subcutaneously developed ulcers of the skin—hence the name.

The Massauah vibrio was isolated by Pasquale (1891) from the faeces of a patient who was probably not suffering from cholera. Resembles the cholera vibrio closely but has four peritrichate flagella—it is therefore not a true *Vibrio*. The Ghinda vibrio was isolated by Pasquale from water—it was regarded as a true cholera vibrio but has since been shown to be distinguished from it by its immunity reactions.

The Nasik vibrio differs in several respects from *V. cholerae*. It is less like a comma and is short and rather fat—arranged singly and with great regularity—filamentous forms are common. The colonies on agar are more opaque than those of cholera. Infundibuliform liquefaction occurs in gelatin—later stratiform—the liquefied gelatin is uniformly turbid and contains no floculi. No growth under anaerobic conditions. Acid in glucose only. I.M. purple and clotted indole—nitrate reduction— $\text{NH}_3 + \text{H}_2\text{S}$ —Catalase++ M.B. reduction—cholera red react on— $\beta$ -haemolysis in horse blood agar plates in 4 days. Café-au-lait growth on potato. Broth cultures are very toxic to rabbits on intravenous injection (Kolle and Schürmann 1912).

*Vibrio fetus*—This organism on account of its special characteristics must be considered separately. It was first isolated and described by M Fadyean and Stockman in 1913 (see Report 1913) who found it in the uterine exudate of aborting sheep. Smith (1918) cultivated the same organism in America from the foetuses of aborting cows (see Chapter 7c)—he named it *V. fetus* (Smith and Taylor 1919). In young cultures it is generally shaped like a comma but later it assumes a spirillar appearance. It is questionable whether this organism should be classified as a *Vibrio* or as a *Spirillum*—its characters partake of both groups. But since Smith has placed it with the vibrios since it has a single polar flagellum and since it is Gram negative it is perhaps best to regard it as belonging to the *Vibrio* group.

Morphologically the smallest forms appear as minute, slender shaped threads—the longest forms may stretch nearly across the field of the microscope. In length it is 1.5 to 5  $\mu$  or more and in breadth about 0.2 to 0.3  $\mu$ . A single organism shows one or two spirals—the length of each spiral is about 2  $\mu$  and the amplitude about 0.5  $\mu$ . In the long forms the spirals are drawn out so that their length is far greater than their breadth. The short forms are sharply curved—the spirals often show an obtuse-angled curve. In young cultures the vibrios are actively motile by a single polar flagellum—in cultures a week old very few are motile. The organism is best stained with alkaline methylene blue, the staining being prolonged over night. It is Gram negative. In old cultures many of the organisms show granular degeneration.

For growth in artificial media, a reduced oxygen pressure is required. When first isolated it will not grow on agar without the addition of blood or some other animal fluid. The growth is very delicate and occurs at the edges of the slope between the agar and the glass—subsequently it spreads round the convexity of the agar. After some months of cultivation in the laboratory a thin surface growth may be obtained. Growth in fluid media—even in blood broth, does not occur till the strain has become thoroughly accustomed to saprophytic conditions. There is no growth in gelatin milk or potato. Sugars are not fermented, and there is no production of indole. In cultures it lives for 2 to 20 weeks at room temperature but dies rapidly in the ice-chest. Dried on threads it lives for less than 3 hours. It is killed by 56° C in 5 minutes. The optimum temperature for growth is 37° C. Antigenically it appears by agglutination to be homogeneous (Smith and Taylor 1919). It is non-pathogenic to laboratory animals. Under natural conditions it gives rise apparently to abortion in cattle and sheep. Experimental inoculation of pure

cultures into pregnant cows may be followed by disease of the fetal membranes (Smith 1919)

A closely related organism, named *Vibrio jejuni*, has been described by Jones and Little (1931), and Jones, Orcutt, and Little (1931). It appears to be responsible for a disease of calves and older cattle, which may occur in epidemic form during the autumn and winter months, and is known as *winter dysentery* or *black scour*. The organisms are most abundant in the jejuni.

## SPIRILLUM

DEFINITION—*Spirillum*.

- Rigid rods of spiral form varying considerably in the number, length, and breadth of the spirals. Usually motile by means of a tuft of polar flagella (3 to 20), which are mostly semicircular in shape. The flagella occur at one or both poles, their number varies greatly and is difficult to determine, since in stained preparations several are often united into a common strand. Generally Gram positive. Some species form a reddish yellow, or greenish yellow pigment. Found in water or putrid infusions.

Type species *Spirillum undula*

Not many organisms in this group have been described. One of the best known is *Spirillum rubrum*, which was isolated by Esmarch (1887) from a mouse that had been decomposing for 3 months under water.



FIG. 105.—*Spirillum rubrum*  
From a broth culture 2 days, 30° C ( $\times 1000$ )

Morphologically the spirilla show considerable variation. Their length may vary from 1 to 50  $\mu$ , and the number of spirals from 1 to about 50. The length of the individual spiral varies according to the species of organism, in some spirilla the spirals are close set, each one being not more than about 1  $\mu$  in length, in others they are looser, and may be 10  $\mu$  or so in length. The width of the organisms

*Vibrio danubius*.—Isolated by Heider (1893) from the Danube canal. Resembles the cholera vibrio closely, but can be differentiated by serological reactions.

*Vibrio helcogenes*.—Isolated by Fischer (1893) from the diarrhoeal faeces of a woman. Some of the mice inoculated subcutaneously developed ulcers of the skin, hence the name.

The Massauah vibrio was isolated by Pasquale (1891) from the faeces of a patient who was probably not suffering from cholera. Resembles the cholera vibrio closely, but has four peritrichate flagella, it is therefore not a true *Vibrio*. The Ghinda vibrio was isolated by Pasquale from water, it was regarded as a true cholera vibrio, but has since been shown to be distinguished from it by its immunity reactions.

The Nasik vibrio differs in several respects from *V. cholerae*. It is less like a comma and is short and rather fat arranged singly, and with great regularity, filamentous forms are common. The colonies on agar are more opaque than those of cholera. Infundibuliform liquefaction occurs in gelatin later stratiform, the liquefied gelatin is uniformly turbid and contains no flocculi. No growth under anaerobic conditions. Acid in glucose only, L.M. purple and clotted, indole—, nitrate reduction—,  $\text{NH}_4^+$  +,  $\text{H}_2\text{S}$ —, Catalase ++, M.B. reduction—, cholera red reaction—,  $\beta$ -haemolysis in horse blood agar plates in 4 days. Case-au lait growth on potato. Broth cultures are very toxic to rabbits on intravenous injection (Kolle and Schurmann 1912).

*Vibrio fetus*.—This organism, on account of its special characteristics, must be considered separately. It was first isolated and described by M'Fadyean and Stockman in 1913 (see Report 1913), who found it in the uterine exudate of aborting sheep. Smith (1918) cultivated the same organism in America from the foetuses of aborting cows (see Chapter 75), he named it *V. fetus* (Smith and Taylor 1919). In young cultures it is generally shaped like a comma, but later it assumes a spirillar appearance. It is questionable whether this organism should be classified as a *Vibrio* or as a *Spirillum*, its characters partake of both groups. But since Smith has placed it with the vibrios since it has a single polar flagellum, and since it is Gram negative, it is perhaps best to regard it as belonging to the *Vibrio* group.

Morphologically, the smallest forms appear as minute, slender, s-shaped threads, the longest forms may stretch nearly across the field of the microscope. In length it is 1.5 to 5  $\mu$  or more and in breadth about 0.2 to 0.3  $\mu$ . A single organism shows one or two spirals the length of each spiral is about 2  $\mu$ , and the amplitude about 0.5  $\mu$ . In the long forms the spirals are drawn out, so that their length is far greater than their breadth. The short forms are sharply curved, the spirals often show an obtuse-angled curve. In young cultures the vibrios are actively motile by a single polar flagellum, in cultures a week old very few are motile. The organism is best stained with alkaline methylene blue the staining being prolonged over night. It is Gram negative. In old cultures many of the organisms show granular degeneration.

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cultures into pregnant cows may be followed by disease of the foetal membranes (Smith 1919)

A closely related organism, named *Vibrio jejuni*, has been described by Jones and Little (1931), and Jones, Orcutt, and Little (1931). It appears to be responsible for a disease of calves and older cattle, which may occur in epidemic form during the autumn and winter months, and is known as *winter dysentery* or *black scours*. The organisms are most abundant in the jejunum.

## SPIRILLUM

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Not many organisms in this group have been described. One of the best known is *Spirillum rubrum*, which was isolated by Esmarch (1887) from a mouse that had been decomposing for 3 months under water.



FIG. 105.—*Spirillum rubrum*  
From a broth culture, 2 days, 30°C ( $\times 1000$ )

Morphologically the spirilla show considerable variation. Their length may vary from 1 to 50  $\mu$ , and the number of spirals from 1 to about 50. The length of the individual spiral varies according to the species of organism, in some spirilla the spirals are close set, each one being not more than about 1  $\mu$  in length, in others they are looser, and may be 10  $\mu$  or so in length. The width of the organisms

varies from about  $0.3$  to  $1.0\ \mu$ , and the amplitude of the spirals from about  $0.8$  to  $2.0\ \mu$ . Even amongst different organisms of the same strain, there is often considerable variation in the number and size of the spirals, together with organisms showing regularly disposed spirals, there may be seen others of two or three times the length, with only one or two irregular undulations. In shape the whole organism may be straight, or it may be bent in one or more places, generally acutely. Filamentous forms are not uncommon. On agar or gelatin the spiral shape may be almost lost, and the organisms closely resemble vibrios. As a rule the curvature is very marked, and there is a tendency for the organisms to be arranged in pairs end to end with the concavities facing in the same direction, so as to present a scalloped appearance, s shaped forms too are common. In young cultures the spirilla are motile—generally by tufts of flagella at the poles. Unlike the spirochaetes, the spirilla stain readily with the ordinary aniline dyes, and are usually Gram positive. Growth is fairly free, though not abundant, on the ordinary media. Most of the water spirilla form a pigment of red, yellow, or greenish yellow colour. The pigment, at least of *Sp. rubrum*, is formed most readily at a low oxygen pressure: it is well marked in the depths of gelatin stab cultures and hardly noticeable on surface growths. The optimum temperature for growth is  $25-30^{\circ}\text{C}$  as a rule. Aerobic conditions are preferred, growth under strict anaerobiosis is very slight. None of the species forms spores, and none is particularly resistant to heat or disinfectants. The biochemical characteristics have not been fully studied. None of the members except *Spirillum minus* (see p 1838) is pathogenic for man or animals. We append a description, based largely on personal observations, of *Spirillum rubrum*.

### *Spirillum rubrum*

*Isolation*.—Esmarch 1887 from a mouse decomposing under water.

*Habitat*.—Water.

*Morphology*.—On solid media the organisms are sharply curved rods,  $2-3\ \mu \times 0.4\ \mu$ , arranged singly and in s shaped or semicircular pairs. In fluid media long spirals are formed  $3-10\ \mu$  or more in length. The axis of the spiral is straight, or bent sharply at a right-angle, the number of spirals varies from about 1 to 8. Ends are sharp, drawn out or sometimes blunt. Long thread like forms also seen. Motile by bundles of flagella at both poles. Gram positive.

*Agar Plate*.—2 days at  $23^{\circ}\text{C}$ . Round,  $0.4\ \text{mm}$ . in diameter, convex, amorphous, almost colourless and transparent colonies with smooth glistening surface and entire edge consistency butyrous, easily emulsifiable. 7 days, rather larger and of a pinkish colour.

*Agar Stroke*.—2 days at  $23^{\circ}\text{C}$ . Poor slightly raised, and almost transparent partly confluent growth with irregular surface and edge formed of single colonies. 7 days slight pinkish coloration.

*Gelatin Plate*.—4 days at  $23^{\circ}\text{C}$ . Small,  $0.3\ \text{mm}$ . in diameter, water-clear, convex colonies, with smooth surface and entire edge. No liquefaction. Deep colonies are pink.

*Gelatin Stab*.—5 days at  $23^{\circ}\text{C}$ . Poor to moderate, filiform growth of very tiny discrete red colonies, extending nearly to bottom of tube. No surface growth, no liquefaction.

*Broth*.—2 days at  $23^{\circ}\text{C}$ . Moderate growth with slight turbidity, and a pale pink flocculo-granular deposit, not disintegrating completely on shaking, no surface growth.

*MacConkey*.—7 days at  $23^{\circ}\text{C}$ . No growth.

- Potato*—7 days at 28° C Poor growth of discrete or partly confluent red colonies.  
*Horse Blood Agar Plate*—2 days at 28° C Very small, low convex colonies, no hæmolysis  
*Resistance*—Not specially resistant Dried on silk threads, the organisms do not survive longer than 6 to 8 days  
*Metabolism* Aerobic, grows very poorly under anaerobic conditions Optimum temperature 25–30° C. often little growth at 37° C. Grows very poorly in peptone water No hæmolysin formed Red pigment formed, best under a low oxygen pressure  
*Biochemical*—No sugars fermented LM unchanged Indole— Cholera red reaction— MR— VP— Nitrates not reduced  $\text{NH}_3$ +  $\text{H}_2\text{S}$ — Catalase+ MB reduction—  
*Pathogenicity*—Non pathogenic to man or animals

Other members of this group that have been described are *Spirillum undula*, *Spirillum serpens*, *Spirillum volutans*, and *Spirillum minus* (see p 1838)

## REFERENCES

- ABDOOSH, Y B (1932) *Brit J exp Path.*, 13, 42.  
 ANDU, A B and NIEKERK, J VAN (1929) *Zbl Bakt.*, 112, 519  
 BELTRANI, I (1926) *J Path Bact.*, 29, 251.  
 BERNARD, P N, GUILLERM, J, and GAILLET, J (1937) *C R Soc Biol*, 128, 180, 303, 391, 478, 508, (1939) *Ibid.*, 130, 23, 147, 228.  
 BOCCIA, I (1911) *Zbl Bakt.*, 60, 431  
 COMBESCO POPESCO, C. and COCORA, I (1936) *C R Soc Biol*, 124, 151  
 CHESTER, C V. (1913) *J infect Dis.*, 12, 472  
 CRENDIROPOULO (1912) *Conseil san maritime quarant. d'Egypte.*  
 DEVEZE, T (1885) *Dtsch. med Wochr.*, 11, 33  
 DIEUDONNÉ. (1894) *Zbl Bakt.*, 18, 363, (1909) *Ibid.*, 50, 107  
 DOORENBOS, W (1936) *C R Soc Biol*, 121, 128, 130  
 DUBAR (1893) *Dtsch med Wochr.*, 19, 799  
 DUNGER, von (1891) *Z Hyg Infektkr.*, 20, 147  
 ESMARCH, E (1887) *Zbl Bakt.*, 1, 225  
 FINKLER and PRIOR. (1884) *Dtsch med Wochr.*, 10, 579, 632  
 FISCHER. (1893) *Dtsch med Wochr.*, 19, 541, 575, 598, 627  
 GAMALÉIA, M N (1885a) *Ann Inst Pasteur*, 2, 482, (1885b) *Ibid.*, 2, 552  
 GARDNER, A D and VENKATRAMAN, K V (1935) *J Hyg, Camb.*, 35, 262.  
 GOHAR, M A (1932) *Brit. J exp Path.*, 13, 371  
 GORDON, M H (1906) *Brit med J.*, 11, 197  
 GOTSCHLICH E (1895) *Z Hyg Infektkr.*, 20, 489, (1906) *Ibid.*, 53, 291  
 GOTSCHLICH, E. and WEIGANO, J (1895) *Z Hyg. Infektkr.*, 20, 378  
 HAHN, M and HIRSCH, J (1929) *Z Hyg Infektkr.*, 110, 355  
 HAYFINK, W M W (1892) *C R Soc Biol*, 44, 635, 671  
 HEIBERG, B (1935) On the classification of *Vibrio cholerae* and the *Cholera like vibrios*.  
 Arnold Busck, Copenhagen, (1936a) *J Hyg, Camb.*, 36, 114, (1936b) *Ibid*, 38, 118  
 HEIDER, A (1893) *Zbl Bakt.*, 14, 341  
 IVANOFF, M (1893) *Z Hyg Infektkr.*, 15, 434  
 JERMOLSKWA, S. (1926) *Zbl Bakt.*, 100, 170  
 JONES, F S and LITTLE, R B (1931) *J exp Med.*, 53, 835 845.  
 JONES, F S, ORCUTT, M., and LITTLE, R. B (1931) *J exp. Med.*, 53, 853  
 KABASHIMA, T (1918) *C R Soc. Biol.*, 81, 618  
 KITASATO, S. (1889) *Z Hyg Infektkr.*, 5, 134  
 KOCH, R (1886) "The Etiology of Cholera," New Sydenham Soc., 115, 327  
 KOLLER, W and SCHÜRMANN, W (1912) See Koller and Wassermann Handb. path Mikrov.  
 (1912–13) 4, 1  
 KUTSCHER, (1893) *Dtsch med Wochr.*, 19, 1301, (1895) *Z Hyg Infektkr.*, 19, 40  
 LANDSTEINER, K. and LEVINE P (1927) *J exp Med*, 46, 212  
 LINTON, R. W. (1935) *Bull. Off int Hyg publ.*, 27, 1104; (1940) *Brit. Acc.*, 4, 2  
 LINTON, R W and MITRA, R. N (1936) *Indian J med Res.*, 24, 323  
 LINTON, R W, MITRA, B N., and MCLICK, D N (1936a) *Indian J med Res.*, 24, 317  
 (1936b) *Ibid.*, 24, 317

- LINTON R. W., MITRA B. N., and SEAL, S. C. (1935-39) *Indian J. med. Res.*, 26, 329  
 LINTON R. W., SEAL, S. C., and MITRA, B. N. (1935) *Indian J. med. Res.*, 25, 57.  
 LINTON R. W., SRIVASTAVA, D. L., and MITRA, B. N. (1934-35) *Indian J. med. Res.*, 22, 633.  
 LINTON R. W., SRIVASTAVA D. L., and SEAL, S. C. (1935) *Indian J. med. Res.*, 25, 569  
 LINTON R. W., SRIVASTAVA D. L., SEAL, S. C., and MOOKERJEE, S. P. (1935-39) *Indian J. med. Res.*, 26, 41  
 LOGHEM, J. J. VAN (1932) *Ned. Tijdschr. Geneesk.*, 76, 1939 (1935) *Bull. Off. int. Hyg. publ.*, 30, 1520.  
 MAASSEN, A. (1902) *Arch. Pathog. exp. appl.*, 18, 21  
 MANWARING, W. H. BOYD W. H. and OKAMI S. (1923) *J. infect. Dis.*, 32, 307  
 METCHNIKOFF, E. (1893) *Ann. Inst. Pasteur*, 7, 562.  
 MITRA, B. N. (1935) *J. trop. Med. (Hyg.)*, 41, 37  
 NAFTER, L. E. and GUPTA, S. K. (1942) *Indian med. Gaz.*, 77, 717  
 NEISSER, M. (1893) *Arch. Hyg.*, 19, 194.  
 NEUFELD and HAENSEL. (1907) *Arch. Pathog. exp. appl.*, 23, 536.  
 NICATI W. and RIETSCHE M. (1934) *C. P. Acad. Sci.*, 99, 925.  
 NOBECCHI, K. (1925) *J. Bact.*, 10, 197  
 PASQUALE, A. (1891) *G. med. Esere.*, 39, 1009  
 PALADINO-BLANDINI A. (1906) *Ann. Igien. (epid.)*, 15, 301  
 PASECHKA, C. L., CHATTERJEE, D. N., and DAS, P. C. (1935-39) *Indian J. med. Res.*, 26, 33.  
 PFEIFFER R. (1902) *Z. Hyg. Infektkr.*, 11, 303 (1894) *Ibid.*, 13, 1 (1905) *Ibid.*, 20, 199  
 POLLITZER, R. (1935-36) *Pap. nat. Quar. Ser.*, Shanghai Ser. vi. p. 70.  
 REPORT. (1913) Rep. Dep. Comm. Epidemic Abortion, Part III. London.  
 RUTTER, M. A. (1907) *Conseil san. mar. et Quarant. d'Egypte*.  
 SCHOLTESS, R. T. (1933) *C. R. Soc. Biol.*, 114, 490, 422.  
 SEAL, S. C. and MITRA, B. N. (1939) *Indian J. med. Res.*, 26, 625.  
 SHOTSHA, A. T. (1931) *Bull. Off. int. Hyg. publ.*, 23, 1022.  
 SMITH, T. (1894) *Zbl. Bakt.*, 16, 324. (1913) *J. exp. Med.*, 23, 701. (1919) *Ibid.*, 30, 313  
 SMITH, T. and TAYLOR, M. S. (1919) *J. exp. Med.*, 30, 299.  
 TAKITA J. (1939) *Kitarato Arch.*, 16, 18.  
 TAYLOR, J. and AHUJA M. L. (1935-36) *Indian J. med. Res.*, 23, 95, 531. (1935-39) *Ibid.*, 26, 1  
 TAYLOR, J., PANDIT S. R., and READ W. D. B. (1937) *Indian J. med. Res.*, 24, 931  
 TAYLOR, J., READ W. D. B. and PANDIT S. R. (1936) *Indian J. med. Res.*, 24, 349  
 VASSILIADIS, P. C. (1937) "Étude sur la bactériologie des vibrions et l'épidémiologie du choléra. Thèse Louvain.  
 VEDDER, A. and DAM, W. VAN (1932) *Zbl. Bakt.*, 126, 145.  
 WHEATY W. B. (1905) *J. infect. Dis.*, 2, 309  
 WHITE, P. B. (1934a) *J. Path. Bact.*, 39, 529 (1934b) *Ibid.*, 39, 530, (1935a) *J. Hyg., Camb.*, 35, 347 (1935b) *Ibid.*, 35, 493 (1937a) *J. Path. Bact.*, 44, 490, (1937b) *Ibid.*, 44, 706 (1938) *Ibid.*, 46, 1 (1940a) *Ibid.*, 50, 160, (1940b) *Ibid.*, 50, 165, (1940c) *Ibid.*, 51, 447 (1940d) *Ibid.*, 51, 449  
 YEN A. C. H. (1932-3) *Proc. Soc. exp. Biol.*, N. Y., 30, 884.  
 ZIMMERMAN E. (1932) *Zbl. Bakt.*, 127, 146 (1933) *Z. Immunforsch.*, 79, 219

## CHAPTER 23

### NEISSERIA

#### DEFINITION.—*Neisseria*

Gram negative cocci usually arranged in pairs. Strict parasites often growing poorly on ordinary media, but growing well on serum media. Frequently pathogenic.

Type species is *N. gonorrhoea*.

The first member of this group to be described was the gonococcus—it was observed by Neisser in 1879 in the pus cells of patients with gonorrhoea and was successfully cultivated by Bumm (1885a, b) and by Leistikow and Loeffler (Leistikow 1882) in 1882. Weichselbaum isolated the meningococcus from the cerebrospinal fluid of patients with cerebrospinal meningitis in the year 1887. In 1895 Jaeger described a similar organism, which he regarded as identical with the meningococcus, but which was almost certainly not this organism—it is now known as the *Diplococcus crassus*. R. Pfeiffer (see Flugge 1896) described the *Micrococcus catarrhalis* in 1896, he found it in the bronchioles and alveoli of children with broncho-pneumonia, it was carefully studied in 1902 by Ghon and H. Pfeiffer. In 1906 von Lingelsheim described a number of Gram negative cocci in the nasopharynx of healthy and diseased persons, these included the *Micrococcus pharyngis siccus*, the *Micrococcus pharyngis cinereus*, the *Diplococcus mucosus*, and the *Micrococcus pharyngis flatus* i, ii, and iii. More recently Branham (1930) has added another member to the group *N. flavescens*. This organism was isolated from the spinal fluid of patients with epidemic cerebrospinal meningitis.

**Habitat**—With the exception of the gonococcus, which is the causative organism of gonorrhoea, those species of Gram negative cocci which have been adequately described are found almost exclusively in the nasopharynx of healthy and diseased persons, or, in the case of the meningococcus, in the meninges and cerebrospinal fluid of patients with cerebrospinal fever.

**Morphology**—The members of the group are all Gram negative cocci, but they differ considerably in their morphology and arrangement and in the ease with which they are decolorized by alcohol. Not only do they differ from one another, but the same organism may vary considerably according to environmental conditions, thus, in the body, the meningococcus and the gonococcus present an almost typical arrangement in the form of diplococci with flattened or slightly concave adjacent sides, but in culture they appear as oval or spherical cocci without the typical diplococcal arrangement. Most of the members of the group are arranged in pairs, tetrads, or small groups, but some members, such as *N. pharyngis*,

appear frequently in the form of dense clumps with occasional isolated organisms. One difference in arrangement that serves to distinguish Gram-negative from Gram-positive diplococci is the way in which the main axis of the oval is directed, with the Gram-negative cocci this axis is always at right angles to the axis joining the two cocci with the Gram-positive cocci it is often coincident with it. In other words, pairs of Gram-negative cocci are usually compressed laterally, Gram-positive cocci often longitudinally. As a rule the Gram-negative cocci are decolorized without difficulty, but some members tend to retain the gentian violet, and so take on an indeterminate colour, which is most confusing. The most notable example of this is Jaeger's coccus or the *Diplococcus crassus*, in a single film Gram-positive and Gram-negative, and numerous other cocci with an indeterminate colour are found living side by side. One reason for the indeterminate staining of some of the Gram-negative cocci is the tendency they have to



FIG. 106.—*Neisseria meningitidis*.

From a serum agar slope culture 4 days 37° C.  
( $\times 1000$ ).

be arranged in groups or dense clumps. Once these have been stained with gentian violet, they are not easily decolorized. Hence it is important always to make films as thin and as uniform as possible. In young cultures the cocci stain fairly evenly, but after about 24 hours autolytic changes set in, with the result that so-called involution forms appear, these are generally large swollen cocci which stain poorly. Both the meningococcus and the gonococcus are characterized as compared with most other neisseriae, by the frequency with which such forms appear, and it may be noted that many of the large swollen forms, in cultures of these species, stain deeply and uni-

formly. J. E. Gordon (1921) described a variety of *N. catarrhalis* in which the degenerative forms began to appear after the 4th hour, but this is unusually early. Some workers have described the presence of Babes-Ernst bodies or melanochromatic granules in members of this group. Elsea and Huxton (1904) state that the meningococcus, when stained with Loeffler's methylene blue often shows a brightly stained central spot, whilst the remainder of the cell is scarcely coloured. With Neisser's stain the granules stain bluish black, the cell body brown. Marx and Worth (1900) found these granules in gonococci, but only in organisms taken from the fluid stage of gonorrhea, they state that the whole cell may appear filled with granules. Capsules are demonstrable in some freshly isolated strains of meningococci (Clapp *et al.* 1935) and in the organism known as *Diplococcus mucosus*.

**Growth Requirements.**—Culturally many of the Gram-negative cocci are characterized by a reluctance to grow on ordinary media especially fluid media. Most of the nasopharyngeal cocci will grow—though often poorly—on nutrient agar,

but the meningococcus will not do so, the addition however, of a small quantity of blood or serum is sufficient to enable growth to occur. The most fastidious is the gonococcus, to cultivate this organism a great variety of media have been devised, the majority depending on the addition of some natural protein such as blood, serum, ascitic fluid, or hydrocele fluid to a basis of nutrient agar. The meningococcus and the gonococcus are not always easy to maintain in culture even though transplants are made every 2 or 3 days the organisms not infrequently die out, and the strains are lost.

All the members of the group are aerobic, little or no growth occurs under strictly anaerobic conditions. Growth of the gonococcus is favoured by the addition of cystine or other source of —SH bodies (McLeod *et al* 1927, Boor 1942). Many workers have stated that the meningococcus and the gonococcus grow best under a lowered oxygen pressure and that their growth is improved by 10 per cent.



FIG 107—*Neisseria gonorrhoea*

From a blood agar slope culture 24 hours  
37° C ( $\times 1000$ )



FIG 108—*Neisseria pharyngis*

From a blood agar slope culture 24 hours  
37° C ( $\times 1000$ )

CO<sub>2</sub> (Wherry and Oliver 1916 Chapin 1918 Kohman 1919, Ruediger 1919 Rock well and McKhann 1921) but numerous other workers have failed to substantiate this (Cook and Stafford 1921 Erickson and Albert 1922 Torrey and Buckell 1922a)

When making experiments on the effect of altering the gaseous constitution of the atmosphere it is very difficult to control all the factors concerned the technique used may, for instance change the moisture content of the atmosphere and the rate of evaporation from the medium. The presence of 10 per cent CO<sub>2</sub> alters the H ion concentration and will interfere with the change in the reaction of the medium that normally occurs during growth, thus CO<sub>2</sub> may be beneficial if the medium has been made too alkaline and it may by its buffering action prevent the accumulation of acid. The failure to standardize these secondary factors is probably sufficient to explain the diverse results obtained by different workers.

The work of McLeod and his colleagues (1934), however does suggest that growth is improved by the addition of 10 per cent CO<sub>2</sub>, particularly that of freshly isolated strains. Glucose and glycerine have little or no beneficial effect, peptone in a 1-3 per cent. concentration seems to be favourable. The optimum H ion concentration for growth is about pH 7.4-7.6, the limits within which growth

will occur are comparatively narrow, but they depend largely on the constitution of the medium. The optimum temperature for all the members is 37° C.; some of them, including the meningococcus and the gonococcus, will not grow at all below 20° C., many of the nasopharyngeal cocci will grow at 22° C., but not always on first isolation. The meningococcus forms a weak hæmolyisin, reaching its maximum in trypticar cultures in about 4 days.

Some of the members—the *pharyngis faræ* group—produce a greenish yellow pigment on solid media, and occasionally a Gram-negative coccus is met with that forms a bright yellow pigment. Some of the Gram negative cocci, particularly the gonococcus and the meningococcus, contain an active autolysin, which is destroyed by heating to 65° C. for half an hour.

**Cultural Characteristics.**—The colonial appearances of all the Gram negative cocci appear to be subject to considerable variation. Two different types of colony of both the meningococcus and the gonococcus have been described (Wassermann 1898, Lipschütz 1904, Atkin 1923, 1925, Cohn 1923); and S. P. Wilson (1923) and G. S. Wilson and Smith (1923) have observed and studied rough and smooth types of numerous nasopharyngeal cocci. In fluid media—broth and serum broth—growth is generally poor, and takes the form of a slight turbidity and a finely granular deposit, which disintegrates hardly at all on shaking; occasionally growth occurs on the surface.

**Resistance.**—The resistance of the Gram negative cocci to inimical agencies is very low. In culture most of them die out in a few days; though if the organisms are seeded into ascitic agar stab tubes—preferably made up with 0.75 per cent. agar—prevented from drying, and kept in the incubator at 37° C., they may live for weeks or even months. Though it is not known with certainty why the Gram-negative cocci die out in culture so quickly, it appears probable that they are killed by the amount of alkali produced; the production of  $\text{NH}_3$  and of alkaline carbonates of organic acids may apparently lower the H ion concentration of the medium to pH 8.6–9.0, and thus bring about the death of the organisms (Phelon *et al* 1927). The meningococcus and the gonococcus are killed by heating to 55° C. in 5 minutes or less, they are very susceptible to desiccation, death occurring usually within an hour or two. Weak disinfectants, such as 1 per cent. phenol or 0.1 per cent.  $\text{HgCl}_2$ , prove fatal in 1 or 2 minutes. The meningococcus and the gonococcus are both sensitive to the sulphonamides and to penicillin.

**Biochemical Reactions.**—Biochemically the members of the group are not very active, the production of acid in glucose, maltose, and sucrose is used as a means of classification. Other sugars, such as galactose, levulose, and dextrin, are used by some workers, but those who have had most experience agree that they are unsatisfactory. Since many species of *Neisseria* will not grow on the ordinary peptone water-sugar medium, it is necessary in testing their sugar reactions to add a small amount of serum, or to grow them on ascitic fluid agar containing litmus and 1 per cent. of the sugar. If serum is used, human or rabbit serum should be chosen, since horse, sheep and ox serum contain maltase, which may lead to a false reaction in the presence of maltose (Roemer 1936, Hendry 1938). Litmus milk is unaltered, except by the *Diplorococcus crassus*, which turns it acid. Indole is not produced. The methyl red test is weakly positive or frankly negative, according to whether or not the organism tested produces acid from glucose; as the increase in H-ion concentration is rarely greater than to pH 6.0, the red colour



developed with methyl red is usually faint. The Voges Proskauer reaction is negative. Nitrates are not reduced. Catalase is produced and all the members that have been tested give the oxidase reaction described by Gordon and MLeod (1928).

**Antigenic Structure**—Most attention has been concentrated on the meningococcus and the gonococcus. The meningococcus has been divided into four antigenic types—Types I, II, III and IV (see pp 538-9) but the results obtained depend largely on the source from which the strains are obtained. In epidemic times most of the cocci isolated can be readily typed but strains isolated from sporadic cases in non epidemic times are frequently inagglutinable with any of the type sera. The gonococcus is even more irregular; clear types are difficult to establish. The majority of the strains appear to be related antigenically and to fall more or less into one or other of two groups (Atkin 1925) (see pp 515-7). Little work has been done on the other Gram negative cocci; one of the chief reasons for this is that most of them are auto agglutinable and homogeneous suspensions cannot be obtained. The complement-fixation test however seems to show that there is a group relationship between *N. catarrhalis* & *N. pharyngis*, the gonococcus and the meningococcus (see Oliver 1929, Price 1933).

Studies on the chemical fractionation of these organisms are still in their infancy. Boor and Miller (1931) and Miller and Boor (1934) amplifying the work of Zozaya (1931) and Zozaya and Wood (1932) (see p 539) have extracted nucleoproteins and polysaccharides from various members of the group. By the precipitation reaction it was found that the nucleoproteins from the meningococcus, the gonococcus and *N. catarrhalis* not only resembled each other closely but also had an affinity with nucleoproteins extracted from pneumococci. Polysaccharides prepared from the meningococcus and the gonococcus reacted in high dilution with Type III antipneumococcal serum as well as with antimeningococcal and antigonococcal serum. The polysaccharide extracted from *N. catarrhalis* reacted with antigonococcal but not with antimeningococcal serum.

**Pathogenicity**—The meningococcus gives rise to rhinopharyngitis to epidemic cerebrospinal meningitis and to post basic meningitis in children. By intraspinal injection of monkeys it is possible to produce a meningitis with pure cultures of the organism. The gonococcus gives rise in human beings to gonorrhoea with all its complications but it is impossible to reproduce this disease in animals.

Towards laboratory animals all the Gram negative cocci behave in much the same way. Injected intraperitoneally in large doses into mice or guinea pigs they cause death in 1 to 3 days. Post mortem there is a small amount of peritoneal exudate and sometimes a little fibrin deposit on the organs; the spleen is slightly enlarged and there is hyperaemia and degeneration of the viscera. The organisms can be cultivated from the peritoneal exudate but rarely from the heart's blood. There is little multiplication of organisms inside the body; no true infection is set up and death occurs from toxæmia. A similar result follows the injection of heat killed organisms though generally a rather larger dose is needed than of living cocci. It seems probable that the toxicity is due to some constituent of the nucleoprotein since nucleoprotein extracted from meningococci and gonococci is almost as toxic to mice as are the dead organisms themselves (Boor and Miller 1934).

## CLASSIFICATION

The Gram negative cocci as a group, have been studied so little that it is impossible to lay down any satisfactory basis for classification. Apart from the meningococcus and the gonococcus, the definition of the different species is far from clear. This is due largely to the fact that the colonial appearances are subject to such great variation that the descriptions given of apparently the same species by different workers are often quite contradictory. The cultural descriptions, for example, of *N. catarrhalis* are most varied (Ghon and Pfeiffer 1902, Dunn and Gordon M. H., 1903, von Lintelsheim 1903, Arkwright 1907, Gurd 1908, Elser and Huntoon 1909, Martin 1911, Netter and Debre 1911, Doptier 1921, Gordon, J. E., 1921), and the only sound basis for identification of this organism appears to be its failure to ferment any sugars. Again, in Germany and America several chromogenic species have been described, forming a greenish-yellow pigment, producing acid in glucose, maltose, and sometimes sucrose, and generally giving a smooth type of colony. In this country a large number of Gram negative cocci have been isolated from the nasopharynx giving the same sugar reactions but quite devoid of pigment. Further it has been found that the



FIG 109—*Neisseria meningitidis*  
Surface colonies on serum agar 48  
hours, 37° C. (× 8).



FIG 110—*Neisseria meningitidis*  
Surface colony on serum agar 7 days,  
37° C. (× 8).

colonies formed are sometimes smooth and sometimes rough, and that an organism which gives a smooth colony on isolation may subsequently give a rough type of colony. The differentiation of those organisms giving rough colonies from *N. pharyngis sicca* is in our experience frankly impossible (Wilson, S. P., 1923, Wilson, G. S. and Smith 1923).

At the moment, therefore, it must be confessed that our ignorance is too great to allow of any satisfactory classification. For provisional purposes the classification on sugar reactions may be used, but this is subject to severe limitations. Briefly, it can be said that *N. catarrhalis* ferments no sugars, the gonococcus ferments glucose, and the meningococcus glucose and maltose, the other nasopharyngeal cocci give varied reactions, some being like the meningococcus and others also fermenting sucrose. The *Diplococcus crassus* can be differentiated by its fermentation of lactose. When first isolated from the body the fermentative reactions of the Gram negative cocci may be irregular. Navarro (1917) for example, found that quite a number of meningococci from the cerebrospinal fluid of children with meningitis failed on first isolation to ferment

either glucose or maltose. We ourselves have isolated organisms from the nasopharynx which fermented maltose, but not glucose. Other workers have observed similar irregularities in the behaviour of this group.

We append a detailed description of *N. meningitidis* and *N. gonorrhoea*, together with some further notes on their differentiation, descriptions of those Gram negative cocci which have received specific names, and a table giving particulars of the main differential criteria that have been relied upon by different workers in subdividing this group. We would add our personal opinion that there is, at present, little justification for the recognition of separate species among the Gram negative cocci of the normal nasopharynx, with the possible exception of *N. catarrhalis*. We should, ourselves, combine the remaining types into a single species, with some appropriate name such as *N. pharyngis*, which we might define as follows.

*Neisseria pharyngis*.—Non motile, Gram negative diplococcus, arranged sometimes in tetrads and often in dense clumps. Grows on agar, giving rise to either rough or smooth colonies, which are generally coherent, tenacious, membranous, and friable—are difficult to emulsify, and are auto-agglutinable when suspended in saline. Grows in serum broth with the production of little or no turbidity, as a rule, and a coarsely granular sediment not disintegrating completely on shaking; a surface ring growth is not infrequently formed, particularly by the rough variants. A yellow, golden yellow, or greenish-yellow pigment may be produced, but is variable in its appearance. The sugar reactions are subject to variation; glucose, maltose, or sucrose may be fermented with the production of acid. Aerobic, will not grow under strictly anaerobic conditions. Growth is best at 37° C, but will generally occur at 23° C. Non pathogenic on subcutaneous injection into mice, large doses intraperitoneally may cause death from toxæmia. The species is subject to great variation in colonial appearance and, apart from the smooth and rough types, a smooth variant may occur that is of butyrous consistency and easy to emulsify and also a mucoid variant containing capsulated diplococci (Wilson, G S and Smith 1928).

### The Meningococcus

**Cultural Characters**.—The meningococcus generally gives rise to a smooth typically lenticular colony. Atkin (1923), however, and more recently Rake (1933), have demonstrated the existence of colonial variants. The appearance of the colony depends on the nature of the medium, the age of the strain, and the antigenic type of the organisms. Freshly isolated strains of Group I generally form smooth colonies, which may be mucoid if the organisms are capsulated; on incubation for some days their edge may become crenated or dentate and secondary papillae may appear on the surface. Organisms of Group II tend to form rather smaller colonies, and may assume a deep yellowish tint on suitable media. Rough colonies, which are generally smaller than those of the smooth form, often appear in strains subjected to laboratory cultivation.

On primary isolation the meningococcus must be provided with such accessory growth factors as are present in blood, serum, milk, and other animal fluids, and in certain vegetable extracts (Lloyd 1916-17). After a few generations on such an enriched medium, it may sometimes be brought to grow on what are described as ordinary culture media, but its vitality under these conditions is uncertain (Murray 1929). Growth is usually favoured by the presence of 5 to 10 per cent. CO<sub>2</sub>. For preservation the meningococcus should be frozen and dried. If this is impossible, it should be maintained in a citric fluid agar stab or on Dorset egg slopes; the tubes should be corked to prevent evaporation and kept in the incu-

bator To conserve the virulence of the organisms they should be subcultured every two days on blood agar slopes, or preferably frozen and dried.

The meningococcus undergoes rapid autolysis this is responsible for the swelling and loss of staining properties in cultures more than a few hours old. This property is destroyed by heating to 60° C for 30 minutes. If the organisms are suspended in saline covered with toluol to prevent contamination and incubated at 37° C autolysis is said to be nearly complete in 4 hours (Flexner 1907a) For this reason all suspensions intended for agglutination should be inactivated by heat

**Antigenic Structure**—Soon after the agglutination test was introduced for the identification of meningococci it was noticed that different strains possessed varying degrees of agglutinability Kutscher (1906) who employed the absorption test observed that there was a marked difference between strains isolated from different sources, but he was unable to classify them by this method In 1909 Elser and Huntoon found that 40 per cent. of meningococci were inagglutinable by a monovalent serum, and that these inagglutinable strains, which they term pseudo-meningococci, exhibited a reduced absorption capacity they further divided the pseudo-meningococci by absorption into two sub-groups. In the same year Dopfer (1909) noticed the presence in nasopharyngeal mucus of cocci resembling the meningococcus in morphology cultural and fermentation reactions, but differing from it in their complete absence of agglutination with a meningococcal serum these organisms he termed parameningococci. Arkwright also in 1909 studied 25 strains of meningococci from cases occurring in epidemic areas, and 20 strains from sporadic cases he noticed not only that by agglutination and absorption the organisms could be roughly divided into groups, but that, serologically the sporadic strains tended to deviate more from the type to which most strains conformed than did the epidemic strains. In 1914 Dopfer and Pauron divided the parameningococci into 3 types,  $\alpha$ ,  $\beta$  and  $\gamma$ . Soon after the commencement of the War Ellis (1915) examined 46 strains from 6 epidemic foci, and found that they fell by agglutination into 2 types, I and II of which Type II was probably identical with Dopfer's parameningococcus. Simultaneously Arkwright (1915) was able to classify 30 out of 35 strains from epidemic cases into 2 main groups, Types I and II of which Type II like Ellis's Type II corresponded to Dopfer's parameningococcus of the remaining 5 strains, 3 were difficult to classify by agglutination, and 2 were intermediate between the two types. Gordon and Murray (1915) by using the absorption test found that 32 strains from the cerebrospinal fluid of epidemic cases fell sharply into 4 groups, which they called Groups I, II, III and IV none of these groups, however showed any relation to Dopfer's parameningococcus. In 1917 Nicolle Debaens and Jouan (1918), using the agglutination test alone were able to classify the meningococci into 4 types, called A B C and D Gordon and Murray's Type I and III strains, as they are now generally referred to, corresponded to Dopfer's Meningococcus and to Nicolle Debaens and Jouan's Type A and their Type II and IV strains to Nicolle Debaens and Jouan's Type B F Griffith (1917) working at the Local Government Board laboratories was able to divide his meningococci into two main groups by simple agglutination, Groups I and II his Group I corresponded roughly with Gordon and Murray's Types I and III and his Group II with their Types II and IV Scott (1917) similarly found that his strains fell into two groups.

Since 1918 observations particularly in the United States, have served to show that no sharp line of demarcation can be drawn between different types of meningococci. Branham Taft and Carlin (1931) and Branham (1932) it is true were able to assign every one of 291 strains of meningococci isolated during a time of epidemic prevalence to one or other of Gordon and Murray's four types, but this was possible only after prolonged study involving examination of their agglutinability their power to absorb agglutinins, and their agglutinogenic capacity. The lack of strict type specificity and the readiness with which many strains

undergo antigenic degradation, render classification by such means arbitrary and unconvincing. A change in the strains used for the preparation of typing sera can easily result in an apparent change in the type of organism under study.

Further observations by Branham and Carlin (1937) and others have led to the broad conclusion that two main groups can be distinguished by agglutination—Group I, which is mainly responsible for epidemic cases and tends to be antigenically homogeneous, and Group II which is mainly responsible for sporadic cases and tends to be antigenically heterogeneous. This concept receives support from other methods of study such as chemical fractionation, and precipitation and capsular swelling reactions.

The complement fixation reaction has been used for classifying meningococci. Nicolle, Debains and Jouan (1918) found this reaction less specific than that of agglutination whereas Bell (see Report 1920) and Butterfield and Neill (1920) regarded it as more specific. Evans (1920) studied the opsonin reactions of antimeningococcal serum. By this means she found that 63 strains fell sharply into 4 groups, there were 4 atypical strains. A fifth group could also be demonstrated, which was closely related to the other four, its members were able to effect a partial absorption of the sera prepared against strains of the other groups.

By chemical fractionation Rake and Scherp (1933a, b) have separated three fractions from meningococci. There is a carbohydrate or "C" substance common to all meningococci and to some other micro organisms, which is probably the same as that described by Zozaya (1931) and Zozaya and Wood (1932). There is a protein or "P" substance, which is also found in gonococci and Type III pneumococci. The third fraction is a sodium salt of a polysaccharide acid (Scherp and Rake 1935) and is responsible for the specificity of Types I and III strains. More recently Menzel and Rake (1942) have brought evidence to show that the specificity of Type II strains is determined by a protein substance. There appears to be no difference between the type specific polysaccharide found in Types I and III.

The presence of the polysaccharide is generally demonstrated by the precipitation reaction, using as antigen a specially prepared extract of the organisms. Petrie (1932) however, has described a simple alternative method. It consists in growing the organisms on agar plates containing the homologous immune serum. Characteristic haloes develop around the colonies. These consist of a precipitate formed by the interaction of the specific polysaccharide, which has diffused out into the medium, with the homologous antibody.

It seems probable that the specific polysaccharide is also responsible for the capsular swelling (Quellung) reaction demonstrated by Clapp, Phillips and Stahl (1935) in smooth Group I strains. A similar reaction with Group II strains was at first thought not to occur, but further observations by Cohen (1940) have shown that certain strains may exhibit capsular swelling and give a typical halo reaction in the presence of serum made with a capsulated Group II strain. Branham and Carlin (1942) have likewise described a separate subgroup of Group II strains, referred to as Group II alpha which are capsulated, give a Quellung reaction, and are strongly antigenic, stimulating the production of antibodies with a specific protective action on mice.

**Pathogenicity**—Mick.—In his original communication Weichselbaum (1887) observes that subcutaneous injection into mice is without effect. Injected intrapleurally with 0.5 ml of a thick suspension from a 24 hours' agar culture the mouse becomes ill, develops paralysis

of the hind limbs, and dies in 1 to 2 days. Post mortem, there is a viscid, often hæmorrhagic, fluid in both pleural cavities, the lungs are hyperæmic in places and may be covered with a false membrane, the spleen is generally enlarged and congested. The organisms are present in enormous numbers in the pus cells of the pleural exudate, and often but not always in the blood and the spleen, in both of which situations they remain mostly extra cellular.

*Intrapertoneal injection* with the same dose kills the mouse in 18 to 24 hours. Post mortem, there is a small amount of sticky fluid in the peritoneal cavity, containing pus cells, the spleen is generally swollen and congested, cocci are found in varying numbers in the exudate, and in small numbers in the spleen and heart's blood.

**GUINEA PIGS**—Weight for weight these animals are said to be somewhat more susceptible than mice (Rist and Paris 1904) but here again subcutaneous injection even of massive doses, fails to give rise to a general infection, at most a small abscess is produced.

*Intrapleural injection* of 1 ml. of a thick suspension of a young agar culture causes death in 1 to 3 days. Post mortem, there is a thick exudate poor in fibrin in both pleural cavities, the lungs are thickened and dark red, the spleen is not enlarged. Cocci are found in the exudate but are generally absent from the spleen and heart's blood.

*Intrapertoneal injection* causes death in 1 to 3 days. Post mortem, there is an exudate, clear or turbid in the peritoneal cavity, on the rolled up omentum and on the anterior surface of the liver there is a deposit of fibrin and pus, there are hæmorrhages into the mesentery and into the visceral and parietal peritoneum, the adrenals are vividly congested and may be hæmorrhagic, the pancreas and surrounding tissues are oedematous. In the pleural cavities there is often an exudate of clear fluid, the lymphatic glands are swollen and congested. Cocci are found in moderate numbers in the peritoneal exudation but are absent from the blood and viscera.

*Sub-dural injection* causes death in 20 to 24 hours, post mortem there is oedema and congestion of the meninges, with pus at the site of injection, there is a large amount of clear fluid in the peritoneum free from cocci (Albrecht and Ghon 1901). There is no multiplication of organisms in the spinal fluid itself.

**RABBITS**—Subcutaneous, intrapleural and intrapertoneal injections are generally without effect.

*Intravenous injection* with 1-4 ml. of a thick suspension of a young agar culture kills the animals in 1 to 4 days. Post mortem, apart sometimes from a few areas of congestion in the lungs there is nothing abnormal to be found. No cocci are present in the blood stream.

*Sub-dural injection* into the skull occasionally causes death. Post mortem there is congestion of the meninges, the cocci may be recovered in culture (Weichselbaum 1887).

*Intracisternal injection* by the sub-occipital route is stated to give rise to cerebrospinal meningitis (Branham and Lillie 1932, Zdrodowski and Voronine 1932). To achieve success virulent cultures and young rabbits (1300-1500 gm.) are desirable. The disease may be acute and prove fatal in 24 hours, or sub-acute and cause death between the 2nd and 7th days. Clinically rigidity of the neck, retraction of the head, spasticity, and sensitive ness to touch, or progressive paralysis may be noted. At necropsy the brain and cord are markedly hyperæmic and are covered with a thin layer of purulent exudate. The cerebrospinal fluid may be almost clear turbid or frankly purulent. Meningococci can be recovered from the spinal fluid and usually also from the blood.

**DOGS**—Weichselbaum (1887) stated that he had succeeded by sub-dural injection, in producing a pachy and lepto-meningitis with acute encephalitis, death occurred from a few hours to the 12th day after inoculation.

**MONKEYS**—Von Lingelsheim (1903) was apparently the first to reproduce the disease in monkeys. After intraspinal injection one monkey became ill in 6 hours, there was retraction of the head and opisthotonos, and death took place in 30 hours. At necropsy

the pia mater was turbid along the vessels with here and there small collections of pus. Meningococci were found in the pus and in the blood.

Flexner (1907b) made a number of experiments on monkeys mostly *Macacus rhesus*. After intraspinal injection of  $\frac{1}{2}$  to 1 agar slope he found that the monkeys became generally weak and apathetic, the head drooped so as to touch the floor of the cage, occasionally however it was retracted, death occurred in 18 hours to 4 days as a rule and was not infrequently preceded by general convulsions. Post mortem the chief lesions were lepto meningitis particularly at the base of the brain, encephalitis and abscesses, hemorrhages into the pia, inflammation of the dorsal root ganglia and acute endarteritis of the vessels. The inflammation of the meninges extended into the membranes covering the olfactory lobes and along the dura mater into the ethmoid plate and nasal mucosa, which was often inflamed and beset with hemorrhages. Diplococci were found in the meningeal exudate and in the nasal mucosa but were not cultivated from the latter situation. They were also present in the sero-purulent fluid in the ventricles.

By giving small repeated doses Flexner succeeded in setting up a chronic meningitis lasting for several weeks. Post mortem there was abundant exudate rich in meningococci, the foramen of Magendie was closed and hydrocephalus and pyrocephalus with ependymitis and dilatation of the ventricles were found. Neither in acute nor chronic cases did the internal organs apart from the central nervous system show any marked changes. Occasionally the organism is found in the blood stream.

Flexner found that not all the monkeys developed the disease. Those that did so generally died within 2 days, or else recovered after a severe illness. From his experiments it is clear that the disease is more acute in monkeys than in man.

McDonald (1908) confirmed Flexner's results in monkeys. Though unable to infect *rhesus* monkeys he obtained successes with *Callithrix* by subdural inoculation of cerebrospinal fluid from human cases of disease.

The experimental lesions which we have discussed are not produced with the regularity that one might expect partly because the meningococci themselves vary considerably in virulence and partly because the susceptibility of different animals—even of the same species—varies within a wide range.

The variations in virulence of the meningococcus depend on the source of origin of the strain, the length of time it has been isolated, the age of the subculture, the nature of the medium (Murray and Ayrton 1924) and doubtless on other factors. When freshly isolated some strains are of sufficient virulence to kill mice inoculated intraperitoneally in a dose of about 100,000 organisms but others are far less virulent. It has however been shown by Miller (1933, 1934–35) that if the culture is suspended in a solution of gastric mucin as few as 2 to 10 organisms of a highly virulent strain suffice to kill a mouse. By frequent mouse passage using organisms suspended in mucin a given strain may be kept at its maximum degree of virulence. Alternatively it should be frozen and dried.

**Toxin Production.**—Flexner (1907a) pointed out that one of the earliest results of intraperitoneal inoculation into guinea pigs was a marked fall of temperature. This is not the normal course in an acute infection and he was led to conclude that the animals died from the effects of a poison liberated from the bodies of the organisms. Albrecht and Ghon (1901) found that filtered cultures were without effect on mice but that 24 hour cultures heated to 65° C for 1 hour when injected intraperitoneally into mice produced death with the same picture as that found after injection of living cocci. Subsequent observers have found that the dose of dead and of living organisms necessary to kill mice is practically identical. Thus Neill and Taft (1920) found that 4,000 million living organisms injected intraperitoneally killed 6 out of 10 mice whereas the same dose of dead cocci killed 5 out of 10. M. H. Gordon

(see Report 1920) showed that when freshly isolated the living cocci were fatal in smaller doses than the dead cocci but after subculture for some time in the laboratory, the lethal doses tended to approximate Petrie (1937), who has made a careful study of the endotoxin, finds that its effect on animals can be reproduced by injections of other Gram negative cocci, and that it belongs to a group of non specific, non antigenic, thermostable, bacterial poisons.

From these results, and from the fact that in animals dying from injection of living cocci the blood and viscera are frequently sterile, it would appear that the main cause of death is a toxæmia Flexner (1907a) found that meningococci undergo very rapid autolysis in culture, as a result of this the endotoxins are liberated from the bodies of the organisms, and it is these which are responsible for the pathogenic effects in animals This view is substantiated by the frequent occurrence of hæmorrhages on the serous membranes, of sterile transudates in the cavities of the body, and of the adrenal hæmorrhages which are found both in animals and in human beings dying from the disease (MacLagan and Cooke 1917, Petrie 1937)

The endotoxin can be extracted from the bodies of the meningococci One of the simplest means (M H Gordon see Report 1920) is to grind 0.05 gm. of dried cocci in an agate mortar with 1.25 ml. of distilled water, to which after a few minutes 1.25 ml. of N/20 NaOH are added, the grinding is continued for about a minute. The cocci pass into solution on the addition of the alkali. The M.L.D. of the fluid thus obtained is generally 0.1 to 0.15 ml.—that is an amount corresponding to about 2 mgm. of the dried cocci (see also Petrie 1937)

Though most workers have regarded the toxin of the meningococcus as essentially an endotoxin there seems to be little doubt that under certain conditions it can readily diffuse out into the medium According to Ferry, Norton, and Steele (1931), hormone broth cultures of pH 6.6 incubated for 4–6 days contain a filtrable toxin specific for each of the serological types of meningococci, as well as a group specific toxin common to all four types Ferry and Schornack (1934) and Maegraith (1935) have shown that these toxins, when inoculated by the intracisternal route into guinea pigs, give rise to convulsions and death within 24 hours. It would serve no useful purpose to discuss how much of the toxic activity of these filtrates is due to substances secreted by the living organisms and how much to substances liberated from the dead organisms There is evidence that the polysaccharide found in Types I and III is soluble in suitable media (Petrie 1932, Kirkbride and Cohen 1934), and it may be that the nucleoprotein to which Boor and Miller (1934) ascribe the toxicity of the meningococci is likewise soluble to a greater or less degree The main conclusion is that for laboratory animals dead cocci are almost as fatal as living cocci and that the tissue reactions are determined by toxic substances liberated from the organisms either before or after their death (For a review of the meningococcus see Branham 1940)

### *Neisseria meningitidis*

*Synonyms*.—*Meningococcus*, *Diplococcus intracellularis meningitidis* of Weichselbaum.

*Isolation*.—From cerebrospinal fluid of patients with meningitis by Weichselbaum in 1887

*Habitat*.—Strict parasite, found in nasopharynx of man.



**Morphology**—Oval or spherical cocci,  $0.8 \times 0.6 \mu$ , often arranged in pairs, with adjacent sides flattened, long axis of oval lies at right angles to axis joining the two cocci. In cultures great variation in size and in depth of staining occurs, due to autolysis in the body the cocci are more regular and are generally intracellular. Non motile; non capsulated. Gram negative.

**Serum Agar Plate**—24 hours,  $37^{\circ}\text{C}$ . Round convex, bluish grey, translucent, amorphous colonies, 1 mm in diameter, with smooth, moist, glistening surface and entire edge, consistency butyrous, easily emulsifiable. Colony is typically lenticular. Later, colonies increase in size, become more yellow and opaque, and may show a granular centre, and a radiate periphery.

**Serum Agar Slope**—24 hours  $37^{\circ}\text{C}$ . Moderate, partly confluent, raised, greyish yellow growth with smooth or irregular surface due to imperfect fusion of colonies. Edge is undulate or made up of single colonies.

**Gelatin Slab**—No growth.

**Serum Broth**—24 hours,  $37^{\circ}\text{C}$ . Poor to moderate turbidity with slight granular or viscous deposit. No surface growth.

**Resistance**—Highly susceptible to inimical agencies. When dried, and kept at room temperature, cocci die in under 3 hours. Killed by moist heat at  $55^{\circ}\text{C}$ . in less than 5 minutes. Killed by 1 per cent phenol in 1 minute and by 0.1 per cent  $\text{HgCl}_2$  almost instantaneously. Sealed cultures kept at  $37^{\circ}\text{C}$  often live for 4 or 5 weeks, and occasionally for 2 or 3 months but when kept at room temperature they generally die in a few days.

**Metabolism**—Optimum H ion concentration is pH 7.4–7.6. Optimum temperature for growth is  $37^{\circ}\text{C}$ ., little or no growth below  $30^{\circ}\text{C}$ . Fails to grow on plain nutrient agar, but grows on tryptic agar, glucose agar, and agar to which blood serum or ascitic fluid has been added. Some strains show a slight formation of yellow pigment. Aerobe, no growth under strictly anaerobic conditions, growth favoured by 5 to 10 per cent  $\text{CO}_2$ . Produces a weak hemolysin.

**Biochemical**—Produces acid, no gas, in glucose and maltose. No change in litmus milk. Catalase +, methylene blue reduction +. MR — or weak +, VP —, indole —,  $\text{H}_2\text{S}$  —.

**Antigenic Structure**—Divided by agglutination and absorption of agglutinins into two main groups, Groups I and II. Some workers divide Group I into Types I and III and Group II into Types II and IV. Group II tends to be more heterogeneous than Group I.

**Pathogenicity**—Responsible for sporadic and epidemic cerebrospinal meningitis in man. Experimentally, it is pathogenic to mice guinea pigs and rabbits, if injected intraperitoneally in fairly large doses, causes death by toxæmia in 1 to 4 days. There is little or no multiplication of the organisms in the body.

### The Gonococcus

**Cultural Characters**—In culture the isolated cocci are round. According to Neisser (1882), as the spherical coccus grows, it becomes oval, division occurs, and two cocci are formed, which cling closely together. These then separate a little, and each one grows and divides again, but in a plane at right angles to that of the first division, so that tetrads result. Each member of the tetrad divides in the same plane as that of the first division. The result is that four pairs of cocci are formed.

The gonococcus is the most difficult member of the group to cultivate. It was first grown by Leistikow and Loeffler (Leistikow 1882) on blood serum gelatin at  $37^{\circ}\text{C}$ ., and by Bumm (1885a) first on coagulated bovine or sheep serum at  $30^{\circ}$ – $34^{\circ}\text{C}$ ., and later (Bumm 1885b) with more success on coagulated human serum. Since then

a host of other media have been introduced reference may be made to a few (Wertheim 1891 Kiefer 1890 Wassermann 189 Thalmann 1900 Lipschutz 1904 Martin 1911 Vedder 1915 Hall 1916 Cole and Lloyd 1917 Thomson 1917 Clark 1920 Swartz and Davis 1920 Buschke and Langer 1921 Cook and Safford 1921 Jenkins 1921 1922 Costa and Bover 1922 Erickson and Albert 1922 Kandiba 1922 Lorentz 1922, Torrey and Buckell 1922a Torrey *et al* 1922 Macnaughton 1923 Lebœuf 1924 Gordon, J 1926)

According to Sordella, Miravent and Negroni (1927) an excellent medium results from adding to nutrient agar 1 per cent. of liver extract. This extract is prepared by macerating ox liver in 5 per cent. NaCl solution for 2-4 hours at 45° C., raising the temperature gently to 60 C., keeping it at 60 C. for 10 minutes, and filtering first through paper then through a Berkefeld candle. The liver extract, if kept in the ice-chest remains potent for months.

As the result of long experience in routine cultivation, McLeod and his colleagues (1934) recommend the use of 10 per cent. heated blood agar of pH 7.4 prepared from broth in which the extraction of the meat has been carried out by Wright's (1933) method. The minimum amount of agar consistent with stability



FIG 111.—*Neisseria gonorrhoea*.  
Surface colonies on serum agar 4 hours,  
37° C. ( $\times 5$ ).



FIG 112.—*Neisseria gonorrhoea*.  
Surface colony on serum agar 5 days,  
37° C. ( $\times 5$ ).

is used. The cultures are incubated in air containing 5 per cent  $\text{CO}_2$ . For ordinary purposes a satisfactory medium is provided by nutrient agar containing 10 per cent ascitic or hydrocele fluid and having a pH of 7.6 the slopes of plates should be moist and the air in the incubator should be saturated with water vapour. Stock cultures are best kept in a similar medium containing 0.5 per cent agar and put up in the form of slabs these tubes should be corked and kept in the incubator. Media with a high amino-acid concentration are not usually satisfactory (Torrey and Buckell 1922a Gordon, J and McLeod 1926) but the addition of —SH groups is beneficial (Boor 1942). Glucose does not improve growth.

Numerous workers (de Christmas 1894 Wassermann 1898 Lipschutz 1904 Gurd 1908 Martin 1911 Cohn 1923) have noticed that the cultural characters of the gonococcus are subject to variation. More recently Atkin (1925) has studied this phenomenon and found that, as with the meningococcus, there is a definite correlation between the serological type of the organism and its colonial appearance. By growing gonococci on thick trypticase plates of pH 7.5 he observed two different types of colony. Type I gave a large irregularly round flattened, translucent colony with an undulate edge and a surface that in 5 days or so

became covered with papillæ, Type II gave a smaller, round low convex or raised, yellowish white, opaque colony, with a slightly uneven surface and an entire or faintly lobate edge, no papillæ were formed. Serologically, colonies of Type I could be differentiated from those of Type II. Atkin found that the organisms in the papillæ of Type I colonies lived about twice as long as did those in the flat part; he therefore concludes that the papillæ arise as a reaction to an unfavourable environment, and represent the first step in the change over to Type II. In some colonies the papillæ actually fuse together, so that they constitute the entire colony, the papillated appearance is thus lost. During the course of subculture in the laboratory, there will be a constant selection of the longer living organisms in the papillæ, so that eventually the strain will consist entirely of these organisms, when this process is complete, the strain will belong culturally and probably serologically, to Type II. Type I colonies are usually observed when pus from acute gonorrhœa is plated out, Type II colonies are usually found in old laboratory cultures, or occasionally in chronic gonorrhœal lesions.

Atkin would regard Type I as being highly parasitic and Type II as a more saprophytic form of the gonococcus. Though he obtained evidence that, in the laboratory, Type I strains gradually acquired the properties of Type II strains he was never able to follow the complete transition, probably this is a matter of months or years. It must not be supposed that gonococci can in practice be divided sharply into two colonial types, between the two main types there are probably numerous sub types, each representing one stage in the process of transition. This accounts for the numerous arbitrary subdivisions that have been made on serological grounds by different workers (see below). But, broadly speaking there appear to be two main centres around which the different strains may be grouped, the type strains can be differentiated both by colonial form and serological behaviour. In these respects the gonococcus closely resembles the meningococcus.

**Antigenic Structure.—AGGLUTINATION.**—The serological study of the gonococci may be said to have commenced with Bruckner and Cristéanu's work in 1906 (1906a). Using immune horse serum prepared by the injection of pure cultures of different strains they found that the gonococci were agglutinated to a titre of about 1-750. A close relationship was established between the gonococcus and the meningococcus both of which were agglutinated to nearly equal titre by a gonococcal serum (Bruckner and Cristéanu 1906b). Vannod in the same year (1906) found that immune gonococcal rabbit serum contained agglutinins for the gonococcus and to a less extent for the meningococcus. Torrey (1907) was the first to use the agglutination test for the serological differentiation of the gonococci. Working with 10 different strains and 8 immune sera he found that there was a difference in the agglutinability of the gonococci, and in conjunction with the absorption of agglutinins test he was able to divide the 10 strains into 3 groups. Such a differentiation was obviously of interest but many subsequent workers failed to confirm this (Wollstein 1907, Vannod 1907, Thomsen and Vollmond 1921, Cook and Stafford 1921, Warren 1921). More recent work has however, tended to show the essential correctness of Torrey's findings. Thus Pearce (1915) drew a distinction on the basis of direct agglutination between strains of gonococci isolated from infants—*vulvovaginitis* and *ophthalmitis*—and those isolated from adults—*acute urethritis*. Hermanns (1921a) studied 85 strains, and using the absorption of agglutinins test he was able to classify them into 6 types, of which Types I and II contained the greatest numbers. Later (1921b) he subdivided his Type II strains into 4 races, *a*, *b*, *c*, and *d*. He found a considerable amount of lability in the antigenic structure of these races, some were simple while others were more complex. Jøtten (1921), by direct agglutination classified 20 out of 27 strains into 4 groups A, B, C, and D, 7 remained ungrouped. The strains falling into Groups A and B were mostly from

severe cases of disease with complications those falling into Groups C and D were mostly from simple cases with no complications. He therefore established a correlation between the agglutinability and the virulence of his strains. Torrey who together with Buckell (1922b) again studied the problem of agglutination was able to a large extent to confirm his original conclusions. They used 77 strains of widely separated origin. By simple agglutination they were unable to obtain any definite grouping, but on the basis of absorption they found that their strains could be divided into 3 groups, which they called regular, intermediate, and irregular. The regular strains were most generalized as regards antigenic properties, the intermediate strains were closely related to the regular types, the irregular strains exhibited marked individual variations. The regular strains were the most complex antigenically and appeared on the whole to be more virulent for man, the irregular strains were less complex and appeared to be less virulent. These findings are analogous to those of Griffith with the meningococcus. Torrey and Buckell however point out that the antigenic structure of the gonococcus is variable and so prone to individualistic expression that grouping of the organisms into sharply defined types is not warranted.

Tulloch (1923a, b) studied 100 strains of gonococci. By simple agglutination he obtained no direct evidence of grouping, but by absorption he was able to classify them as follows:

Type I	72	main sub-group	
" I	7	lesser	"
" I	3	"	"
" I	5	"	"
" I	5	"	"
Unplaced	8		

It will be seen that he classifies 92 per cent. in one group, of which 72 strains fall into one sub-group. Tulloch states that Gordon found 25 out of 30 strains by absorption of agglutinins to belong to a well-defined sub-group.

Atkin (1925) was able to classify gonococci into two serological types. Most strains isolated from cases of acute urethritis could be classified in Type I, whereas strains isolated from chronic infection such as cervicitis or arthritis, generally belonged to Type II. Many strains agglutinated to a greater or less extent with serum of each type. Atkin suggests that in the body during the process of a chronic infection, or in the laboratory during long periods of subcultivation, Type I may gradually change into Type II. The evidence for this, however, is admittedly inconclusive. Atkin's work does seem to reconcile to some extent the varying results of different workers, and to agree in particular with the findings of Torrey and Buckell.

Summarizing we may say that the serological classification of the gonococcus is beset with difficulties, that there are probably two main types, of different degrees of antigenic complexity, that between these two main types there are a number of intermediate types containing one or more antigens common to both main types, and that recently isolated strains from acute forms of the disease appear to belong chiefly to Type I while old laboratory strains or strains isolated from chronic disease tend to belong to Type II.

Precipitins—Torrey (1907) prepared a precipitinogen by filtering a 5-weeks broth culture through a layer of sterile talc on filter paper. With this he obtained a precipitin reaction with immune gonococcal rabbit serum. Bruckner and Cristeannu macerated a culture in 0.15 per cent. NaOH which dissolved the gonococci in a few minutes, the solution was filtered through porcelain. They found a close relation between the gonococcus and the meningococcus, thus anti-gonococcal horse serum contained precipitins for both organisms, similarly with anti-meningococcal goat serum.

**Pathogenicity**—The gonococcus is not only a strict parasite, but it is a specific ally human parasite. Of the numerous attempts which have been made to reproduce gonorrhoea in animals other than man, not one has been successful (Leistikow 1882, Neisser 1882, Bumm 1885a, Nicolaysen 1897). Neither inoculation of pus nor of pure cultures on to the mucosa of the urethra or conjunctiva gives rise to disease, even in the anthropoid apes. Experiments on the lower animals, however, have shown that the injection of gonococci directly into the peritoneum or the blood stream frequently proves fatal, and that severe local inflammation may be set up by injecting the organisms into the anterior chamber of the eye or into a joint cavity.

**MICE**.—*Subcutaneous* injection of pure cultures is without effect. *Intraperitoneal* injection of a saline suspension of a young serum agar culture is often fatal in 24 hours. Post mortem there is slight congestion of the peritoneum, and sometimes a small amount of viscous exudate containing pus cells. Gonococci are found in varying numbers, both intra- and extra-cellular in position. They may frequently be cultivated from the peritoneum and occasionally from the heart's blood. With smaller doses, many of the mice do not succumb for 2 to 3 days, at necropsy in these mice it is rare to find gonococci microscopically, and cultures are uniformly sterile. Some mice survive without showing signs of illness. Almost exactly the same results follow the injection of cultures which have been killed by heating to 70° C for 1 hour (Wassermann 1893). There is no increase of virulence by passage.

**GUINEA PIGS**.—*Intraperitoneal* injection of 5 ml. of a 6-days serum broth culture (Nicolaysen 1897) or of a 24-hours' growth on a Blake bottle of serum glucose agar (Wollstein 1907) kills the animals in 24 hours as a rule. Post mortem there is congestion of the serosa with small hemorrhages, a little clear or turbid fluid in the peritoneal cavity, oedema of the pancreas and surrounding tissues, congestion or hemorrhage into the adrenals, a layer of fibrin and pus over the liver, spleen and omentum, sometimes clear fluid in the pleural cavities. Films from the peritoneum and omentum show varying numbers of polymorphonuclears, and diplococci situated intra- and extra-cellularly. Cultures from the peritoneum are generally positive. It will be seen that the post mortem findings are similar to those following injection of guinea-pigs with meningococci, as a rule however larger doses of gonococci are required to produce the same effect. After some generations *in vitro* the gonococci lose their virulence, and become innocuous to guinea pigs.

**RABBITS**.—*Subcutaneous* injection of 10 ml. of a 4 to 5-days' serum broth culture (Maslovski 1900) gives rise to slight inflammatory swelling after 24 hours, later suppuration occurs, so that in 10 days a small abscess is formed, containing thick, sterile pus. The temperature rises somewhat, and the animal loses weight. The same result follows the injection of heat-killed cultures. Smaller doses are without effect.

*Intraperitoneal* injection of large doses of gonococci, washed off young serum agar cultures, kills the animal in 24 hours. Post mortem, there is some peritoneal reaction, and the organisms may be cultivated from the peritoneum and occasionally from the heart's blood. Bruckner and Cristeannu (1906c) claimed to have raised the virulence to such an extent that a rabbit injected intraperitoneally with 1/20 of a serum agar slant died in 2 to 10 hours. These results have not been confirmed.

*Intravenous* injection produces fatal results with smaller doses. The gonococci may be recovered from the blood after 24, sometimes after 48 hours. The results of different workers are, however at variance. Vannod (1907), for example injected living cocci in a dose of 5 ml. of an ascitic peptone broth culture intravenously into rabbits, and obtained practically no reaction.

Maslovski (1900) injected a few drops of a 3-day's serum broth culture into the anterior chamber of the eye of rabbits. The following day there was diffuse turbidity of the cornea,

accompanied by hypopyon, in the pus cells gonococci could not be detected microscopically, but could be cultured for the first two days

Nicolaysen (1897) injected an aqueous suspension of gonococci into the knee joint of a rabbit. Arthritis followed with abundant purulent exudate, which persisted for a week, no organisms could be demonstrated in it, however. The same result occurred when a culture killed by heat at 70° C for 1 hour was injected

From these results it is seen that the gonococcus does not live for long in the animal body, cultures are rarely positive after 2 days, and then only when the injections are made directly into serous cavities or the blood stream. In fact it is doubtful whether true proliferation occurs at all. De Christmas (1897), for instance, found that if the cocci were introduced, enclosed in collodion sacs, into the peritoneal cavity of laboratory animals, they failed to grow. The reaction following the injection of gonococci is due therefore, not to a true infection, but to a toxic action of the organisms. That this is true is abundantly clear from the numerous experiments on gonotoxin

**Toxin Production.**—De Christmas (1897, 1900) recorded experiments from which he concluded that the gonococcus forms a true exotoxin. His results have not, however, been confirmed by other workers. Wassermann (1898), for instance, found that heat killed cultures were as fatal to mice, injected intraperitoneally, as living cultures, on the other hand, filtered cultures proved innocuous, unless a filtrate of a 2 to 3 weeks' culture was used; even then, the filtrate was never as toxic as the whole culture. He grew the gonococci in 33 per cent ascitic broth in a thin layer of fluid in flat bottomed flasks. After a week the cocci were collected, dried, and ground in an agate mortar, weighed quantities of the powder were suspended in water, and sterilized by steaming or in the autoclave. Injected intraperitoneally into mice this powder was fatal in a dose of 0.01 gm, death occurring in 24 hours or later. Injected into rabbits or guinea pigs subcutaneously, it caused wide spread doughy infiltration, often passing on to necrosis. Injected into the anterior chamber of the eye of rabbits, it caused corneal turbidity, hypopyon, and sometimes complete destruction of the eye. He was unable to extract the toxin by heat, by distilled water or by N/10 NaOH, the cocci still remained toxic. The toxin was not destroyed in the bacterial bodies by drying or by heat at 120° C, it was not destroyed by absolute alcohol, or by prolonged boiling, nor was he able to immunize rabbits or mice against it. These results point strongly to the conclusion that the toxin is an endotoxin—a body contained in the cell substance, and adhering strongly to it.

Schäffer (1897) found that filtered ascitic broth cultures, 2 to 6 days old, were without effect when injected in large doses—3–10 ml—into guinea pigs and rabbits. Maslovsky (1900) likewise found that the filtrate of a 9-days' serum broth culture, injected subcutaneously into rabbits, gave rise to nothing but a slight rise of temperature and loss of weight. He concluded too that the toxic substance was an endotoxin. Scholtz (1900) came to the same conclusion. On the other hand, Vannod (1907) working with filtrates of 17–20 days' cultures in de Christmas's medium—concentrated veal broth containing 75 per cent of ascitic fluid—found that they were fatal to rabbits injected intraperitoneally. Post mortem, there was a purulent effusion into the peritoneal cavity, the serosa was congested and covered with purulent deposits, the suprarenals were enlarged and hyperæmic. Vannod, however, used large and repeated doses—a total of 18 to 24 ml. in 10 days to 3 weeks.

We may conclude that the gonococcus contains a toxin which can be extracted by grinding the dried organisms and suspending the resultant powder in water. In cultures in fluid media the toxin may be liberated from the cocci by autolysis, after 2 or 3 weeks' incubation, the organisms may have autolysed to such an extent that a certain amount of toxin may be present in filtrates of the cultures. The available evidence leaves little doubt that the toxin belongs to the class of so called endotoxins.

*Neisseria gonorrhoeae*

*Synonym.*—*Gonococcus*.

*Isolation.*—First described by Neisser in 1879, first cultivated by Bumm (1885a, b), and Leistikow and Loeffler (Leistikow 1882).

*Habitat.*—Strict parasite of man. Found in genito-urinary system of patients suffering from gonorrhoea.

*Morphology.*—Oval or spherical coccus,  $0.8 \mu \times 0.6 \mu$ , frequently arranged in pairs, with adjacent sides flattened or slightly concave, resembling a pair of kidney beans, long axis of oval lies at right angles to axis joining the two cocci. In cultures great variation in size and in depth of staining occurs, due to autolysis, in the body the cocci are more regular, and are generally intracellular. Non motile, non-capsulated, Gram negative.

*Serum Agar Plate.*—24 hours  $37^{\circ} \text{C}$  Round, convex or slightly umbonate, greyish white translucent, amorphous colonies, 0.5–1 mm. in diameter, with smooth, glistening surface and entire edge, consistency butyrous or slightly viscid, fairly easily emulsifiable. Later, colonies increase in size, and may develop a roughened surface and a crenated edge.

*Serum Agar Slope.*—24 hours  $37^{\circ} \text{C}$  Rather poor, partly confluent, raised, greyish yellow growth with smooth surface, edge entire or formed of single colonies. Consistency often viscid.

*Gelatin Slab.*—No growth.

*Serum Broth.*—24 hours  $37^{\circ} \text{C}$  Very poor growth with little or no turbidity, and a slight granular deposit, partly disintegrating on shaking.

*Resistance.*—Highly susceptible to inimical agencies. When dried, the cocci succumb in an hour or two. Killed by moist heat at  $55^{\circ} \text{C}$ . in less than 5 minutes, and at  $42^{\circ} \text{C}$  in 5–15 hours (Carpenter *et al* 1933). In serum cultures they are killed by 1/4000  $\text{AgNO}_3$  in  $7\frac{1}{2}$  minutes, and in pus in 2 minutes. Sealed cultures kept at  $37^{\circ} \text{C}$ . may live for 4 or 5 weeks, when kept at room temperature, they die in a day or two.

*Metabolism.*—Optimum H ion concentration for growth is pH 7.5. Optimum temperature for growth is  $37^{\circ} \text{C}$  no growth under  $30^{\circ} \text{C}$ . or over  $33.5^{\circ} \text{C}$ . Fails to grow on plain agar as a rule, requires the presence of serum blood, ascitic fluid, or hydrocele fluid, glucose is not beneficial. Aerobic, but growth is said to be improved by a lowered oxygen pressure, by presence of —SH groups, and by 10 per cent of  $\text{CO}_2$  in the atmosphere. Little or no growth under strictly anaerobic conditions.

*Biochemical.*—Produces acid, no gas, in glucose. No change in litmus milk. Catalase—M.B. reduction—M.R.—, V.P.—, indole—,  $\text{H}_2\text{S}$ —

*Antigenic Structure.*—No clear definition into separate serological types, most strains appear by agglutination and absorption of agglutinins to belong to one or other of two main groups, numerous other less important groups.

*Pathogenicity.*—Responsible for gonorrhoea and ophthalmia neonatorum in man. Experimentally, it proves fatal to mice, guinea-pigs, and rabbits, if injected in large doses intraperitoneally, there is little or no multiplication of the organisms in the body.

Differentiation of the *Gonococcus* from the *Meningococcus*.—Morphologically the two organisms are very similar. In the body, both occur chiefly in pairs situated intracellularly. It is sometimes stated that the adjacent sides of the meningococci are flattened whereas those of the gonococci are concave, thus leaving an oval space between the two organisms. Possibly the meningococcus is slightly larger than the gonococcus in the body, though smaller in a 24-hours' culture in the laboratory (Wollstein 1907). Culturally the gonococcus is more dysgonic, it grows more slowly, forms smaller colonies, and grows on a narrower range of media than the meningococcus, the colonies are slightly viscous and do

not emulsify so readily, colonies of the meningococcus, on the other hand, are butyrous and emulsify with the greatest of ease. Biochemically, the meningococcus produces acid in glucose and maltose, the gonococcus in glucose only as pointed out on page 531 a medium containing human or rabbit serum should be used for testing the fermentation of maltose. The agglutination test is only of limited value, since there exists a group relationship between the two organisms (Bruckner and Cristianu 1906b, Wollstein 1907, Elser and Huntoon 1909, Gordon, M. H., 1925). The meningococcus is more toxic to animals, tested by intra peritoneal injection on mice or guinea pigs a smaller dose of meningococci than of gonococci is required, whether alive or dead, to cause death.

*N. catarrhalis*.—This organism has been variously described by different workers. Ghon and Pfeiffer (1902), who first studied it fully, stated that in sputum it occurred in pairs, tetrads, or occasionally small groups, the organisms were shaped like coffee-beans, and were both intra and extra cellular. In culture they appeared larger, were generally in tetrads, and stained evenly. On agar after 24 hours, the colonies resembled in size those of *Streptococcus pyogenes*; they were convex, whitish grey, with a glistening surface and an eaten edge. After 3 or 4 days they were 3 to 4 mm in diameter, and were differentiated into a prominent, more elevated, opaque, slightly brownish centre, and a thinner grey, transparent, wave-like periphery with a crenated edge; the consistency was friable, and in saline they auto-agglutinated. A more or less similar description was given by von Lingelsheim (1906), he stated that the colony was smaller than that of a meningococcus colony, and that even on ascitic agar the diameter never exceeded 1-2 mm. Elser and Huntoon (1909) described two types of colony, one resembling Ghon and Pfeiffer's description the other like a small meningococcus colony. J. E. Gordon (1921) described four types of colony: (1) like Ghon and Pfeiffer's, (2) similar to the first, but coloured pale yellow, (3) small, flat, grey, translucent, amorphous colonies with a smooth glistening surface and entire edge, (4) almost pin point, transparent glistening colonies with a smoothly rounded edge. Morphologically, these consisted of giant cocci showing metachromatic staining. It seems clear that the colonial appearance of *N. catarrhalis* is subject to variation, in all probability both smooth and rough types are formed, similar to those of the meningococcus and the gonococcus. In gelatin stab culture there is a poor growth confined to the upper part of the tube, there is no liquefaction. There is no turbidity in broth, but a granular deposit is formed, if the tube is kept still, a surface membrane may appear. Growth occurs within a range of 18° to 42° C, the optimum being at 37° C. There is no development under anaerobic conditions. Growth is favoured by blood, serum, and ascitic fluid, but not by glycerine. The organism appears to be more resistant than the meningococcus or gonococcus, cultures are said to live for 4 or 5 months at 21° C, if prevented from drying, the organisms may live in dried sputum for 27 days, they are killed by heating to 65° C for 30 minutes. No sugars are fermented. The virulence of this organism to laboratory animals is low. The rabbit is resistant, but guinea pigs injected intraperitoneally with large doses—half to one agar slope—die of toxæmia in about 24 hours. At post mortem there is a mild degree of peritonitis, slight enlargement of the spleen, and hyperæmia and degeneration of the viscera, the organisms may be recovered from the peritoneal exudate, but rarely from the heart's blood. Heat killed cultures are almost as fatal as living ones.



*N. pharyngis* flava, I, II and III—A number of Gram negative cocci have been described whose characteristic feature is the formation of greenish yellow colonies on agar or ascitic agar von Langelsheim (1906) and some other workers have called them *Micrococcus* or *Diplococcus pharyngis flarus*, and have differentiated them on the basis of colonial appearance and sugar reactions into three groups, I, II and III (Martin 1911, Report 1916, Dopter 1921). Elser and Huntoon (1909) have called them chromogenic cocci, and have divided them similarly into three groups I, II and III. J. E. Gordon (1921) has likewise called them chromogenic cocci, and has divided them into six groups, I to VI. Elser and Huntoon's chromogenic III and Gordon's chromogenic III agree with von Langelsheim's flava III in fermenting glucose and maltose only, Elser and Huntoon's chromogenic II and Gordon's IV agree with von Langelsheim's I and II in fermenting glucose, maltose and

lævulose; Elser and Huntoon's chromogenic I agrees with Gordon's chromogenic V in fermenting glucose, maltose, lævulose, and sucrose; but has no counterpart in von Langelsheim's classification. Over and above these are Gordon's chromogenic I, which ferments glucose only, his chromogenic II, which ferments glucose and lævulose, and his chromogenic VI, which ferments glucose, maltose, lævulose, sucrose, and lactose. Undoubtedly one of the reasons for the discrepancies in the sugar reactions is due to the fact that von Langelsheim, who described the flava group read all his reactions after 24 hours, this probably explains why he never observed the fermentation of sucrose. In colonial appearance von Langelsheim's flava I and III resemble the meningococcus, and flava II *N. catarrhalis*, Elser and Huntoon's chromogenic II

agrees with von Langelsheim's flava I and III. Gordon describes his organisms as giving colonies either like the meningococcus or like *N. catarrhalis*, his group VI gives pale yellow colonies larger and more opaque than the others. In the face of these divergencies in cultural and biochemical reactions, it is clear that no fixed types can be described. Our own work has shown that the cultural appearances of the Gram negative cocci from the nasopharynx are subject to great variation, and most workers agree now that lævulose is an unreliable sugar. Division therefore of these organisms either on



FIG 113—*Neisseria pharyngis*  
Surface colony on agar, 24 hours  
37° C. (× 8) Smooth type



FIG 114.—*Neisseria pharyngis*  
Surface colonies on agar, 5 days  
37° C. showing differentiation.  
Secondary rough type (× 8)



FIG 115.—*Neisseria pharyngis*  
Surface colonies on agar, 5 days,  
37° C., showing formation of second-  
ary papillae (× 8)

colonial appearance or on the basis of levulose fermentation is most unsatisfactory. We must await further work before a classification can be attempted.

*N. pharyngis sicca*—According to von Langelshiem (1906) this organism consists of fine Gram negative diplococci. On agar it gives rise to an irregularly round raised, opaque, slightly yellowish colony, up to 3 mm. in diameter, with a dull, dry, deeply furrowed surface, and a crenated edge, the colony is very firm, often adherent to the medium, difficult to disintegrate, and impossible to emulsify, it is so coherent that it can be picked up bodily. von Langelshiem says that it produces acid in glucose, maltose, and levulose, but other workers have found that it also ferments sucrose (Elaer and Huntoon 1909, Gordon J. E., 1921). It seems doubtful whether this organism should be regarded as a distinct species. Our own work suggests that it is merely a rough variant of one of the other nasopharyngeal cocci which ferments glucose, maltose, and sucrose. We have observed the formation by cocci giving these sugar reactions of smooth colonies when first isolated from the nose, and the appearance in later cultures of typically rough colonies indistinguishable from those described as being characteristic of *N. pharyngis sicca*.

*N. pharyngis cinerea*—von Langelshiem (1906) described this organism as consisting of plump cocci arranged in pairs or more usually loose heaps. On agar it forms small round, grey or greyish white colonies, 1-1.5 mm. diameter, with an entire edge, under a low magnification their colour is brownish and they appear coarsely granular. Some authors state that the colonies are dry, brittle, and opaque (Netter and Debré 1911). It ferments no sugars. This organism closely resembles *N. catarrhalis* and is probably merely a variety of it, it corresponds closely in description to Gordon's *N. catarrhalis* sub-group III (Gordon J. E., 1921).

*Diplococcus mucosus*—This organism, which was isolated by von Langelshiem (1906, 1908) from the nasopharynx and cerebrospinal fluid is distinguished by its capsulation, growth on plain agar, growth at room temperature, and formation of mucinous colonies. Microscopically it consists of small diplo- and tetrads surrounded by true capsules. Colonies are 1.5-4 mm. in diameter, convex, yellowish grey, opaque, mucinous, and easily emulsified. Gelatin is not usually liquefied. McFarlan's (1941) and Bray and Crick shank's (1943) strains grew on MacConkey's agar, fermented glucose only, and did not liquefy gelatin. Cowan's (1939) two strains grew on MacConkey's agar, produced late acid from lactose, and acid and clot in litmus milk. Most strains seem to be pathogenic for mice. The systematic position of this organism is still in doubt. It may be noted that an organism resembling *Diplococcus mucosus* is not infrequently present in infected wounds and burns.

*Diplococcus crassus*—This organism is not easy to define. It was first described by Jaeger (1895) as a meningococcus, but as it was said to form long chains in culture, to be Gram positive in the cerebrospinal fluid and in pure cultures, to be able to grow on gelatin at room temperature, and to be highly resistant to drying it is certain that it was not the meningococcus (Jaeger 1903a). On agar it forms rather small, greyish white, granular colonies, 1-1.5 mm. in diameter, with an entire edge. Growth occurs at 20° C. Microscopically it consists of plump diplo- and tetrads, some of which are Gram positive and some Gram negative, it appears in reality to be a Gram positive organism that is very easily decolorized. Its sugar reactions differentiate it from all the other members of the group, since in addition to fermenting glucose, maltose, levulose, and sucrose, it ferments lactose. It is said to be sometimes agglutinated by anti-



FIG 116—*Neisseria pharyngis*  
Surface colony on agar 24  
hours, 37° C × 8 showing  
primary rough type of  
colonial variant

TABLE 35

Organism	Morphology	24-48 hour Colony on Aseptic Agar	Serum Broth	Growth on Agar	Growth at 22°C	Dextrose	Maltose	Sucrose	Remarks
<i>Neisseria meningitidis</i>	Oval or spherical coccus, often in pairs, with adjacent sides flattened (Fig. 1)	Round, convex, bluish grey, translucent, amorphous, 1-2 mm in diameter, with smooth glistening surface and entire edge. Butyrous and easily emulsifiable	Moderate turbidity, and slight granular deposit	±	-	A	A	-	Two or four main antigenic types
<i>Neisseria gonorrhoeae</i>	Oval or spherical coccus, often in pairs, with adjacent sides, concave	Round, convex, or slightly umbonate, greyish white, translucent, amorphous, 1 mm in diameter, with smooth glistening surface and entire edge; slightly viscid and fairly easily emulsifiable	Little or no turbidity; slight granular deposit	-	-	A	-	-	Growth is slower and poorer than meningococcus. Probably 2 main antigenic types.
<i>Neisseria catarrhalis</i>	Spherical or oval coccus arranged singly, in pairs, in tetrads, or in clumps	<i>Smooth Type</i> Round, low convex, greyish white, translucent, and amorphous, with smooth, glistening surface and entire edge <i>Rough Type</i> Larger, raised, whitish grey, with slightly uneven surface and entire edge; later differentiated into a raised, brownish, more opaque centre, and a thinner greyish translucent periphery with a crenated edge; friable and difficult to emulsify	No turbidity; coarse, granular deposit; sometimes a surface pellicle and ring growth	±	±	-	-	-	Auto agglutinable
<i>N. pyrogena</i>	Small diplococci, uniform in size, and arranged in dense clumps	Irregularly round, raised, greyish-white, opaque, 2-3 mm in diameter; with dry, deeply wrinkled surface and an undulate edge; very tough and brittle consistency; adherent to medium; impossible to emulsify		+	+	A	A	A	

<i>N. flava</i> i	Similar to the meningococcus	Round low convex, greenish yellow, translucent, amorphous, 0.5-2 mm in diameter, with smooth, glistening surface and entire edge. Consistency butyrous, emulsifies fairly well	No turbidity. granular deposit, often a surface pellicle	+	+	A	A	-	
<i>N. flava</i> ii	Similar to <i>N. catarrhalis</i>	Round low convex, opaque, yellow, 0.5-1.0 mm in diameter, with smooth surface and entire edge. Brittle, and difficult to emulsify	Like <i>N. flava</i> i	-	-	A	A	A	
<i>N. flava</i> iii	Similar to the meningococcus	Resembles colony of meningococcus, but slightly yellow in colour. butyrous, and fairly easily emulsifiable	Like <i>N. flava</i> i	±	±	A	A	-	
<i>Dyscococcus mucosus</i>	Cocci arranged in pairs and tetrads, surrounded by capsules	Luxuriant round convex greyish translucent, and mucoid, 3-4 mm in diameter with smooth glistening surface and entire edge, viscous consistency	Turbidity and surface ring growth	+	+	A	-	-	Pathogenic for mice
<i>Dyscococcus crassus</i>	1 lump cocci in pairs, tetrads, or chains. Gram staining in later minute	Round, convex, greyish white, granular and translucent 0.5-1.5 mm in diameter with entire edge		+	+	A	A	A	Acid in lactose
<i>N. flavescens</i>	Similar to the meningococcus	Like those of the meningococcus, but less moist and characterized by a golden yellow pigmentation		+	+	-	-	-	Very poor growth on dextrose agar. Antigenically homogeneous and different from <i>N. meningitidis</i> and <i>N. catarrhalis</i>

- FLÜGGE, C (1896) "Die Mikroorganismen" Leipzig
- GHOF, A and PRIGER, H (1902) *Z klin Med*, 44, 262
- GORDON, J (1926) *J Path Bact*, 20, 319
- GORDON, J and M LEON, J W (1926) *J Path Bact*, 29, 13, (1928) *Ibid*, 31, 185
- GORDON, J E (1921) *J infect Dis*, 29, 462
- GORDON, M H (1925) *Spec Rep Ser med Res Coun, Lond*, No 98, p 105
- GORDON, M H and MORRIS, E G (1915) *J R Army med Cps*, 25, 411
- GRIFFITH, F (1917) *Rep loc Govt Bd publ Hlth New Ser*, No 111, p 62
- GURD, F B (1908) *J med Res*, 18, 291
- HALL, I C (1916) *J Bact*, 1, 343
- HENDRY C B (1938) *J Path Bact*, 46, 383
- HERMANIKS, J (1921a) *J infect Dis*, 28, 132, (1921b) *Ibid* 29, 11
- JAEGER, H (1895) *Z Hyg InfektKr*, 19, 351, (1899) *Dtsch med Wschr*, 25, 472, (1903a) *Zbl Bakt*, 33, 23, (1903b) *Ibid*, 33, 681, (1903c) *Z Hyg InfektKr*, 44, 225
- JEVENS, C E (1921) *J Path Bact*, 24, 166 (1922) *Ibid*, 25, 165
- JÖTTEN, K W (1921) *Z Hyg InfektKr*, 92, 9
- KANDLER, L (1922) *Z Hyg InfektKr*, 96, 347
- KIRBY, (1895) *Berl klin Wschr*, 32, 332
- KIRKBRIDE M B and COHEN, S M (1934) *Amer J Hyg*, 20, 444
- KOHMAN, E F (1919) *J Bact* 4, 571
- KUTSCHER, K (1906) *Dtsch med Wschr*, 22, 1071
- LEBEUF, F (1924) *C R Soc Biol*, 90 768
- LEISTIKOV (1882) *Berl klin Wschr* 19, 500
- LINGELSHIEIM, W von (1903) *Dtsch med Wschr*, 31, 1217 (1906) *Klin Jb* 15, 373 (1908) *Z Hyg InfektKr*, 59, 457
- LIPSCHÜTZ, B (1904) *Zbl Bakt* 36, 743
- LLOYD, D J (1916-17) *J Path Bact* 21, 113
- LORENTZ, F H (1922) *Munch med Wschr*, 69, 1695
- McFARLAN, A M (1941) *J Path Bact* 53, 446
- MACLAGAN, P W and COOKE W E (1917) *J R Army med Cps*, 29, 228
- MACNAUGHTON F G (1923) *J Path Bact* 26, 297
- McLEOD, J W, COATES J C, HATFIELD, F C, PRIESTLEY, D P, and WHEATLEY, R. (1934) *J Path Bact*, 39, 221
- McLEOD, J W, WHEATLEY, B and PHILON H V (1927) *Brit J exp Path* 8, 25
- M DONALD, S (1908) *J Path Bact* 12, 442
- MAEGBAITH, B (1935) *Brit J exp Path*, 16, 109
- MARTIN W B M. (1910) *J Path Bact*, 14, 136, (1911) *Ibid*, 15, 76
- MAEY, H and WOUTER, F (1900) *Zbl Bakt*, 28, 1 33 65 97
- MASLOVSKI (1900) *Zbl Bakt* 27, 541
- MEYER, A E O and RAKE G (1942) *J exp Med*, 75, 437
- MILLER C P (1933) *Science*, 78, 340, (1934-35) *Proc Soc exp Biol, NY* 32, 1136 1138 1140
- MILLER C P and BOOR, A K. (1934) *J exp Med*, 59, 75
- MURRAY, E G D (1929) *Spec Rep Ser med Res Coun Lond* No 124 p 15
- MURRAY, E G D and AYRTON R (1924) *J Hyg, Camb*, 23, 23
- NABARRO D (1917) *Rep loc Govt Bd publ Hlth, New Ser*, No 114 p 207
- NEILL, M H and TAFT, C E (1920) *Bull US Hyg Lab*, No 124 p 93
- NEISSER A. (1879) *Zbl med Wiss*, 17, 497 (1882) *Dtsch med Wschr* 8, 279
- NETTER, A and DERRÉ, R. (1911) 'La Ménigite Cérébro spinale' Paris
- NICOLAYSEN, L. (1897) *Zbl Bakt*, 22, 305
- NICOLLE N, DEBAINS E, and JOUAN C (1918) *Ann Inst Pasteur* 32, 160
- OLIVER J O (1929) *J Hyg, Camb*, 29, 259
- PEARCE L. (1915) *J exp Med*, 21, 289
- PETRIE G F (1932) *Brit J exp Path*, 13, 380 (1937) *J Hyg Camb*, 37, 42
- PHILON H V, DUTHIE, G M, and M LEON, J W (1927) *J Path Bact* 30, 133
- PRICE, I N O (1933) 'The Complement Fixation Test for Gonorrhoea' London County Council
- RAKE, G (1933) *J exp Med* 57, 549
- RAKE, G and SCHERR, H W (1933a) *J exp Med* 58, 341 (1933b) *Ibid*, 58, 361
- Report (1916) *Spec Rep Ser med Res Coun, Lond* No 2, (1920) *Ibid*, No 80
- RIST, E and PARIS, A (1904) *Bull Inst Pasteur*, 2, 338
- ROCKWELL, G E and MCKHANN, C F (1921) *J infect Dis*, 28, 249
- ROSEH A B (1936) *Pers comm*
- RUEDIGER E H (1919) *J infect Dis*, 24, 370
- SCHÄFFER J (1897) *Fortschr Med*, 15, 813
- SCHERR, H W and RAKE G (1935) *J exp Med*, 61, 753
- SCHOLTZ, W (1900) *Zbl Bakt*, 27, 162
- SCOTT, W M (1917) *Pep. loc. Govt Bd. publ Hlth New Ser*, No 114, p 111.

meningococcal serum (Jaeger 1903b, c, Netter and Debre 1911, Dopter 1921). According to von Langelsheim (1906) an antimeningococcal serum agglutinates *Dip. crassus* almost or quite to titre, whereas an anti-crassus serum agglutinates the meningococcus to only  $\frac{1}{2}$ – $\frac{1}{16}$  titre. According to Jaeger (1899) dried cultures remain alive for 3 or 4 months.

*N. flavescens*.—This organism was isolated by Branham (1930) from the spinal fluid of a number of patients suffering from epidemic cerebrospinal meningitis. It differs from the meningococcus chiefly in its production of pigment, its lack of fermentative action, and its antigenic constitution. In the spinal fluid it appears in the form of Gram negative oval-shaped cocci arranged in flattened pairs, individual cells vary in size and depth of staining, giant forms are common. The organisms grow well on blood agar and semi-solid agar, but poorly on dextrose agar. They produce a golden yellow pigment. They are without fermentative action on any of the usual carbohydrates. They are not agglutinated by type antimeningococcal sera, but constitute among themselves a serologically homogeneous group. Their ability to give rise under favourable conditions to meningitis appears to be unquestioned.

## REFERENCES

- ALBRECHT H and GHOF, A. (1901) *Wien Klin. Wochs.*, 14, 954.  
 ARKWEIGHT, J. A. (1907) *J. Hyg., Camb.*, 7, 145, (1909) *J. Hyg., Camb.*, 9, 104, (1915) *Brit. med. J.*, ii, 885.  
 ATKIN, E. E. (1923) *Brit. J. exp. Path.*, 4, 325, (1925) *Ibid.*, 6, 235.  
 BOOR, A. K. (1912) *Proc. Soc. exp. Biol., N. Y.*, 50, 22.  
 BOOR, A. K. and MILLER, C. P. (1934) *J. exp. Med.*, 59, 63, (1944) *J. infect. Dis.*, 75, 47.  
 BRANHAM, E. (1930) *Publ. Hlth Rep., Wash.*, 45, 845, (1932) *J. Immunol.*, 23, 49, (1940) *Bact. Rev.*, 4, 59.  
 BRANHAM, S. E. and CARLIN, S. A. (1937) *J. Bact.*, 34, 270, (1942) *Proc. Soc. exp. Biol., N. Y.*, 49, 141.  
 BRANHAM, S. E. and LILLIE, R. D. (1932) *Publ. Hlth Rep., Wash.*, 47, 2137.  
 BRANHAM, S. E., TAFT, C. E., and CARLIN, S. A. (1931) *Publ. Hlth Rep., Wash.*, 46, 897.  
 BRAY, P. T. and CRICKSHANK, J. C. (1943) *Brit. med. J.*, i, 601.  
 BRUCK, C. (1906) *Deutsch. med. Wochs.*, 32, 1363.  
 BRUCKNER, J. and CRISTEANU, C. (1906a) *C. P. Soc. Biol.*, 60, 846, (1906b) *Ibid.*, 60, 907, (1906c) *Ibid.*, 60, 942.  
 BUMM, E. (1885a) *Deutsch. med. Wochs.*, 11, 508, (1885b) *Ibid.*, 11, 910.  
 BUSCHKE, A. and LANGER, E. (1921) *Deutsch. med. Wochs.*, 47, 65.  
 BUTTERFIELD, C. T. and NEILL, M. H. (1930) *Bull. U.S. Hyg. Lab.*, No 124, p. 11.  
 CARPENTER, C. M., BOAK, R. A., MUCCI, L. A., and WARREN, S. L. (1933) *J. Lab. clin. Med.*, 18, 981.  
 CASPER, W. A. (1937a) *J. Bact.*, 34, 303, (1937b) *J. Immunol.*, 32, 421.  
 CHAPIN, C. W. (1918) *J. infect. Dis.*, 23, 342.  
 CHRISTMAS, J. DE. (1897) *Ann. Inst. Pasteur*, 11, 609, (1900) *Ibid.*, 14, 331.  
 CLAPP, F. L., PHILLIPS, S. W., and STAHL, H. J. (1930) *Proc. Soc. exp. Biol., N. Y.*, 33, 302.  
 CLARK, G. W. (1920) *J. Bact.*, 5, 99.  
 COHEN, S. M. (1940) *J. infect. Dis.*, 67, 74.  
 COHN, A. (1923) *Klin. Wochs.*, 2, 573.  
 COLE, S. W. and LLOYD, D. J. (1917) *J. Path. Bact.*, 21, 267.  
 COOK, M. W. and STAFFORD, D. D. (1921) *J. infect. Dis.*, 23, 561.  
 COSTA, S. and BOYER, L. (1922) *C. R. Soc. Biol.*, 87, 806.  
 COWAN, S. T. (1933) *Lancet*, ii, 1002.  
 DOTTER, C. (1909) *C. R. Soc. Biol.*, 67, 74, (1921) "L'Infection Méningococcique" Paris.  
 DOTTER and PATROV (1914) *C. R. Soc. Biol.*, 77, 231, 292.  
 DUNN, R. A. and GORDON, M. H. (1900) *Brit. med. J.*, ii, 421.  
 ELLIS, A. W. M. (1915) *Brit. med. J.*, ii, 880.  
 ELZER, W. J. and HUNTOON, F. M. (1909) *J. med. Res.*, 20, 371.  
 ERICSSON, M. J. and ALBERT, H. (1922) *J. infect. Dis.*, 30, 268.  
 EVANS, A. C. (1920) *Bull. U.S. Hyg. Lab.*, No 124, p. 45.  
 FERRY, N. S., NORTON, J. F., and STEELE, A. H. (1931) *J. Immunol.*, 21, 293.  
 FERRY, N. S. and SCHORNACK, P. J. (1934) *J. Immunol.*, 28, 143.  
 FLEISHER, S. (1907a) *J. exp. Med.*, 9, 105, (1907b) *Ibid.*, 9, 142.

- FLÖGGE, C. (1896) \* Die Mikroorganismen Leipzig
- GHOV, A. and FRISCHER, H. (1900) *Z. Klin. Med.* 44, 202
- GORDON, J. (1915) *J. Path. Bact.* 20, 719
- GORDON, J. and M. LEON, J. W. (1916) *J. Path. Bact.* 29, 13 (1923) *Ibid.* 31, 185
- GORDON, J. L. (1921) *J. infect. Dis.* 29, 462
- GORDON, M. H. (1925) *Spec. Rep. Ser. med. Res. Coun., Lond.* No. 93, p. 105
- GORDON, M. H. and MURRAY, L. G. (1915) *J. R. Army med. Cps.* 25, 411
- GRIFFITH, F. (1917) *Rep. loc. Govt. Bd. pub. Hlth.* New Ser., No. 111, p. 52
- GRUB, F. B. (1905) *J. med. Res.* 18, 201
- HALL, I. C. (1916) *J. Bact.* 1, 313
- HENRY, C. B. (1935) *J. Path. Bact.* 40, 347
- HERMAN, J. (1921a) *J. infect. Dis.* 23, 132, (1921b) *Ibid.* 29, 11
- JAEGER, H. (1895) *Z. Hyg. Infectkr.* 19, 351, (1899) *Dtsch. med. Wochr.* 25, 471, (1903a) *ZM. Bakt.* 33, 23, (1903b) *Ibid.* 33, 681 (1903c) *Z. Hyg. Infectkr.* 44, 225
- JENKINS, C. L. (1921) *J. Path. Bact.* 24, 160, (1922) *Ibid.* 25, 103
- JÖTTEN, A. W. (1921) *Z. Hyg. Infectkr.* 92, 9
- KANDLER, L. (1927) *Z. Hyg. Infectkr.* 90, 347
- KIRCH, (1895) *Berl. Klin. Wochr.* 32, 332
- KIRKBRIDE, M. B. and CONYER, S. M. (1931) *Amer. J. Hyg.* 20, 441
- KOHMAN, L. F. (1919) *J. Bact.* 4, 571
- KOTSCHER, A. (1906) *Dtsch. med. Wochr.* 32, 1071
- LEDEBT, F. (1921) *C. R. Soc. Biol.* 90, 68
- LEISTIKOW, (1892) *Berl. Klin. Wochr.* 19, 500
- LINGENBERG, W. von (1903) *Dtsch. med. Wochr.* 31, 1217 (1908) *Klin. Wb.* 15, 373 (1909) *J. Hyg. Infectkr.* 39, 457
- LIPSCHITZ, H. (1904) *ZM. Bakt.* 36, 743
- LYON, D. J. (1916) *J. Path. Bact.* 21, 115
- LORENTZ, I. H. (1922) *Dtsch. med. Wochr.* 69, 1035
- McFARLAN, A. M. (1911) *J. Path. Bact.* 53, 416
- MACLAGAN, P. W. and COOKE, W. L. (1917) *J. R. Army med. Cps.* 29, 228
- MACNAUGHTON, F. G. (1923) *J. Path. Bact.* 26, 297
- McLEOD, J. W., COATES, J. C., HARTOLI, I. C., FRIENTLEY, D. I. and WHEATLEY, B. (1934) *J. Path. Bact.* 29, 21
- McLEOD, J. W., WHEATLEY, B. and PHILSON, H. V. (1927) *Brit. J. exp. Path.* 8, 10
- McDONALD, S. (1908) *J. Path. Bact.* 12, 442
- MARGRAITH, R. (1935) *Brit. J. exp. Path.* 16, 109
- MARTIN, W. B. M. (1910) *J. Path. Bact.* 14, 136 (1911) *Ibid.* 15, 76
- MARY, H. and WOTHE, F. (1900) *ZM. Bakt.* 28, 1 33 65 97
- MASTOVSKI, (1900) *ZM. Bakt.* 27, 541
- MEYER, A. L. O. and LAKE, G. (1917) *J. exp. Med.* 75, 437
- MILLER, C. I. (1933) *Science* 78, 311 (1934-35) *Proc. Soc. exp. Biol. A* 1 32, 1136 1137 1140
- MILLER, C. I. and BOOR, A. K. (1934) *J. exp. Med.* 59, 75
- MURRAY, L. G. D. (1909) *Spec. Rep. Ser. med. Res. Coun., Lond.* No. 121 p. 17
- MURRAY, L. G. D. and AYTTON, R. (1914) *J. Hyg., Camb.* 23, 23
- NABARRO, D. (1917) *Rep. loc. Govt. Bd. pub. Hlth.* New Ser. No. 114 p. 207
- NEILL, M. H. and TAYLOR, C. L. (1920) *Bull. U.S. Hyg. Lab.* No. 124 p. 43
- NEISSER, A. (1879) *ZM. med. Wochr.* 17, 497, (1880) *Dtsch. med. Wochr.* 8, 279
- NETTER, A. and DEBERG, R. (1911) *La Meningite Cérébro-spinale* Paris
- NICOLAYEV, L. (1897) *ZM. Bakt.* 20, 305
- NICOLLE, N., DEBAIN, F. and JOCAN, C. (1918) *Ann. Inst. Pasteur* 32, 150
- OLIVER, J. O. (1929) *J. Hyg., Camb.* 20, 209
- PEARCE, L. (1915) *J. exp. Med.* 21, 289
- PETRIE, G. F. (1932) *Brit. J. exp. Path.* 13, 380 (1937) *J. Hyg. Camb.* 37, 42
- PHILSON, H. V., DETRIZ, G. M., and MILLER, J. W. (1927) *J. Path. Bact.* 30, 133
- PRICE, F. V. O. (1933) "The Complement Fixation Test for Gonorrhea" London County Council.
- PAKE, G. (1933) *J. exp. Med.* 57, 549
- PAKE, G. and SCHERF, H. W. (1933a) *J. exp. Med.* 58, 341, (1933b) *Ibid.* 58, 301
- Report. (1916) *Spec. Rep. Ser. med. Res. Coun., Lond.* No. 2, (1920) *Ibid.* No. 50
- RIST, L. and LARIN, A. (1904) *Bull. Inst. Pasteur* 2, 338
- ROCKWELL, G. F. and MCHUGHAN, C. I. (1921) *J. infect. Dis.* 28, 249
- ROSEN, A. B. (1936) *Proc. comm.*
- RÜDIGER, E. H. (1919) *J. infect. Dis.* 24, 378
- SCHÄFFER, J. (1897) *Fortschr. Med.* 15, 813
- SCHERF, H. W. and PAKE, G. (1935) *J. exp. Med.* 61, 753
- SCHULTZ, W. (1900) *ZM. Bakt.* 27, 102
- SCOTT, W. M. (1917) *Rep. loc. Govt. Bd. pub. Hlth.* New Ser., No. 114, p. 111

- SORDELLI A., MIRAVENT J. M., and VEGRONI, P. (1926) *Rev Inst. bact., B Aires*, 4, 635.  
 STOKINGER, H. E., ACKERMAN, H., and CARPENTER, C. M. (1944) *J. Bact.*, 47, 141.  
 SWARTZ, E. O. and DAVIS, D. M. (1920) *J. Amer. med. Ass.*, 75, 1124.  
 TEAGUE O. and TORREY J. C. (1907) *J. med. Res.*, 17, 223.  
 THALMANN (1900) *Zll. Bakt.*, 27, 828.  
 THOMSEN O. and VOLLMOND, E. (1921) *C. P. Soc. Biol.*, 84, 328.  
 THOMSON D. (1917) *Brit. med. J.*, 1, 869.  
 TORREY J. C. (1907) *J. med. Res.*, 18, 329. (1908) *Ibid.*, 19, 471.  
 TORREY J. C. and BUCKELL, G. T. (1922a) *J. infect. Dis.*, 31, 125. (1922b) *J. Immunol.*, 7, 303.  
 TORREY, J. C., WILSON M. A., and BUCKELL, G. T. (1922) *J. infect. Dis.*, 31, 148.  
 TULLOCH, W. J. (1923a) *J. State Med.*, 31, 501. (1923b) *J. E. Army med. Cps* 40, 12, 93.  
 VANNOD, T. (1906) *Diach. med. Wochr.*, 32, 1984. (1907) *Zll. Bakt.*, 44, 10, 110.  
 VEDDER, E. B. (1915) *J. infect. Dis.*, 18, 385.  
 WARREN S. H. (1921) *J. Path. Bact.*, 24, 424.  
 WASSERMANN A. (1897) *Berl. klin. Wochr.*, 34, 685. (1898) *Z. Hyg. Infectkr.*, 27, 293.  
 WATABE, T. (1910) *J. infect. Dis.*, 7, 159.  
 WEICHELBAUM A. (1887) *Fortschr. Med.*, 5, 573, 620.  
 WENTHEIM, E. (1891) *Diach. med. Wochr.*, 17, 1351.  
 WHERRY W. B. and OLIVER, W. W. (1916) *J. infect. Dis.*, 19, 253.  
 WILSON, G. S. and SMITH, M. M. (1923) *J. Path. Bact.*, 31, 59.  
 WILSON, S. P. (1923) *J. Path. Bact.*, 31, 47.  
 WOLLSTEIN M. (1907) *J. exp. Med.*, 9, 588.  
 WRIGHT H. D. (1933) *J. Path. Bact.*, 37, 25.  
 ZDRODOWSKI, P. and VOROVINE, E. (1932) *Ann. Inst. Pasteur*, 48, 61.  
 ZOZAYA, J. (1931) *J. exp. Med.*, 54, 723.  
 ZOZAYA J. and WOOD, J. E. (1932) *J. infect. Dis.*, 50, 177.



## CHAPTER 24

### STREPTOCOCCUS

#### DEFINITION—*Streptococcus*

Spherical or ovoid cells, arranged in short or long chains, or in pairs. Usually non motile. Non sporing. Most species Gram positive. Some species form capsules. Growth tends to be relatively slight on artificial media, and some species grow poorly in the absence of added native proteïn. Several species produce characteristic changes in media containing blood. Various carbohydrates are fermented with the production of acid. Most species fail to liquefy gelatin. Most species are aerobic and facultatively anaerobic, some are anaerobic. Many species are normally parasitic on man or animals, some species are highly pathogenic, and some produce soluble toxins.

Type species *Streptococcus pyogenes*

The term *Streptococcus* was first used by Rosenbach in 1884, when describing a coccus, growing in chains, that had been isolated from suppurative lesions in man. To this organism he gave the name *Streptococcus pyogenes*. A chain-forming coccus had, however, been described by Fehleisen in the previous year as the causative organism of erysipelas (Fehleisen 1883), and Pasteur, Chamberland and Roux, in 1881, had described a septicæmic infection in rabbits, resulting from the inoculation of these animals with human saliva, which probably affords the earliest recorded reference to the pneumococcus, although no clearly identifiable description of this species was published prior to the independent studies of Fraenkel and of Weichselbaum in 1886. In 1887 Nocard and Mollereau reported the experimental production of mastitis in the cow and goat, by the inoculation into the udder of a streptococcus isolated from the milk of a cow suffering from that disease. In 1888 Schutz described a streptococcus that he had isolated from the lesions of strangles in the horse. In more recent years, chain forming cocci have been isolated from a variety of pathological conditions in man and animals, from the mouth or from the feces of healthy subjects, from milk and various milk products, and from other sources.

The tendency to grow in chains of varying length gives to the members of this group a very characteristic morphology, and they possess in common other characters that appear to justify their inclusion in a single bacterial genus. The Committee of the Society of American Bacteriologists (Winslow *et al* 1920) separated the pneumococcus from the main streptococcal group, by forming a genus *Diplococcus*, with *Diplococcus pneumoniae* as the type species. It appears to the authors, for reasons which will become apparent, that this separation is undesirable, and the summary of generic characters as set out by the American Committee has been modified in the required sense. As so modified, and with other minor emendations, including the substitution of *Str. pyogenes* for *Str. hemolyticus* as the name of the type species, the description of these generic characters is as given above.

The many attempts that have been made to evolve a satisfactory classification of the streptococci provide an admirable example of the difficulties with which the systematic bacteriologist is faced. As will be seen, there is no single criterion on which reliance can be placed, even in making a primary division into sub-groups that are themselves to be further divided by the application of other tests. It happens that, in this particular genus, one of the most useful differential criteria is provided by the changes induced by the growing organisms in media containing blood, but we shall note that this test, valuable as it is, cannot be too rigidly applied. Here, as elsewhere, we have to apply a variety of criteria selected, on the usual basis of statistical empiricism, as differentiating between groups each of which possesses several highly correlated characters. Here, as elsewhere, we find

that the method of antigenic analysis is playing an increasingly important part in the differentiation and identification of those ultimate types, or varieties, for which we need distinguishing names or labels.

The most convenient method of discussing this problem will be to take various characters in turn, and see how far they enable us to differentiate between one species, or type, and another.

**Morphology**—Taking the genus as a whole, the characters that might be regarded as supplying possible differential criteria are (1) the length of the chains formed, (2) the shape of the individual cells forming them, (3) the

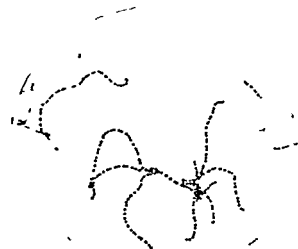


FIG 117—*Str. pyogenes*

From 24 hours culture on agar showing long chains  
( $\times 1000$ )

presence or absence of capsules, and (4) in the light of certain recent observations, the very occasional presence of flagella.

In the earlier days of bacteriology much attention was paid to chain formation as a differential criterion, and such names as *Streptococcus brevis*, *Streptococcus longus*, *Streptococcus longissimus*, and *Streptococcus conglomeratus* were coined to denote strains with the corresponding tendencies to grow in short chains, long chains, or chains which were very long or tangled. It has, however, come to be generally recognized (Thalman 1912, Brown 1919) that these characters are often variable within a single strain, so that, while the modal chain length of any species may be sufficiently characteristic to deserve inclusion in a description of the specific characters, it is quite useless for purposes of classification, and misleading when employed for purposes of nomenclature. Some species or groups, such as *Str. pneumoniae* and the enterococcus, usually occur in pairs or very short chains and never form chains of any considerable length. Others, such as *Str. pyogenes*,

usually occur in chains of eight or more cocci and often form long chains. Others again, such as the strains of the viridans type, are markedly variable in this respect, occurring both as very short chains, or pairs, and as chains of enormous length.



Fig. 118—*Viridans streptococci*.

From 24 hours culture in broth, showing short chains ( $\times 1000$ )



Fig. 119—*Str. pneumoniae*

From 24 hours culture on agar showing diplococci and short chains ( $\times 1000$ )

The shape of the individual cocci forming the chain also varies and there is a tendency for the cells of short-chained streptococci to be ovoid, with the long axis in the axis of the chain. When, as in the pneumococcus, a majority of the cocci occur in pairs, each cell of a pair may be definitely lanceolate, the blunt ends being adjacent. The individual cells of long chained streptococci tend to approach more closely to the spherical form, or they may sometimes be compressed in such a way that the longer axis of the cell lies at right angles to the axis of the chain as a whole. Cell shape, like modal chain length, may vary from one species of streptococcus to another, but it forms no better criterion for systematic purposes. Occasional strains may take on a diphtheroid type of morphology, which may lead to errors in identification (see Lamanna 1944).

Capsule formation, when it occurs, is of greater value for classification. It is almost constantly shown by *Str. pneumoniae* when growing within the tissues (see Fig. 120) and is absent in most other species, though not in all. Seastone (1931, 1913) however, has reported the presence of capsulation in *Str. pyogenes* during the first 2-2½ hours of growth in serum broth, though the capsules have usually disappeared by the 3rd or 4th hour. Morison (1940) likewise finds that all recently isolated strains of this organism are

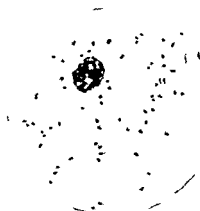


Fig. 120—*Str. pneumoniae*

In peritoneal exudate of mouse showing capsulation ( $\times 1000$ ).

capsulated to some degree. Capsulation is, however, very irregular, occurring in some types and not in others, and seems to be affected by the amount of hyaluronic acid in the substrate and the amount of hyaluronidase secreted. Generally speaking, capsulation and the secretion of hyaluronidase appear to be mutually exclusive (McClellan 1911, 1912), and there is no constant association of capsulation with virulence.

From time to time, accounts have appeared in the literature of motile streptococci, but many of these have been based on somewhat inadequate observations, and the genus *Streptococcus* has, until recently, been regarded by most bacteriologists as composed exclusively of non flagellated species. It is certain that flagellated forms are very rare, but two observers (Koblmüller 1935, Pownall 1935) have recently given careful and detailed descriptions of motile strains of streptococci. In each case the organism described was apparently a motile variant of the species, or group, that is generally known by the name of "enterococcus."

Streptococci stain readily with the ordinary dyes, none of them is acid fast, and the great majority are frankly Gram positive. Some strains tend to lose the Gram stain if decolorization is prolonged, a few species or varieties have been described as frankly Gram negative.

**Cultural Requirements**—Some species of streptococci, such as *Str. pneumoniae* and to a less extent *Str. pyogenes*, grow poorly on the simpler media of the laboratory when first isolated, though they can usually be trained to grow on these media after a limited number of subcultures. The growth of these species is markedly improved by the addition to the medium of such materials as blood or serum. A few species or types, such as the enterococci, grow well in the presence of bile or bile salts, while most do not.

The optimal temperature for growth is, with most parasitic species, in the neighbourhood of 37° C. Some species found in milk have an optimum about 30° C. The range over which growth occurs is, for the more sensitive species, somewhat restricted, 42° C. or thereabouts marks the upper limit, growth is usually slow at temperatures below 30° C. and often ceases below 20° C. Some species, on the other hand, including certain streptococci found in milk, grow actively at temperatures of 45° C. or over (see Sherman and Stark 1931). Such thermophilic types are of considerable economic importance in relation to pasteurization (see Chapter 93).

Most species are aerobic and facultatively anaerobic. Some are strict anaerobes, or microaerophilic. It will be more convenient to discuss the other characters of these anaerobic streptococci, which have not yet been studied in any detail, in a separate section of this chapter (p. 596). Pneumococci grow best in an atmosphere containing 10 per cent. carbon dioxide, some strains are said not to grow at all in its absence (see Fleming 1941, Kempner and Schlayer 1942). Glutamine is said to be required for the growth of Group A but not of Group B hæmolytic streptococci (Fildes and Gladstone 1939). For a fuller account of the growth requirements see p. 66.

These cultural requirements, while supplying useful ancillary evidence in identification, do not, except in the case of the anaerobic species and the markedly thermophilic streptococci, afford an adequate basis for any primary classification.

**Growth Characters**—On solid media the streptococci tend to form small, discrete, slightly raised colonies, 1 mm. or less in diameter. The modal colonial form varies in different species, and it may in some cases be sufficiently characteristic to assist

in identification, particularly in recently isolated strains. Many strains of pneumococci, for instance, give characteristic "draughtsman" colonies, with an entire, sharply raised edge and a central depressed area. A Type III pneumococcus frequently gives a characteristic watery, or mucoid, colony. One type of hæmolytic streptococcus gives easily recognizable minute, clear colonies, and so on.

Many species grow poorly in gelatin, and in a gelatin stab such growth as occurs is mainly confined to the track of the needle. The only streptococci, apart from some of the anaerobic species, that are known to liquefy gelatin are certain strains of enterococci. When liquefaction occurs it is usually infundibuliform in type.

In broth or other liquid media many species of streptococci give a granular growth, the medium remaining clear and the granular masses collecting as a powdery deposit, or adhering to the sides of the tube. Although this type of growth shows a characteristically high frequency among certain species of streptococci, especially when first isolated, it is by no means constant, and the degree of granularity may vary over a wide range. In some cases a granular deposit may be associated with a

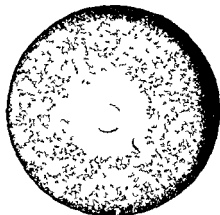


FIG 121—*Str pyogenes*

Surface colony on blood agar plate, showing zone of hemolysis round colony ( $\times 8$ )

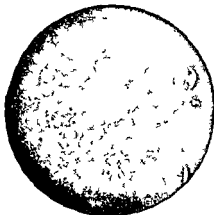


FIG 122—*Str pneumoniae*

Surface colonies on blood agar plate showing zones of discoloration and partial hemolysis round colonies ( $\times 8$ )

varying degree of turbidity of the medium, in others the growth, as a whole, may be distinctly though finely granular, the granules remaining dispersed throughout the medium. Any one strain may undergo marked changes in this respect on subculture, if a strain which, on first isolation, gives a markedly granular growth, is subjected to repeated subculturing at short intervals, it is often possible to produce a diffuse growth within a limited number of generations. Granularity is usually more evident in cultures grown at  $37^{\circ}\text{C}$  than at  $22^{\circ}\text{C}$ . In all cases the type of growth in a fluid medium is closely associated with the character of chain formation. A strain which is forming long chains will give a typically granular growth, if the growth becomes diffuse it will be found that the average chain length has diminished. Those species in which the diplococcal form predominates, as for instance *Str pneumoniae* or enterococci, usually give diffuse, non granular growths in fluid media. A description of the variations in morphological appearance, colonial form, and type of growth in broth met with among the streptococci is given on page 591.

**Metabolic Activities**—Most of the early attempts to classify the streptococci were based on their fermentation reactions in an empirically selected series of substrates. The behaviour of the different members of this genus in blood containing media has, however, come to occupy so important a place as a criterion for primary classification, that it will be convenient to deal with it first.

#### Classification Based on Changes Produced in Blood.

**Blood Plates**—Marmorek (1895) first noted the ability of certain strains of streptococci to lyse red blood corpuscles *in vivo* and *in vitro*, but it was Schottmüller (1903) who proposed that the ability to produce hæmolytic *in vitro* should be adopted as a differential criterion for purposes of classification. He noted that certain strains of streptococci produced clear zones of lysis when grown on blood agar plates, while others gave colonies which were surrounded by zones of greenish discoloration. For the former type Schottmüller proposed the name *Str. hæmolyticus*, for the latter *Str. viridans*. These observations formed the basis of a system of classification and nomenclature which has been developed by many



FIG. 123—*Str. pyogenes*

Deep colony in blood agar plate showing wide zone of complete hæmolytic and the sharply differentiated margin of the colony— $\beta$  hæmolytic ( $\times 8$ )



FIG. 124—Viridans streptococci.

Deep colonies in blood agar plate showing zone of discoloured cells round colony, obscuring margin, and zone of incomplete lysis beyond— $\alpha$  hæmolytic ( $\times 8$ )

subsequent investigators. It has, indeed, been shown that the green producing streptococci are not devoid of hæmolytic activity, though the zones which they produce on blood agar plates are different in kind, as well as in extent, from those formed round the colonies of the frankly hæmolytic strains. Mandelbaum (1907) emphasized the importance of microscopical examination of the colonies formed on blood agar plates, and noted that, while the colonies of the long pathogenic streptococci were surrounded by clear colourless zones, those of the viridans type, and also those of the pneumococcus, were surrounded by a zone of discoloured, non hæmolyzed corpuscles lying immediately next the colony, and an outer narrow hæmolyzed zone containing only corpuscular shadows.

These phenomena were studied in much greater detail by Smith and Brown (1915), and by Brown (1919). The monograph by Brown, in which the appearances met with and the factors which determine or modify them are fully discussed, contains an admirable review of the literature dealing with the classification of streptococci up to 1919. In this monograph great emphasis is laid on the importance of employing a uniform and standardized technique, and, in particular, on the

superiority of poured plates and the observation of deep colonies, over plates which have been inoculated by surface spreading only

The medium recommended by Brown consists of veal peptone agar containing 5 per cent of horse blood. The agar is stored in tubes in 12 ml amounts, when required for use a tube is melted and cooled to 45° C, 0.66 ml of horse blood is added and evenly mixed with the agar, the medium is inoculated with a loop or two of a 24 hours' broth culture that has been so diluted as to give about 100 colonies and is then poured into a Petri dish 9 cm in diameter thus giving a layer about 2 mm thick. The plates after preliminary drying in the inverted position with the plate tilted on the lid are incubated at 37° C for 24 hours. They are then examined and the appearances noted. They are re-examined after another 24 hours' incubation at 37° C and finally after a further 24 hours in the ice chest. For the recognition of hemolytic and non hemolytic strains in primary mixed cultures the use of blood agar plates with a surface inoculation usually suffices and has certain advantages, but for the critical determination of the type of hemolysis given by any strain once it has been isolated in pure culture the technique recommended by Brown should be strictly adhered to at least in regard to the medium used and the examination of deep colonies. The use of horse blood is particularly important. It is well known that the red corpuscles of different animal species vary widely in their resistance to different hemolytic agents. In spite of this fact many workers have used the blood of the rabbit or ox or some other animal, in testing the hemolytic activity of various species or strains of streptococci, and it is probable that some at least of the discrepancies met with in the literature are due to this variable factor. It may, at times be desirable to use the blood or red cells of some particular species in a given series of observations on streptococcal hemolysis, but if the results are to be used for purposes of identification or classification they should always be controlled by parallel tests made with horse blood.

Brown records four different types of reaction in blood agar plates, which he designates as follows

$\alpha$  A somewhat greenish discoloration and partial hemolysis of the blood corpuscles immediately surrounding the colony, forming a rather indefinitely bounded zone 1-2 mm in diameter outside of which is a second, narrow, clearer not discoloured zone. Under the microscope many corpuscles are seen in the inner zone and these are obviously discoloured, the discoloration varying in degree with different strains of streptococci. Very few corpuscles remain in the outer clearer zone, and these are never discoloured. These typical appearances may fail to appear after 24 hours, or even after 48 hours incubation, at the end of which time the narrow outer zone of hemolysis may not have developed. In such cases this zone makes its appearance during the subsequent 24 hours in the ice chest. If a plate which has developed the typical appearances, is reincubated for 24 to 48 hours and then placed in the ice chest for a further 24 hours a double series of rings will frequently develop so that the colony is surrounded by a hazy discoloured ring, a clear hemolysed ring, a second hazy ring and a second clear ring. By repeating the whole process it is sometimes possible to develop three or more series of such rings.

$\beta$  The colonies are surrounded by sharply defined clear colourless zones of hemolysis 2-4 mm in diameter. Under the microscope no corpuscles can be seen within this zone. The zones of  $\beta$  hemolysis develop more rapidly than those of the  $\alpha$  type. They are often well developed after 18 hours incubation. They extend slightly between the 24th and 48th hour but show no qualitative changes. They undergo no alteration or extension during the subsequent 24 hours in the ice chest.

$\alpha$  ( $\alpha$  prime) The colonies are surrounded by a zone of hemolysis which is slightly hazy and less sharply limited than in the case of true  $\beta$  hemolysis. The colony itself is not sharply defined and examination with the microscope shows that the hemolysed zone contains throughout, a moderate number of unaltered corpuscles which are most numerous

in the immediate neighbourhood of the colony. There is no discoloration. Unlike the zones of  $\beta$  hæmolytic a considerable extension of the zones may occur during the 24 hours in the ice-box. It is noted that some strains which produce  $\alpha'$  hæmolytic on horse blood agar may produce typical  $\alpha$  hæmolytic on rabbit blood agar.

$\gamma$  The colonies develop in the blood agar without any change in the surrounding medium.

The  $\beta$  hæmolytic strains of Smith and Brown correspond to Schottmüller's *Str. hæmolyticus*. The  $\alpha$  hæmolytic strains may be regarded as equivalent to his *Str. viridans*, though it may be noted that strains are encountered that produce the characteristic green coloration without the formation of a detectable zone of hæmolytic. The significance of the  $\alpha'$  type of hæmolytic is not clear. It seems to be of infrequent occurrence. There seems no good reason for attaching the label  $\gamma$  to those streptococci that cause no change in blood media. Strains of this type have sometimes been referred to as "indifferent streptococci."

**Soluble  $\beta$ -hæmolytic.**—The terms  $\alpha$  and  $\beta$  hæmolytic have attained general currency in bacteriological literature and serve a useful purpose. No effort has been made however to reconcile this usage with that of the terms "hæmolytic" and "non hæmolytic" as applied to streptococci. By a "hæmolytic" streptococcus is meant almost always a strain that causes  $\beta$  hæmolytic on blood agar. By a "non hæmolytic streptococcus" is meant a strain that either produces  $\alpha$  hæmolytic, or gives rise to no change at all. Some workers would confine the term "hæmolytic streptococcus," or at least the specific name *Str. hæmolyticus* if that be used, to strains that, in addition to causing  $\beta$  hæmolytic in blood agar, can be shown to produce a soluble hæmolytic, and this is an aspect of the problem that must be discussed in more detail.

Marmorek (1895) showed that cultures of certain strains of streptococci in a fluid medium had the power of lysing added blood corpuscles, and Besredka (1901) obtained hæmolytic filtrates from cultures of streptococci in heated rabbit serum. Braun (1912) reported that all strains of streptococci which produced hæmolytic on rabbit blood agar plates gave rise to a filtrable hæmolytic when grown in rabbit serum broth. The factors which determine this hæmolytic production have been studied by McLeod (1912), Meader and Robinson (1920) and de Kruijff and Ireland (1920). All these observers noted that hæmolytic production was absent or minimal in cultures grown in plain broth without the addition of serum. More recently Todd (1932) and Todd and Hewitt (1932) have shown that potent hæmolytic filtrates may be obtained by growing hæmolytic streptococci in a medium containing yeast extract, or in a special broth medium, sterilized by filtration instead of by autoclaving and containing dextrose, sodium bicarbonate and sodium phosphate.

De Kruijff and Ireland have carried out careful quantitative studies of the rate of hæmolytic production, and of its inactivation in the culture medium. They found that the hæmolytic titre of the supernatant fluid from their cultures after centrifugalization reached its maximum after 8 hours incubation at 37° C., and then rapidly declined. In many cases no hæmolytic could be detected after 14 hours, though, when the whole culture was tested instead of the supernatant fluid, some lytic action might persist up to the 24th hour. A hæmolytic filtrate is completely inactivated by heating at 55° C. for 30 minutes, and may lose most of its activity when incubated at 37° C. for 2 hours or more (see McLeod 1912).

It has been shown by Neill and Mallory (1926) that streptolysin, when exposed to air at relatively low temperatures, undergoes an oxidation that is readily reversible by suitable chemical reagents. The hæmolytic is active in the reduced form, inactive in the oxidized. At higher temperatures (55° C.) an irreversible inactivation occurs.

Todd (1934, 1935, 1939) has demonstrated the production of two kinds of hæmolytic. One, the O lysin, is oxygen-labile at ordinary temperatures, but can be re-activated by



reduction with 0.1 per cent sodium hydrosulphite, provided that it is protected from the air, it remains stable in the ice chest for years. The other, the S lysin, which can be extracted readily from streptococci by shaking with serum, is inactivated, like the O lysin, by incubation at 37° C for 2-4 hours but unlike the O lysin cannot be re-activated by reduction, it is very sensitive to both heat and acid and can be preserved only by storage at very low temperatures (-73° C). The O lysin produces hæmolysis rapidly and is antigenic in the free state. The S lysin produces hæmolysis more slowly and is antigenic only when present in the organisms. Each is neutralized by a separate antibody. The O lysin is formed by strains belonging to Group A, C (human) and G. The S lysin appears to be formed by strains of all groups, but the type of lysin produced is group specific, an antiserum to the S lysin of Group A strains failing to neutralize S lysin formed by other groups. Herbert and Todd (1944) have met with strains of streptococci producing only S and others only O hæmolysin. On blood agar the S strains formed  $\beta$  hæmolytic colonies on the surface and in the depth, both aerobically and anaerobically. The O strains on rabbit blood agar—but not on horse blood, which contains O antilysin—formed  $\beta$  hæmolytic colonies only in the depth and only under aerobic conditions. The S lysin was purified and found to be a lipo protein hapten, incapable of stimulating the formation of antibodies.

The extreme lability of both O and S lysins in broth cultures at 37° C renders the ordinary titration method of assessing their potency unreliable. Special precautions have to be taken to prevent destruction of either lysin before we can assert that any given strain which produces  $\beta$  hæmolysis on blood agar plates is incapable of forming a filtrable hæmolysin in a fluid medium.

Apart from the difficulties caused by the lability of the lysins, it may be noted that some strains of *Str. pyogenes* produce  $\alpha$  hæmolytic colonies on blood agar or sometimes completely non hæmolytic colonies.

The first type has been studied by various workers. Fry (1933) showed that certain strains which formed  $\alpha$  hæmolytic colonies on aerobic blood agar plates produced typical  $\beta$  hæmolytic colonies when incubated anaerobically. Fuller and Maxted (1939) showed that two or three factors were concerned in this result. Under aerobic conditions  $\beta$  hæmolytic colonies were formed if all reducing sugar was removed from the blood agar or if catalase was added to the medium. They bring evidence to suggest that in the  $\alpha$  hæmolytic colonies peroxide is formed before the hæmolysin with the result that a green zone is produced. If the formation of the peroxide is prevented by anaerobic incubation or is neutralized by catalase then the hæmolysin produces typical  $\beta$  hæmolytic colonies. The mode of action of the reducing sugar in the medium is not quite clear, but it appears to inhibit the formation of the hæmolysin. It may be noted that Fry's strains formed soluble hæmolysin in serum broth under aerobic conditions. For practical purposes it is advisable to incubate primary plate cultures anaerobically, or both aerobically and anaerobically.

The second type has been reported by Coburn and Paul (1941) and Colebrook and his colleagues (1942). In a ward outbreak of streptococcal respiratory infection, Coburn and Paul found that the causative organism Type 12, occurred in two forms. One form gave rise to typical hæmolytic colonies at 37° C, the other gave rise to hæmolytic colonies only at 22° C. The second variant appeared to be possessed of a higher degree of infectivity than the normal form. Colebrook's observations were made in a surgical ward in which typical  $\beta$  hæmolytic colonies belonging to Group A Type 12 had been giving rise to septic complications. A series of cases was studied in which completely non hæmolytic streptococci, both aerobically and anaerobically, were isolated from the wounds. These organisms proved also to belong to Group A Type 12. They formed no S hæmolysin, but most of them, when specially tested, were found to produce O hæmolysin. The frequency with which such strains occur is at present unknown, but they are believed to be uncommon.

$\alpha$ -haemolysin.—When we turn to those streptococci that produce  $\alpha$  haemolysis on blood agar plates, we have two mechanisms to consider the reaction that causes the lysis of the red cells, and the reaction that causes the green coloration.

Cole (1914) described the presence in pneumococci of a labile intracellular haemolysin which was liberated from the cells on autolysis. It was commonly stated by subsequent workers that the pneumococcus produced no filtrable haemolysin in fluid media, and the agent causing  $\alpha$  haemolysis was supposed to be of some quite different nature. It has, however, been quite clearly demonstrated within recent years (see Neill 1926, Sickles and Coffey 1928, Cowan 1934, Todd 1934) that the pneumococcus, when grown under suitable conditions, produces a soluble haemolysin of the oxygen sensitive, heat sensitive type, which undergoes reversible oxidation at low temperatures. Whether other streptococci that produce  $\alpha$  haemolysis would also elaborate a filtrable haemolysin under suitable conditions is at present unknown.

Until recently the most widely accepted view in regard to the green coloration associated with  $\alpha$  haemolysis was that it was due to the formation of methaemoglobin or of some closely allied substance (Schnabel 1921, McLeod and Gordon 1922, Rother 1925). It was shown by McLeod and Gordon that the pneumococcus produces hydrogen peroxide and that hydrogen peroxide will discolour heated blood agar, in which the blood catalase has been inactivated. The mechanisms involved in the production of hydrogen peroxide by the pneumococcus, or by pneumococcal extracts, have been studied in considerable detail by Avery, Morgan and Neill (Avery and Morgan 1924, Morgan and Avery 1924, Avery and Neill 1924a, b, c, Neill and Avery 1924a, b, 1925, Morgan and Neill 1924, Neill 1925). The systems involved appear to include the catalysed oxidation reduction and peroxidase mechanisms discussed in Chapter 3, and the actual course of the reaction seems to be determined in the main by the oxygen pressure to which the reacting system is exposed. It is, however, clear that the formation of methaemoglobin is not itself the cause of the green pigmentation, unless the apparent greenness is due to an optical illusion resulting from a colour contrast, and, in view of the amount of catalase present in unheated blood, it is difficult to believe that hydrogen peroxide is the active agent in cultures on unheated blood agar plates.

This problem has been brought nearer solution by the studies of Hart and Anderson (1933) (see also Anderson and Hart 1934a). Working with the pneumococcus, they found that when small quantities of laked blood were added to broth cultures in the presence of an alkaline buffer solution, an olive green precipitate was formed. This could be separated, washed, and dissolved in dilute alkali to give a green solution. Crystalline haemoglobin, or methaemoglobin gave the same green pigment when incubated under suitable conditions with pneumococcal cultures. The spectroscopic and chemical analysis of this pigment suggest that it is an iron-containing derivative of haemoglobin. It is rapidly bleached by hydrogen peroxide, but it is not affected by reducing agents. An identical, or very similar, green pigment can be obtained by incubating laked blood, haemoglobin, or methaemoglobin with various chemical reducing systems, such as ascorbic acid, cysteine glucose, etc. From the results obtained with autolysed bacterial cells, washed bacteria, bacterial extracts, etc., it would seem that the green pigment results from the activity of a bacterial oxidation reduction system, one component of which is intracellular. This system is not peculiar to the pneumococcus, it is shared, not only by those streptococci that produce the green pigment on blood agar plates, but by *Str. pyogenes* which gives  $\beta$  haemolysis, and by the enterococci which usually produce no change on unheated blood media. It is also possessed by unrelated bacteria, such as *Staph. aureus* and *Bact. coli*. The production of green pigmentation by some species and not by others would seem to be due, not so much to the presence or absence of the necessary enzyme system as to secondary factors, themselves determined by the metabolic activities of the bacteria concerned, which sometimes permit this system to function, and sometimes suppress it. We have already noted that some strains of haemolytic streptococci may produce  $\beta$  haemolysis under anaerobic

conditions while giving typical  $\alpha$  haemolysis with green coloration when cultivated aerobically

Summarizing the observations recorded above we may say that a study of the appearance of the colonies on blood agar plates together with a test for the production of a filtrable haemolysin enables us to divide streptococci into three main categories

(1) *Haemolytic streptococci*—These produce  $\beta$  haemolysis on blood agar plates. They may be differentiated into two sub groups (a) those that produce a filtrable haemolysin, and (b) those that do not. Among strains of *Str. pyogenes* two variants are known. One produces  $\alpha$  haemolytic colonies aerobically but  $\beta$  haemolytic colonies anaerobically. It forms a soluble haemolysin in broth aerobically. The other forms completely non haemolytic colonies but produces a soluble haemolysin of the O type.

(2) *Streptococci giving  $\alpha$  haemolysis*

(3) *Streptococci that have no action on blood media under the usual conditions of testing*

**Classification Based on Fermentation Reactions**—The capacity of different species or types of streptococci to ferment different substrates played a very large part in the earlier attempts to separate this group of bacteria into its natural elements. The classification of Andrewes and Horder (1906) based largely on the earlier studies of Gordon (1902-03 1903-04 1905) and the more extensive and detailed classification proposed by Holman (1916) (see also Floyd and Wolbach 1914 Lyall 1914, Broadhurst 1915) depended almost wholly on a selected series of fermentation tests. It is hardly necessary to-day to set out these classifications in their original form, since few would now accept them as affording an adequate basis for the differentiation of named species or types. This does not of course mean that these fermentation tests are of no value in classifying the streptococci. The reverse is the case. In more recent years, however, the tendency has been to employ fermentation reactions as ancillary rather than as primary differential criteria. When a particular substrate has been found to be of value in differentiating between certain related species or types it has been used for this purpose but has not necessarily been employed in the differentiation of other species within the genus. It will therefore be more convenient to consider the exact systematic significance of these fermentation tests after we have discussed certain other differential criteria and in particular the results obtained by antigenic analysis but it will be useful to summarize here the observations that have been made on the correlation between enzymic activities and natural habitat.

The early observations that streptococci isolated from pathogenic lesions in man usually ferment lactose and salicin seldom if ever mannitol raffinose or inulin and give acid without clot in milk has been amply confirmed. The value of inulin fermentation as a differential test for the identification of the pneumococcus has also been firmly established. It is clear that this species almost always ferments this fructosan while most other streptococci fail to do so. There has been a tendency to exclude from the species *Str. pneumoniae* any strain that fails to ferment inulin but it is doubtful as Berger and Silberstein (1926) point out whether this test can be applied with such complete rigidity. As regards other substrates the pneumococcus ferments lactose and usually raffinose but not salicin or mannitol. Milk is acidified and frequently clotted.

There is some measure of agreement that the streptococcus commonly found in the human mouth which is of the  $\alpha$  haemolytic type ferments lactose raffinose and salicin, but not mannitol and usually forms a clot in milk. In addition it has been pointed

out by Niven, Smiley and Sherman (1942) that, in general, the  $\alpha$ -haemolytic streptococci fail to hydrolyse arginine with the production of ammonia, whereas the  $\beta$ -haemolytic streptococci and the enterococci are able to do so.

There is also agreement that the fermentation of mannitol is characteristic of the streptococci that normally inhabit the human intestine (see Winslow and Palmer 1910, Fuller and Armstrong 1913, Hopkins and Lang 1914, Dible 1921, Bagger 1926, Meyer 1926, Meyer and Schönfeld 1926, Downie and Cruickshank 1928). These streptococci usually ferment salicin and lactose, seldom inulin or raffinose, and usually clot and decolorize litmus milk. A few of them are peculiar in liquefying gelatin. In relation to this group of streptococci an additional fermentation test was introduced by Rocharix (1924)—the fermentation of aesculin in a bile-containing medium. This reaction has been studied by Meyer and Schönfeld (1926) and by Weatherall and Dible (1929). It has been found by many workers to be of considerable differential value; but, as Weatherall and Dible point out, the inclusion of bile salts in the medium removes it from the ordinary category of fermentation tests, since bile salts inhibit the growth of many species of streptococci (see below) and in their absence aesculin is attacked, though often slowly, by many non-faecal species or types.

Comparing the streptococci of man with those from other animals, several points of interest have emerged.

Winslow and Palmer (1910), while confirming the frequency of mannitol-fermenting streptococci in human faeces, noted their rarity in faeces from the cow or horse. They found also that raffinose-fermenting strains, while relatively uncommon in human faeces, were very common in the faeces of cattle. Fuller and Armstrong (1913) examined 349 strains of streptococci isolated from faeces—123 from man, 129 from the horse and 97 from cattle. They found that 65 per cent. of the faecal streptococci from man fermented mannitol, as compared with 2-3 per cent. of the bovine or equine faecal strains. Raffinose fermenting streptococci, on the other hand, were not found among the human faecal strains, while 12 per cent. of the equine faecal strains, and 73 per cent. of the bovine faecal strains fermented this sugar. The equine faecal strains were characterized by a frequent failure to ferment lactose.

The fact that streptococci isolated from horses, whether from suppurative lesions or from the mouths or intestines of normal animals, frequently fail to ferment lactose has been noted by many observers (see Jones 1919).

The importance of milk as a human food, and of the cow as a stock animal, has led to a careful and detailed study of the streptococci that occur in normal milk, or in the milk from diseased animals, or that have been isolated from the udder in acute or chronic mastitis.

A type of streptococcus that is almost constantly present in milk, and has been given the name *Str. lactis*, resembles in many ways the streptococcus commonly found in human faeces; and it will therefore be more convenient to deal with its fermentation reactions when considering its probable relation to that organism in a later section.

In regard to those streptococci that are associated with mastitis in the cow, acute or chronic, certain additional criteria, based on fermentation reactions, have been introduced within recent years in an attempt to differentiate the characteristically bovine strains from those of human origin. Among these are (1) the final pH produced in glucose broth (Avery, R. C., and Cullen 1919, Ayers and Mudge 1922, Frost *et al.* 1927, Minett *et al.* 1929, Avery, R. C. 1929a, b, Minett and Stableforth 1931, 1934, Lancefield 1933, Hare and Colebrook 1934); and (2) the capacity to hydrolyse sodium hippurate (Ayers and Rupp 1922 and other references above). It has been found that mastitis strains of human origin produce a final pH of 5.0-5.6 in glucose broth, and fail to hydrolyse sodium hippurate. Bovine strains, on the other hand, produce a final pH of 4.2-4.8 in glucose broth, and hydrolyse sodium hippurate. It has also been found that the fermentation of trehalose and sorbitol afford a valuable criterion in the examination of a particular group of haemolytic streptococci that are found both in men and animals—mainly horses. The human strains in the group usually ferment trehalose but not sorbitol; the animal strains ferment sorbitol but not trehalose (Ogura 1929, Edwards 1932, 1933, Minett 1935a, b).

The behaviour of rough variants appears to be inconstant. Many probably most, rough strains are bile soluble (Griffith 1923 Peimann 1927 Downie *et al.* 1931) but some are less readily lysed than normal, smooth strains, and a few are apparently quite insoluble. It seems clear that this test properly performed is one of the most reliable at our disposal. A smooth strain that proves insoluble in sodium deoxycholate should not be given the title *Str. pneumoniae* without a clear statement that because of this important divergence from the specific characters, the diagnosis must be regarded as uncertain.

The use of bile has provided another differential test within this group. Weissenbach (1918) noted that enterococci grew well in a medium containing 10 per cent. of bile while *Str. pyogenes* and other streptococci tested by him failed to grow in this medium. The observation that enterococci grow freely in high concentrations of bile has been confirmed by many subsequent observers but it would appear that this character is not peculiar to this species, or group. The ability of other species and types to grow in bile-containing media (10 per cent. and 40 per cent.) has been studied by several workers (see Belenky and Popowa 1929 Minett and Stableforth 1931 Lancefield 1933 Hare and Colebrook 1934 Hare 1935 Hare and Maxted 1935 Colebrook, Maxted and Johns 1935). The possible significance of this character in the classification of streptococci other than the enterococci will be more conveniently discussed in relation to the results obtained by antigenic analysis.

Another test that has been applied in the classification of this group is the ability to reduce and thus decolorize methylene blue added to milk, usually in a concentration of 1:5000 (Avery R.C. 1929a, b). This test would appear to depend in part on the ability of the organism to multiply in the presence of methylene blue in part on the Eh that the growing organism induces. Many of the strains that reduce methylene blue have been isolated from animal sources or from milk. There is general agreement that strains of hemolytic streptococci isolated from severe human infections fail to do so. Enterococci, however, reduce methylene blue actively (Kleckner 1935), even in a concentration of 1:1000. Here again, the significance of this test must be considered in relation to the antigenic structure of the various species or types.

A fourth test that is of considerable value from the point of view of classification is that of heat resistance. It has long been known that heat resistant streptococci occur in milk (see Avers and Johnson 1910 1913 Avers, Johnson and Davis 1915) and Loran (1914) recorded the presence of heat resistant streptococci in the human faeces. Houston and McCloy (1916) noted that heat resistance was characteristic of enterococci, and this observation has been amply confirmed by Dible (1921) and by many subsequent observers. It is usual to employ exposure to a temperature of 60° C. for 30 to 35 minutes as an arbitrary test of heat resistance. The problem that arises in this connection is the identity or non-identity of the milk and faecal strains of heat resistant streptococci and this will be considered in a later section. Other tests which will be considered under the classification of the enterococci, comprise ability to grow at 45° C. at pH 9.6 and in the presence of 6.5 per cent. NaCl.

**Antigenic Structure** It will be convenient to begin our discussion of antigenic structure with the pneumococcus in part because this is a well-defined species in which the complications referred to above do not arise in part because it was in fact one of the first species in which a division into immunologically different races or types was clearly demonstrated and in part because the chemical study of antigenic structure was initiated with this organism and we still know far more about the chemical differences that determine the antigenic specificity of the various types of pneumococci than we do about similar differences in any other bacterial species.

**The Antigenic Structure of the Pneumococcus.**—Neufeld and Handel (1909) first demonstrated the existence of antigenically different types of pneumococci. They studied the protective effect of different antipneumococcal sera in mice and

found that a given serum would protect against the homologous strain of pneumococcus but not against heterologous strains. Dochez, Avery and their colleagues (Dochez and Gillespie 1913, Dochez and Avery 1915, Avery 1915, Avery *et al* 1917) later studied the antigenic relationships of a large collection of pneumococci using the methods of direct agglutination and agglutinin absorption. They confined their attention, for the most part, to strains isolated from cases of lobar pneumonia in man, and among these they were able to recognize three well-differentiated types, Types I, II and III, leaving a large heterogeneous group unclassified. Lister (1916) carried out a similar study on strains isolated from cases of pneumonia among the mine workers in South Africa. He was able to differentiate several antigenic types, which he labelled with letters instead of numbers. These observations have since been confirmed and extended by workers in many parts of the world, the three classical American types being generally accepted as the standard of reference, and it was not long before we obtained a reasonably adequate picture of the distribution of these three types in cases of lobar pneumonia, in other pneumococcal infections, in healthy contacts and in the population at large. For many years, however, no serious attempt was made to analyse the unclassified heterologous group which formed a considerable proportion (25-50 per cent) of the strains isolated from cases of lobar pneumonia, and the majority of those isolated from the upper respiratory tract of normal persons. It was the custom to refer to such strains as belonging to Group IV, an unfortunate nomenclature that became frankly misleading when the label was changed to "Type IV", which not infrequently happened.

Within recent years Cooper and her colleagues (Cooper, Edwards and Rowenstem 1929, Cooper *et al* 1932, Cooper and Walter 1935) have made a detailed study of this previously unclassified group. Among strains isolated from lobar and bronchopneumonia from various other pneumococcal infections and from normal persons they have identified 29 new antigenic types making 32 in all, including the classical Types I, II and III. Most of these types are, it should be noted, sharply differentiated from each other, so that they may be identified by direct agglutination. Since Cooper's study, further types have been described by other workers (see Kauffmann *et al* 1910, Walter *et al* 1911, Merch 1912). Unfortunately two systems of labelling have been used: the one making use of numbers irrespective of antigenic components shared with other types, the other using letters in addition to numbers to bring out antigenic components possessed by different types in common. Lally (1914) has described the cross reactions that may be met with, and has suggested the use of a series of Arabic numbers ranging from 1 to 74 to cover the types known at present. We may be quite sure that there are still new types of pneumococci to be identified, and new labels to be given, but it seems likely that our present 74 types include most of those that are parasitic in man. Gundel and Schwarz (1932), for instance, report that, of 364 strains of pneumococci, isolated from sick or healthy children or adults and containing no examples of the classical Types I, II and III, only 3 per cent were unassignable to one or other of Cooper's new Types IV-XXXII.

It has long been recognized that the antigenic behaviour of intact pneumococci in the normal smooth state is probably determined by the nature of the capsules surrounding the bacterial cells. Neufeld (1902) for instance noted that the capsules of pneumococci, when acted upon by a specific antiserum, become greatly swollen, and within recent years, it has been shown that this phenomenon can be utilized in the identification of pneumococcal types (Neufeld and Ftinger Tulczyńska 1931, Ftinger Tulczyńska 1932, Armstrong 1932, Logan and Smeall 1932, Sabin 1933, Beckler and MacLeod 1934, Cooper and Walter 1935).

There is also (Avery and Heidelberger 1923) a nucleo-protein antigenic component, precipitable from extracts by acetic acid. It is probably situated deeply within the intact bacterial cell. It is shared by all pneumococci and by many other bacteria including all those species of streptococci that have been examined.

The picture of the antigenic structure of the species *Str. pneumoniae* that emerges from these studies may be tentatively outlined as follows. There is a central protoplasmic portion of the cell which, in its antigenic relationships is neither species nor type specific. Situated probably at the cell surface, there is another component, mainly carbohydrate in nature, but containing nitrogen and phosphorus, that is specific for *Str. pneumoniae* as a species. External to this, in the normal smooth forms, there is a capsule, composed wholly or in part of a polysaccharide that is specific for each pneumococcal type. There are, we must suppose, over seventy of these capsular polysaccharides within the pneumococcal species, probably there are many more. The antigenic behaviour, and to some extent the virulence, of the intact pneumococcal cells are, it should be noted, determined by these capsular antigenic components, so that they are of particular importance to the medical bacteriologist.

**The Antigenic Structure of the Hæmolytic Streptococci.**—When we come to study the antigenic structure of the hæmolytic streptococci we are on much more difficult and debatable ground, in part because, as we have already indicated it is by no means easy to define exactly what we mean by a hæmolytic streptococcus, in part because, if we accept the usual definition—the occurrence of  $\beta$  hæmolysis on a blood agar plate—we shall include in our hæmolytic group, not one species, but several, in part because the technical difficulties of antigenic analysis are far greater than in the case of the pneumococcus. In spite of all these difficulties a great advance in our knowledge has been made during recent years, and, though we cannot as yet present any clear and detailed picture, we can provide a sketch plan which, with the necessary modifications, will certainly provide the basis for any future, and more complete, classification.

To obtain a clear picture of the present position, it will be better to disregard the sequence in which our knowledge has been reached.

**Group Relationships.**—The observations of Hitchcock (1924) and of Lancefield (1928, 1933, 1941) have revealed the presence of a number of different serologically active polysaccharides in hæmolytic streptococci from different sources. Instead of being responsible, as in pneumococci, for type-specificity, each polysaccharide is common to a group of organisms derived from a particular source. Thus, the majority of strains isolated from pathological lesions in man share the same polysaccharide and are classified as Group A. Strains from mastitis in the cow possess another polysaccharide and are classified as Group B. Strains from infections in lower animals possess still another polysaccharide and are classified as Group C and so on. Altogether 12 groups have now been recognized, each with its peculiar polysaccharide antigen (Table 37).

The association between the type of polysaccharide and the source of the strains is close, but not absolute. Human beings, for example, may be infected occasionally with organisms belonging to Groups B, C, D, and G, though Group A strains far outnumber the rest. Moreover, when infection with these other types occurs, it is often confined to one part of the body. Thus, Group B strains are seldom found except in infections of the female genital tract, Group D strains are restricted mainly to cystitis and wound infections, and Group G strains to genito-urinary and occasional throat infections.

A great advance was made in the study of the antigenic structure of the pneumococci and indeed of bacteria in general when Avery Heidelberg and their colleagues attacked the problem from the chemical side (Heidelberg and Avery 1923 1924 Avery and Heidelberg 1923 1925 Avery et al 1925 Heidelberg et al 1925 Avery and Morgan 1925 Heidelberg 1927 Heidelberg and Coebel 1927). By suitable methods of extraction followed by fractional precipitation it was found possible to separate the capsular components that determine type specificity in a state of chemical purity. These components were found to be complex polysaccharides, and some of the main chemical and physical characters of the capsular polysaccharides of the three classical types have been determined (see Table 36). Solutions of these polysaccharides it will be noted give specific precipitation in high dilution when mixed with the corresponding antisera.

TABLE 36

CHARACTERS OF THE TYPE SPECIFIC ANTIGENS OF PNEUMOCOCCI AFTER AVERY AND HEIDELBERGER

Type	Optical Rotation.	Per cent Nitrogen	Substances obtained on Hydrolysis.	Dilution giving Specific Precipitation.
I	+ 300°	5	Galacturonic acid, and amino-sugar derivative	1:6 000 000
II	+ 74°	0	Glucose	1:5 000 000
III	- 33°	0	Glucose and Glucuronic acid	1:6 000 000

The chemical constitution of the capsular antigens of the new types of pneumococci differentiated by Cooper and her colleagues has yet to be determined though a start has been made with this work (see Heidelberg and Kendall 1931 Brown and Robinson 1943). We may however safely assume that each pneumococcal type is characterized by a specific capsular polysaccharide that determines its antigenic behaviour. We may identify these types either by agglutination or by the capsule-swelling reaction or by precipitation tests carried out with an autolysate or extract of the pneumococcal cells.

Some at least of these pneumococcal polysaccharides may exist in immunologically different forms or may be altered during the process of chemical extraction and purification. Thus the studies of Enders (1930) and of Wadsworth and Brown (1931) showed the presence in Type I pneumococci of a specific antigenic component that differed in its immunological reactions from the specific capsular polysaccharide as ordinarily prepared. Avery and Goebel (1933) were able to show that this component is an acetylated form of the Type I capsular polysaccharide and that it is apparently in this form that the polysaccharide exists in the normal bacterial cell. The acetyl groups are removed by the methods of extraction and purification that had been commonly employed leaving a deacetylated polysaccharide that is still specific for the Type I pneumococcus, but has lost certain antigenic activities possessed by the normal acetylated form. It is important to note that the type-specificity of the capsular antigen is not destroyed by this particular chemical change, and it seems quite likely that similar minor alterations in chemical structure may be induced during the extraction and purification of many other antigenic components.

This polysaccharide capsular component is not of course the only antigenic constituent of the pneumococcal cell. Tillet, Goebel and Avery (1930) have isolated another component that gives all the usual reactions of a polysaccharide, is not inactivated by peptic or tryptic digestion and yields about 30 per cent of reducing sugar on hydrolysis. It contains about 6.1 per cent of nitrogen and differs from the capsular polysaccharides in containing phosphoric acid. It is not type specific, but appears to characterize the pneumococcus as a species.



There is also (Avery and Heidelberger 1923) a nucleo protein antigenic component precipitable from extracts by acetic acid. It is probably situated deeply within the intact bacterial cell. It is shared by all pneumococci and by many other bacteria including all those species of streptococci that have been examined.

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TABLE 37

GROUP AND TYPE SPECIFIC ANTIGENS OF HÆMOLYTIC STREPTOCOCCI (MODIFIED FROM LANCEFIELD 1941)

Group Specificity		Type Specificity		
Serological group	Group-specific Carbohydrate substances.	Specific types recognized.	(Type-specific substances).	
			Designated	Chemical nature.
A	Polysaccharides immunologically distinct for each group	At least 30	"M" "T"	Proteins Undetermined
B		Several	S	Polysaccharides
C		Several	—	Proteins
D, E, F, G, H, K, L, M, N		Several	S	Polysaccharides

The association between the type of polysaccharide and the hæmolytic power of the organisms is variable. So far as we know most Group A strains give rise to typical  $\beta$  hæmolytic colonies on blood agar plates but as already pointed out a few strains have been reported that are hæmolytic only under anaerobic conditions or are frankly non hæmolytic. In human infections we are generally on safe ground in disregarding non hæmolytic streptococcal colonies except under very unusual conditions they are unlikely to be responsible for the ordinary manifestations of hæmolytic streptococcal infections. In Group B however the position is different. Stableforth (1932) found that about half the bovine strains he studied were non hæmolytic even though he could detect no immunological difference between the hæmolytic and the non hæmolytic members of the group. Among the other groups non hæmolytic strains have been found with varying frequency. Special strains in Group C produce  $\alpha$  hæmolytic colonies and one group Group N comprising strains of *Str. lactis* appears to give rise to uniformly non hæmolytic colonies. The evidence suggests that the possession of the group polysaccharide is a more constant and fundamental character than the ability to lyse blood cells. It is therefore becoming common to pay greater attention to the type of polysaccharide present in a given strain than to its hæmolytic power though in practice especially in medical laboratories the observation of hæmolytic colonies constitutes the first important differential criterion of presumptive pathogenicity.

The polysaccharides themselves which are sometimes referred to as C substances appear to form an integral part of the bacterial cell. To obtain them in solution the organisms have to be disrupted. For this purpose either Lancefield's (1933) acid-extraction method or Fuller's (1938) formamide method may be used. Their presence can be demonstrated by a simple precipitation reaction using specially prepared antiserum. They seem to play no part in the agglutination of streptococci. There is little information yet on their chemical structure but the polysaccharide of Group A strains is known to yield reducing sugars on hydrolysis to be non toxic for animals and to be antigenic only when in combination with the cellular protein.

#### Type Relationships

*Group A*—The findings recorded by Dochez, Avery and Lancefield (1919), Bliss (1920, 1922), Gordon (1921), Eagles (1924) and Stevens and Dochez (1925a, b)

made it possible to differentiate and identify many different antigenic types among hæmolytic streptococci isolated from human infections, and the more detailed and extensive studies of Griffith (1926, 1927, 1928, 1934, 1935), Smith (1926, 1927), James (1926), McLachlan and Mackie (1928), Gunn and Griffith (1928), and Dora Colebrook (1935) have served to establish many of these types on a satisfactory serological basis. Griffith, who has been the main contributor to this particular problem, has differentiated by means of the slide agglutination reaction 27 types of pathogenic hæmolytic streptococci isolated from various lesions in man. Of these, 23 were later found to be members of Group A, Types 7, 20 and 21 fell into Group C, and Type 16 into Group G. Before he died, Griffith added three more types to Group A, bringing the total numbered types up to thirty. Further types have since been added.

Lancefield (1928, 1933) attacked the problem from another angle. She extracted type specific antigens from the cocci by hot acid, and identified them by the precipitin test. On the whole, her results agreed with those of Griffith, but some discrepancies occurred. Further observations (Lancefield 1940, 1943, Swift *et al* 1943, Lancefield and Stewart 1944, Watson and Lancefield 1944, Stewart *et al* 1944, Zittle 1942, Elliott 1943, Krumwiede 1943) have shown that two separate antigens are concerned in the type specificity of Group A strains. One referred to as M, is a protein; the other, referred to as T, is of undetermined nature. The M antigen is present only in mucoid or matt colonies (see p. 595) and is most abundant in freshly isolated pathogenic strains; the T antigen is found not only in mucoid and matt, but also in the degraded glossy colonies containing avirulent organisms. Antibody to M seems to be responsible for the M precipitin reaction, for type-specific protection, and as a rule for part of the type-specific agglutination of matt variants. Antibody to T appears to be solely responsible for type specific agglutination of glossy variants and mainly responsible for type specific agglutination of matt variants, but to be unconcerned in protection. The M antigen can be destroyed by peptic or tryptic digestion; heat killed enzyme treated organisms stimulate the production of T antibodies alone, so that pure T antibody can be obtained and the distribution of T antigen determined. In most types M and T antigens occur together, but in a few types one or other is missing or is shared with some other type—hence the occasional discrepancies between the results of agglutination and precipitation tests.

For example, Types 10 and 12 contain serologically identical M antigens but different T antigens. Closely related T antigens occur in Types 15, 17, 19, 23 and 30 and another set of related T antigens occurs in Types 4, 24, 26, 28, 29, and 46. In some types strains are encountered that contain no T antigen at all. Moreover, the amount of M and T antigen that is formed or at least that can be detected by agglutination methods depends on the temperature of incubation. There is evidence to suggest that the M substance is formed best at 37° C., but is liable to inactivation at this temperature—possibly through enzyme action (Elliott 1944).

An additional complication to the precipitin reaction is that extracts may contain variable amounts of non specific nucleo-proteins, which are common to hæmolytic streptococci, pneumococci, and streptococci of the viridans group. It may also be noted that there is at least one other non type specific protein ('Y') about which little is as yet known. It will therefore be clear that the type determination of Group A streptococci is far from straightforward, nevertheless their general antigenic constitution is now fairly well understood.

**Group B**—The existence of specific types in Group B was first established by Stableforth (1932), who found three serological types among strains from bovine mastitis, these were sharply distinguished by their precipitation and agglutination reactions. The type-specific antigen was found by Lancefield (1934, 1938) to be, not a protein as in Group A, but a polysaccharide. By precipitation tests she was able to define four specific types. British and Australian investigators (Stableforth 1937, Stewart 1937, Simmons and Keogh 1940) have carried this study further by separating off, mainly by the agglutination and absorption of agglutinins technique, a number of sub-types. What the antigen or antigens concerned in the agglutination test may be is still a matter for conjecture, but so far as the four main types are concerned, there seems to be agreement between the results of the precipitation and agglutination tests (Slanetz and Naghski 1940, Simmons and Keogh 1940). The notation is still confused, but there is much to be said for Stableforth's suggestion of numbering the main types, as in Group A, and giving small letters to the sub-types, e.g. Type 1a, 1b, 1c, 1d, 2a, 3a, 3b, etc. Again there is disagreement on the identity or non identity of Group B strains of human and bovine origin. Further work will be required before discrepancies of this sort can be cleared up.

**Group C**—It has already been mentioned that three of the types of hæmolytic streptococci identified by Griffith, namely Types 7, 20, and 21, belonged to Group C. Five more types have now been differentiated by the agglutination test among human strains (Simmons and Keogh 1940), and five among equine strains (Bazelev and Battle 1940). There is reason to believe that the type-specific antigen is of protein nature.

**Group D**—This group contains a number of organisms of the enterococcus type, the specific or varietal nature of which will be discussed later (p 582). By agglutination several types have been distinguished by Japanese and other workers (for references see Grumbach and Schnetz 1938, Ehrismann 1943). Lancefield (1941) states that she has been able to define three distinct types. The type-specific antigens appear to be of polysaccharide nature.

**Other Groups**—One type has been found in Group E (Lancefield 1941). In Group F Bliss (1937) established four types by precipitation and agglutination reactions, and in Group G one type, which was identical with F Type 1. Simmons and Keogh (1940) were able to distinguish a number of serological types among both the large-colony and the so-called minute forms of streptococci in Group G. One type has been found in Group K (Lancefield 1941).

**Antigenic Structure of the  $\alpha$ -Hæmolytic Streptococci**—We know little, as yet, about the antigenic structure of the many species and varieties of  $\alpha$  hæmolytic streptococci that have been described. It seems quite clear that the viridans group of streptococci contains no such polysaccharide group antigens as have been found in the hæmolytic streptococci. On the other hand, specific types can be recognized within certain species. Sherman, Niven and Smiley (1943), for example, using the precipitation technique, have found that strains of *Str. salivarius* fall into two main types and an unknown number of other types. Solowey (1942) likewise who studied 103 strains of viridans type from subacute bacterial endocarditis and 99 strains from human throats and extracted teeth, was able to distinguish at least eight different types. The systematic relationship between the different organisms has still to be worked out.

## Properties Classification and Nomenclature of the Hæmolytic Streptococci

Before describing the pathogenicity of the various streptococci it will be convenient to summarize the main properties of the different groups and to discuss the nomenclature of the organisms within each group. It must be emphasized once again that not all strains falling into Lancefield's groups form  $\beta$  hæmolytic colonies on blood agar. The heading of this section is not strictly accurate but until some more suitable term is proposed we shall include  $\alpha$  hæmolytic and non-hæmolytic organisms possessing the same group polysaccharide as hæmolytic organisms under the general designation of hæmolytic streptococci.

### Group A

*Streptococci* belonging to this group share in common the Group A carbohydrate component described by Lancefield. They are divisible into a number of different serological types either by agglutination reactions with absorbed sera or by precipitin reactions carried out with type specific antisera and suitably prepared bacterial extracts. The type specific antigens concerned are protein in nature and soluble in dilute acids. Of the 27 types described by Griffith (1935) it would appear that four (Types 7, 20, 21 and 16) do not possess the group specific carbohydrate and should therefore be excluded from this group. Types 7, 20 and 21 belong, antigenically to Group C. Type 16 to Group G (Hare 1936).

Group A streptococci in addition to producing  $\beta$  hæmolysis in blood agar plates appear without exception to form a soluble hæmolysin.

In regard to their other biological and biochemical characters, Group A streptococci produce a final pH of 5.0-5.6 in dextrose broth; they do not hydrolyse sodium hippurate; they do not reduce methylene blue in milk; they ferment trehalose but not sorbitol; they may or may not grow in 10 per cent bile agar and seldom grow in 40 per cent bile agar. These are the main differential group characters. In addition Group A strains almost always ferment salicin and lactose; seldom mannitol and very seldom raffinose or inulin. With the exception of Type 3 strains (Fuller and Maxted 1939; Hadley *et al.* 1941) peroxide is generally produced. A biochemical classification based on the fermentation of starch, mannitol, and cellobiose which is in concordance with serological typing has been proposed by Keogh and Simmons (1940).

The strains that have been adequately identified as belonging to this group have in the main been derived from infections in man—tonsillitis, scarlet fever, cellulitis, erysipelas, puerperal fever, other types of septæmia, acute broncho-pneumonia, otitis media and so on. They have also been isolated from the throat, nasopharynx or nose in normal persons. They have occasionally been isolated from cases of mastitis in cattle but in such instances there have usually been grounds for suspecting a human source of infection.

It is clear that streptococci falling into Group A possess all the characters that have been attributed to the classical *Str. pyogenes* and the group itself is so well differentiated that it clearly requires a distinctive label. Should it be given specific rank? It is certain that we cannot include in a single species all the antigenic groups of hæmolytic streptococci (A-N). The only question is into how many different species they should be divided. The balance of the evidence at present available appears to us to be in favour of recognizing the Group A strains as constituting a species in the generally accepted bacteriological sense. Its relation to the strains falling into other antigenic groups will be discussed in succeeding sections. There remains the question of the correct specific name. The name *Str. hæmolyticus* has obtained wide currency. It was used by the Committee of the Society of American Bacteriologists to denote the type species (Winslow *et al.* 1920) and we adopted it in the first edition of this book.

But the knowledge that has accumulated during recent years seems to us to render the name inconvenient and misleading. If we adhered to it we should no longer mean by *Str. haemolyticus* a haemolytic streptococcus or even a streptococcus giving  $\beta$  haemolysis but only a particular group of streptococci falling into that category. It therefore seems to us wiser to revert to the name *Str. pyogenes* defining that name as equivalent to a streptococcus of Group A with the proviso that this definition may well have to be modified in the future.

Before leaving this group and turning to others we may note briefly the existence of certain strains that have been isolated from milk and from the throats of infected persons during milk borne epidemics of tonsillitis (Davis and Rosenow 1910, Davis 1910, 1909, Stokes and Hachtel 1910, Capps and Davis 1914). These streptococci differ from the classical *Str. pyogenes* only in giving mucoid or semi-mucoid colonies on blood agar plates and in showing well-defined capsules. Because of this difference Davis allotted to them a separate specific name *Str. epidemicus*. It seems very doubtful however whether this procedure is justified. Although *Str. pyogenes* is not a capsulated species in the same sense as *Str. pneumoniae* it not infrequently forms recognizable capsules when growing in the tissues, or during the first few hours of its growth in serum broth. Seelemann and Hadenfeldt (1933) compared a large number of strains bearing the label *Str. epidemicus* with typical strains of *Str. pyogenes* and were unable to distinguish between them. On the basis of the available evidence we think that the name *Str. epidemicus* should be provisionally discarded and that the strains bearing that label should be included in the species *Str. pyogenes*. It may be noted that there is some tendency for other organisms to produce capsules when growing in milk.

Similarly there seems to us no adequate reason for allotting different specific names to strains that while possessing all the essential characters of *Str. pyogenes* differ from one another in regard to certain fermentation reactions. We should not therefore recognize a *Str. infrequens* fermenting mannitol as well as salicin or a *Str. anginosus* fermenting neither of these sugars. It is indeed by no means certain that all the strains to which these names have been attached are true haemolytic streptococci of the *pyogenes* type.

### Group B

The streptococci of this group share in common a group specific antigen that is carbohydrate in nature and differs from the group-specific antigen of Group A, or of any of the groups subsequently described. Group B streptococci are further divisible by precipitation or agglutination on tests into four main antigenic types, and by absorption of agglutinin tests into a number of sub types. The type-specific antigens within this group are apparently not acid soluble proteins but complex carbohydrates of a different chemical structure from the carbohydrate that determines group-specificity.

Streptococci falling within this group may or may not produce  $\beta$  haemolysis in blood agar plates. It would appear (see Stableforth 1939) that something over half of these strains are  $\beta$  haemolytic but the exact proportion is still a subject of controversy. Brown (1937, 1939) has shown that the haemolytic strains are characterized by the formation of a double zone of haemolysis which is best seen around deep colonies in rabbit blood agar incubated for 48 hours at 37 °C and refrigerated overnight. Although many haemolytic Group B strains yield a soluble haemolysin it would seem that this character is much less constant than with Group A strains and the titres of the filtrates obtained are usually much lower.

In regard to their other biological and biochemical characters Group B strains differ from Group A strains in that they produce a lower final pH in glucose broth (4.2-4.8), hydrolyse sodium hippurate and usually grow both on 10 per cent and 40 per cent bile agar. They resemble Group A strains in failing to reduce methylene blue in milk and in fermenting trehalose but not sorbitol. Sucrose and glycerol are usually fermented (Summons and Keogh 1940, Gunsalus and Sherman 1943) but mannitol, raffinose, sorbitol,

and inulin are not, the reactions in lactose and salicin are variable (Brown 1939). It may be noted that a considerable proportion of Group B strains form pigmented cells, usually yellow or red in colour (Orla-Jensen 1919; Plummer 1941).

The great majority of Group B strains that have been adequately identified have been isolated from cases of mastitis in cattle in many cases under conditions which have made it almost certain that they were the primary cause of the disease. They have occasionally been isolated from the normal human throat and vagina. They are very rarely pathogenic for man.

Group B, like Group A, clearly demands a label, and we feel that it may provisionally be accorded specific rank. We follow Klimmer and Haupt, and Minett and his colleagues in giving it the name *Str. agalactiæ* (Kitt 1893) (see Klimmer and Haupt 1930, Minett 1935b) in preference to the name *Str. mastitidis contagiosa* assigned by Nocard and Mollereau (1887) to streptococci isolated from a similar source. There can be no doubt that in defining this species antigenic structure should take precedence over hæmolytic production as a systematic criterion, and that the specific name should be applied to the non hæmolytic as well as to the hæmolytic strains. We should therefore, describe hæmolytic production as being an almost constant character within the species *Str. pyogenes* but a variable character within the species *Str. agalactiæ* whether the non hæmolytic strains of *Str. agalactiæ* should be classed as a distinct variety the future must decide.

#### Group C

The strains that fall within this group share a common group-specific polysaccharide antigen. The existence of a number of antigenic types can be demonstrated by agglutination. Of Griffith's original series Types 7, 20 and 21 were later found to belong to this group. Five further types among human strains have been differentiated by Simmons and Keogh (1940) and five among equine strains by Bazeley and Battle (1940). The type specific antigen appears to be of protein nature. On blood agar the colonies tend to be surrounded by a rather larger zone of hæmolytic than colonies of Group A, the outer edge of the hæmolytic zone being hazy. Colonies of the typical *Str. equi* type are large, honey-coloured and of very viscous consistency; if closely adjacent they readily flow together forming a streak of sticky material. Most but not all strains form a filtrable hæmolytic.

In regard to their other biological and biochemical characters Group C strains display certain significant differences among themselves. They produce in glucose broth a final pH intermediate between that produced by Group A strains on the one hand and by Group B strains on the other: the range covered by the group as a whole appears to vary between pH 4.5 and pH 5.4. No Group C strains hydrolyse sodium hippurate. Many of them grow on 10 per cent bile agar but few on 40 per cent bile agar. On the basis of their action on trehalose and sorbitol Group C streptococci can be differentiated into three sub-groups (Edwards 1934). One of these sub-groups ferments neither trehalose nor sorbitol, another ferments sorbitol but not trehalose, a third ferments trehalose but not sorbitol.

There is a correlation between fermentation reactions and habitat that gives to these sub-groups an importance that they would not otherwise possess. Most of the strains belonging to the trehalose negative sorbitol negative sub-group have been isolated from horses, particularly from cases of strangles, though they have also been obtained from infections in other animals. They are further differentiated from the groups of streptococci that have been described above by their failure to ferment lactose. They share with these other groups the ability to ferment salicin and the inability to reduce methylene blue in milk. This sub-group clearly corresponds in its biochemical characters to the

*Sir equi* of many authors. According to Bazeley and Battle (1940) all strains of this sub group fall into one serological type—Type 1. The second sub group ferments sorbitol but not trehalose, and fails to reduce methylene blue in milk. Serologically, it comprises two types. Type 2 is the more numerous, and is distinguished by fermenting lactose. It has been isolated from respiratory catarrh of horses, and from a variety of lesions in horses, cattle, guinea-pigs, rabbits and so on. Type 3 does not ferment lactose, and was found by Bazeley and Battle only in equine respiratory catarrh.

- There is as yet no evidence that streptococci belonging to this, or to the preceding, sub group are pathogenic for man.

The third sub group is characterized by the fermentation of trehalose but not of sorbitol. The strains belonging to this sub-group have been derived both from animal and human sources, and it seems clear that some strains at least are pathogenic for man. Three of Griffith's 27 types of human pathogenic, hæmolytic streptococci belong to this C sub-group, not to Group A. All strains of this sub group are recorded as fermenting salicin, but the action on lactose appears to vary. The results recorded by Edwards (1934) and by Hare (1935) suggest that many of the animal strains fail to ferment lactose, while almost all the human strains act on this sugar. Among equine strains Bazeley and Battle (1940) found that the great majority did not ferment lactose, these fell into their serological Type 4. A few, which did ferment lactose, fell into Type 5. Simmons and Keogh (1940) who studied 169 strains of human origin falling into the trehalose positive sorbitol negative sub group were able to divide them into seven further sub groups on the basis of lactose, asculin, amygdalin and raffinose fermentation; these bore some relation to the eight serological types—Types 7, 20 and 21 and five new types—which they were able to distinguish by agglutination.

The labelling of Group C hæmolytic streptococci presents a problem of some difficulty. There is no specific name that can be applied to the group as a whole. One sub-group of strains, however, does seem to merit special attention, namely the group of trehalose, sorbitol and lactose-negative strains, responsible for strangles in horses, which produce characteristic colonies, and fall into Bazeley and Battle's serological Type 1. If Bazeley and Battle's findings are confirmed, then it may be justifiable to apply the specific designation *Sir equi* to this sub group. The term *Sir dysgalactæ* is in common use among veterinary bacteriologists to denote a non hæmolytic streptococcus responsible for some cases of acute or subacute mastitis in cattle (see Minett 1936, Stableforth 1942). According to Stableforth (1945) this organism contains the Lancefield Group C carbohydrate, and should therefore be included with the other strains that we have just described. Its exact relationship, however, to these strains is still under study and the specific name that it has been awarded should be regarded as provisional only.

### Group D

Great confusion has existed in the past between (a) the hæmolytic streptococci falling into Group D, (b) the enterococci—usually non hæmolytic—isolated from human faeces, and (c) the lactic streptococci—also non hæmolytic—so frequently present in milk.

*Group D hæmolytic streptococci*—Lancefield (1933–1941) has recognized a group of hæmolytic streptococci that are characterized by the possession of a specific polysaccharide antigen. These organisms are generally known as Group D hæmolytic streptococci. They have been isolated mainly from cheese and from human faeces. Morphologically, they tend to assume the diplococcal rather than the streptococcal formation. On horse blood agar they give rise to  $\beta$  hæmolytic colonies. Some, at least, of the strains are capable of producing a filtrable hæmolysin under special conditions (Todd 1934, Plummer 1941).



They give a low final pH in glucose broth (4.0-4.8). They grow freely on 10 per cent and 40 per cent bile agar. They reduce methylene blue in milk. They are heat resistant, withstanding a temperature of 60°C for 30 minutes. They fail to hydrolyse sodium hippurate. They ferment lactose, salicin, nearly always mannitol, and usually but not always trehalose and sorbitol.

*Enterococci*.—Many bacteriologists, especially of the French school, have long recognized the occurrence in the human intestine of a characteristic streptococcus usually occurring in pairs of ovoid cocci, sometimes in short chains. The characteristic morphology of this organism was described by Thiercelin in 1899, who called it the *Enterococcus*, and under that name it has made frequent appearances in later literature.

The interrelation of the streptococci of the human intestine has been studied by Dible (1921), whose paper on this subject affords an admirable example of the methods which should be employed in differentiating bacterial groups. He tested 134 strains of streptococci from human faeces as regards their behaviour in a large number of biological tests including heat resistance (see Houston and McCloy 1916) and chain formation as well as various biochemical reactions, and measured the association between different pairs of reactions by calculating a statistical coefficient of association. Using one such coefficient which gives the value of +1 where the association between two characters is absolute, 0 where there is no association, and -1 where the characters are mutually exclusive, he obtained the following values for the association between a particular series of characters among his 134 strains:

Heat resistance and mannitol fermentation	+ 0.93
" " " raffinose fermentation	- 0.85
" " " chain formation	- 0.96
Mannitol fermentation and chain formation	- 0.93

Heat resistance, as here designated, was tested by the ability of the various strains to survive heating at 60°C. for 30 minutes. Those organisms were classed as chain formers which showed any wide departure from the diplococcal form. Dible thus succeeded in demonstrating the existence in human faeces of a characteristic group of organisms which possessed a predominantly diplococcal morphology, were unusually resistant to heat, almost always fermented mannitol, and very seldom fermented raffinose. It may be added that the streptococci belonging to this group constantly fermented salicin, very seldom fermented inulin, and gave good growth on gelatin at 22°C. About 10 per cent of them liquefied gelatin. None of them caused hæmolysis in blood agar, or produced a green pigment. In a later communication (Weatherall and Dible 1929) it was noted that some strains of enterococci, having all the characters referred to above, produced areas of hæmolysis when grown on blood agar plates, but no filtrable hæmolyse could be obtained by the usual methods of cultivation. It seems clear that these hæmolytic strains correspond to the 'Group D' hæmolytic streptococci referred to above.

It has already been noted that streptococci from the human faeces grow freely in the presence of bile (Weissenbach 1918) and reduce methylene blue in milk. They have also been shown to produce a low final pH in glucose broth (Sherman and Stark 1931), to grow at both 10° and 45°C, and to develop in skimmed milk containing 1/1,000 methylene blue. Most strains belonging to this group produce no change on a blood agar medium, though a few have been described as forming  $\alpha$  hæmolytic or  $\beta$  hæmolytic colonies. According to Ehrismann (1943) the effect on blood is variable.

*Lactic streptococci*.—Günther and Thierfelder (1895) described the occurrence in milk of an organism which was responsible for spontaneous souring and clotting. They described the organism as a short bacillus, but Heinemann (1906), when investigating the bacterial flora of milk some ten years later, pointed out that a particular streptococcus, which was almost constantly present in fresh milk, was probably identical with the organism of Günther and Thierfelder. Baehr (1910) confirmed the frequent presence of this streptococcus and noted that it produced a large amount of acid, and rapid clotting. Ruediger

Both tests should be conducted in glucose lemco broth. In the 45° C test the temperature of the water bath must be controlled to  $\pm 0.1^\circ \text{C}$ , in the pH 9.6 test the reaction of the medium must be adjusted immediately before use the tubes should be incubated in an aneroid jar containing soda lime and the pH of uninoculated control tubes should be checked by the glass electrode immediately before and after incubation. Provided that the tests are carried out under as standard conditions as possible and that complete uniformity in every respect is not insisted on it is often possible to allocate individual strains to one or other group (Table 38).

TABLE 38  
DIFFERENTIATION OF *Str. faecalis* AND *Str. lactis*

	Haemolysis on horse-blood agar	Mannitol	Sorbitol	Sucrose	Growth				Serological group	Source
					Resistant at 60° C for 30 min	at 45° C	at pH 9.6	in 6.5 NaCl		
<i>Str. faecalis</i>	Variatile	+	+	+	+	+	+	+	D	Mainly faeces
<i>Str. lactis</i>	Usually none	±	—	—	—	—	—	—	V	Milk and cheese

Since the lactic streptococci belong to a different serological group (Group N) they need not be considered here. Group D hæmolytic streptococci and enterococci require a little further discussion. Numerous species have been recognized and named among these organisms.

*Str. zymogenes* for example of MacCallum and Hastings (1899) which is hæmolytic ferments mannitol liquefies gelatin and digests casein appears to correspond to the former group and *Str. faecalis* of Anfrues and Horder (1906) to the latter group. *Str. liquefaciens* of Orla-Jensen (1919) resembles *Str. faecalis*, but liquefies gelatin and digests casein. *Str. durans* of Sherman and Wing (1937) is  $\beta$  hæmolytic grows at 50° C but does not usually ferment either mannitol or sorbitol. *Str. glycerinnaceus* differs from *Str. faecalis* in fermenting glycerol. *Str. bovis* and *Str. anulaceus* (see Orla-Jensen 1919 Ayers and Mudge 1923) resemble each other in fermenting raffinose but not mannitol their relation however, to the enterococci is still in doubt.

In considering the nomenclature of this group we must determine whether any single species can be defined rigidly enough to enable it to be recognized and distinguished from other species by tests that yield consistent results. Unless we can do this, the use of a specific name tends to be misleading rather than helpful. We think that it is now possible to define broadly *Str. faecalis* in this way. Ignoring the absence of hæmolysis, we see that the enterococci differ in no important respect from Group D hæmolytic streptococci. We should therefore define *Str. faecalis* as a coccus arranged in pairs or very short chains growing in the presence of bile salt, usually resisting heat at 60° C. for 30 minutes, almost always fermenting lactose, mannitol, salicin and usually sucrose trehalose and sorbitol but not raffinose or inulin reducing the dye and forming a solid clot in litmus milk and possessing the specific Group D polysaccharide subsidiary characters being the failure of most strains to hydrolyse sodium hippurate the production of a low pH (4.0-4.8) in glucose broth and the ability to grow at temperatures between 10° and 15° C, in skimmed milk containing 1:1000 methylene blue, in lactose

(1912) noted the absence of hæmolysis on blood agar plates. Sherman and Albus (1918) in a careful comparative study of strains of this organism and of other strains referred to as *Str. pyogenes* noted the following points of difference. The lactic-acid streptococcus grew predominantly as diplococci or short chains, it clotted milk within 24 hours, it produced high acidity in milk (0.75 per cent. or more measured as lactic acid), it grew well at 10° C., but very poorly at 43° C. and it rapidly reduced methylene blue, litmus, indigo carmine and neutral red. In each of these characters it differed sharply from *Str. pyogenes*. Ayers, Johnson and Mudge (1924) report that the lactic-acid streptococcus produces high acidity, rapidly clots and decolorizes litmus milk, and reduces methylene blue, or Jann's green. This particular streptococcus has for long enjoyed specific rank, under the title *Str. lactis*, but it will be noted that many of its most striking characteristics are shared by enterococci. Ayers and Johnson (1924) carried out a careful comparative study of these two types, testing them as regards their reaction on blood agar, their morphology, their ability to withstand heating at 60° C., their reaction in litmus milk and Jann's green medium, their fermentation reactions, and the final pH attained. They were unable to detect any difference in behaviour except that enterococci appeared to form acid somewhat less vigorously than *Str. lactis*. Klockner (1935) also concludes that it is not possible to differentiate with certainty between lactic-acid streptococci and enterococci, though there are minor points of difference. Many strains of lactic acid streptococci, for instance, fail to ferment mannitol.

The origin of these streptococci in milk is an unsolved problem. It seems clear that they are not normal inhabitants of the cow's udder, but find their way into the milk from some outside source (Sherman and Albus 1918; Ayers, Johnson and Mudge 1924). Stark and Sherman (1935) have recorded their common occurrence on certain plants.

The relation between these three groups of streptococci has become clarified as the result of recent work. In the first place it has been shown by Gaham and Bartley (1939), Sherman, Smiley and Niven (1940), Seelemann and Nottbohm (1940), Shattock and Mattick (1943) and Ehrsmann (1943) that the enterococci possess the specific carbohydrate antigen of Group D hæmolytic streptococci, but that the lactic streptococci do not. It would appear that the enterococci differ from Group D hæmolytic streptococci only in their failure as a rule to produce characteristic  $\beta$  hæmolysis in blood agar. In the second place, careful comparison between strains of enterococci and of lactic streptococci has revealed differences in their resistance to heat and in certain growth characters that appear to be of some classificatory value. Sherman and his co-workers have been most active in this field. In 1934, Sherman and Stark found that enterococci withstood exposure in sterile skimmed milk to a temperature of 65° C. for 30 minutes, grew vigorously at 45° C., and developed in a lactose agar medium having a pH of 9.6 or containing 6.5 per cent. sodium chloride, whereas lactic streptococci did none of these things.

Tests of this sort, in which the result depends on a number of different factors besides the main one under examination, are seldom satisfactory, and it is not surprising therefore that the findings of subsequent workers have varied considerably. Shattock and Mattick (1943) find that resistance tests are best carried out in broth at 60° C., that special precautions have to be taken to maintain a pH of 9.6 in lactose agar, and that growth of enterococci in the presence of 6.5 per cent. NaCl is often so poor as to render this test of little value. Ehrsmann (1943) finds the heat-resistance test unreliable, some true enterococci proving susceptible, and some faecal streptococci other than those belonging to Group D proving resistant. Hobbs (1939) working in our laboratory, noted a number of discrepancies in the behaviour of *faecalis* and *lactis* strains in the Sherman set of tests and Ehrsmann (1943) in Germany made similar observations on faecal streptococci. According to Shattock (1945) the only two reliable tests are growth at 45° C. and at pH 9.6.

Both tests should be conducted in glucose leuco broth. In the 45° C test the temperature of the water bath must be controlled to  $\pm 0.1^\circ \text{C}$ , in the pH 9.6 test the reaction of the medium must be adjusted immediately before use, the tubes should be incubated in an anaerobic jar containing soda lime, and the pH of uninoculated control tubes should be checked by the glass electrode immediately before and after incubation. Provided that the tests are carried out under as standard conditions as possible, and that complete uniformity in every respect is not insisted on it is often possible to allocate individual strains to one or other group (Table 38).

TABLE 38  
DIFFERENTIATION OF *Str. faecalis* AND *Str. lactis*

	Haemolysis on horse- blood agar	Man- nitol	Sor- bitol	Suc- rose	Resist- ance 60 C 30 min.	Growth			Sero- logical group	Source
						at 45° C	at pH 9.6	in 6.5% NaCl		
<i>Str. faecalis</i>	Variable	+	+	+	+	+	+	+	D	Mainly faeces
<i>Str. lactis</i>	Usually none	±	—	—	—	—	—	—	N	Milk and cheese

Since the lactic streptococci belong to a different serological group (Group N), they need not be considered here. Group D hæmolytic streptococci and enterococci require a little further discussion. Numerous species have been recognized and named among these organisms.

*Str. zymogenes*, for example, of MacCallum and Hastings (1899), which is hæmolytic, ferments mannitol, liquefies gelatin and digests casein, appears to correspond to the former group, and *Str. faecalis* of Andrewes and Horder (1906) to the latter group. *Str. liquefaciens* of Orla-Jensen (1919) resembles *Str. faecalis*, but liquefies gelatin and digests casein. *Str. durans* of Sherman and Wing (1937) is  $\beta$  hæmolytic, grows at 50° C but does not usually ferment either mannitol or sorbitol. *Str. glycerinatensis* differs from *Str. faecalis* in fermenting glycerol. *Str. bovis* and *Str. inulinaceus* (see Orla-Jensen 1919; Ayers and Mudge 1923) resemble each other in fermenting raffinose but not mannitol, their relation however, to the enterococci is still in doubt.

In considering the nomenclature of this group, we must determine whether any single species can be defined rigidly enough to enable it to be recognized and distinguished from other species by tests that yield consistent results. Unless we can do this, the use of a specific name tends to be misleading rather than helpful. We think that it is now possible to define broadly *Str. faecalis* in this way. Ignoring the absence of hæmolysis, we see that the enterococci differ in no important respect from Group D hæmolytic streptococci. We should therefore define *Str. faecalis* as a coccus arranged in pairs or very short chains, growing in the presence of bile salt, usually resisting heat at 60° C for 30 minutes, almost always fermenting lactose, mannitol, salicin, and usually sucrose, trehalose and sorbitol, but not raffinose or inulin, reducing the dye and forming a solid clot in litmus milk, and possessing the specific Group D polysaccharide, subsidiary characters being the failure of most strains to hydrolyse sodium hippurate, the production of a low pH (4.0–4.8) in glucose broth, and the ability to grow at temperatures between 10° and 45° C, in skimmed milk containing 1:1,000 methylene blue, in lactose

broth containing 1-15 000 potassium tellurite, and on lactose agar at a pH of 9.6 or containing 6.5 per cent. NaCl. It may be noted that *Str. faecalis* possesses the unusual property among streptococci of being insensitive to penicillin (Fleming 1939). It seems to us very doubtful whether any other members of this group can be assigned specific rank, and we should agree with Ellis (1937) and Mattick (1943) in regarding for the present the *ymogenes liquefaciens* and *durans* organisms as varieties of *Str. faecalis*. *Str. glycyminaceus* has no claim even to a varietal name. The position of *Str. bovis* must await further observations.

Within the enterococcus group an attempt has been made to distinguish antigenic types, but so far little progress has been made. Lancefield (1941) states that she has been able to define three types, differing apparently in the nature of their polysaccharide antigen and Grumbach and Schnetz (1935) claim to have distinguished seven types by agglutination. Whether these types occur within the species *Str. faecalis* as we have defined it, or correspond to any of its varieties, it is at present impossible to say.

#### Group E

The few strains belonging to this antigenic group were isolated from cows' milk by Lancefield (1933). In view of the small number of strains examined, it is too early to give any generalized description of the other group characters. Lancefield (1941) has recognized one antigenic type.

#### Group F

This group has been differentiated by Lancefield and Hare (1935) and by Hare (1935). It possesses a characteristic group-specific polysaccharide antigen, by means of which it may be identified. Ellis (1937) has established four antigenic types by the use of precipitation and agglutination tests.

Group F strains grow slowly on blood agar plates, forming minute non-point transparent colonies surrounded by a narrow zone (1.5-1.8 mm. in diameter) of  $\beta$ -haemolysis. They are identical with the strains described by Long and Ellis (1934) as "minute haemolytic streptococci". They do not, when tested by the usual methods, form a filtrable haemolysin acting on horse blood, but according to Plummer (1941) they do form a filtrable haemolysin for sheep blood.

Group F strains produce a final acidity in glucose broth of pH 4.8-5.2. They do not hydrolyse sodium hippurate. They do not reduce methylene blue in milk. They do not grow either on 10 per cent. or on 40 per cent. bile agar. Some but not all strains ferment trehalose, none ferments sorbitol. All that have been tested ferment lactose and salicin.

The strains that have been identified as belonging to this group have been derived mainly from the human throat. There is some evidence that they may be responsible for occasional cases of tonsillitis, and perhaps for other infections of the respiratory tract.

The provisional label for this group is clearly "Group F haemolytic streptococci", with a proviso that we are not insisting on the demonstration of the formation of a filtrable haemolysin before admitting a streptococcus to the haemolytic class.

#### Group G

The strains of this group share a common group-specific polysaccharide antigen (Lancefield and Hare 1935; Plummer 1935; Lancefield 1941). Hare (1935) notes that some sera produced by the injection of Group C strains tend to give cross-precipitation with extracts from Group G strains—a circumstance that appears to be due to their possession of a common protein antigen (Lancefield 1941). Organisms from matt colonies of Groups C and G strains resemble each other in fermenting trehalose but not sorbitol, and in producing fibrinolysin and streptolysin O. A number of antigenic types have been dis-

tinguished by Simmons and Keogh (1940) among both the large colony and the so called minute forms of streptococci in this group, and Bliss (1937) has recognized one type, which was identical with her Group I Type 1

As regards their other biological and biochemical properties there appears to be a conflict of evidence in regard to the final pH attained in glucose broth. Plummer (1935) gives this as pH 4.4-4.6, which would place these strains in the "high acid" group, but Lancefield and Hare (1935) and Hare (1935) give figures of pH 4.6-5.2. Group G strains do not hydrolyse sodium hippurate. The majority are able to multiply on 10 per cent bile agar, but few on 40 per cent bile agar. They do not reduce methylene blue in milk. They ferment trehalose, but not sorbitol. All strains tested have fermented lactose, but the fermentation of salicin appears to be less constant than with Groups A, B and C.

Most of the strains that have been identified as belonging to this group have been isolated from man, a few from the monkey or the dog. Several of the human strains have been derived from normal persons, but it seems clear that some, at least, are pathogenic for man, though there is a suggestion that they seldom cause very severe infections. One of Griffith's 27 types—Type 16—of human pathogenic hemolytic streptococci falls into this group.

The labelling of this group should clearly be provisional. It shows obvious relationships to Group C, and, apart from its antigenic structure, to Group A. There seems no reason, at the moment, to allot to it any specific name. "Group G hemolytic streptococci" will serve our immediate purpose.

#### Groups H and K.

These two additional antigenic groups have been differentiated by Hare (1935). They appear to differ from each other, and from the other groups of hemolytic streptococci that have been described, in regard to their group-specific antigens. Those strains that have been examined have shown other cultural or biochemical characters that may have differential significance, but so few strains belonging to these two groups have yet been studied that it would be premature to describe their characters in any detail. They have all been isolated from the nose or throat of normal persons, and there is as yet no evidence that they are pathogenic. One antigenic type has been recognized in Group H. (Lancefield 1941).

#### Groups L and M

These groups have been differentiated by Fry (1941), who has been kind enough to supply us with the following information. Most of the original strains studied were isolated by Dr. Tom Hare from animal sources.

The majority of Group L strains came from dogs and pigs, but two (Hooper and Krone) were isolated from the human throat by White, Rudd and Ward (1939) in Australia. The colonies are small, but have a wide zone of  $\beta$  hemolysis. A soluble hemolysin is formed. There is no growth on bile agar, and no hydrolysis of sodium hippurate. Trehalose is fermented and sometimes lactose, but not mannitol, salicin, or sorbitol.

Strains belonging to Group M came almost exclusively from the tonsil of the dog; no human strains have yet been identified. Growth is extremely poor and the organisms rapidly die out. On blood agar the colonies are very small, but are surrounded by a wide zone of  $\beta$  hemolysis. A soluble hemolysin is formed, but it is weak and acts slowly. Lactose is fermented, but not trehalose, mannitol, salicin or sorbitol. The formamide method of extraction cannot be used for this group, as heating much above 100° C. destroys the precipitinogen.

#### Group N

We have already pointed out, when discussing Group D streptococci (p. 582) that the lactic streptococci can be distinguished from the enterococci by a series of metabolic and heat resistance tests, and by their failure to form the group specific polysaccharide

antigen common to the hemolytic and non hemolytic members of Group D. The observations of Sherman, Smiley and Niven (1940), of Seelemann and Nottbohm (1940) and of Shattock and Mattick (1943) have shown that the lactic streptococci form a group specific antigen of their own, and it therefore seems appropriate, in spite of the absence of hemolysis caused by these organisms, to include them in the Land field series and assign them to Group A. There seems no reason why the chief representative of this group, the properties of which have already been described, should not be awarded specific rank and referred to as *Str. lactis*. Whether the closely allied organism, called by Orla-Jensen (1919) *Str. cremoris* is to be treated similarly is less clear. According to Yawger and Sherman (1937) it differs from *Str. lactis* in the slightly larger size of its cells, its greater tendency to form chains, its lower optimal temperature, its greater susceptibility to methyl blue, its failure to form ammonia in a 4 per cent peptone medium and certain other minor respects. These differences seem to be far more suggestive of environmental variation than of fixed hereditary characters and it would be wise for the present to regard *Str. cremoris* as no more than a variant of *Str. lactis*.

**The Classification of the  $\alpha$ -hemolytic Streptococci.**—It has been noted in preceding sections that streptococci giving  $\alpha$  hemolysis with a characteristic green coloration on blood agar plates, failing to produce a soluble hemolysin, usually fermenting raffinose but not mannitol, and possessing certain other characters in common can constantly be isolated from the human mouth and throat and from the faeces of cattle. The problem that confronts us is whether these streptococci form a group or a species and, if a group, whether the species of which that group is formed are sufficiently well differentiated to be allotted specific names.

Ayers and Mudge (1923) express the view that the  $\alpha$  hemolytic streptococci of the bovine intestine differ in certain minor characters from the  $\alpha$  hemolytic streptococci of the human mouth and throat, and reference to several of the papers quoted above will reveal a tendency to accord the bovine strains specific rank under the name *Str. bovis*. It is however by no means clear that this procedure is justified or on what differential characters the proposed nomenclature is to be based. Indeed there is some evidence to suggest that *Str. bovis* may be related to the Group D hemolytic streptococci (see Shattock and Mattick 1943).

There is another streptococcus falling into this group that seems to merit separate consideration. Freudenreich (1897) isolated a streptococcus from Kefir, a form of fermented milk. This streptococcus has the usual characters of the viridans type (Sherman 1921, Ayers *et al.* 1921, Ayers and Rupp 1922). When tested in the ordinary way, with a Durham fermentation tube, this organism, like other streptococci, produces acid but no gas from various substrates. If however, it is tested in the Eldridge fermentation tube in which it is grown in a shallow layer of fluid medium, freely exposed to the air, and the  $\text{CO}_2$  evolved is taken up by a standard solution of lithium hydroxide, exposed in a connected tube of the same kind, as large an amount of  $\text{CO}_2$  is evolved from lactose as is given off when that sugar is fermented by *Bact. coli*. It would seem that the almost anaerobic conditions existing in the closed Durham fermentation tube inhibit the production of  $\text{CO}_2$  by the Kefir streptococcus. It is difficult to assess the real significance of this observation since we have no knowledge of the way in which most species of non gas producing bacteria would behave if tested in the Eldridge tube, instead of in the closed Durham or Smith tubes in which they have in fact been tested, but no  $\text{CO}_2$  was formed in the Eldridge tube by such other strains of streptococci as were tested by Ayers and his colleagues including strains isolated from the bovine faeces, and the lactic acid streptococcus.

There can we think, be little doubt that the  $\alpha$  hemolytic streptococci of the viridans type will ultimately be separated into a number of distinct species or types,

but we doubt very greatly whether it is yet possible to define with any exactitude either an inclusive species *Str viridans*, or any of the fermentative types that have in the past been given specific names. We may, however, mention some of the properties attributed to the main types.

*Str salivarius*—This organism occurs in the human mouth and intestine. It is said to form short chains, to grow at 45° C but not at 10° C, to produce very little greening on blood agar, to produce a low final acidity (pH 4.0–4.4) in glucose broth, to clot milk to form a soluble levan from sucrose and raffinose, to give rise to large mucoid colonies on agar containing 5 per cent of either of these sugars, to ferment lactose, raffinose and salicin, and to hydrolyse aesculin but not sodium hippurate (Niven *et al* 1941; Sherman *et al* 1943). Most workers record it as being without action on inulin but Sherman, Niven and Smiley (1943) include this sugar among those that are fermented.

*Str mitis*—This organism also occurs in the human mouth. According to Sherman, Niven and Smiley (1943) it comprises a much less homogeneous group of strains than *Str salivarius*. It does not usually grow at 45° C; it forms good  $\alpha$  hæmolytic colonies on blood agar, it does not produce so low an acidity in glucose broth as *Str salivarius*; it often fails to clot milk, it synthesizes no polysaccharide from sucrose or raffinose; it does not form mucoid colonies on 5 per cent sucrose agar, it usually ferments lactose and salicin but not as a rule raffinose or inulin, and it fails to hydrolyse sodium hippurate or, with some exceptions, aesculin.

*Str equinus*—This organism is found in the intestine of the horse. It is said to grow at 45° C, to produce good greening on blood agar, to produce no polysaccharide from sucrose or raffinose, to hydrolyse aesculin but not hippurate, to grow on blood agar containing 30 per cent bile, and to ferment salicin but not lactose. It is usually reported as fermenting raffinose, but Sherman, Niven and Smiley (1943) disagree with this statement.

*Str bovis*—This organism has already been mentioned under Group D hæmolytic streptococci and on p. 588.

*Str acidominimus*—This organism was described by Ajers and Hudge (1922) who isolated it from cows' milk and faeces. It is also found in the vagina of the cow (Smith and Sherman 1939). It forms  $\alpha$  hæmolytic colonies on blood agar, it fails to grow at 10° C or 45° C, it has very weak fermentative properties producing a final pH in glucose broth of about 6.2, it has some hydrolysing effect on hippurate but not usually on aesculin; it ferments lactose and sucrose, and sometimes mannitol but not as a rule salicin or raffinose, it has little or no action on litmus milk and in milk containing 1–10,000 m.ethylene blue it fails to grow.

*Str thermophilus*—This organism produces completely non hæmolytic colonies on blood agar, but may conveniently be mentioned here. It was described by Orla-Jensen (1919) as one of the organisms that grows actively in milk at a temperature of 50° C. It does not grow at 10° C, it is not destroyed by heating to 63° C for 30 minutes; it forms long chains in milk, its colonies are of the pin-point type; it ferments lactose and sucrose, but not salicin, and it clots milk but has only slight reducing action on the litmus. Whether disaccharides are fermented by this organism without preliminary hydrolysis to mono-saccharides has been discussed by Wright (1936) and Sherman and Stark (1938).

Omitting *Str thermophilus*, which is non hæmolytic, we feel that it is wiser at present to use the non-committal group term '*viridans streptococci*' for these organisms rather than the specific name *Str viridans*. The strongest claim to specific rank appears to be possessed by *Str salivarius*, and if the findings of Sherman and his colleagues are confirmed, namely that all strains of this species form a soluble polysaccharide from sucrose, it may be well to admit this claim. The other named organisms, however, are the object of so many discrepant reports that any attempt to accord them specific rank would in our opinion be premature.



### Pathogenicity and Toxin Production.

The various species of streptococci described above include some of the most important pathogens of man, and are responsible for many infections of economic importance among animals.

*Str. pyogenes* gives rise to numerous pyogenic and septicæmic infections in man (see Chapter 67), as well as being the cause of scarlet fever (see Chapter 66).

It is pathogenic for a number of laboratory animals, including the rabbit, the mouse and the guinea pig, but the virulence of different strains for these animals is by no means uniform, and the guinea pig is often relatively resistant. To obtain a highly virulent strain for the mouse, or for the rabbit, it is often necessary to test a large number of strains of human origin, or to adapt a strain to the new host by repeated passage. When a highly virulent strain has been obtained, its intravenous injection leads to a fatal septicæmia, frequently associated in the rabbit with suppurative lesions in the joints, and sometimes with an ulcerative endocarditis. Intraperitoneal injection is followed by a suppurative peritonitis leading to a septicæmic infection, while subcutaneous injection is followed by a localized abscess, with or without a subsequent generalization. With some strains, so far at least as the rabbit is concerned, intradermal injection is followed by a spreading erysipelatous infection of the skin, or by a more severe dermal infection leading to necrosis, and an infection of the latter type not infrequently terminates as a septicæmia.

*Str. pyogenes* is of particular interest to the pathologist in that it combines the capacity for tissue invasion with the production of filtrable exotoxins.

One of these toxins, streptococcal hæmolyxin, has already been considered. That this substance is toxic in the animal body, as well as being lytic for red blood corpuscles *in vitro*, there can be no doubt. Its minimal lethal dose is large (5–10 ml for the rabbit), but when administered intravenously in this amount it kills the animal within 24–36 hours with an associated hæmoglobinuria, and with evidence of intravascular hæmolyxin at necropsy (McLeod and McNeé 1913, Channon and McLeod 1929). Weld (1934, 1935) more recently described preparations of streptolysin, obtained by extracting the cocci with serum that are fatal for mice in a dose of 0.1 ml. Streptococcal hæmolyxin is, as we have seen, thermolabile, being inactivated at 58° C in 30 mins. As already noted, Todd (1934) has demonstrated the presence in lytic filtrates of two hæmolyxins, one oxygen sensitive but reactivable by reduction and another which is oxygen stable but is very sensitive to heat and to acid. Both are toxic, the S type causing death by intravascular hæmolyxin, the O type probably causing death by some other means. Specific antibodies, having protective properties and showing no cross-neutralization, can be prepared against each type of hæmolyxin (Todd 1935b).

In addition to their action on red blood corpuscles, filtrates of broth cultures of *Str. pyogenes* have a destructive action on polymorphonuclear leucocytes. This was first demonstrated by van der Velde (1894) in his studies on experimental infections with streptococci, and he named the active principle *leucocidin*. It was subsequently shown by Neisser and Wechsberg (1900, 1901) that this action could be demonstrated *in vitro*, since the streptococcal filtrates, in killing the leucocytes, deprive them of their power of reducing methylene blue. It was early noted that streptococcal leucocidin is relatively thermolabile, and most authors (see Nakayama 1920, Channon and McLeod 1929) have concluded that its thermolability is of the same order as that of streptococcal hæmolyxin. Channon and McLeod conclude, from this and other observations, that streptococcal hæmolyxin and leucocidin are identical, but Nakayama regards them as different, a view that is put forward more strongly by Evans (1931) and by Gay and Oram (1933), who dispute the previous findings in regard to the thermolability of streptococcal leucocidin, and state that it withstands heating at 70° C for 30 minutes. Todd (1942) has brought evidence to suggest that the leucocidin is identical with the oxygen labile streptolysin O, and that it plays no part in the determination of virulence to mice. This problem must be regarded as awaiting final solution.

with Group C streptococci, but not with Group A streptococci. There is some evidence that the virulence of Group A strains is more closely related to the type-specific M protein than to the possession of a capsule (see Lancefield 1941). The importance of hyaluronidase is likewise doubtful. Neither Crowley (1944) who studied 376 strains of hemolytic streptococci from human sources, nor Humphrey (1944) who studied 81 strains of pneumococci from consecutive cases of pneumonia, could find any relation between the amount of hyaluronidase produced and the apparent virulence of the strain.

In summary *Str. pyogenes* produces the following toxins or aggressive substances (see Chapter 4) which, in one way or another, determine or are associated with its pathogenic activity: (a) hemolysin, (b) leucocidin, (c) erythrogenic toxin, (d) fibrinolysin, (e) hyaluronic acid, (f) hyaluronidase, (g) type-specific M protein.

*Str. agalactiae*.—Our knowledge of the pathogenicity and toxigenicity of this species is, as yet, very incomplete. It produces mastitis in cattle, but, so far as our present knowledge goes, is not pathogenic for man. Its pathogenicity for the mouse or the rabbit, is low (Minett and Stableforth 1931, Minett 1935). Some but not all strains produce a filtrable hemolysin, which Todd (1934) has found to be of the oxygen-stable, non-antigenic type. Whether *Str. agalactiae* produces a leucocidin is not known. It has been shown by Smith (1929) that strains of hemolytic streptococci isolated from cat le may produce an erythrogenic toxin that is neutralized by scarlatinal antitoxin, but there is no evidence that these strains were *Str. agalactiae*. No erythrogenic toxin was produced by the mastitis strains examined by Minett and Stableforth (1931). *Str. agalactiae* does not produce a fibrinolysin active against human fibrin (Lancefield and Hare 1935, Hare 1935), or against cattle fibrin. It produces hyaluronidase, but seldom forms capsules, when it does, the capsules do not consist of hyaluronic acid (McClellan 1941).

*Str. pneumoniae*.—The pneumococcus is an important pathogen of man, giving rise to pneumonia, particularly the lobar form, sinusitis, otitis media, less frequently meningitis, suppurative arthritis, or peritonitis, and occasionally other infections.

It is highly pathogenic for the mouse and rabbit, rather less so for the guinea pig. The cat, dog and chicken are relatively resistant. It is a characteristically invasive organism causing a fatal bacteremic infection when injected intravenously, an acute peritonitis followed by bacteremia when injected intraperitoneally, localized suppuration followed by generalization when injected subcutaneously and a spreading inflammatory lesion followed by generalization when injected intradermally in the rabbit. Different serological types of pneumococci may show differences in their virulence for different laboratory animals: for instance many strains of Type III pneumococci are relatively avirulent for the rabbit (Tillett 1927). Again, different strains belonging to the same type may show wide variations in virulence as judged by their minimal lethal dose. With a strain of maximal virulence the injection of  $1 \times 10^8$  pneumococci into the peritoneum of a mouse will cause an infection that leads to death within 1–48 hours.

In contrast to *Str. pyogenes* the pneumococcus is not an actively toxigenic organism, in the sense of producing filtrable toxins. Some observers have indeed regarded it as being devoid of this capacity and as affording a typical example of a bacterium the pathogenicity of which is determined entirely by its powers of invasion, associated, of course, with the effects of those presumptive "endotoxins" (see Chapter 4) that must always be concerned in invasive bacterial infections. There is little doubt that this contrast is a true one, in the sense that filtrable toxins play a much more prominent part in *Str. pyogenes* infections than in those due to pneumococci, but the difference is not so absolute as it has been supposed to be.

We have seen that *Str. pneumoniae* produces a filtrable hemolysin when grown under optimal conditions. This is of the oxygen-sensitive type, and is antigenic. There is evidence (Todd 1934) that it bears some antigenic relationship to the oxygen-sensitive hemolysin of *Str. pyogenes* but is certainly not identical with it. Oram (1934) has described the presence in pneumococcal filtrates of a leucocidin, active for rabbit leucocytes. Like the streptococcal leucocidin described by Gav and Oram (1933), this substance is

relatively thermostable. It is not inactivated in one hour at 70°C, but is completely inactivated at 85°C for a similar period.

Several observers (Julianelle and Reimann 1926, Reimann and Julianelle 1926, Maur 1928, Pittman and Falk 1930, Goodner 1931) have noted purpuric lesions following the injection of pneumococcal extracts and autolysates. The active agent would appear to be an 'endotoxin,' i.e. some constituent of the bacterial cell, rather than a soluble toxin. It may be noted that Avery and Goebel (1933) report that mice may develop purpura after injections of the acetylated form of the Type I pneumococcal polysaccharide, while this effect is not produced by the deacetylated form.

Pneumococci produce hyaluronidase in varying amount, but there appears to be no relation between their activity in this respect and their virulence (Humphrey 1944). Whether they form a fibrinolysin against human fibrin is not certainly known. The formation of the characteristic fibrin network in the alveoli in lobar pneumonia in man would seem to make it unlikely that the pneumococci concerned act vigorously on human fibrin, but it would appear that they are able to attack rabbit fibrin or fibrinogen. Goodner (1931, 1933) has recorded that the oedema fluid withdrawn from the spreading oedematous lesions caused by the injection of virulent pneumococci into the rabbit's skin not only fails to clot, but retards the coagulation of normal rabbit's blood and that a similar anticoagulant property is possessed by pneumococcal extracts and autolysates. The active substance is relatively thermostable, since it withstands heating to 70°C. for 15 minutes. Such autolysates when injected into the skin together with a slightly virulent strain of pneumococcus greatly extend the area of the lesion produced.

None of the other groups, species, or types of streptococci that we have described above have been studied, in regard to their pathogenicity for laboratory animals in the same detail as *Str. pyogenes* and *Str. pneumoniae*. A few brief notes will give most of the information that we possess.

**Streptococci of Group D.**—The organisms of this group are far less pathogenic than *Str. pyogenes* or *Str. pneumoniae*. They have however, occasionally been isolated from the blood stream in man usually in cases of subacute endocarditis (see Chapter 68) rarely in other conditions. They are normal inhabitants of the intestinal tract and are not infrequently present in suppurative abdominal lesions though their pathogenic role is often doubtful. They are also not uncommonly present in infections of the urinary tract. Their virulence for laboratory animals is usually low. Dible (1921) tested 83 faecal strains by injection into mice and 6 of these showed some degree of pathogenicity. It is very probable that such pathogenicity as this group possesses is confined to particular species, or strains, and that many strains such for example as most of those isolated from milk or cheese are altogether non pathogenic. It may, perhaps be noted that the hæmolytic strains belonging to this group do not produce a fibrinolysin active against human fibrin.

**Streptococci of the Viridans Group.**—The position of this group, in regard to pathogenicity, is very similar to that of Group D. Streptococci of the viridans type are of low virulence, both for man and for animals. In man they are frequently isolated from localized septic lesions in connection with the teeth and gums and they are the most frequent cause of subacute bacterial endocarditis. Here again, it is probable that different species or types within the group vary considerably in their pathogenicity for different animal species. Many of them are probably quite avirulent. In none of them is the virulence high.

Very brief notes must suffice for the remaining labelled groups of hæmolytic streptococci, since our knowledge of their pathogenic potentialities is as yet in its earliest infancy. The data available are contained in the recent papers by Lancefield, Hare, and others, to which frequent reference has already been made.

**Hæmolytic Streptococci of Group C** are certainly pathogenic. They have been frequently isolated from suppurative and acute inflammatory lesions in horses, cattle, guinea pigs and other animals. They have also been isolated from human infections, but their pathogenicity for man appears to be lower than that of *Str. pyogenes*. The human pathogenic strains that have been examined produce a fibrinolysin active against human fibrin; the strains derived from animals do not.

**Hæmolytic Streptococci of Group F**—The strains that have been identified as belonging to this group have been derived from minor infections of the respiratory tract in man, and from the normal human throat. Their possible pathogenic role must be regarded as *sub judice*, but their virulence would seem in any case, to be of a low order. They do not produce a fibrinolysin active on human fibrin.

**Hæmolytic Streptococci of Group G**—The streptococci of this group are certainly pathogenic. They have been isolated from tonsillitis, endocarditis and urinary infections in man (Macdonald 1939; Rantz 1942), from pneumonia in the monkey and from otitis in the dog. They have also been isolated from the normal human throat. Such evidence as is available suggests that Group G, like Group C strains, have a definitely lower virulence for man than has *Str. pyogenes*. Group G strains produce a fibrinolysin acting on human fibrin.

**Hæmolytic Streptococci of Groups E, H and K**—There is as yet no evidence that the streptococci belonging to these groups are pathogenic.

**Hæmolytic Streptococci of Groups L, M, and N**—Organisms belonging to Groups L and M appear to be pathogenic for certain animals, especially the dog. Group N strains are apparently non-pathogenic.

### Variation in the Characters of Streptococci.

It is not always possible from the records to identify the strain in which particular variations have been observed with one or other of the groups, species or types that have been defined in this chapter. It is, for instance, sometimes impossible to tell whether the term "hæmolytic streptococci" or *Str. hæmolyticus* is equivalent to *Str. pyogenes*, in the sense in which we have used that term. In almost all the instances given below, however, the identity of the strain or strains concerned is not in doubt, and the reservation that it is necessary to make is little more than formal.

**Variations in *Str. pyogenes***—There are many reports in the literature of the appearance of non-hæmolytic or  $\alpha$  hæmolytic variants in cultures derived from an originally hæmolytic strain. These reports have at times been regarded as invalidating hæmolysin production as a differential test, but in view of our more detailed knowledge of the factors that determine the action of streptococci on red blood corpuscles, it is clearly unnecessary to assume that the appearance in a culture of a  $\beta$  hæmolytic streptococcus of a variant that gives typical  $\alpha$  hæmolysis or no hæmolysis at all on the surface of an aerobic blood agar plate affords an instance of the mutation of *Str. pyogenes* into a streptococcus of the viridans type or into a completely non-hæmolytic form.

An illuminating example is given by Todd (1928b). By repeated mouse passage he was able to obtain from a typical  $\beta$  hæmolytic strain of streptococcus a variant that produced no hæmolysis at all on the surface of aerobic blood agar plates. When grown anaerobically this variant maintained full hæmolytic activity. Moreover the aerobically non-hæmolytic variant not only inactivated its own hæmolysin when exposed to a free supply of oxygen but under the same conditions inactivated the hæmolysin produced by the original hæmolytic strain, if the latter was grown in symbiosis with it. The appearance of non-hæmolytic variants in an originally hæmolytic strain has, it may be noted, been recorded by many other workers (see p. 567).

*Str. pyogenes* varies considerably in colonial appearance. Adopting the terminology of Dawson, Hobby and Olmstead (1938), we may recognize the following forms: (a) mucoid colonies, probably corresponding to the pseudo glossy colonies of Todd and Lancefield (1928). The organisms tend to be uniform in size, arranged in pairs or short chains, and to show capsulation when young. In broth, growth is diffuse or finely granular; (b) matt colonies, probably representing an intermediate stage between the mucoid and the smooth colony forms. The organisms are slightly pleomorphic, non capsulated, and are arranged in short chains or small clumps. The growth in broth is coarsely granular but not flocculent; (c) smooth colonies, corresponding to the glossy colonies of Todd (1928a). The organisms tend to be of uniform size and to be arranged in short chains. Growth in broth is diffuse or finely granular; (d) rough colonies, first described by Eagles (1923), flat and very irregular in outline. The organisms are large, pleomorphic, and arranged in long chains. Growth in broth is flocculent. The type specific acid-soluble protein M is usually present in the mucoid, matt, and smooth forms, but not in the rough form. Its loss is associated with an absence of virulence to mice (Todd 1928a, Todd and Lancefield 1928, Lancefield and Todd 1928), but its presence does not necessarily signify that the strain is virulent. The change from mucoid or matt to rough or smooth corresponds to the S  $\rightarrow$  R variation that occurs in many other bacterial species. According to Todd (1930), who studied the influence of oxygen pressure on this change, repeated subculture on solid media under aerobic conditions favours the appearance of smooth avirulent variants, but cultivation under anaerobic conditions prevents it. The effect of aerobiosis is apparently dependent on the formation of bacterial peroxide, the smooth variants being more resistant to this agent than the original matt virulent forms. In broth cultures the effect of increasing oxygen pressure by aeration is somewhat different; matt avirulent variants tend to appear under anaerobic conditions, whereas the virulence of the original matt strain is maintained when the culture is freely aerated even though smooth variants make their appearance. The change from matt to smooth is not associated with a loss of the power to produce hæmolyisin.

**Variations in other Groups of Hæmolytic Streptococci**—Variant colonies of more or less similar type to those found in *Str. pyogenes* have been described in other groups of streptococci. The reader will find an illustrated description of them in the article by Dawson, Hobby and Olmstead (1938). In addition, "minute" or dwarf colonies have been observed in Groups C and G by Long and Bliss (1934), Lancefield (1941) and Morton and Sommer (1944). Though, in general, the fermentation reactions of streptococci are relatively stable, differences have been noted in the same species associated with variations in colony forms. For instance, in Group C, the smooth and dwarf colony forms are said to ferment lactose and trehalose, but not sorbitol or mannitol. On the other hand, the mucoid colony forms ferment sorbitol and mannitol, but not lactose or trehalose (Morton and Sommer 1944).

**Variations in *Str. pneumoniae***—The more important types of variation that are encountered in the pneumococcus have already been noted in Chapter 9. The S  $\rightarrow$  R variation is here associated with the loss of the characteristic capsule, and with it the polysaccharide antigen that confers type specificity. Here, as elsewhere, the rough variants of pneumococci usually retain the characteristic bile solubility. They also retain the power of producing a hæmolyisin and a leucodæm (Oram 1934).

We have also referred in Chapter 9 to the important observations of Griffith (1928) on the conversion of a smooth strain of pneumococcus, belonging to a particular antigenic type, through the corresponding rough variant to a smooth strain belonging to a different antigenic type, and we have noted that these observations have been confirmed by several subsequent workers. Up to the present time this remains the only instance in which a transmutation of one normal bacterial type into another has been demonstrated under experimental conditions, and it affords no grounds for the assumption that such types are unstable under natural conditions.

*In regard to the other groups or species referred to in the present chapter we know too little as yet of the variations to which they are subject to attempt any systematic description of them. The description of variants that fit into no general scheme tends to confusion rather than to the clarification of knowledge. We must not of course ignore the fact that such variations occur, but to assess their true significance we must wait until we can allot them their proper place in the picture of bacterial structure which is being slowly but surely pieced together by modern methods of study.*

#### **The Anaerobic Streptococci.**

All the species or groups of streptococci described above are aerobic and facultatively anaerobic. Cocci growing in short or long chains have however been isolated which are either strictly anaerobic or grow only under micro-aerophilic conditions.

antigenic analysis, using antisera to eight of the strains showed considerable diversity of antigenic structure, but many group reactions. The two main types differentiated on colonial appearances were shown to be antigenically distinct. Finally, those strains that were tested for heat resistance were found to be killed by heating to 53-60°C for 30 minutes. Stone (1940), who studied 26 strains from parturient women divided them into three groups according to their growth in 10 or 40 per cent bile and was able to demonstrate the presence of at least two acid extractable antigens having some relation to those met with in the non-anaerobic types of streptococci.

It may be noted that the pathogenicity of anaerobic streptococci for laboratory animals appears to vary widely. Wegehans (1909) produced small abscesses in the peritoneal cavity of mice. Marwedel and Wehrsig (1915), with one strain examined, produced an acutely fatal infection in a guinea pig. Prévot (1925) records pathogenic lesions of a suppurative gangrenous or oedematous type sometimes fatal, with most of his strains. Harris and Brown (1929) found that three of 57 strains derived from cases of puerperal fever killed mice within 24 hours. Colebrook and Haro (1933) tested seven puerperal strains by subcutaneous injections into mice. Two of them gave rise to small caseous foci at the site of inoculation, but none of the mice died.

*Anaerobic pneumococci have also been described (Smith 1936)*

We append a summarized description of the more important species, or groups, to which names, or labels, can at the moment be attached.

#### SPECIES, GROUPS AND TYPES

##### *Str. pyogenes*

**MORPHOLOGY**—Cocci, usually spheroidal, about 0.5-0.75  $\mu$  in diameter arranged in chains of varying length, but usually including ten or more cocci. Capsules usually absent or poorly developed in tissues, frequently present in young serum broth cultures of freshly isolated strains. Non motile. No spores. Gram positive, not acid fast.

**GROWTH REQUIREMENTS**—When first isolated may grow poorly on ordinary nutrient agar. Growth is markedly improved by the addition of blood or serum. Optimal temperature 37°C. Grows at temperature slightly over 40°C. Poor growth below 20°C and usually fails to grow at 10°C. Aerobic and facultatively anaerobic.

**TYPE OF GROWTH**—On solid media after 24 hours incubation the colonies are small about 0.5-0.75 mm in diameter, opaque, slightly raised, circular with an entire margin, a slightly granular surface, and a granular structure, when viewed by transmitted light showing some differentiation into a more opaque central portion and a more translucent periphery. After further incubation (48 to 72 hours) the colony may extend in diameter and become differentiated into a raised central portion, smooth or contoured, and a flatter peripheral zone. The colonial forms presented by different strains are subject to considerable variation, and a plate from a single strain may show colonies of very varying appearance, particularly with regard to the smoothness, contouring or granularity of the surface and the degree of differentiation between the central and peripheral zones.

Streaked cultures on solid media give a relatively scanty growth with a tendency for a majority of the colonies to remain discrete. The growth emulsifies easily but usually gives a granular suspension.

**In blood agar plates**.—The colonies are surrounded by a zone of  $\beta$  haemolysis (see above) best seen in the deep colonies. A filtrable haemolysis is formed in fluid cultures which is oxygen labile. An oxygen stable haemolysis is also produced.

**In broth, or serum broth**—When first isolated, the growth may be finely granular or may form a powdery deposit at the bottom of the tube or cling to its sides. Turbidity of the medium may be moderate or slight. After subculture, and particularly when repeatedly subcultured at short intervals the turbidity may increase markedly and the granular deposit decrease, or disappear, but the turbidity almost always remains of a finely granular

In regard to the other groups or species, referred to in the present chapter, we know too little as yet of the variations to which they are subject to attempt any systematic description of them. The description of variants that fit into no general scheme tends to confusion rather than to the clarification of knowledge. We must not, of course, ignore the fact that such variations occur, but to assess their true significance we must wait until we can allot them their proper place in the picture of bacterial structure, which is being slowly but surely pieced together by modern methods of study.

### The Anaerobic Streptococci.

All the species, or groups, of streptococci described above are aerobic and facultatively anaerobic. Cocci growing in short or long chains have, however, been isolated, which are either strictly anaerobic, or grow only under micro-aerophilic conditions.

These anaerobic streptococci are of considerable importance to the medical bacteriologist, since some, at least, are certainly pathogenic for man, and the studies of recent years have shown that organisms of this type are a frequent source of severe puerperal infection (see Chapter 66). Apart from puerperal sepsis, and puerperal septicaemia, most of the strains of anaerobic streptococci that have been isolated have been derived from suppurative or gangrenous lesions, which have often been noted as producing a foul or foetid odour (see Veillon 1893, Kröning 1895, Sternberg 1900, Rust 1901, Lowkewicz 1901, Silberschmidt 1902, Warwedel and Wehrsig 1915, Küssling 1924, 1929, Prévot 1924, 1925, 1933). The importance of these organisms in relation to puerperal septicaemia was first insisted on by Schottmüller (1910, 1923), though their presence in the genital tract during the puerperium had been noted by several earlier workers. Schottmüller's observations have since been extended and confirmed by many subsequent observers (Bingold 1921, 1932, Lehmann 1926, Harns and Brown 1929, Colebrook 1930, Colebrook and Hare 1933). It would seem (Nativg 1905, Wegelius 1909, Rosowsky 1912, Soule and Brown 1932, White, E. 1933) that these anaerobic streptococci form part of the normal flora of the female genital tract, and it seems possible (White, E. 1933) that this is their principal normal habitat. Such attempts as have been made to isolate them from the normal human throat or intestine have been unsuccessful.

It is certain that the anaerobic streptococci comprise many different groups, species or types, but the data available are as yet far too scanty to permit of any systematic classification or nomenclature. Reference to the papers by Prévot, and by Colebrook and Hare will afford descriptions of several of the strains that have been isolated, and of certain differential criteria on which a future classification may in part be based.

It may be noted that many of these cocci are very small ( $0.3-0.4 \mu$ ), but the size tends to vary considerably in subculture (Colebrook and Hare 1933). Many but not all strains form abundant gas in fluid cultures, differing sharply in this way from the aerobic and facultatively anaerobic species that we have described above. Many, but not all strains produce an extremely foul odour.

Colebrook and Hare (1933) have studied the growth of 60 strains of anaerobic streptococci on blood agar and have thus been able to distinguish four different types on the basis of rate of growth, colony form, and changes produced in the medium. Only two of the 60 strains produced hæmolytic colonies, three others gave characteristic coal black colonies. The remaining 55 strains produced no change in the medium. About half the strains tested failed to ferment any test substrate, with the remainder it was not found possible to correlate the fermentation reactions with the colonial characters. A preliminary attempt at



antigenic analysis, using antisera to eight of the strains showed considerable diversity of antigenic structure, but many group reactions. The two main types differentiated on colonial appearances were shown to be antigenically distinct. Finally, those strains that were tested for heat resistance were found to be killed by heating to 58-60° C for 30 minutes. Stone (1940), who studied 26 strains from parturient women divided them into three groups according to their growth in 10 or 40 per cent bile, and was able to demonstrate the presence of at least two acid extractable antigens having some relation to those met with in the non anaerobic types of streptococci.

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Anaerobic pneumococci have also been described (Smith 1936).

We append a summarized description of the more important species or groups to which names, or labels, can at the moment be attached.

#### SPECIES GROUPS AND TYPES

##### *Str. pyogenes*

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Streaked cultures on solid media give a relatively scanty growth with a tendency for a majority of the colonies to remain discrete. The growth emulsifies easily but usually gives a granular suspension.

**In blood agar plates**—The colonies are surrounded by a zone of  $\beta$  haemolysis (see above) best seen in the deep colonies. A filtrable haemolysin is formed in fluid cultures which is oxygen labile. An oxygen stable haemolysin is also produced.

**In broth or serum broth**—When first isolated the growth may be finely granular or may form a powdery deposit at the bottom of the tube, or cling to its sides. Turbidity of the medium may be moderate or slight. After subculture and particularly when repeatedly subcultured at short intervals the turbidity may increase markedly, and the granular deposit decrease, or disappear, but the turbidity almost always remains of a finely granular

type, and the growth seldom or never becomes evenly diffuse. There is no pellicle formation.

*On gelatin* the growth is slight, and the colonies are minute and punctiform.

*In gelatin stab* there is a slight growth along the track, with minimal growth on the surface. The gelatin is not liquefied.

*On potato* growth is very slight, and often not detectable by the naked eye.

**HEAT RESISTANCE, AND VIABILITY**—*Str. pyogenes* is killed by heating to 55° C. for 30 minutes. It tends to die out in subculture unless preserved under particularly favourable conditions, but it remains viable for long periods in the dry state.

**BIOCHEMICAL REACTIONS**—*Str. pyogenes* does not hydrolyse sodium hippurate, does not reduce methylene blue in milk, produces a final pH of 5.0-5.6 in glucose broth, ferments trehalose, lactose, saccharose, salicin and occasionally mannitol, with the formation of acid but no gas, does not ferment sorbitol, inulin or raffinose. Produces acid in litmus milk, but no coherent clot. Does not liquefy gelatin. Does not reduce nitrates. Does not form indole. Is not soluble in bile. Its growth is inhibited by bile.

**ANTIGENIC STRUCTURE**—*Str. pyogenes* possesses the group-specific polysaccharide antigen of Lancefield's Group A hemolytic streptococci. It is differentiated into a large number of antigenic types by type-specific protein antigens, over 30 of these types have so far been identified by agglutination and absorption and by precipitation tests.

**PATHOGENICITY AND TOXIN PRODUCTION**—*Str. pyogenes* produces a variety of infections in man and more rarely in domestic animals. Some strains are highly pathogenic for the mouse or the rabbit, less so for the guinea pig. It produces a soluble hemolysin, a leucocidin, an erythrogenic toxin and a fibrinolysin acting on human fibrin.

### *Str. agalactiae*

**MORPHOLOGY, GROWTH REQUIREMENTS AND TYPE OF GROWTH**—In these characters *Str. agalactiae* does not differ significantly from *Str. pyogenes*, in many strains, however, the cells are yellow or red in colour.

**ACTION ON BLOOD**—Between one-third and one-half of the strains of *Str. agalactiae* that have been examined produce  $\beta$  hemolysis in blood agar plates. These strains produce a filtrable hemolysin of the oxygen-stable type.

**HEAT RESISTANCE**—*Str. agalactiae* is killed by heating to 60° C for 30 minutes.

**BIOCHEMICAL REACTIONS**—*Str. agalactiae* hydrolyses sodium hippurate, it does not reduce methylene blue in milk, it produces a final pH of 4.2-4.8 in glucose broth, it produces acid in trehalose, sucrose, glycerol, and usually in lactose and salicin, but not in sorbitol, mannitol, raffinose or inulin, it forms acid and clot in milk, it does not liquefy gelatin. It is not soluble in bile. Its growth is not inhibited by 10 per cent bile, and usually not by 40 per cent.

**ANTIGENIC STRUCTURE**—*Str. agalactiae* possesses the group-specific polysaccharide antigen of Lancefield's Group B streptococci. It is separable into different antigenic types, four of which have so far been identified by type-specific antigens which appear to be carbohydrate not protein in nature. Sub types of the four main types have been recognized.

**PATHOGENICITY AND TOXIN PRODUCTION**—*Str. agalactiae* is an important cause of mastitis in cattle. Its pathogenicity for laboratory animals is low. Some, if not all, strains produce a filtrable hemolysin. It does not produce a fibrinolysin acting on human fibrin. Whether it produces a leucocidin is unknown. There is no evidence that it produces an erythrogenic toxin.

### *Str. pneumoniae*

**MORPHOLOGY**—Ovoid or lanceolate cocci, arranged in pairs or short chains, when in pairs, the adjacent ends of the cocci are usually bluntly rounded, the opposite ends

more acutely pointed. Some strains (particularly Type III) tend to form longer chains. As seen in films from the tissues, the pneumococcus shows a well marked capsule, and this capsule is frequently retained in cultures on suitable media. Non motile. No spores. Gram positive. Not acid fast.

**GROWTH REQUIREMENTS**—Grows poorly on ordinary media, especially when first isolated; the addition of blood or serum to the medium greatly improves growth. Optimal temperature 37° C, range of growth more restricted than with other species of *Streptococcus*. Usually no growth on gelatin at 20° C. Aerobic and facultatively anaerobic. Growth of some strains improved by incubation in 10 per cent CO<sub>2</sub>.

**TYPE OF GROWTH**—On solid media. Small raised, circular colonies, 0.5–1 mm in diameter, with a smooth surface, an entire edge, and very little differentiation. On a favourable medium, such as blood agar, the colonies are often characteristic, the surface is flat and smooth, and the edges are sharply and steeply raised from the surface of the medium. In some cases the edge may be raised above the surface of the colony, forming a raised circumferential ring. Several adjacent colonies may become confluent forming a raised area of growth with a flat, even surface and a sharply delimited edge. With longer periods of growth (48–72 hours) the central portion of the colony often undergoes autolysis. Some strains (particularly Type III) give characteristic mucoid colonies. Old laboratory strains of pneumococci, particularly when grown on a relatively unfavourable medium such as ordinary nutrient agar, often give smaller colonies which lack the characteristic appearance of a recently isolated strain grown on a favourable medium. The consistency of the colonies is butyrous, and the growth emulsifies easily. In blood agar plates, the colonies are surrounded by a zone of  $\alpha$  hemolysis showing the characteristic green coloration. In a suitable fluid medium, a filtrable hemolysin is formed, which is of the oxygen labile type.

In broth, or serum broth, *Str. pneumoniae* gives a diffuse turbid growth, with a slight deposit, increasing on prolonged incubation. No pellicle is formed.

On gelatin very slight growth, usually none at or below 20° C.

Gelatin stab—very slight growth along track, with minimal surface growth. No liquefaction.

Potato—growth slight, or absent.

**HEAT RESISTANCE AND VIABILITY**—*Str. pneumoniae* is sensitive to heat, being killed at a temperature of 55° C in 20 minutes or less. It is a relatively delicate organism and dies out rapidly in artificial cultures unless maintained under particularly favourable conditions, as, for instance, in semi solid agar to which blood has been added.

**BIOCHEMICAL ACTIVITIES**—The pneumococcus produces acid, but no gas, from lactose, saccharose, and inulin, and usually from raffinose. Salicin is rarely fermented, when fermented, acid is not usually produced for some days. Mannitol is not fermented. Milk is acidified, and frequently clotted. Nitrates are not reduced. Indole is not formed. Gelatin is not liquefied. The pneumococcus is soluble in bile.

**ANTIGENIC STRUCTURE**—The pneumococcus possesses a species specific carbohydrate antigen, the presence of which is not detected by agglutination reactions carried out with normal smooth forms. The species is divided into a number of antigenic types by type specific polysaccharide antigens contained in the capsules. Over seventy of these types have so far been identified.

**PATHOGENICITY AND TOXIN PRODUCTION**—The pneumococcus causes pneumonia and certain other infections in man. It is highly pathogenic for mice and slightly less so for rabbits. Guinea pigs are rather more resistant, and cats, dogs, fowls and pigeons much more resistant.

The pneumococcus produces a soluble hemolysin, and a leucocidin. It also produces a substance acting on rabbit fibrin or fibrinogen, and preventing the formation of a clot.

## Streptococci of the Viridans Group

**MORPHOLOGY**—Cocci in short or long chains, spheroidal or ovoid, when ovoid, long axis in axis of chain. Non-capsulated. No spores. Non motile Gram positive Not acid fast.

**GROWTH REQUIREMENTS**—Most strains grow more readily on ordinary media than does *Str pyogenes* or *Str pneumoniae* but growth is usually improved by addition of blood or serum. Some strains, on the other hand, grow very poorly. Optimal temperature for most strains 37° C—the range of temperature for growth extends further than that of *Str pyogenes* or *Str pneumoniae* in the downward direction. Aerobic and facultatively anaerobic.

**TYPE OF GROWTH**—On solid media the colonies do not differ in any distinctive way from those of *Str pyogenes* (see above). With streak cultures, the growth may be slightly more profuse and more confluent.

In blood agar plates—The colonies are surrounded by a zone of  $\alpha$  hæmolysis, showing the characteristic green coloration.

In broth—The type of growth varies with chain length. Many strains or varieties grow in short chains and produce a uniform but slightly granular, turbidity in broth with little or no deposit, but some strains grow in long chains and give growths which are indistinguishable from those of *Str pyogenes*.

On gelatin or in gelatin stab, the growth does not differ from that of *Str pyogenes* except that it may be slightly more profuse.

On potato growth is slight and often not detectable by the naked eye.

**HEAT RESISTANCE**—Most strains are killed by heating at 55–56° C for 30 minutes. The general vitality is greater than that of *Str pyogenes*.

**BIOCHEMICAL REACTIONS**—Milk is acidified and often clotted. Most strains produce acid from lactose and saccharose often from raffinose and/or salicin rarely from inulin or mannitol. Nitrates are not reduced. Indole is not formed. Gelatin is not liquefied. Not soluble in bile.

**PATHOGENICITY AND TOXIN FORMATION**—Viridans streptococci form no soluble toxin, nor hæmolyxin. Usually non pathogenic for laboratory animals other than the rabbit in which some strains give rise to arthritis and valvular lesions. They are a common cause of subacute ulcerative endocarditis in man.

**DIFFERENTIATION WITHIN THE GROUP**—The viridans group of streptococci certainly contains more than a single species. It is possible that the common streptococcus of the human mouth which has the peculiar property of producing a soluble levan from sucrose and raffinose and of forming large mucoid colonies when grown on agar containing 5 per cent of these sugars deserves specific rank with the title *Str salivarius*, that the common streptococcus of bovine faeces should be recognized as *Str bovis*, and that a streptococcus isolated from Kefir which produces CO<sub>2</sub> from lactose when grown in Eldridge tubes should be known as *Str Lefir*. Our knowledge is however, not yet sufficient to allow us to define species or to assign specific names with any degree of certainty.

*Str faecalis*

**MORPHOLOGY**—Ovoid cocci, growing in pairs or short chains. Some strains resemble the pneumococcus in morphology, but possess no capsule. More rarely, the appearance may be almost bacillary. Most strains non motile, but a few motile strains have recently been described. No spores are formed. Gram positive and not acid fast.

**GROWTH REQUIREMENTS**—Grows well on the ordinary laboratory media. Optimal temperature about 37° C but grows well up to 45° C and down to 10° C. Aerobic and facultatively anaerobic.

**TYPE OF GROWTH** On solid media the colonies are somewhat larger than those of the species referred to above. After 24 hours the colonies are usually 0.75 mm in average

diameter, and, on longer incubation increase to a diameter of 1.2 mm. The colonies are smooth, circular, low convex in elevation, with an entire edge, they have a homogeneous, or slightly granular structure and show little differentiation. In streaked cultures the colonies tend to be confluent, and growth may appear as a uniform film. The growth is easily emulsified.

*In blood agar*—some strains, corresponding to Lancefield's Group D hemolytic streptococci, give  $\beta$  hemolysis. They form no filtrable hemolysin when tested by the ordinary methods, but a hemolysin of the oxygen stable type may be demonstrated by special cultural methods. Most strains are non-hemolytic.

*On MacConkey agar* small pink colonies are formed.

*In broth* there is an abundant diffuse growth, with a very slight deposit. No pellicle formation occurs.

*On gelatin*, there is a good growth with colonies very similar to those produced on agar. Growth occurs at 10° C. Most strains fail to liquefy the gelatin but a few do so.

*In gelatin stab*, there is good growth along the track with little surface growth. Some strains produce liquefaction which is usually infundibuliform.

Grows in medium containing 10 or 40 per cent bile in lactose agar of pH 9.6 or containing 6.5 per cent NaCl in lactose broth containing 1:15,000 potassium tellurite and in milk containing 1:1,000 methylene blue.

**HEAT RESISTANCE AND VIABILITY**—Heat resistant. Withstands a temperature of 60° C. for 30 minutes. Survives in culture for a long time. Insoluble in bile. Insensitive to penicillin.

**BIOCHEMICAL ACTIVITIES**—Usually fails to hydrolyse sodium hippurate. Produces a final pH of 4.0–4.8 in glucose broth. Produces acid in lactose, mannitol, salicin and usually sucrose, trehalose and sorbitol but not in raffinose or inulin. Reduces the dyes and produces acid and clot in litmus milk. Most strains reduce nitrates. Strains of the *liquefaciens* and *zymogenes* varieties liquefy gelatin and digest casein. Some strains produce  $H_2S$ .

**ANTIGENIC STRUCTURE**—Possesses the Group D specific polysaccharide. There appear to be several antigenic types.

**PATHOGENICITY AND TOXIN PRODUCTION**—Most strains appear to be non-pathogenic or to possess a pathogenicity of a low order. They occasionally cause urinary infections in man or infections in relation to the intestinal tract. They have occasionally been isolated from the blood stream and are the cause of some cases of subacute endocarditis. Most strains are non-pathogenic for laboratory animals. A few show some degree of pathogenicity. There is no evidence that any of the streptococci form a filtrable toxin. Such strains as have been examined do not form a fibrinolysin acting on human fibrin.

**DIFFERENTIATION WITHIN THE GROUP**—There appear to be a number of varieties of this organism such as those called *zymogenes liquefaciens* and *durans* (see p. 585). Whether these have any special habitat apart from the human intestine is not known.

### Str. faecalis

This organism is found in milk and milk products and on certain plants. Its differentiation from *Str. faecalis* has been discussed in the body of the chapter (see p. 582). Suffice it to say that the main distinguishing characters are as follows: it does not grow at 45° C., it does not grow in lactose agar of pH 9.6 or containing 6.5 per cent NaCl, it is killed by a temperature of 60° C. within 30 minutes, it has no action on blood, mannitol, sucrose, and sorbitol are less readily fermented, and the organism possesses the Group N, not the Group D specific polysaccharide. A *cremoris* variety has been described.

The characters of the other labelled groups or types of streptococci, as far as we yet know them, have been summarized in the body of this chapter.

## REFERENCES

- ANDERSON A. B. and HART, P. D. A. (1934a) *J. Path. Bact.*, **39**, 463. (1934b) *Lancet*, ii, 359.
- ANDREWS, F. and HORDER, T. (1906) *Lancet*, ii, 703, 773, 832.
- ARMSTRONG, R. P. (1932) *Brit. med. J.*, i, 187.
- AVERT, O. T. (1915) *J. exp. Med.*, **22**, 804.
- AVERT, O. T., CHICKERING, H. T., COLE, R., and DOCHET, A. P. (1917) *Monogr. Rockefeller Inst. med. Res.*, No. 7.
- AVERT, O. T. and CULLEN, G. E. (1919) *J. exp. Med.*, **29**, 215. (1923) *Ibid.*, **33**, 199.
- AVERT, O. T. and GOSSEL, W. F. (1933) *J. exp. Med.*, **58**, 731.
- AVERT, O. T. and HEIDELBERGER, M. (1923) *J. exp. Med.*, **38**, 81. (1925) *Ibid.*, **42**, 367.
- AVERT, O. T., HEIDELBERGER, M., and GOSSEL, W. F. (1925) *J. exp. Med.*, **42**, 709.
- AVERT, O. T. and MORGAN, H. J. (1924) *J. exp. Med.*, **39**, 233, 289. (1925) *Ibid.*, **42**, 347.
- AVERT, O. T. and NEILL, J. M. (1924a) *J. exp. Med.*, **39**, 347. (1924b) *Ibid.*, **39**, 357. (1924c) *Ibid.*, **39**, 543.
- AVERT, R. C. (1924a) *J. exp. Med.*, **50**, 463. (1924b) *Ibid.*, **50**, 78.
- AYERS, S. H. and JOHNSON, W. T. (1910) *Bull. U.S. Dep. Agric.*, B.A.I., 126. (1913) *Ibid.*, 161. (1914) *J. infect. Dis.*, **34**, 49.
- AYERS, S. H., JOHNSON, W. T., and DAVIS, B. J. (1918) *J. infect. Dis.*, **23**, 200.
- AYERS, S. H., JOHNSON, W. T., and MUDGE, C. S. (1914) *J. infect. Dis.*, **34**, 29.
- AYERS, S. H. and MUDGE, C. S. (1922) *J. infect. Dis.*, **31**, 40. (1923) *Ibid.*, **33**, 153.
- AYERS, S. H. and PETER, P. (1922) *J. infect. Dis.*, **30**, 353.
- AYERS, S. H., RUFF, P., and MUDGE, C. S. (1921) *J. infect. Dis.*, **29**, 233.
- BAHR, J. (1910) *Arch. Hyg., Berl.*, **72**, 91.
- BAGGER, S. V. (1926) *J. Path. Bact.*, **29**, 225.
- BAILEY, P. L. and BATTLE, J. (1940) *Aust. vet. J.*, **16**, 140.
- BECKLEY, E. and MACLEOD, P. (1934) *J. clin. Invest.*, **13**, 901.
- BELENKY, D. E. and POPOWA, N. N. (1929) *Zbl. Bakt.*, **113**, 22.
- BERGER, E. and SILBERSTEIN, W. (1926) *Klin. Wochr.*, **5**, 2307.
- BESKEDKA, A. (1901) *Ann. Inst. Pasteur*, **15**, 830.
- BINGOLD, K. (1921) *Fortschr. Arch.*, **232**, 22. (1932) *Dtsch. med. Wochr.*, **57**, 443.
- BLISS, E. A. (1937) *J. Bact.*, **33**, 623.
- BLISS, W. P. (1920) *Johns Hopk. Hosp. Bull.*, **31**, 173. (1922) *J. exp. Med.*, **38**, 575.
- BRACH, H. (1912) *Zbl. Bakt.*, **62**, 353.
- BROADBENT, J. (1915) *J. infect. Dis.*, **17**, 277.
- BROWN, J. H. (1919) *Monogr. Rockefeller Inst. med. Res.*, No. 9. (1937) *J. Bact.*, **34**, 35. (1939) *Ibid.*, **37**, 133.
- BROWN, R. and POBINSON, L. K. (1943) *J. Immunol.*, **47**, 7.
- CAPPS, J. A. and DAVIS, D. J. (1914) *Arch. intern. Med.*, **14**, 630.
- CHAIN, E. and DUTHIE, E. S. (1930) *Nature, Lond.*, **144**, 977. (1940) *Brit. J. exp. Path.*, **21**, 324.
- CHANNON, H. A. and McLEOD, J. W. (1929) *J. Path. Bact.*, **32**, 283.
- CORVEN, A. F. and PAULL, R. H. (1941) *J. exp. Med.*, **73**, 501.
- COLE, R. (1914) *J. exp. Med.*, **20**, 346.
- COLERBROOK, DORA C. (1935) *Spec. Rep. Ser. med. Res. Coun., Lond.*, No. 205.
- COLERBROOK, L. (1930) *Brit. med. J.*, ii, 134.
- COLERBROOK, L. and HARR, R. (1933) *J. Obstet. Gynaec.*, **40**, 609.
- COLERBROOK, L., ELLIOTT, S. D., MAXTED, W. R., MORLEY, C. W., and MORTON, M. (1942) *Lancet*, ii, 30.
- COLERBROOK, L., MAXTED, W. R., and JONES, A. M. (1933) *J. Path. Bact.*, **41**, 521.
- COOPER, G., EDWARDS, M., and ROSENSTEIN, C. (1929) *J. exp. Med.*, **49**, 461.
- COOPER, G., ROSENSTEIN, C., WALTER, A., and PETER, L. (1930) *J. exp. Med.*, **55**, 531.
- COOPER, G. and WALTER, A. W. (1935) *Amer. J. path. Hlth.*, **25**, 463.
- COWAN, S. T. (1934) *J. Path. Bact.*, **38**, 61.
- CROWLEY, N. (1944) *J. Path. Bact.*, **58**, 27.
- DAVIS, D. J. (1912) *J. Amer. med. Ass.*, **58**, 1832. (1929) *Ibid.*, **83**, 978.
- DAVIS, D. J. and POSENOW, E. C. (1912) *J. Amer. med. Ass.*, **58**, 773.
- DAWSON, M. H., HOBBS, G. L., and OLMTREAD, M. (1933) *J. infect. Dis.*, **62**, 133.
- DEHL, J. H. (1921) *J. Path. Bact.*, **24**, 3.
- DICK, G. F. and DICK, G. H. (1924a) *J. Amer. med. Ass.*, **82**, 265. (1924b) *Ibid.*, **83**, 84. (1925a) *Ibid.*, **84**, 802. (1925b) *Ibid.*, **84**, 1477.
- DOCHET, A. P. and AVERT, O. T. (1915) *J. exp. Med.*, **21**, 114.
- DOCHET, A. P., AVERT, O. T., and LANCEFIELD, R. C. (1919) *J. exp. Med.*, **30**, 179.
- DOCHET, A. P. and GILLESPIE, L. P. (1913) *J. Amer. med. Ass.*, **61**, 77.
- DOWNIE, A. W. and CRICKSHANK, J. (1923) *Brit. J. exp. Path.*, **9**, 171.
- DOWNIE, A. W., STENT, L., and WHITE, S. M. (1931) *Brit. J. exp. Path.*, **12**, 1.
- DUBIN REYNOLDS, F. (1933) *J. exp. Med.*, **58**, 161.

- EAGLES, G H (1924) *Brit J exp Path.*, 5, 199, (1928) *Ibid.*, 9, 330  
 EDDY, B E (1944) *Publ Hlth Rep.* Wash., 59, 449-451, 1041  
 EDWARDS, P R (1932) *J Bact.*, 23, 259, (1933) *Ibid.*, 25, 527, (1934) *Ibid.*, 27, 527  
 EHRLICH, O (1943) *Arch Hyg.* Berl., 129, 116  
 ELLIOTT, S D (1943) *Brit J exp Path.*, 24, 159, (1944) *Pers comm*  
 ENDERS, J F. (1930) *J exp Med.*, 52, 235  
 ETINGER TULCZYNSKA, R (1932) *Z Hyg Infektkr.*, 114, 769  
 EVANS, A C (1931) *Publ Hlth Rep.* Wash., 46, 2539  
 FALK, I S and YANG, S Y (1926) *J infect Dis.*, 38, 1  
 FEHLRESEN (1883) "Aetiology des Erysipels" Berlin  
 FIELDS P and GLADSTONE, G P (1939) *Brit J exp Path.* 20, 334  
 FLEMING A (1932) *J Path Bact.*, 35, 831, (1941) *Lancet*, 1, 110  
 FLOYD, C and WOLBACH, S B (1914) *J med Res.*, 29, 493  
 FRANKEL, A (1896) *Z klin Med.*, 10, 401, (1896) *Ibid.* 11, 437  
 FREUDENREICH, E von (1897) *Zell Bakt.*, 11te Abt., 3, 47  
 FROST, W D, GUMM, M, and THOMAS, R C (1927) *J infect Dis.*, 40, 698  
 FRY, R M (1933) *J. Path Bact.*, 37, 337, (1941) *Pers comm*  
 FULLER, A T. (1938) *Brit J exp Path.*, 19, 130  
 FULLER, A T and MAXTED, W R. (1939) *J Path Bact.*, 49, 83  
 FULLER, C A and ARMSTRONG, V A (1913) *J infect Dis.*, 13, 442  
 GARNER, R L and TILLET, W S (1934) *J exp Med.*, 60, 239  
 GAY, F. P. and GRAM, F (1933) *J Immunol.*, 25, 501  
 GOODNER, K. (1931) *J exp med.*, 54, 847, (1933) *Ibid.*, 58, 153  
 GORDON, M H (1902-3) *Rep loc. Gort Bd publ Hlth.*, 32, 421, (1903-4) *Ibid.*, 33, 388, 422, (1905) *Lancet*, 1, 1400, (1921) *Brit med J.*, 1, 632  
 GRAHAM, N C and BARTLEY, F O (1939) *J Hyg.* Camb., 39, 538  
 GRIFFITH, F. (1923) *Rep publ. Hlth med Subj.*, Lond., No 18, (1926) *J Hyg Camb.*, 25, 385; (1927) *Ibid.*, 26, 303, (1928) *Ibid.*, 27, 113, (1934) *Ibid.*, 34, 542 (1935) *Ibid.*, 35, 23  
 GRUMBACH, A. and SCHWETZ, A (1938) *Schweiz. Z allg Path Bakt* 1, 59  
 GUNDEL, M. and SCHWAB, K T (1932) *Z Hyg Infektkr.*, 113, 498  
 GUNN, W. and GRIFFITH, F (1928) *J Hyg.* Camb., 28, 250  
 GUTSALUS, I C and SHERMAN, J M (1943) *J Bact.*, 45, 155  
 GÜTHER, C and THIERFELDER, H (1895) *Arch Hyg.* Berl., 25, 164  
 HADLEY, F. P., HADLEY, P., and LEATHEN, W W (1941) *J infect Dis.* 68, 264  
 HARE, R. (1935) *J. Path Bact.*, 41, 499  
 HARE, R and COLEBROOK, L. (1934) *J Path Bact.*, 39, 429  
 HARE, R and MAXTED, W R (1935) *J Path Bact.*, 41, 513  
 HARRIS, J W and BROWN, J H (1929) *Johns Hopk Hosp Bull.*, 44, 1  
 HART, P D A and ANDERSON, A B (1933) *J Path Bact.*, 37, 91  
 HARTLEY, P (1928) *Brit J exp Path.*, 9, 259  
 HEIDELBERGER, M (1927) *Physiol Rev.*, 7, 107  
 HEIDELBERGER, M and AVERY, O T (1923) *J exp Med.*, 38, 73, (1924) *Ibid.*, 40, 301  
 HEIDELBERGER, M and GOEBEL, W F (1927) *J biol Chem.*, 74, 613  
 HEIDELBERGER, M, GOEBEL, W F, and AVERY, O T (1925) *J exp Med.*, 42, 727  
 HEIDELBERGER, M and KENDALL, F E (1931) *J exp Med.*, 53, 625  
 HEINEMANN, P G (1906) *J infect Dis.*, 3, 173  
 HERBERT, D and TODD, E W (1944) *Brit J exp Path.*, 25, 242  
 HIRST, G K (1941) *J exp Med.*, 73, 493  
 HITCHCOCK, C H (1924) *J exp Med.*, 40, 445  
 HOBBS, R C (1939) *J Dairy Res.*, 10, 35  
 HOLMAN, W L (1916) *J med Res.*, 34, 377  
 HOPKINS, J G and LANG, A (1914) *J infect Dis.*, 15, 67  
 HOTTELE, G A and PAPPENHEIMER, A M (1941) *J exp Med* 74, 545  
 HOUSTON, T and McCLOY, J M. (1916) *Lancet*, 1, 632  
 HUMPHREY, J H (1914) *J Path Bact* 56, 273  
 JAMES, G R (1926) *J Hyg.* Camb., 25, 415  
 JONES, F S (1919) *J exp Med.*, 30, 159  
 JULIANELLE, L A and REIMANN, H A (1926) *J exp Med.*, 43, 87  
 KAUFFMANN, F, MORCH, E and SCHMIDT, K. (1940) *J Immunol.*, 39, 397  
 KEMPFER, W and SCHIAFFR, C (1942) *J Bact.*, 43, 387  
 KENDALL, F E, HEIDELBERGER M and DAWSON, M H (1937) *J biol Chem.*, 118, 61  
 KEOGH, E V and SIMMONS R. T (1940) *J Path Bact.*, 50, 137  
 KISSLING, K. (1924) *Munch. med Wochr.*, 71, 1457, (1929) *Ibid.*, 76, 1163  
 KITT (1893) see Minett (1935b)  
 KLECKNER, A L. (1935) *J Lab clin Med.*, 21, 111  
 KLEIN, S J (1933) *J Bact.*, 26, 215, (1935) *Ibid.*, 30, 43

- KLEIN, S. J. and STOVER, F. M. (1931) *J. Bact.* 22, 387.  
 KUMMER, M. and HAUFF, H. (1930) *Erythr. Hyg.* 11, 354.  
 KORNBLUMER, L. O. (1935) *Zbl. Bakt.* 123, 310.  
 KRÖSTIG (1935) *Zbl. Gynäk.* 19, 409.  
 KRUIP, P. H. DE and IRELAND, P. M. (1930) *J. infect. Dis.* 28, 255.  
 KREMWITZER, E. (1943) *J. Bact.* 48, 117.  
 LAMANTA, C. (1944) *J. Bact.* 47, 327.  
 LANCEFIELD R. C. (1925) *J. exp. Med.* 47, 91, 469, 481, 843, 857, (1933) *Ibid.* 57, 571.  
 (1934) *Ibid.* 59, 441, (1935) *Ibid.* 67, 25, (1940) *Ibid.* 71, 521, 539, (1941) *Harvey Lectures Ser.* 36, 251, (1943) *J. exp. Med.* 78, 465.  
 LANCEFIELD, R. C. and HARR, R. (1935) *J. exp. Med.* 61, 335.  
 LANCEFIELD, R. C. and STEWART, W. A. (1941) *J. exp. Med.* 79, 79.  
 LANCEFIELD R. C. and TODD, E. W. (1925) *J. exp. Med.* 43, 769.  
 LEBMANN, W. (1926) *Munch. med. Wochschr.* 73, 233.  
 LISTER, P. S. (1916) *Publ. S. Afr. Inst. med. Res.* No. 8.  
 LOGAN W. R. (1914) *J. Path. Bact.* 18, 527.  
 LOGAN, W. R. and SWEALL, J. T. (1932) *Brit. med. J.*, 1, 183.  
 LONG, P. H. and BLISS, E. A. (1934) *J. exp. Med.* 60, 619.  
 LOWENWITZ, X. (1901) *Arch. med. Exp.* 13, 633.  
 LYALL, H. W. (1914) *J. med. Res.* 30, 497.  
 MACCALLUM W. G. and HASTINGS, T. W. (1909) *J. exp. Med.* 4, 521.  
 McCLEARY, D. (1941) *J. Path. Bact.* 53, 15, (1942) *Ibid.* 54, 254.  
 MACDONALD I. (1929) *Med. J. Aust.* 11, 471.  
 MCLACHLAN, D. G. S. and MACKIE, T. J. (1928) *J. Hyg. Camb.* 27, 225.  
 McLEOD, J. W. (1912) *J. Path. Bact.* 16, 321.  
 McLEOD, J. W. and GORDON, J. (1922) *Biochem. J.* 16, 499.  
 McLEOD, J. W. and McNEIL, J. W. (1913) *J. Path. Bact.* 17, 524.  
 MAIR, W. (1917) *J. Path. Bact.* 21, 305, (1925) *Ibid.* 31, 215, (1929) "A System of Bacteriology" *Med. Res. Coun.*, London, 2, 165.  
 MARDELBAUM, M. (1907) *Z. Hyg. Infektkr.* 58, 25.  
 MARMONIER, A. (1925) *Ann. Inst. Pasteur*, 9, 593.  
 MARWEDL and WEHSELO (1915) *Munch. med. Wochschr.* 62, 1023.  
 MEADER, P. D. and ROBINSON, G. H. (1929) *J. exp. Med.* 52, 639.  
 MEYER, K. (1926) *Zbl. Bakt.* 99, 416.  
 MEYER, K. and PALMER, J. W. (1935) *J. Ind. Chem.* 114, 629.  
 MEYER, K. and SCHÖNFELD, H. (1926) *Zbl. Bakt.* 99, 402.  
 MINETT, F. C. (1935a) *J. Path. Bact.* 43, 357, (1935b) *Proc. 24 Int. Vet. Cong.*, p. 511, (1935c) *J. Hyg. Camb.* 35, 504.  
 MINETT, F. C. and STABLEFORTH, A. W. (1931) *J. comp. Path.* 44, 114, (1934) *J. Dairy Res.* 5, 223.  
 MINETT, F. C., STABLEFORTH, A. W., and EDWARDS, S. J. (1929) *J. comp. Path.* 42, 215, (1936) *J. Hyg. Camb.* 35, 504.  
 MORCH, E. (1942) *J. Immunol.* 43, 177.  
 MORGAN, H. J. and AVERY, O. T. (1924) *J. exp. Med.* 33, 335.  
 MORGAN H. J. and NEILL, J. M. (1924) *J. exp. Med.* 40, 269.  
 MORISOV J. E. (1940) *J. Path. Bact.* 51, 401.  
 MORTON H. E. and SOMMER, H. E. (1944) *J. Bact.* 47, 123.  
 NAKAYAMA, Y. (1920) *J. infect. Dis.* 27, 85.  
 NASTIG, H. (1905) *Arch. Gynäk.* 78, 701.  
 NEILL, J. M. (1925) *J. exp. Med.* 41, 229, 535, (1929) *Ibid.* 44, 199.  
 NEILL, J. M. and AVERY, O. T. (1924a) *J. exp. Med.* 33, 757, (1924b) *Ibid.* 40, 405, 423, (1925) *Ibid.* 41, 255.  
 NEILL, J. M. and MALLORY, T. B. (1926) *J. exp. Med.* 44, 241.  
 NEISSER, M. and WECHSELBERG, F. (1900) *Munch. med. Wochschr.* 47, 1261, (1901) *Z. Hyg. Infektkr.* 26, 299.  
 NEUFELD, F. (1900) *Z. Hyg. Infektkr.* 34, 454, (1902) *Ibid.* 40, 54.  
 NEUFELD, F. and ETINGER-TULCHINSKY, R. (1931) *Z. Hyg. Infektkr.* 112, 402.  
 NEUFELD, F. and HINDEL, L. (1909) *Arch. Roentgenröntgenstr.* 34, 293.  
 NICOLLE, M. and ADRIEN, (1907) *Ann. Inst. Pasteur*, 21, 20.  
 NILES C. F., SMILEY, K. L., and SHESMAN, J. M. (1911) *J. Bact.* 41, 479, (1912) *Ibid.* 43, 651.  
 NOCARD and HOLLEREAU (1887) *Ann. Inst. Pasteur*, 1, 109.  
 OGURA, K. (1929) *J. Jap. Soc. vet. Sci.* 8, 174.  
 ORAM, F. (1934) *J. Immunol.* 26, 233.  
 ORLA-JENSEN S. (1919) "The lactic acid bacteria." Copenhagen.  
 PASTEUR, L., CHAMBERLAND, C., and ROUX, E. (1881) *C. R. Acad. Sci.* 92, 159.  
 PITTMAN, M. and FALK, I. S. (1930) *J. Bact.* 19, 327.



- PLUMMER, H (1935) *J Bact.*, 30, 5, (1941) *J Immunol.* 42, 91  
 POWELL, M (1935) *Brit J exp Path.*, 16, 185  
 IRÉLOT, A. (1924) *Les Streptococcus Anaérobies*, Paris, (1925) *Ann Inst Pasteur*, 39, 417, (1913) *Ann Sci Nat (Series Biologiques)* 15, 163  
 IULVERTZ, R. J. V. (1928) *Brit J exp Path.*, 9, 276.  
 RANE, L. and WYMAN, L. (1937) *J Immunol.*, 32, 321  
 RANTZ, L. A. (1912) *J infect Dis.*, 71, 81  
 REIMANN, H. A. (1927) *J exp Med.*, 45, 807  
 REIMANN, H. A. and JILLIANEUX, L. A. (1926) *J exp. Med.*, 43, 97  
 PIST, E. (1901) *Zbl. Bakt.*, 30, 287  
 ROCHAIX, A. (1924) *C. P. Soc Biol.*, 90, 771  
 ROSENBAUM, F. J. (1884) 'Mikroorganismen bei den Wundinfektionskrankheiten' Wiesbaden.  
 ROSOWSKY, A. (1912) *Zbl Gynak.*, 39, 4  
 POTTER, W. (1925) *Dtsch med Wochr.*, 51, 1031  
 RUDNIGER, G. F. (1912) *Science* 35, 223  
 SABIN, A. B. (1933) *J Amer med Ass* 100, 1584  
 SCHWABEL, A. (1921) *Z Hyg Infectkr.*, 83, 175  
 SCHOTTENFELDER, H. (1903) *Munch. med Wochr.*, 50, 849 909, (1910) *Mitt Grenz geb med Chir.*, 21, 450, (1928) *Munch. med Wochr.*, 75, 1580 1634  
 SCHÜTZ, W. (1888) *Arch. exp. appl. Tierheilk.*, 14, 450  
 SEASTONE, C. V. (1931) *J Bact.*, 28, 481 (1913) *J exp Med.*, 77, 21  
 SEELEMAN, M. and HALENBERG, A. (1932) *Zbl Bakt.*, 126, 71  
 SEELEMAN, M. and NOTTBOUM, H. (1910) *Zbl Bakt* 140, 142  
 SHATTOCK, P. M. F. (1914) *Lancet* comm.  
 SHATTOCK, P. M. F. and MATTHEW, A. T. R. (1913) *J Hyg Camb* 43, 173  
 SHERMAN, J. M. (1921) *J Bact.*, 6, 127  
 SHERMAN, J. M. and ALBERT, W. B. (1918) *J Bact.*, 3, 153  
 SHERMAN, J. M., NIXON, C. F., and SMILEY, K. L. (1913) *J Bact* 45, 249  
 SHERMAN, J. M., SMILEY, K. L. and NIXON, C. F. (1910) *J Dairy Sci* 23, 529  
 SHERMAN, J. M. and STARK, P. (1931) *J Bact.*, 22, 275 (1934) *J Dairy Sci* 17, 529 (1934) *J Bact.*, 38, 77  
 SHERMAN, J. M. and WING, H. (1937) *J Dairy Sci.*, 20, 165  
 SICKLES, G. M. and CORREY, J. M. (1928) *J infect. Dis.*, 43, 490.  
 SILBERSCHNIDT, W. (1902) *Z Hyg Infectkr.*, 41, 427  
 SIMMONS, R. T. and KROGH, F. V. (1910) *Aust J exp Biol med Sci.*, 18, 151  
 SLAVETT, L. W. and NACHSKI, J. (1910) *J infect Dis.*, 66, 80  
 SMITH, F. (1931) *Brit J exp Path.*, 17, 329  
 SMITH, F. R. and SHERMAN, J. M. (1933) *J infect Dis* 65, 301  
 SMITH, J. (1926) *J Hyg, Camb.*, 25, 167, (1927) *Ibid* 26, 420, (1929) *J Path Bact.*, 32, 401  
 SMITH, T. and BROWN, J. H. (1915) *J med Res.*, 31, 47  
 SOLOWAY, M. (1912) *J exp Med.*, 76, 109  
 SOULE, S. D. and BROWN, T. A. (1912) *Amer J Obstet Gynec.*, 28, 532  
 STABLEFORTH, A. W. (1932) *J comp Path.*, 45, 183 (1937) *J Path Bact* 45, 267 (1942) *Proc R Soc Med.*, 35, 625 (1945) *Lancet* comm.  
 STARK, P. and SHERMAN, J. M. (1935) *J Bact.*, 30 639  
 STERNBERG (1900) *Wien Klin Wochr.*, 13, 531  
 STEVENS, F. A. and DOCHER, A. R. (1926a) *J exp Med.*, 43, 379, (1926b) *Ibid*, 44, 439  
 STEWART, D. F. (1937) *J Path. Bact.*, 45, 279  
 STEWART, W. A., LANCEFIELD, R. C., WILSON, A. T., and SWIFT, H. F. (1911) *J exp Med.*, 79, 91  
 STOKES, W. P. and MACHTEL, F. W. (1912) *Publ Hlth Rep.*, Wash., Nov 22  
 STONE, M. L. (1910) *J Bact.*, 39, 509  
 SWIFT, H. F., WILSON, A. T., and LANCEFIELD, R. C. (1913) *J exp Med.*, 78, 127  
 THALMANN (1912) *Zbl Bakt.*, 66, 240  
 THIERCELYN (1899) *O R Soc Biol.*, 5, 269  
 TILLET, W. S. (1927) *J exp Med.*, 45, 1093, (1935) *J Bact.*, 29, 111  
 TILLET, W. S., EDWARDS, L. B., and GARNER, L. L. (1931) *J clin Invest.*, 13, 47  
 TILLET, W. S. and GARNER, R. L. (1933) *J exp Med.*, 58, 485  
 TILLET, W. S., GOEBEL, W. F., and AVERY, O. T. (1930) *J exp Med.*, 52, 95  
 TODD, E. W. (1928a) *Brit J exp Path.* 9, 1, (1928b) *J exp Med.*, 48 493, (1930) *Brit J exp Path.* 11, 368 469, 480, (1932) *J exp Med* 55, 267 (1934) *J Path Bact.*, 39, 239, (1934a) *Ibid* 47, 423, (1934b) *Brit J exp Path.*, 19, 367, (1939) *J Hyg, Camb.*, 39, 1, (1942) *Brit J exp Path.*, 23, 136  
 TODD, F. W. and HEWITT, L. F. (1932) *J Path Bact.*, 35, 973  
 TODD, E. W. and LANCEFIELD, R. C. (1928) *J exp Med.*, 48, 751

- TODD E W LAURENT I J M., and HILL, V G (1933) *J Path Bact.*, 26, 201  
 VELLON M A (1893) *C P Soc. Biol* 40 807  
 VELDE, H VAN DER. (1894) *La Cellule* 10 401  
 WADSWORTH A and BROWN P (1931) *J Immunol* 21, 245  
 WALTER A W GUEIN V H BEATTIE M W COYLER, H V., and BUCCA, H B (1941)  
*J Immunol* 41, 225  
 WATSON I F and LANCEFIELD P C (1944) *J exp Med* 79 89  
 WEATHERALL, C and DILL, J H (1929) *J Path. Bact.* 32, 413  
 WEGELIUS W (1909) *Arch Gynaek* 88 249  
 WEISELBAUM A (1886) *Med Jb* 1 463  
 WEISSENBACH R J (1918) *C R Soc Biol* 81, 559 819  
 WELD J T (1934) *J exp Med* 59 83 (1935) *Ibid* 61, 43.  
 WHITE, C FULD G V and WARD H H (1933) *Med J Aust* 2 98  
 WHITE F (1933) *J Obstet Gynaec* 40 630  
 WHITE S M (1929) *Biochem J* 23, 1165  
 WINSLOW C E A BROADHURST J BUCHANAN P E KEUKWIEDE, C., ROGERS, L A.,  
 and SMITH G H (1920) *J Bact.* 5 191  
 WINSLOW C E A and PALMER, G T (1910) *J infect. Dis.*, 7 1  
 WRIGHT H D (1936) *J Path Bact* 43, 487  
 LAWGER, F S and SHERMAN J M (1937) *J Dairy Sc.*, 20 20.  
 ZITTEL C A (1942) *J Immunol* 43 31

## CHAPTER 25

### STAPHYLOCOCCUS, MICROCOCCUS, SARCINA, RHODOCOCCUS, AND LEUCONOSTOC

#### STAPHYLOCOCCUS

##### DEFINITION —*Staphylococcus*

Spherical or ovoid, non motile, Gram positive cells, arranged in grape like clusters on solid media, and in pairs small groups or short chains in liquid media. On agar the growth is of a golden, white, or yellow colour. Great variation in biochemical activities, hæmolytic power and pathogenicity. Actual or potential parasites.

Type species is *Staphylococcus aureus* Rosenbach.

**HISTORY**—The presence of micrococci in pus was noted by Koch in 1878, they were cultivated in a liquid medium by Pasteur in 1880, they were shown by Ogston in 1881 to be constantly present in acute and chronic abscesses, they were cultivated by him in eggs, and were found to be pathogenic to mice and guinea pigs; but it was left to Rosenbach in 1884 to make a thorough study of the staphylococci, to obtain pure cultures on solid media, and to divide them into two species—*Staphylococcus pyogenes aureus* and *Staphylococcus pyogenes albus*. In the following year Passet (1885) added another species—*Staphylococcus pyogenes citreus*. In 1887 Biondi isolated two types from saliva, both pathogenic for laboratory animals, one of these was apparently identical with an organism described as *M. tetragenus* by Koch and Gaffky (Gaffky 1883), who had found it in the sputum of patients suffering from pulmonary tuberculosis, the other was distinguished from *Staphylococcus aureus* by its diminutive size— $0.3-0.5 \mu$  in diameter—and the slowness with which it liquefied gelatin, to this he gave the name of *Staphylococcus salivarius pyogenes*. Welch (1891) noticed a white staphylococcus in stitch abscesses following the suturing of operation wounds, this he called the *Staphylococcus epidermidis albus*. Andrewes and Gordon (1905-6) who investigated a large number of cocci from different sources, found a special type commonly present in saliva which they named the *Staphylococcus salivarius*, this differed in many respects from the *Staphylococcus salivarius pyogenes* of Biondi, they also found a coccus of peculiar characteristics present in scurf, but did not identify it by a special name. Winslow and Rogers (1906), in an attempt to arrive at a classification on a statistical basis, conducted a painstaking investigation into the *Coccaceæ*, and proposed a division into six genera, from which the original genus *Staphylococcus* was omitted. Later, however, Winslow, Rothberg and Parsons (1920) modified this classification, and reinstated the *Staphylococcus* in its old place, dividing the genus into six species.

The differentiation between these two types morphologically, therefore, is not always easy, on a single medium alone it may be impossible, but if the appearance of the coccus is studied in both liquid and solid media, not much difficulty will be experienced. The differential points are (1) that the chains formed by staphylococci rarely contain more than four members and (2) that the clusters formed by streptococci generally consist of aggregations of chains, the chain is the fundamental unit of the streptococcus.

**Staining Reactions**—The staphylococci stain well with most of the aniline dyes and are uniformly Gram positive. It is not uncommon, however, to see them described as being sometimes negative. Winslow, Rothberg and Parsons (1920), in a study of 180 strains, encountered 5 Gram negative strains and the same authors make the generalization that, whereas the orange and white cocci are Gram positive, the yellow and red ones, including the *Sarcinae*, are Gram negative. This discrepancy can be explained by the facts firstly that different types of staphylococci do vary in their resistance to decolorization, and secondly that many strains which are Gram positive in an 18 or 24 hours culture become Gram negative as they grow older. To obtain uniformity, therefore, it is essential to use a young culture—never more than a day old—and not to prolong unduly the process of decolorization. If these precautions are taken, it will be found that almost without exception the staphylococci, at any rate on first isolation as also the *sarcinae*, react positively to Gram's stain.

**Cultural Reactions**—The staphylococci are among the easiest of micro organisms to cultivate *in vitro*. Though some develop more slowly than others particularly *Staphylococcus citreus*, they all give abundant growths.

In nutrient broth after 24 hours at 37° C there is a moderate to dense turbidity, with a moderate deposit of a powdery nature, which, on shaking swirls up and disappears completely, increasing the turbidity. This is the usual picture. But some types, the salivary staphylococci for example, form a thick, weedy, glutinous deposit, leaving the supernatant fluid clear. After 2 days incubation, a surface ring growth is generally present. In no case is there any distinctive pigment formation in fluid media.

On nutrient agar, there is produced within 1 to 2 days a moderately thick, raised confluent growth with a moist, glistening, smooth or somewhat honey combed surface—due to the imperfect fusion of individual colonies, in most cases it is of butyrous consistency and easy to emulsify, but in the case of the salivary staphylococci it is glutinous, adherent to the medium and more difficult to emulsify. Pigment production is most obvious on agar at 22° C.

On gelatin plates at 22° C development is slower, there being often no visible growth for 2 to 3 days, the colonies which are then formed are small and relatively unpigmented, later a zone of liquefaction may appear around them.

In gelatin stab cultures there is a filiform growth reaching to the bottom of the tube, and a surface growth of variable degree. Liquefaction may or may not occur, when it does, the usual type given by the white and golden cocci is

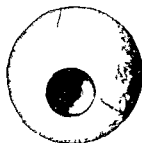


FIG 126—*Staphylococcus aureus*

Surface colony on agar, 24 hours, 37° C (× 8)

**Habitat**—There are but few situations from which staphylococci may not be isolated. In the animal body they are normally found in the nose, on the skin, in saliva, in the intestinal contents, and in feces, they are frequently isolated from suppurative processes (see Pathogenicity), and they are present in varying numbers in air, water, milk, sewage, and on all articles liable to come in contact with these substances. Their main habitat appears to be the nose. Several workers (see Hallman 1937, McFarlan 1933, Gillespie *et al.* 1939) have shown that they are present in the anterior nares of a high proportion of normal persons and that 30 to 60 per cent of persons are nasal carriers of potentially pathogenic staphylococci. Relatively harmless members of the *albus* species are commonly present on the skin, but the more dangerous *aureus* species is found on the hands of only a small proportion of healthy persons who are usually shown, on investigation, to be heavy nasal carriers (Gillespie *et al.* 1939, Miles *et al.* 1944). *Staph. aureus* has been reported in the milk of a high proportion of nursing mothers (Report 1942, Duncan and Walker 1942), but whether this is a normal condition or whether it occurs only in infected maternity wards is not clear. Ubiquitous as staphylococci are their natural habitat is the animal body, and it is the animal body that furnishes the main supply to the outside world.

**Morphology**—The staphylococci consist of round or somewhat oval cells, having an average diameter of  $0.8-1.0 \mu$ . The size is variable, not only from one species to another, but in members of the same species, it depends partly on the age of the culture and the nature of the medium on which it is grown. Some species are generally smaller than others, thus the average diameter of *Staphylococcus aureus* is  $0.7-0.9 \mu$ , whereas the salivary staphylococci are said to be larger,  $1.0-1.2 \mu$ . All the members of the group are non motile, non-flagellated and non sporing. They are usually described as non-capsulated.

Lyons (1937), however, states that capsules are demonstrable in 3-hour broth cultures, but disappear as the culture gets older.

The true staphylococci are arranged in grape-like clusters, that is to say they form groups, the members of which are disposed in three planes of space without regard to any definite configuration. This distribution is best appreciated when a hanging drop preparation is examined—especially if a stereoscopic microscope is employed. The characteristic grouping into clusters is more evident on solid than in liquid media. Indeed in broth it is common to find the cocci occurring not only in groups, but in pairs and in



FIG. 125.—*Staphylococcus aureus*

From an agar culture 24 hours 37° C. ( $\times 1000$ )

short chains. They are then liable to be mistaken for streptococci. This confusion may be accentuated owing to the fact that streptococci on solid media tend to lose their capacity for forming chains, and may develop in small clusters

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FIG 126—*Staphylococcus aureus*

Surface colony on agar, 24 hours 37° C (× 8)

infundibuliform or saccate, by the yellow cocci crateriform. The time of its appearance is likewise subject to a considerable amount of variation, it may be noticeable after 2 days or it may be a fortnight or even longer before it becomes apparent. Speaking broadly, it may be said that the pyogenic staphylococci liquefy gelatin early, the saprophytic ones late.

On *potato* there is a moderate confluent growth. *Loeffler's serum* is a good nutritive medium on which pigment is well developed. On *MacConkey's* neutral red lactose bile salt agar, the colonies are very small, pale pink after 24 hours, and deep pink after 48 hours.

Working with pure cultures, Chapman and Berens (1935) and Chapman (1936) find that on proteose peptone lactose agar containing a concentration of 1-300 000 crystal violet, white, violet, or orange colonies with a violet fringe may be formed. As a rule, the strains giving rise to either of the latter two types of colony are hæmolytic, produce coagulase, and are toxic to rabbits on intravenous inoculation, while strains forming white colonies are negative in all these respects. More recently, Chapman and his colleagues (1937) have described a proteose peptone lactose agar, pH 8.6 containing 0.017 per cent. bromthymol blue, that is said to inhibit the growth of non pathogenic staphylococci, and Chapman (1944) has described an alkaline bromthymol blue agar containing potassium tellurite for isolation of staphylococci from the faeces.

**Variant colonial types**—including rough and G forms—have been described by Hoffstadt and Youmans (1932), Bigger, Boland, and O'Meara (1927), and Swingle (1935).

**Metabolism**—The staphylococci are facultative anaerobes, growing best in the presence of oxygen. On agar plates incubated anaerobically, the growth tends to spread out on the surface of the medium, so that the colonies instead of being convex, are flat and effuse. Lubinski (1894) was the first to point out that no pigment is produced under anaerobic conditions, though, if a culture which has been incubated anaerobically is exposed to the air, it soon develops its characteristic colour. He found, however, that continuous anaerobic cultivation generally caused a strain to lose its power of producing pigment—a power which was not regained by subsequent cultivation in the presence of oxygen.



FIG. 127 — *Staphylococcus aureus*  
Agar stroke culture  
24 hours 37° C.

The limits of temperature between which growth is possible are wide, varying from about 12° to 40° C, but as a rule development is most rapid at 37° C. A slightly alkaline medium of pH 7.4-7.6 is preferable for the initiation of growth, but some growth will occur even at pH 4.0-5.0. Growth is slightly increased by the addition of blood and glucose to the medium, serum has no beneficial effect. According to Hucker (1924a), the staphylococci are unable to use ammonium salts as their sole source of nitrogen. Hughes (1932) and later Knight (1935), both of whom have studied the nutritive requirements of *Staph. aureus*, describe the existence of an essential growth factor, which subsequent work (Knight 1937) has shown to contain vitamin B<sub>1</sub> and nicotinic acid. The organisms can now be grown aerobically on a medium containing only known chemical constituents,

namely a mixture of glucose, salts and fourteen amino acids, supplemented by the two vitamin constituents just mentioned (Fildes *et al* 1936, Knight 1937). According to Klügler, Grossowicz and Bergner (1943), nicotinic acid is the essential factor in the glucose metabolism of *Staph aureus*, thiamin ( $B_1$ ) acts as a catalyser in the oxidation of pyruvic acid. Riboflavin can apparently be synthesized by the organisms themselves (O'Kane 1941). As with other parasitic bacteria, organic sulphur is necessary for growth, this is usually obtained in the form of cystine, but methionine and sodium dithiodiacetate can be substituted (Fildes and Richardson 1937, Gladstone 1937). No growth occurs in the complete absence of  $CO_2$  (Gladstone *et al* 1935). (See also Chapter 3).

Hewitt (1930) has shown that *Staph aureus*, when grown in ordinary infusion broth, brings about a rapid fall in electrode potential to between  $E_h - 0.1$  and  $-0.2$  volt. Owing to lack of peroxide formation the potential remains at a low level for a long time, showing no tendency to rise as it does with streptococci. In glucose broth the potential does not fall as low as in ordinary broth.

**Pigment Production.**—As already mentioned, the staphylococci are active pigment producers. *Staphylococcus aureus* forms a golden, *Staphylococcus citreus* a lemon yellow pigment, while cultures of *Staph albus* are of a porcelain white colour. The development of the pigment and its actual tint depend, however, on several factors. As with many other organisms the optimum temperature for pigment production does not coincide with the optimum temperature for growth: more pigment is produced at  $22^\circ C$  than at  $37^\circ C$ , if cultures that have been incubated at  $37^\circ C$  are subsequently left at room temperature, the colour is seen to deepen. Gelatin and broth are unsuitable. Oxygen is requisite for its development, under anaerobic conditions the growth is colourless. Carbon dioxide is said to favour its production, provided oxygen is also present (Lubinski 1894). An agar medium containing 33 per cent milk is recommended for its study (see Christie and Keogh 1940).

A very important point to remember is that a given strain of staphylococcus under conditions of artificial cultivation may lose its power of producing pigment. The cause of this loss is unknown. It is most noticeable in the case of *Staphylococcus aureus*, which, when freshly isolated from the animal body, gives a rich golden pigment, but often loses this character on prolonged cultivation. This variation adds greatly to the difficulty of classification, and it must be emphasized that in the study of a particular strain, the property of pigment formation should be noted as soon after isolation as possible (Dudgeon 1908).

**Gelatin Liquefaction.**—There is a considerable amount of discrepancy in the reports of various authors as to the power of staphylococci to liquefy gelatin. Of 41 white strains examined by Gordon (1903-4), 24 liquefied gelatin. Kutscher and Konrich (1904) reported upon 57 strains of staphylococci, and found that all liquefied gelatin. Similarly with Klopstock and Bockenheimer (1904) who examined 30 strains, and with Fraenkel and Baumann (1905) who examined 36 strains. Dudgeon (1908) found that 44 out of 46 *aureus* strains and 35 out of 56 *albus* strains liquefied gelatin while Winslow, Rothberg and Parsons (1920), in examining 180 strains found that 67 per cent of the *aureus* and 47 per cent of the *albus* strains liquefied gelatin.

These discrepancies are to be explained partly by possible differences in the cocci examined and partly by the length of time during which growth was observed.



There is general agreement that staphylococci isolated from pathological sources liquefy gelatin more frequently than those isolated from water, air, skin, etc.

There is likewise general agreement that the orange cocci are more rapid liquefiers than the white, and the white cocci more rapid than the yellow.

Summing up, it may be said that *Staphylococcus aureus* liquefies gelatin almost always, *Staphylococcus albus* frequently, and *Staphylococcus citreus* sometimes.

**Resistance to Heat and Disinfectants.**—The staphylococci are among the more resistant of the non-sporing organisms. In broth or agar tubes sealed with paraffin and kept in the ice-chest, cultures may remain alive for months. Dried on threads they retain their vitality for 3 to 6 months, and from dried pus they have been cultivated after 2 to 3 months. Many of them are heat-resistant, in that they will withstand a temperature of 60° C. for half an hour. In pure culture they resist a concentration of 1 per cent. phenol for 15 minutes, but are killed by a concentration of 2 per cent. Mercuric chloride is a poor disinfectant for staphylococci; to kill them in 10 minutes a 1 per cent. solution is required. Many of the aniline dyes exert a strongly bactericidal action on the staphylococci—as indeed they do on most Gram-positive organisms. This selective action is made use of in certain technical procedures, such as the isolation of *Br. abortus* from milk, where it is endeavoured by the incorporation of a dye—gentian violet or crystal violet—to inhibit the growth of Gram positive organisms. Use is also made of the great susceptibility of staphylococci to the violet dyes in the isolation of streptococci, whose susceptibility to these dyes is very much less, Garrod (1942) (see p. 135). Other dyes, of which malachite green appears to be the strongest and acid fuchsin the weakest, are also employed, usually in a concentration of about 1/10,000 (Oesterlin 1925). Most strains of staphylococci are sensitive to penicillin.

**Biochemical Reactions.**—The ability of the staphylococci to ferment sugars varies greatly according to the strain employed. For this reason it is not possible to classify them on this basis with anything like the same precision as, for example, the coliform group of bacilli. As a rule the golden cocci have the greatest fermentative power, the white are less active. There is a wealth of literature on the fermentative capacities of the staphylococci, with a corresponding difference of opinion amongst the various authors as to the importance of the different sugars. Thus, Andrewes and Gordon (1905-6) lay stress on the reactions in maltose, lactose, glycerol and mannitol. Winslow, Rothberg and Parsons (1920), on the other hand, come to the conclusion that the only sugar of differential value is lactose. Working with *Staphylococcus aureus* and *albus*, these authors found that 68 per cent. of the strains formed acid from glucose, 63 per cent. from maltose, 61 per cent. from sucrose, and 49 per cent. from lactose; salicin, inulin and raffinose were rarely fermented, mannitol and dulcitol never. With these findings most authors disagree, particularly with regard to mannitol, which is generally held to be attacked by *Staphylococcus aureus*, and frequently by *Staphylococcus albus* (Dudgeon and Simpson 1923). It is quite clear, however, that it is impossible to dogmatize on the reactions of any one strain. Dudgeon (1908), who examined 121 *aureus* and *albus* strains on a large number of sugars, found that very few agreed in giving identical results.

Similarly with *lactus milk* the reactions are variable. Studying 150 *aureus* and *albus* strains, Winslow, Rothberg and Parsons (1920) found that 75 produced acid, clot and peptonization, 60 acid, generally clot, and no peptonization, 23 alkali and peptonization, 16 alkali but no peptonization, while 7 produced no change.

These findings are in agreement with those of other authors, except with regard to peptonization, which is less commonly reported.

The *proteolytic* activity of staphylococci is not very strong. Some strains are fibrinolytic (Sasaki and Fejgin 1937, Neter 1937) and some, particularly those of canine origin, can digest coagulated horse serum (Minett 1936). *Lipase* production has been reported by Orcutt and Howe (1922) and Christie and Graydon (1940). Most *aureus* strains produce hyaluronidase.

The *methyl red* test is generally positive with the *aureus* and *albus* strains, negative with the *citreus* strains. The *Indole Proskauer* reaction is given by most strains of *Staphylococcus aureus*. According to Winslow, Rothberg and Parsons (1920), most strains of staphylococci reduce nitrates to nitrites. Hucker (1924a) on the contrary, found that, though 49 out of 50 *aureus* strains reduced nitrates, only 23 out of 152 *albus* strains were able to do so. *Hydrogen sulphide* is stated by Andrewes and Gordon (1905-6) to be formed in small quantity by the pyogenic staphylococci, in greater quantity by *Staphylococcus albus*. We have been unable to confirm this. *Ammonia* is produced by 89 per cent of the golden and white strains (Winslow *et al.* 1920). *Indole* is apparently never produced (Hucker 1924a).

**Antigenic Structure**—Studying agglutination of staphylococci by immune rabbits' sera, the early workers (Kolle and Otto 1902, Otto 1903, Proschner 1903, Kutscher and Konrich 1904, Veiel 1904, Klopstock and Bockenheimer 1904, and Koch 1908) reported that they could be sharply divided into two types—the pathogenic and the saprophytic types. The great majority of strains isolated from purulent lesions in the human body were agglutinated by a serum prepared against one such strain, whereas the strains isolated from saprophytic sources were not agglutinated. The difference in titre to which agglutination occurred rather suggested that there might be one or more sub groups within the main types. This suggestion was confirmed by later workers who employed the more delicate test of absorption of agglutinins. Thus Julianelle (1922) found that the staphylococci could be divided into three types with two sub groups, whereas Hine (1923), working with 81 strains, was able to classify them into two main types, each of which had at least three sub groups. There was evidence to suggest that the pathogenic strains formed a more homogeneous serological group than the saprophytic.

More recently Yonemura (1936) examined 324 strains from pathological sources and was able by the use of absorbed sera to divide them into nine main types: all but 22 of the strains, however, fell into three of these types. Blair and Hallman (1936) recognized three types by the absorption of agglutinins method. Cowan (1938, 1939a) likewise showed that it was possible by slide agglutination, using absorbed sera, to distinguish three main types among the pathogenic staphylococci; there is reason to believe, however, that each of these contains sub types distinguishable by means of absorbed sera (Christie and Keogh 1940). Working in India, Goyle and Minchin (1940) were able to classify only 33 per cent of pathogenic strains by Cowan's method. Considerably more work will have to be done before the serological typing of staphylococci becomes of much practical use in epidemiological investigations.

Julianelle and Wieghard (1934, 1935) and Wieghard and Julianelle (1935) as the result of *chemical fractionation* of staphylococci have isolated two polysaccharides each containing about 4 per cent nitrogen and distinguished from each other by optical rotation and by the type of sugar produced on hydrolysis. The first polysaccharide was extracted from pathogenic (A), the second from non pathogenic

(B) strains. The sera of rabbits that had been inoculated intravenously with whole cocci reacted specifically to a high titre with the polysaccharides when tested by the precipitation reaction. The polysaccharides were non antigenic and non toxic for rabbits and mice but Type A gave rise on intradermal inoculation of patients with staphylococcal infections to an immediate skin reaction of the wheal and erythema type. Besides the carbohydrates a complex protein substance was isolated from both pathogenic and non pathogenic strains of staphylococci. This protein though non toxic to rabbits and mice was antigenic giving rise to precipitins that reacted however only with the protein and not with the polysaccharide substances. On intradermal inoculation into susceptible patients the protein evoked a skin reaction of the delayed inflammatory type. Serological observations showed that the protein was responsible for the species-specificity of staphylococci while the type-specificity was determined by the soluble specific carbohydrate substances. For the differentiation of pathogenic from non pathogenic strains it was essential to employ the precipitation reaction, using as an antigen either the extracted polysaccharide the supernatant fluid of centrifuged young broth cultures or an acid extract of the sedimented organisms. The agglutination reaction was affected by the group protein and was unfitted for type differentiation. This work has been confirmed by Hegemann (1937) and Peragallo (1937) but Thompson and Korazo (1937) have brought evidence of the existence of at least one further polysaccharide which they term Type C, and Verwey (1940) claims to have isolated a type-specific protein from staphylococci. Durfee (1942) found that all strains agglutinating with Cowan's sera formed Juhanelle's polysaccharide Type A.

In general, it may be said that the precipitin reaction is more suited for the differentiation of pathogenic from non pathogenic strains and the agglutination reaction for the sub-division of the pathogenic strains into types.

Staphylococci are known to be frequent carriers of bacteriophage. Fik (1942) took advantage of this to develop a cross-culture method of bacteriophage typing. By this means he was able to classify 44 strains of *Staph aureus* into 3 different groups. Our own observations have shown that if potent lytic filtrates are prepared, staphylococci may be typed by a method similar to that used for the Vi phage-typing of typhoid bacilli. Already 21 different types have been established, and the method has proved of considerable value in epidemiological inquiries (Wilson and Atkinson 1945).

**Toxin Production.**—When grown under suitable conditions certain strains of staphylococci give rise to a filtrable toxin having a series of effects which though described a long time ago by such workers as van der Velde (1894) Denys and Havet (1898) von Lingelheim (1899) Kraus and Clairmont (1900) and Neisser and Wechsberg (1901) have received intensive study during recent years. This activity followed largely on the reinvestigation of the problem by Parker in 1924. A toxic filtrate is hæmolytic especially towards rabbit cells—it has a destructive action on leucocytes when injected intradermally into the skin of the rabbit or the guinea pig it gives rise to necrosis and when injected intravenously into the rabbit or mouse it causes acute and fatal toxæmia.

Toxin formation is a property of pathogenic strains and is therefore limited mainly to the *aureus* type. Considerable variation exists between different strains and if toxin is required on a large scale for immunological or other purposes it is important to select a strain with a high toxigenic capacity. Various methods are

used in the production of the toxin. In a fluid medium it develops rather slowly. Thus Neisser and Wechsberg (1901), by testing broth cultures filtered at intervals, found that the toxin, as judged by the hæmolytic titre, was first demonstrable on the 4th day of incubation at 37° C, and that it rose to a maximum between the 10th and 14th days, after which it gradually diminished. Burnet's (1930) technique of growing the organisms on 0.8 per cent nutrient agar for 24 hours in air containing 10-20 per cent CO<sub>2</sub>, and extracting the toxin from the agar with saline, is very satisfactory, and is widely used, either in its original or a slightly modified form (Parish and Clark 1932, Dolman 1932). The medium used should have a reaction of between pH 6.0 and 7.0 (Walbum 1922). Though the toxin can traverse a Seitz filter, some of its activity is lost in passage, and it is therefore advisable to separate the organisms from the toxin by centrifugation. A good toxin should hæmolyse a 1 per cent suspension of rabbit cells in a dose of 0.0005-0.002 ml, 0.001-0.005 ml should cause necrosis on intradermal inoculation, and 0.25-0.8 ml per kilo injected intravenously should kill a rabbit within a few minutes (Burnet 1929, Gross 1931c, Parish and Clark 1932). Satisfactory toxins can be prepared by growth in a chemically defined medium (Gladstone 1933, Smith and Price 1933a).

Though Burnet (1929), Gross (1931c) and Gengou (1932) formed the opinion that the various activities of a toxic filtrate were due to one and the same toxin, subsequent workers have inclined to the opposite view. Without prejudicing the issue, we shall, for the sake of simplicity, describe each of the manifestations separately.

**$\alpha$ -Hæmolsin**—This is active against rabbit, but not against human, red corpuscles and causes lysis rapidly at 37° C. It has some action on sheep red corpuscles, but this is destroyed by heating at 57° C for 30 minutes (Flaum 1933). It can be specifically neutralized in multiple proportions by an antiserum. Its resistance to heat is peculiar since it is inactivated more readily by lower than by higher temperatures (Arrhenius 1907, Landsteiner and von Rauchenbichler 1909, Seiffert 1935-36, Kodama and Nisiyama 1938, Rigdon 1933a, Beumer 1930, Fulton 1943). According to Arrhenius (1907) lysin heated to 70° C loses a great part of its hæmolytic activity, but regains it if it is heated for a further 5 minutes to 100° C. This fundamental observation was confirmed by Landsteiner and von Rauchenbichler (1909) using a temperature of 65° C for 30 minutes to inactivate the lysin. As the result of experimental observations they were led to believe that at the lower temperature the lysin enters into combination with a protein constituent, derived from the broken down organisms, the medium, or added serum, to form an inactive compound that can be destroyed by heating to a higher temperature, thus liberating the lysin. The failure to realize this unusual reaction to heat has led to much confusion by workers who have assumed that because the lysin is apparently destroyed by heating for 2 hours at 57° C, 1 hour at 60° C, or 30 minutes at 65° C, it is necessarily destroyed by boiling. According to Burnet (1931) Kodama and Nisiyama (1938), and Fulton (1943), a strong toxin is not destroyed completely even in 30 minutes at 100° C. Its activity is said to be inhibited by ascorbic acid in a concentration of 30 mgm per ml (Mercier 1938), and by azochloramid (Heise and Starin 1940). There is a close parallelism between the  $\alpha$  hæmolsin content of a filtrate and (1) the dermonecrotic and lethal factors (see Pantou and Valentine 1932, Levine 1939), (2) the ability of a given strain to reduce methylene blue (McBroom 1937), and (3) the leucocidin content as estimated by the Neisser Wechsberg method (Wright 1936), though this is still subject to dispute (see Flaum 1933). Morgan and Graydon (1936) have brought evidence to show that in most toxic filtrates the  $\alpha$  lysin contains two antigenically distinct components referred to as  $\alpha_1$  and  $\alpha_2$ , which have different combining powers for antitoxin, both are dermonecrotic. Treatment of the toxin with 0.2-0.5 per cent formol at 37° C leads

in the course of a few days to a disappearance of its hemolytic and toxic properties, but not to that of its flocculating capacity with antitoxin (see Burnet 1931). Toxoid so prepared is antigenic and is sometimes used as a vaccine. Strains producing  $\alpha$  lysin are predominantly of human origin.

**$\beta$ -lysin.**—This acts on sheep, ox and human, but not on rabbit, red corpuscles, and causes lysis only after the tubes have stood at room temperature or in the ice-chest over night—the so-called "hot-cold" lysis (Bogger 1933, Glenn and Stevens 1935). It is more resistant to formalin than the  $\alpha$  lysin, and is usually stated to be more resistant also to heat, the degree of destruction at 57° C. in half-an-hour being much less. According, however, to Kodama and Kodama (1939) it resembles  $\alpha$  lysin in being inactivated more readily at low than at high temperatures. It is antiserally distinct from the  $\alpha$  lysin, and can be specifically neutralized by a suitable antiserum. It is much less toxic than the  $\alpha$  lysin to rabbits, guinea-pigs and mice (Bryce and Rountree 1935). Intradermal inoculation into guinea-pigs gives rise to no more than a transient erythema (Smith and Price 1935a). Strains producing  $\beta$ -lysin are predominantly of bovine origin (see Minett 1936, Slanetz 1942).

**$\gamma$ -lysin.**—Smith and Price (1935b) have described the production by a strain of staphylococcus of a  $\gamma$ -toxin, which causes rapid lysis of the red corpuscles of a variety of animals including the rabbit and sheep, and delayed lysis of rat and guinea-pig corpuscles. It appears to be antigenically distinct from the  $\alpha$  and  $\beta$ -toxins though it may have some relationship to the  $\alpha$ -toxin.

**Leucocidin.**—The study of the leucocidin content of toxic filtrates has been carried out by two different methods. In the Neisser Weichberg (N W) method the leucocidin is titrated by its ability to inhibit reduction of methylene blue by healthy rabbit leucocytes. The Panton Valentine (P V) method, described by Panton and Valentine in 1937 and modified slightly from that of van der Velde (1944) depends on direct microscopic observation of the destructive action of the toxin on human leucocytes. According to the observations of Valentine (1936) and Wright (1936) it would appear that the N W leucocidin is identical with the  $\alpha$  hemolysin, but that the P V leucocidin is different from it. Flaum (1935) on the other hand, observed that a practically pure  $\beta$ -lysin might have a strong leucocidal effect, as tested by the N W method, and concluded that the N W leucocidin is not always identical with the  $\alpha$  lysin. He suggests, however, that the P V leucocidin may be identical with the  $\beta$  lysin. In his experience it proved to be more heat stable than the leucocidin associated with the  $\alpha$  lysin. Proom (1937) considers that the  $\alpha$  lysin is able to interfere with the respiratory activity of the leucocytes, as measured by the N W technique, but that it has not the same destructive action on the cell and nucleus as is manifested by the P V leucocidin. Weld and Mitchell (1942) state that the  $\alpha$  lysin and the N W leucocidin agglutinate rabbit leucocytes, and that the agglutination reaction can be used instead of methylene blue reduction for measuring the N W leucocidin.

**Enterotoxin.**—Some strains of staphylococci, mostly of the *aureus* species, produce under favourable conditions an enterotoxin that is capable of giving rise to acute food poisoning in man (see Chapter 7). Our information on the properties of this substance is confusing and incomplete. There is no satisfactory method of titrating it, and there is some doubt about its heat stability. It is prepared best by growth in a semi-solid agar medium, such as that described by Dolman and Wilson (1938, 1940). The cultures should be incubated for about 40 hours in an atmosphere containing 10–30 per cent. CO<sub>2</sub>, and should then be passed through cheese cloth and fine filter paper and centrifuged at high speed. The clear supernatant fluid, which contains the enterotoxin, should be sterilized by gradocol filtration. Tests for its presence and its potency were originally made by feeding filtrates to human volunteers or to monkeys, but Dolman, Wilson and Cockcroft (1936) pointed out that, provided the  $\alpha$  and  $\beta$  toxins were destroyed by heat or formalin, or were neutralized by antiserum, the enterotoxin could be demonstrated by its ability to give rise to vomiting and diarrhoea in kittens injected intraperitoneally.

Inoculation of 1-3 ml of a potent filtrate into a kitten weighing 350-700 gm is followed as a rule within half-an-hour by lassitude and weakness, unsteadiness of gait and paroxysmal vomiting of the projectile type associated with diarrhoea. The kitten may be ill for several hours but usually recovers completely.

How far the kitten test can be regarded as a specific reaction to the enterotoxin is doubtful. Several workers (Rigdon 1938b, Jones and Lochhead 1939, Hammon 1941) have reported non-specific reactions after the intraperitoneal injection of control materials. Moreover, Fulton (1943) working in our laboratory found little relationship between the ability of a filtrate to give rise to vomiting in kittens on intraperitoneal injection and to vomiting in man when taken by the mouth. Other methods of demonstrating the enterotoxin have been suggested such as the intravenous inoculation of rabbits (Kupchik 1937) of kittens (Davison, Dack and Cary 1938, Davison and Dack 1939) or of monkeys (Segalove and Dack 1941) or the feeding of kittens (Dolman, Wilson and Cockcroft 1936) but none has so far proved as reliable as the feeding of human volunteers.

The nature of the enterotoxin is still in doubt. Dolman (1943) regards it as distinct from the  $\alpha$  and the  $\beta$  toxins. He finds that a pure  $\beta$  toxin when injected intraperitoneally into the kitten gives rise to early and repeated vomiting coming on sometimes within 5 minutes and followed by death some hours later. Vomiting caused by the enterotoxin on the other hand begins later and is not fatal. Though Slanetz (1942) suggests that the  $\beta$  toxin and the enterotoxin are identical this seems improbable because a pure  $\beta$  toxin is innocuous to man by the mouth (Fulton 1943, Dolman 1943). For the separation of the enterotoxin from the  $\alpha$  toxin, reliance has been placed on the difference in their heat stability. The  $\alpha$  toxin is said to be destroyed by heating at 65° C in 30 minutes whereas the enterotoxin is said to withstand boiling for 15-30 minutes or longer. As Fulton (1943) however, points out, this is not a reliable method of differentiation because the  $\alpha$  toxin though inactivated by heating at 65° C. is moderately resistant to boiling (see p. 615). It is at present very difficult to say whether the enterotoxin is distinct from the other toxins or whether it is related to the  $\alpha$  toxin but the observation by Dolman (1944) of a strain that produced enterotoxin in the complete absence of  $\alpha$  or  $\beta$  toxin suggests that the enterotoxin is distinct from the hæmolytic toxins. The mode of action of the enterotoxin is not yet understood. The observations of Bayliss (1940) on the cat and of Richmond, Reed, Shaughnessy and Michael (1942) on the rabbit suggest that its action is peripheral rather than central and that it affects either the sensory nerve endings or the smooth muscle of the small intestine.

*Staphylocoagulase*—The ability of certain staphylococci to coagulate citrated or oxalated plasma was first described by Loeb (1903-04) and confirmed by Much (1908). Later it was studied by a number of other workers (von Daranyi 1926, Gross 1931a, b, Stephan 1931, Vanbreuseghem 1931, Chapman *et al.* 1934, Walston 1935).

For its demonstration about 0.1 ml of an overnight broth culture of *Staph. aureus* or of a broth suspension of an agar slope culture made up to the same density as a broth culture is mixed with 0.5-1.0 ml of a freshly prepared 1/10 dilution of human or rabbit plasma in saline. The mixture is incubated at 37° C for 3-6 hours. If no clot has appeared by this time it should be left overnight at room temperature and again examined. The plasma of other animals such as the horse, ox, sheep or guinea pig may be used instead. If kept undiluted in the ice chest under sterile conditions citrated plasma remains suitable for several months (Fisk 1940). Since plasma may undergo spontaneous coagulation a control tube containing plasma alone diluted with saline should always be put up, as well as tubes inoculated with a known coagulase positive and a known coagulase negative strain. Culture media containing fermentable carbohydrates should be avoided, since Neter (1937) has shown that under these conditions many strains form an anti coagulase

which may inhibit the clotting of the plasma. Other sources of error have been pointed out by Gillespie (1943). According to Duncan and Walker (1942) the result of the test is influenced by the proportion of culture to plasma in the mixture. The optimal proportion can be determined in the usual way by titrating a series of increasing dilutions of culture against a fixed volume of plasma, or of increasing dilutions of plasma against a fixed volume of culture. Tests for the presence of coagulase may also be made by the slide method (Cadness Graves *et al.* 1943).

Though there is a high correlation between coagulase and  $\alpha$  haemolysin production there seems little doubt that these two substances are distinct. The haemolysin is destroyed in half an hour by exposure to a temperature of 56° C, whereas the coagulase is not (Vanbreuseghem 1934, Cruickshank 1937, Smith and Hale 1944). The haemolysin is absorbed by red corpuscles, the coagulase is not. An antibody to the haemolysin can readily be obtained by the inoculation of rabbits but not to the coagulase (Gross 1931a). Coagulase is formed almost exclusively by strains of *Staph aureus* or by *albus* variants of *aureus* strains. Cruickshank (1937) maintains that coagulase production constitutes the most convenient and reliable single test for estimating the pathogenicity of a given strain. This contention is supported by a number of subsequent workers (Chapman *et al.* 1938, Marcuccio 1938, Gillespie, Devenish and Cowan 1939, Fairbrother 1940, Christie and Keogh 1940).

The method of coagulase action is not yet clearly understood. According to Smith and Hale (1944) coagulase itself is a thermostable substance, filtrable through a gradocol membrane having an A.P.D. of 0.31  $\mu$  but completely held back by one of 11  $\mu$ . It appears to be the precursor of a thermolabile thrombin like substance the production of which depends on the participation of an activator present in the plasma of some animals but deficient or lacking in others. The staphylocoagulase reaction resembles normal thrombin formation from prothrombin under the influence of thrombokinase except that calcium is not required.

**Pathogenicity.**—The staphylococci can be fairly sharply divided into pathogenic and non pathogenic types. Thus the great majority of strains isolated from suppurative lesions in the animal body are found to be pathogenic for rabbits, and to a less extent for mice and guinea pigs. On the other hand, the great majority of strains isolated from normal skin, air, water, dust, etc. are harmless to these animals. Sometimes the virulence of the pathogenic strains diminishes on prolonged cultivation but this is not always so, even after years of sub culture in the laboratory the virulence may remain intact. Moreover, by passage through rabbits it is generally possible to raise the virulence of a strain which has become temporarily avirulent, with the saprophytic strains this is impossible. According to Lubinski (1894) the virulence of *Staph aureus* for rabbits can be increased by growth under anaerobic conditions, growth in pure oxygen was said to have the reverse effect.

**Man.**—The staphylococci which are responsible for disease in the human body generally belong to the *aureus* or *albus* varieties, only occasionally can *Staphylococcus citreus* be incriminated. *Staphylococcus aureus* is more pathogenic to man than *Staphylococcus albus*, it gives rise to the severer lesions such as osteomyelitis, pyaemia—sometimes associated with an infective endocarditis—mastitis, boils and abscesses in various parts of the body, and on occasion to a peculiarly fatal form of broncho pneumonia (Finland, Peterson and Strauss 1942) whereas *Staphylococcus*

*albus* is responsible for the milder inflammatory lesions such as acne pustules, stye abscesses and other minor suppurative conditions of the skin. Staphylococci are frequently found in conjunction with other organisms particularly in the chronic stages of gonorrhœa, in bronchitis in post influenza pneumonia and in catarrhal conditions of the nose and respiratory passages. Their exact significance in these cases is difficult to assess but it is probable that they assist these other organisms in giving rise to suppuration.

Several observers have made personal experiments on themselves to test the pathogenicity of the staphylococci in pure culture. Thus Garre (see Neisser 1912) found that by rubbing staphylococci into the skin of his arm he was able to produce boils which took a considerable time to heal. (For further information on Pathogenicity to man, see Chapter 67.)

**Experimental Inoculation**—The numerous experiments which have been conducted on the pathogenicity of the staphylococci suffice to show that the only laboratory animals that can be artificially infected with ease are the rabbit, the mouse, and the guinea pig. Of these undoubtedly the most susceptible is the rabbit.

Not all strains are equally pathogenic. As a rule the cocci which are isolated directly from suppurative processes in the body prove virulent, whereas those isolated from skin, air, water, etc. are avirulent. The most pathogenic are the golden strains; many of the white strains are pathogenic though to a less degree, the yellow cocci are generally non pathogenic.

**RABBITS**—The subcutaneous injection of 1 ml. of a 24 hours broth culture of *Staph aureus* gives rise to a local abscess from which the organisms can be recovered. If the culture is mixed with an equal amount of 2 per cent melted agar and inoculated intracutaneously into the rabbit's back a spreading necrotic lesion occurs with little pus formation (Jackson, Nicholson and Holman 1940).

Intravenous injection of 0.1 to 0.5 ml. of a strain of *Staph aureus* recently isolated from a suppurative focus generally proves fatal in 24 to 48 hours. Post mortem there are hæmorrhages and bloody exudations on the serous membranes and parenchymatous degeneration of the glandular organs; the cocci can be recovered from the blood stream.

Intravenous injection of a smaller dose—about 0.01 to 0.05 ml.—gives rise to a pyæmic condition accompanied by loss of weight and general weakness and proving fatal in 1 to 6 weeks. Post mortem multiple small or large circumscribed abscesses are found particularly in the kidneys and less frequently in the myocardium, lungs, spleen, bone marrow and costal cartilages. Sometimes vegetations develop on the mitral and tricuspid valves and the chordæ tendinæ without any artificial wounding of these structures. Acute osteomyelitis of the long bones not infrequently develops. The cocci can be recovered from the suppurative lesions. In animals that recover reparative processes occur.

*Staphylococcus albus* is usually much less pathogenic. Strains recently isolated from suppurative foci may cause death on intravenous injection of 1–2 ml. of a 24 hours broth culture. Strains isolated from saprophytic sources are non pathogenic unless given in large doses when death occurs apparently from toxæmia.

Staphylococci which have been killed by heat at 60° C. for 2 hours if given in large doses—2–4 agar slopes—at repeated intervals of 10 days may give rise to progressive cachexia with death in 2 or 3 weeks. Post mortem much the same changes are found as those following acute death from a living culture, namely a hæmorrhagic exudate in the peritoneal cavity, serous hæmorrhages, and parenchymatous degeneration of the glandular organs (Koch 1908).



MICE AND GUINEA PIGS are much less susceptible than rabbits, and though death may follow the intraperitoneal injection of virulent cultures, there is frequently no more than a local abscess formation from which the animal recovers.

**Protection Experiments.**—Procher (1903) who attempted to prepare an immune serum suitable for prophylactic and therapeutic use obtained some hopeful results by the injection of living virulent staphylococci into goats and horses. 15 ml. of immune goat serum given subcutaneously protected rabbits against 0.5 ml. of a virulent broth culture injected intravenously 24 hours later. But if given at the same time as or subsequently to the injection of the cocci it had but little or no effect. In spite of numerous studies by such workers as Forssman (1935-41) Blair and Hallmann (1935) Ki chung and Farrell (1936) Lyons (1937) Downie (1937) Smith (1937) Cowan (1939b) and Valentine and Butler (1939) the mechanism of immunity in natural and experimental infections of man and animals with staphylococci still remains obscure. Serum prepared by the injection of rabbits or horses with toxoid and later with toxin, has a high antitoxic titre, and may save the life of test animals inoculated prophylactically but it seems to have little or no direct bactericidal effect so that local abscess formation is not necessarily prevented. An international standard for staphylococcal antitoxin has now been laid down: one unit is contained in 0.2316 mgm. of the standard preparation (Smith and Ipsen 1937). The potency of the serum is treated by the hæmolytic method using rabbit red corpuscles, by the intracutaneous injection of guinea pigs, and by the intravenous injection of mice.

#### CLASSIFICATION

Little useful purpose would be served by describing the numerous attempts that have been made to provide a satisfactory classification for the staphylococci. Those who are interested should consult the studies of Posenbach (1884) Winslow and Rogers (1906) Dudgeon (1908) Ehrler (1913) Winslow Rothberg and Parsons (1920) Hucker (1924a) and Dudgeon and Simpson (1925). Nearly all workers are agreed that there is a gradation from the actively fermentative, gelatin liquefying pathogenic group of which the type is *Staph. aureus* down to the weakly fermentative, gelatin non liquefying saprophytic group of which Gordon's scurf staphylococcus is a typical representative. No sharp line or cleavage occurs, and no single property can be regarded as satisfactory as a basis for classification. Though there is something to be said for dividing the staphylococci into a pathogenic group *Staph. pyogenes* (Cowan 1937) and a non pathogenic group *Staph. saprophyticus* (Fairbrother 1940) on the basis of coagulase production, we doubt whether there is much to be gained, partly because the production of golden pigment and coagulase are fairly highly correlated, partly because coagulase production has not yet been established as a certain indicator of potential pathogenicity and partly because pathogenicity is not a good criterion on which to establish specific differences. It is true that we recognize the name *S. repococcus pyogenes* but here the differentiation is based on the firmer ground of antigenic structure. Until the serological study of the staphylococci has progressed further we think it best to keep to the time-honoured names of *Staph. aureus*, *Staph. albus* and *Staph. citreus* and taking account of more recent knowledge to define them tentatively as follows:

*Staphylococcus aureus* is a pathogenic species producing suppurative lesions of

varying severity in man and animals; usually it forms a golden-yellow pigment, but *albus* variants may be thrown off either in the body or *in vitro*. It produces toxic filtrates with hæmolytic, leucocytolytic, necrotizing, and lethal properties for the rabbit, and sometimes with irritating properties for the gastro-intestinal tract of man, it almost always liquefies gelatin, ferments lactose and mannitol, and coagulates plasma, it contains a specific polysaccharide not possessed by non-pathogenic staphylococci, its antigenic structure differs from that of the following species

*Staphylococcus albus* is feebly pathogenic, or non pathogenic, it is normally present on the skin, in the hair, and apparently in air, water and dust. It gives porcelain-white or indifferently coloured colonies, but never produces a yellow or golden pigment. It forms no toxin, or does so less frequently than *Staph aureus*, though white variants of *Staph aureus* are often highly toxigenic. It frequently liquefies gelatin, but less constantly than *Staph aureus*. It often ferments lactose, but not mannitol, it forms no coagulase, and it appears to contain, as a rule, a polysaccharide different from that in virulent *Staph aureus* strains.

The third species, *Staphylococcus citreus*, is a non pathogenic saprophyte. It produces a distinctive lemon yellow pigment, it is doubtful whether it ever forms a toxin; it liquefies gelatin less frequently and less rapidly than the preceding species. It has little or no fermentative ability, and it forms no coagulase.

We append, for purposes of reference, a detailed description of *Staph aureus*, together with some of the characters ascribed to those types of staphylococci which have, at various times, received specific names.

### *Staphylococcus aureus* Rosenbach

*Isolation*—First described fully by Rosenbach (1884)

*Habitat*—Actual or potential parasite found in suppurative lesions of man, in the nose on the normal skin, and in cow's milk.

*Morphology*—Spherical cells, 0.8–1.0  $\mu$  in diameter, in cultures on solid media the cocci are arranged in grape like clusters, in broth they occur as small groups, pairs and short chains of not more than four members. Stain well with the usual aniline dyes. Non motile, Gram positive, non acid fast.

*Agar Plate*—24 hours, 37° C. Circular colonies, 1–2 mm in diameter, low convex, amorphous, opaque, and of a golden colour, having a smooth glistening surface and an entire edge, butyrous in consistency and easily emulsifiable. No differentiation visible.

*Agar Stroke*—24 hours, 37° C. Abundant, confluent, raised, golden yellow, opaque growth, with a glistening smooth or slightly contoured surface, and an entire or slightly undulate edge.

*Gelatin Stab*—5 days, 22° C. Abundant filiform growth reaching to bottom of stab surface growth about 5 mm in diameter, liquefaction of infundibuliform or saccate type.

*Broth*—24 hours, 37° C. Moderate uniform turbidity with a moderate, powdery deposit, disintegrating readily on shaking, slight ring growth at surface.

*Rabbit Blood Agar Plate*—48 hours, 37° C. Good growth. Blood is partly or completely hæmolyzed around colonies.

*MacConley's Agar*—24 hours, 37° C. Tiny, convex, pinkish colonies about 0.5 mm in diameter. Later they increase somewhat in size, and take on a deep red colour.

MICE AND GUINEA PIGS are much less susceptible than rabbits and though death may follow the intraperitoneal injection of virulent cultures there is frequently no more than a local abscess formation from which the animal recovers.

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*Gelatin Slab*—5 days, 22° C. Abundant filiform growth reaching to bottom of slab. Surface growth about 5 mm in diameter, liquefaction of infundibuliform or saccate type.

*Broth*—24 hours, 37° C. Moderate uniform turbidity with a moderate, powdery deposit, disintegrating readily on shaking, slight ring growth at surface.

*Rabbit Blood Agar Plate*—48 hours, 37° C. Good growth. Blood is partly or completely hæmolyzed around colonies.

*MacConkey's Agar*—24 hours, 37° C. Tiny, convex, pinkish colonies about 0.5 mm in diameter. Later they increase somewhat in size, and take on a deep red colour.

*Potato*.—24 hours, 37° C. Poor, yellowish, effuse growth. 5 days, moderate, confluent, slightly raised, golden yellow growth.

*Loeffler's Serum*.—24 hours, 37° C. Good, raised, confluent, golden yellow growth.

*Resistance*.—May withstand moist heat at 60° C. for 30 minutes, generally killed in one hour. Destroyed by 2 per cent. phenol in 15 minutes.

*Metabolism*.—Aerobic, facultatively anaerobic. Growth occurs best at 37° C., limits 12° to 45° C. Optimum pH for growth is 7.4-7.6. Pigment formed most readily at 22° C., is not formed in cultures grown anaerobically. Filtrable hemolysin produced,  $\alpha$ -lysin acts rapidly on rabbit and sheep red corpuscles;  $\beta$ -lysin acts slowly on sheep corpuscles—hot-cold lysin. Coagulase produced which clots citrated human or rabbit plasma.

*Biochemical Reactions*.—Ind, no gas, in glucose, maltose, mannitol, lactose, and sucrose. L.M. acid, clot, and sometimes peptonization. Indole—. M.P.—. V.P.—. Nitrates reduced to nitrites. M.B. reduction—. H<sub>2</sub>S—. NH<sub>3</sub>—.

*Antigenic Structure*.—Contains a polysaccharide A, demonstrable by precipitation. Divisible by agglutination with absorbed sera into three main types and a number of sub-types.

*Pathogenicity*.—Forms exotoxin with hemolytic, leucocytolytic, skin necrosing, and lethal properties for the rabbit. Some strains also form an enterotoxin acting on man. *Staph. aureus* is frequently responsible for suppurative lesions in the human body, such as boils and abscesses, acute osteomyelitis, infective endocarditis, pyæmia, etc. Experimentally, it is pathogenic for rabbits, less so for mice and guinea pigs. 0.1-0.5 ml. of a 24-hours' broth culture injected intravenously into a rabbit is generally fatal in 24 to 48 hours; post mortem, hemorrhages on the serous membranes, the coxi can be recovered from the blood stream. A smaller dose may not prove fatal for 1 to 6 weeks; post mortem, multiple abscesses are frequently seen, especially in the kidneys, less frequently in the myocardium, lungs, spleen, bone marrow, and costal cartilages.

*Staphylococcus albus*.—Resembles *Staph. aureus* in many respects. For chief differences, see p 621.

*Staphylococcus citreus*.—Resembles *Staph. aureus* in many respects. For chief differences, see p 621.

TABLE 29

	Parent.	Glucose.	Maltose.	Mannitol.	Lactose.	Sucrose.	Gel. Dp.	Path. to Man.
<i>Staph. aureus</i> Rosenbach	Golden	A	A	A	A	A	—	High
<i>Staph. aureus</i> Schroeter	Golden	A	A	—	—	A	—	High
<i>Staph. epidermidis</i> Gordon	White	A	A	—	A	A	+	Feeble
<i>Staph. candidus</i> Cohn	White	A	A	+	A	A	—	Feeble
<sup>1</sup> <i>Staph. schweitzeri</i> Gordon	White	A	A	—	—	A	—	—
<i>Staph. candidus</i> Flagg	White	A	A	+	—	A	—	Feeble
Scarl. <i>staphylococcus</i> Gordon	White	A	—	A	—	—	—	—
<i>Staph. citreus</i>	Lemon yellow	—	—	—	—	—	—	—

<sup>1</sup> Viscid growth in broth.

## MICROCOCOCCUS, SARCINA, RHODOCOCOCCUS AND LEUCONOSTOC

In their monograph on the *Systematic Relationships of the Coccaceæ*, C E A Winslow and Anne R Winslow (1908) emphasize that the white and golden cocci belong to a series of cocci, including the streptococci which are essentially parasitic in nature and active in fermentative power, whereas the yellow and red forms, including the sarcinæ, are generally saprophytic in nature and possess a restricted fermentative power

## Micrococcus

DEFINITION—*Micrococcus*

Spherical or ovoid cells, non motile, arranged in pairs tetrads or groups, but not in grape like clusters or chains Generally Gram positive Grow freely on ordinary media. Sometimes produce a yellowish pigment Gelatin liquefaction is not constant, and is usually slow Fermentative activities weak. Usually non pathogenic to man or animals

Type species is *Micrococcus luteus* Cohn.

The term *Micrococcus* was first used in a generic sense by Cohn (1875), who applied it to small spherical or oval non motile organisms which occurred in chains or groups This, it will be observed, was a comprehensive description it was not long in fact before certain members, such as the streptococci and the staphylococci, were removed from it, and awarded generic names of their own The term *Micrococcus*, as now used, is defined above

As with most saprophytic organisms, the micrococci have not been studied so fully as the pathogenic cocci—the streptococci and the staphylococci It must therefore be made clear that, in the following account, we record the characters of certain micrococci which have been from time to time described by different workers, without committing ourselves to any judgment as to their claims to be accorded specific rank In general, we have been guided in our selection by Hucker (1924b, 1929), who uses pigment production, gelatin liquefaction nitrate reduction, and ability to use ammonium salts as the sole source of nitrogen as his main differential criteria To his reports we would refer the reader who requires more detailed information

(1) *Micrococcus luteus* (Schroeter) Cohn.—Isolated by Schroeter (1875) from a potato on which a yellow growth was found, is a non motile, Gram positive coccus 1.0-1.2  $\mu$  in diameter, occurring in pairs, tetrads or small groups. It grows well on ordinary media giving a smooth, lemon yellow layer Gelatin is not liquefied though on this point various workers disagree Nitrates are not reduced The optimum temperature of growth is 25°C Its fermentative powers are weak, acid being formed in glucose not in lactose It is commonly found in air, water, milk and milk products The pigment is insoluble in water

(2) *Micrococcus varians* (Dyar) Migula (1900) is similar to *M. luteus*, but is differentiated from it by its ability to reduce nitrates

(3) *Micrococcus conglomeratus* Flugge (Migula 1900) Forms large clumps of organisms, often in pairs Generally Gram positive Liquefies gelatin and reduces nitrates Forms an abundant light yellow growth on agar Can utilize ammonium salts as the sole source of nitrogen

(4) *Micrococcus flavus*—Isolated by Prove (1884-87) from human urine About 0.8  $\mu$  in diameter, occurring singly, in pairs or short chains On gelatin gives colonies 5 mm in diameter by the 3rd day Liquefies gelatin about the 12th day Does not reduce nitrates. Forms a considerable amount of slimy matter Is said to ferment certain

carbohydrates. Grows between 6°C and 36°C optimum temperature 22°C. Is distinguished by the fact that it forms a yellow pigment only when it is exposed to light—sunlight or diffused daylight when kept in the dark the growth is colourless.

(5) *Micrococcus coronatus* Flugge.—Described by Flugge (1896). A coccus 1  $\mu$  in diameter occurring in groups or short chains, Gram positive. On gelatin plates in two days it forms whitish points around these there appears a fresh growth which later becomes separated from the colony by a clear ring of liquefaction. The central part assumes a dark brown the peripheral a yellowish brown colour. It is the halo around the colony which gives the organism its name. Found in air.

(6) *Micrococcus caseolyticus* Evans (1916). This organism is similar in many respects to *M. coronatus*. It occurs in large clumps is generally Gram positive and is found in milk, milk products and in the udder of the cow. It produces a luxuriant white growth on agar slants. It liquefies gelatin and reduces nitrates. It produces an acid curd in milk and peptonizes it rapidly. It generally utilizes ammonium salts as the sole source of nitrogen and usually produces acid in glucose and lactose.

(7) *Micrococcus radiatus* Flugge.—Described by Flugge (1896). Small coccus, less than 1  $\mu$  in diameter occurring in small groups, or sometimes short chains. On gelatin plates the colonies after 2 days are about 1 mm in diameter whitish in colour with a slightly irregular edge. During the next two days outgrowths occur radiating from the centre in an orderly manner so that the colony assumes an appearance not unlike that of a starfish. Meanwhile liquefaction sets in and the centre sinks gently into the medium. The ends of the outgrowths are joined together by a ring of growth, from which after another two days fresh radiations may arise and from these yet a third set. When fully grown, the colony has a diameter of 1.0–1.5 cm. In a gelatin stab the colonies down the stab send out horizontal shoots giving the growth



FIG. 128.—*Micrococcus tetragenus*.  
From an agar culture 24 hours 3°C  
( $\times 1000$ )

a feathery appearance. On the surface there is a slow liquefaction of infundibuliform type.

(8) *Micrococcus ureæ* Cohn (1875) is a spherical organism occurring singly in pairs, or in short chains. Diameter 0.8–1.0  $\mu$ . Gram positive though often weakly so. On agar it forms whitish, low convex, opaque colonies. Gelatin is not liquefied as a rule. Nitrates not reduced. Can utilize urea and ammonium salts as sole source of nitrogen. Found in stale urine.

(9) *Micrococcus freudenreichii* is closely related to *M. ureæ*. It is a facultative parasite. Cells occur singly or in clumps. Generally Gram positive. Liquefies gelatin, but does not reduce nitrates. Does not usually produce sufficient acid to curdle milk. Can utilize ammonium salts but not urea, as the sole source of nitrogen. Is one of the commonest non-pigment producing micrococci in milk and dairy utensils.

(10) *Micrococcus tetragenus* Gaffky.—First described by Gaffky in 1883. It was isolated by Koch from lung cavities in patients with pulmonary tuberculosis. Gram-positive spherical organism, dividing in two planes at right angles to each other so that tetrad forms are produced. In the human and animal body a capsule is formed. Grows freely on ordinary media. Optimum temperature for growth is 37°C. The growth on agar is of a glutinous consistency often adherent to the medium, and difficult to emulsify.

Colonies are whitish in colour. Zone of lysis around colonies on blood agar. In broth a thick, weedy, glutinous deposit is formed, the supernatant fluid being comparatively clear. Gelatin is not liquefied. Nitrates are not reduced. Acid is said to be produced in glucose, maltose, lactose, and sucrose. The organism is occasionally pathogenic to man and may even give rise to septicæmia. One of its most notable characteristics, differentiating it from other micrococci, is its high pathogenicity for the mouse. Subcutaneous or intraperitoneal inoculation leads to the production of a septicæmia, which proves fatal, according to circumstances, in 1 to 8 days. At post mortem there may be small abscesses in the spleen. The cocci are found in large numbers in the blood and tissues. Guinea pigs are susceptible, though less so than mice. Non-capsulated avirulent and other variants have been described by Wreschner (1921) and Reimann (1936).

(11) *Micrococcus buccalis* is a small, non motile, Gram positive coccus about  $0.5 \mu$  in diameter, isolated by Ozaki (1915) from the mouth. It is peculiar in being an obligate anaerobe. In stab glucose agar it gives a filiform growth with gas formation. There is turbidity in dextrose broth and a grayish sediment. It ferments glucose, maltose, lactose, and sucrose, but not mannitol, with the production of acid and gas. There is no liquefaction of gelatin.  $H_2S$  positive, indole negative. Its optimum temperature is  $37^\circ C$ , there is no growth under  $20^\circ C$ . It is non pathogenic to laboratory animals. This coccus differs in several important points—notably its active fermentative powers and its anaerobic nature—from most other micrococci, but as no special group is available for it as yet, we choose to place it here.

Other anaerobic micrococci have been described at various times, notably the *Micrococcus gingivalis* Ozaki (1912), the *Micrococcus minimus* Gioelli (1907) and the gas producing organism *Micrococcus aerogenes* Schottmuller (1912). (For the last three references see Ozaki 1915.)

## Sarcina

### DEFINITION.—*Sarcina*

Same definition as *Micrococcus*, except that cell division occurs, under favourable conditions, in three planes, so that cubical packets are formed.

The type species is *Sarcina ventriculi* Goodsir.

The first description of the sarcinæ was by Goodsir (1842), who found an organism arranged in cubical packets in the stomach of a patient suffering from gastric disease, to this he gave the name *Sarcina ventriculi*. Schroeter (1875) was the next to describe a sarcina—the *Sarcina aurantiaca*—and since then several others have been isolated, some from suppurative processes but the majority from non pathological sources, including soil (see Smit 1933).

The following is a description of certain of the sarcinæ that have been described as separate species.

*Sarcina ventriculi*, Goodsir.—This, the first sarcina described, was not cultured till long after Goodsir (1842) had noticed it microscopically in the stomach contents.

**Morphology**—Spherical coccus,  $0.8-1.0 \mu$  in diameter, arranged in cubical packets and groups. In liquid media it occurs in pairs, small groups, and packets. Non motile. Gram positive, but decolorizes easily. Non acid fast.

**Agar Plate**.—48 hours,  $37^\circ C$ . Circular colonies, 1 mm in diameter, convex, amorphous, opaque, pale-yellow, with a smooth surface and entire edge. rather viscous in consistency, and easily emulsifiable.

**Agar Slope**.—48 hours,  $37^\circ C$ . Moderate, confluent, raised opaque creamy yellow growth with a smooth or contoured surface and an undulate edge.

**Gelatin Stab**.—Moderate filiform growth, no liquefaction.

**Broth**.—48 hours,  $37^\circ C$ . Moderate uniform turbidity with a viscous deposit, disintegrating on shaking. No surface growth.



*Metabolic and Biochemical Activities.*—Aerobic, facultatively anaerobic. Optimum temperature for growth 22–30° C. Pigment formed most readily at 22° C. No hæmolysin formed. Ferments no sugars. L.M. unchanged. M.R. —. V.P. —. M.R. reduction —.

*Pathogenicity.*—Non-pathogenic.

*Sarcina lutea* Schroeter.—This organism, which was described by Schroeter (1873) as the *Bacteridium aurantiacum*, is a Gram-positive sarcina, giving a chrome-yellow growth on culture media. On agar the colonies are raised, yellow, coarsely granular, with an entire margin and a moist glistening surface. On potato they are dry, dull and granular. In broth there is an abundant yellow sediment with no turbidity. The optimum temperature of growth is 25° C. It is found in air, soil and water.

*Sarcina aurantiaca* Flügge.—Described by Flügge (1896). Gram-positive spheres developing in packets. On gelatin plates forms slowly-growing orange-yellow colonies gradually liquefying gelatin. On agar slope, gives a thick reddish-yellow growth composed of single colonies; similarly on potato. In broth gives a turbidity with abundant sediment. Grows best at room temperature. Found in air and water.

*Sarcina ureæ* Beijerinck (1901) is a spherical organism, 0.7–1.2  $\mu$  in diameter, occurring singly, in pairs, and in packets. Said to be motile by peritrichate flagella and to form spores. Gram-positive. Resists heating to 100° C. for 5 minutes. Gelatin not liquefied. Found in stale urine (see Gibson 1935).

*Sarcina conjunctivæ* Verderame.—Isolated by Verderame (1911) from the conjunctival sac of a girl suffering from acute conjunctivitis. Named by him *Sarcina conjunctivæ citrea*. Is stated to be Gram-negative. On agar after 24 hours the colonies are pinhead in size, round, bright-yellow, opaque, with an entire edge, and of butyrous consistency; after 48 hours they are 4 mm. in diameter and lemon-yellow in colour. On an agar slope there is a thick creamy layer of growth after 2 days. Similar growth, but more abundant on ascitic agar. In gelatin stab there is a surface growth of 2–3 mm. in diameter after 24 hours, and a filiform growth of fine, light, greyish-yellow, opaque colonies; no liquefaction. On potato an abundant, creamy, lemon-yellow growth of rather dry appearance. In broth after 24 hours there is a very light turbidity, with a suspension in the liquid of fine floculi which are easily broken up on shaking, and a cloudy viscous deposit of light yellow colour. Optimum temperature 37° C., but grows well at 14–18° C. Facultative anaerobe. Forms acid in glucose, levulose, maltose, lactose, sucrose and inulin, not in mannitol or galactose. H<sub>2</sub>S positive. Indole negative. Nitrates not reduced. Non-pathogenic to mice or guinea-pigs.

Numerous other sarcinae have been described, such as *Sarcina citrea* Menge in 1892, and an unnamed Gram-negative *Sarcina* isolated by Nagano (1902) from the pus of an ovarian abscess. Orskov (1930) draws attention to a sarcina found in the mouth and throat which, though non-motile itself, gives rise to motile cocci, each provided with a flagellum. The cocci themselves are not reproducible. For this remarkable organism the name *Sarcina mirabilis* is suggested.

### *Rhodococcus*

*DEFINITION.*—*Rhodococcus*.

Spherical or ovoid cells occurring in groups or regular packets. Usually Gram-positive, but are easily decolorized. Growth on agar abundant with formation of red pigment. Weak fermentative powers. Gelatin rarely liquefied. Nitrates generally reduced. Saprophytes.

Type species is *Rhodococcus rhodochrous* Zopf. The term *Rhodococcus* was first introduced by Zopf in 1891 (see Buchanan 1925).

*Rhodococcus rhodochrous* Zopf.—Found in water. Spherical organism, about 0.8–1.0  $\mu$  in diameter, occurring singly, in pairs and in small groups. Gram-positive. Gives a confluent, raised growth of a carmine hue on agar. Thick, rose-red pellicle in broth,

with a red flocculent sediment (Bergey 1923) L.M. slightly alkaline Carmine-red streak on potato Does not liquefy gelatin but generally reduces nitrates Aerobic Optimum temperature 25° C Non pathogenic

*Rhodococcus cinnabareus* Flugge—Described by Flugge as a large spherical Gram positive coccus occurring in twos threes and fours grows very slowly On gelatin after 4 days the colonies are 0.5–1.0 mm. in diameter at first they are brick red later cinnabar-red in colour In gelatin stab small white colonies are formed down the stab in 4 to 5 days on the surface there is a large red knob of growth no liquefaction On potato the growth is even slower Optimum temperature 25° C Does not reduce nitrates and cannot utilize ammonium salts as the sole source of nitrogen Found in air and water

*Rhodococcus roseofulvus* Flugge—Described by Babes as the cause of red sweat (see Flugge 1896) The cocci are oval in shape 1  $\mu$  long by 0.6–0.8  $\mu$  broad, and are bound by gelatinous material into a reddish zoogloeal mass In the body they surround the hairs particularly of the axilla, and impart a red coloration to the sweat Gram positive Grows on egg white at 37° C forming a red pigment Does not liquefy gelatin Generally curdles milk and causes slight peptonization Produces acid in glycerol and mannitol

*Rhodococcus agilis* Ali-Cohen—Isolated by Ali-Cohen (1889) from drinking water and named *Micrococcus agilis* Peculiar in being motile possessing one or two flagella Occurs mostly in pairs sometimes in short chains or in tetrads Gram positive 1  $\mu$  in diameter Grows in all media at room temperature forming a rose-coloured pigment Liquefies gelatin slowly Optimum temperature 25° C Found in water

### Leuconostoc

#### DEFINITION—*Leuconostoc*

Spherical or oval cells arranged in pairs and chains the cocci are surrounded by a gelatinous envelope which unites them into zoogloeal masses Usually Gram positive but decolorize easily Saprophytes usually growing in cane sugar solutions

Type species is *Leuconostoc mesenteroides* van Tieghem

The first organism of this group was described by Cienkowski in 1878 as the *Ascococcus mesenteroides* This name was amended by van Tieghem in 1878 to *Leuconostoc mesenteroides* (see Flügge 1896) According to Hucker and Pederson (1930) to whom reference should be made for more detailed information organisms of the genus *Leuconostoc* are found in slimy sugar solutions in fermenting vegetables and in milk and milk products Morphologically they are intermediate between the streptococci and the lactobacilli They all produce about 45 per cent lactic acid from glucose 20 per cent CO<sub>2</sub> and 25 per cent volatile products including acetic acid and ethyl alcohol They form mannitol from fructose and sucrose and a levulan or dextran from sucrose

*Leuconostoc mesenteroides* (Cienkowski) van Tieghem—A spherical coccus 0.9–1.2  $\mu$  in diameter occurring in pairs and in chains. Usually Gram positive The chains are surrounded by a thick tough gelatinous coating which is stated by von Schebler to consist of dextrin the aggregation of several chains within their envelopes gives rise to large compact gelatinous zoogloeal masses. Develops on the surface of parsnip-root and beetroot solutions in the form of thick cakes of cartilaginous consistency (Flügge 1896) It likewise thrives on grape-sugar and cane-sugar solutions provided nitrate and phosphate are added. Cultivated in peptone water or in gelatin to which lactose or maltose has been added it is morphologically similar to a streptococcus no gelatinous envelope being formed but if glucose or cane sugar is incorporated in the gelatin then the characteristic zoogloeal masses appear On either of these media there appears in 10 to 14 days a thick whitish mass of confluent colonies having a glassy surface looking

rather like a layer of crystals. During the first week the growth is dry and of cartilaginous consistency, but later it becomes softer and moister, and assumes an almost pulp-like consistency. Ferments glucose, maltose, lactose, sucrose, mannitol and either arabinose or xylose, with the formation of acid and gas. Facultative anaerobe. Optimum temperature 25°C. Is found in fermenting vegetable material and in sugar solutions.

Hucker and Pederson (1930) recognize two other species. *Leuconostoc dextranicum* ferments sucrose, but not pentoses, produces a moderate amount of slime from sucrose, and may be associated with either vegetable or dairy products.

*Leuconostoc citrovorum* does not ferment sucrose or pentoses, produces no slime from sucrose, and is generally associated with milk or milk products.

## REFERENCES

- ALI COHEN, CH. H. (1889) *Zbl. Bakt.*, 6, 33.  
 ANDREWES, F. W. and GORDON, M. H. (1905-6) *Rep. loc. Govt. Bd. publ. Hlth. Suppl.*, p. 543.  
 ARBEVIUS, S. (1907) *Immunochemie*. Akad. Verlagsgesell., Leipzig.  
 BAYLISS, M. (1940) *J. exp. Med.*, 72, 669.  
 BELJERINCK, M. W. (1901) *Zbl. Bakt.*, IIte Abt., 7, 33.  
 BERGEY, D. H. (1923) *Manual of Determinative Bacteriology*. Baltimore.  
 BEUMER, J. (1939) *C. R. Soc. Biol.*, 130, 233.  
 BIGGER, J. W. (1933) *J. Path. Bact.*, 36, 87.  
 BIGGER, J. W., BOLAND, C. R., and O'MEARA, R. A. Q. (1927) *J. Path. Bact.*, 30, 261.  
 BIONDI, D. (1887) *Z. Hyg. Infectkr.*, 2, 194.  
 BLAIR, J. E. and HALLMAN, F. A. (1935) *Proc. Soc. exp. Biol.*, N. Y., 33, 392. (1936) *J. Bact.*, 31, 81.  
 BRYCE, L. M. and ROUNTREE, P. M. (1936) *J. Path. Bact.*, 43, 173.  
 BUCHANAN, R. E. (1925) *General Systematic Bacteriology*. Baltimore.  
 BURDET, F. M. (1929) *J. Path. Bact.*, 32, 717. (1930) *Ibid.*, 33, 1. (1931) *J. Path. Bact.*, 34, 759.  
 CADNESS GRAVES, B., WILLIAMS, R., HARPER, G. J., and MILES, A. A. (1943) *Lancet*, 1, 736.  
 CHAPMAN, G. H. (1936) *J. Bact.*, 22, 199. (1944) *J. Bact.*, 47, 211.  
 CHAPMAN, G. H. and BERENS, C. (1935) *J. Bact.*, 29, 437.  
 CHAPMAN, G. H., BERENS, C., NELSON, E. L., and CURCIO, L. G. (1938) *J. Bact.*, 35, 311.  
 CHAPMAN, G. H., BERENS, C., PETERS, A., and CURCIO, L. (1934) *J. Bact.*, 28, 343.  
 CHAPMAN, G. H., LIER, C. W., BERENS, C., and CURCIO, L. (1937) *J. Bact.*, 33, 533.  
 CHRISTIE, R. and GRAYDON, J. J. (1940) *Aust. J. exp. Biol.*, 19, Pt. 1, 9.  
 CHRISTIE, R. and KROGH, E. V. (1940) *J. Path. Bact.*, 51, 169.  
 COHN, F. (1875) *Oechsle Beitr. Biol. Pflanz.*, 1, Hft. 2, 127.  
 COWAN, S. T. (1938) *J. Path. Bact.*, 46, 31. (1939a) *Ibid.*, 48, 169. (1939b) *Ibid.*, 48, 545.  
 CRUICKSHANK, R. (1937) *J. Path. Bact.*, 45, 295.  
 DARÁNYI, J. V. (1926) *Zbl. Bakt.*, 99, 74.  
 DAVISON, E. and DACK, C. M. (1939) *J. infect. Dis.*, 64, 302.  
 DAVISON, E., DACK, C. M., and CART, W. E. (1938) *J. infect. Dis.*, 62, 219.  
 DEVYS, J. and HAVET, J. (1898) *La Cellule*, 10, 7.  
 DEVENISH, E. A. and MILES, A. A. (1939) *Lancet*, 1, 1088.  
 DOLMAN, C. E. (1932) *Canad. publ. Hlth. J.*, 23, 125. (1943) *Ibid.*, 34, 45. (1944) *Ibid.*, 35, 337.  
 DOLMAN, C. E. and WILSON, R. J. (1938) *J. Immunol.*, 35, 13. (1940) *Canad. publ. Hlth. J.*, 31, 68.  
 DOLMAN, C. E., WILSON, R. J., and COCKCROFT, W. H. (1936) *Canad. publ. Hlth. J.*, 27, 489.  
 DOWNIE, A. W. (1937) *J. Path. Bact.*, 44, 573.  
 DUDGEON, L. S. (1908) *J. Path. Bact.*, 12, 242.  
 DUDGEON, L. S. and SIMPSON, J. W. H. (1923) *J. Hyg., Camb.*, 27, 160.  
 DUNCAN, J. T. and WALKER, J. (1942) *J. Hyg., Camb.*, 42, 474.  
 DUFFY, O. (1942) *J. Bact.*, 44, 589.  
 EVANS, A. C. (1916) *J. infect. Dis.*, 18, 437.  
 FAIRBROTHER, R. W. (1940) *J. Path. Bact.*, 50, 83.  
 FIELDS, P. and RICHARDSON, G. M. (1937) *Brit. J. exp. Path.*, 18, 292.  
 FIELDS, P., RICHARDSON, G. M., KNIGHT, B. C. J. G., and GLADSTONE, G. P. (1936) *Brit. J. exp. Path.*, 17, 481.

- FINLAND, M., PETERSON, O. L., and STRAUSS, E. (1942) *Arch intern Med.* 70, 183  
 FISK, A. (1940) *Brit J exp Path.*, 21, 311  
 FISK, R. T. (1942) *J infect Dis.*, 71, 153, 161  
 FLAUM, A. (1938) *Acta path microbiol Scand.* Suppl. 35  
 FLUGGE, C. (1896) "Die Mikroorganismen," Part II, p 96, 3rd edit. Leipzig  
 FORSSMAN, J. (1935-1941) *Acta path microbiol Scand.* Numerous references  
 FRAENKEL, C. and BAUMANN (1905) *Munch med Wochr.*, 52, 937  
 FULTON, F. (1943) *Brit J exp Path.*, 24, 65  
 GAFFKY (1883) *Arch. klin Chir.*, 28, 495  
 GARROD, L. P. (1942) *Brit med J.*, 1, 290  
 GENCOU, O. (1932) *Ann Inst Pasteur*, 48, 135  
 GIBSON, T. (1935) *Arch. Mikrobiol.*, 6, 73  
 GILLESPIE, E. H. (1943) *Mon. Bull. Emerg. publ. Hlth. Lab. Serv.*, 2, 19  
 GILLESPIE, E. H., DEVENISH, E. A., and COWAN, S. T. (1939) *Lancet* ii. 8.0  
 GLADSTONE, G. P. (1937) *Brit J exp Path.*, 18, 322, (1938) *Ibid.* 19, 208  
 GLADSTONE, G. P., FIELDS, P., and RICHARDSON, G. M. (1935) *Brit J exp Path.*, 16, 335  
 GLENNY, A. T. and STEVENS, M. F. (1935) *J Path. Bact.*, 40, 201  
 GOODSER, J. (1842) *Edin. med surg J.*, 57, 430  
 GORDON, M. H. (1903-4) 33rd ann. Rep. loc. Govt Bd publ. Hlth. Suppl. p 388  
 GOYLE, A. V. and MINCHIN, R. L. H. (1940) *Indian J. med Res.*, 27, 611  
 GROSS, H. (1931a) *Zbl. Bakt.*, 122, 354, (1931b) *Ibid.*, 123, 212, (1931c) *Z. Immunforsch.* 73, 14.  
 HALLMAN, F. A. (1937) *Proc. Soc. exp. Biol. N. Y.*, 38, 789  
 HAMMON, W. M. (1941) *Amer J. publ. Hlth.*, 31, 1191  
 HEGEMANN, G. (1937) *Zbl. Bakt.*, 140, 108  
 HEISE, M. D. and STARIN, W. A. (1940) *J infect Dis.*, 67, 70  
 HEWITT, L. F. (1930) *Biochem. J.*, 24, 676  
 HINE, T. G. M. (1922) *Lancet*, ii. 1380  
 HOFFSTADT, R. E. and YOUNG, G. P. (1932) *J infect Dis.*, 51, 216.  
 HUCKER, G. J. (1924a) *N. Y. St. agric. Exp. Sta. Tec. Bull.*, No 100, (1924b) *Ibid.*, No 102  
 (1928) *N. Y. St. agric. Exp. Sta. Tec. Bull.*, No 135  
 HUCKER, G. J. and PEDERSON, C. S. (1930) *N. Y. St. agric. Exp. Sta. Tec. Bull.* No 167  
 HUGHES, T. P. (1932) *J. Bact.*, 23, 437  
 JACKSON, S. H., NICHOLSON, T. F., and HOLMAN, W. L. (1940) *J. Path. Bact.* 50, 1  
 JONES, A. H. and LOCHHEAD, A. G. (1939) *Food Res.*, 4, 203  
 JULIANVILLE, L. A. (1922) *J infect Dis.*, 31, 256  
 JULIANVILLE, L. A. and WIGGARD, C. W. (1934) *Proc. Soc. exp. Biol. N. Y.*, 31, 947  
 (1935) *J exp. Med.*, 62, 11, 31  
 KITCHING, J. S. and FARRELL, L. N. (1936) *Amer J. Hyg.*, 24, 268  
 KLIGLER, I. J. (1913) *J infect Dis.* 12, 432  
 KLIGLER, I. J., GROSSOWITZ, N., and BERGER, S. (1943) *J. Bact.*, 46, 399  
 KLOPFSTOCK and BOCKENHEIMER. (1904) *Arch. klin Chir.*, 72, 325  
 KNIGHT, B. C. J. G. (1935) *Brit J exp Path.*, 16, 315 (1937) *Biochem. J.* 31, 731  
 KOCH, J. (1908) *Z. Hyg. InfektKr.*, 58, 287  
 KODAMA T. and KODAMA, T. (1939) *Kitasato Arch.*, 18, 36  
 KODAMA T. and NISYAMA S. (1938) *Kitasato Arch.*, 15, 247  
 KOLLE W. and OTTO, R. (1902) *Z. Hyg. InfektKr.*, 41, 389  
 KRAUS, R. and CLAIRMONT, P. (1900) *Wien klin Wochr.*, 13, 49  
 KUPCHIK, G. T. (1937) *J infect Dis.* 61, 320  
 KUTSCHER and KOWRICH, F. (1904) *Z. Hyg. InfektKr.*, 48, 249  
 LANDSTEINER, K. and RAUCHENBUCHLER, R. VON (1909) *Z. Immunforsch.* 1, 439  
 LEVINE B. S. (1939) *J Path. Bact.*, 48, 291  
 LINGELSHREIN, W. VON (1899) *Beitr. exp. Ther.*, Heft. 1, 49  
 LOEB, L. (1903-4) *J med Res.*, 10, 407  
 LUBINSKY, W. (1894) *Zbl. Bakt.*, 18, 769  
 LYONS C. (1937) *Brit. J exp Path.*, 18, 411  
 MCBROOM J. (1937) *J infect. Dis.*, 60, 364  
 MCFARLAN, A. M. (1938) *Brit med J.*, ii. 939  
 MARCUCCIO, G. (1938) *Riv. sanit. sicil.* 26, 621, 625 6<sup>th</sup>  
 MERCIER, P. (1938) *C. R. Soc. Biol.*, 127, 297  
 MIGULA, W. (1900) "System der Bakterien," IIte Band, p 1. Gustav Fischer. Jena  
 MILES A. A., WILLIAMS R. E. O. and CLAYTON COOPER, B. (1944) *J Path. Bact.*, 58, 513  
 MINNETT, F. C. (1936) *J Path. Bact.*, 42, 247  
 MORGAN F. G. and GRAYDON, J. J. (1936) *J Path. Bact.*, 43, 335.  
 MUCH, H. (1908) *Biochem. Z.*, 14, 143  
 NAGANO, J. (1902) *Zbl. Bakt.*, 32, 327

- NEISSER, M. (1919) See Kofke and Wassermann, "Hdb. der path. Mikroorg.," 11te Abt. (1912 13), 4, 356.  
 NEISSER, M. and WECHBERG F. (1901) *Z Hyg InfektKr.*, 28, 909.  
 NETER, E. (193) *J Bact.*, 24, 943.  
 OGSTON A. (1891) *Brit. med. J.*, 1, 369.  
 OSTERLIN E. (1925) *Zbl. Bakt.*, 94, 313.  
 O KANE, D J (1941) *J Bact* 41 441  
 OSCUTT M. L. and HOWE, P. E. (1927) *J exp. Med.*, 35, 409.  
 ORSKOV J. (1930) *Acta path. Microbiol. scand. Suppl.*, 3, 519.  
 OTTO, R. (1903) *Zbl. Bakt.*, 34, 44.  
 OZAKI, Y. (1915) *Zbl. Bakt.*, 78, 118.  
 PANTON P. and VALENTINE, F. C. O. (1932) *Lancet*, 1, 506.  
 PARISH, H. J. and CLARK, W. H. M. (1937) *J. Path. Bact.*, 35, 251.  
 PARKER, J. T. (1924) *J exp. Med.*, 40, 61.  
 PASSET (1885) *Fort. Med.*, 3, 33.  
 PERAGALLO I. (1937) *G. Bakt. Immun.*, 18, 577.  
 PROOM H. (193) *J Path. Bact.*, 44, 423.  
 PRÖSCHER. (1903) *Zbl. Bakt.*, 34, 43.  
 PROVIA, O. (1884) *Cohns Beitr. Biol. Pflanz.*, 4, 400.  
 REIMAN, H. A. (1936) *J Bact* 31, 333 40.  
 Report (1942) *Mos. Bull., Emory pub. Hlth Lab. Ser.* 1, Feb., p. 9.  
 RICHMOND J. J. PEED C. L., SHATKENESEY H. J., and MICHAEL, V. (1941) *J. Bact.*, 44, 901.  
 PIGDON P. H. (1937) *J Lab. clin. Med.*, 24, 140. (1937) *Proc. Soc. exp. Biol. N. Y.*, 38, 8.  
 ROSENBERG, F. J. (1884) "Mikroorganismen bei den Wundinfektionskrankheiten des Menschen." Wiesbaden.  
 SAKEL, S. and FEJGIN B. (193) *C. R. Soc. Biol.* 126, 139.  
 SCHROETER, J. (1885) *Cohns Beitr. Biol. Pflanz.*, 1, Heft. 2, 109.  
 SEGALOVE, M. and DACK, G. M. (1941) *Food P.*, 6, 177.  
 SEITZERT W. (1935-36) *Zbl. Bakt.*, 135 100.  
 SLANETZ, L. W. (1947) *J Bact.*, 43, 100.  
 SMIT J. (1933) *J Path. Bact.*, 36, 433.  
 SMITH, M. L. (1937) *J Path. Bact.*, 45 300.  
 SMITH, M. L. and IPSEN J. (193) *Quart. Bull. Hlth. Org. L.A.*, 7, 845.  
 SMITH M. L. and PRICE, S. A. (1937) *J Path. Bact* 47 361 (1938) *Ibid* 47 379.  
 SMITH, W. and HALE, J. H. (1944) *Brit. J. exp. Path.*, 25, 101.  
 STEPHAN F. (1934) *Z Hyg InfektKr.*, 116, 550.  
 SWINGLE, E. L. (1935) *J Bact.*, 29 46.  
 THOMPSON P. and KHORAZO D. (1937) *J Bact.*, 34, 69.  
 VALENTINE, F. C. O. (1936) *Lancet* 1, 576.  
 VALENTINE, F. C. O. and BUTLER, E. C. B. (1939) *Lancet*, 1, 973.  
 VANERUSSEHEM, R. (1934) *C. P. Soc. Biol.*, 116, 650.  
 VIEL, F. (1904) *Munch. med. Wochs.*, 51, 13.  
 VELDIE, H. VAN DER. (1894) *La Cellule*, 10, n. 401.  
 VERDERAME, P. (1911) *Zbl. Bakt.*, 59 377.  
 VERWEY W. F. (1940) *J exp. Med.*, 71, 633.  
 WALBYM, L. E. (1922) *Biochem. Z.*, 123, 367.  
 WALSTON H. D. (1935) *J Hyg., Camb.*, 35, 549.  
 WELCH, W. H. (1891) *Int. J. med. Sci.*, 102, 439.  
 WELD J. T. and MITCHELL, L. C. (1947) *Proc. Soc. exp. Biol. N. Y.*, 49 3 0.  
 WIEGHARD C. W. and JULIANVILLE, L. A. (1935) *J exp. Med.*, 62, 23.  
 WILSON G. S. and ATKINSON J. D. (1943) *Lancet*, 1, 64.  
 WINSLOW C. E. A. and ROGERS, A. F. (1906) *J. infect. Dis.*, 3, 48a.  
 WINSLOW C. E. A., POTTSBERG W., and PARSONS, E. I. (1920) *J. Bact.*, 5, 145.  
 WINSLOW C. E. A. and WINSLOW A. R. (1908) "The Systematic Relationship of the Coccaceae." New York.  
 WRESCHNER, H. (1921) *Z Hyg InfektKr.*, 93, 4.  
 WRIGHT J. (1936) *Lancet* 1, 1002.  
 YOVENURA V. (1938) *Z. Immunforsch.*, 89 392.

## CHAPTER 26

### CHROMOBACTERIUM AND ACHROMOBACTERIUM

#### CHROMOBACTERIUM

##### DEFINITION —*Chromobacterium*

Small, non sporing, aerobic rods, usually motile and usually Gram negative producing a yellow, red, or violet pigment, which is generally insoluble in water Saprophytic, commonly found in water or soil

Type species. *Chromobacterium violaceum*

Numerous organisms of this group have been isolated at various times from water, soil, sewage, and occasionally from contaminated food stuffs *Chr violaceum* was described by Bergonzoni in 1881 (see Report 1920) *Chr prodigiosum* was first observed by Bizio in 1823 (see Breed and Breed 1924), who found it in "bleeding polenta" *Chr aquatilis* was isolated from water by Frankland and Frankland in 1889

The classification of these organisms presents considerable difficulties The American Committee of Bacteriologists has created a genus, which they call *Erythrobacillus*, for the inclusion of those bacilli which produce a red or pink pigment The genus *Chromobacterium* they reserve for those bacilli which produce a violet pigment As these organisms have received relatively little attention from bacteriologists, and as in consequence their properties have been incompletely studied, it would seem advisable for the moment to group all the aerobic non sporing pigment forming rods into the single genus *Chromobacterium* With regard to the naming of the individual species within the genus, there is considerable confusion One and the same organism has frequently been described under two names, and specific names have been given to organisms which appear to be merely varieties of existing species We shall therefore describe in some detail those organisms which undoubtedly deserve specific rank, and shall refer briefly to others whose claim to this distinction is more doubtful

**Morphology.**—The organisms of this group are small rods, varying in length from about 1.0 to 3.0  $\mu$ , and in breadth from about 0.5 to 0.7  $\mu$  *Chr prodigiosum* may be regarded as an exception, since it is usually described as a small cocco-bacillus It is necessary to point out, however, that the size of this organism is subject to considerable variation, and that even on the same type of medium a single strain may at one time give rise to cocco bacilli, and at another to rod forms indistinguishable from those formed by other members of *Chromobacterium* (Fig 129) Motility is a frequent characteristic, and is dependent on the possession of peritrichate or sometimes polar flagella Most members of this group are Gram negative

**Cultural Reactions.**—Growth occurs readily on the ordinary media, a particularly heavy growth is generally observed in broth



FIG 129—*Chromobacterium prodigiosum*

From an agar culture 24 hours 37° C (× 1000)

Left—Strain showing coccobacillary forms

Right—Same strain from a different culture showing definite rod

The colonies on agar are usually homogeneous for the first day or two and then become differentiated into a convex pigmented and relatively opaque centre and an effuse colourless almost transparent periphery with an irregularly crenated edge. This differentiation results from the secondary outgrowth around the original colony of a film which may be so thin as to escape observation unless a careful search is made for it.

In agar stroke cultures there is a raised confluent pigmented growth with a smooth or beaten copper surface and an undulate or lobate edge.

In gelatin stab culture there is a moderate filiform growth extending to the bottom of the tube usually succeeded by liquefaction which may occur rapidly or slowly. In general the liquid is turbid and granular and in the upper centimetre or so is pigmented; below this out of contact with the air the growth is colourless. A surface pellicle or ring growth strongly pigmented and a slightly pigmented deposit at the bottom of the liquefied gelatin are not uncommon.

In broth there is a dense turbidity generally with a pigmented ring growth or surface pellicle and a slightly pigmented deposit which frequently becomes viscous and is difficult to disintegrate on shaking. The broth itself often remains colourless. Multiplication of the bacilli seems to occur chiefly near the surface; a membrane forms in contact with the air, sinks to the bottom and is replaced by a fresh membrane. This accounts for the heavy deposit that is generally present after a week.

Blood serum is frequently liquefied.

Resistance—The organisms of this group are non sporing and show no par

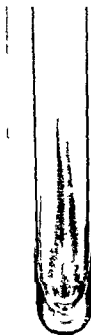


FIG 130—*Chromobacterium prodigiosum*  
Agar stroke culture  
24 hours 37° C

ticular resistance to heat or disinfectants. They appear to be killed by a temperature of 55° C in 1 hour.

**Metabolism and Biochemical Characters**—All the members are markedly aerobic, growing best in the presence of an ample supply of oxygen. Growth under anaerobic conditions is poor, and is not accompanied by pigment formation.

Proteins are broken down readily as a rule, gelatin, casein and often blood serum are digested. Carbohydrates are attacked freely by some sparingly by others, generally with the production of acid only.

Growth can be obtained on synthetic media devoid of protein. The optimum temperature of growth is between 25° and 37° C, but some strains will grow as low as 0° C.

All form catalase and reduce methylene blue. Indole is rarely produced. The M R and V P reactions are generally negative. Nitrates are often reduced to nitrites, and ammonia is generally formed from peptone. Formation of  $H_2S$  is slight or absent.

Pigment is produced only in the presence of oxygen, and at a suitable temperature. The optimum temperature for pigment formation does not necessarily correspond with that for growth. Thus many organisms grow best at 30–37° C, but form little or no pigment, whereas at a lower temperature growth is poorer but pigment formation is abundant. Pigment is developed best on the surface of solid media, in broth and in the depths of stab cultures there is little or none formed. Potato is a medium that may be specially recommended for the study of pigment production. As a rule pigment is formed most abundantly on primary isolation, after subculture in the laboratory for some time, the power to form it may diminish seriously, or be altogether lost. Not all bacilli in a given culture produce the same amount of pigment, some colonies are deeply coloured others are faintly or partly coloured while still others are completely colourless.

Sullivan (1905, 6) states that the formation of pigment is dependent on the reaction of the medium, the temperature, the free access of oxygen and the presence of certain salts. In synthetic media, asparagin, succinic lactic and citric acids, when combined with an ammonium base allow of growth and pigment formation. Malic, tartaric, and oxalic acids combined with ammonia, allow of growth but not of pigment production. Acetic, uric, and formic acids, combined with ammonia, are unfavourable for both growth and pigment production. (This refers to solutions of 0.4 per cent or under.) The salts favouring the production of pigment serve either to provide nutrient material or to fix the acid produced, or to afford material for essential syntheses.

**Pathogenicity**—The organisms of this group are essentially saprophytic. A few doubtful cases of suppuration in man have been described (see Schutz and Laun 1933), but generally speaking these organisms do not give rise to natural disease in man or in animals. On experimental inoculation into laboratory animals they prove harmless except in very large doses.

**Variation**—It is convenient to mention here that the characters of a single strain are liable to considerable variation. Its cultural appearances are by no means uniform, colonies may vary in their size, shape, opacity, surface, and consistency.

In their ability to produce pigment there is likewise great variation. From 7 different strains of *Chr prodigiosum* and *Chr kieltense*, Eisenberg (1914) isolated



no fewer than 22 variants differing in colour from dark red to red, orange-red, pink, pale-pink, pinkish white, and colourless, most of these remained stable on subculture. From *Chr violaceum* 5 variants were obtained. Eisenberg found that variants appeared most readily in ageing cultures, and more rapidly in fluid than in solid media.

**Group Producing a Violet Pigment**—*Chr violaceum*, the chief member of the group is a common inhabitant of water. The violet pigment is soluble in alcohol, but insoluble in water, chloroform or benzol. For its formation in synthetic media both magnesium sulphate and a phosphate are required. On subculture the organism frequently loses its pigment forming power. We append a detailed description of this organism, together with brief accounts of some of the other violet pigment producing members.

### *Chromobacterium violaceum*

*Synonym*—*B. violaceus*

*Isolation*—Described by Bergonzoni in 1881 (see Text).

*Habitat*—Water

*Morphology*—Rods  $1.5-3.0 \mu \times 0.6 \mu$ , axis straight sides parallel, ends rounded, arranged singly. Motile by peritrichate flagella. Non-sporing, non-capsulated. Gram negative non-acid fast. May show bipolar staining.

*Agar Plate*.—2 days at  $37^{\circ} \text{C}$ . Round colonies  $\frac{1}{2}$ –1 mm. in diameter, low convex violet coloured translucent with a smooth glistening surface and an entire edge, butyrous consistency and easily emulsifiable, structure appears floccular. After 5 days colonies are larger 1–2 mm. in diameter have a finely lobate edge, and are differentiated into a dark convex centre, and a paler flattened, radiate periphery.



FIG 131—*Chromobacterium violaceum*

Surface colony on agar 48 hours  $22^{\circ} \text{C}$ . ( $\times 8$ )

*Agar Slope*.—2 days  $25^{\circ} \text{C}$ . Moderate growth, raised, violet, opaque with glistening smooth or beaten-copper surface and an undulate edge. Medium unchanged.

*Gelatin Stab*.—5 days  $20^{\circ} \text{C}$ . Moderate, filiform growth to bottom of stab, violet surface growth, slow infundibuliform or saecate liquefaction, Liquefied portion is granular and violet.

*Broth*.—2 days  $25^{\circ} \text{C}$ . Abundant growth, with dense colourless turbidity, a violet ring growth, and a moderate viscous deposit disintegrating on shaking. Mawkish odour.

*Blood Serum*.—9 days  $25^{\circ} \text{C}$ . Moderate confluent growth colourless, with an uneven surface. Digestion is very slow, or may be absent.

*Potato*.—6 days at  $25^{\circ} \text{C}$ . Abundant, raised, confluent, creamy growth of violet colour, with smooth or contoured glistening surface.

*Persistence*.—Killed by  $55^{\circ} \text{C}$ . in 1 hour.

*Metabolism*.—Aerobic slight colourless growth anaerobically. Opt. temp.  $25^{\circ} \text{C}$ . Pigment formation best at  $25-30^{\circ} \text{C}$ . Pigment soluble in alcohol, insoluble in chloroform or water.

*Nutritional*.—Grows well on ordinary media. In synthetic media, pigment is formed only in the presence of  $\text{MgSO}_4$  and a phosphate.

*Biochemical*.—Sometimes acid in glucose and maltose. Indole— or slight +. M.R. — V.P. — Nitrate reduction +  $\text{NH}_3$  + Catalase + M.B. reduction +  $\text{H}_2\text{S}$  slight +. L.M. 6 days  $25^{\circ} \text{C}$ . alkaline violet ring growth, later may be coagulated, and slowly peptonized.

*Pathogenicity*—Non pathogenic to man and animals.

*Chr janthinum*, found in water and soil, is said to be motile by one or two polar flagella. Colonies on agar are at first milky white but later become violet. Liquefaction of gelatin is slow. On broth it forms a violet surface pellicle. Optimum temperature 30° C. It is doubtful whether this organism is a distinct species.

*Chr amethystinum* is a non motile bacillus found in water. On agar the growth is first non pigmented, later it assumes a dark violet colour with a wrinkled surface of metallic lustre. Gelatin is liquefied, a violet surface pellicle forms on the liquid. In broth a surface pellicle appears, the fluid itself is coloured brown. Optimum temperature 30° C.

*Chr cœruleum* is said to be motile by polar flagella. Colonies are spreading and of a bluish grey colour. Slow liquefaction of gelatin. Greyish surface pellicle on broth. Milk is rendered sky blue at the surface and is digested with an alkaline reaction. The pigment is soluble in water and alcohol but not in ether or chloroform. Optimum temperature 30° C.

**Group Forming a Pink or Red Pigment**—The most important member of this group is *Chr prodigiosum*. It was first demonstrated by Bartolomeo Bizio in 1823 as a cause of "bleeding polenta" (Breed and Breed 1924). Infections of meat, fish, bread and other articles of food with this organism from time to time give rise to alarm. Klein (1894) describes an instance where the food in a large mercantile establishment in London became contaminated with it. In spite of the fact that the residents consumed pink meat for some days, they suffered from no apparent ill effects. The pigment is soluble in absolute alcohol, ether, chloroform, benzol and carbon disulphide, but is insoluble in water. An alcoholic solution when acted on by mineral acids is turned first carmine red, then reddish violet. Alkalies turn it brownish yellow, and chlorine water after turning it reddish brown then golden yellow, finally decolorizes it. In the spectrum blue and violet are completely absorbed and an absorption band is seen in the green. Wrede (1930) gives its empirical formula as  $C_{22}H_{22}N_2O$  and says that only one of the nitrogen atoms will combine with acids. The pigment is destroyed in a few days by sunlight. It is very resistant to reduction but is fairly easily oxidized. In synthetic media it is formed only in the presence of magnesium sulphate and a phosphate, preferably potassium phosphate (Kuntze 1900, and Sullivan 1905-06). Its formation is said to be inhibited by the presence of calcium salts (Bordet 1930). It is produced most readily at 22° C, and according to Amako (1930) there is a close parallelism in cultures between the catalase content and the amount of pigment formed.

Hefferan (1904) whose detailed study is one of the most valuable on this group of bacteria proposed the following classification.

**Pigment insoluble in water**

Group I *Chr prodigiosum*, *Chr indicum*, *Chr kielsense* etc. Pigment red at first, later becomes darker—carmine or violet red.

Group II *Chr rubricum*, *Chr ruber*, etc. Develop more slowly. Pigment is orange red or yellow red, never becoming darker.

Group III *Chr mycoides roseum*, etc. Pigment is salmon pink, coral pink, rose or flesh coloured.

**Pigment soluble in water**

Group IV *Chr lactis erythrogenes*, *Chr rubefaciens* etc. Rose red pigment.

We append a detailed description of *Chr prodigiosum* and *Chr indicum*. For a description of other members of this group, see Hefferan (1904)

### *Chromobacterium prodigiosum*

*Synonyms*.—*Serratia marcescens* Bizio *Micrococcus prodigiosus* Cohn.

*Isolation*.—First described by Bizio in 1823 (see Text).

*Habitat*.—Water and air, found on bread, meat, milk, potatoes and other food stuffs.

*Morphology*.—Tiny rods often oval or cocco-bacillary,  $0.7-1.0 \mu \times 0.7 \mu$ , sides convex, ends rounded, arranged singly and in groups, irregularity in shape frequent, there being generally some definitely bacillary forms present. Motile by 2-4 peritrichate flagella. Non-sporing, non-capsulated. Gram negative, non-acid fast, bipolar staining not uncommon.

*Agar Plate*.—2 days,  $25^{\circ}C$  Round colonies, 1-2 mm. in diameter, low convex, smooth, glistening paint like surface, entire edge, amorphous structure, butyrous consistency, easily emulsifiable. Individual colonies vary from colourless to bright red in colour, all are translucent. 5 days Colonies are larger, 2-4 mm. in diameter,

surface radially striated, edge finely lobate, differentiated into a low convex, red, opaque centre, and an effuse transparent, colourless periphery.

*Agar Slope*.—Abundant, raised, opaque scarlet growth, with smooth paint like surface and lobate edge. Medium unchanged.

*Gelatin Stab*.—Abundant, filiform growth to bottom of tube. Liquefaction infundibuliform reaching to bottom of tube in 5 days at  $20^{\circ}C$ , liquid is pink at upper part and turbid, red ring growth at surface, pink floccular deposit, rest of growth uncoloured.

*Broth*.—2 days,  $25^{\circ}C$  Luxuriant pink growth, with dense turbidity, a red ring growth, and a moderate, reddish, powdery or finely granular deposit mostly disintegrating on shaking. After 5 days

there is a red surface pellicle and a heavy flocculo-granular deposit. Mawkish odour, like trimethylamine.

*Blood Serum*.—7 days,  $25^{\circ}C$  Profuse growth, medium mostly liquefied, fluid is red and turbid.

*Potato*.—6 days at  $25^{\circ}C$  Abundant, thick, raised, confluent growth of dark maroon colour, surface papillate and slightly glistening. Potato not coloured.

*MacConkey Plate*.—24 hours at  $37^{\circ}C$  Circular, low convex, opaque, amorphous, pink colonies, 1 mm. in diameter, with smooth glistening surface and entire edge.

*Resistance*.—Destroyed by  $55^{\circ}C$ . for 1 hour.

*Metabolic*.—Aerobe, slight colourless growth anaerobically. Opt. temp.  $25-30^{\circ}C$ . Grows at  $37^{\circ}C$ , but no pigment produced. Pigment blood red, soluble in chloroform not in water.

*Nutritional*.—Grows well on ordinary media. In synthetic media pigment is formed only in the presence of  $MgSO_4$  and a phosphate.

*Biochemical*.—Acid or acid and gas in glucose, maltose, mannitol, sucrose and salicin, occasionally in lactose. Indole — M.R. — V.P. + Nitrates reduced.  $NH_3$  production + Catalase + M.B. reduction ++ Starch diastase —  $H_2S$  + I.M. clotted in 2 days, peptonized in 5 days, fluid pinkish.

*Pathogenicity*.—Non pathogenic to man, and to animals except in enormous doses.



FIG 13<sup>a</sup>—*Chromobacterium prodigiosum*

Surface colony on agar 24 hours,  $37^{\circ}C$  ( $\times 8$ )

*Chromobacterium indicum*

*Isolation*—By Koch from the stomach of an ape.

*Morphology*—Slender, often curved, bacillus,  $2-4\ \mu$  long by  $0.6\ \mu$  broad, motile by peritrichate flagella. Gram-negative. Non acid fast.

*Cultural Characters*—Grows readily on ordinary media.

*Agar plate*—2 days at  $25^{\circ}\text{C}$  Colonies are circular, 1 mm. in diameter, amorphous, low convex, translucent, and pink, with a smooth surface and entire edge, butyrous in consistency and easily emulsifiable. After 5 days, there is usually a thin, transparent, colourless peripheral fringe with a finely lobate edge.

*Gelatin*—5 days at  $20^{\circ}\text{C}$  Abundant growth, complete liquefaction of gelatin, upper  $\frac{1}{2}$  cm. of the liquefied gelatin is densely turbid and bright red in colour, pinkish deposit at bottom of tube.

*Broth*—2 days,  $25^{\circ}\text{C}$  Luxuriant growth, with dense turbidity, a pink surface pellicle and ring growth, and an abundant, moderately granular, pink deposit, which disintegrates partly on shaking.

*Potato*—6 days,  $25^{\circ}\text{C}$  Moderate, slightly raised, confluent growth of café au lait colour, having a smooth or finely granular, contoured, moist, glistening surface.

*Loeffler's serum*—5 days  $25^{\circ}\text{C}$  Abundant, confluent, raised, pinkish growth, with a glistening contoured surface, slight digestion, increasing with further incubation.

*Biochemical Reactions*—Same as *Chr prodigiosum*

*Pathogenicity*—Nil.

*Chromobacterium kielense*—This organism, which was isolated from water, resembles *Chr prodigiosum* very closely, it is not clear, in fact, that it is a separate species. It is said uniformly to produce both acid and gas in carbohydrate media but as many strains of *Chr prodigiosum* also produce gas, this distinction is an imperfect one.

*Group Forming a Yellow or Orange Pigment*—*Chr aquatile*, an organism isolated by the Franklands (1889) from deep wells in the Kent chalk, is a typical example of this group. The yellow pigment is insoluble in water but dissolves in alcohol, ether and chloroform. In non albuminous media it is developed slowly, but it appears rapidly in a peptone solution containing magnesium sulphate and dipotassium hydrogen phosphate.

We append a detailed description of *Chr aquatile* and *Chr typhi flavum*, together with brief notes on a few of the other members of this group. The identity of *Chr aquatile* is not very certain, since the original description was incomplete. The present description is founded on a study of the strain of *Chr aquatile* obtained from the National Collection of Type Cultures, London. *Chr typhi flavum* is the name we suggest for the organism commonly known as *Bact typhi flavum*. Though suspected by certain German workers of being a pigmented variant of the true typhoid bacillus, the evidence, which has been critically reviewed by Cruickshank (1935), is quite insufficient to establish any such relationship.

*Chromobacterium aquatile*

*Synonym*.—*Bacillus aquatilis* Frankland.

*Habitat*.—Water.

*Morphology*—Slender rod-shaped organism,  $2.5\ \mu \times 0.6\ \mu$ , axis straight, sides parallel, ends rounded, arranged in bundles, length irregular. Motile by peritrichate flagella. Non sporing, non-capsulated. Gram negative, non-acid fast.

**Agar Plate.**—2 days at 25° C. Rounded colonies, 1-2 mm. in diameter, low convex, opaque, yellowish-grey colour, with a dull, dry, granular or russe surface, and an entire edge, consistency slightly membranous but emulsification fairly easy. After 5 days, colony shows differentiation into finer granular, yellowish-brown, convex centre, and a clear almost transparent, effuse, and sometimes radiate periphery which has an erose or villous edge.

**Agar Slope.**—Abundant, confluent, opaque, yellowish, raised growth, with smooth shining surface and a lobate or villous edge.

**Broth.**—2 days at 25° C. Slight turbidity, with slight powdery deposit. After 5 days the growth is more abundant; there is a very slightly granular turbidity, and sometimes a surface pellicle and ring growth; moderate membranous deposit, disintegrating incompletely on shaking.

**Gelatin Slab.**—5 days at 20° C. Good growth, extending to bottom of tube; gelatin shows commencing infundibuliform liquefaction, the liquefied gelatin shows a floccular turbidity, and is covered with a granular pellicle.

**Potato.**—6 days at 25° C. Abundant, slightly raised, confluent, greyish-brown growth, with a dry, dull, worm-eaten surface.

**Loeffler's Serum.**—5 days at 25° C. Good, confluent, raised, yellowish-white growth, with wrinkled surface. 14 days, partial digestion.

**Metalloids.**—Strict aerobe; no growth under anaerobic conditions. Yellow pigment formed on agar, soluble in alcohol, ether, and chloroform, but insoluble in water. Optimum temperature for growth 25-30° C.

**Biochemical Characters.**—Acid in glucose, maltose, mannitol, and sucrose. LM clot, peptonization, and partial decoloration in 3 days. Ind. —. M.P. —. V.P. —. Nitrates reduced to nitrites.  $\text{NH}_4\text{Cl}$  —.  $\text{H}_2\text{S}$  —. M.B. reductive —. Catalase —.

**Pathogenicity.**—Nil.

#### *Chromobacterium typhi-farum*

**Synonym.**—*Bacterium typhi farum* (Dresel and Stahl 1928).

**Habitat.**—Air grass, plants. Has been found in normal human faeces and urine.

**Morphology.**—Slender, rod-shaped organism,  $1.3 \mu \times 0.5-0.7 \mu$ ; axis straight, sides parallel, ends rounded, sometimes filaments up to 15-20  $\mu$  in length; arranged singly, in pairs end-to-end or in groups; in fluid media sausage-like aggregations may occur. Bodily mobile when grown at 22° C, but poorly or not at all at 37° C. Flagella peritrichate. Non-sporing. Non-encapsulated. Gram negative; occasional bipolar staining. Non-acid fast.



FIG. 133.—*Chromobacterium typhi-farum*

Surface growth on agar plate, showing the peculiar appearance caused by aggregations of the bacteria. One-brover body is clearly visible 24 hours, 37° C.

**Agar Plate.**—24 hours at 37° C. Ready growth of colonies 1-2 mm. in diameter, round, low convex, amorphous, smooth, glistening, opaque, with entire edge, consistency between, emulsifiability easy. Odour or rusty yellow pigment, not diffusing into the medium.

5 days.—Colonies are larger, 2-4 mm., differentiated frequently into a central plateau, often with a granular surface, and a smooth bevelled periphery; edge sometimes crenated.

In the centre of the colonies aggregated masses with radiating extensions of a granular nature, or beaver bodies with a clear-cut margin, may often be seen with the lens by transmitted light. The

granular structures are aggregations of organisms, known to the German workers as *Bakterien-verbänden* or *synplasmata* (FIG. 133). The beaver bodies represent downgrowths of the colony into the medium.

- Agar Slope*—24 hours at 37° C Abundant, confluent, smooth, glistening, yellow, opaque growth, with entire or slightly undulate edge
- 5 per cent Glycerine Agar Slope*—Growth similar to that on agar slope but very mucoid in character
- Agar Shake*—Yellow surface growth with a few small colonies throughout the medium
- Agar Pour Plate*—Deep colonies are biconvex, sometimes with lateral knobs or projections
- Broth*—24 hours at 37° C Uniform turbidity with a powdery white or yellowish deposit easily dispersed on shaking After 5 days, increased turbidity with a fine surface scum or ring
- Horse blood Agar Plate*—No hæmolyms Colonies as on agar plate
- MacConkey's Agar*—24 hours at 37° C Growth hardly perceptible 5 days—Irregularly round colourless colonies with a rough surface and crenated edge
- Loeffler's Serum Slope*—24 hours at 37° C Good, confluent, glistening yellow growth with smooth or slightly contoured surface and undulate edge No liquefaction
- Potato*—3 days at 37° C Abundant confluent, yellow, glistening mucoid growth
- Gelatin Stab*—Filiform growth along line of inoculation After 6-10 days liquefaction begins and progresses till the medium is entirely liquefied in the next 10-15 days Liquefaction is infundibuliform, a yellow surface pellicle forms, and later sinks to the bottom
- Resistance*—Destroyed by 55° C in 10 minutes
- Metabolism*—Aerobic Poor unpigmented growth anaerobically Grows freely at 20-37° C, with optimum nearer to 37° C Pigment ochre or rusty yellow insoluble in water and chloroform, partly soluble in alcohol and ether
- Biochemical Reactions*—Acid in glucose, mannitol sucrose, salicin rhamnose arabinose, xylose, maltose usually later Lactose, inositol and dulcitol not fermented Litmus milk neutral or transient acidity, becoming alkaline after 3-7 days, occasional soft clot Indole—MR + VP—Nitrates reduced Very slight H<sub>2</sub>S production at 22° C Catalase +
- Antigenic Structure*—The organisms are antigenically heterogeneous Both as regards flagellar and somatic antigens, sera prepared against any one strain will agglutinate the homologous strain and usually several other strains There is a tendency for the organisms to fall roughly into antigenically similar groups, the H antigens being more cosmopolitan than the O
- Pathogenicity*—It has been suggested that the organism is a variant of *Salmonella typhi*, of potential pathogenicity to man but this has not been substantiated Pathogenic to mice only on injection of enormous doses

There are several organisms in water and in soil belonging to this group, amongst which may be mentioned

*Chr ochraceum*—Motile by polar flagella Infundibuliform liquefaction in gelatin with a pale yellow, later ochre coloured, deposit On agar and potato a thin, ochre-yellow streak.

*Chr fuscum*.—Non motile, liquefies gelatin slowly or not at all On agar and potato gives a thick, wrinkled, chrome-yellow growth.

*Chr aurantiaceum*—Motile by peritrichate flagella No liquefaction of gelatin On agar and potato forms a light orange growth

*Chr denitrificans*—Described by Burri and Stutzer (1895) who isolated it from horse faeces, called by them *B denitrificans* I. Appears to be common in the soil Rods with rounded ends, 1.5-2.5  $\mu$   $\times$  0.75  $\mu$ . Actively motile Gram negative Grows freely on ordinary media, more quickly at 37° C than at room temperature, and is aerobic Colonies on agar are very thin and membranous, having a thicker centre, generally circular, but may have a lobate or irregularly erose edge. In broth there is a dense turbidity in 24 hours, with a reddish white deposit disintegrating on shaking, surface

ring growth Produces large amount of gas in nitrate broth, reducing the nitrate to free nitrogen. Non pathogenic This organism is not to be confused with *Ps denitrificans* (q v Chapter 21)

## ACHROMOBACTERIUM

### DEFINITION—*Achromobacterium*

Motile or non motile, Gram negative rods, usually small to medium in size, forming no pigment on agar, and varying in their fermentative ability Optimum temperature for growth about 25° C, but often good growth at 37° C Saprophytic; commonly found in water, soil, and milk.

Organisms of this group are widespread in nature, but have so far received little systematic study The public health bacteriologist meets them mainly in the analysis of water, milk, food, and soil, where they attract attention by their frequency on agar and gelatin plates incubated at 22° C or sometimes at 37° C They are mainly responsible for the formation of slime on stored meat (Haines 1933) They are differentiated from the *Chromobacterium* group mainly by their failure to form pigment Discussion of their classification would at present serve no useful purpose, and all that we need do here is to give a brief account of their more common characteristics

*Morphologically* they are Gram negative, motile or non motile rods, often of the size of cocciform bacilli, but varying considerably in thickness, rather fat rods, and fat cocco bacilli are quite common

The optimum temperature for growth is about 20–25° C At 37° C growth is generally stated to be slight or absent, but in our experience abundant growth of these organisms is by no means infrequent in cultures made from certain foods Most strains grow at 0° C (Coyne 1933), Haines (1933) found that the generation time in broth at this temperature was about 9 hours The colonial appearances vary After 24 hours at 22° C colonies on agar are about 0.5 mm in diameter, circular, smooth, convex, greyish white, and translucent with an entire edge, after 5 days they are 3–5 mm in diameter, raised or low convex, greyish, opaque, with a smooth surface and entire edge, or sometimes with a beaten-copper surface and an irregular edge Mucoid colonies are not uncommon, and seem to be formed most frequently by the short fat cocco-bacillary type Colonies with central crateriform depressions, draughtsman like colonies, colonies with a roughish surface, and colonies showing radial striation are sometimes met with An aromatic odour may be noticeable

In broth there is a uniform turbidity of varying degree with a powdery, granular, or viscous deposit On potato a layer of growth is formed, which is sometimes mucoid or creamy, and which often takes on a café au lait appearance after a week or two

Most, but not all, strains seem to grow in MacConkey's bile-salt medium In liquid MacConkey they give rise to turbidity, but not usually to acid production; on solid MacConkey they form small yellowish colonies, which after 5 days at 22° C may reach a diameter of 1–2 mm Their growth is not inhibited by concentrations of brilliant green and sodium tetrathionate, such as are used in the isolation of *Salmonella* organisms and as they form non lactose fermenting colonies on MacConkey's medium, they may cause trouble in the search for pathogenic organ

isms in foods such as synthetic cream and made up meat dishes suspected of being responsible for conveying food poisoning or enteric fever

Litmus milk may be unchanged. More often it is turned alkaline or peptonized. Some strains produce acid and curdle the milk. The litmus is often reduced to within a few millimetres of the surface.

Sugar reactions are variable. Most strains have little or no fermentative ability but some produce acid in glucose or in glucose, maltose, mannitol, sucrose and salicin, less often in lactose. There is no record of gas production. Some strains give a positive MR or VP reaction or both. Nitrates are not uncommonly reduced to nitrites but indole is rarely formed. Haines (1933) who studied 132 strains isolated from meat found that 67 strains fermented glucose and maltose, 59 fermented glucose only, whereas 6 failed to ferment even glucose. Gelatin may or may not be liquefied. In Haines's series 115 strains liquefied gelatin in 1 to 2 days and 17 strains liquefied it slowly or not at all.

Little or nothing is known of their antigenic characteristics. It is probable that they are all non pathogenic.

#### REFERENCES

- AMARO T. H. (1930) *Zbl. Bakt.* **116**, 494-499.  
 BORDET P. (1930) *Ann. Inst. Pasteur* **45**, 26.  
 BREED R. S. and BREED M. E. (1924) *J. Bact.* **9**, 545.  
 BURRI R. and STUTZER A. (1895) *Zbl. Bakt. IIte Abt.* **1**, 257-350.  
 COYNE F. P. (1933) *Proc. roy. Soc. B* **113**, 196.  
 CRUICKSHANK, J. C. (1935) *J. Hyg. Camb.* **35**, 354.  
 DRESEL E. G. and STICKL O. (1928) *Dtsch. med. Woch.* **54**, 517.  
 EISENBERG P. (1914) *Zbl. Bakt.* **73**, 466.  
 FRANKLAND G. C. and FRANKLAND P. F. (1889) *Z. Hyg. InfektKr.* **6**, 373.  
 HAINES R. B. (1933) *J. Hyg. Camb.* **33**, 175.  
 HEFFERAN M. (1904) *Zbl. Bakt. IIte Abt.* **8**, 311, 397, 456, 520.  
 KLEIN E. (1894) *J. Path. Bact.* **2**, 217.  
 KUNTZE W. (1900) *Z. Hyg. InfektKr.* **34**, 169.  
 Report (1920) *Rep. Comm. Amer. Bacteriologists* *J. Bact.* **5**, 191.  
 SCHÜTZ F. and LAUN H. (1933) *Zbl. Bakt.* **129**, 124.  
 SULLIVAN M. X. (1905-8) *J. med. Res.* **14**, 109.  
 WREDE F. (1930) *Z. Hyg. InfektKr.* **111**, 531.



## CHAPTER 27

### PROTEUS AND ZOPFIUS

#### PROTEUS

##### DEFINITION—*Proteus*

Highly pleomorphic rods, filaments and curved cells being common in young cultures Gram negative Actively motile Characteristic spreading growth on moist media Often liquefy gelatin and often produce vigorous decomposition of proteins Ferment glucose and usually sucrose but not mannitol or lactose with production of acid and gas

Type species *P. vulgaris* Hauser

**Habitat**—Organisms of the *Proteus* group have been known since the earliest days of bacteriology They are widely distributed in nature, and constitute an important part of the flora of decomposing organic matter of animal origin They are constantly present in rotten meat and in sewage, and very frequently in manure. Though often demonstrable in the faeces of man and animals, they are rarely found in large numbers except when the normal intestinal mechanism is deranged They are not uncommon in garden soil and on certain vegetables, such as melons and celery (Cantu 1911), but it seems probable that their access to these materials results largely from contamination with sewage or manure

Besides their wide saprophytic existence, *Proteus* bacilli are able under certain conditions to grow in the animal body and even to give rise to pathological disturbances The role they play in summer diarrhoea is not yet entirely clear, but there is no doubt that in some outbreaks of this disease they multiply enormously in the intestinal canal, particularly of infants This holds particularly true of Morgan's bacillus which in the light of Rauss's (1936) recent work, must be regarded as belonging to the *Proteus* group They are primarily responsible for some cases of cystitis, and they are to be met with as secondary invaders in infections of the bladder and in wounds Many strains, referred to as *Proteus X* strains, have been isolated from the urine, faeces, or blood of patients suffering from typhus fever, though their exact relationship to the aetiological agent of this disease is still obscure (see Chapter 83)

**Morphology**—The organisms are rod shaped, but are subject to great variation in size In agar cultures after 24–48 hours, the majority are of the coliform type, 1–3  $\mu$  long by 0.4–0.6  $\mu$  wide, though short fat cocco-bacillary forms are not uncommon (Figs. 136–137) In young rapidly growing cultures, however, in which swarming (see p. 643) is apparent, many of the organisms are long, curved, and filamentous reaching 10, 20, or even 30  $\mu$  in length (Fig. 135) There is no very characteristic arrangement, the bacilli are distributed singly, in pairs, in short chains, in small bundles, or in larger bundles in which the members tend to be arranged concentrically, more or less simulating the isobars in a diagram of a

**cyclone** There is some variation in depth of staining. Except for non flagellated O variants, all members are actively motile by peritrichate flagella in young cultures. Neither spores nor capsules are formed. The reaction to Gram's stain is uniformly negative.

**Cultural Characters**—Growth occurs freely on the usual media. One of the most characteristic properties of *Proteus* strains is their ability to 'swarm' on solid media. Cantu (1911) who made a study of this feature, found that if an organism of the *Proteus* group was inoculated into the water of condensation of an agar slope a rapid growth occurred which spread over the whole surface, producing a uniform layer hardly distinguishable from the medium. This process has been more fully described by Moltke (1927, 1929). Swarming may be defined as a progressive surface spreading by the microbes from the edge of the parent colony. It is best studied by touching the centre of an agar plate with a needle dipped in a *Proteus* culture. First of all a colony develops and then after about 6 hours at 37° C a thin, effuse, ground glass type of growth appears round the edge of the colony, and rapidly spreads over the whole plate (Fig 134). If it is examined under the microscope it is seen that, when swarming commences long



FIG 134—*Proteus vulgaris*  
showing swarming on agar 6  
hours 37° C ( $\times 8$ )



FIG 135—*Proteus vulgaris*  
From an agar culture 6 hours 37° C showing  
long filaments ( $\times 1000$ )

(Figs 136 and 137) The property of swarming is observed only on the surface of solid media. In the depths of an agar shake or pour plate culture the colonies are compact. This circumstance may be made use of in the isolation of organisms mixed with *Proteus bacilli* (Fry 1932). Other methods for the inhibition

slender rods in continuous motion break away from the periphery of the colony and after travelling some distance from the parent colony join neighbouring lateral offshoots to form arches which are rapidly filled with other rods from within. Whole rafts of rods tear loose from the peninsula so formed and work across the agar so that in a short time the colony is surrounded by an archipelago of islands and solitary organisms all constantly in motion. The very long rod forms are the predominant feature in the picture: they form arches, islands, spirals and question mark forms (Fig 135). Once the plate is completely covered with swimmers the long rods are replaced by quite short forms

of swarming consist in (a) the addition to the medium of certain narcotic drugs such as chloral hydrate, morphine and sodium phenylethylbarbiturate (Kramer and Koch 1931 Lode and Howard 1932), chloral should be employed in a final concentration of 1/500-1/1000, this method suffers from the disadvantage that many strains are relatively insensitive to the drugs used, (b) the incorporation of 5-6 per cent alcohol in the medium (Floyd and Dack 1939), this method is effective, but if used for the isolation of streptococci has the drawback of lysing the blood in the agar and inhibiting the growth of the streptococci, (c) the addition to the medium of sodium azide in a final concentration of 1/5000-1/10 000 (Snyder and Lichstein 1940, Lichstein and Snyder 1941), in the higher concentration not only is spreading prevented, but the growth of *Proteus* is almost completely inhibited the growth of streptococci is good, though the zone of haemolysis around  $\beta$  lytic colonies appears green and around  $\alpha$  lytic colonies brown, (d) the use of 6 per cent agar (Hayward and Miles 1943) Swarming is also suppressed



FIG 136—*Proteus vulgaris*

From an agar culture 24 hours, 37° C  
showing chiefly rod forms ( $\times 1000$ )



FIG 137—*Proteus vulgaris*

From an agar culture, 48 hours, 37° C.,  
showing short rods only ( $\times 1000$ )

on some of the media, such as Wilson and Blair's bismuth sulphite agar and Leifson's desoxycholate citrate agar, used for the isolation of the pathogenic intestinal Gram negative bacilli. According to Lominski and Lendrum (1942) some 'surface active agents,' like the alkyl sulphates, have a strong anti-swarming power<sup>1</sup>

Swarming is due essentially to the active motility of the bacilli. Non motile O variants give rise to compact colonies which according to Felix (1922), may be of three types (1) smooth, translucent, homogeneous, with an entire edge, (2) granular and opaque with an irregular edge, (3) tiny colonies, barely visible to the naked eye after 24 hours.

Observation clearly shows that swarming on an agar plate is a discontinuous phenomenon. On a plate inoculated centrally and incubated at 37° C, a thin layer of bacteria is present at the site of inoculation after about 4 hours. Swarming then begins, and in 6 hours the breadth of the growth is 1-1.5 cm. Further progress outwards ceases but the layer of growth becomes thicker. After 8 hours swarming again starts, and a fresh ring of growth appears. The alternation of swarming and

<sup>1</sup> Certain quinone compounds have also been found to be effective in suppressing the growth of *Proteus* (see p 1578)

rest occurs regularly, a fresh ring of growth being formed about every 4 hours till the plate is covered. It is due to this periodic extension that the surface of the growth appears rippled or contoured (see Russ Münzer (1935)).

Though the property of swarming is generally considered to be peculiar among aerobic bacteria to the *Proteus* group it has been pointed out by Rauss (1936) that strains of Morgan's (1906) bacillus, which have not hitherto been included in this group, also exhibit the ability to swarm under appropriate conditions. On ordinary agar at 37° C this organism forms circumscribed colonies, but on solid media containing only 1 per cent agar incubated at 20–28° C, characteristic swarming occurs. Variant forms are described having a less marked power to spread, and these give rise to colonies distinguished by varying degrees of peripheral spread as well as by their structural appearance.

In broth *Proteus* gives rise to a uniform turbidity accompanied by a slight to moderate powdery deposit and a faint ammoniacal smell. Cantu (1911) states that a surface pellicle is never formed, while Wenner and Rettger (1919) and Jacob (1932) say that a thin fragile pellicle may develop in older cultures. Gelatin plate colonies are very characteristic (Cantu 1911).

**Resistance**—Few observations appear to have been made on the resistance of *Proteus* bacilli. Our own limited experience has shown that they are readily destroyed by heat and disinfectants. Exposure to moist heat at 60° C for 1 hour is sufficient to sterilize a broth culture. A 1 per cent phenol solution inoculated with one million organisms per ml, is found to be sterile within 30 minutes.

**Metabolism**—The members of this group are aerobes and facultative anaerobes. Growth under strict anaerobic conditions is very poor, and certain enzymic activities may be suppressed. The optimum temperature for growth is about 34–37° C, though rapid multiplication occurs above 20° C. The limits of growth are between about 10° C and 43° C.

Reports on the hæmolytic activity of *Proteus* are discrepant. Wenner and Rettger (1919) obtained uniformly negative, and Norton, Verder and Ridgway (1928) uniformly positive results. In neither of these reports is the type of blood mentioned. Taylor (1928), using human blood, observed hæmolysis regularly within 24 hours in 1 per cent blood broth, but not on 10 per cent blood agar plates. Jacob (1933) used 5 per cent rabbit blood agar plates, and found that all strains produced  $\beta$  hæmolysis in 24–48 hours. As with many other organisms it seems probable that the nature of the blood is an important factor in determining the result.

The main features that distinguish *Proteus* bacilli from other Gram negative gelatin liquefying rods are the production of  $H_2S$  and the active decomposition of urea (Moltke 1927). According to Wolf (1918–19), urea is broken down readily as much as 45 per cent of the total nitrogen of urine being transformed into ammonia. The production of indole and the digestion of serum proteins varies with different strains, as a rule these two properties are negatively correlated. The power to digest serum proteins is often lost during cultivation in the laboratory. Catalase is formed, but the oxidase reaction described by Gordon and McLeod (1928) is negative.

Since the *Proteus* bacilli are found most constantly in decomposing animal matter, they are generally regarded as putrefactive organisms. Rettger and Newell (1912–13) dispute this. They define putrefaction as a particular process of protein decomposition which is brought about through the agency of bacteria with the evolution of foul smelling

products which are characteristic of ordinary cadaveric decomposition" Amongst these decomposition products they consider mercaptan and the oxy-acids to be of particular significance indole skatole and  $H_2S$  are less characteristic. According to this definition the *Proteus* group would be classed amongst the non putrefactive bacteria. The connotation that Rettger and Newell attach to the term putrefaction is that it is essentially an anaerobic process it is dependent therefore on the anaerobic growth of bacteria. *Proteus* bacilli, when grown under anaerobic conditions do not digest proteins, and therefore cannot be regarded as capable of causing true putrefaction. They are however frequently associated with the anaerobes in putrefying organic material and no doubt assist these greatly by using up oxygen and rendering the conditions suitable for their growth. It may be noted that this definition would not meet with universal acceptance.

**Biochemical Reactions**—Acid and gas are formed from glucose sucrose, glycerol xylose and almost always from salicin Mannitol, lactose, dulcitol starch, dextrin sorbitol and raffinose are never fermented. The action on maltose is variable and is of value in classification Moltke (1927) who examined 194 strains found that 37 fermented maltose, and 157 did not. The maltose positive strains fermented sucrose and salicin within 24 hours, while the maltose negative strains took 3–15 days to ferment sucrose and 10–21 days to ferment salicin. All but one of the maltose positive strains produced indole, while the maltose-negative strains uniformly failed to do so. A negative correlation was observed between the fermentation of maltose and the ability to digest coagulated horse serum. In the definition of the group by Winslow and his colleagues (1920) it is stated that the gas produced from glucose and sucrose consists entirely of  $CO_2$ . Mendel (1911) however gives the gas ratio as  $H_2$ ,  $CO_2$  = 6·8 : 1, while Jacob (1932) gives it as 3·4 : 1. These latter figures agree with our own findings. The fermentative activity of *Proteus* is fairly constant, but occasional strains are anaerogenic on isolation (Edwards 1912) and others lose their power to attack certain sugars on prolonged cultivation. *Proteus morganii* is restricted in its fermentative power from the start. The action on litmus milk is subject to some variation. Usually a true rennet clot is produced which retracts and squeezes out whey. Digestion of the clot sets in and is accompanied by progressive alkalinization of the medium. The litmus is reduced. The clot is often digested completely within a week, and the reaction is strongly alkaline. Slight initial acidity is sometimes observed, and not infrequently proteolysis may overshadow coagulation so that there is not time for a definite clot to form. Most strains attack milk rapidly, imparting to it a yellow colour a bitter soapy flavour and a foul odour but some strains have little or no coagulative or proteolytic effect (Plahn 1937). The reaction to the methyl red and Voges Proskauer tests varies with different strains. Nitrates are reduced to nitrites. Methylene blue is decolorized slowly in broth cultures. Urea is decomposed with the production of ammonia—a reaction that may be used in the differentiation of *Proteus* from *Salmonella* (Ferguson and Hook 1943). Phenylalanine is broken down with the formation of phenylpyruvic acid (Henriksen and Closs 1938).

**Antigenic Structure**—Our knowledge is both deficient and conflicting. Most workers (Cantu 1911 Wenner and Rettger 1919 Taylor 1923) who have prepared immune sera against different strains have found that a given serum may agglutinate either the homologous strain only or a number of heterologous strains as well. Any simple subdivision by agglutination or absorption of agglutinins has proved impossible. Moltke (1927) who paid special attention to the H and O antigens

found that by direct agglutination the swarming strains could be divided into 3 main groups and that by absorption these main groups could be divided into a number of sub groups. The non swarming strains differed from the swarming strains in their absence of an H antigen and differed among themselves in the type of their O antigen. The relation of the *vulgans* to the Y strains is still somewhat doubtful. The O antigens of the X strains differ from those of the *vulgans* strains but there appears to be a certain group relationship between the H antigens (Yacob 1932). Among the O antigens of the X strains there are at present three fairly well-defined groups represented by the strains OX 2 OX 19 and OY K (see Chapter 83). White (1933) has brought evidence to show that the O antigen of OX 19 contains two receptors one of which is alkali labile and is mainly responsible for agglutination of this organism by an antiserum prepared against it the other of which is alkali stable and is responsible for the reaction of the bacillus with the sera of patients suffering from typhus fever—the Weil Felix reaction. Meisel and Mikulaszek (1933) and Castaneda (1934 1935) have reported the extraction of soluble specific polysaccharide substances from *Proteus* X strains. Castaneda's results agree closely with those obtained by White. They show that the alkali stable polysaccharide referred to as X is common to both *Proteus* X 19 and *Rickettsia prowazekii* (see Chapter 39) while the alkali labile polysaccharide referred to as P is specific to *Proteus* Y 19.

Our own observations on a limited number of strains suggest that there is a wide variety of H and O antigens in *Proteus vulgaris* the H antigens being distributed to some extent independently of the O antigens.

With regard to *Proteus morganii* Rauss states that the H antigen tends to be group-specific and the O antigen type specific. In an examination of 48 strains 7 types of H antigen were differentiated but as many as 17 types of O antigen. A relationship was found between the H receptor of one group of *morganii* and a strain of *Proteus vulgaris*.

**Pathogenicity**—*Proteus* bacilli are frequently found in and appear to be responsible for a number of inflammatory and suppurative conditions in man. They are a very common cause of cystitis and may be isolated in pure culture from the urine of infected patients. They are not uncommonly found in abscesses either alone or in combination with other organisms. Metchnikoff and his co workers (see Chapter 71) found them almost constantly in the faeces of infants with summer diarrhoea. They are frequently present usually as secondary invaders in wounds and burns where they probably favour the development of the pathogenic anaerobes. And they have been isolated from a variety of conditions such as volvulus peritonitis croupous pneumonia acute gastro-enteritis of the food poisoning type (Wichels and Barner 1925 Plahn 1937 Cooper et al 1941) empyema gangrene of the lung and septicæmia. Jensen (1913) considers them responsible for one form of epidemic calf dysentery and Wynn (1898) has encountered them in an epidemic disease of fish in Lake Zurich. They are also responsible for the black rot of eggs.

An organism described as *P. hydrophilus* has been held responsible for the Red leg disease of frogs but as this organism is monotrichate does not swarm on agar and does not decompose urea it is very doubtful whether it should be included in the *Proteus* group (see Kulp and Borden 1949).

The relation of *Proteus* Y 19 to typhus fever is discussed in Chapter 83.

Their pathogenicity to laboratory animals is variable. virulent strains on introduction into the tissues are able to proliferate and invade the blood stream.

Strains of lower virulence cause chronic inflammatory processes, either of the suppurative or of the infective granuloma type (Larson and Bell 1913, Wenner and Rettger 1919), the latter are best seen after intraperitoneal injection

The inoculation intraperitoneally of 0.5-1.0 ml. of a 24 hour broth culture of a virulent strain generally proves fatal to rats and mice in 18-48 hours, and to guinea pigs and rabbits in 1-7 days. Severe infections in rabbits are said to be characterized by extreme emaciation (Larson and Bell 1913). Though invasion of the tissues may occur on parenteral inoculation, Jensen (1913) states that in naturally infected calves, even when the organisms are seething in the gut, the blood and tissues remain sterile

**Classification**—Hauser (1885) divided the *Proteus* group into three species (1) *Proteus vulgaris*, Gram negative, liquefies gelatin, peptonizes fibrin, produces indole and has a variable action on glucose and sucrose (2) *Proteus mirabilis*, Gram negative, more highly pleomorphic, and liquefies gelatin more slowly (3) *Proteus zenkeri*, Gram positive, does not liquefy gelatin, does not form indole, and fails to attack sugars. *Proteus zenkeri* was found to be very similar to an organism described two years previously by Kurth (1883) under the name of *B. zopfii*, as both of these organisms are Gram positive, they had to be transferred to a separate genus, which is known as *Zopfius*

Hauser's subdivision of the *Proteus* group on the basis of morphology, rate of liquefaction of gelatin, and indole production, has been found impracticable. The morphology is variable, depending especially on the medium and the age of the culture. The rate of liquefaction of gelatin is likewise variable, it is rapid with newly isolated strains, and is often much slower, or even absent, with strains that have been long under artificial cultivation. Indole production used to be tested by the nitroso indole reaction, but as Berthelot (1914) has shown, this reaction is untrustworthy, and is given by indolacetic acid as well as by indole, when tested by Ehrlich's reagent, using the ether extraction method, it is found that the results are different, and that indole production is not nearly so constant a feature of *Proteus* as it was originally considered to be

The most striking characteristics of the members of this group are their ability to swarm on solid media, their production of  $H_2S$ , their decomposition of urea, their liquefaction of gelatin, and their failure to ferment lactose or any of the polyhydric alcohols. It should, however, be added that O variants occur which have lost their power of swarming, and that old strains may no longer liquefy gelatin

The recent observations of Rauss (1936) suggest very strongly that Morgan's bacillus which has hitherto occupied an invidious position in the *Salmonella* group, is closely related to *Proteus*. Its ability to swarm under suitable conditions, its frequent fermentation of xylose and its occasional fermentation of sucrose, its production of indole and  $H_2S$ , its formation of alkali in litmus milk, the greater group specificity of its H and the greater type specificity of its O antigens, the group relationship of at least one of its H antigens to *Proteus*, its growth only under favourable conditions in the intestine of human beings, and its general pathogenicity for experimental animals—all bring it closely into line with organisms of the *Proteus* group. Its failure to liquefy gelatin or constantly to ferment sucrose must be considered in relation to the negative reactions obtained in these two respects with known *Proteus* strains, particularly those that have been long cultivated in the laboratory. Intermediate types that peptonize milk, and the organisms described by Magath (1928), which were isolated from cystitis and which liquefied gelatin but

did not ferment sucrose, seem to show that too much stress should not be laid on any single biochemical characteristic. Rauss's conclusions are supported by the observations of Henniksen and Closs (1938), who found that both *Proteus* and Morgan's bacillus were able to break down phenylalanine with the formation of phenylpyruvic acid.

Until a more thorough study has been made, we consider that the most convenient method of classifying the *Proteus* group is on the basis of maltose fermentation and gelatin liquefaction, though we hesitate to ascribe specific names to the members of the sub groups. The X strains belong mostly to the maltose-positive sub group, but the Kingsbury strain differs in this respect, as well as in its failure to liquefy gelatin.

(For review of the *Proteus* group, see Cantu 1911, Berthelot 1914, Wenner and Rettger 1919, Besson and Ehringer 1923, Moltke 1927, Taylor 1928, Jacob 1932, Mello 1938)

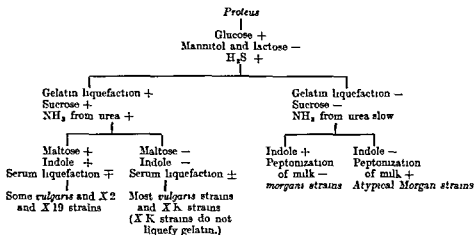


FIG. 138

This diagram must be regarded as merely tentative and subject to both exceptions and alterations.

### Proteus vulgaris

*Synonym.*—*B. proteus vulgaris*

*Isolation.*—By Hauser in 1885 from putrefying material.

*Habitat.*—Putrefying animal and vegetable matter, often in faeces, soil and infected wounds.

*Morphology.*—Straight or slightly curved rods,  $1.0-2.5 \mu \times 0.4-0.6 \mu$ , with parallel sides and rounded ends, arranged singly, in pairs end to end, and in short chains. In young swarming cultures, long curved filamentous forms are common. Considerable variation in length, ovoid forms in pairs may be seen, and in old cultures large bloated forms. Staining is fairly uniform, though variations in depth occur. Actively motile by numerous peritrichate flagella though slightly motile forms with 4 flagella, two at each end and non motile forms devoid of flagella may occur. Gram negative.

*Agar Plates.*—24 hours,  $37^{\circ} C$ . The whole plate is covered with a slightly raised layer of growth, which, but for a faint rippling or contouring of the surface and a marked



odour is easily overlooked. Sometimes indefinite primary colonies are seen, of variable diameter having a smooth or slightly ringed draught-man-like surface and an entire edge. The whole growth is translucent of the same colour as the medium butyrous, and easily emulsifiable. The complete layer of growth over the whole plate is due to swarming of the bacilli (see text). Non flagellated O forms give rise to compact colonies.

*Agar Slope*.—If the organisms are inoculated into the condensation water they swarm rapidly and in about 8 hours at 3 ° C. form a uniform, slightly raised, translucent growth with a glistening faintly contoured surface over the whole slope. There is a thick turbid growth in the water of condensation itself.

*Gelatin Stab*.—24 hours 22° C. Good filiform growth, consisting of discrete and confluent colonies, extending to the bottom of the tube. Smooth raised surface growth 2 or 3 mm. in diameter. Crateriform liquefaction is generally visible after 24 hours in newly isolated strains. Liquefaction later becomes stratiform and is complete in 4 to 6 days. With old laboratory strains liquefaction is slower and may not be complete for 3 weeks. Sometimes the power to liquefy gelatin is lost altogether. Occasionally very fine tangled branches grow out from the filiform stab. Sometimes the liquefaction is infundibuliform or saccate.

*Broth*.—24 hours 3° C. Moderate growth with a slight to moderate uniform turbidity and a moderate powdery deposit disintegrating completely on shaking. No surface growth. The growth increases only slightly on further incubation.

*Glucose Agar Shake*.—24 hours 3° C. Profuse growth of tiny colonies throughout medium, and layer of growth over whole surface. Numerous bubbles of gas throughout medium sometimes blowing the agar up to the plug.

*Horse Blood Agar Plates*.—24 hours 3° C. Uniform growth over whole surface with indefinite single colonies. The blood is cleared, translucent and of a slightly brownish colour.  $\beta$  haemolysis on 5 per cent. rabbit blood agar plates.

*MacConkey Plates*.—24 hours 3° C. Good growth of colourless, discrete or partly confluent colonies. The colonies may be smooth, but more often have a slightly roughish surface with an irregularly crenated, radially striated edge.

*Loeffler's Serum*.—Spreading growth over whole surface. On further incubation the serum is liquefied partly or completely but this power of liquefaction is confined chiefly to maltose-negative strains. Newly isolated strains are more active liquefiers than old laboratory strains.

*Dorset Egg*.—24 hours 3° C. Spreading growth over whole surface. On further incubation digestion occurs, but does not usually proceed to completion.

*Cooked Meat Medium*.—5 days 3° C. Good growth with some bubbles of gas. No blackening or visible digestion occurs.

*Potato*.—5 days 3 ° C. Raised, confluent glistening, greyish-brown growth. The potato itself takes on a café-au-lait colour.

*Resistance*.—Not specially resistant. Killed by moist heat at 50° C. in 1 hour.

*Metabolism*.—Aerobe and facultative anaerobe. Growth under anaerobic conditions is poor only a very thin, effuse barely visible growth is formed on agar in 4 days at 3 ° C. No digestion of protein media occurs under anaerobic conditions. Optimum temperature for growth 34°–3° C. Good growth occurs at 20° C. A haemolysin is formed, acting on rabbit blood. No pigment formed, except the café-au-lait pigment on potato. Growth is improved by the addition of glucose and of nitrates. No soluble toxin formed.

*Biochemical*.—All strains produce acid and gas in glucose galactose glycerol, and sucrose. Nearly all strains ferment salicin, and some ferment maltose. Lactose mannitol, and mannose are never attacked. Old laboratory strains may lose their power of fermenting sucrose. *Litmus mull.* Alkaline some strains coagulate the casein

and then digest it, others digest it without preliminary coagulation, the litmus is reduced. Indole is produced by the maltose-fermenting, but not by the maltose-negative strains.  $H_2S$  + +  $NH_3$  + +. Catalase + +. Methylene blue reduction +. Nitrates reduced to nitrites. MR + VP +, if Barritt's method is used. Urea is decomposed with the formation of  $NH_3$ .

*Antigenic Structure*—Incompletely worked out. By direct agglutination the swarming strains can be divided into 3 main groups, but several smaller groups are present, by absorption the main groups can be divided into sub-groups. No apparent relationship between the serological and the biochemical grouping. The O antigens tend to be type specific. Among the X strains the OX 2, OX 19, and OX K strains are distinct.

*Pathogenicity*—Produces no specific infection under natural conditions, but is frequently found in erythema, infantile diarrhoea, and suppurative lesions generally. Is probably responsible for one form of calf dysentery. Virulence to laboratory animals is variable. Highly virulent cultures inoculated intraperitoneally into rabbits, rats, or guinea pigs cause death in a few hours, presumably from toxæmia. Less virulent cultures cause emaciation with death in a week or more after intraperitoneal inoculation, and abscesses and inflammation lasting for months after subcutaneous inoculation. In fatal cases the organisms can generally be recovered from the blood and viscera.

*Proteus morgani* was isolated by Morgan (1906) from the stools of patients with summer diarrhoea. It is motile by 25-30 peritrichate flagella. Motility is sometimes lost after long cultivation, but it may sometimes be restored by passage through broth at 20° C. Though not swarming at 37° C. on ordinary agar it swarms readily at 20-25° C. on 1 per cent agar. Variant types, however, occur which are less actively motile, and which give rise to characteristic streaming colonies. The general cultural characters resemble those of the coliform group. Acid and a small amount of gas are produced in glucose peptone water within 24 hours. Xylose is often fermented with the production of acid only, while occasional strains are said to produce acid and a small amount of gas in sucrose after 10 days. Gelatin is not liquefied, but both indole and  $H_2S$  are formed abundantly. Litmus milk is turned alkaline. About 30 per cent of strains give rise to a hæmolyxin for sheep red cells. Antigenically, most workers (Lewis 1911-12, Kligler 1919, Thorjatta 1920, Jordan, Crawford, and McBroom 1935) including ourselves, have noted the extraordinary heterogeneity of members of this species. Rauss (1936) who has made a careful study of this question, finds that the H antigen tends to be group specific and the O antigen type-specific. Seven H receptors and 17 O receptors were differentiated in 49 strains. One of the H antigens in *P. morgani* is similar to one of the H antigens in *P. vulgaris*. The organism seems to be mainly parasitic and potentially pathogenic, assuming a considerable rôle in some outbreaks of infantile diarrhoea. It has been isolated from paratyphoid like fevers (Havens and Mayfield 1930). Infections in birds, mammals, and reptiles are not uncommon (Lovell 1929), while in mice it may give rise to spontaneous epidemics of enteritis (Wilson 1927), especially in the late summer and autumn months. Experimentally, it produces a rapidly fatal infection in mice on intraperitoneal inoculation. It does not produce a soluble toxin.

Other types of bacilli were isolated by Morgan (1906-1907) which are sometimes called after him, and which differ from his No. 1 bacillus in their motility, their action on milk, or some other characteristic, but the term Morgan's bacillus is generally used to indicate the organism we have just described.

## ZOPFIUS

DEFINITION—*Zopfius*.

Long rods, occurring in evenly curved chains. Gram positive. Motile. Spider-web growth on solid media. Facultative anaerobes. Carbohydrates and gelatin not attacked. Hydrogen sulphide not formed.

Type species, *Zopfius zopfii* (Kurth) Wenner and Retger isolated originally from the intestinal tract of hens.

Organisms of this group are differentiated in several ways from those of the *Proteus* group. Apart from being Gram-positive, they ferment no carbohydrates, they form no  $H_2S$ , they do not liquefy gelatin, and they do not exhibit the phenomenon of swarming.

Morphologically rods are formed, about  $3.5 \mu$  long by  $0.8 \mu$  broad, having rounded ends and parallel sides, and occurring in long evenly curved chains. Filamentous forms are common. The organisms are motile by peritrichate flagella (Fig. 12 p. 30).

On agar they form small indistinct colonies having on magnification a spider web appearance. Sometimes the colonies are thinnest at the centre and are surrounded by arborescent tufts. In gelatin stab arborescent lateral branches, interlacing freely grow out from the stab. There is a slow moderate growth in broth, with occasionally a thin fragile pellicle. Optimum temperature for growth is  $25^\circ C$ . Growth occurs freely between  $20^\circ$  and  $30^\circ C$ . but is very poor at  $3^\circ C$ . No sugars are fermented, and there is only a scanty growth in litmus milk with no visible change in the medium. On potato there is a moderate growth the medium is darkened. Gelatin, serum, and egg are not digested. No indole or  $H_2S$  is formed, but there is some production of  $NH_3$ .

## REFERENCES

- BETHKELOT, A. (1914) *Ann. Inst. Pasteur* 28, 839-913.  
 BESSON, A. and EHRINGER, G. (1923) *Paris med.*, 1, 22a.  
 CANTU, C. (1911) *Ann. Inst. Pasteur* 25, 852.  
 CASTANEDA, M. R. (1934) *J. exp. Med.*, 60, 119. (1935) *J. exp. Med.*, 62, 289.  
 COOPER, K. E., DAVIES, J. and WISEMAN, J. (1941) *J. Path. Bact.*, 52, 91.  
 EDWARDS, J. L. (1942) *J. Hyg. Camb.* 42, 233.  
 FELIX, A. (1929) *Z. Immunforsch.* 35, 57.  
 FERGUSON, W. W. and HOOK, A. E. (1943) *J. Lab. clin. Med.*, 28, 1-15.  
 FLOYD, T. M. and DACK, G. M. (1939) *J. infect. Dis.* 64, 499.  
 FRY, R. M. (1939) *Brit. J. exp. Path.*, 13, 456.  
 GORDON, J. and McLEOD, J. W. (1933) *J. Path. Bact.*, 31, 185.  
 HAUSER, G. (1885) "Ueber Fäulnisbakterien." *Leipzig*.  
 HAYES, L. C. and MAYFIELD, C. R. (1930) *J. prev. Med.*, 4, 179.  
 HAYWARD, N. J. and MILLS, A. A. (1943) *Lancet* ii, 116.  
 HENRIKSEN, S. D. and CLOSS, E. (1931) *Acta path. microbiol. scand.*, 15, 101.  
 JENSEN, C. O. (1913) "Handbuch der pathogenen Mikroorganismen." Kollé and Wassermann, 9te Aufl., 6, 121.  
 JORDAN, E. O., CRAWFORD, R. R., and McBROOM, J. (1930) *J. Bact.*, 29, 130.  
 KLIGLER, I. J. (1919) *J. exp. Med.*, 29, 531.  
 KRÄMER, E. and KOCH, F. E. (1931) *Z. Bakt.* 120, 452.  
 KUTY, W. L. and BORDEN, D. G. (1942) *J. Bact.*, 44, 673.  
 KURTH, H. (1883) *Bd. Ztg.* 41, 369-393, 409-415.  
 LARSON, W. P. and BELL, E. T. (1913) *J. infect. Dis.* 13, 510.  
 LEWIS, G. J. (1911) 19<sup>th</sup> 41st Ann. Rep. Soc. Govt. Bd. M.O.s Suppl. 465.  
 LICHTSTEIN, H. C. and STYDER, M. L. (1941) *J. Bact.* 42, 63.  
 LOPE, A. and HOWARD, A. (1932) *Zbl. Bakt.*, 124, 533.  
 LOMINCKI, I. and LENDEUM, A. C. (1942) *J. Path. Bact.* 54, 421.  
 LOVELL, P. (1929) *J. Path. Bact.*, 32, 79.  
 MACGATH, T. B. (1929) *J. infect. Dis.* 43, 181.  
 MENDEL, H. and MIKULASZEK, E. (1933) *C. R. Soc. Biol.* 114, 364.  
 MELLO, J. de T. (1935) *Ann. Fac. Med. S. Paulo* 14, 4.  
 MENDEL, J. (1911) *Zbl. Bakt.*, 11te Abt., 29, 290.

- MOLTYE, O. (1927) "Contributions to the characterization and systematic classification of *Escherichia coli* (Haeuser) Levin and Munksgaard" Copenhagen (1929) *Zbl. Bakt.*, 111: 379.
- MORAN, H. DE L. (1906) *J. L. med. J.* 1: 204. (1907) *Ibid.* 11: 16.
- NORTON, J. L., VERDER, L., and RIDGWAY, C. (1928) *J. infect. Dis.* 43, 458.
- PLINY, G. (1917) *J. L. med. J.* 11: 116-117, 96, 196.
- RATON, A. J. (1936) *J. Ind. Bact.*, 42, 183.
- REITZEL, L. F. and NEWELL, C. L. (1917) 13) *J. Biol. Chem.* 13: 341.
- REITZEL, A. (1935) *Zbl. Bakt.* 133, 214.
- REYER, M. L. and LICHTEN, H. C. (1917) *J. infect. Dis.* 67: 113.
- TAYLOR, J. F. (1924) *J. Path. Bact.*, 31: 89.
- TRUETT, T. (1920) *J. Bact.* 5, 67.
- WENNER, J. J. and REITZEL, L. F. (1919) *J. Bact.*, 4, 331.
- WHITE, I. B. (1933) *Brit. J. exp. Path.* 14, 145.
- WICKEL, P. and HARRIS, W. (1925) *Med. Klin.*, 21: 1850.
- WILSON, G. C. (1927) *J. Hyg. Camb.* 26, 170.
- WINSLOW, C. E. A., ISHAI-KHAY, J., BUCHANAN, R. L., KACHWISSE, C., ROGERS, L. A., and SMITH, G. H. (1920) *J. Path. Bact.*, 5, 121.
- WOLF, C. G. I. (1918-19) *J. Path. Bact.*, 22, 459.
- WYLL, G. (1925) *Z. Hyg. Inf. Dis.* 27, 143.
- YACOB, M. (1937) *Indian J. med. Res.*, 19: 787.

## CHAPTER 28

### BACTERIUM

#### DEFINITION—*Bacterium*

Gram negative non sporing rods often motile with peritrichate flagella. Some species capsulated. Easily cultivable on ordinary laboratory media. Aerobic and facultatively anaerobic. All species ferment dextrose with the formation of acid or acid and gas. Many species are active fermenters of a wide range of carbohydrates and allied substrates. Typically intestinal parasites of man and animals, though some species may occur in other parts of the body on plants, or in the soil. Many species are pathogenic.

Type species *Bacterium coli*

#### Classification and Nomenclature

In the past the generic term *Bacterium* has been used to comprise a broad group of Gram negative non sporing rods occurring in the intestinal canal of man and animals and on plants and living either a saprophytic commensal or pathogenic existence. It was early realized that the members which were pathogenic to man and animals differed from most of the non pathogenic forms in failing to ferment lactose. Of recent years the non lactose-fermenting group has been subdivided mainly on the basis of biochemical and antigenic characters into a number of sub groups which have been given the generic names of *Eberthella*, *Salmonella* and *Shigella*. Of these *Eberthella* which comprises the typhoid bacillus is so closely related to the *Salmonella* group that we can see no useful purpose in maintaining its separate identity. We shall therefore deal in the following chapters with the *Salmonella* group which comprises the typhoid paratyphoid and food poisoning organisms and the *Shigella* group containing the dysentery bacilli. A residue of strains fermenting lactose late weakly or not at all and usually regarded as non pathogenic is classified in the relatively indeterminate group of paracolon bacilli and will be considered in the present chapter.

The classification and nomenclature of the lactose-fermenting organisms are subject to wide variation in opinion. On the one hand there is a school led by American workers (see Bergey *et al* 1939) who would do away completely with the genus *Bacterium* and substitute for it a number of genera—*Escherichia*, *Aerobacter* and *Klebsiella*—to include organisms of animal origin, and a single genus—*Erwinia*—to comprise the plant pathogens. On the other hand there is a school represented largely by workers in Great Britain but not without considerable support in the United States which adopts for the present a conservative attitude and prefers to include all these organisms in a single genus—*Bacterium*. With this latter school we would identify ourselves.

In the definition of the genera mentioned above reliance is placed for differential purposes largely on habitat, biochemical characters and pathogenicity.

Of these neither habitat nor pathogenicity though useful adjuncts can be regarded as satisfactory basal criteria for classification. The separation therefore of the group *Erwinia* from other coliform organisms merely on the basis of plant pathogenicity is not sound taxonomical practice. If this property was closely associated with some more valid criterion there would be something to be said for recognizing the *Erwinia* group, but though Elrod (1912) maintains that the secretion of an enzyme protopectinase which is responsible in plants for the production of soft rot is characteristic of the *Erwinia* group it is clear that this is merely another way of saying that these organisms are pathogenic to plants. Elrod's own observations on the biochemical characters of the plant pathogens support those of Dowson (1939) in showing that these organisms cannot be distinguished satisfactorily from coliform organisms of animal origin. Until some more stable differential character is found we prefer therefore to regard the coliform organisms of plant and animal origin as belonging to a single group.

The division of the organisms from animal sources into *Escherichia*, *Aerobacter* and *Klebsiella* is in our opinion hardly more fortunate. The first two groups are morphologically and culturally indistinguishable and their differences in biochemical behaviour though worthy of specific recognition are not of sufficient constancy or importance to serve as criteria for generic differentiation. *Klebsiella* which comprises capsulated organisms found in the respiratory tract is perhaps even less justified. Practically all coliform bacilli seem to be capable of forming capsules under favourable conditions and in our experience there is no way of distinguishing a non motile capsulated Gram negative bacillus in the respiratory tract from a similar organism in the intestinal tract. Neither habitat nor capsule formation is sufficiently constant or distinctive to justify the recognition of the genus *Klebsiella*.

We would plead therefore for the maintenance of the *Bacterium* group as a convenient repository for a wide range of Gram negative non sporing bacilli that cannot be classed at present in either the *Salmonella* or *Shigella* groups. As knowledge increases it is probable that further genera will be split off. But international agreement is essential if the new genera are to receive universal recognition without it they will merely add to the existing confusion.

The strict taxonomist is worried by the fact that the type species *Bacterium triloculare* Ehrenberg 1828 is no longer recognizable and that the genus *Bacterium* is therefore invalid. On the other hand the organism that has for years been regarded by bacteriologists as typical of this genus is the common coliform organism of the intestine *Bacterium coli*. We see no reason why the genus should not be re-defined with this organism as the type species so as to include the numerous Gram negative non sporing bacilli that cannot be assigned to other recognized genera. Whether this genus is admitted as valid by the taxonomists or is accepted temporarily for purposes of convenience (see Breed and Conn 1936) is a matter more of academic than practical importance. Our tentative definition of this genus follows closely that given by the American Committee (Winslow *et al* 1917 1920).

**Morphology**—Neither the shape size structure or arrangement of the bacterial cells nor the appearances presented by cultures on the ordinary solid or liquid media afford any adequate criteria for the differentiation of species within this group.

The modal form of the individual cell is that of a bacillus 2 to 3  $\mu$  in length and 0.6  $\mu$  in breadth with parallel sides and rounded ends (see Fig 139). By the usual methods of examination the cell appears to be almost devoid of internal

is far too irregular and variable a character in the *Bacterium* group to serve as a criterion for specific separation

Some species, such as *Bact friedlanderi*, are normally capsulated, and this character, when it occurs, has some differential value, but those bacilli which are normally capsulated and form mucoid colonies when first isolated from the tissues, frequently lose the property of capsule formation during subculture on artificial media, while other normally non capsulated species may acquire a capsule under particular conditions. It may be noted that many coliform strains isolated from milk are normally capsulated. Most of these strains belong to the intermediate aerogenes cloacae group, but some undoubted strains of *Bact coli* form a capsule, and give rise to a mucoid growth on solid media. It may further be noted that the presence of a capsule is not incompatible with active motility

**Conditions of Growth**—The members of this group grow readily on the ordinary nutrient media of the laboratory, without the addition of any accessory substances

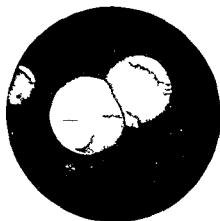


FIG 140—*Bact coli*  
Colonies on agar plate after 24 hours  
( $\times 8$ )



FIG 141—*Bact coli*  
Larger and flatter type of colony on agar  
plate after 24 hours ( $\times 8$ )

They are aerobic, and facultatively anaerobic, though the growth is usually far less copious under the latter conditions. The optimum temperature is for most species, in the neighbourhood of  $37^{\circ}\text{C}$ , and the range over which growth occurs is fairly wide, extending for most species from about  $42^{\circ}\text{C}$  as an upper limit to  $18^{\circ}\text{C}$  or lower

There are, however, certain differences in behaviour that are of significance from the systematic point of view. Thus, *Bact aerogenes* grows very poorly or not at all at a temperature of  $44^{\circ}\text{C}$ , and differs in this respect from the closely related *Bact coli*. Moreover, many strains of *Bact aerogenes* have their optimum growth temperatures nearer  $30^{\circ}\text{C}$  than  $37^{\circ}\text{C}$ .

It happens that the nutritional requirements of several species within this genus and the enzymic mechanisms that they employ in their attack on various substrates have been studied in considerable detail by the methods that have recently been introduced in the investigation of the biochemical activities of bacteria. Many of the results obtained in these studies have been described and discussed in Chapter 3. In the present chapter we may therefore confine our attention to such

structure It stains evenly, it forms no spores, and it shows no granules. It is Gram negative, and non acid fast This modal form is, however, widely departed from as regards the shape and size of the individual cells. Some strains are almost coccoid in form, others show long, sometimes filamentous bacilli. There is a tendency for the cocco-bacillary or the elongated, form to predominate in any single strain, but some cultures show a wide diversity in this respect Cell length is, indeed, a highly variable character in this group, and it is possible, as Barber (1907) has shown, to obtain long-celled strains of *Bact coli* by simple selection of individual cells for successive subculture

Rudimentary branching, with the formation of Y forms, followed by division at each of the three points of the Y, has been described by Hort (1920), and by Gardner (1925), as an occasional happening in some species of *Bacterium*

Many species are motile, other species are non motile By the usual methods of staining the flagella of motile species appear to be numerous and to have a



FIG 139—*Bact coli*  
From 24-hours culture on agar ( $\times 1000$ )

peritrichous arrangement How far this appearance corresponds to reality is doubtful. Pijper (1938) for example, using solar dark ground illumination, finds that the typhoid bacillus—an organism morphologically indistinguishable from *Bact coli*—possesses only two flagella, attached one on each side of the body near the middle They are broadly coiled spiral structures, which, when in action become entwined to form a long tail by which the organism propels itself through the surrounding medium There is evidence that each of these flagella is made up of a number of extremely fine threads but, according to Pijper, there are only two flagella to each organism, with single opposite points of attachment

The peritrichous appearance disclosed by flagella stains is regarded as an artefact How far Pijper's conclusions will be confirmed it is impossible to say Whether there are two flagella or several flagella is not likely to give rise to confusion so long as it is recognized that they are attached to the sides of the rod and not as in *Pseudomonas*, to the end (see Fig 13 p 31)

Before the *Salmonella* and the *Shigella* groups were separated from the *Bacterium* group motility was of some importance in distinguishing for instance, the typhoid and paratyphoid bacilli, which are nearly always motile, from the dysentery bacilli which are consistently non motile In the present *Bacterium* group, however, motility is of far less differential value It is true that some members, like *Bact coli*, are usually motile, and that other members, like *Bact aerogenes*, are usually non motile, but the property of motility or non motility is a characteristic of the individual strain, not of the species as a whole Moreover, motile organisms may give rise to non motile variants, though the reverse phenomenon, of normally non motile species giving rise to motile variants, does not appear to occur Motility



is far too irregular and variable a character in the *Bacterium* group to serve as a criterion for specific separation

Some species, such as *Bact friedlanderi* are normally capsulated and this character, when it occurs has some differential value, but those bacilli which are normally capsulated and form mucoid colonies when first isolated from the tissues, frequently lose the property of capsule formation during subculture on artificial media while other normally non-capsulated species may acquire a capsule under particular conditions. It may be noted that many coliform strains isolated from milk are normally capsulated. Most of these strains belong to the intermediate aerogenes-cloacæ group but some undoubted strains of *Bact coli* form a capsule, and give rise to a mucoid growth on solid media. It may further be noted that the presence of a capsule is not incompatible with active motility

**Conditions of Growth**—The members of this group grow readily on the ordinary nutrient media of the laboratory without the addition of any accessory substances

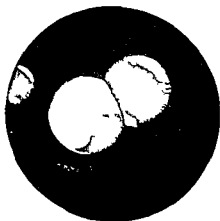


FIG 140—*Bact coli*  
Colonies on agar plate after 24 hours  
( $\times 8$ )



FIG 141—*Bact coli*  
Larger and flatter type of colony on agar  
plate after 24 hours ( $\times 8$ )

They are aerobic, and facultatively anaerobic though the growth is usually far less copious under the latter conditions. The optimum temperature is for most species in the neighbourhood of  $37^{\circ}\text{C}$ , and the range over which growth occurs is fairly wide extending for most species from about  $42^{\circ}\text{C}$  as an upper limit to  $18^{\circ}\text{C}$  or lower

There are, however certain differences in behaviour that are of significance from the systematic point of view. Thus *Bact aerogenes* grows very poorly or not at all at a temperature of  $44^{\circ}\text{C}$  and differs in this respect from the closely related *Bact coli*. Moreover, many strains of *Bact aerogenes* have their optimum growth temperatures nearer  $30^{\circ}\text{C}$  than  $37^{\circ}\text{C}$ .

It happens that the nutritional requirements of several species within this genus and the enzymic mechanisms that they employ in their attack on various substrates have been studied in considerable detail by the methods that have recently been introduced in the investigation of the biochemical activities of bacteria. Many of the results obtained in these studies have been described and discussed in Chapter 3. In the present chapter we may therefore confine our attention to such

reactions as are of value in identifying the different species within the genus or in distinguishing between them.

**Type of Growth**—The type of growth given by the various species within this genus is very similar. When normal smooth strains are grown in broth a uniform turbidity develops increasing rapidly during the first 12 to 18 hours of growth and then more slowly up to 48 to 72 hours. Pellicle formation is rare and when present is very slight. A slight deposit forms as growth increases and this is easily dispersed on shaking the tube.

On agar the colonies are relatively large with an average diameter of 2-3 mm. but vary considerably in size. They may be circular raised and low convex, with an entire edge and smooth surface they may be flatter with a more irregular surface and a more effuse and irregular edge or they may assume the typical vine-leaf form which is commonly described as characteristic of *Salmon typhi*. Even with freshly isolated strains the range of variation is wide and when old laboratory strains are under examination the most varied colonial forms may be seen. Apart from the possible appearance of rough variants a single strain may show several different types of colony if successive subcultures in broth are interspersed with platings and subculture of individual colonies.

As an exception to this general rule we may note that members of the Friedlander group when freshly isolated give rise to typically mucoid colonies. The differential value of this characteristic is diminished by the fact that certain other members particularly those belonging to the intermediate-aerogenes-cloacae group may similarly give rise to this type of growth. It is very common in strains isolated from milk and is often lost on subculture in the laboratory.

There are a few other growth characters which possess some differential value. Thus we may note the classical difference between the growth of *Salmon typhi* and *Bacter coli* on potato the former being colourless and barely visible the latter displaying a characteristic yellowish tint (Fremlin 1893). Again there is the so-called 'nail head' growth of Friedlander's bacillus when grown in stab culture in gelatin due to the raised circular convex growth which sometimes develops on the surface above the inoculation track but this phenomenon is inconstant. Chromogenesis is not a common property of the group but appears to be met with occasionally in individual members, such as the organism described by Parr (1937) under the name of *Bacter aureus*. It is a moot point, however whether such organisms should not be classified in the genus *Chromobacterium*.

**Resistance to Heat, and to various chemical Substances.**—Most members of this group are killed by exposure to a temperature of 50° C. for about 1 hour or of 60° C. for 15-20 minutes. So far as they have at present been studied the various species within the genus do not differ from one another in any significant way. On the whole the typical fecal coli strains tend to have a slightly higher resistance to heat than the closely related members of the intermediate-aerogenes-cloacae group. A small proportion of them are not completely destroyed by exposure to 60° C. for 30 minutes in broth or to pasteurization at 63° S° C. for the same time in milk (Henneberg and Wendt 1930; Wilson *et al* 1930). Towards chlorine in water the aerogenes type tends to be slightly more resistant than the coli or intermediate types (Bardsley 1930b). In raw water stored under atmospheric conditions coli form bacilli may remain alive for weeks or months. On the whole *Bacter aerogenes* tends to survive rather longer than *Bacter coli* but the results are influenced among other things by the temperature (see Platt 1935; Raghavachari and Iyer 1939b).

In faeces stored at 0° C coliform organisms may be demonstrated for a year or more the coli type being often gradually supplanted by the intermediate and aerogenes types (Parr 1938)

There are certain chemical substances which exert a definitely selective bactericidal or inhibitory action

The typhoid bacillus is less resistant to the lethal action of mineral acids than is the colon bacillus Winslow and Lochridge (1906) showed that the bactericidal effect was due to the action of the dissociated hydrogen ions and found that the concentration required to bring about a 99 per cent reduction in the viable organisms in a bacterial suspension was 2.94 per million in the case of *Salm typhi* and 7.49 per million in the case of *Bact coli*

Malachite green in suitable concentration kills *Bact coli* or inhibits its growth without exerting the same effect on *Salm typhi* (Loeffler 1903 1906 Lentz and Tietz 1903 1905) There are other green dyes that have a similar selective action, and more recent studies (see Browning Gilmour and Mackie 1913 Krumwiede and Pratt 1914) have shown that brilliant green gives the best differential results To this dye the bacilli of the paratyphoid group are most resistant the typhoid bacillus is somewhat less resistant while the dysentery bacilli and still more the members of the *Bact coli* group are very susceptible

Caffeine (Roth 1903 Hoffman and Ficker 1904) and lithium chloride (Gray 1931 Havens and Mayfield 1933) are other substances that inhibit the growth of *Bact coli* in concentrations that have no effect on the typhoid bacillus, while cholesterol (see Manfredi 1917) appears to inhibit the growth of typhoid or paratyphoid bacilli in concentrations that permit the growth of *Bact coli*

Sodium deoxycholate in the presence of sodium citrate inhibits the growth of coliform bacilli while having little effect on organisms of the *Salmonella* and *Shigella* groups (Leifson 1935 Hynes 1942) Potassium tellurite in suitable concentration in the presence of iron alum is said to inhibit coliform and salmonellae but not the Flexner dysentery bacilli (Wilson and Blair 1941) Selenium salts were found by Haendel (see Guth 1916 Leifson 1936) to inhibit coliform more than typhoid bacilli and tetrathionate was found by Muller (1923) (see also Knox Gell and Pollock 1943) to have much the same effect

Differences of this kind have not however been employed for the purposes of identification or classification They have on the other hand been extensively exploited in devising selective or enrichment media for the isolation of the pathogenic species from faeces or water They are considered from this point of view in Chapters 69 and 92

**Biochemical Activities**—From the first isolation of *Bact coli* by Escherich fermentation tests were found to provide the readiest method of distinguishing one species of *Bacterium* from another It was soon found for instance that *Bact coli* actively fermented lactose while *Salm typhi* did not (Chantemesse and Widal 1887, Smith 1890), and the production of acid and gas from glucose by *Bact coli* but of acid alone by *Salm typhi* was pointed out by Chantemesse and Widal in 1891 The addition of a suitable indicator to the test media to register acidity (Wurtz 1892) and the introduction of the simple fermentation tube as a test for gas production (Smith 1890 1893 Durham 1898) greatly increased the facility with which a large series of comparative qualitative tests could be carried out To dextrose and lactose other test substances have from time to time been added such as the hexoses fructose levulose and galactose, the disaccharides maltose and saccharose, the trisaccharide raffinose, polysaccharides such as dextrin starch and inulin, the pentoses arabinose and xylose the methyl pentose, rhamnose, the hexahydric alcohols dulcitol and mannitol the glucoside salicin, and the cyclohexanehexol inositol The reaction in litmus milk the presence or

absence of indole production in peptone water, and the production of hydrogen sulphide have served as additional differential criteria, and other tests, such as the final pH attained in a dextrose-containing medium the nature and amount of the gases evolved, or the production of some particular fermentation product, have been employed as aids to differentiation within particular sub-groups.

It soon became clear that the presence or absence of the power to ferment lactose, originally noted as differentiating *Bact coli* from *Salm typhi*, corresponded to a fundamental line of cleavage within this group. The lactose fermenters were, for the most part, found to be normal inhabitants of the intestinal tract of man or the higher animals or to exist on various plants or in the soil. They were active fermenters of many carbohydrates, including polysaccharides, they tended to clot milk, as well as acidify it, they frequently formed indole, and they tended to reduce various dyes (Dunbar 1892, Rothberger 1898). The non lactose fermenters tended, as a class, to comprise the pathogenic species, producing intestinal infections in man and animals, and the range of their fermentative activity tended to be less extensive than that of the lactose fermenters, though most species attack a considerable number of substrates.

This early division of the genus into two broad sub groups on the basis of the lactose fermentation has stood the test of time, although there are a few species or types for which some intermediate position must be found. As already noted, the genera *Salmonella* and *Shigella* have been created for the more important of the non lactose fermenters. These organisms will be considered in separate chapters. In the remainder of the present chapter we shall devote ourselves to a consideration of the lactose fermenting organisms together with the indeterminate group of paracolon bacilli, which appear to be more nearly related to the coliform bacilli than to the *Salmonella* or *Shigella* organisms. Though no clear line of demarcation can be laid down between the coliform bacilli and the Friedlander group of bacilli it will be convenient to describe them separately.

### The Coli-Aerogenes Group

**Biochemical Differentiation.**—In his original description of *Bact coli*, Escherich noted the occurrence of two types, one of which, *Bact coli*, formed relatively long rods was motile and clotted milk slowly, while the other, *Bact lactis aerogenes*, formed shorter, plumper rods, was non motile, and clotted milk more actively. Kruse (1894) emphasized the heterogeneity of the group covered by the term "*B coli*" as usually employed, pointing out that it included a variety of related species, widely distributed as intestinal parasites and in water and soil. The use of a relatively small series of fermentation tests, including especially dextrose, lactose, sucrose starch inulin action on litmus milk, and indole formation, resulted in the recognition of certain primary divisions within this group (Reif 1896, Grumbert and Legros 1900, Durham 1901, Jordan 1903). One of the groups so defined fermented polysaccharides such as starch and inulin, and usually failed to form indole, this corresponded with the *Bact lactis aerogenes* type of Escherich and became established as a separate species, the *lactis* being usually omitted from the name. The second and third groups differed from *Bact aerogenes* in failing to ferment starch and inulin, and in forming indole in peptone water. They differed from each other in regard to their action on saccharose. One, corresponding to the existing strains of Escherich's *Bact coli commune*, failed to ferment this sugar, the other fermented it, and Durham (1901), who found it to occur more frequently than the saccharose negative

type, named it *Bact coli communis*. The application of a more extended series of tests resulted in further subdivision of this group, and elaborate classifications were suggested by various observers on the basis of the results obtained (Bergey and Deehan 1908, MacConkey 1905, 1909, Jackson 1911). It may be noted that one important correlation between biochemical activity and natural habitat had already been detected. The *Bact aerogenes* type was found to be a relatively infrequent inhabitant of the intestine, but was frequently isolated from certain grasses and from the soil, while *Bact coli commune* and *Bact coli communis* were noted to be typically intestinal parasites (Winslow and Walker 1907). This correlation was of practical as well as of theoretical importance. The presence or absence of "B coli" in water supplies, and the relative number of this organism if present, soon came to be recognized as a very valuable indication of the presence and degree of faecal pollution (see Chapter 92), and it became very desirable, apart from any question of systematic classification, to differentiate between those types which were of intestinal origin, and those which might occur in unpolluted waters. The investigations of those who have been primarily concerned with the practical aspects of the bacteriological analysis of water supplies have added materially to our knowledge of this group.

Apart from the merely positive or negative results, as regards acid or gas production in the various sugars, certain observations made in the earlier days had indicated a difference in kind between the fermentation of one and the same carbohydrate by different strains of bacilli of the colon type. Thus Smith (1895) using the method of the fermentation tube, noted that gas was produced more rapidly and in greater amount by *Bact aerogenes* than by *Bact coli*, and a rough estimation of the ratio of  $\text{CO}_2$  to  $\text{H}_2$  in the gas evolved showed that this was higher with the former organism than with the latter. He noted also that the degree of final acidity was lower with *Bact aerogenes* than with *Bact coli*. In both respects *Bact cloaca*, a coliform organism isolated from sewage by Jordan (1890) and differentiated from all other types of coliform bacilli by its power of liquefying gelatin, corresponded with *Bact aerogenes*. Russell and Bassett (1899) confirmed the differential value of a high or low  $\text{CO}_2$ ,  $\text{H}_2$  ratio, and noted that the high ratio strains appeared to be normal soil forms, rather than intestinal parasites. This question was placed on an entirely new footing by the careful quantitative studies of Harden and his colleagues (Harden 1901, 1905; Harden and Walpole 1906) who showed that strains of coliform bacilli were divisible into two well defined classes. In one, typified by *Bact coli*, the  $\text{CO}_2$ ,  $\text{H}_2$  ratio of the gas evolved gave a value closely approximating unity. In the other, typified by *Bact aerogenes*, the  $\text{CO}_2$ ,  $\text{H}_2$  ratio gave a value of 2:1, or thereabouts. These observations have been amply confirmed by later observers.

Voges and Proskauer (1898) had described a colour reaction given by certain bacteria, but not by *Bact coli*. It is obtained by adding a few drops of a strong solution of potassium hydrate to a culture grown in a dextrose medium. In a positive reaction a red, fluorescent coloration appears, which may develop relatively slowly. The nature of this reaction was elucidated by Harden and his colleagues (Harden 1906, Harden and Norris 1911) who showed that it depends on the production of acetylmethylcarbinol ( $\text{CH}_3\text{CHOHCOCH}_3$ ), this, in the presence of alkali and of atmospheric oxygen is oxidized to diacetyl ( $\text{CH}_3\text{COCOCH}_3$ ) which reacts with the peptone of the broth to give the red colour.

This reaction had been applied to the examination of the colon group by some

of the observers referred to above, and it had been noted that *Bact. aerogenes* as opposed to *Bact. coli* gave a positive reaction (Durham 1901) MacConkey (1909) observed the great preponderance of Voges Proskauer negative types among strains isolated from the faeces positive reactions were given by 11 of 173 strains isolated from human faeces, 8 of 67 strains from the faeces of the horse and none of 57 strains from faeces of the calf goat pig or goose

The fundamental importance of the Voges Proskauer reaction, and of the  $\text{CO}_2$   $\text{H}_2$  ratio as compared with the presence or absence of fermentation in particular carbohydrates other than lactose was not however realized by the earlier investigators so that the reaction was simply assigned a place among some selected series of tests and V P positive and V P negative strains were often allocated to the same sub group though it was noted by Howe (1904) during the examination of strains of lactose-fermenting bacilli derived from water that there was almost perfect correlation between a positive V P reaction and the ability to produce large amounts of gas from dextrose

Petruschky (1899 1890) made the first attempt to measure by titration the degree of acidity produced by various members of the coli typhoid group while Smith (1895) as noted above called attention to the low acid production of *Bact. aerogenes* as compared with *Bact. coli* A great advance in the investigation of this aspect of bacterial metabolism was marked by the introduction of indicators which rendered possible the ready determination of the hydrogen ion concentration attained during the bacterial fermentation of any test substance. Clark and Lubs (1915) on this basis, devised the methyl red test for the differentiation of members of the coli typhoid group The addition of this indicator to five-day cultures in dextrose phosphate peptone water distinguishes between those strains which produce and maintain a high concentration of hydrogen ions, and those which produce an initial lower concentration of hydrogen ions and then cause reversion towards neutrality by the further decomposition of the organic acids to carbonates, and perhaps by the formation of ammonium compounds from proteins The former type such as *Bact. coli* give a red coloration and are referred to as methyl red positive the latter, such as *Bact. aerogenes* give a yellowish colour and are referred to as methyl red negative It soon became clear that there was a very high negative correlation between the methyl red test and the Voges Proskauer reaction (Levine 1916a b) and a series of intensive studies soon placed on a firm foundation the conclusion, already propounded as a tentative hypothesis, that the lactose-fermenting coliform bacilli could be divided into two primary divisions on the basis of the  $\text{CO}_2$   $\text{H}_2$  ratio the Voges-Proskauer reaction, and the methyl red test The first of these, containing strains giving a  $\text{CO}_2$   $\text{H}_2$  ratio of about 2 : 1 V P positive and M.R. negative comprised the great majority of the strains isolated from plants grain, and unpolluted soil or water Such strains were relatively infrequent in material obtained from the intestines of man or animals This group could be further subdivided, on the basis of gelatin liquefaction, into the non liquefying form *Bact. aerogenes* and the much less common liquefying form *Bact. cloacae* The second group containing strains giving a  $\text{CO}_2$   $\text{H}_2$  ratio of approximately 1 : 1 V P negative and M.R. positive, contained the great majority of those strains isolated from the intestines of man or animals, as exemplified by *Bact. coli commune* or *Bact. coli communis* This group was found to be further divisible on the basis of the ordinary fermentation tests, along lines which will be considered later (Keyes 1909 Rogers *et al.* 1914 1915 1918 Johnson 1916

Hulton 1916, Levine 1916c, d, 1917, Burton and Rettger 1917, Chen and Rettger 1920, Winslow *et al* 1919, Bardsley 1926, 1934, Pawan 1931)

Besides the division rendered possible by the tests we have just outlined into a *coli* group on the one hand and an *aerogenes-cloacae* group on the other further work revealed the occurrence of a third group of strains possessing properties intermediate between those of the two main groups. This group is, as yet, not completely defined, and is therefore most conveniently referred to as the "intermediate" group. Brown (1921) drew attention to the usefulness of a medium containing citrate for the differentiation of *Bact coli* from *Bact aerogenes*. Koser (1923, 1924, 1926a, b) devised a synthetic medium in which citrate was provided as the sole source of carbon. He found it possible to differentiate coliform bacilli into a MR +, VP -, citrate - *coli* type, a MR -, VP +, citrate + *aerogenes* type, and a MR +, VP -, citrate + intermediate type. Examination of 104 soil strains from fields subjected to only chance pollution showed that 23.1 per cent were of the *coli*, 67.3 per cent of the *aerogenes*, and 7.7 per cent of the intermediate type. Further work in numerous countries soon revealed the value of this test in differentiating the intermediate from the *coli* group (see Bardsley 1926, 1934, Pawan 1931).

As pointed out by Vaughn Mitchell and Levine (1939) and Levine (1941) it is advisable to carry out the methyl red test on a culture incubated for 5 days at 30° C and the Voges Proskauer test on a culture incubated for not more than 2 days at 30° C. At 37° C cultures of some strains of the *aerogenes cloacae* group may fail to revert to alkaline and acetylmethylcarbinol is either not formed or is destroyed (see Tittler 1938). Some of the discrepancies in the literature are doubtless ascribable to carrying out these tests under unfavourable conditions. Special precautions have also to be taken with the citrate test. It may be noted that, according to a number of authors (Kline 1935, Stuart Griffin and Baker 1939, Parr 1939, Griffin and Stuart 1940) some strains of *Bact coli* may acquire the power of utilizing citrate.

In Table 40 we have summarized the results recorded by various observers (see Bardsley) with regard to the percentage of strains isolated from different sources, which give the particular reactions to which we have referred. We have included the indole reaction because recent work suggests that it is particularly significant in relation to habitat. It will be noted that figures are available for all tests only in the case of the strains derived from animal faeces, but it may safely be assumed that the strains with a high gas ratio would have given positive VP and negative MR reactions, and *vice versa*.

TABLE 40

SHOWING THE PERCENTAGE OF LACTOSE FERMENTING COLIFORM BACILLI FROM VARIOUS SOURCES WHICH GIVE THE REACTIONS INDICATED (VARIOUS AUTHORS)

Source	Percentage.					
	CO <sub>2</sub> H <sub>2</sub>		MR. +	VP +	Indole +	Citrate +
	2 1	1 1				
Faeces (human and animal)	14.37%	85.62%	90.40%	7.87%	93.03%	6.71%
Sewage (and similar material)	—	—	82.74%	17.26%	—	—
Soil	—	—	14.70%	80.24%	33.40%	89.6%
Grasses and grains	95.18%	4.82%	—	—	—	—

Numerous other tests have been used for the differentiation of members of this group

The fermentation of cellobiose a glucoside derived from cellulose (see Jones 1924 Jones and Wise 1926) has been recommended for distinguishing *Bact aerogenes* and intermediate strains which ferment this substance from *Bact coli* which fails to do so (see Koser 1906c Tittsler and Sandholzer 1936) Too much reliance should not be placed upon the result of this test in the identification of individual strains Betty Smith (1942a) for example who examined 600 strains of coliform bacilli found that about 10 per cent of *Bact coli* strains fermented cellobiose and that about one third of the intermediate and *aerogenes* strains were without action upon it

The production of  $H_2S$  in a proteose peptone ferric citrate agar medium was found by Vaughn and Levine (1934) to be characteristic of intermediate strains Further observations by the same authors (1936a b) showed that if cysteine was incorporated in the medium a high proportion of all coliform strains produced  $H_2S$  and that even without cysteine the result was greatly influenced by the concentration of agar If this test is to be of differential value strict attention must be paid to the exact composition of the medium

Lefson (1933) observed an almost perfect correlation between the V P reaction and the fermentation of sodium malonate

The fermentation of polysaccharides such as starch by the *aerogenes* group was pointed out by Durham (1901) and confirmed by Levine (1918) It has been made use of by W J Wilson (1933) in the preparation of a medium for distinguishing *Bact aerogenes* from *Bact coli* On the other hand a medium containing sodium sulphate and rosolic acid in certain proportions and another containing hexamine are said to favour *Bact coli* at the expense of *Bact aerogenes* (Wilson W J 1933)

For the differentiation of *Bact aerogenes* from *Bact cloacae* gelatin liquefaction and the production of acid and gas from glycerol and starch are of chief value (Levine 1918) *Bact aerogenes* is gelatin - glycerol + starch +, and *Bact cloacae* gelatin +, glycerol - starch Intermediate strains are said to be differentiated from strains of *coli* type not only by their production of  $H_2S$  but also by their ability to form trimethyleneglycol in the anaerobic fermentation of glycerol (see Werkman and Gillen 1932)

Besides the effect on carbohydrates nitrogen utilization has been explored in an attempt to discover reliable tests Koser (1918) Chen and Rettger (1920) and numerous subsequent workers (see Bardley 1926 1934) found that *aerogenes* strains were able to utilize uric acid as their sole source of nitrogen whereas strains of the *coli* and intermediate types were not More recently Mitchell and Levine (1938) have examined other substances such as yeast nucleic acid allantoin hydantoin uracil, and urea They find that *aerogenes* strains can make use of all these substances as their sole source of nitrogen but that intermediate strains can use only urea and *coli* strains only uracil

Considerable help is afforded by a test introduced as long ago as 1904 by Eijkman (1904 1914) who found that *coli* but not *aerogenes* strains were able to form gas in a glucose broth medium incubated at  $46^\circ C$  This test has had a chequered career having been reported on favourably by some workers and utterly condemned by others Recent work (see Levine *et al* 1934 Wilson *et al* 1935) has rendered it clear that the success of the test depends on exact standardization of the incubating temperature which must be adjusted to  $43^\circ-45^\circ C$  in the medium itself The only satisfactory means of doing this is to incubate the tubes in a constant temperature water bath The differential value of the test is greatly enhanced by the replacement of glucose by lactose Our own experience suggests that MacConkey's lactose bile salt broth is the medium of choice Using this medium in the way described it was found that of 193 M R +, V P - citrate -, indole + *coli* strains 180 gave a positive  $44^\circ C$  MacConkey reaction whereas of 303 other strains 40 of which



belonged to the indole-negative *coli* group only 10 did so. The value of the method has now been amply confirmed (see Mackenzie and Hilton Sergeant 1938, Dodgson 1938, Bardsley 1938a, Clegg and Sherwood 1939, Perry 1939, Raven Peden and Wright 1940, Ferramola 1940, Clegg 1941, Sherwood and Clegg 1942, Batty Smith 1942b, Stuart *et al.* 1942). Two discordant reports on the value of the test have appeared. Raghavachari and Iyer (1939a) found that about 50 per cent. of aerogenes like strains in Madras waters and Boizot (1941) about 10 per cent. in Singapore waters were able to produce gas in MacConkey broth at 44° C. Possibly this may be due to a higher optimal temperature for growth of *aerogenes* strains in the tropics than in temperate climates. With this reservation however we may say that the 44° C. MacConkey test appears to be of greater value than any other single test in picking out typical faecal *coli* strains.

There can, we think, be no doubt that the primary division of the lactose fermenters must be made on the basis of the gas ratio, methyl red, Voges Proskauer, and citrate tests. By this means we obtain a primary classification into *coli*, intermediate, and *aerogenes* groups. The secondary division is more difficult. From an ecological standpoint we believe that the indole 44° C. MacConkey and gelatin liquefaction tests afford the most satisfactory grouping, but from a systematist's point of view there is much to be said in favour of classification on sugar reactions. Taking the first method of grouping we may note that about 93 per cent. of strains from human and animal faeces produce indole, and that about 95 per cent. give a positive 44° C. MacConkey reaction. On the other hand indole negative and 44° C. MacConkey negative faecal strains are uncommon. A positive indole test given by a citrate negative strain or a positive 44° C. MacConkey test given by any strain is therefore strongly suggestive of its faecal origin. Though both intermediate, *aerogenes* and *cloacae* strains are often present in mammalian faeces in small numbers (Cruickshank and Cruickshank 1931, Parr 1939) the balance of evidence strongly suggests that in this country and in the United States (see Griffin and Stuart 1910) these organisms are primarily of non faecal origin. Their main habitat is still in doubt, but they are found most often in soil, grains, grasses, food stuffs and decaying vegetation. The classification reached by this method is of special value in the interpretation of water analysis tests and is depicted in Chapter 92.

With regard to the second method of grouping the test substances that have been accorded special prominence are saccharose, dulcitol and salicin, the value of the last being emphasized by Kligler (1914a, b) and by Levine (1917). Winslow, Kligler and Rothberg (1919) in their excellent review of the classification of the whole *coli* typhoid group conclude that the types set out in Table 41 under their appropriate names are the only ones which are sufficiently well established to merit separate consideration.

It will be noted that although the reactions in dulcitol and adonitol are included in the table the species are adequately defined by the reactions in saccharose and salicin together with the presence or absence of motility. It will also be noted that *Bact. coscoroba* differs from *Bact. coli communis* in being non motile, *Bact. immobile* from *Bact. coli commune* in the same way, and *Bact. grunthal* from *Bact. acidi-lactici* only in being motile. Winslow and his colleagues express the view that the presence or absence of motility taken alone does not justify specific differentiation, and they suggest the recognition of four species: *Bact. neapolitanum*, *Bact. coli communis*, *Bact. coli commune* and *Bact. acidi-lactici*, regarding the *coscoroba*, *immobile* and *grunthal* types as varieties of the corresponding species. This appears to us a wise and conservative view. We should ourselves prefer to narrow the limits still further regarding *Bact. coli* as a single species and placing the *neapolitanum*, *communis*, *commune* and *acidi-lactici* types as varieties.

TABLE 41  
FERMENTATIVE TYPES OF *Bact coli* (Winslow et al)

	Saccharose	Salicin	Dulcitol	Adonitol	Motility
<i>Bact neapolitanum</i>	A G	A.G	—	—	—
<i>Bact coli communis</i>	A G	—	A.G	—	+
<i>Bact coscoroba</i>	A G	—	A G	—	—
<i>Bact coli commune</i>	—	A.G	A.G	—	+
<i>Bact immobile</i>	—	A.G	A.G	—	—
<i>Bact acidi lactici</i>	—	—	—	A G	—
<i>Bact grunthal</i>	—	—	—	A.G	+

**Serological Differentiation**—Several attempts have been made to study the antigenic structure of various strains of *Bact coli* by the method of agglutination. The results display an extreme heterogeneity of antigenic factors (Mackie 1913, Mirra 1936, Bredenbrocker 1938, Stuart et al 1940). In some instances, as in the hæmolytic strains of *Bact coli* isolated by Dudgeon from cases of acute urinary infection, there is evidence that antigenically homogeneous groups may be related to particular infective conditions (Dudgeon et al 1921, 1923). The same suggestion is apparent in the work of Lovell (1937) on *Bact coli* strains isolated from calves that had died from white scours. Using the precipitin test, he obtained evidence of the existence of two antigens: one, a soluble specific carbohydrate substance, associated with the capsule and probably similar to or identical with that previously described by D. E. Smith (1927), the other, of undetermined nature, contained in the body of the bacillus. Of 110 strains from 45 calves, he was able to place 79 in one of eight different types. It is interesting to note that some coliform strains contain O or H antigens found in members of the *Salmonella* group (see Chapter 30). The detailed antigenic analysis of the coliform bacilli remains for future study.

Stamp and Stone (1944) have described the presence of a common antigen—referred to as the  $\alpha$  antigen—in many members of the coliform and paracoliform group. It is more heat stable than the H antigen, withstanding a temperature of 75° C for 1 hour, but less so than the O antigen, being inactivated at 100° C in 15 minutes. It is destroyed by exposure for 1 hour to alcohol at 65° C. It is commonest in recently isolated strains, and tends to be lost on subculture. Agglutinins to the  $\alpha$  antigen are not uncommon in rabbit sera, and may lead to confusion in the identification of suspected pathogenic organisms.

**Pathogenicity**—The majority of the lactose-fermenting coliform bacilli appear to be non pathogenic under ordinary conditions. The MR —, VP + group have their normal habitat on plants, the MR +, VP —, citrate + group appear to live in the soil, while the MR +, VP —, citrate — forms are normal intestinal parasites. Under abnormal conditions these bacilli may cause acute or chronic infective lesions in the urinary tract, or elsewhere (see Chapter 67). It seems probable also that they may sometimes play a part in the causation of enteritis, in man or in animals, though their role appears generally to be a secondary one. There is, however, some evidence that, in newly born animals, the advent to the intestine of *Bact coli*, or of particular races of this organism, may result in a local enteritis or even general septicæmia. Smith and Liddle (1922) and Smith and Orcutt (1925), for example, attributed the disease in newly born calves known

as *white scours* and the fatal septicæmia that may sometimes accompany it to infection with *Bact. coli* (see also Lovell 1937). They found that the disease was prone to occur in calves fed on milk instead of colostrum. It is possible that the neonatal diarrhoea of human infants may result from the access to the intestine of especially virulent forms of *Bact. coli*, but the available evidence is as yet insufficient to afford conviction (see Chapter 71). According to Gwatkin, LeGard and Hadwen (1938) bovine mastitis may occasionally be due to coliform bacilli.

The pathogenicity of most strains for laboratory animals is low. Very large doses administered intraperitoneally to the mouse, or intravenously to the rabbit may prove fatal, but it seems likely that death results in these cases from a toxæmia rather than from a true infection. Occasionally strains of greater virulence are found.

Several observers have described the presence of soluble toxic substances in young broth cultures of *Bact. coli* (see for instance Steinberg and Ficker 1926, Smith and Little 1927, Smith D.I. 1927, Rennebaum 193). But most of these toxins have proved to be heat stable and there is no reason to believe that they differ from the toxic components that can be extracted in larger quantity from the bacterial cells. Since there is at the moment no clear evidence that the toxic substances that can be extracted from the colon bacillus differ in their action from those of other species within this genus, a discussion as to their probable nature may be deferred to a later section.

### The Friedländer Group

There are certain species of lactose fermenting coliform bacilli which cannot readily be placed in any of the groups which we have differentiated above. There are, for instance, the so-called capsulated bacilli including Friedländer's pneumobacillus, Abel's bacillus of ozæna, the bacillus of rhinoscleroma and others. There appears to be a preponderance of opinion that the organisms of this group are in some way related to *Bact. aerogenes* principally because that organism is sometimes capsulated, but the fermentation reactions as described by those who have isolated and studied these capsulated coliform bacilli appear to be extremely variable and several observers have recorded the fermentation of saccharose but not of lactose. The balance of evidence suggests that these strains should be included in the genus *Bacterium* but it is convenient in the meantime to consider them in a separate subsection of this group.

#### GROUP CHARACTERISTICS

Short non motile non sporing capsulated Gram negative rods, giving a profuse mucoid growth on solid media and usually fermenting carbohydrates with the production of acid and gas. Usual habitat, respiratory tract of man and certain animals.

Under the general term *B. mucosus capsulatus* a large number of organisms have been described with the characteristics enumerated above. V. Frisch in 1882 isolated a capsulated bacillus from patients with rhinoscleroma. In 1883 Friedländer cultivated a similar organism—generally known as Friedländer's bacillus or *Bact. friedländeri*—from the lungs of patients who had died of pneumonia. Loewenberg, in 1894 and Abel in 1896 cultivated a similar organism from the nasal secretion of patients with ozæna. Besides these, several organisms have been described by other workers such as *B. pseudopneumonicus* Passet, *B. canalis*

*capsulatus* Mori *Proteus hominis capsulatus* Bordoni Uffreduzzi *B capsulatus* Pfeiffer (Pfeiffer 1889) *B mucosus capsulatus* Paulsen *B crassus sputigenus* Kreibohm *B buccalis muciferens* Miller the bacillus of sputum septicæmia Miller, the granuloma bacillus (Small and Julianelle 1923) *Bacterium mucogenum* (Edwards R T 1905) *B capsulatus* (Wright and Mallory 1895) and *Klebsiella paratyphica* an organism isolated by Wallace Cahn and Thomas (1933) from a paralytic tick borne disease of moose (For references see Fricke 1896 Bamforth 1928) The *ozæna* bacillus described by Abel must not be confused with the non-capsulated coccobacillus described by Perez (1899) (See Chapter 19)

**Habitat**—Friedlander's bacillus appears to lead a parasitic existence. The commonest situation in which it is found is the respiratory tract of man. It is an occasional inhabitant of the nasopharynx, it is found in diseased conditions of the nose such as *ozæna* and rhinoscleroma and it is sometimes present in the lungs in pneumonia influenza bronchitis bronchiectasis tuberculosis and



FIG 142—*Friedlander's Bacillus*

From an agar culture, 24 hours 3 °C  
showing capsules ( $\times 1000$ )

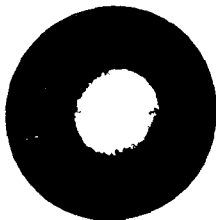


FIG 143—*Friedlander's Bacillus*

Surface colony of mucoid type on agar 24  
hours 37° C ( $\times 8$ )

other diseases. It has been isolated from a large number of suppurative conditions in different parts of the body, such as pleurisy appendicitis, cystitis pyelonephritis ulcerative endocarditis endometritis, brain abscess and general septicæmia (Perkins 1904) and was found by Dudgeon (1926) in 5.5 per cent. of faeces examined from normal and abnormal conditions.

**Morphology**—In the body the organism generally takes the form of short ovoid diplobacilli surrounded by large capsules looking not unlike pneumococci. In culture it is pleomorphic. The usual form is a short straight thick rod about  $1.2 \mu$  long and  $0.8 \mu$  wide with parallel or bulging sides and rounded or slightly pointed ends. The bacilli are usually in pairs end-to-end or arranged singly. Besides this form there are several others of most varied appearance—thick curved sausage forms clubbed forms long sinuous filamentous forms long straight rods—staining regularly or irregularly. In most strains the organisms are surrounded by a capsule apparent in cultures which can be demonstrated by Gram's stain or by any of the ordinary capsule stains. In some strains however the organisms, instead of being individually capsulated, are embedded in an interstitial mucoid

substance which stains less intensely than the bacilli. There are moreover, non capsulated variants which morphologically and culturally resemble the coliform bacilli, these often become predominant in old cultures. The organisms are non motile and non sporing. They stain easily, sometimes show bipolar staining and are Gram negative.

The capsule of these organisms is formed not only in the animal body but in culture. To demonstrate it in body fluids it is sufficient to fix it by heat and stain with carbol fuchsin or methylene blue. To demonstrate it in culture it is advisable to suspend the bacilli in a low dilution—not higher than 1/5—of serum, dry in air and fix in a saturated solution of corrosive sublimate before staining (Toenniesen 1912).

Chemical examination of the capsule has revealed the presence of a nitrogen free carbohydrate material which according to Toenniesen (1921) and Kramár (1921) is a polysaccharide of galactose, but Heidelberger, Goebel and Avery (1925) regard it as a polysaccharide containing glucose. The capsule appears to consist of gum. This substance may be separated from the bacilli in the following manner (Toenniesen 1921). The organisms are dried, suspended in water to which 1 per cent KOH is added, and heated until the gum covering passes into solution. On cooling the undissolved bacilli form a sediment and the opalescent supernatant fluid is pipetted off. This is acidified with acetic acid and 3 volumes of 91 per cent alcohol are added when a heavy precipitate of the gummy substance occurs, this precipitate is purified by dissolving it in distilled water and reprecipitating it with acid and alcohol. In the pure state it is a loose snow white powder, giving an opalescent solution in distilled water acids or alkalis, it is free from nitrogen and does not reduce Fehling's solution. It has special antigenic properties which will be referred to later.

**Cultural Reactions.**—One of the characteristic features of this group is the luxuriant, greyish white, mucoid almost diffuent growth on agar. This results no doubt from the presence of the gummy envelope around the bacilli containing a high proportion of water—92 per cent (Toenniesen 1921). The condensation water on an agar slope is converted into a greyish white mucoid mass. In broth the organisms grow freely, giving rise after a few days to a marked viscosity so that the medium takes on the consistency of melted gelatin. Great stress used to be laid on the nail headed growth in stab gelatin cultures, a circular convex growth may occur on the surface with a filiform growth in the stab the whole resembling a round headed nail. This appearance is not constant and is given only by some strains, it depends too on the amount of gelatin in the culture, with small amounts about 4 per cent, the surface growth is flat (Friedländer 1883). There is no liquefaction of the gelatin but often a large napiform bubble of gas accumulates just beneath the surface, giving on first sight the appearance of liquefaction. On potato there is a moderate creamy yellow mucoid growth later turning to a buff or light café-au lait colour.

The cultural appearances of the Friedländer group are subject to considerable variation. This is due to the production of variants which have different forms of growth from the parent strain. Toenniesen (1913) found that when the usual mucoid type was kept in the incubator and subcultured every few days non mucoid variants rapidly appeared. As the original colonies grew older white, more or less translucent peripheral sectors developed, which on microscopical examination were found to consist largely of non capsulated bacilli, these could

be subcultured, and obtained pure. The growth of these non-capsulated bacilli was no longer mucoid, but resembled the growth of *Bact. coli*. Regression to the mucoid type might occur—often suddenly. As well as this non-mucoid type Toennissen (1914) later observed three other variants. The mucoid capsulated type may be regarded as the smooth form, and the non-mucoid, non-capsulated type as the rough form of Friedlander's bacillus (Julianelle 1936). Dissociation may take other forms. Thus according to our own observations, secondary colonies may appear in the substance or on the surface of the original colonies, or there may develop a jelly-like translucent peripheral fringe showing slight radial striation, or sometimes the whole colony may dry up and wither away, leaving an effuse transparent layer looking like ground glass—aptly called by Collins (1931) suicide colonies. Beham (1912) Hadley (1937) and Goelings (1933) have recorded similar observations.

**Resistance and Metabolism.**—The organisms are killed by moist heat at 55° C. in half an hour. They may survive drying for months (Loewenberg 1934). If kept at room temperature cultures remain viable as a rule for weeks or months. They are aerobic; growth under strictly anaerobic conditions is very poor. There is no hæmolysis of horse or sheep's red cells. The optimum temperature for growth is 37° C. the limits are 12° and 43° C. Some strains form a slightly brownish pigment, most easily produced by growth on potato.

**Biochemical Reactions.**—The fermentation of sugars by members of this group is subject to considerable variation (Clairmont 1902, Perkins 1904, 1907, Edwards 1906, Pare 1912, Fitzgerald 1914, Coulter 1917, Small and Julianelle 1923, Bamforth 1928, Edwards 1928, 1930, Julianelle 1930, 1933, Elbert and Guerkess 1930, Hay 1932, Wallace Cahn and Thomas 1933, Morris and Julianelle 1934, Kiewe and Hsu 1933, Wilson *et al.* 1933, Gossens 1934, Osterman and Rettger 1941). Many strains produce acid and gas in glucose, maltose, mannitol, lactose, sucrose and salicin. Several, however, do not attack lactose and others either do not ferment sucrose or ferment it late. Gas may be formed rapidly or not for several days; some strains do not form gas. Occasionally no sugars are fermented, but this lack of fermentative ability is uncommon except in strains that have been subcultured for a long time in the laboratory. In litmus milk also the reaction of different strains varies. Generally acid and clot are formed, but many strains do not produce sufficient acid to precipitate the caseinogen, while some strains produce no change at all. The litmus is occasionally decolorized.

There is fairly general agreement that biochemical reactions do not afford an adequate basis for classification, since most workers have been unable to discover any constant relationship between the biochemical activities of a given strain and its source of origin or antigenic structure. Goelings (1934) and Wielenga (1937) however consider it possible to distinguish three species on the basis of biochemical activity: (a) The least active and the least variable comprises the rhinoscleroma strains, which have the following reactions: they form no gas from carbohydrates; they produce acid in glucose, maltose, mannitol, and adonitol in 24 hours, in sucrose within 10 days, and sometimes in glycerol; they do not ferment lactose, dulcitol, or amygdalin within 3 days; they have no action on litmus milk; they usually produce alkali in neutral peptone water in 10 days; they are M.R. positive and V.P. negative; they do not grow in citrate or *d* tartrate; they do not grow in fresh ox bile; and they form no H<sub>2</sub>S. (b) The second group comprising the oxæna strains is similar in most respects but there is more variability between the different members. Most strains fail to produce gas from carbohydrates, but some do; they produce acid and sometimes gas from glucose, maltose, mannitol, and adonitol in 24 hours; many strains from lactose or amygdalin within 3 days, but the majority fail to ferment sucrose within 10 days and none ferments dulcitol; many strains produce acid and some-

times clot in litmus milk, they usually produce alkali in neutral peptone water in 10 days, they are M R positive and V P negative some strains grow in citrate, most strains grow in fresh ox bile, and many strains form  $H_2S$  (c) The third according to Goslings (1931), comprises the Friedländer and *Bact. aerogenes* strains. These are the most active biochemically, growing freely in fresh bile producing gas from carbohydrate media fermenting lactose and amygdalin within 3 days and sucrose within 10 days, turning neutral peptone water either slightly acid or alkaline reducing litmus, utilizing sodium citrate and *d* tartrate but not forming  $H_2S$ . De Graaff (1936) and Wielenga (1937) would distinguish Friedländer's bacillus from *Bact. aerogenes* though they are not in complete agreement on the criteria that should be used for this purpose.

Of reactions not studied by Goslings or Wielenga the indole test is generally negative, nitrates are reduced to nitrites, ammonia is formed from peptone, methylene blue is generally reduced in broth, gelatin is not liquefied, and there is usually an abundant production of catalase. All these reactions, however apply mainly to Friedländer and *aerogenes* strains, the behaviour of rhinoscleroma and ozena strains has been less thoroughly studied. According to Hay (1932) one of the most characteristic features of the *mucosus capsulatus* group is their ability to ferment inositol but this is likewise shared by *Bact. aerogenes*.

**Antigenic Structure**—Till quite recently no satisfactory division of the Friedländer group had been made on the basis of antigenic structure. The main difficulty was due to the fact that, though injection of the capsulated bacilli into rabbits is able to give rise to an agglutinating serum, this serum has little action except on non-capsulated organisms, several attempts were therefore made to rid the bacilli of their capsules. Porges' method (1905) was one of the most successful. He suspended an agar culture in 10 ml of saline filtered through paper mixed it with a quarter of its volume of N/4 HCl solution, and heated for 15 minutes, it was cooled rapidly, and neutralized with N/4 NaOH solution. The resultant suspension was homogeneous and non viscous, and agglutinated with a specific immune serum. Though this method undoubtedly removes the capsules it often renders the bacilli spontaneously agglutinable or agglutinable by normal serum. Streit (1906) found that if the bacilli were grown on potato or potato agar they gradually lost their capsules, and became more agglutinable. Small and Julianelle (1923) obtained the same result by growing them on agar for 24 hours storing the cultures in the ice chest, and subculturing monthly, after 1 to 2 years many of the strains had lost their capsules. Agglutination tests made with non capsulated bacilli obtained in these ways gave, however, very inconstant results nor could a method of analysis in which the natural capsular antigens were disregarded be accepted as satisfactory (Streit 1906 Beham 1912 Fitzgerald 1914 Coulter 1917 Small and Julianelle 1923).

Further work in America (Avery *et al* 1925, Heidelberger *et al* 1925 Julianelle 1926a b, c) has largely cleared up the confusion. It would appear that the immunological reactions of the Friedländer group are similar to those of the pneumococci, depending on the presence in the cell of two entirely different factors—a polysaccharide in the capsule responsible for the type-specificity, and a nucleoprotein in the soma responsible for the species specificity. According to Julianelle (1926a) there are three serological types distinguishable by agglutination absorption, precipitation, and protection tests, and a heterogeneous group (X) of strains that have not yet been antigenically differentiated. If a serum is prepared against Type A by injection of heat killed encapsulated organisms it will agglutinate encapsulated bacilli of its own type, but not those of any other type,

similarly with Types B and C. The serum contains an antibody that reacts specifically with the polysaccharide fraction in the capsule and as the polysaccharide varies in the different types the serum against each type is specific. The polysaccharide of Types A and B has been isolated and has been found to flocculate in the presence of a specific serum. If the bacilli are deprived of their capsules they lose their specificity and agglutinate equally though only to a low titer with sera prepared against any type.

The nucleo-protein can be separated from the dissolved bacilli by precipitation with acetic acid in the cold. It gives rise to a species antibody which does not react with the capsulated bacilli or with the polysaccharide but which agglutinates capsule-free cells of any type and reacts with the nucleo-protein of any type.

A serum prepared by injection of smooth, capsulated bacilli contains antibodies to the polysaccharide and the nucleo-protein, a serum prepared by injection of rough non-capsulated bacilli contains only one antibody—active against the nucleo-protein. For the classification of the Friedlander group into types it is essential to use smooth bacilli both for the preparation of sera and for agglutinating antigens. Unless this rule is strictly observed the results will be unsatisfactory. Failure to realize this principle probably accounts for the discrepant results of the earlier workers.

The capsulated bacilli stimulate the production not only of type-specific agglutinins and precipitins but also of type-specific protective bodies. Thus a serum prepared by injection of capsulated bacilli of Type A will protect mice against intraperitoneal injection of Type A, but not against injection of bacilli of Types B or C. The nucleo-protein does not stimulate the production of protective bodies—a serum, therefore prepared by injection of capsule-free bacilli of any type is unable to protect mice against infection with capsulated bacilli of any type. It is important to note that the polysaccharide in the pure state, though precipitable by immune anti-S serum of the homologous type is unable to stimulate the production of immune bodies, it must be present in combination with the proteins of the cell. The nucleo-protein, on the other hand, is able by itself to stimulate the production of non-specific antibodies. The polysaccharide is present in quite young cultures of Friedlander's bacillus so that filtrates of these cultures may be used as antirens in the precipitin test.

A few tabular results may make these relations clear

TABLE 42

ANTI-SMOOTH SERUM ACTING ON SMOOTH CAPSULATED BACILLI AND ON ROUGH NON-CAPSULATED BACILLI OR ON SMOOTH BACILLI THAT HAVE BEEN ARTIFICIALLY DEPRIVED OF THEIR CAPSULES.

	Anti-S Serum		
	Type A.	Type B.	Type C.
Type A S	+++	—	—
Type B S	—	++	—
Type C S	—	—	++
Type A R	±	±	±
Type B P	±	±	±
Type C P	±	±	±



TABLE 43

ANTI ROUGH SERUM OR AN ANTI PROTEIN SERUM ACTING ON SMOOTH CAPSULATED AND ROUGH NOT CAPSULATED BACILLI

	Anti R or Anti P Serum		
	Type A	Type B	Type C
Type A S	--	—	—
Type B S	—	—	—
Type C S	—	—	—
Type A R	+++	+++	+++
Type B R	+++	+++	+++
Type C R	+++	+++	+++

It is of interest to note that the B type of Friedländer's bacillus is similar immunologically to Type II pneumococcus. The specific polysaccharide in the B type in conjunction with the protein fraction stimulates the formation of an immune serum that will agglutinate pneumococcus Type II and will protect mice against infection with it. Similarly pneumococcus Type II serum will agglutinate Friedländer Type B and protect mice against infection with it. The polysaccharides in the two organisms appear to be closely alike though not absolutely identical (Avery *et al.* 1925). A Friedländer Type B organism will not however absorb the agglutinins from pneumococcus Type II serum nor a pneumococcus Type II organism from a Friedländer Type B serum. This probably indicates that though the polysaccharides are alike the protein fractions of the organisms are different.

According to Tomášek (1925) Quast (1926) Prica (1930) and Neuber (1934) the *rhinoscleroma* bacillus can be distinguished by agglutination and complement fixation from Friedländer's and the *ozæna* bacillus. Morris and Julhelle (1934) however who examined 10 strains of the *rhinoscleroma* bacillus were unable to distinguish this organism antigenically from strains of Friedländer's Type C. This observation is supported to some extent by Goslings and Snijders (1936) who found that all *rhinoscleroma* strains had the same capsular and the same somatic antigens. The capsular antigen was practically indistinguishable from the Type C Friedländer antigen.

According to Julhelle (1935) the *ozæna* bacillus can be distinguished by agglutination and absorption of agglutinins from Friedländer and *rhinoscleroma* bacilli. Of 19 *ozæna* strains studied 12 fell into one group 2 into another and the remaining 5 were antigenically heterogeneous. Goslings and Snijders (1936) likewise concluded that there were two and possibly three different capsular antigens among *ozæna* strains. These they labelled D, E and (F). All strains however appeared to share a common somatic antigen. Wielenga (1937) reports on three strains that possessed the somatic antigen of *Bact. ozænae* and the Type C capsular antigen of *Bact. friedländeri*.

Prašek and Prica (1933) state that they have been successful in extracting a soluble specific carbohydrate containing substance which is probably a galactan from the capsules of the *ozæna* and the *rhinoscleroma* bacillus. The substances from the two organisms were quite distinct and showed no cross precipitation when tested against the heterologous immune sera. They were likewise distinct from the polysaccharide extracted from a Friedländer's bacillus.

The relation between Friedländer and *aerogenes* strains is still obscure. Julhelle (1937) who studied 3 strains of *Bact. aerogenes* found that they shared a common somatic

antigen, which was different from the Friedlander somatic antigen, but that each had a distinct capsular antigen. One of the capsular antigens was specific, one was related to pneumococcus Type II and one to both pneumococcus Type II and Friedlander Type B. Goslings and Snijders (1936), Wiekema (1937), and Oortman and Reijer (1941) likewise found considerable antigenic complexity in the *aeropyrus* group. According to Goslings and Snijders, even the somatic antigen varies in different strains of *Bact. aeropyrus*, being sometimes similar to that of the rhinocleroma bacillus, sometimes to the osena bacillus, and sometimes to neither. The capsular antigen of one strain was similar to that of Friedlander Type B, but the rest were different from any of the A to F capsular antigens.

Studying capsulated strains of diverse origin, Edwards (1939) was able to divide them into three groups by agglutination: (1) strains, chiefly from human pneumonia and pleurisy, identical with Julianelle's Type A, (2) strains from infections of man, together with some strains of *Bact. aeropyrus* isolated from soil, water or milk, and an occasional strain of human origin, identical with Type B, (3) some soil strains of *Bact. aeropyrus* and an unusual granuloma strain. Julianelle (1931) obtained evidence that most Type A strains were of human, and most Type B strains of animal origin.

It is clear that the relationship of these various capsulated organisms to each other will not be understood until a sufficient number of fully representative strains of each group have been examined by serological and other methods.

The smooth type is capsulated, the rough type is not. It might therefore be thought that virulence depends on capsule formation. Toennessen (1914) discusses this possibility, but concludes that the association between capsule formation and virulence is fortuitous. He isolated, for example, one variant which, though non capsulated, was highly virulent. He states that old cultures of the smooth type, in which the capsules have largely become autolysed, have just the same virulence for mice as fresh young capsulated cultures. Moreover, Julianelle's Friedlander Type C strains, though capsulated, were non virulent. It would appear, therefore, that capsule formation is often associated with, but is not essential to virulence.

After subcutaneous injection of a very small dose—about 0.0000001 ml. of a 24 hours' broth culture of a virulent strain—into mice, the animals die in 12 to 72 hours. Post mortem, there is a local exudate, the focal glands are swollen, and the spleen is enlarged. Capsulated bacilli are found in the blood and viscera (Pfeiffer 1889, Fricke 1896, Toennessen 1914).

Guinea pigs are refractory to subcutaneous, but succumb to intraperitoneal injection, death occurring in 12 to 72 hours. The fatal dose is about 0.01 ml. of a 24 hours' broth culture. Post mortem, there is a viscous exudate in the peritoneum, the spleen may be enlarged, and the suprarenals hæmorrhagic. The bacilli are found in large numbers in the blood and viscera.

Rabbits appear to be more resistant, but they succumb after intravenous or intra-peritoneal injection with a dose of about 0.1 ml. of a broth culture. Intraperitoneal inoculation is likewise fatal to pigeons.

**Classification.**—The demarcation of this group from other groups of capsulated bacilli, and the subdivision of the group itself, are both in a very unsatisfactory state. In the first place it is doubtful what relation capsulated organisms of the *aerogenes* and intermediate type have to Friedlander's bacillus. It is usual to regard *Bact. aerogenes* as a saprophyte of grains, the intermediate types of coliform bacilli as saprophytes of soil, and the Friedländer *ozæna* rhinoscleroma group as parasites of man and animals. But the fact that most Friedlander strains of respiratory origin are indistinguishable from strains of intermediate I type (see Chapter 92) and that many strains found in cystitis appear to be identical with *Bact. aerogenes* renders dangerous any attempt to separate these organisms on the basis of habitat alone. It is difficult to avoid the conclusion that all these organisms should be classified in a single group, but whether that group should be called *Aerobacter*, *Encapsulatus*, or *Klebsiella* is very doubtful. For the moment we prefer to keep them within the wide *Bacterium* genus.

Attempts to make subdivisions within the group must necessarily await the definition of the group itself. Though Goslings (1934) and Wielenga (1937) are convinced of the value of biochemical tests in distinguishing between the scleroma *ozæna*, and *pneumonia* types, it is clear from a careful study of their papers that apart perhaps from the scleroma group, there is so much variation between the different members within each group as to make the classification of any individual strain of unknown origin often impossible. At the moment a study of antigenic structure seems to hold out the only promise of throwing any light on this problem. A careful comparison of adequate numbers of freshly isolated strains from different sources is urgently called for. Until this is done, it will be impossible to decide whether *ozæna* and rhinoscleroma strains are specifically distinct from strains of Friedlander, or whether they are merely types of the same species differing in the polysaccharide constituents of their capsule. Any such attempt should include a thorough study of intermediate and *aerogenes* strains.

Mention should perhaps be made here of an organism that appears to be responsible for joint ill of foals, and that is referred to by a variety of names, such as *B. nephritidis equi*, *B. equarius*, *Bact. viscosum equi*, and *B. pyosepticus equi*. Morphologically, this organism is a pleomorphic Gram negative bacillus occurring singly, in streptococcus-like chains, and as filaments. Some authors describe it as capsulated, others as non-capsulated. It forms tenacious colonies on agar, gives rise to a very viscous sediment in broth, gives a nail-head growth in gelatin stab without liquefaction, ferments glucose, maltose, mannitol, lactose, and sucrose with the production of acid but not gas, is M.R. —, V.P. —, citrate —, indole —, reduces nitrates to nitrites, produces acid, and sometimes clot, in litmus milk, is antigenically heterogeneous, and is non pathogenic to laboratory animals. The normal form on isolation is said to be mucoid, but a non mucoid variant is sometimes cultivated directly from foals, though more often it is seen only as the result of *in vitro* variation. (For references see Edwards 1931, 1932.) The exact relationship of this organism to the members of the Friedlander group is doubtful.

A detailed description of Friedlander's bacillus is given on p. 6-0

### The Paracolon Group

There remain a number of organisms which, for one reason or another, cannot be included in any of the lactose-fermenting groups already described, or in either of the non lactose-fermenting groups to be considered in the next two chapters. These organisms ferment lactose late, weakly, irregularly, or not at all. Some constantly give rise to non lactose-fermenting variants. Some produce gas abundantly, and some in only small quantity, others are completely anaerogenic. A few species are pathogenic for man; others are under suspicion, and others again are almost certainly non pathogenic. Some are found in faeces, some in water, some in soil, and some in other situations.

This heterogeneous collection of organisms we propose to refer to as paracolon bacilli. They constitute a group that appears to be intermediate between the coliform bacilli on the one hand and the non lactose-fermenting *Salmonella* and *Shigella* bacilli on the other. From the coliform bacilli they are distinguished mainly by their late fermentation of lactose, or by their failure to ferment it at all. From *Salmonella* and *Shigella* they differ mainly in fermenting either lactose, sucrose or salicin and in their lack of the particular antigens that characterize these two groups.

Paracolon bacilli have frequently been isolated from the faeces of persons suffering from enteric like infections enteritis and cystitis. Occasionally an organism belonging to this group has been isolated from the blood stream. In very many instances, however, they have been cultivated from the faeces of normal persons (see Sandiford 1935), and it seems doubtful whether they have any real significance as primary infecting agents in epidemic infections of the enteric or dysenteric type, though there can be no doubt that some species at least possess pathogenic potentialities when they invade the tissues from the intestinal tract. The great majority of the organisms included in this group have it may be noted, been isolated in the tropics, and there seems no doubt (Sandiford 1935) that they are a more common constituent of the normal intestinal flora under tropical than under temperate conditions.

The classification of these strains raises problems of considerable difficulty. Various schemes have been suggested (Chalmers and Macdonald 1916, Castellani and Chalmers 1920, Castellani 1935), but these are not in accordance with the general lines that we have discussed above, and in the authors' opinion lay too much stress on minor differences in fermentative ability. Stuart, Wheeler, Rustigian and Zimmerman (1943), who have subjected several members of this group to the series of tests used for differentiating

the coliform bacilli—MR, VP, indole, citrate, and cellobiose—find that they can be divided roughly into three main groups according to whether they resemble the *coli*, intermediate, or *aerogenes* types. Within the separate subdivisions many of the strains were found to be antigenically similar, but a number remained that could not be classified by serological means. For the sake of example the fermentation reactions of a few of the paracolon bacilli that have received names are set out in Table 44. For a fuller description of their characters the reader is referred to papers by Castellani (1902, 1907, 1912, 1938), Castellani and Chalmers (1920), Archibald (1911), Chalmers and Macdonald (1916), but it must be understood that the nomenclature of all these organisms may have to be revised when a satisfactory basis of classification is eventually laid down.

More recently Seritt (1945), who studied 108 strains of paracolon bacilli mostly isolated from cases of infantile diarrhoea, found that they could be classified into four groups on their reactions in sucrose and dulcitol. Serologically five main antigens could be defined though many strains were heterogeneous. Some strains besides containing one or more of the A-E series of antigens, were characterized by the additional inclusion of a *Shigella alkalescens* antigen, or of antigens found in the Flexner or Newcastle groups.

It is of interest to note that Dudgeon has recorded a considerable series of cases of acute urinary infection, associated with a pyrexial reaction simulating enteric fever and caused by a late lactose fermenting coliform bacillus. Lactose is fermented slowly, saccharose is unchanged, mannitol and dulcitol are fermented with the production of acid and gas, and litmus milk is rendered acid and usually clotted (Dudgeon 1924, Dudgeon and Pulvertaft 1927). It may be noted that this organism is hæmolytic for human red cells, and that Dudgeon and his colleagues (Dudgeon *et al.* 1921, 1922) have shown that strains of *Bact. coli* from the faeces, or from the urine in cases of cystitis may be divided into hæmolytic and non hæmolytic types. The hæmolytic strains are particularly frequent in certain types of acute urinary infection. It may clearly be necessary to elaborate our classification when these or other tests have been applied to the group as a whole. Paracolon bacilli have likewise been isolated by Webb (1937) in Mauritius from the urinary tract of patients suffering from enteric like fevers, pyelitis, or cystitis but they differed in some respects from the organism described by Dudgeon. Agglutinins were usually demonstrable in the serum of the patients in titres varying from 1/25 to 1/250.

The strains that have from time to time been described as *Bact. coli anaerogenes* on account of their failure to produce gas from certain carbohydrates are probably related either to some members of this group, or to the late lactose fermenting types of the *Shigella* group.

The curious organism described by Massani (1907) as *Bact. coli mutabile* is itself a non lactose-fermenting strain, but is characterized by the property of giving rise to lactose-fermenting mutants, which show no tendency to revert to the parent form. This species has been discussed in some detail in Chapter 9. It may be noted that Dulaney and Michelson (1935) have recently described a severe epidemic of diarrhoea in infants apparently caused by this organism. It is clearly allied to the paracolon group of *Bacteria*, though the exact relationship remains obscure.

It will be convenient to describe at this point an organism originally isolated by Castellani (1912), and studied in more detail by Khaled (1923). This species *Bact. asiaticum* is a non lactose-fermenter, and on this criterion would be excluded from the coliform group. In saccharose, however, it forms both acid and gas, and in its ability to attack this sugar it differs sharply from the gas forming non lactose fermenting bacilli of the paratyphoid group that will be considered in Chapter 30. It differs from them also in its ability to form indole. *Bact. asiaticum* is a motile bacillus having the usual characters of the genus. It ferments dextrose, mannitol and saccharose with the formation of acid and gas but produces no change in lactose or dulcitol. It acidifies, but does not clot milk. It usually forms indole. It does not liquefy gelatin. It appears to be a cause of enteric-like infections in man particularly in the tropics.

TABLE 44  
THE FERMENTATION REACTIONS OF SOME PARATYPIC BACTERIA

	Gas	Reaction	Glucose	Maltose	Dextrin	Mannitol	Inulin	Sucrose	Galactose	Milk	Starch	Glycerol	Cellulose	Mucic
<i>Bact. coagulans</i>	O or S	O	AG	AG	AG	AG	AG	AG	AG	A (S)	-	-	-	-
							(S)			alk				
<i>Bact. grisea</i>	A	O	AG	AG	O	O	AG	AG	A	alk	-	-	-	-
	(G.v.a.)						(S)			alk (S)				
<i>Bact. Hesperianus</i>	AG	O	AG	AG	AG	AG	O	AG	A	-	-	-	-	-
	(Slow)													
<i>Bact. weissenbergi</i>	A	AG	AG		A	A			A	-	-	-	-	-

Notes.—O or S = no fermentation, or very slight.

AG (?) = slight acid and gas.

A (G.v.a.) = acid, very little gas.

A, alk, or alk. or D = slight acid, or alkaline, or decarboxylated.

A alk (S) = acid, reverting to slight alkaline.

*Growth Requirements.*—*Bact coli* grows readily on all ordinary laboratory media. The optimum temperature for growth is in the neighbourhood of 37°C, but growth occurs over an extended range from about 15 to 45°C.

*Biochemical.*—Produces acid and gas in dextrose, maltose, mannitol, lactose, xylose, rhamnose, and arabinose, but not in dextrin, starch, inositol or as a rule cellobiose. Sucrose, salicin, raffinose, glycerol, and dulcitol are attacked by some strains but not by others. Acidifies and clots milk, reduces nitrates, and nearly always forms indole. MR +, VP —, Koser's citrate —, gas production in MacConkey's broth at 44°C +. H<sub>2</sub>S is not usually formed, but the result depends to some extent on the medium.

*Antigenic Structure.*—*Bact coli* is an antigenically heterogeneous species, the antigenic structure of which has not yet been studied in any detail.

*Pathogenicity.*—*Bact coli* is a normal inhabitant of the intestine of man and other animals. In certain circumstances it may play a pathogenic role, sometimes in the intestine itself, more commonly in organs or tissues anatomically related to it, such as the gall bladder. It is a frequent cause of infection of the urinary tract in man.

*Varieties.*—On the basis of the fermentation reactions, *Bact coli* may be divided into the following varieties:—

- var commune* ferments salicin but not saccharose
- var communis* ferments saccharose but not salicin
- var neapolitanum* ferments both saccharose and salicin
- var aculi-lactici* ferments neither saccharose nor salicin

Some authorities recognize further varieties which are differentiated by being non-motile.

#### *Bact. aerogenes*

*Bact aerogenes* differs from *Bact coli* in the following points. Morphologically the rods are often shorter and plumper and they are occasionally capsulated. They may be motile or non motile. The colonies on agar are more convex, smoother and often mucoid. The deposit in broth is often more viscous. Growth is rather more abundant at lower temperatures, less abundant at temperatures over 37°C, and usually very slight or absent at temperatures of 42–44°C. Inositol, cellobiose, dextrin and starch are frequently fermented. Most strains fail to form indole. There is a more abundant formation of gas but a lower acidity. The CO<sub>2</sub>:H<sub>2</sub> ratio is high, approximately 2:1. The Voges-Proskauer reaction is positive. The methyl red reaction is negative. Growth occurs with citrate as the only source of carbon. Gas is not produced in MacConkey broth at 44°C. The normal habitat of this species is not known with certainty. It is common on grains and plants, and is often found though usually only in small numbers in the faeces of man and animals, the balance of evidence however is against its being primarily an intestinal parasite.

#### *Bact. cloacae*

*Bact cloacae* resembles *Bact aerogenes* except that it is usually motile, is seldom capsulated, liquefies gelatin, but does not ferment glycerol or starch. Its distribution in nature appears to be the same.

#### Intermediate Forms

The existence of strains that are intermediate between *Bact coli* and *Bact aerogenes* has been noted in the text, and their characters have been described (see p. 663). They are distinguished (1) from *Bact coli* mainly in being citrate positive, in fermenting cellobiose, in being usually indole negative, and in failing to form gas in MacConkey broth at 44°C, and (2) from *Bact aerogenes* in being MR + VP —, and in failing to utilize uric acid as their sole source of nitrogen. In this country they appear to be distributed chiefly in the soil.

## Friedländer's bacillus

*Synonyms*—*Pneumobacillus*, *Bact. friedländeri*, *Bact. pneumoniae* *B. pneumoniae* *B. mucosus capsulatus* *Encapsulatus pneumoniae*, *Klebsiella pneumoniae*

*Isolation*.—Isolated by Friedländer in 1883 from the lungs of patients dying of pneumonia.

*Habitat*.—Chiefly a parasite. Found in the nose, mouth, and intestine of normal persons, in the lungs of patients with pneumonia, influenza and tuberculosis, and other respiratory diseases, and in suppurative conditions of other parts of the body.

*Morphology*.—Short, thick, oval rod, about  $1\frac{1}{2}\mu$  long and  $0.5-0.8\mu$  broad. Axis straight, sides parallel or bulging ends rounded, arranged singly and in pairs end-to-end. Some strains are highly pleomorphic, curved rods, sausage forms, and long wavy filaments being found in culture, in the body diplobacilli, very like pneumococci, are commonest. Considerable variation in staining, particularly of pleomorphic forms. Non-motile. A capsule is present even in cultures, it contains a nitrogen-free polysaccharide of glucose. Gram-negative.

*Agar Plate*.—24 hours at  $37^{\circ}\text{C}$  Round, amorphous, convex, greyish-white, faintly translucent mucoid colonies, 1-2 mm. in diameter, with smooth, glistening surface and entire edge, consistency mucoid, emulsifiability easy. 7 days, larger, from 3-10 mm. in diameter, raised, with flattened surface, sometimes studded with secondary colonies, and slightly undulate edge, sometimes differentiated into an opaque porcelain white centre and a less opaque, jelly-coloured periphery showing more translucent radial sectors. A non-mucoid variant occurs.

*Agar Slope*.—24 hours at  $37^{\circ}\text{C}$  Abundant, raised, faintly translucent, greyish-yellow, mucoid, almost diffuent growth, with glistening smooth or beaten-copper surface, and entire or undulate edge.

*Gelatin Slab*.—7 days at  $20^{\circ}\text{C}$  Moderate, filiform, greyish-white growth, confluent in the upper part, extending to bottom of tube. Convex, mucoid surface growth about 3 mm. in diameter—nail-headed appearance. No liquefaction, even after 4 weeks, but a large napiform gas bubble often appears near the surface. Sometimes numerous lateral outgrowths occur from the stab after 3 or 4 weeks.

*Broth*.—24 hours at  $37^{\circ}\text{C}$  Moderate growth with moderate uniform turbidity, and a slight, powdery deposit disintegrating easily. Ring growth at surface. 7 days, heavy turbidity with abundant, viscous deposit, marked ring growth. The culture is viscous in consistency.

*Glucose Agar Shake*.—24 hours at  $37^{\circ}\text{C}$  Multiple tiny colonies throughout medium, more numerous near the top. Medium is torn into rifts with gas.

*Horse Blood Agar Plate*.—24 hours at  $37^{\circ}\text{C}$  Convex, milky white colonies, 1 mm. in diameter, with smooth surface and entire edge. No hæmolysis, but plate is browned.

*MacConkey's Agar*.—24 hours at  $37^{\circ}\text{C}$  Peddlish colonies, 1-3 mm. in diameter. 7 days, round, convex or umbonate, red colonies with smooth surface and entire or lobate edge.

*MacConkey's Fluid Medium*.—7 days at  $37^{\circ}\text{C}$  Moderate turbidity, magenta colour, sometimes gas formation.

*Potato*.—24 hours at  $37^{\circ}\text{C}$  Yellowish, confluent, mucoid growth. 7 days, abundant raised, mucoid, creamy, buff yellow, or café-au-lait growth, with smooth, slightly pitted, or nodular surface.

*Persistence*.—Killed by moist heat at  $55^{\circ}\text{C}$ . in half an hour. Cultures at room temperature live for months.

*Metabolism*.—Aerobe. Very slight growth in 10 days under strict anaerobic conditions. Grows luxuriantly in culture. No hæmolysis of sheep's or horse's red cells. Tendency to form brownish pigment on potato.

*Biochemical*.—Highly variable. Acid, and generally gas, in glucose, maltose, mannitol, lactose, sucrose, and salicin. Inositol is said to be fermented. L.M. acid, or acid and clot. Indole  $\mp$ , M.R.  $\pm$ , V.P.  $-$ , nitrates reduced,  $\text{NH}_3$   $\pm$ .



H<sub>2</sub>S —, MB reduced, catalase ++, Koser's citrate +, gas in MacConkey broth at 44° C negative (For biochemical characters of rhinoscleroma and ozæna strains, see p 670)

**Antigenic Structure**—By agglutination, absorption, and precipitin tests there are three types, A, B, and C of Friedländer's bacillus, and a heterogeneous group X. The specificity of the types is determined by the carbohydrate fraction in the capsule. The B type is similar immunologically to Type II pneumococcus. (For antigenic structure of rhinoscleroma and ozæna strains see p 673)

**Pathogenicity**—Is found in lesions of the respiratory tract and in suppurative conditions generally in the human body. Experimentally, it is highly pathogenic to mice on intraperitoneal or intravenous injection. Rabbits are less susceptible, but often succumb to intravenous or intraperitoneal injection. Types A and B are highly virulent for mice, Type C is avirulent. After intraperitoneal injection of a very small quantity of broth culture mice die in 18 to 24 hours, PM viscous exudate in peritoneum, capsulated bacilli numerous in blood, lungs and spleen.

### The Paracolon Group

The organisms of this group which differ from *Bact coli* in fermenting lactose late, weakly, irregularly, or not at all and from the *Salmonella* and *Shigella* groups in fermenting either lactose, sucrose, or salicin, in usually producing indole, and in their antigenic structure, have been described on pp 670-8

### REFERENCES

- ABEL, R. (1896) *Z Hyg InfektKr.*, 21, 89  
 ANDERSON, A., DeMOLVEREUX, W. A. and GOODPASTURE, E. W. (1945) *J exp. Med.* 81, 25  
 ARCHIBALD, R. G. (1911) *4th Rep Wellcome trop Res Lab*  
 AVERY, O. T., HEIDELBERGER, M., and GOEBEL, W. F. (1926) *J exp Med.*, 42, 709  
 BANFORTH, J. (1928) *J Hyg, Camb.*, 27, 343  
 BARBER, M. A. (1907) *Kansas Univ Sci Bull.*, 4, 1  
 BARDLEY, D. A. (1926) *J Hyg, Camb.*, 25, 11, (1934) *Ibid.* 34, 38, (1935a) *Ibid.* 38, 309, (1935b) *Ibid.* 38, 721  
 BATTY SMITH, C. G. (1942a) *J Path Bact.*, 54, 45, (1942b) *J Hyg, Camb.*, 42, 55  
 BEHAM, L. M. (1912) *Zbl Bakt.*, 66, 110  
 BERKEY, D. H., BREED, R. S., MURRAY, E. G. D. and HITCHENS, A. P. (1939) *Manual of Determinative Bacteriology* 5th Edit. Baillière Tindall and Co. London.  
 BERKEY, D. H. and DEEMAN, S. J. (1908) *J med Res.*, 19, 175  
 BHATTAGAR, S. S. and SINGH, K. (1935) *Indian J med Res.*, 23, 337  
 BOZOT, G. E. (1941) *J Hyg, Camb.*, 41, 566  
 BREIDENBÖCKER, J. (1938) *Arch Hyg Berl.*, 119, 289  
 BREED, R. S. and COOK, H. J. (1936) *J Bact.*, 31, 517  
 BROWN, H. C. (1921) *Lancet*, i, 22  
 BROWNING, C. H., GILMOUR, W., and MACKIE, T. J. (1913) *J Hyg Camb.*, 13, 335  
 BURTON, L. V. and RETTOER, L. F. (1917) *J infect Dis.* 21, 162.  
 CASTELLANI, A. (1900-14) *Ceylon med Rep Colombo* (1907) *J Hyg, Camb.*, 7, 1, (1912) *J trop Med Hyg.*, 15, 161, (1929) *Ibid.*, 41, 325, 344, 362  
 CASTELLANI, A. and CHALMERS, A. J. (1920) *Ann Inst Pasteur*, 34, 600  
 CHALMERS, A. J. and MACDONALD, N. (1916) *Lancet* ii 139  
 CHANTENESSE, A. and VIDAL, F. (1887) *Arch Physiol norm path.*, 9, 217, (1891) *Bull Méd.*, 5, 935  
 CHEN, C. C. and RETTOER, L. F. (1920) *J Bact.*, 5, 253  
 CLARKE, P. (1902) *Z Hyg InfektKr.*, 39, 1  
 CLARK, W. M. and LURS, H. A. (1915) *J infect Dis.*, 17, 160  
 CLEGG, L. F. L. (1941) *J Path. Bact.* 53, 51  
 CLEGG, L. F. L. and SHERWOOD, H. P. (1939) *J Hyg, Camb.*, 39, 361  
 COLLINS, G. (1924) See HADLEY, P. (1927) *J infect Dis.*, 40, 1  
 COULTER, C. B. (1917) *J exp Med.*, 26, 763  
 CRUICKSHANK, J. and CRUICKSHANK, R. (1931) *A System of Bacteriology*, "Med Res Coun, Lond.", 8, 353  
 DODGSON, R. W. (1938) *Proc R Soc Med.*, 31, 920

- DOWSON, W J (1939) *Zbl Bakt., nte Abt.*, 100, 177  
 DUDGEON, L S (1924) *J Hyg., Camb.*, 22, 348, (1926) *Ibid.*, 25, 119  
 DUDGEON, L S and PULVERTAFT, P J V (1927) *J Hyg., Camb.*, 26, 285  
 DUDGEON, L S, WORDLEY, E., and BAWTREE, F (1921) *J Hyg., Camb.*, 20, 137, (1922) *Ibid.*, 21, 168.  
 DULANEY, A D and MICHELSON, I D (1935) *Amer J publ Hlth.*, 25, 1241  
 DUNBAR, W (1892) *Z Hyg InfektKr.*, 12, 485  
 DURHAM, H E (1898) *Brit med J.*, 1, 1387, (1901) *J exp Med.*, 5, 353  
 EDWARDS P R. (1928) *J Bact* 15, 245, (1929) *Ibid.*, 17, 339, (1931) *Kentucky agric Exp Sta Bull.*, No 320, (1932) *J infect Dis.*, 51, 268  
 EDWARDS, R T (1906) *J infect Dis.*, 2, 431  
 ELKMAN, C (1904) *Zbl Bakt.*, 37, 436-742, (1914) *Ibid.*, 11to Abt., 39, 75  
 ELBERT B J and GUERKESS, W M. (1930) *Ann. Inst Pasteur.*, 44, 548  
 ELBOD R P (1947) *J Bact.*, 44, 433  
 ESCHERICH, T (1880) *Fortschr Med.*, 3, 515, 547  
 FERRAMOLA, R (1940) *Amer J publ Hlth.*, 30, 1083  
 FITZGERALD, J G (1914) *J infect Dis.*, 15, 268  
 FREMLIN, H (1893) *Arch Hyg., Berl.*, 19, 293  
 FRICKE C (1896) *Z Hyg InfektKr.*, 23, 380  
 FRIEDLÄNDER, C (1883) *Fortschr Med.*, 1, 715.  
 GARDNER, A D (1925) *J Path Bact.*, 28, 189  
 GOSLINGS, W R O (1934) *Zbl Bakt.*, 123, 33, (1935) *Ibid.*, 124, 193  
 GOSLINGS W R O and SWIDERS, E P (1936) *Zbl Bakt.*, 136, 1  
 GRAAFF, W C DE (1936) *Ant v Leeuwenhoek ned Tijdschr Hyg.*, 3, 18  
 GRAY, J D A (1931) *J Path Bact.*, 34, 335  
 GRIFFIN, A M and STUART, C A (1940) *J Bact.*, 40, 83.  
 GRIMBERT, L. and LEGROS G (1900) *C R Soc. Biol.*, 52, 491  
 GUTH F (1916) *Zbl Bakt.*, 77, 457  
 GWATRIN, R, LEGARD, H. M., and HADWEN, S (1938) *Canad J comp Med.*, 2, 155  
 HADLEY, P (1927) *J infect. Dis.*, 40, 1  
 HARDEN, A. (1901) *J chem Soc.*, 79, 610, (1905) *J Hyg., Camb.*, 5, 488, (1906) *Proc. roy Soc., B.*, 77, 424  
 HARDEN, A and NORRIS, D (1911) *Proc roy Soc., B.*, 84, 492.  
 HARDEN, A and WALFOLE, G S (1906) *Proc roy Soc., B.*, 77, 399  
 HAVENS, L C. and MAYFIELD, C. R (1933) *J infect Dis.*, 52, 157  
 HAY H R (1932) *J Hyg., Camb.*, 32, 240.  
 HEIDELBERGER, M., GOEBEL, W. F., and AVERY, O T (1920) *J exp Med.*, 42, 701  
 HENYEBERG W and WENDT H. (1935) *Zbl Bakt., nte Abt.*, 93, 39  
 HOFFMAN, W and PICKER, M. (1904) *Hyg Rundsch.*, 14, 1  
 HORT, E C. (1920) *J Hyg., Camb.*, 18, 369  
 HOWE F (1904) *Zbl Bakt.*, 36, 454  
 HULTON, F (1916) *J infect Dis.*, 19, 606.  
 HYNES, M (1942) *J Path Bact.*, 54, 193  
 JACKSON, D D (1911) *J infect Dis.*, 8, 241  
 JAMPOLIS, M., HOWELL, K. M., CALVIN, J K., and LEVENTHAL, M. L. (1932) *Amer J. Dis Child.*, 43, 70  
 JOHNSON, B R (1916) *J Bact.*, 1, 96  
 JONES, H N (1924) *Science.*, 60, 455  
 JONES, H N and WISE, L E (1926) *J Bact.*, 11, 359  
 JORDAN, E O (1890) *Report on Water Supply and Sewerage, Mass Bd Hlth.*, Pt. 2, p 836, (1903) *Science.*, 3, 1  
 JULIANELLE, L A (1926a) *J exp Med.*, 44, 113, (1926b) *Ibid.*, 44, 683, (1926c) *Ibid.*, 44, 735, (1930) *Ibid.*, 52, 539, (1935) *J Bact.*, 30, 535, (1937) *J Immunol.*, 32, 21.  
 KEYES, F G (1909) *J med Res.*, 21, 69  
 KHALED, Z (1923) *J Hyg., Camb.*, 21, 362  
 KLEWE, H and HSÜ, M. (1935) *Z ImmunForsch.*, 88, 481  
 KLIGLER, I J (1914a) *J infect. Dis.*, 14, 81, (1914b) *Ibid.*, 15, 187  
 KLINE, E K. (1935) *Amer J publ Hlth.*, 25, 833  
 KNOX, R, GELL, P G H and POLLOCK, M. R (1943) *J Hyg., Camb.*, 43, 147  
 KOSER, S A (1918) *J infect. Dis.*, 23, 377, (1923) *J Bact.*, 8, 493, (1924) *Ibid.*, 9, 59, (1926a) *J Amer Wat Wks Ass.*, 15, 641, (1926b) *J. Bact.*, 11, 400, (1926c) *J. infect. Dis.* 38, 506  
 KRAMÁR, E (1921) *Zbl Bakt.*, 87, 401  
 KUCHWILDE, C and PRATT, J S (1914) *J exp Med.*, 19, 501  
 KRUSE W (1894) *Z Hyg InfektKr.*, 17, 1  
 LEIFSON, E (1933) *J Bact.*, 26, 329, (1935) *J Path Bact.*, 40, 581, (1936) *Amer J. Hyg.*, 24, 423

- LENTZ, O and TIETZ, J (1903) *Munch med Wochr*, 50, 2139. (1905) *Klin Jb*, 14, 405
- LEVINE, M (1916a) *J Bact*, 1, 87. (1916b) *Ibid*, 1, 153. (1916c) *J infect Dis*, 18, 358. (1916d) *Ibid*, 19, 773. (1917) *Amer J publ Hlth*, 7, 784. (1918) *J Bact*, 3, 253. (1941) *Amer J. publ Hlth*, 31, 351
- LEVINE, M, EPSTEIN, S. S. and VACHN, R. H (1934) *Amer J publ Hlth*, 24, 505
- LOEFFLER, F. (1903) *Dtsch med Wochr*, 29, 36. (1906) *Ibid*, 32, 289
- LOEWENBERG (1894) *Ann Inst Pasteur*, 8, 292
- LOVELL, R (1937) *J Path Bact*, 44, 125
- MACCONKEY, A (1905) *J Hyg, Camb*, 5, 333. (1909) *Ibid*, 9, 86
- MACKENZIE, E F W and HILTON SERGEANT, F C (1938) *J R Army med Cps*, 70, 14 73
- MACHIE, T J (1913) *J Path Bact*, 18, 137
- MAFFREDI, L (1917) *Rif Med*, 33, No 35 849
- MASSINI, R (1907) *Arch Hyg, Berl*, 61, 250
- MIRRA, G (1936) *Ann Med nat colon*, 42, 205
- MITCHELL, N B and LEVINE, M (1938) *J Bact*, 36, 587
- MORRIS, M C and JULIANELLE, L A (1934) *J infect Dis*, 55, 150
- MULLER, L (1923) *C R Soc Biol*, 89, 434
- NEUBER, E (1934) *Arch Derm Syph, Wien*, 170, 154
- OSTERMAN, E and REYTOER, L F (1941) *J Bact*, 42, 609, 721
- PAGE, C G (1912) *J med Res*, 26, 489
- PARR, L W (1937) *Proc Soc exp Biol, N Y*, 35, 563. (1939) *Amer J publ Hlth*, 28, 445. (1939) *Bact Rev*, 3, 1
- PAVAN, J L (1931) *J trop Med Hyg*, 34, 229, 267, 288, 310, 317 345, 360 380, 391, 413
- PEREZ, F (1899) *Ann Inst Pasteur*, 13, 937
- PERKINS, R G (1904) *J infect Dis*, 1, 241. (1907) *Ibid*, 4, 51
- PERRY, C A (1939) *Food Res*, 4, 381
- PETRUSCHKE, J (1889) *Zbl Bakt*, 6, 625, 657. (1890) *Ibid*, 7, 49
- PREIFFER (1889) *Z Hyg InfektKr*, 6, 145
- PLJFER, A. (1938) *J Path Bact*, 47, 1
- PLATT, A E (1935) *J Hyg, Camb*, 35, 437
- POGES, O (1905) *Wien klin Wochr*, 18, 691
- PRÁŠEK, E and PRICA, M (1933) *Zbl Bakt*, 128, 381
- PRICA, M (1930) *Zbl Bakt*, 115, 334
- QUAST, G (1920) *Zbl Bakt*, 97, 174
- RAGHAVACHARI, T N S and IYER P V S (1939a) *Indian J med Res*, 26, 867 (1939b) *Ibid*, 28, 877
- RAVEN, C, PEDEV, D, and WRIGHT, H D (1940) *J Path Bact*, 50, 287
- REYK, E (1896) *Ann Inst Pasteur*, 10, 242
- RENNENBAUM, E H (1935) *J Bact*, 30, 625
- ROGERS, L A, CLARK, W M and DAVIS B J (1914) *J infect Dis*, 14, 411
- ROGERS, L A, CLARK, W M, and EVANS, A C (1914) *J infect Dis*, 15, 99. (1915) *Ibid*, 17, 137
- ROGERS, L A, CLARK, W M, and LUBS, H A (1918) *J Bact*, 3, 231
- ROTH, E (1903) *Hyg Rundsch*, 13, 489
- ROTHBERGER, C J (1898) *Zbl Bakt*, 24, 513
- RUSSELL, H L and BASSETT, V H (1899) *Proc Amer publ Hlth Ass*, 25, 570
- SANDIFORD, B R (1935) *J Path Bact*, 41, 77
- SEVITT, S (1945) *J Hyg, Camb*, 44, 37
- SHERWOOD, H P and CLEGG, L F L (1942) *J Hyg, Camb*, 42, 45
- SMALL, J C and JULIANELLE, L A (1923) *J infect Dis*, 32, 456
- SMITH, D E (1927) *J exp Med*, 46, 155
- SMITH, T (1890) *Zbl Bakt*, 7, 502. (1893) *Misc Invest infect parasit Dis Dom Animals*, 8<sup>o</sup> Washington, 53. (1895) *Amer J med Sci*, 110, 283
- SMITH T and LITTLE, R B (1922) *J exp Med*, 38, 181 453. (1923) *Ibid*, 37, 671. (1927) *Ibid*, 46, 123
- SMITH, T and ORCUTT, M L (1925) *J exp Med*, 41, 89
- STAMP and STOVE, D M (1944) *J Hyg, Camb*, 43, 206
- STEINBERG, B and ECKER, E E (1926) *J exp Med*, 43, 443
- STREIT, H (1906) *Zbl Bakt*, 40, 709
- STUART, C A, BAKER, M, ZIMMERMAN, A, BROWN, C, and STONE, C M (1940) *J Bact*, 40, 101.
- STUART, C A, GRIFFIN, A M, and BAKER, M E (1938) *J Bact*, 36, 391
- STUART, C A, WHEELER, K M, RUSTIGIAN, R, and ZIMMERMAN, A (1943) *J Bact*, 45, 101
- STUART, C A, ZIMMERMAN, A, BAKER, M, and RUSTIGIAN, R (1942) *J Bact*, 43, 537
- TITSLER, R P (1933) *J Bact*, 35, 157
- TITSLER, R P and SANDHOLZER, L A (1936) *J Bact*, 31, 301

- TORNWISSEN E. (1912) *Zbl. Bakt.*, 65, 23. (1913) *Ibid.*, 63, 301. (1914) *Ibid.*, 73, 241.  
(1921) *Ibid.*, 85, 223.
- TOMÁSEK, V. (1925) *Zbl. Bakt. Ref.*, 78, 364.
- VAUGHN, R. and LEVINE, M. (1936a) *J. Bact.*, 31, 24. (1937) *Ibid.*, 32, 65.
- VAUGHN, R., MITCHELL, N. B., and LEVINE, M. (1937a) *J. Amer. Water Works Ass.*, 31, 992.
- VOGES, O. and PROSKAUER, R. (1894) *Z. Hyg. Inf. Bkt.*, 28, 21.
- WALLACE, G. L., CAHN, A. E., and THOMAS, L. J. (1923) *J. Hyg. Camb.*, 53, 206.
- WEER, J. L. (1937) *J. Hyg. Camb.*, 57, 37.
- WEBSTER, L. T. (1924) *J. exp. Med.*, 47, 605. (1937) *Ibid.*, 52, 499.
- WERKMAN, C. H. and GILLES, G. F. (1932) *J. Bact.*, 23, 16.
- WIELINGA, D. K. (1937) "Onderzoekingen over het Sclerotium respiratorium en de Group der Kapselhakten." N V Noord-Hollandische Geneesmaatschappij, Amsterdam.
- WILSON, G. A., TWIDG, R. S., WRIGHT, E. C., HENNEY, C. R., COWELL, M. P., and MATH, L. (1935) *Spec. Rep. Ser. med. Res. Coun., Lond.*, No. 24.
- WILSON, W. J. (1935) *J. Hyg. Camb.*, 33, 404.
- WILSON, W. J. and BLAIR, E. M. Mc V. (1941) *Dis. med. J.*, n. 561.
- WINSLOW, C. E. A., BRADDEST, J., BUCHANAN, R. E., KIRKWHIRE, C., ROGERS, L. A. and SMITH, G. H. (1917) *J. Bact.*, 2, 405. (1920) *Ibid.*, 5, 191.
- WINSLOW, C. E. A., KILGIER, I. J., and POTTSFORD, W. (1919) *J. Hyg.*, 4, 429.
- WINSLOW, C. E. A. and LOCKHART, E. E. (1906) *J. Hyg. Camb.*, 3, 44.
- WINSLOW, C. E. A. and WALKER, L. T. (1917) *Science*, 28, 78.
- WRIGHT, J. H. and MALLORY F. B. (1897) *Z. Hyg. Inf. Bkt.*, 22, 207.
- WURTZ, R. (1897) *Arch. Med. exp.*, 4, 5a.

## CHAPTER 29

### SHIGELLA

#### DEFINITION—*Shigella*

Gram negative non motile rods 2-3  $\mu$  long by 0.5-0.7  $\mu$  broad. Non capsulated, non sporing. Ferment a variable number of carbohydrates with the production of acid. Lactose is not attacked except by a few species and then not for two days or more. Reduce nitrates to nitrites, form ammonia but not hydrogen sulphide, are Voges Proskauer negative, and fail to grow in Koser's citrate. Facultative anaerobes. Some species are antigenically related. At least one species produces a toxin. Most species are pathogenic to man, giving rise to dysentery or sometimes acute gastro enteritis. Found as a rule in the intestinal tract of human dysentery patients and contacts.

Type species, *Shigella shiga*

Without entering here into the vexed question of classification (see p. 696), we propose to discuss in this chapter the following organisms or groups of organisms: (1) *Sh. shiga*, described by Shiga (1898a, b, 1901) in Japan and Kruse (1900-1901) in Germany, (2) *Sh. schmitzi*, isolated by Schmitz (1917) in Rumania and called *B. ambiguus* by Andrewes (1918), (3) Other non mannitol fermenters described by Dudgeon and Urquhart (1919) in Macedonia, by Large and Sankaran (1934) and by Sachs (1913) in India by Hazen (1933) in the United States and by Ali (1933) in Egypt, and referred to as para Shiga bacilli, (4) *Sh. flexneri* described by Flexner (1900a, b) in the Philippines, and Strong and Musgrave (1900) in Manila, (5) The organism isolated by Clayton and Warren (1929a, b) known as the Newcastle bacillus, by Downie, Wade and Young (1933) known as the Manchester bacillus, and by Boyd (1931, 1932) in India known as 88, (6) A group of mannitol fermenters resembling *Sh. flexneri* biochemically, but differing from it antigenically, described by Boyd (1931, 1932, 1936, 1938) in India under the names 170, P238, and D1, (7) *Sh. alkalescens*, described by Andrewes (1918), (8) *Sh. sonnei* a late lactose fermenting organism defined by Sonne (1915) in Denmark, though almost certainly described by previous workers in the United States and Germany (see Koser *et al.* 1930, Boylen 1934) and probably identical with Kruse's Type E bacillus, (9) *Sh. dispar*, a late lactose fermenting organism described by Andrewes (1918), and possibly related to the organisms described by Castellani (1907, 1912b) as *B. ceylonensis* B and *B. madampensis*. A few other organisms of less importance will receive occasional mention.

**Morphological and cultural Characters**—Dysentery bacilli are non motile, Gram negative organisms indistinguishable morphologically from members of the *Bacterium* group. Their cultural characteristics are likewise insufficiently distinctive to require separate description from those of the *Bacterium* and *Salmonella* groups. It may be noted, however, that the colonies of Sonne's bacillus tend to

be larger and more opaque than those of the Shiga Flexner types. On primary cultivation from the faeces this organism forms colonies on agar that are circular convex, 2 mm. in diameter colourless fairly translucent with a smooth surface and entire edge when viewed against a dark background they appear whitish and relatively opaque. Some colonies show at one or more portions of the periphery a small tangled hair like projection giving to the colony a bursting bombshell appearance (Braun and Weil 1925). Sometimes on first isolation but nearly always in early subculture a second type of colony is seen which is larger raised translucent with a coarsely ground-glass surface and an irregularly undulate or crenated edge when viewed against a dark background it appears clearer and less opaque than the first type. These two types of colony which consist of organisms differing antigenically and in certain other respects have been described by several workers (Mits 1921 Orskov and Larsen 1925 Leuchs and Plochmann 1925 Frye 1927 Large 1929 Cann and de Navasquez 1931 Thjotta and Waaler 1932 Johnston and Kaake 1932 Waaler 1932 Glynn and Starkey 1939). They are generally regarded as corresponding to smooth and rough forms though as Bridges (see Report 1939) points out the antigenic change that accompanies the colonial transition resembles in some respects the specific-group rather than the smooth rough type of variation.

Morphological and cultural variants of the dysentery bacilli have been described by several workers. Differences have often been noted in the antigenic structure saline and acid agglutinability and pathogenicity for laboratory animals of the rough and smooth variant. Indeed as in members of the *Salmonella* group antigenic structure is of far more importance in defining smooth and rough types than the colonial appearance. (For references to variation see Arkwright 1931 Carver 1931 Isabolinsky 1936 Koser and Stron 1930 Braun and Baake 1930 Kobayashi et al 1931 Johnston and Kaake 1932 Wyckoff 1933 Wasler 1933.)

**Resistance and Metabolism.**—The members of this group are not specially resistant. They are killed by a temperature of 50° C in 1 hour by 0.5 per cent phenol in 6 hours and by 1 per cent. phenol in about 15–30 minutes. When dried on linen and kept in the dark at room temperature they survive for from 5 to 46 days (Vaillard and Doptier 1903 Roelcke 1933). In garden earth at room temperature in the dark they survive for 9 to 13 days (Roelcke 1933). In naturally infected faeces kept alkaline and prevented from drying they may remain alive for some days, but in stools that are allowed to become acid through growth of coliform or other bacilli, they perish often in a few hours. On the whole *Sonnea* bacillus is more resistant to inimical agents generally than the Shiga or Flexner bacillus.

They are aerobes and facultative anaerobes. Their optimum temperature is about 34° C. According to Braun and Weil (1925) *Sonnea* bacillus grows as readily at 45° C as at 34° C and more readily at 10° C. than most coliform bacilli. Shiga bacillus is characterized by its inability to form catalase.

With the exception of some strains of *Sh. alkalescens* none of the members appears capable of producing a haemolysin active against sheep corpuscles.

**Biochemical Reactions.**—*Sh. shiga*, *Sh. sonnei* and members of the para Shiga group produce acid from glucose the remaining members, with the exception of some strains of the Newcasttle bacillus also ferment mannitol. Hence, the primary classification of the group is into mannitol and non-mannitol fermenters. *Sh. sonnei* and *Sh. dispar* produce acid from lactose the fermentation, however

is slow and is not usually apparent for 2 to 10 days. Lactose fermenting papillae often develop on the original lactose-negative colonies obtained by primary plating. Sucrose is also fermented late, though, according to Forsyth (1933), *Sh. dispar* acidifies sucrose before lactose. Sucrose is attacked by a small minority of Flexner strains after subculture in the laboratory for a variable length of time, but never on first isolation. Dulcitol is fermented rapidly by *Sh. alkalescens*, and slowly by the Newcastle bacillus and the *dispar* like organism known as *Bact. ceylonense* B. The Newcastle bacillus is distinguished from other members by its ability to form gas in glucose and dulcitol. The quantity of gas is very small, often amounting to not more than a bubble in a Durham fermentation tube and may be evident only on first isolation. The closely allied Manchester bacillus forms a trace of gas in mannitol as well as in glucose and dulcitol, but the antigenically similar organism met with in India and in the United States, known as Type 88 forms no gas at all. *Sh. alkalescens* is distinguished from *Sh. flexneri* by the fermentation of dulcitol and xylose, but according to de Azavedo (1939a) the substance of greatest differential value is 5 per cent glycerol, which is always attacked by the *alkalescens* and not by the *flexneri* type. *Sh. alkalescens* is said by Wood and Keeping (1914) to be unique among the dysentery bacilli in its ability to form trimethylamine from choline. *Sh. dispar* ferments xylose and sorbitol, thus differing from *Sh. sonnei*, it must be pointed out, however, that xylose fermenting strains of *Sh. sonnei* do occur (Bojlin 1931, Cruickshank and Swyer 1910), though the great majority of strains isolated in Great Britain and the United States lack the power to ferment this sugar. In litmus milk there is generally a slight acidity for a few days. The reaction may remain permanently acid and go on to clotting, as with *Sh. sonnei* and *Sh. dispar*, or it may revert to neutral, as with *Sh. shigae*, *Sh. schmitzi*, and *Sh. flexneri*. Many strains of *flexneri*, after a preliminary acidity turn milk alkaline. *Sh. alkalescens* produces an initial and lasting alkalinity.

Indole formation is of some differential importance, serving to distinguish Schmitz's from Shiga's bacillus, and *Sh. dispar* from *Sh. sonnei*. The Newcastle bacillus does not form indole, but *Sh. alkalescens* does. As regards the Flexner group, Gettings (1919) found that of 285 strains tested, 158 produced indole and 127 did not. The methyl red test is of limited value, if the cultures are incubated at 37° C, it may help to distinguish the positive *Sh. dispar* from the negative *Sh. sonnei*, but, according to Bamforth (1931), *Sh. sonnei* may give a positive result if it is incubated at 30° C.

All strains reduce nitrates to nitrites, none forms  $H_2S$ , none grows in Koser's citrate, and none gives a positive Voges Proskauer reaction. A list of biochemical reactions is given in Table 45. (For reference to the more recent studies on these reactions see Lester 1926, Smith and Fraser 1928, Kernin 1928, Nelson 1930, Bojlin 1930, 1934, Johnston and Brown 1930, Buchanan and Roux 1930, Koser *et al* 1930, Cann and de Navasquez 1931, Welch and Mickle 1932, 1934, Downie *et al* 1933, Forsyth 1933, Bamforth 1934, Whitehead and Scott 1934, Large and Sankaran 1934, Mandry 1935, McGinnes *et al* 1936, Boyd 1931, 1932, 1936, 1938, Hazen 1938, Ali 1938, Hardy *et al* 1940, Sachs 1943).

**Antigenic Structure**—The serological behaviour of the dysentery bacilli is complicated. Of the non mannitol fermenters the Shiga group is homogeneous, all strains of *Sh. shigae* are agglutinated by a specific serum prepared against any one strain. An anti shigae serum has some agglutinating action on Schmitz's bacillus and on some strains of the Flexner group. A serum prepared against Schmitz's

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TABLE 45

## FERMENTATION REACTIONS OF DYSENTERY BACILLI

With the possible exception of the Newcastle bacillus which has been reported by one worker to be mobile all strains are non-motile reduce nitrates are V.P. reduce methylene blue in broth cultures and form  $\text{NH}_3$  none forms  $\text{H}_2\text{S}$  or grows in flower a citrate

Organism	Acid to					Nitrogen	Soluble	Lithmus	Indole	Milk	Casein	Remarks
	Glucose	Mannitol	Lactose	Sucrose	Dextrose							
<i>Sh. dysenteriae</i>	+	-	-	-	-	-	-	sl. A	-	-	-	Produces exotoxin
<i>Sh. sonnei</i>	+	-	-	-	-	+	+	sl. A	+	-	+	
<i>Para Sh. dysenteriae</i>	+	-	-	-	-	-	+	sl. A	+	+	+	May be subcultured on lysate of arabinose and in bile Maltose may be fermented
<i>Sh. newcastle</i>	+ g	+	-	-	+ R late	-	-	sl. A → sl. alk	-	+	+	This Newcastle type does not ferment mannitol the Maltose ester type does, Type 88 forms no gas
<i>Sh. flexneri</i>	+	+	-	-	-	+	+	sl. A → sl. alk	+	+	+	Does not ferment glycerol Sucrose may be fermented by all laboratory cultures
<i>Sh. sp. ?</i> by Tyson 1928 DL, 1924	+	+	-	-	± late	+	+	sl. A → sl. alk	-	+	+	Type 170 is dialed negative, and turns milk first acid then alkaline
<i>Sh. allactosea</i>	+	+	-	-	-	+	+	Alk	+	+	+	Always ferments glycerol in 10 days but not dextrin Ferments trimethylamine from choline
<i>Sh. sonnei</i>	+	+	+ late	+ late	-	+	+	Alk chit	-	-	+	
<i>Sh. dysenteriae</i>	+	+	+ late	+ late	+	+	+	Alk chit	+	+	+	

sl = slight A = acid Alk = alkaline g = small amount of gas v = variable ± More often than not T Usually not

bacillus will agglutinate a Shiga bacillus to  $\frac{1}{2}$  or  $\frac{1}{4}$  titre, but antigenically Schmitz's bacillus and Shiga's bacillus are easily distinguishable, a Shiga bacillus cannot absorb the agglutinins from a serum prepared against Schmitz's bacillus, nor a Schmitz bacillus from an anti shiga serum. According to Schutze (1944) Shiga strains vary in their agglutinability, hypoagglutinable strains may be rendered more agglutinable by growth at 20-26° C, by heating in saline to 60° C for 1 hour, or by adding 0.5 per cent phenol.

Schmitz's bacillus used to be regarded as antigenically homogeneous, but the observations of Boyd (1935) suggest that the freshly isolated organisms contain two antigens which, for convenience, may be referred to as type and group. It is believed that after some time in the laboratory the type antigen is lost and the group alone remains. A serum, therefore, prepared against freshly isolated bacilli will agglutinate both recent and old strains, a serum prepared against old strains will have little or no action on recent strains. The picture seems to be very similar to that which will be described below for the 103 type of Flexner's bacillus, though it is still doubtful whether the variation is to be regarded as of the type and group or the smooth and rough order.

Little is yet known of the antigenic structure of the para Shiga group of dysentery bacilli. Dudgeon and Urquhart (1919), who studied 11 strains found them to be antigenically alike. Similar findings were reported by Hazen (1938). Ali (1938), however, was able by agglutination and absorption of agglutinins to distinguish four serological groups among 8 strains, and Sachs (1943) was able to distinguish eight serological groups among 107 strains (see also Large and Sankaran 1934, Christenson and Gowen 1944). It would appear that the para Shiga group of bacilli is far from being antigenically homogeneous. None of the members appears to be related to either *Sh. shiga* or *Sh. schmitzi*.

The mannitol fermenting group has long been recognized as antigenically heterogeneous. The work of Gettings (1919), Murray (1918), and Andrewes and Inman (1919) afforded a picture of the Flexner bacillus as containing at least four antigenic components.

According to Andrewes and Inman, each of these components which they refer to as V, W, X and Z, is represented to some extent in every strain, but in any given strain there is usually a preponderance of one antigen over the rest. In certain races, V, W, and Z, there is so great a preponderance of a single antigenic component, different in each instance, over the rest as to make them behave like distinct serological types, each race requires its own antiserum for adequate agglutination. The X race is peculiar in that it will not agglutinate with any sera but its own, yet it is able to give rise to a serum that will agglutinate not only X races, but also Z, and, to a certain extent, V races. The agglutinins corresponding to each of these four types cannot be more than partially absorbed by the others. Andrewes and Inman found at least two sub races, VZ and WX, these were members of the V and W races respectively, but contained so large a proportion of a second antigenic constituent as to modify their serological behaviour. One race, which is called Y, and which corresponds to the original Y strain of Hiss and Russell (1903), contains a more evenly balanced mixture of V, W, and Z components, with a small amount of X. For this reason a serum prepared against a Y strain is more cosmopolitan than the rest, having a wide range of agglutination (Fig 144).

This conception has been challenged by the work of Boyd (1931, 1932, 1936,



FIG 144.—DIAGRAM REPRESENTING THE VARIATION IN ANTIGENIC STRUCTURE OF *Shigella flexneri*  
(After Andrewes.)

1938) in India. Studying a strain of dysentery bacillus known as 103, which was biochemically similar to Flexner's bacillus, but which, when newly isolated, was not agglutinated by antisera to the V, W, Y, and Z races and only very feebly by an X antiserum, Boyd observed that after some time in artificial culture it gave rise to two types of colony. One of them, referred to as 103A, was in all respects identical with the smooth circular colonies of the freshly isolated strain and was virtually inagglutinable by V to Z antisera. The other, referred to as 103B, was slightly larger than normal and somewhat rough in outline and was agglutinated readily by antisera of the V to Z group. This variant differed further from 103A in that it bred true, whereas 103A behaved like the parent strain in consistently giving rise to a variant of the 103B type. Further study showed that a serum prepared against 103A agglutinated both 103A and 103B, but that a serum prepared against 103B had practically no action on 103A. Absorption of agglutinin experiments confirmed the suggestion that 103A contained two antigens—a type and a group—but that 103B contained only the group antigen. Since 103B grew uniformly in broth and was perfectly stable in saline, Boyd concluded that the variation he had observed was more of the phasic than of the smooth-rough order. It differed from the type-group variation seen in the flagellar antigens of organisms of the *Salmonella* genus in that it was irreversible. The time at which the A → B variation occurs varies greatly from strain to strain. Sometimes the parent culture may remain stable for months or years, at others it throws off B variants on first subculture or even in the body.

Similar observations were made with other strains of Flexner's bacillus, such as P119 and the 88 strain of the Newcastle-Manchester bacillus. Absorption of agglutinin experiments seemed to show that the V, W, and Z races each contained

a specific and a group antigen. The Y race contained group antigen only. The group antigen was not completely homogeneous; in fact, Boyd obtained evidence of at least six different components. The X race was at first thought to be similar to the others, but later Boyd (1940) came to regard it not as a separate race but as an incomplete variant of Z. It will be seen that in contradistinction to Andrewes and Inman, who regarded V, W, X, and Z as each containing the same four antigens, one of which alone was dominant in each race, Boyd believes that V, W, and Z share some components of a common group antigen but that their specific antigens are distinct (Fig 145). On

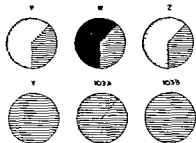


FIG 145.—DIAGRAM REPRESENTING THE VARIATION IN ANTIGENIC STRUCTURE OF *Shigella flexneri*.

The different components of the composite group antigen has been omitted for the sake of simplicity.  
(After Boyd.)

this view, Boyd (1940) would classify all strains containing the group antigen, including the Newcastle bacillus, as belonging to the Flexner variety, numbering the different races according to their specific antigen, and would include other strains devoid of the group antigen but biochemically similar to Flexner's bacillus in a new species for which the name of *boyd* has been suggested

Thus

	Old designation	New designation
<i>Sh flexneri</i>	V	I
" "	W	II
" "	Z	III
" "	103	IV
" "	P119	V
" "	Newcastle-Manchester-88	VI
<i>Sh boyd</i>	170	I
" "	P288	II
" "	D1	III

It seems to us very doubtful whether the Newcastle bacillus should be included in the Flexner group. Though it may share some of the component antigens of this group, it differs sufficiently in its biochemical characters to justify its being allocated provisionally to a species of its own.

Boyd's work, which is of fundamental importance, has been substantially confirmed by Wheeler (1944 *a, b*) in the United States. Wheeler has extended the six component factors of the Flexner group antigen to nine, he agrees that Type X has no specific antigen and therefore cannot be regarded as a type, and he finds that the specific antigen of the W race may be combined with two different sets of group antigen components.

Weil, Black and Farsetta (1944), neglecting the presence of minor antigens found no group antigen common to the Flexner types and therefore proposed that the Flexner and Boyd groups should be regarded as belonging to a single series and classified according to the nature of the primary antigen. Fourteen types were distinguished in this way, labelled I to XIV. Neither Wheeler nor Weil and his colleagues seem to have recognized the form of variation described by Boyd in his strains 103 and P119. Weil worked with a relatively few strains, many of them isolated a considerable time previously, and it would therefore be unwise to lay too much stress on his findings. All that we can say at present is that Boyd's classification, though admittedly imperfect, appears to provide us with the best working model for the serological examination of the Flexner dysentery group. Takita (1937) observed in Flexner bacilli a variation similar to that in 103, but proceeding in the opposite way. It is probable that the discrepancy is apparent not real, depending on the reverse way in which the variants were labelled.

Before leaving Flexner's bacillus it may be noted that several workers have studied its complex antigenic constitution, particularly Kruse and his colleagues (1907) and Lentz and Prügge (1931) in Germany, and Aoki (1921, 1923) in Japan. Sartorius and Reploh (1931, 1932), Clauberg (1932), Kemper (1933), and Nagakura (1937) have endeavoured to relate the different types established by various workers to each other, but with only partial success. Further observations will be required before we reach a satisfactory understanding of the problem that this organism presents.

It is now known that some strains of the Flexner Y variety contain somatic antigens that are found in the *Salmonella* group. The presence of the VLXIII combination has been demonstrated by Bornstein, Saphra and Daniels (1941) who point out that care must be taken in the identification of strains on the basis of agglutination.

The antigenic structure of *Sh. alcalescens* is still in doubt. According to Neter (1933) and Neter and Heide (1940) it contains two antigenic components, one specific and the other shared with the Flexner bacillus. De Assis (1939b) found that two serological types could be established distinguished by their specific antigens, but that both shared the same group antigen with *Sh. flexneri*. Archer (1942) noticed that *Sh. alcalescens* might form two types of colony: against a dark ground one appeared clear, the other opaque. Clear colonies were readily agglutinated, opaque colonies were hypoagglutinable but were rendered almost completely agglutinable by boiling. Boyd (1933) observed the occurrence of cross agglutination between *Sh. alcalescens* and an organism known as P274 which was biochemically similar to Flexner's bacillus but was not antigenically related to it. Rothstadt, Fenner and Baker (1949) in Australia have brought strong evidence to show that P274 is capable of giving rise to dysentery and have suggested that it should be classified with the Boyd variety. These results are conflicting. On the one hand there are observations pointing to the possession by *Sh. alcalescens* of the group Flexner antigen and on the other to its relationship to the P274 type. From a study of this organism Stuart, Rutiglian, Zimmerman and Corrigan (1943) conclude that there is a species-specific antigen A, a major antigen B found also in paracolon and coliform bacilli, a minor antigen C found in *Sh. flexneri* and in paracolon and coliform bacilli, and minor antigens D and E found again in paracolon and coliform bacilli. These workers observed a series of strains showing biochemical reactions ranging from those of the typical *Sh. alcalescens* on the one hand to those of *Bact. coli* on the other and their results render it doubtful how far *Sh. alcalescens* can as yet be defined as a species. An antigenic relationship between *Sh. alcalescens* and some paracolon bacilli was also noted by Sevitt (1945).

Sonne's bacillus presents less difficulty. As has been mentioned, this organism forms two types of colony which, as Large (1909) showed, are serologically distinct. A serum prepared against organisms in the smooth colony phase agglutinates both the smooth and the rough colony organisms, whereas a serum prepared against organisms in the rough colony phase agglutinates only the rough colony organisms. There is a close analogy with the 103 strain of Flexner's bacillus. For practical identification of the organism it is important that a serum should be used containing both types of agglutinins. *Sh. dispar* is antigenically heterogeneous though Carpenter (1944) found that 38 out of 44 strains he studied fell into one type.

It may be noted that the agglutination of dysentery bacilli occurs rather slowly, and that it is advisable to incubate all tests for 4-6 hours at 50° C. and to delay the final reading till the tubes have been left at room temperature overnight. This holds particularly for the *newcastle*, *alcalescens sonnei* and *dispar* types.

Apart from the serological differentiation of the dysentery bacilli, certain indirect tests may be used to distinguish between the types, such as Michaelis's (1917) acid agglutination test and susceptibility to action of the bacteriophage. The acid agglutination test depends on the different H<sup>+</sup> ion concentrations necessary for flocculation. Using the particular range employed by Michaelis, Andrewes and

Inman (1919) found that Schmitz's bacillus, *Sh. alkalescens*, and *Sh. dispar* were agglutinated, whereas *Sh. shigae* and *Sh. flexneri* were not. The test is only a rough one, and it is doubtful whether the information it furnishes justifies its use under ordinary conditions. On the other hand, the susceptibility of different types of dysentery bacilli to the action of certain phages appears to be very much more specific. According to Burnet and McKee (1930), whose article should be consulted for further details, bacteriophages that are active against Flexner bacilli can be divided into four main groups. One of these groups is capable of lysing bacilli only in the smooth phase, while the other three may or may not lyse smooth strains but generally lyse rough strains of all types. Characteristic differences in their sensitivity to a series of phages are presented by the V, W, X, Y, and Z types of *Sh. flexneri*.

**Chemical Fractionation.**—Early observations on the chemical structure of the antigens of the dysentery bacillus revealed the presence of specific polysaccharides in *Sh. shigae* (Kurauchi 1929, Meyer 1930, 1931, Morgan 1931) *Sh. flexneri*, and *Sh. sonnei* (Kurauchi 1929). The more recent work of Boivin and Mesrobian (1937a-b, 1938), Mesrobian and Boivin (1937), and Haas (1937, 1938a), carried out by use of the trichloroacetic acid technique, shows that the smooth, but not the rough somatic antigen of the Shiga and Flexner bacilli contains a polysaccharide hapten, which is linked to nitrogenous and lipid compounds to form a complete antigen. Similar polysaccharides have been demonstrated in *Sh. schmitzi* (Haas 1938c) *Sh. sonnei* (Haas 1938b), and *Sh. alkalescens* (see Weil 1943). The still more recent studies of Morgan (1930, 1937) and Morgan and Partridge (1940, 1941), who used the less drastic method of extraction of acetone treated bacterial cells with diethylene glycol seem to show that the complete smooth somatic antigen of Shiga's bacillus consists of a specific polysaccharide hapten, a non antigenic conjugated protein and a non antigenic phospholipin of the cephalin type. The polysaccharide is strongly dextrorotatory and yields 97 per cent of reducing sugars on acid hydrolysis. When combined with the conjugated protein, it forms a powerful antigen. It is interesting to note that the conjugated protein extracted from Shiga's bacillus appears to be identical with that found in the somatic antigen of *Salmonella typhi*. The polysaccharide not only forms a precipitate in the presence of a specific antiserum but neutralizes specifically the hemolytic action of *shigae* heterophile antibody (see Chapter 8) on sheep red corpuscles in the presence of complement (Meyer and Morgan 1935).

**Toxin Formation by Dysentery Bacilli.**—A large amount of work has been carried out on the formation of toxin by members of the dysentery group, particularly by *Sh. shigae*.

In 1903 Conradi prepared an autolysate of dysentery bacilli—probably Shiga's bacillus—which he found to be toxic for rabbits and guinea pigs. An 18 hours culture was suspended in saline and incubated for 24 to 48 hours at 37.5°C, after centrifugalization the yellowish supernatant fluid was removed, diluted with 5 times its volume of saline, and filtered through a Berkefeld candle, the filtrate was tested for sterility, and then concentrated to 1/10–1/50 of its bulk at 35°C. This product when injected intravenously into rabbits or intraperitoneally into guinea pigs in a dose of 0.1 ml., proved fatal in about 48 hours. In rabbits death was preceded by diarrhoea, collapse, and paralysis of the legs, in guinea pigs by a rapid fall of temperature and collapse. At necropsy Conradi found in both animals congestion of the intestine, mucus and blood adhering to the mucosa, and frequently small hemorrhages of the mucous and the serous coats. When a smaller dose was injected into rabbits the animals lived for 4 to 6 days, and he found that post mortem the mucosa of the last third of the large intestine was swollen, blackish red in colour, and ulcerated in several places.

Neisser and Shiga (1903) confirmed Conradi's results, and noted in addition that the

toxic substances were precipitated by alcohol and ether, and largely destroyed by heat at 75° C. Todd (1904) in this country was able to show that 4 to 6 weeks' cultures of Shiga's bacillus contained a soluble toxin, which was highly active on rabbits and horses, but to a much less extent on guinea-pigs, rats, and mice. Flexner's bacillus proved incapable of giving rise to a soluble toxin. Dopter (1905), studying the histological appearances of rabbits dying of paralysis subsequent to injection with 24-hours' broth cultures of *Sh. shigae*, observed definite lesions in the spinal cord, consisting chiefly of chromatolysis of the anterior horn cells, sometimes with small interstitial haemorrhages and focal necroses of the grey matter. The lesions occurred as frequently after the injection of toxin as of bacilli. Further work by Kraus and Dörr (1905) led to the conclusion that Shiga's bacillus gave rise to two toxins: (1) a soluble toxin, present in 8-10-days' broth cultures and in filtered saline suspensions of 24-hours' agar cultures; this was fatal to rabbits but not to guinea-pigs, and gave rise to the production of a specific neutralizing antitoxin; (2) an insoluble toxin present in the bacterial bodies, which was fatal both to rabbits and to guinea-pigs. No soluble toxins were found in cultures of Flexner's bacillus.

Flexner and Sweet (1906), using a modification of Conrad's method, obtained a toxin from 24-hours' agar cultures of Shiga's bacillus, which, injected intravenously into rabbits, gave rise to diarrhoea, paralysis, convulsions, and death. The paralysis began in the upper limbs and extended at times to the lower limbs. Sometimes the animals survived for 10 days after the extremities were paralysed; they lay on one side in a position of opisthotonos. Post mortem, small haemorrhages were seen in the brain, and softening of the grey matter in the spinal cord. In the intestine there was congestion of the serosa; the walls of the gut, especially of the caecum and appendix, were thickened and oedematous, the mucosa was yellowish-white and thrown into deep folds, which were sometimes covered by a pseudo-membrane or stippled with haemorrhages. The mesenteric glands were swollen, oedematous, and congested. Heat at 81° C. for 1 hour destroyed the toxin. These observations were largely confirmed by Bessau (1911), who concluded that there were two different toxins—one a paretic or neuro-toxin causing paralysis of the muscles, the other a marasmic or intestinal toxin causing a fall in temperature, diarrhoea, and chronic marasmus. The paretic toxin was neutralized by antitoxin, the marasmic toxin was not. Further, rabbits were affected by both toxins, whereas in guinea-pigs the paretic toxin was without effect.

Bessau's conclusions were supported by the observations of Oltzky and Kligler (1920), who concluded that *Sh. shigae* formed two toxins: one a neurotoxin acting on the central nervous system of the rabbit and identified by Oltzky and Kligler as an exotoxin; the other having a specific affinity for the intestine and regarded by Oltzky and Kligler as an endotoxin.

This conception was challenged by Okell and Blake (1930), who found that toxin was not liberated from the cell bodies in the absence of autolysis, and therefore concluded that there was only one toxin produced, namely an endotoxin. The more recent chemical work, however, of Boivin and Mesrobian (1937a-A, 1938) leaves little doubt that the dual conception is correct. By means of the trichloroacetic acid method these workers were able to demonstrate the presence in cultures of Shiga's bacillus of (1) a thermostable glycolipoid somatic antigen, which they regarded as an endotoxin having an enterotoxic effect, and (2) a thermolabile protein substance, which they regarded as an exotoxin having a neurotropic effect. The exotoxin is specific to Shiga's bacillus, and may be formed by either antigenically smooth or rough strains of this organism (Haas 1937, Istrati 1938, Steabben 1943, Olitzki *et al.* 1943); the endotoxin is similar to the smooth somatic antigens found in other members of the dysentery group (Haas 1938a, b, c) and in the *Salmonella* group (see p. 724), and is absent from antigenically rough strains.

It is perhaps unfortunate that the terms endotoxin and exotoxin have been used to refer to the two types of toxic substances produced by Shiga's bacillus. The so-called exotoxin is closely bound up with the cell bodies and, as the observations of Okell and Blake (1930) showed, (see also Olitzki, Bend-reky and Koch 1943), is not excreted



by the living bacilli but appears free in the culture medium only after autolysis of the dead cells has begun. Again the implied distinction between enterotoxin and neurotoxin is misleading since it is apparent that the so called exotoxin besides causing paralysis of the limbs leads also to serious changes in the intestine perhaps by causing vasoconstriction of the intestinal blood vessels (Penner and Bernheim 1949). For the present it is better to refer to the so called exotoxin by Steadman's term *neuro-enterotoxin*.

Summarizing our present knowledge we may say that *Sh. shigæ* gives rise to a specific soluble neuro enterotoxin which is present in broth cultures about a week old in filtered autolysates of 24 hour agar cultures and in the dried bacterial bodies. It is destroyed by a temperature of 75°-80° C. maintained for an hour. If prepared by a method favouring bacillary autolysis (Hansen 1936 Takita 1939) it proves fatal to mice inoculated intravenously in a dose of about 0.001 to 0.01 ml. It is fatal to the rabbit causing paralysis of the limbs diarrhoea and collapse but has less action on the guinea pig in which diarrhoea and collapse alone are produced. It is a protein and may be precipitated by trichloroacetic acid (Boivin and Mesrobian 1937a) and to a less extent by the addition of 40 per cent solid ammonium sulphate (Blake and Okell 1929). It gives rise to and is neutralized by a specific antitoxin which combines with it in constant proportions. In addition, smooth strains of *Shigæ* bacillus contain a toxic somatic antigen which is a lipopolysaccharide conjugated with a protein resembling similar smooth antigens in the *Salmonella* and in other members of the *Shigella* group.

With the partial exception of Schmitz's bacillus the high toxicity of *Sh. shigæ* is not rivalled by other dysentery bacilli. The ground up bacterial bodies of *Sh. flexneri*, *Sh. sonnei* and *Sh. dysenteriae* prove fatal on intravenous inoculation of rabbits but only in a dose that is about 20 times greater than the corresponding fatal dose of dried *Shigæ* cells. Schmitz's bacillus is variable in its toxicity. According to Buchwald (1939) it may give rise to a thermolabile neurotoxin precipitable by trichloroacetic acid from old broth cultures and causing paralysis of the extremities when inoculated into mice and rabbits but it appears to be rather less potent than that formed by *Shigæ* bacillus. *Sh. alkalescens* is said to be non toxic.

**Pathogenicity**—*Sh. shigæ*, *Sh. schmitzi*, *Sh. flexneri*, the Newcastle bacillus, *Sh. sonnei* and a number of types described by Boyd (1938, 1940) undoubtedly give rise to dysentery in man. The role of *Sh. alkalescens* is still doubtful though it may be responsible for infections of the urinary tract. *Sh. dysenteriae* appears to be non pathogenic. The evidence in favour of the pathogenicity of some members of the para *Shigæ* group is strong but further observations are desirable. (For fuller information see Chapter 70.)

With the exception of captive monkeys which may carry *Sh. flexneri* in the gut (see Lovell 1929) and which may sometimes as on a vitamin deficient diet (Verder and Petran 1937 Janota and Dach 1939) develop dysenteric symptoms (Preston and Clark 1938 David and Schurl 1939) and with the possible exception of dogs which may be infected with either *Shigæ* or *Flexner's* bacillus (Dold 1916) animals do not appear to suffer from dysentery. It is not possible to reproduce the typical disease as it occurs in man by experimental inoculation or feeding of the ordinary laboratory animals. Nevertheless many dysentery organisms especially *Sh. shigæ* are toxic to rabbits horses and mice and to a less extent to guinea pigs. After subcutaneous inoculation into rabbits dogs and young pigs

the living bacilli may become localized in the intestine and give rise to catarrhal and necrotic lesions which often prove fatal (Vaillard and Dopfer 1903)

**RABBITS.**—A small dose—0.01 mm of a 24-hours agar culture of *Sh. dysenteriae* injected intravenously proves fatal in 1 to 4 days. Death is preceded by diarrhoea, paresis or total paralysis of the extremities, and collapse. Post mortem, there may be hemorrhages into the subcutaneous tissue and peritoneum, the intestine, especially the caecum and colon, is congested and may show submucous hemorrhages. The mucosa itself is congested, cedematous, and sometimes studded with petechiae (Vaillard and Dopfer 1903, Amako 1904). In the lumen of the gut there is often mucus or bloody fluid. A similar picture is seen after injection of dead bacilli in larger quantity or of toxin. If a smaller dose of bacilli is given, there may be time for necrosis and actual ulceration of the intestine to occur. The living bacilli can be recovered from the mucosa and from the corresponding mesenteric glands. Subcutaneous injection has much the same effect as intravenous, but the animals survive longer. The lesions following injection of Flexner's and Sonne's bacillus are not unlike those produced by *Sh. dysenteriae*, if a sufficient dose is given, but they are rarely so severe.

**MICE.**—0.1 mm of a 24-hours culture of *Sh. dysenteriae* injected intraperitoneally or subcutaneously kills the animal in 1 to 4 days. At necropsy there may be no evident change or there may be catarrhal inflammation of the intestine with watery mucus in the gut. *Sh. flexneri* and *Sh. sonnei* often prove fatal on intraperitoneal inoculation of large doses.

**GUINEA PIGS.**—These animals are less susceptible with respect to *Sh. dysenteriae* than are rabbits and mice. The lesions produced by subcutaneous or intraperitoneal injection of living bacilli vary. After a fatal dose there may be no marked macroscopic changes, or there may be intestinal lesions similar to those found in rabbits. Death may be produced by large intraperitoneal doses of Flexner's or Sonne's bacillus.

**OTHER ANIMALS.**—By giving a cat  $\frac{1}{2}$  drop of croton oil, and injecting a whole agar slope of Shiga's bacillus directly into the stomach, Shiga (1895) succeeded in setting up diarrhoea for a week—the animal passed crv, slimy stools, from which the bacilli could always be cultivated. It died 4 weeks later, at necropsy there was congestion of the rectal mucosa, and a covering of mucus over the whole of the large gut. The bacillus was recovered from the caecum and large intestine. Most workers have failed completely to reproduce true dysenteric lesions in cats, dogs, rabbits, or monkeys either by injection *per os* or *per rectum* though the feeding of monkeys with large doses of Flexner's bacillus may give rise to severe dysenteric symptoms (see Dack and Petran 1934).

**Classification.**—Space does not permit of a description of the numerous attempts that have been made to afford a satisfactory classification of the dysenteric bacilli. These may be summarized by saying that Lentz (1902) in Germany first perceived the difference between the mannitol and the non-mannitol fermenters, that Hiss (1904) in the United States, later supported by Boylston (1934) in Denmark, suggested a classification on fermentation reactions—that Kruse and his colleagues (1907) in Germany realized the value of serological methods of classification, and had the merit of pointing out the antigenic complexity of the mannitol fermenting group—that Shiga (1906) and Amako (1906) in Japan combined fermentation and serological methods, that Gettings (1919) Murray (1916) and Andrewes and Inman (1919) in England arrived independently at results agreeing closely with each other and showed that the non-mannitol fermenting Shiga bacillus was antigenically distinct and homogeneous, whereas the mannitol fermenting bacilli, of which Flexner's bacillus was the main example, were heterogeneous and divisible into four types according to the preponderance of one or other of the antigenic components V, W, X, and Z, and that Boyd (1931, 1932, 1935, 1936, 1940) reached a different conclusion on the antigenic structure of the mannitol

fermenting group finding that some strains shared a group antigen and that others did not.

All workers are now agreed that the main lines of cleavage are (1) between the mannitol and the non mannitol fermenters and (2) in the mannitol fermenting group between the non lactose and the late-lactose fermenters. Opinion varies mainly in respect of the differentiation of antigenic types from species and the nomenclature to be employed.

In the non mannitol fermenting group *Sh. schmitzi* is distinguished from *Sh. shigae* by fermenting rhamnose producing indole and being antigenically distinct. The Newcastle bacillus may or may not ferment mannitol: the non mannitol fermenting strains can be distinguished readily from *Sh. shigae* or *Sh. schmitzi* by their production on first isolation of a bubble or two of gas in tubes of glucose and dulcitol and by their different serological behaviour. Besides these species there is a group of organisms at present ill-defined which resemble *Sh. shigae* or *Sh. schmitzi* biochemically but differ from both in their antigenic structure. These organisms have been described by Dudgeon and Urquhart (1919) as para-Shiga bacilli. Since they vary in their fermentation of arabinose their production of indole and their serological reactions it is impossible to say at present whether they should be classified as a single species *Sh. parashigae* and subdivided into types or whether more than one species will be required. We prefer to leave the matter open and refer to them simply as para-Shiga bacilli.

In the non lactose fermenting subdivision of the mannitol fermenting group *Sh. alkalescens* differs from most strains of *Sh. flexneri* in fermenting dulcitol, xylose, sorbitol and according to de Assis (1939a) most important of all glycerol. It is moreover susceptible to acid agglutination. The *flexneri* types are peculiarly difficult to classify satisfactorily at present. The old division on antigenic structure into V, W, X, and Z types has been challenged by Boyd (1938, 1940) who does not recognize the X type and who proposes to replace the letters V, W and Z by the Roman numerals i, ii, iii and to include under the term *Bact. dysenteriae* Flexner three other types iv, v and vi corresponding to his strains 103, P119 and 88. All of these types according to Boyd share a common group antigen. Other strains biochemically similar to Flexner's bacillus but not containing the Flexner group antigen he would put into a separate species labelled by his own name, and divide them on an antigenic basis into *Bact. dysenteriae* Boyd Types i, ii and iii corresponding to his strains 170, P288 and D1.

This raises serious difficulties both of classification and of nomenclature. Leaving nomenclature until later we think that there is a good precedent for classifying antigenic variants of the same species into types distinguished by numerals, for this reason we would welcome the replacement of the letters V, W, and Z by numerals. We think also that there is justification for classifying biochemically similar but antigenically distinct organisms as separate species and we see no objection to the use of the term *boydii* as a specific name. When however we consider the constitution of the *flexneri* species we are faced with two difficulties: namely should the X type be omitted and should strain 88, which is a mannitol fermenting variant of the Newcastle bacillus be included? In his 1938 paper Boyd concluded provisionally that X contained a distinctive type antigen, and it was not till 1940 that he decided that this type was merely a variant of Z. Previous workers have had little difficulty in recognizing the X type and the evidence on which it has been degraded might be considered as unconvincing.

still, in our opinion, good reason for continuing this mode of classification, but there is now perhaps reason for proceeding still further in the taxonomic differentiation of these organisms. As we have already explained in the previous chapter, the term *Salmonella* has got for practical purposes to be accepted generically, in spite of the logical inconsistencies which its adoption leads to. If we accept this, then we are forced to separate off the dysentery bacilli from the *Bacterium* genus, and classify them under a separate generic name, such as *Shigella*. The specific names can then follow easily. American workers have adopted the specific name *ambigua* in place of *schmitzi* and of *paradysenteriae* in place of *flexneri*.

Both Schmitz's and Flexner's bacillus are so well known by the names of their discoverers that it is a pity not to perpetuate this association in the nomenclature of the dysentery group. The strict systematist may object that the terms *ambigua* and *paradysenteriae* have priority, but this argument has little power to move us. The primary purpose of nomenclature is utility, and to insist on an inapt and uninformative specific name merely on grounds of botanical convention is to forget that the Sabbath was made for man and not man for the Sabbath. The classification and nomenclature that we would suggest as being the most valuable for teaching purposes is given diagrammatically in Fig 146.

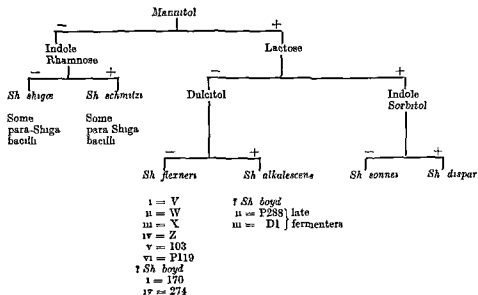


FIG 146.—TENTATIVE CLASSIFICATION OF THE DYSENTERY BACILLI  
 Note—*Shigella newcastle* may or may not ferment mannitol and has been omitted from the figure

For two recent reviews of the dysentery bacilli reference may be made to Neter (1942) and Weil (1943)

#### REFERENCES

- ALI, M. (1938) *J. Egypt med. Ass.* 21, 650  
 AMANO, T. (1903) *Z. Hyg. Infektkr.* 60, 93  
 ANDREWS, F. W. (1918) *Lancet* i 560  
 ANDREWS, F. W. and INMAN, A. C. (1919) *Spec. Rep. Ser. med. Res. Coun., Lond.* No 42  
 AOKEI, K. (1921) *Tokoku J. exp. Med.* 2, 142, (1923) *Ibid.* 4, 12

- ARCHER, G T L. (1942) *J. R. Army med. Cps.*, 79, 109.
- ARKWRIGHT, J A. (1921) *J. Path. Bact.*, 24, 36.
- ASSIS, A DE (1939a) *O Hospital*, 15, 447. (1939b) *Ibid.*, 15, 655.
- BAMFORTH, J (1934) *J. Hyg., Camb.*, 34, 69.
- BECK, A. and BUCKLE, D F. (1939) *J. Hyg., Camb.*, 39, 311.
- BESSAU, G. (1911) *Zbl. Bakt.*, 57, 27.
- BLAKE, A. V. and OKELL, C. C. (1929) *J. Path. Bact.*, 32, 121.
- BOVIN, A. and MESSEGEANT, L. (1937a) *C. R. Acad. Sci.*, 204, 302. (1937b) *Ibid.*, 204, 1759. (1937c) *C. R. Soc. Biol.*, 124, 442. (1937d) *Ibid.*, 124, 1078. (1937e) *Ibid.*, 125, 706. (1937f) *Ibid.*, 126, 222. (1937g) *Ibid.*, 126, 323. (1937h) *Ibid.*, 126, 602. (1938) *Ibid.*, 128, 446.
- BOULÉ, H. (1930) *C. P. Soc. Biol.*, 103, 613. (1934) "Dysentery in Denmark." Bianco Lunos Bogtrykkeri A/S, Copenhagen.
- BORNSTEIN, S., BAFERA, I., and DANIELS, J B. (1941) *J. Immunol.* 42, 401.
- BOYD, J S K. (1931) *J. R. Army med. Cps.*, 57, 161. (1932) *Ibid.*, 59, 241, 331. (1933) *Ibid.*, 64, 289. (1936) *Ibid.*, 66, 1. (1939) *J. Hyg., Camb.*, 38, 477. (1940) *Trans. P. Soc. trop. Med. Hyg.*, 33, 503.
- BRAY, H. and BAKER, F. (1930) *Zbl. Bakt.*, 115, 462.
- BRAY, H. and WEIL, A. J. (1928) *Zbl. Bakt.*, 109, 16.
- BUCHANAN, G. and PORT, P. (1930) *J. med. 4<sup>th</sup> S. Africa* 4, 60.
- BUCHWALD, H. (1939) *Z. Immunforsch.*, 96, 440.
- BURNET, F. and MCKIE, M. (1930) *J. Path. Bact.*, 33, 637.
- CANN, L. W. and NAVASQUEZ, S DE. (1931) *J. Hyg., Camb.*, 31, 301.
- CARPENTER, P L. (1944) *J. Bact.*, 47, 419.
- CARTER, J R. (1921) *Lancet*, 1, 657.
- CASTELLANI, A. (1907) *J. Hyg., Camb.*, 7, 1. (1912a) *J. trop. Med. Hyg.*, 15, 162. (1912b) *Zbl. Bakt.*, 65, 262.
- CHRISTENSEN, W B. and GOWEN, G H. (1944) *J. Bact.*, 47, 171.
- CLAUBERG, K. W. (1932) *Zbl. Bakt.*, 124, 23.
- CLAYTON, F H. A. and WARREN, S H. (1929a) *J. Hyg., Camb.*, 28, 355. (1929b) *Ibid.*, 29, 191.
- COVADI, H. (1903) *Deutsch. med. Wochr.*, 29, 26.
- CRUICKSHANK, R. and SWYER, R. (1940) *Lancet*, ii, 803.
- DACK, G M. and PETRAN, E. (1934) *J. infect. Dis.*, 55, 1.
- DAVID, H. and SCHIEL, A. (1939) *Zbl. Bakt.*, 144, 43.
- DOLD, H. (1916) *Deutsch. med. Wochr.*, 42, 811.
- DORTCH, C. (1905) *Ann. Inst. Pasteur*, 19, 303.
- DOWNIE, A. W., WADE, E., and YOUNG, J A. (1933) *J. Hyg., Camb.*, 23, 196.
- DUDGEON, L S. and URGHART, A. L. (1919) *Spec. Rep. Ser. med. Res. Coun.*, No. 40 p. 25.
- FLEISHER, S. (1900a) *Zbl. Bakt.*, 28, 625. (1900b) *Bull. Johns Hopk. Hosp.*, 11, 231.
- FLEISHER, S. and SWEET, J E. (1906) *J. exp. Med.*, 8, 514.
- FORSYTH, W L. (1933) *J. trop. Med. Hyg.*, 36, 60.
- FITZ, G M. (1927) *J. Hyg., Camb.*, 26, 271.
- GETTINGS, H S. (1919) *Spec. Rep. Ser. med. Res. Coun. Lond.*, No. 30.
- GLYNN, J H. and STARKER, D H. (1939) *J. Bact.*, 37, 315.
- HAAS, R. (1937) *Z. Immunforsch.*, 91, 204. (1938a) *Ibid.*, 92, 305. (1938b) *Ibid.*, 94, 239. (1938c) *Ibid.*, 94, 480.
- HANSEN, A. (1936) *Biochem. Z.*, 287, 30.
- HARDY, A. V., WAIT, J., KOLODNY, M H., and DECAPITO, T. (1940) *Amer. J. publ. Hlth.*, 30, 53.
- HAYES, E. L. (1938) *J. infect. Dis.*, 63, 330.
- HISS, P H. (1904) *J. med. Res.*, 13, 1.
- HISS, P H. and FUSSELL, P F. (1903) *Med. News* 82, 209.
- ISABOLINSKY, M. and GITOWITZ, W. (1906) *Zbl. Bakt.*, 97, 140.
- ISTAT, G. (1938) *C. R. Soc. Biol.*, 128, 1010.
- JANOTA, M. and DACK, G M. (1939) *J. infect. Dis.*, 65, 219.
- JOHNSTON, M. M. and BROWN, A. (1930) *Canad. publ. Hlth J.*, 21, 394.
- JOHNSTON, M. M. and KAAKE, M J. (1932) *Canad. publ. Hlth J.*, 23, 109.
- KEMPER, F. (1933) *Zbl. Bakt.*, 130, 260.
- KERRIN, J C. (1908) *J. Hyg., Camb.*, 28, 4.
- KOBAYASHI, R., OKUBO, H., OHYO, J., IRI, M., NAKAMURA, B., MACHIDA, S., KOBAYASHI, E., MATSUMOTO, I., and MATSUTANA, S. (1931) *Kiassato Arch.* 8, 99.
- KOSER, S A., REITER, D O., BORTVICKER, E., and SWINGLE, E. L. (1930) *J. proc. Med.*, 4, 477.
- KOSER, S A. and STYRON, N C. (1930) *J. infect. Dis.*, 47, 443.
- KRAUS, R. and DÖRR, R. (1905) *Wien. H. u. Wochr.*, 18, 1077.
- KRUSE, W. (1900) *Deutsch. med. Wochr.*, 26, 637. (1901) *Ibid.*, 27, 370, 306.

- KRUSE, RITTERSHAUS, KEMP, and METZ. (1907) *Z Hyg InfektKr.*, 57, 417
- KURACHI K. (1929) See Ando, K. (1929) *J Immunol*, 17, 555
- LARGE, D T M (1929) *J R Army med Cps* 52, 1
- LARGE, D T M and SANKARAN, O K (1934) *J R Army med Cps*, 63, 231
- LEVY, (1902) *Z Hyg InfektKr.*, 41, 559
- LEVY, O and PRUDON, R (1931) Kalle, Kraus and Uhlenhuth's "Handbuch der pathogenen Mikroorganismen" Gustav Fischer, Jena, 3te Aufl., 3, 1377
- LESTER, V (1926) *Acta path microbiol scand.*, 3, 696
- LEUCHS, J and FLOCHMANN, E (1927) *Zbl Bakt.*, 104, 347
- LOVELL, R (1929) *Proc roy Soc Med.*, 22, 820
- MCGIVRES O F., MCCLEAY, A L., SPINDLER, F., and MAXCY, H F (1936) *Amer J Hyg* 24, 552
- MANDRY, O C (1935) *Puerto Rico J publ Hlth* 10, 308
- MESROBIAN, L. and BOIVIN, A (1937) *C R Soc Biol*, 124, 439
- MEYER, K (1930) *Z ImmunForsch.*, 68, 98, (1931) *Ibid.*, 69, 134 499
- MEYER, K and MORGAN W T J (1935) *Brit J exp Path.*, 16, 476
- MICHAELIS, L. (1917) *Dtsch med Wschr.*, 43, 1506
- MITA, K. (1921) *J infect Dis.*, 29, 580
- MORGAN, W T J (1931) *Brit J exp Path.*, 12, 62, (1936) *Biochem J*, 30, 969, (1937) *Ibid.*, 31, 2003
- MORGAN, W T J and PARTIDGE S M (1940) *Biochem J*, 34, 169, (1941) *Ibid.*, 35, 1140
- MURRAY, E G D (1918) *J R Army med Cps* 31, 257, 353
- NABARRO D (1923) *J Path Bact* 26, 429, (1927) *Ibid* 30, 176
- NAGAKURA K. (1937) *Z ImmunForsch.*, 90, 139
- NEISSER, M and SHIGA, K. (1903) *Dtsch med Wschr.*, 29, 61
- NELSON, R L. (1930) *J Bact.*, 20, 183
- NETER, E (1938) *J Immunol* 35, 339, (1942) *Bact Rec* 6, 1
- NETER, E and HEIDE, A M (1910) *Amer J Hyg.*, 31, B, 69
- OKELL, C C and BLAKE A V (1930) *J Path Bact.*, 33, 7
- OLITSKY, P K and KLIGLER, I J (1920) *J exp Med* 31, 19
- OLITSKY, L., BENDERSKY, J., and KOCH P K (1917) *J Immunol* 48, 71
- ORSKOV, J and LAESSEN A (1923) *J Bact* 10, 473
- PENNER, A and BERNHEIM, A I (1942) *J exp Med* 76, 271
- PRESTON, W S and CLARK, P F (1938) *J infect Dis.*, 63, 238
- Report (1939) Antigenic variation in the organisms of bacillary dysentery etc  
Standards Laboratory, Oxford
- ROELCKE K. (1938) *Z Hyg InfektKr.*, 120, 307
- ROTHSTADT, L. F., FENNER F., and BAKER B A (1912) *Pers comm*
- SACHS A (1943) *J P Army med Cps* 80, 91 [Wrongly described as Sachs II]
- SANTORINI F and REFLOH H (1931) *Klin Wschr.*, 10, 2216 (1932) *Zbl Bakt* 126, 10
- SCHMITZ, H F F (1917) *Z Hyg InfektKr.*, 84, 449
- SCHÜTZE H (1944) *J Path Bact.*, 56, 200
- SEVITT, S (1945) *J Hyg, Camb* 44, 37
- SHIGA, K. (1898a) *Zbl Bakt.*, 23, 699, (1898b) *Ibid.*, 24, 617 670 913 (1901) *Dtsch med Wschr.*, 27, 741, 765 783, (1908) *Z Hyg InfektKr.*, 60, 75.
- SMITH, J and FRASER, A M (1929) *J Path. Bact.*, 31, 511
- SONNE, C. (1915) *Zbl Bakt* 75, 408
- STABBEK, D (1943) *J Hyg Camb.*, 43, 83
- STRONG, R P and McORAVE W E (1900) *J Amer med Ass.*, 35, 498
- STUART, C. A., RUSTIGIAN, R., ZIMMERMAN, A and CORRIGAN F V (1943) *J Immunol* 47, 425
- TAKITA, J (1937) *J Hyg, Camb.*, 37, 271, (1939) *Kitasato Arch.*, 18, 174
- THOJOTA T and WAALER, F (1932) *J Bact.*, 24, 301
- TODD, C. (1904) *J Hyg, Camb.*, 4, 480
- VAUILLARD, L. and DORTCH C (1903) *Ann Inst Pasteur.*, 17, 463.
- VERDER, F. and PETRAN, E. (1937) *J infect Dis.*, 60, 193.
- WAALER, E. (1935) "Studies on the Disassociation of the Dysentery Bacilli. I Kommission  
Hov Jacob Dybwad, Oslo
- WEIL, A J (1943) *J Immunol*, 46, 13
- WEIL, A J., BLACK J and FARSETTA K (1944) *J Immunol.*, 49, 321 341
- WELCH H and MICKLE F L (1932) *Amer J publ Hlth*, 22, 263, (1934) *Ibid.*, 24, 219
- WHEELER, K M (1914a) *J Immunol*, 43, 87 (1914b) *Amer J publ Hlth.*, 34, 621
- WHITHEAD, H and SCOTT W M (1931) *Lancet*, ii 248
- WILSON, G C., TRIGG, P S., WRIGHT R C., HENDRY, C. B., COWELL, M P and MAIER, J (1935) *Spec. Rep Ser med Res Coun. Lond.*, No. 206.
- WOOD A. J. and KEEFING F E. (1944) *J Bact.*, 47, 309
- WYCKOFF, R. W G (1933) *J exp Med.*, 57, 163

## CHAPTER 30

### SALMONELLA

#### DEFINITION—*Salmonella*

Gram negative non sporing rods usually 1-3  $\mu$  long and 0.5-0.7  $\mu$  broad. Primarily intestinal parasites widely distributed in man mammals and birds. With few exceptions all species are motile by peritrichate flagella. Easily cultivable on ordinary media. Aerobic and facultatively anaerobic. Apart from a few species that form acid only acid and gas are produced from glucose mannitol, dulcitol, and sorbitol. Lactose, sucrose, adonitol, and except rarely salicin are not fermented. Indole and acetylmethylcarbinol are not formed. Gelatin is seldom liquefied.  $H_2S$  production is usual. The species are closely related to each other by somatic and flagellar antigens. Most species are diphasic. Pathogenic for man animals birds or all three giving rise to food poisoning enteritis or typhoid like infections.

**Nomenclature**—In Chapter 28 we have already mentioned our decision to split off the *Salmonella* and *Shigella* sub-groups from the wide group of Gram negative non sporing rods previously classified under the genus *Bacterium*. The justification for this decision is one of expediency. That the close relation of their antigenic components and the type of disease to which they give rise serve to differentiate the salmonella and the dysentery bacilli from the ordinary coliform bacilli will not be questioned. But whether it is justifiable to give these sub-groups generic rank and to distinguish each of the antigenic types by a specific name when in the *Streptococcus* group the main sub-groups are not given generic rank and the antigenic types are treated as varieties and labelled by numbers is very questionable. That the term species is being used with two different connotations is clear enough, and we make no attempt to defend such inconsistency. On the other hand we may plead that bacterial taxonomy is still in a process of evolution. That there is as yet no general agreement on the definition of the terms genus and species and that until a final ruling is laid down by some properly constituted international committee we as writers of a text book of bacteriology must be free to select such names from among those that have been proposed as will best serve to aid the student in the recognition of the various groups of bacteria that he has to study.

So far as the term *Salmonella* is concerned a special sub-committee of the International Society of Microbiology (Report 1934) recommended the adoption of the terminology introduced by Kauffmann which recognized the generic status of the *Salmonella* group and the specific rank of each of the antigenically distinguishable types. Since then the number of recognized types has more than doubled. There are now over 130 specifically named serological types around which an extensive literature has grown up. It is therefore almost inconceivable that any international committee on nomenclature appointed in future would

suggest such changes in definition as would necessitate the degradation of the generic term *Salmonella* to specific rank, and the numbering as varieties of all the present named species. Convenience and expediency must be our justification for adopting the recommendations of the *Salmonella* sub-committee in face of the obvious illogicality of using a different system of nomenclature for members of the *Streptococcus* group.

If, however, we decide to employ *Salmonella* as a generic term, we must take care to define it as far as possible on the same general principles as have guided us in defining other genera. The lines of demarcation in biology can rarely be sharp. It is dangerous to rely, therefore, on any single character in the identification of a bacterium. Here we find ourselves in conflict with Kauffmann (1911), who lays down the following definition: "*Salmonella* bacteria are Gram negative bacteria which, on the ground of their antigenic structure, can be included in the Kauffmann-White scheme." If we rely solely on antigenic constitution, as Kauffmann does, we shall be logically compelled to include in the *Salmonella* genus every organism, no matter how different it may be in other characters, that contains a single antigen hitherto recognized in the scheme of classification drawn up by Kauffmann and White. Already numerous strains of coliform and paracolonic bacilli have been found to contain one or more of the H or O antigens of *Salmonella* (Habs and Arjona 1935, Gard 1937, Gard and Eriksson 1939, Schiff, Bornstein, and Saphra 1941, Saphra and Silberberg 1942, Peluffo, Edwards, and Bruner 1942, Leon 1942, Wheeler *et al* 1943, Edwards, Cherry and Bruner 1943); and some of the *Salmonella* O antigens have been recognized in strains of Flexner dysentery bacilli (Bornstein, Saphra, and Daniels 1941), and even in members of the *Pasteurella* group (Schütze 1928, Pirotsky 1938). To transfer these organisms, which differ in fermentation and pathogenic characters, to the *Salmonella* genus, merely because their constituent antigens happen to share some of the necessarily limited groupings with those common among the salmonellae, is not only to lose all sense of proportion, but to sanction a principle in bacterial nomenclature that cannot but lead to progressively increasing confusion.



and carnivores are not uncommonly infected. Except for one or two members, like the typhoid bacillus, that are non pathogenic to animals, man does not seem to act as an important reservoir of infection, the invading organisms are quickly thrown off, and the chronic carrier state is unusual.

**Morphology.**—The shape, size, structure, and arrangement of the bacterial cells do not differ materially from those in the *Bacterium* group. The usual length is 2–3  $\mu$  and the usual width 0.6  $\mu$ , but considerable deviation from these modal values is found under different environmental conditions and in different cultural variants. With the exception of *Salm gallinarum* and its variant *pullorum*, all species are motile, though individual non motile strains of the typhoid bacillus, for example, may be encountered occasionally in the body, and non motile variants may be thrown off under cultural conditions in the laboratory. Motility, however, is such a general characteristic that failure of a strain to exhibit it on primary isolation from the body must be regarded as almost sufficient in itself to exclude it from the *Salmonella* group. Whether the flagella are peritrichate, which has

been the conclusion drawn by most workers from a study of fixed and stained bacilli, or whether, as appears possible from Pijper's (1933, 1940) studies of living typhoid bacilli by sunlight darkground photomicrography, they are disposed one on each side of the bacillus, is disputable, and is for our purpose more of academic than of practical importance.

Capsules are not ordinarily formed, but many species, notably *Salm paratyphi B*, may give rise to mucoid colonies in which the individual organisms are surrounded by a polysaccharide-containing capsular material (Fletcher 1918). The formation of a capsule by the typhoid bacillus has been described by Kühnemann (1911), Carpano (1913), Marrassini

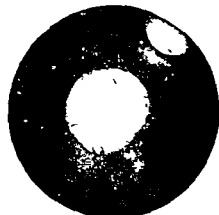


FIG 147.—*Salm. enteritidis*

Colony on agar plate after 24 hours ( $\times 8$ )

(1913), Shimidzu (1913), and Gay and Clappole (1913).

**Cultural Characters.**—The members of this group grow readily on ordinary nutrient media and cannot be distinguished from coliform bacilli. A few species, however, such as *paratyphi A*, *abortus-oris*, *typhi suis*, *sendai*, and *pullorum*, grow less abundantly and form but a thin layer of growth on an agar slope. On brilliant green agar plates the difference is particularly noticeable, the growth being both slower and less abundant than that of other members of the group. *Salm. typhi* and *Salm. rostock* likewise grow poorly on brilliant green agar, though developing fairly well on ordinary nutrient agar. In broth, smooth strains give rise to a uniform turbidity, increasing rapidly during the first 12 to 18 hours of growth, and then more slowly up to 48 to 72 hours. Pellicle formation is rare, and when present is slight. A deposit forms as growth increases, this disperses readily on shaking leading to an increase in the turbidity of the culture.

On agar, the colonies are relatively large, with an average diameter of 2–3 mm, but they vary considerably in size. They may be circular and low convex with a smooth surface and entire edge, they may be flatter with a less regular surface

and a more effuse serrated edge, or they may assume the vine leaf form which used to be regarded as characteristic of *Salm typhi*. Dwarf colony forms are sometimes met with. They were first described by Jacobsen in 1910, and have since been reported on by several workers (Mellon and Jost 1926, W J Wilson 1933, Morris, Sellers and Brown 1941, Morris, Barnes, and Sellers 1943). On ordinary agar the colonies after 24 hours' incubation are only about 0.2-0.3 mm in diameter. According, however, to the original observations of Jacobsen (1910) which have since been confirmed many times, colonies of more normal size are formed on media containing assimilable sulphur compounds.

As has already been noted in the previous section certain species notably *Salm paratyphi B*, give rise under favourable conditions to a mucoid growth. Sometimes the colonies are mucoid after 24 hours' incubation, they are about twice the size of normal colonies and resemble large drops of mucilage (Fletcher 1918). More often the mucoid appearance is developed as a secondary phenomenon after prolonged incubation. Thus, if an agar plate is inoculated in three or four



FIG 148—*Salm typhi*

Surface colonies on agar 24 hours  
at 37° C ( $\times 8$ )



FIG 149—*Salm typhi murium*

Surface colonies on agar 24 hours  
at 37° C ( $\times 8$ )

places with the point of a needle, and after one day's incubation at 37° C is left at room temperature for a few days, large colonies are formed characterized by a depressed centre surrounded by a luxuriant mucoid wall. The 'mucoid wall test', or *Schleimwall Versuch* described originally by Müller (1910), is of some differential value, being common with most freshly isolated d tartrate negative strains of *Salm paratyphi B* and generally negative with *Salm typhi murium* (Kauffmann 1941). The mucoid material contains a polysaccharide (Birch Hirschfeld 1935) which appears to be antigenically homogeneous no matter by what species of *Salmonella* it is formed.

**Resistance to Heat and to various Chemical Substances**—Most members of this group are killed by exposure to a temperature of 50° C for about 1 hour or of 60° C for 15-20 minutes. Many observations have been made on the resistance of salmonellae to different chemical reagents chiefly in an endeavour to prepare selective media on which the growth of coliform bacilli would be inhibited. Malachite green in suitable concentration kills *Bact coli* or inhibits its growth without exerting the same effect on *Salm typhi* (Loeffler 1903, 1906, Lentz and Tietz 1903, 1905). There are other green dyes that have a similar selective action,

and the studies of Browning Gilmour, and Mackie (1913) and of Krumwiede and Pratt (1914) have shown that brilliant green gives the best results. To this dye bacilli of the paratyphoid group are most resistant, the typhoid bacillus is somewhat less resistant, whereas the dysentery bacilli and still more the coliform and paracolon bacilli are very susceptible. Caffeine (Roth 1903, Hoffman and Ficker 1904) and lithium chloride (Gray 1931, Havens and Mayfield 1933) are other substances that inhibit the growth of *Bact. coli* in concentrations that have no effect on the typhoid bacillus. Sodium tetrathionate is now being extensively used to favour the growth of salmonellae at the expense of the coliform bacilli (Muller 1923, Schafer 1934-35, Jones 1936)—an action that appears to be due not to any inhibitory action it possesses on coliform bacilli, but to the ability of most organisms of the *Salmonella* group to reduce this substance and use it as

a source of energy (Pollock, Knox, and Gell 1942). Sodium desoxycholate in the presence of certain other substances, inhibits the growth of coliform but not of dysentery or salmonella bacilli and is being used increasingly in the preparation of selective media for the isolation of intestinal pathogens (Leifson 1935, Hynes 1942). Selenium salts are also of value for the same purpose (Guth 1916, Leifson 1936).

**Biochemical Activities.**—As stated in the definition of the genus, the members of this group do not ferment lactose, sucrose or adonitol. Salicin also is rarely fermented, and then only late. On the other hand glucose, mannitol, dulcitol, and sorbitol, and almost invariably maltose and dextrin, are fermented. Apart from a few species, like *Salm. typhi* and *Salm. gallinarum*, gas is formed, though anaerogenic variants are not uncommon. Among the ordinary sugars, arabinose, xylose, trehalose and inositol are useful for the differentiation of species. As examples, we may quote the failure of *Salm. cholerae* *sus* to ferment arabinose or trehalose, which is of value in distinguishing it from *Salm. paratyphi* C, and the failure of *Salm. zagreb* to ferment inositol, which is of value in distinguishing it from *Salm. saint-paul*. The power to ferment rhamnose often varies with different strains of the same species, this is sometimes made use of in the separation of epidemiological types, as, for instance, in *Salm. typhi* *murum* (Edwards and Bruner 1940a).

Also of value in the differentiation of species are the organic acids which were introduced by Brown, Duncan, and Henry (1924, 1926). Those commonly used are *d* tartrate, *l* tartrate, *l* tartrate sodium citrate, and sodium mucate. Failure to ferment *l* tartrate is fairly common, the other four acids are acted upon by most species.

In the past considerable attention has been devoted to certain special reactions. One of these first described by Stern (1916) consists in growing the organisms under test in a fuchsin sulphite glycerol meat-extract medium. Some organisms known as Stern positive produce in this medium a deep lilac colour within 3 days. If the medium is pink to red after a week, the strain is regarded as 'Stern negative'. An intermediate group of strains sometimes regarded as giving a weakly positive reaction, turn the medium deep red, purple or lilac in between 3 to 7 days. The reaction is apparently due to the formation of an aldehyde. It is certainly not due solely to acid formation.



FIG. 150.—*Salm. typhi*  
24 hours culture  
on agar slope

TABLE 46

FERMENTATION REACTIONS OF *SALMONELLA* GROUP (MODIFIED FROM KAUFFMANN 1941)

Species	Gas	Anal. inosc.	Dulc. 4	Inocul. 4	Rhamnose	Trehalose	Xylose	d-tartrate	l-tartrate	s-tartrate	Citrate	Mucate	Stern 6, 9, 10	H <sub>2</sub> S	Gelatin liquefaction
<b>SEROLOGICAL GROUP A</b>															
<i>Salm. paratyphi A</i>	+	+	+	-	+	+	-	-	-	-	-	-	-	+	-
<b>SEROLOGICAL GROUP B</b>															
<i>Salm. paratyphi B</i>	+	+	+	v	v	+	+	v	v	v	+	+	v	+	-
<i>Salm. abony</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-
<i>Salm. typhi murium</i>	+	+	+	v	v	v	v	v	+	v	+	+	+	+	-
<i>Salm. stanley</i>	+	+	+	-	+	+	+	+	+	-	+	+	v	+	-
<i>Salm. heidelberg</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-
<i>Salm. chester</i>	+	+	+	v	+	+	+	+	+	+	+	+	+	+	-
<i>Salm. n. san-diego</i>	+	+	+	-	+	+	+	+	x	-	+	v	+	+	-
<i>Salm. saint-paul</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-
<i>Salm. zagreb</i>	+	+	+	-	+	+	+	+	+	+	+	+	+	+	-
<i>Salm. kaposvar</i>	+	+	+	-	+	+	+	+	+	v	+	+	-	+	-
<i>Salm. reading</i>	+	+	+	v	v	+	+	+	x	-	+	+	v	+	-
<i>Salm. derby</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-
<i>Salm. essen</i>	+	+	+	-	+	+	+	+	+	+	+	+	+	+	-
<i>Salm. budapest</i>	+	+	+	v	+	+	+	+	+	+	+	+	v	+	-
<i>Salm. californica</i>	+	+	+	+	+	+	+	+	+	-	+	+	+	+	-
<i>Salm. brandenburg</i>	+	+	+	-	+	+	+	+	+	+	+	+	v	+	-
<i>Salm. bispebjerg</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-
<i>Salm. abortus-equi</i>	+	+	+	-	+	+	+	+	x	+	+	-	+	-	-
<i>Salm. abortus-ovis</i>	+	x	v	-	x	-	v	+	x	x	+	-	-	x	-
<i>Salm. altendorf</i>	+	+	+	+	+	+	+	+	+	-	+	+	+	+	-
<i>Salm. salinae</i>	+	+	+	-	+	+	+	+	+	+	+	+	+	+	+
<i>Salm. abortus-bovis</i>	+	+	+	-	v	+	+	+	+	-	+	+	+	+	+
<i>Salm. bredeney</i>	+	+	+	-	+	+	+	+	x	-	+	+	+	+	-
<i>Salm. schleissheim</i>	+	+	-	-	+	+	v	+	x	-	+	+	+	+	+
<b>SEROLOGICAL GROUP C</b>															
<i>Salm. paratyphi C</i>	+	+	+	-	+	+	+	+	+	+	+	-	+	+	-
<i>Salm. cholerae-suis</i>	+	-	x	-	+	-	+	+	-	v	+	x	-	x	-
<i>Salm. cholerae-suis</i> var <i>kummenlof</i>	+	-	x	-	+	-	+	v	x	x	+	x	-	+	-
<i>Salm. typhi-suis</i>	+	+	+	-	+	+	+	-	+	+	+	+	-	+	-
<i>Salm. thompson</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-
<i>Salm. cardiff</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
<i>Salm. virchow</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
<i>Salm. oranienburg</i>	+	+	+	-	+	+	+	+	x	-	+	+	v	+	-
<i>Salm. potadam</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	-
<i>Salm. bareilly</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
<i>Salm. harford</i>	+	+	+	-	+	+	+	+	+	+	+	+	+	+	+
<i>Salm. mukawanima</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
<i>Salm. monterideo</i>	+	+	+	v	+	+	+	+	v	-	+	+	+	+	+
<i>Salm. oslo</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
<i>Salm. amersfoort</i>	+	+	+	-	+	+	+	+	+	+	+	+	+	+	+
<i>Salm. braenderup</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
<i>Salm. tennessee</i>	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+
<i>Salm. newport</i>	+	+	+	-	+	+	+	+	+	+	+	+	+	+	+
<i>Salm. kolibus</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
<i>Salm. bovis morbificans</i>	+	+	+	+	+	+	+	+	+	v	+	+	+	+	+
<i>Salm. muenchen</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+
<i>Salm. oregon</i>	+	+	+	+	+	+	+	v	+	+	+	+	v	+	+
<i>Salm. mexicana</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+

TABLE 46—continued

FERMENTATION REACTIONS OF SALMONELLA GROUP (MODIFIED FROM KAUFFMAN 1941)

Species	Ga	Arabinose	Dulcitol	In which	Rhamnose	Trehalose	Xylose	d-tartrate	l-tartrate	ε-tartrate	Citrate	Mucate	Serine hydrolysis	H <sub>2</sub> S	Starch liquefaction
<b>SEROLOGICAL GROUP C—</b>															
continued															
<i>Salm. manhattan</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. narashino</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. glostrup</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. litchfield</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. duesseldorf</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. bonariensis</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. amherstiana</i>	+	+	+	+	+	+	—	+	+	—	+	+	+	+	—
<b>SEROLOGICAL GROUP D</b>															
<i>Salm. typhi</i>	—	+	+	—	—	+	+	+	—	—	+	+	—	+	—
<i>Salm. enteritidis</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
var danyers	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
var essen	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
var chaco	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. dublin</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. rostock</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. moscow</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. blegdam</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. beria</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. eastbourne</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. oendou</i>	—	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. anarimon</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. dar-es-salaam</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. goettigen</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. panama</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. clabornei</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. jariana</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. pullorum</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. gallinarum</i>	—	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<b>SEROLOGICAL GROUP E</b>															
<i>Salm. london</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. gire</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. uganda</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. anatum</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. ouenater</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. nyborg</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. vejle</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. amager</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. zanibar</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. shangana</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. meleagridis</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. lezington</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. newington</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. seldania</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. new-brunswick</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. illinois</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. taksony</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. senftenberg</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
var newcastle	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. miloese</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. ansbury</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—

TABLE 46—continued

FERMENTATION REACTIONS OF *SALMONELLA* GROUP (MODIFIED FROM HAUFFMANN 1941)

Species	Gas	Arabinose	Dulcitol	Inositol	Rhamnose	Trehalose	Xylose	d-tartrate	L-tartrate	s-tartrate	Glycerol	Mucate	Stern glycerol	H <sub>2</sub> S	Gelatin liquefaction
<b>OTHER SEROLOGICAL GROUPS</b>															
<i>Salm. aberdeen</i>	+	+	+	—	+	+	+	+	+	+	+	+	++	+	—
<i>Salm. rubislaw</i>	+	+	+	+	+	+	+	+	+	+	+	+	++	+	—
<i>Salm. soli</i>	+	+	+	—	+	+	+	+	+	+	+	+	++	+	—
<i>Salm. poona</i>	+	+	+	—	+	+	+	+	+	+	+	+	++	+	—
<i>Salm. worthington</i>	+	+	x	+	+	+	+	+	+	+	+	+	++	+	—
<i>Salm. wickita</i>	+	+	+	—	+	+	+	+	+	+	+	+	++	+	—
<i>Salm. habana</i>	+	+	+	+	+	+	+	—	+	+	+	+	++	+	—
<i>Salm. heves</i>	+	+	+	—	+	+	—	+	+	+	+	+	+	+	—
<i>Salm. carrau</i>	+	+	+	—	+	+	+	+	x	—	+	+	++	+	—
<i>Salm. onderstepoort</i>	+	+	+	—	+	+	+	+	+	+	+	+	++	+	—
<i>Salm. florida</i>	+	+	+	—	+	+	+	+	+	+	+	+	++	+	—
<i>Salm. madelia</i>	+	+	+	—	+	+	+	+	—	—	+	+	++	+	—
<i>Salm. huttingfoss</i>	+	+	+	+	+	+	+	+	+	+	+	+	++	+	—
<i>Salm. gaminsara</i>	+	+	+	+	+	+	+	+	+	+	+	+	++	+	—
<i>Salm. szentes</i>	+	+	+	—	+	+	+	+	+	+	+	+	++	+	—
<i>Salm. lirkoe</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. cerro</i>	+	+	+	—	+	+	x	+	+	—	+	+	++	+	—
<i>Salm. kentucky</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. minnesota</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. tel-aviv</i>	+	+	+	—	+	+	+	+	+	+	+	+	++	+	—
<i>Salm. ballerup</i>	+	+	+	—	+	+	+	+	+	+	+	+	++	+	—
<i>Salm. hormachae</i>	+	+	+	—	+	+	+	+	+	+	+	+	++	+	—
<i>Salm. urbana</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. adelade</i>	+	+	+	—	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. inverness</i>	+	+	+	+	+	+	+	+	+	+	+	+	+	+	—
<i>Salm. arizona</i>	+	+	—	+	+	+	+	—	—	—	+	+	++	+	±

Gas { + = normal amount  
± = small or absent

Arabinose xylose and organic acids { + = positive within 3 days.  
+1 = positive usually 4-8 days  
x = late or irregularly positive often negative  
v = variable with different strains

Stern & glycerol { ++ lilac in 1-3 days  
+ = purple or lilac in 4-8 days

Gelatin { + = rapid liquefaction.  
± = slow liquefaction.

The second reaction was described by Bitter Weigmann, and Habs (1926). It is essentially a test of the ability of the organism to grow in a synthetic medium containing ammonium salts as the main source of nitrogen. A 1 per cent solution of sugar of which the most generally useful is rhamnose is added to test the fermentative power of the strain under these conditions. Strains that grow readily ferment the sugar and are referred to as ammonium strong. Strains that grow poorly and fail to ferment the sugar are referred to as ammonium weak. The result of the test may be negative not because the organism cannot grow in an ammonium medium but because like some strains of *Salm. paratyphi B* it fails to ferment rhamnose. In practice the inoculated medium is incubated for 20 hours at 37° C., methyl red is then added to determine

whether or not the pH has been lowered to the point at which this indicator gives its characteristic red colour. Instead of the liquid medium a citrate agar medium introduced by Simmons (1926) may be employed. This has been extensively used by Hohn and Herrmann (1936 c). According to Kauffmann (1941) less reliance can be placed on the ammonium reaction than on the fermentation of organic acids. The ammonium reaction and the reaction in Stern's medium are however useful at times, as, for instance in the differentiation of the fermentative types of *Salm. enteridis* (Kauffmann 1935b).

In litmus milk the great majority of species produce transitory acidity followed by permanent alkalinity in 3 days but with a few species like *Salm. paratyphi A*, *Salm. abortus-ovis*, *Salm. typhi suis*, *Salm. typhi*, *Salm. sendai* and *Salm. pullorum* the reaction in 3 days is neutral or slightly acid. On the whole little information of differential value is gleaned from a study of the reaction in litmus milk and Kauffmann (1941) has now ceased to include this medium in his series of routine tests.

With the exception of variant colonies of *Salm. eastbourne* and very occasional variants of *Salm. enteritidis* and *Salm. panama* (see Seligmann and Saphra 1943) indole production is uniformly negative. The majority of strains form  $H_2S$  but *Salm. typhi-suis*, *Salm. sendai*, *Salm. gallinarum* var. *dunburg* and *Salm. senftenberg* var. *newcastle* fail to do so.  $H_2S$  formation is variable with *Salm. typhi* and *Salm. paratyphi A*, some strains being positive and others negative. Gelatin liquefaction is very uncommon, but *Salm. abortus-ovis* and *Salm. schleissheim* liquefy it rapidly and *Salm. dar-es-salaam* and *Salm. arizonae* very slowly. Urea is not decomposed.

**Antigenic Structure.**—As has already been noted, the classification of the *Salmonella* group is now firmly based on the antigenic structure of the species of which it is composed. The arrangement of the various antigenic components in the cells and flagella of the salmonellae has already been described in Chapter 8 in relation to the general problem of antigenic structure and in Chapter 9 in relation to the study of bacterial variation. Before considering the different species that have been differentiated by this method of analysis, we may however recall the relevant facts and consider some of them in greater detail.

In the *Salmonella* group of bacilli we are dealing in the main with flagellated organisms. We therefore have to consider both the H (flagellar) and O (somatic) antigens. By appropriate methods we can test for these components separately. A formalized broth culture of a flagellated species readily agglutinates in the presence of the homologous H agglutinin but responds very poorly if at all to the homologous O agglutinin. The reason for this behaviour is, perhaps, that the formalin fixes the flagella over the bacterial surface in such a way that the somatic antigens are no longer exposed to the action of the O agglutinins. To test for the O antigens we can employ a bacterial suspension that has been treated with hot alcohol thus removing or inactivating the H antigens.

In respect of their flagellar antigens many of the species with which we are concerned are diphasic—that is to say their flagella may assume two alternative forms. Originally the two phases were referred to as *specific* and *group* but later when another type of phase variation was discovered the so-called  $\alpha$   $\beta$  variation this nomenclature became no longer generally applicable. It is now usual to call the two phases 1 and 2 and to restrict the terms *specific* and *group* to organisms in which Phase 1 contains a, b, c etc. antigens and Phase 2 1, 2, 3 etc. antigens. Speaking generally Phase 1 tends to contain more specific, and Phase 2 less specific antigens but it will be convenient to reserve a full description of

the variations in the antigenic constitution of the two phases till a little later in this chapter (p 716) It is sufficient to remark here that in Phase 1 there are one or more antigenic components, and in Phase 2 there are two or more

The somatic antigens are monophasic, though occasionally a given strain may differ from the type species in its lack of one of the two, three, or four antigenic components which most members possess

The identification and labelling of the *Salmonella* antigens was initiated by White (1926, 1929a, b), and continued and extended by Kauffmann (1929a, b, 1930b, 1931, 1934a) In their earlier studies the two investigators used a different system of labelling, so that descriptions given in the English and German papers during the period can be correlated only by the aid of an appropriate key giving the equivalent numbers and letters in the two systems (see Lovell 1932a) Recently however, the terminology introduced by Kauffmann has been adopted for general use by a special sub-committee of the International Society for Microbiology (Report 1934), and each member of the *Salmonella* group is now allotted an antigenic formula based on this system of notation

The somatic (O) antigens are accorded Roman numerals The flagellar antigens of Phase 1 are accorded small letters, these antigens have already illustrated the limitations of an alphabetical notation by exceeding twenty six in number By convention, those discovered later than the antigen that received the label y have been accorded an additional distinguishing numeral, z<sub>1</sub>, z<sub>2</sub> and so on The flagellar antigens of Phase 2 are labelled in two different ways At first they were accorded arabic numerals, but later it was found that the second phase of certain species uniformly contained the antigens e and n, often associated with x or with one of the z series of antigens It thus happens that Phase 2 may contain antigenic components of the 1, 2, 3 series or of the e n series The position is confusing, because both the e and some of the z antigenic components may be found in the first phase More recently, strains have been described in which Phase 2 contains neither the 1, 2, 3 nor the e, n series, but instead antigenic components that are characteristic of Phase 1

A few examples may be given to illustrate the use of this notation. The formula for *Salm paratyphi* A, which is a monophasic flagellated bacillus existing only in Phase 1 is [I] II, XII a — The square brackets indicate that the I somatic antigen may be missing The three dots after XII mean that traces of other somatic antigens are present, but are excluded from the antigenic formula for the sake of simplicity The a represents the flagellar antigen, and the dash following it shows that there is no second phase The formula for *Salm paratyphi* B is [I] IV, [V] XII b ↔ 1 2 The square brackets show that antigenic components I and V may be missing, b is the flagellar antigen of Phase 1, and 1, 2 the flagellar antigens of Phase 2 Again the dots following XII and 2 indicate an abbreviation of the antigenic formula The double tipped arrow shows that the two flagellar phases are reversible The formula for *Salm amersfoort* is VI<sub>1</sub> VI<sub>2</sub> VII d ↔ e, n, x It will be seen that there are two components to the VI somatic antigen, and that, as indicated by the presence of the e n antigens in Phase 2, the organism shows the α β type of phase variation (see p 716) The formula for *Salm meleagridis* is III Δ XXVI e, h ↔ 1 w Phase 2 contains neither the 1, 2, 3 nor the e n series of antigens, but two antigenic components 1 and w, that are normally found in Phase 1 of other organisms *Salm gillinarum* which is non motile has the simple formula [I] IX, XII —, the two dashes showing that there is neither a first nor a second flagellar phase An organism so far met with only in Phase 2, is *Salm abortus equi*, which has the formula IV, XII — e, n, x, the dash shows that Phase 1 is absent



One of the most interesting features of the *Salmonella* group is the way in which the same antigenic components recur in different combinations. Thus different O antigens are found in combination with the same H antigens. The same set of Phase 2 antigens are replaced by different Phase 1 antigens when different diphasic species change from the group to the specific or from the  $\beta$  to the  $\alpha$  phase. The same set of antigenic components recurs in different combinations in Phase 2 of different species and so on. The implications of these facts in relation to the evolution of the *Salmonella* group are discussed by White (1936).

**The Kauffmann White Diagnostic Scheme**—It is clear that the observed distribution of these antigenic components forms a basis for a natural scheme of classification and this has been adopted in the Kauffmann White scheme proposed for international adoption by the *Salmonella* Sub-committee (Report 1934). It seems reasonable to regard the antigenic structure of the bacterial cell as more fundamental than the antigenic structure of the appended flagella. The *Salmonella* group as a whole has therefore been divided into sub-groups each of which shares a common somatic antigen. Where more than one somatic antigen is present one of these antigens is regarded as determining the sub-group to which the species concerned shall be allocated. Thus, Group B consists of those organisms that possess the O antigen IV or the antigens IV or V. Group D consists of those organisms that possess the O antigen IX, and so on. The group letter it should be noted forms no constituent part of the name of any species or of its antigenic formula. The groups display the natural relations of the different salmonellae but the antigenic components that determine those relationships are labelled according to the Kauffmann convention. It should perhaps be recalled that there is a fundamental difference between this scheme of classification and that adopted in describing the antigenic structure of other bacterial groups. A

Group B haemolytic streptococcus for instance means a haemolytic streptococcus that possesses the Group B antigen, and that antigen has no other label.

It must be made clear that the Kauffmann White scheme is essentially a scheme for the differentiation in practice of the various species. It is not, nor does it pretend to be a record of the complete antigenic structure of each organism. The antigenic constitution of most organisms is far more complex than is suggested by the formulae given in the table. Only the major antigens or those antigens that are of differential importance are recorded. It follows that owing to the possession of minor somatic or flagellar antigens cross agglutination may occur between organisms which, judged by the table possess no common factor. It follows too that as further members of the *Salmonella* group are discovered alterations will have to be made in some of the present formulae if the diagnostic value of the table is to be preserved.

Another warning must be issued. Many of the antigenic components identified by a single numeral or letter are themselves complex, consisting of two or more fractions. For instance the V and VI somatic antigens each comprise two portions labelled  $V_1$  and  $V_2$  and  $VI_1$  and  $VI_2$ . The VII somatic antigen contains three portions,  $XII_1$ ,  $XII_2$ , and  $XII_3$ . The d flagellar antigen contains five partial antigens d,  $d_1$ ,  $d_2$ ,  $d_3$  and  $d_4$ . The fact therefore that two species are represented in the table as containing for example the d antigen, does not mean that their d antigens are necessarily identical. One of the species may contain one pair of the d fractions and the other species another pair so that though both species will be agglutinated by an anti-d serum prepared against all the fractions,

TABLE 47

KAUFFMANN WHITE SCHEME.

Group	Type	O Antigen.	H Antigen.	
			1 Phase.	2 Phase
Group A	<i>S paratyphi A</i>	[I] II XII	a	—
Group B	<i>S paratyphi B</i>	[I], IV [V], XII	b	1 2
	<i>S abony</i>	[I] IV V XII	b	e n x
	<i>S typhi murium</i>	[I], IV [V], XII	i	1 2 3
	<i>S stanley</i>	IV V XII	d	1 2
	<i>S hesdelberg</i>	IV V XII	r	1 2 3
	<i>S chester</i>	IV [V], XII	e h	e n x
	<i>S san-diego</i>	IV [V] XII	e h	e n x
	<i>S salinatus</i>	IV XII	i e h	e n x <sub>11</sub>
	<i>S saint paul</i>	I IV V XII	e h	1 2 3
	<i>S -agreb</i>	IV V XII	e h	1 2
	<i>S laposrar</i>	IV V XII	e (l)	1 3
	<i>S koeln</i>	IV V XII	v	1 2 3
	<i>S reading</i>	IV XII	e h	1 5
	<i>S derby</i>	[I] IV XII	f g	—
	<i>S kaapstad</i>	IV XII	e l	1 7
	<i>S essen</i>	IV XII	g m	—
	<i>S budapest</i>	I IV XII	g t	—
	<i>S cal fornia</i>	IV XII	g m t	—
	<i>S brandenburg</i>	IV XII	l v	e n x <sub>1</sub>
	<i>S bapchberg</i>	I IV XII	a	e n x
	<i>S abortus eq u</i>	IV XII	—	e n x
	<i>S abortus-ovis</i>	IV XII	c	1 6
	<i>S arechavaleta</i>	IV [V] XII	a	1 7
	<i>S altendorf</i>	IV XII	c	1 7
	<i>S abortus bovis</i>	[I] IV XXVII XII	b	e n x
	<i>S schwarzengrund</i>	I IV XXVII XII	d	1 7
	<i>S bredeney</i>	I IV [XXVII] (XII)	l v	1 7
	<i>S achleissheim</i>	IV XXVII XII	b x <sub>11</sub>	—
Group C	<i>S paratyphi C</i>	VI <sub>1</sub> VI <sub>2</sub> VII (VI)	c	1 5
	<i>S cholerae suis 1</i>	VI <sub>1</sub> VII	c	1 5
	<i>S cholerae-suis 2</i>	VI <sub>2</sub> VII	c	1 5
	<i>S typhi suis</i>	VI <sub>1</sub> , VI <sub>2</sub> , VII	c	1 5
	<i>S thompson</i>		k	1 5
	<i>S cardiff</i>		k	1 10
	<i>S turchone</i>		r	1 2 3
	<i>S infantis</i>		r	1
	<i>S oran end ury</i>		m t	—
	<i>S potadam</i>		l v	e n x <sub>11</sub>
	<i>S bareilly</i>		y	1 5
	<i>S hartford</i>		y	e n x
	<i>S mikawas ma</i>		y	e n x <sub>11</sub>
	<i>S montevideo 1</i>	VI <sub>1</sub> , VII	g m s	—
	<i>S oelo</i>		a	e n x
	<i>S montevideo 2</i>		g m s	—
	<i>S amersfoort</i>		d	e n x
	<i>S brænderup</i>		e h	e n x <sub>11</sub>
	<i>S georg a</i>	VI VII	b	e n x <sub>1</sub>
	<i>S concord</i>	VI VII	l v	1 2 3
	<i>S tennessee</i>	VI VII	x <sub>11</sub>	—
	<i>S guers</i>	VI, VIII	e l	1 2
	<i>S newport</i>		e h	1 2 3
	<i>S kottbus</i>		e h	1 5

TABLE 47 (continued)

Group	Type	O-Antigen	H Antigen	
			1 Phase	2 Phase
Group C (cont'd)	<i>S. bovis moribundus</i>	VI, VIII	r	1 5
	<i>S. muenchen</i>		d	1 2
	<i>S. oregon</i>		d	1 2 3
	<i>S. mexicana</i>		d	1 2 4
	<i>S. manhattan</i>		d	1 5
	<i>S. narashino</i>		a	e n x
	<i>S. glostrup</i>		z <sub>10</sub>	e n z <sub>11</sub>
	<i>S. litchfield</i>	VI, VIII VI VIII VIII (VIII)	1 v	1 2, 3
	<i>S. duesseldorf</i>		z <sub>1</sub> z <sub>11</sub>	—
	<i>S. donar ensis</i>		i	e n x
	Unnamed type		e h	1 2
Group D	<i>S. virginea</i>	IX, XII [VI]	d	—
	<i>S. amherstiana</i>		1 (v)	1 6
	<i>S. typhi</i>		d	—
	<i>S. enteritidis</i>		g m	—
	<i>S. dublin</i>		g p	—
	<i>S. rostock</i>		g p u	—
	<i>S. moscow</i>		g q	—
	<i>S. blegdam</i>		g m q	—
	<i>S. pensacola</i>		g m t	—
	<i>S. berta</i>		f g t	—
	<i>S. eastbourne</i>		e h	1 5
	<i>S. sendai</i>		a	1 5
	<i>S. loma linda</i>		a	e n x
	<i>S. onartimon</i>		b	1 2
	<i>S. dar-es-salaam</i>		1 w	e n
	<i>S. goettingen</i>		1 v	e n z <sub>1</sub>
	<i>S. durban</i>		a	e n, z <sub>11</sub>
	<i>S. panama</i>		1 v	1 5
	<i>S. italica</i>		1 v	1 11
	<i>S. claibornes</i>		h	1 5
Group E	<i>S. gallinarum</i>		a	—
	<i>S. napolitana</i>		1 z <sub>11</sub>	e n x
	<i>S. javanica</i>		1 z <sub>11</sub>	1 5
	<i>S. canastota</i>		z <sub>11</sub>	1 5
	<i>S. pullorum</i>		—	—
	<i>S. london</i>	III V XVI	1 v	1 6
	<i>S. greigii</i>		1 v	1 7
	<i>S. uganda</i>		1 z <sub>11</sub>	1 5
	<i>S. anatum</i>		e h	1 6
	<i>S. muenster</i>		e h	1 5
	<i>S. nyborg</i>		e h	1 7
	<i>S. vejle</i>		e h	1 2 3
	<i>S. amager</i>		y	1 2 3
	<i>S. zanibar</i>		h	1 5
	<i>S. shangana</i>		d	1 5
	<i>S. melengrindae</i>		e h	1 w
	<i>S. lexington</i>		z <sub>1</sub>	1 5
	<i>S. wellerstedtii</i>		r	z <sub>1</sub>
	<i>S. newington</i>	III V	e h	1 6
	<i>S. selandica</i>		e h	1 7
	<i>S. new-brunswick</i>		1 w	1 7
	<i>S. cambridge</i>		1 w	e h
	<i>S. illinois</i>		z <sub>1</sub>	1 5

TABLE 47 (continued)

Group	Type	O Antigen	H Antigen	
			1 Phase	2 Phase
Group E (contd.)	<i>S. talsony</i>	I III XIX	1	2 <sub>4</sub>
	<i>S. senftenberg</i>		g s, t	—
	<i>S. nilotica</i>		d	2 <sub>4</sub>
	<i>S. amsbury</i>		z <sub>27</sub>	—
Other Groups	<i>S. pretoria</i>	XI	k	1 2 3
	<i>S. aberdeen</i>	XI	i	1 2 3
	<i>S. rubislaw</i>	VI	r	e n x
	<i>S. solt</i>	VI	y	1 5
	<i>S. grumpensis</i>	XIII XXII	d	1 7
	<i>S. poona</i>	XIII XXII	z	1 6
	<i>S. borbeck</i>	XIII XXII	l v	1 6
	<i>S. scorth ngto</i>	I XIII XXIII	l w	z
	<i>S. ic ch la</i>	I XIII XXIII	d	—
	<i>S. haba a</i>	I XIII XXIII	f g	—
	<i>S. m ss ss ppt</i>	I XIII XXIII	b	1 5
	<i>S. ke es</i>	(I) VI XIV XXIV	d	1 5
	<i>S. carra</i>	VI XIV XXIV	y	1 7
	<i>S. n adel a</i>	(I) VI XIV XXV	v	1 7
	<i>S. onderstepoort</i>	(I) VI XIV XXV	e (h)	1 2
	<i>S. florida</i>	(I) VI XIV XXV	d	1 7
	<i>S. sunda all</i>	(I) VI XIV XXV	z	e n x z <sub>14</sub>
	<i>S. horsham</i>	(I) VI XIV XXV	l, v	e o x
	<i>S. hillingfoss</i>	XVI	b	e n x
	<i>S. gam nara</i>	XVI	d	1 7
	<i>S. arentes</i>	XVI	k	1 2 3
	<i>S. kirkee</i>	XVII	b	1 2
	<i>S. cerro</i>	XVIII	z <sub>4</sub> z <sub>23</sub> z <sub>24</sub>	—
	<i>S. kentucky</i>	(VIII) XX	i	z <sub>4</sub>
	<i>S. m nnesota</i>	XXI XXVI	b	e n x
	<i>S. tel-aviv</i>	XXVIII	y	e n x
	<i>S. pomona</i>	XXVIII	y	1
	<i>S. ballerup</i>	XXIX (VI)	z <sub>14</sub>	—
	<i>S. hormæches</i>	XXIX (VI)	z <sub>20</sub>	—
	<i>S. i rbana</i>	XXX	b	e n x
	<i>S. arizona</i>	XXXIII	z <sub>4</sub> z <sub>23</sub> z <sub>24</sub>	—
	<i>S. adela de</i>	XXXV	fg	—
	<i>S. inverness</i>	XXXVIII	k	1 6
	<i>Bact. coli 1</i>	XXXI (VI)	—	1 2
	<i>Bact. coli 2</i>	XXXII	—	1 5
	<i>Bact. coli 3</i>	IV V VII	z <sub>20</sub>	—
	<i>Bact. coli 4</i>	IV (XXVII) VII	z <sub>21</sub>	—
	<i>Bact. coli 5</i>	(I) VI XIV XXV	z <sub>22</sub>	—

Numerous other coliform and paracolon bacilli containing O or H antigens found in the *Salmonella* group have been described. *Salmonella* O antigens have likewise been met with in members of the Flexner dysentery and the *Pasteurella* groups.

- [ ] These antigens may be missing  
 ( ) — Only a part of the antigen is present  
 = Very much abbreviated formula.  
 = Not fully investigated

absorption of such a serum with one species will not remove the agglutinins for the other species. The g and t antigens are likewise complex (Edwards, Bruner, and Hinshaw 1940). In the descriptions of the individual species we have recorded many of the minor and the partial antigens that do not appear in the Kauffmann-White scheme.

This is not the place to describe the practical identification of *Salmonella* species. For information on this subject readers are referred to publications by Kauffmann (1939b, 1941) and Edwards and Bruner (1942c).

### Antigenic Variation.

The variations that occur within this group have been studied by many observers, and these studies have been particularly fruitful in adding to our knowledge of the mechanisms of bacterial variation as a general phenomenon. For this reason they have been discussed in some detail in Chapters 8 and 9. There is no need to repeat that discussion here, but for the sake of clarity and completeness, it will be well to give a brief resumé of the antigenic variations that may be met with in the *Salmonella* group.

**Phase variation affecting the H antigens.**—In *specific-group phase variation*, which was described by Andrewes (1922, 1923), the flagella may assume two alternative antigenic forms. The antigenic components of the specific phase are either peculiar to the particular species concerned or are shared by only a few other species. On the other hand, the group phase contains antigenic components that are shared by many other species. Any given culture of a particular diphasic strain may consist entirely of bacilli in the specific phase or entirely of bacilli in the group phase, or may contain representatives of both phases. A bacillus in one phase, though always capable of giving rise to descendants in the alternative phase, usually maintains a constant phase over a number of generations. If therefore, we prepare plate cultures of a diphasic organism and make numerous subcultures, each from a single colony, we should as a rule obtain some suspensions in the specific, others in the group phase. If these are killed by the addition of formalin after 18–24 hours' growth, there will usually not have been time for any change in phase to occur.

The  $\alpha$   $\beta$  phase variation, which was described by Kauffmann and Mitsui (1930), consists in a variation that does not differ in principle from specific-group variation. Instead, however, of Phase 2 containing the antigenic components 1, 2, 3, etc., it contains the antigenic components e, n, etc. It will be noted that the antigens in Phase 1 are similar in both types of variation, being of the a, b, c type, but that in Phase 2 they differ.

A special type of  $\alpha$   $\beta$  phase variation has been described by Edwards and Bruner (1933), which may perhaps be regarded as a third type. It was found that, in *Salm. thompson*, Phase 2, instead of containing the 1, 2, 3 or the e, n . . . type of antigens, contained an antigenic component of the a, b, c series, which had previously been met with only in Phase 1. In other words, organisms of this type have what, for practical purposes may be regarded as two alternative first phases.

It must be realized that, in organisms showing diphasic variation, strains may occur which remain persistently in one phase—usually Phase 2—and in which the alternative phase can be demonstrated only by growth in the presence of immune serum containing agglutinins to the Phase 2 antigens. *Salm. thompson*, for example, is almost invariably in the non-specific phase on first isolation and the k antigen of Phase 1 may not be demonstrable till after two or three subcultures in the presence of an anti-group serum.

**Artificial phase variation** is a phenomenon that appears at present to be more of academic than of practical importance though its significance must remain doubtful till further work has been carried out. It was observed by Kauffmann (1936c) that, if a certain strain of *Salm. typhi* was cultured in broth containing agglutinating serum to the normal

d antigen this antigen was lost and replaced by a new antigen j which had never been met with in typhoid bacilli under natural conditions. Similarly, the b, z<sub>12</sub> antigens of *Salmonella schlesheim* may be replaced by a new antigen z<sub>4</sub>. In both these organisms which are monophasic the antigen appearing in the induced phase belongs to the series, a, l, c commonly found in Phase 1. Bruner and Edwards (1911a) however, were able to induce in the monophasic *Salmonella paratyphi A* a second phase containing the l, 5 antigens which are characteristic of the group series of antigens of Phase 2. An even more remarkable series of induced variations was recorded by Edwards and Bruner (1939) in *Salmonella abortus-equi*. This organism exists normally only in Phase 2 in which it possesses the antigens e, n, x. By growth in appropriate sera a Phase 1, containing the antigen a, was induced and a Phase 3, containing the antigen z<sub>4</sub>. All three phases were reversible. It is by no means always easy to decide whether a new phase that appears as the result of growth in immune serum is to be regarded as an artificial phase or as the alternative phase of an organism that is generally, though not invariably monophasic. Legitimate difference of opinion may exist on this point. For example *Salmonella cholerae-suis* var *kunzen-dorf* was for many years regarded as a well-established monophasic type occurring in the group phase. When it was found, however, by Bruner and Edwards (1939) that a specific phase could be induced containing the same antigen e as *Salmonella cholerae-suis* it was decided to omit the *kunzen-dorf* type from the Kauffmann-White scheme on the ground that it was merely a group phase variant of the parent strain.

**Variation in the Vi antigen.**—There is a type of variation that concerns the Vi antigen (see p. 1825). The H and O antigens remain unaffected. Kauffmann (1930a) found that typhoid bacilli might exist in three forms: (1) the V form which contains the full quota of Vi antigen and is inagglutinable by O serum, (2) the VW form which contains some Vi antigen but not enough to inhibit O agglutination and (3) the W form which contains no Vi antigen and is fully agglutinable by O serum. A similar variation in the Vi antigen may be observed in *Salmonella paratyphi C* and *Salmonella bollingeri*. The V → W variation appears to be irreversible (Craig and Branlon 1936b).

**Variations in the O antigens.**—In some strains containing two or more O components some colonies may be found in which one of the components is missing or developed to only a slight extent. The two antigenic components most likely to be affected are I and VII. Within the same strain there are I + +, I + and I ± colonies, the tendency being for the I + + to pass over to the I ± type. The VII antigen is complex, containing VII<sub>1</sub>, VII<sub>2</sub> and VII<sub>3</sub> fractions. According to Kauffmann (1941) it is the VII<sub>2</sub> fraction that is most likely to be lost or weakened. A rather different type of variation is noticed in organisms like *Salmonella paratyphi B* and *Salmonella typhimurium* containing the IV-V antigenic components. In some strains the V antigen is missing not merely from certain colonies but from the whole strain. Strains of *Salmonella paratyphi B* showing this so-called loss variation seem to be commoner in carriers than in acute cases and strains of *Salmonella typhimurium* showing the same loss variation are according to Edwards and Bruner (1940a) usually derived—at least in the United States—from pigeons.

**The OH → O variation.**—Flagellated strains may occasionally give rise to variants that contain only the O antigen. As a rule, this type of variation is irreversible. Some strains like *Salmonella gallinarum* and *Salmonella pullorum* are permanently non-flagellated. The OH → O variation cannot readily be induced in the laboratory. The production of O forms by growth on phenolized agar is not a true impressed variation since there is a rapid reversion to the normal flagellated form when the organisms are inoculated on to ordinary media.

**The smooth → rough variation.**—This type of variation has already been described at sufficient length in Chapter 9. So far as the *Salmonella* group is concerned, it consists essentially in a loss of the normal smooth somatic antigen with the appearance of a new, rough antigen having far less specific properties than the smooth antigen and conferring on the bacilli a sensitivity to salt solution which renders them unstable in suspension.

The flagellar antigens are unaffected. This type of variation seems to occur rather more frequently in nature, is common in strains maintained through many generations on ordinary laboratory media, and can readily be induced by several different procedures—the prolonged incubation of a broth culture, growth in the presence of antibodies acting on the smooth somatic antigens, subjection to the action of an anti-smooth phage, and so on. To prevent its occurrence strains should be dried from the frozen state *in vacuo*, or, if this is not practicable, kept on a dry Dorset egg medium in the ice-chest and subcultured as infrequently as possible. Moist solid media and sugar-containing media should be avoided.

It is of interest to note that the rough polysaccharide antigen appears to be limited to members of the *Salmonella* group. It is possessed by all those strains that have been examined but not by other species of bacteria, such as coliform or dysentery bacilli (White 1929a).

Many of the observations recorded by workers in this field indicate quite clearly that the S → R variation is not an "all-or none" process so far as the culture as a whole is concerned. It is gradual or step-like. A smooth strain may lose some of its normal somatic antigen and uncover some of its rough antigen, or develop it to replace the smooth so that it will respond to both an anti-smooth and an anti-rough serum. As has already been noted (see Chapters 8 and 9), variation by loss may proceed further than the R form, giving rise to the  $\rho$  forms of White (1932), in which the R polysaccharide is lost.

Variation of the mucoid or M antigen.—Mention has already been made (p. 705) of the formation by some members of the *Salmonella* group, notably *Salm. paratyphi* B, of mucoid colonies containing a nitrogen free polysaccharide (Burch Hirschfeld 1935). This so-called mucoid or M antigen (Kauffmann 1935a, 1936a) is formed best at room temperature, is resistant to heating at 60° C for 1 hour and to formalin, but is destroyed partly or completely by heating at 100° C for 2 hours, and by treatment with 96 per cent alcohol or normal HCl at 37° C for 20 hours. The presence of the antigen is best demonstrated by the use of living suspensions and an anti M serum. The mixtures should be incubated for 2 hours at 37° C and overnight at room temperature. Sera usually have a low titre seldom exceeding 1/160. Only one M antigen has yet been met with.

Variation of the X antigen.—The X antigen was first observed by Topley and Avrton (1924) in suspensions of *Salm. typhi-murum*, *Salm. newport*, and *Salm. enteritidis* which had been grown in broth for some days at 37° C, but has since been found to develop in almost all the salmonellae studied. Like the M antigen it is non-specific. It is thermostable resisting heat at 100° C for 30 minutes, and is unaffected by 0.25 per cent formalin. It occurs in both rough and smooth strains. Its appearance is most common in old broth cultures though it may be present in broth cultures grown for the normal length of time or even in suspensions from agar, especially if the surface is moist. Agglutination resembles the granular somatic type, but the flakes are as a rule slightly coarser and tend to lie along the sides of the agglutination tube. They become apparent in 2 to 4 hours, but continue to increase for some time. High titre serum is easy to prepare by the inoculation of rabbits with a culture rich in X agglutino-gen. The importance of the X antigen lies in the confusion it may cause in routine serum agglutination tests should any of the suspensions contain this antigen or any of the sera contain anti X agglutinins. In practice, few workers seem to have met with the X antigen, but in one or two laboratories its presence has caused considerable trouble. It is always wise to make certain that standard agglutinable suspensions are free from it (see Cruickshank 1939).

Fermentative Varieties of Antigenic Strains.—Another point that perhaps needs some comment is the recognition of fermentative varieties that are antigenically identical. As a systematic procedure this is a reversal of the common finding, in which different antigenic types can be recognized in a species that is homogeneous as regards its enzymic activities. Many of the fermentative differences that have

been accepted as defining separable varieties are very slight. Sometimes they are revealed only by the use of a special medium or a special reaction. The definition and labelling of these fermentative types has been carried out by a few observers, particularly by Kauffmann who would extend the process even further than many other workers would be prepared to do (see Kauffmann and Buron 1935). The significance of these types must we think remain very doubtful until we have far more extensive data than are at present available. At the same time the existence of the differences in enzymic activity can clearly not be ignored, if only because the evidence that exists to-day scanty as it is suggests that they are correlated with natural habitat and natural pathogenicity.

To take as an example the fermentative varieties of *Salmonella enteritidis* Kauffmann (1935b) records the origin of small samples of these strains. Of 8 strains of *Salmonella enteritidis* examined by him 7 were isolated from man one from a guinea pig. Of 29 strains of *Salmonella enteritidis* var. *danskyi* 13 strains came from human infections, 9 from rats or from the Ratin virus used for the extermination of these vermin. The 8 strains of *Salmonella enteritidis* var. *chaco* all came from infected soldiers in the Chaco war. Of 7 strains of *Salmonella enteritidis* var. *essen* some came from human infections, some from infections in ducks. Again strains of *Salmonella paratyphi* B that ferment *D* tartrate were found by Kristensen and Kauffmann (1937) in patients suffering from enteritis, strains causing typical paratyphoid fever do not usually ferment *D* tartrate. Strains of *Salmonella typhimurium* that infect European ducks and chickens do not as a rule ferment rhamnose in opposition to those found in the United States which are rhamnose positive (Edwards and Bruner 1940a).

If the fermentative abilities described are found to be constant for any particular strain and if the varieties differentiated by them are found to show significant differences in their natural distribution there will clearly be a good case for providing them with separate labels. At the moment it is wisest to regard these labels as provisional.

**The Bacteriophage Method of typing Salmonella Strains**—Of far greater value though of more restricted application is the method of subdividing antigenic types into sub types by means of the bacteriophage method of typing introduced by Craigie and Brandon (1936a, b) and Craigie and Yen (1938). It was known that salmonellae were susceptible to the action of bacteriophages and that these were closely related to the type of O antigen present, but owing to the wide communal sharing of the same O antigen bacteriophages of this sort were useless for the differentiation of sub types. Soon after the discovery by Felix and Pitt (1931a, b) of the Vi antigen of the typhoid bacillus several workers established the existence of bacteriophages acting specifically on the V form of the bacillus. The special contribution of Craigie and his co-workers was their observation of a peculiar adaptability possessed by one particular anti Vi phage. When this phage was grown on typhoid strains of different origin, races of bacteriophage were obtained that were found to have developed a high degree of specificity for the particular strain on which they had been propagated. By means of a series of bacteriophages prepared in this way, Craigie and Yen were able to classify nearly all of 592 strains of typhoid bacilli into six bacteriophage types. The evidence obtained from a study of the origin of the strains revealed a high degree of correlation between the bacteriophage type and the epidemic source. These observations have since been abundantly confirmed so that the bacteriophage method of typing is rapidly becoming a routine procedure in the investigation of outbreaks of typhoid fever. The degree of specificity seems to be almost complete and the results are more constant and reliable than those of typing by the fermentative method.



As already mentioned, however the method is subject to limitations. In the first place not all strains of typhoid bacilli possess a Vi antigen or possess it in sufficient quantity to enable the bacteriophage to act satisfactorily. For example in Craigie and Yen's series of 700 strains, 42 belonged to the W form and 72 to the VW form. The remaining 586 strains belonged to the V form, possessing a fully developed Vi antigen—of these 9·6 per cent. were successfully typed. The proportion of W and VW forms was higher in this series, which contained many strains that had been isolated months or years previously, than is commonly found in routine laboratories handling freshly isolated strains, of which usually only about 5 per cent. are found to be lacking in Vi antigen (Craigie and Brandon 1936; Bauss 1939). Another limitation is that only species of *Salmonella* containing the Vi antigen can be typed. This is a very serious limitation since according to our present knowledge the distribution of this antigen is restricted to *Salm. typhi*, *Salm. paratyphi* C, *Salm. gallinarum* and *Salm. formicæ*.

More recent observations, however suggest that some other species possess either a Vi antigen or another type of antigen which is susceptible to differential attack by the bacteriophage. Felix and Callow (1943) for example have been able to develop a series of bacteriophages acting on *Salm. paratyphi* B and have by this means succeeded in dividing strains of this organism into four bacteriophage types, which correspond closely with the epidemiological types.

The technical methods used in the bacteriophage typing of typhoid and paratyphoid bacilli cannot be described here. Table 4—however, illustrates the susceptibility of typhoid bacilli to the different types of bacteriophage.

It will be noticed that Type A strains are sensitive to all bacteriophage type strains, but that the remaining types possess a fairly high degree of specificity. Further types and sub-types are constantly being described. Types L and M and sub-types B<sub>1</sub>, B<sub>2</sub>, D<sub>1</sub>, E<sub>1</sub>, and F<sub>1</sub> have since been added and Felix (1943) in this country has reported three new sub-types D<sub>2</sub>, D<sub>3</sub>, and L<sub>2</sub>, and a new type labelled provisionally No. 91. As more strains of typhoid bacilli are studied from different environmental conditions, it is probable that even further bacteriophage types will be discovered. Sufficient, however are already known to render this method of study profoundly valuable in the field of epidemiological inquiry. More recently Craigie (1949) has modified his original scheme but the principles of his method remain unaltered.

**Chemical Structure of the Salmonella antigens.**—We may add a brief note on our knowledge such as it is, of the chemical constitution of the antigens on which the present classification of the *Salmonella* group is based. Of the nature of the flagellar antigens we as yet know nothing beyond the hints conveyed by heat lability and sensitiveness to extraction with alcohol.

Of the somatic antigens we know rather more. Furth and Landsteiner (1925; 1929) isolated several different chemical components from bacilli of the *Salmonella* group. Some of these were polysaccharides. The studies of White (1929a, b; 1931) make it clear that the somatic antigens—those labelled I, II, III and so on in our antigenic formulae—are polysaccharides, or have a polysaccharide component and further evidence on similar lines has been recorded by subsequent observers (Carper 1929-30; Comblesco et al. 1930; Basilewsky and Péménil 1933). More recent studies by Freeman, Challinor and Wilson (1940) and Morgan and Partridge (1949) on *Salm. typhi* suggest that the O antigen of this organism is a polymolecular complex formed of a specific polysaccharide, a protein, and a

phospholipin. The protein component appears to be chemically and immunologically very similar to the conjugated protein prepared by Morgan and Partridge (1910) from the specific somatic antigen of *Sh shiga*. According to Freeman (1913) the somatic antigen of *Salm typhi murium* chemically resembles that of *Salm typhi*.

TABLE 43

REACTIONS OF TYPE STRAINS OF *Salm typhi* TO CRITICAL TEST DILUTIONS OF TYPE BACTERIOPHAGE STRAINS (FROM CRAIGIE AND YEN 1938)

Type of <i>Salm typhi</i> .	Type of Bacteriophage.										
	A	B	B <sub>1</sub>	C	D <sub>1</sub>	D <sub>2</sub>	E	F	G	H	J
A	CL	CL	CL	CL	CL	CL	CL	CL	CL	CL	CL
B <sub>1</sub>	—	CL	+++	±	—	—	±	+++	+	—	+++
B <sub>2</sub>	±	—	CL	—	+	—	—	+	—	—	—
C	—	—	±	CL	±	—	±	±	±	—	±
D <sub>1</sub>	—	—	—	±	CL	+++	—	—	—	—	—
D <sub>2</sub>	—	—	—	—	±	CL	—	—	—	—	—
E	—	—	—	—	—	—	CL	—	—	—	—
F	—	—	—	—	—	—	—	CL	—	—	—
G	—	—	—	—	—	—	—	—	CL	—	—
H	—	—	—	—	—	—	—	—	—	CL	—
J	—	—	—	—	—	—	—	—	—	—	CL

CL = Confluent Lysis    +++ = Numerous discrete plaques of subnormal size  
± = Few plaques subnormal in size except in Type C strains

**Pathogenicity and Toxin Production**—As has already been pointed out (p. 703) members of the *Salmonella* group are widely distributed in the animal kingdom. Broadly speaking the type of disease with which these organisms are associated is either an enteritis or a septicæmia. In man a few species like *Salm typhi* and *Salm paratyphi B* give rise to a disease characterized by a fairly long incubation period and the predominance of septicæmic over intestinal symptoms. The great majority however of the *Salmonella* species produce acute gastro enteritis of the food poisoning type in children and adults or acute enteritis in infants characterized by a short incubation period and the predominance of intestinal over septicæmic symptoms. Nevertheless a disease apparently starting as an enteritis may become a septicæmia which runs a typhoid like course or is associated with localized manifestations of disease in the meninges bones joints or other

situation. As in animals and birds a proportion of persons who become infected may harbour the organism for a varying length of time without showing any signs of disease. Though certain types of *Salmonella* are far commoner in man than others it is now clear that practically every species so far described is capable of infecting human beings. Even *Salm. gallinarum* which has been regarded as restricted to fowls has been isolated from a human case of disease. It is impossible here to describe in detail the natural pathogenicity of members of the *Salmonella* group for man and animals but some information in this respect will be found in the descriptions of the individual species on pp 726-44.

Experimental observations on the pathogenicity of *Salmonella* for laboratory animals have been confined to a relatively few species, such as *Salm. typhi*, *Salm. typhi-murium* and *Salm. enteritidis*. It will be sufficient in this chapter to give a brief description of the effects produced in experimental animals by some of the better known species, and a short discussion of the nature of the toxic substances that these organisms produce.

### *Salm. typhi*

If massive doses of living typhoid bacilli are administered by the mouth to chimpanzees, it is possible to produce a disease that is very similar to typhoid fever in man (Mitschukoff and Besredka 1911).

Administration of typhoid bacilli by the mouth to ordinary laboratory animals (rabbit, guinea pig, rat or mouse) does not give rise to an infection of this type, or usually to any harmful result at all.

The intraperitoneal or intravenous injection of living typhoid bacilli in adequate doses induces a fatal infection and the bacilli can be recovered from the blood and tissues post mortem. The effect of such injections would seem to be in part toxic, in part dependent on the multiplication of the bacteria in the body. With some strains of typhoid bacilli it may be necessary to inject 1 000 million living bacilli or more into the peritoneum of a mouse to produce a fatal result. A low multiple of this dose (5 000-10 000 million bacilli) will usually cause a purely toxic death when the bacilli have been killed by heat before inoculation. A highly virulent strain will induce a fatal infection following the intraperitoneal injection of 50 million bacilli, or even a little less. But there is no evidence that the typhoid bacillus possesses an ability to multiply freely in the blood or tissues of small laboratory rodents when injected in small doses, such as would certainly prove fatal in the case of a frankly invasive organism.

There is nothing characteristic in the findings at necropsy in a mouse or guinea-pig that has died as the result of an intraperitoneal injection of living typhoid bacilli. There is, of course, the usual inflammatory reaction in the peritoneum, sometimes associated with subserous hæmorrhages and the organism which has become generalized throughout the body may be recovered from the blood or from any of the tissues. Following the intravenous injection into the rabbit of doses of living typhoid bacilli too small to produce a rapidly fatal infection, the bacilli tend to localize themselves in certain situations, particularly in the gall bladder. But experiments of this type will be more conveniently considered in Chapter 69 in relation to the pathogenesis of enteric infections in man.

### *Salm. typhi murium*

The effects produced by the administration of living cultures of this organism to the small rodents of the laboratory are entirely different from those produced by the typhoid bacillus. We are dealing with a natural pathogen of these animals, which gives rise in them to a characteristic disease, usually known as mouse typhoid. This disease is produced when living cultures of *Salm. typhi murium* are given by the mouth as well as when they are administered by subcutaneous or intraperitoneal injection, though the time to death is longer in the former case than in the latter. The organism has a very definite

invasive power for the tissues of mice and of other laboratory rodents. A virulent strain will kill 50 per cent or more of mice injected intraperitoneally with a dose of 100 bacilli, as compared with the 50-100 million that are required in the case of a virulent typhoid bacillus. Mice dying within 2 to 3 days after the injection of a moderate dose of a virulent strain will be found to have succumbed to an acute septicæmia with few obvious lesions, but mice dying after the more usual period of 5 to 10 days often show characteristic lesions, including a varying degree of splenic enlargement often associated with the presence of small necrotic foci, larger and very characteristic necrotic lesions in the liver, and sometimes scattered pneumonic patches in the lungs associated with a scanty pleural exudate. These lesions have been described in some detail by many observers (see, for instance, Seiffert, Jahneke and Arnold 1923). The spread of infection from the intestine, and the subsequent involvement of the various tissues, have been studied and described by Muller (1912) and by Orskov and his colleagues (Orskov, Jensen and Kobayashi 1928, Orskov and Moltke 1928, Orskov and Lassen 1930). These experiments will be considered in some detail in Chapter 47.

One other point should be emphasized. The disease produced in the mouse by *Salm. typhi* shows no tendency to spread by contact from mouse to mouse. The disease produced by *Salm. typhi murium* is highly contagious, and there are few laboratories that have not experienced serious epidemics of this infection among their normal stock of mice or guinea pigs.

#### *Salm. enteritidis*

It need only be said that this species, particularly the *danzysz* variety, behaves in much the same way as *Salm. typhi murium*. Whether all varieties of *Salm. enteritidis* are equally pathogenic for small rodents, as judged by experimental infection in the laboratory, we do not know.

#### *Salm. paratyphi B*

The main interest of this species, from our present point of view, is that it occupies a position in some ways intermediate between that of *Salm. typhi* and *Salm. typhi murium*. It is a natural pathogen of man but not of rodents, but antigenically it is very closely related to the mouse typhoid bacillus. When injected into mice it kills them in far smaller doses than does *Salm. typhi*, though it is less virulent than *Salm. typhi murium*. When administered by the mouth, as Orskov and his colleagues have shown, it has a limited ability to invade the tissues and multiply in them, but it rarely gives rise to fatal infections.

It is probable that many other species of *Salmonella* will be found to possess high or moderate virulence for laboratory animals, and in some instances we already know this to be the case. *Salm. cholerae suis*, for instance, is highly virulent for the rabbit. But the mapping out of the relative virulence, or pathogenicity, of all the 130 or more species or varieties of *Salmonella* that have so far been described, and of the new arrivals that may well be imminent, will clearly be a laborious undertaking, and one that is perhaps not very likely to be embarked on in the near future.

**The Toxins of the Salmonella Group**—As we have noted above, the injection of killed typhoid bacilli, in adequate dosage, will produce toxic death in the usual laboratory animals, and the "toxins" of the typhoid bacillus were studied by several of the earlier bacteriological workers (Pfeiffer 1894, Sanarelli 1894, Chantemesse 1897, Brieger 1902).

Macfadyen and Rowland (1901, 1903), by grinding typhoid bacilli in the frozen state and subsequently extracting them with 0.1 per cent KOH, prepared a solution which was highly toxic for rabbits but only slightly toxic for guinea pigs. Meyer and Bergell (1907) obtained an extract of typhoid bacilli that was toxic for rabbits by suspending the bacilli from agar cultures in slightly alkaline distilled water, and filtering the extracts through Chamberland candles after they had stood for 48 hours at room temperature.

Conradi (1906) found that a filtrate from an autolysed saline suspension of *Salm typhi* after evaporation at 35° C. to one-tenth of its original volume was toxic for guinea pigs in a dose of 0.2 ml. Yamanouchi (1909) reported that filtrates of cultures of *Salm typhi* in 5 per cent peptone water were toxic for rabbits in a dose of 0.5-1 ml. per kilo body weight. Arima (1912) grew *Salm typhi* on weakly alkaline agar, suspended the growth in saline and later separated the bacillary bodies by centrifugation. The supernatant fluid was pipetted off and examined separately. The bacillary bodies were washed three times ground up, and extracted with saline. Both solutions were toxic for rabbits, less so for guinea pigs and mice. Douglas (1921) showed that tryptic digests of acetone-extracted typhoid bacilli were highly toxic. The statements with regard to the thermostability of these toxic extracts or filtrates are somewhat conflicting. But most workers report them as heat resistant. Taken as a whole these observations indicate that typhoid bacilli elaborate toxic substances which are, in the main, retained within the bacterial cell though they are liberated into the surrounding medium when autolysis occurs. There is, at the moment, no evidence that the typhoid bacillus produces an exotoxin in the ordinary sense of that term.

Most other salmonellae that have been examined behave, in this respect, in the same way as *Salm typhi* though the toxicity of killed cultures or extracts of the different species may show significant differences in toxicity for laboratory animals, particularly the rabbit (White 1926, 1929b).

The two natural pathogens of rodents, *Salm typhi-murium* and *Salm enteritidis*, may be considered in rather more detail from this point of view, since it happens that we have recently obtained some knowledge of the constitution of one, at least, of their toxic products.

Ecker (1917), Ecker and Richardson (1923), Branham (1923), Ecker and Rumington (1923) and Menten and King (1930) noted the toxicity for rabbits or mice of broth filtrates of *Salm typhi-murium* and of certain other salmonellae, and Casper (1928-29) prepared a toxic extract of the bacterial bodies by extraction with anti-formin. Ecker and Rumington (1923), Menten and King (1930) and Casper (1928-29) were able, by various methods of adsorption or precipitation, to obtain a partially purified toxic fraction from these crude filtrates or extracts. The fractions so obtained were thermostable. The purification was very incomplete, and though qualitative tests indicated the presence of polysaccharide constituents, no serious attempt was made to determine the chemical constitution of the toxic components. Martin (1934) showed that the greater part of the toxic material present in a broth culture of *Salm typhi-murium* was contained in the bacterial cells, and that these constituted the most favourable starting material for obtaining a purified toxin. His method of purification is considered below.

Independent studies on the immunological properties of chemical fractions isolated from *Salm typhi-murium* and *Salm enteritidis* by Borvin and his colleagues (1933, 1934, 1935) and by Raistrick and Topley (1934), Delafield (1934) and Martin (1934) have thrown further light on this problem. These studies were undertaken, in the main, with the object of obtaining a chemically pure immunizing substance from these organisms. The point that concerns us here is that fractions obtained by both groups of workers were found to be highly toxic.

Borvin and his colleagues obtained their fractions by extraction with trichloroacetic acid, followed by alcohol precipitation. Raistrick and Topley extracted the bacilli with acetone, digested them with trypsin and submitted the tryptic digest to fractional precipitation with alcohol.

The final product obtained by both groups of workers was free from protein, as judged by the ordinary chemical tests, but contained a polysaccharide component that gave rise to reducing sugars on hydrolysis. Nitrogen and phosphorus were also present, and the results of micro-combustion analysis, taken with various qualitative tests, suggested that the active fraction consisted of a complex polysaccharide combined with a phosphatide. The polysaccharide can be separated from the phosphatide residue by heating with weak acetic acid. When this is done the polysaccharide is entirely, and the phosphatide

almost non toxic. The unhydrolysed toxic material as prepared and tested by Martin (1934), had an average lethal dose for the mouse of 0.5 mgm. The substance obtained by Boivin and his colleagues is reported as being slightly more toxic.

A point of considerable interest in regard to these findings is that there seems little doubt that the toxic substance is itself the somatic antigen of the bacterial cell. Rabbits immunized with the purified fractions produce characteristic somatic agglutinins. The polysaccharides isolated from salmonella organisms by previous workers and identified with the somatic antigens or with their haptens constituents were in most cases found to be non toxic. But this was probably because the method of preparation had resulted in the splitting of the polysaccharide from the phosphatide component.

The same series of experiments have given an indication of one of the ways in which these toxic substances produce their effects. It had already been shown by several workers (see Chapter 44) that natural infections in man and experimental infections in animals might be associated with an increase in the concentration of sugar in the blood. and Menten and King (1930) had found that rabbits injected with the fraction isolated by them from *Salm typhi-murium* developed hyperglycaemia.

DeLafield (1934) who had already made a careful study of the chemical changes occurring in the blood of the rabbit following the injection of killed suspensions of various bacteria (see Chapter 44) was able to show that the purified fractions prepared by Raistrick and Topley induced a marked hyperglycaemia followed in many cases by a fall in blood sugar far below the normal level. These findings were confirmed by Boivin and Mesrobian (1934b).

These observations suggest that the toxic substances isolated from *Salm typhi-murium* and *Salm enteritidis* are representatives of a group of antigenic components possessing very similar toxic properties but differing widely in the chemical structure that determines their antigenic specificity and that bacterial components of this type are very widely distributed among different bacterial groups. DeLafield (1931, 1932) found that a wide variety of Gram negative bacteria including *Bact coli*, *Bact aerogenes*, *Sh shiga*, *Salm typhi*, *Salm typhi-murium*, *H influenza*, *H bronchiseptica*, *Past muriseptica*, *Proteus vulgaris* and the meningococcus induced hyperglycaemia in rabbits while the Gram positive species studied did not, and Boivin and Mesrobian (1934b) have isolated fractions of the same type as those obtained from *Salm typhi-murium* from a variety of bacilli of the coli typhoid group though the toxicity of most of these fractions has not yet been determined.

There is of course no reason to suppose that these particular substances are the only toxic constituents of the bacteria in which they occur. It is possible that there are many others. They do however, afford examples of endotoxins that are definable chemical substances and not merely crude extracts containing all the multitudinous components of the bacterial cells from which they were derived.

We append a detailed description of the general characters of a typical member of the *Salmonella* group—*Salm paratyphi B*—and a summarized description of each of the different species.

### *Salm paratyphi B*

*Synonyms*—*Bact paratyphosum B*, *Bact schottmulleri*.

*Isolation*—From cases of enteric infection in man by Achard and Bensaude (1896) and Schottmuller (1900, 1901).

*Habitat*—Almost entirely a parasite of the human intestinal tract.

*Morphology*—Bacilli with parallel sides and rounded ends usually 2-3  $\mu$  long and 0.6  $\mu$  broad. Gram negative, non acid fast. Non sporing, usually non capsulated but may form a mucoid envelope. Actively motile, flagella are stated to be peritrichate but of this there is some doubt.

- Agar Plate*—24 hours at 37° C. Commonest type of colony is circular low convex greyish yellow faintly translucent, 1-3 mm. in diameter with smooth surface and entire edge butinous in consistency and emulsifies easily. Surface may be irregular though smooth, and edge may be coarsely undulate or finely but irregularly indented. Colonies that are left at room temperature, after preliminary incubation at 37° C., may develop a so-called mucoid wall.
- Agar Slope*—24 hours at 37° C. Abundant raised, faintly translucent, greyish yellow growth with glistening smooth or beaten-copper surface and entire or undulate edge.
- Gelatin Slab*—7 days at 20° C. Good filiform growth, with moderate growth on the surface. No liquefaction.
- Broth*—24 hours at 37° C. Abundant growth with uniform turbidity which continues to increase slightly for 2-3 days. Slight to moderate powdery deposit demonstrating readily on shaking. No surface growth.
- MacConkey's Agar*—24 hours at 37° C. Yellow colonies, 1-3 mm. in diameter often of vine-leaf type.
- Desferrioxal Citrate Agar*—24 hours at 37° C. Colourless colonies, 1-2 mm. in diameter sometimes with a black centre.
- Wilson and Blair's Agar*—48 hours at 37° C. If closely packed, colonies are about 1 mm. in diameter and appear faintly greenish. If discrete with room to spread, they are larger flatter and darkish in colour with a metallic sheen.
- Resistance* Killed by moist heat at 55° C. in half an hour. Cultures at room temperature or preferably on egg medium in the ice-chest, live for months.
- Metabolism* Aerobic and facultative anaerobic. Grows between about 15° and 42° C., best at 37° C. May form a mucoid, capsular material at room temperature. Grows well in Bacterium medium containing ammonium salts as the main source of nitrogen. No hemolysin formed.
- Biochemical*—Acid and gas in glucose, maltose, mannitol, sorbitol, dulcitol, arabinose, trehalose and xylose, but not in lactose, sucrose, salicin or adonitol. Reactions variable in rhamnose and inositol, in *d*-l and *s*-tartrates, and in citrate, mucic acid generally fermented in one day. Variable reaction in Stern's medium. Occasional variants fail to form gas. Fermentative sub-division based mainly on failure to ferment *d*-tartrate or rhamnose. H<sub>2</sub>S— Indole negative. M.R.— V.P.— Nitrates reduced. NH<sub>3</sub>— M.B. reduced. Catalase— Koser's citrate— Gas formation in MacConkey broth at 44° C. negative.
- Antigenic Structure*—Diphasic flagellated bacillus. Antigenic formula [II], IV, [V], XII.  $b \leftrightarrow 1-2$ . Strains lacking the V somatic antigen—the *odona* variety—are said to be commoner in carriers than in acute cases.
- Pathogenesis*—Gives rise as a rule to paratyphoid fever in human beings, but may be responsible for acute gastro-enteritis, followed or not after some days by the usual febrile disease. Strains from cases of gastro-enteritis often ferment *d*-tartrate which strains from cases of paratyphoid fever seldom do. Has occasionally been isolated from domestic animals, but is not known to cause disease in them under natural conditions. Inoculated intraperitoneally into mice in a dose of about 500 million bacilli, it produces death in 1 to 3 days. Post-mortem there is some peritonitis and enlargement of the spleen. The bacilli can be recovered in culture from the spleen and heart blood.

## GROUP A.

*Salmonella paratyphi* A 4.F [II], II XII a—

Isolated from enteric fever in man (Gwyn 1893, Schottmüller 1903, 1911 Brown and Kayser 1902). Causes enteric fever but not acute gastro-enteritis in man. Has once been isolated from a pig (Broudin 1907), but is not a natural pathogen of animals. Growth is poorer than that of most other salmonellae. Different strains vary in their H<sub>2</sub>S pro-

duction. Occasional strains fail to form gas. By growth in immune serum an artificial second phase containing the 1, 5 antigens, and an artificial third phase containing antigen  $z_{11}$ , have been demonstrated by Bruner and Edwards (1941a).

## GROUP B

*Salm paratyphi B* A F [I], IV, [V], XII       $b \longleftrightarrow 1, 2$

Isolated from cases of enteric fever in man (Achard and Bensaude 1896, Schottmüller 1900, 1901). The I antigen is present only in some strains. The V antigen is generally present, but may be missing, particularly in strains from carriers (Kauffmann 1934c), such strains have been named *Salm paratyphi B* var *odense*. Strains in which the non-specific phase is missing are sometimes referred to as *Salm paratyphi B* var *java*. Causes enteric fever in man, and sometimes an acute, but mild enteritis. The strains causing enteric fever form a mucoid wall and do not ferment *d* tartrate, the strains causing simple enteritis do not form a mucoid wall but do ferment *d* tartrate. The *jawa* variety is distinguished by its inability to ferment *s* tartrate, it appears to be unusually common in Panama (Edwards and Bruner 1943). By means of suitable bacteriophages several sub types have been distinguished by Felix and Callow (1943). Kristensen and Bojlén (1929) have described a number of fermentative sub types based on the use of rhamnose and inositol. May rarely fail to form gas. *Salm paratyphi B* is mainly a human pathogen but its isolation has occasionally been recorded from the mesenteric nodes of normal pigs (Hormaeche and Salsamendi 1936, 1939) from animals slaughtered for human consumption (Bartel 1938), and from chickens (Edwards and Bruner 1943).

An organism, described by Gail (1938) as a new *Salmonella* type under the name of *Salm abortus-canis*, with the antigenic formula IV, V, XII       $b \longleftrightarrow z_1, z_6$  is regarded by Kauffmann (1941) not as a true type but as an artificial variant of *Salm paratyphi B*.

*Salm abony* A F [I], IV, V, XII       $b \longleftrightarrow e, n, x$

Isolated from the faeces of normal persons by Rauss in Budapest, and described by Kauffmann (1940a). Is distinguished from *Salm abortus-bovis* by absence of the XXVII antigen by its fermentation of inositol and of *s* tartrate and by its failure to liquefy gelatin. *Salm schlesheim*, with which it may be confused, contains the XXVII somatic and the  $z_{12}$  flagellar antigen fails to ferment dulcitol inositol or *s* tartrate and liquefies gelatin. *Salm. abony* can be distinguished from the *d* tartrate positive strains of *Salm paratyphi B* only by the differences in its Phase 2 flagellar antigens. Its pathogenicity for man is doubtful.

*Salm typhi-murium* A F [I], IV, [V], XII       $i \longleftrightarrow 1, 2, 3$

A natural pathogen of rodents, particularly mice, in which it causes a typhoid like disease (Loeffler 1892). This organism is identical with *Bact aertrycke* which was originally isolated from a case of acute gastro-enteritis (food poisoning) in man (de Nobelet 1898). It is under the name of *Bact aertrycke* that it appears in almost all the recent and current literature, so that this synonym is an important one to remember. It is also identical with *B pestis canis* described by Wherry (1908) as the cause of an epidemic disease in guinea pigs, and with the bacillus that Nocard (1893) isolated from a parrot suffering from psittacosis and named *Bact psittacosis* under the mistaken impression that it was the cause of that disease. The same organism is frequently referred to in German literature as the "Breslau bacillus". Strains in which the V antigen is missing are sometimes referred to as the *copenhagen* (Kauffmann 1934c) or the *storr*s (Edwards 1935) variety. This type is common in American pigeons. Those in which the non specific phase alone can be demonstrated are sometimes referred to as the *binns* variety (Schutze 1920), it should be noted however, that by growth of the *binns* variety in the presence of group serum Bruner and Edwards (1939) were able to demonstrate the existence of a specific phase in all strains. It shares its H antigens with *Salm aberdeen*. Numerous fermentative sub types have been described. In Europe strains from ducks and chickens usually fail to ferment rhamnose, inositol, *d* tartrate, or citrate, but non rhamnose fermenting strains



are uncommon in American ducks (Edwards and Bruner 1940a) Occasional strains fail to form gas

*Salm typhi* murium is pathogenic for man as well as for animals and is the most frequent cause of outbreaks of salmonella food poisoning It commonly gives rise in man to an acute gastro-enteritis, not infrequently fatal, but it occasionally causes a prolonged fever of the enteric type Although it has most often been recorded as causing epidemic disease in mice and rats (see, for instance, Meyer and Matsumura 1927), it is naturally pathogenic for many other animal species In addition to causing infections in guinea-pigs and parrots, it has been isolated from epidemics in sheep (Bruns and Gasters 1920, White 1929b, Lovell 1932b), chicks (Doyle 1927, Edwards 1929, 1939), pigeons (Beaudette 1926, Lesbouvries and Verge 1932, Cernaiani and Popovici 1933), turkeys (Rettger *et al* 1933), canaries (Beaudette 1926), and silver foxes (Benedict *et al* 1941) In Germany, it is by no means uncommon in cattle, in which it may cause severe enteritis (Lütje 1937, Bartel 1938) It also causes infections in ducks, and has been isolated from ducks' eggs (Scott 1932, 1933, Dalling and Warrack 1932, Lovell 1932b), and from American dried egg It has been demonstrated in the mesenteric glands of normal pigs (Hormaeche and Salsamendi 1936, 1939, Scott 1940, Varela and Zozaya 1942, Rubin *et al* 1942), and has sometimes been isolated from pigs that have died of swine fever (see Lovell 1932b). It may be responsible for enteritis in cats (van Dorssen 1937).

*Salm stanley* A.F. IV, V, XII  $d \longleftrightarrow 1, 2 \dots$

Isolated from cases of food poisoning and examined by Schütze (1920), Savage and White (1925) White (1926), and Kauffmann (1931) (See also Boecker and Silberstein 1932, Kauffmann 1930a, 1934a) The partial antigens present in the specific phase are d and  $d_2$  of which the d is shared with *Salm amersfoort*, *Salm muenchen*, *Salm typhi* and *Salm gaminara* It is not known to be a natural pathogen of animals, but it has been isolated from imported American spray-dried egg in Great Britain

*Salm. heidelberg* A.F. IV, V, XII  $r \longleftrightarrow 1, 2, 3 \dots$

Isolated from a case of food poisoning by Habs (1933) (See also Kauffmann 1931a, Kauffmann and Silberstein 1934) It has since been found in the mesenteric lymph nodes of normal pigs in Mexico (Varela and Zozaya 1942)

*Salm chester* A.F. IV, [V] XII  $e, h \longleftrightarrow e, n, x \dots$

Isolated by Grace of Chester from the faeces of patients suffering from gastro-enteritis in a mental institution, and described by Kauffmann and Tesdal (1937-38) Has since been found in other European countries (Kauffmann 1941) Has been isolated from cases of infantile diarrhoea in Uruguay (Hormaeche, Peluffo and Aleppo 1940), from cases of gastro-enteritis in the United States (Bornstein, Saphra and Strauss 1941) and in Panama (Edwards and Bruner 1943), and from the mesenteric nodes of healthy pigs in Uruguay (Hormaeche and Salsamendi 1939) It is indistinguishable from *Salm san-diego* except in the  $\beta$  phase, in which *chester* is said to have the formula  $e, n, x, z_{11}, z_{17}, z_{19} \dots$  and *san-diego*  $e, n, z_{11}, z_{17}, z_{19} \dots$

*Salm san-diego* A.F. IV, [V] XII  $e, h \longleftrightarrow e, n, z_{11} \dots$

Isolated originally from an outbreak of food poisoning Described by Kauffmann (1940d) Also found in a case of enteritis in the United States, and in New York sewage (Bornstein and Saphra 1942) in a healthy pig in Uruguay (see Kauffmann 1941), and in fowls in the United States (Edwards and Bruner 1943). For differentiation from *Salm chester*, see above

*Salm sahnatis* A.F. IV, XII  $d, e, h \longleftrightarrow d, e, n, z_{11} \dots$

Isolated from rat faeces near Salinas, California. Described by Edwards and Bruner (1942a) Is antigenically complex, having d and e in both phases. By cultivation in the presence of agglutinating serum to *Salm. typhi* the d antigen is lost from each phase, and the organism becomes biochemically and antigenically indistinguishable from *Salm san-diego*

*Salm saint-paul* A.F. I, IV, V, XII e, h  $\longleftrightarrow$  1, 2, 3

Isolated from the liver of a turkey poult in the United States Described by Edwards and Bruner (1940b) Also isolated from faeces and urine of a patient with an intermittent fever in New York City (Bornstein and Saphra 1942)

*Salm zagreb* A.F. IV, V, XII e, h  $\longleftrightarrow$  1, 2

Isolated at Zagreb Described by Kauffmann (1941)

*Salm kaposvar* A.F. IV, V, XII e, (h)  $\longleftrightarrow$  1, 5

Isolated from faeces of 3 members of a family who were suffering from gastro-enteritis and described by Rauss (1941) in Hungary Phase 1 is similar to that of *Salm onderstepoort* in that only part of the H antigen is present

*Salm koeln* A.F. IV, V, XII  $\gamma \longleftrightarrow$  1, 2, 3

Not to be confused with the *koeln* variety of *Salm dublin*

*Salm reading* A.F. IV, XII e, h  $\longleftrightarrow$  1, 5

Isolated from the Reading water supply (Schutze 1920) Has since been isolated from faeces of cases or carriers in relation to outbreaks of gastro enteritis (Kauffmann 1930a, 1931, 1934a), Boecker and Silberstein 1932), from the mesenteric nodes of healthy pigs (Scott 1940) and from an epidemic among laboratory guinea pigs (see Lovell 1932b)

*Salm derby* A.F. [I] IV, XII f, g —

Isolated from cases of food poisoning at Derby by Peckham (1923) (See also Savaga and White 1925, White 1926, Kauffmann 1931, 1934a) Kauffmann (1937a) pointed out that in some strains the I antigen was absent Has since been isolated from cases of infantile diarrhoea (Hormaeche, Peluffo and Aleppo 1936, 1940) from the mesenteric nodes of healthy pigs (Hormaeche and Salsamendi 1936, 1939, Edwards, Bruner and Rubin 1940, Rubin *et al.* 1942, Varela and Zozaya 1942) from turkeys (Edwards 1939), and from imported American spray dried egg in Great Britain Occasional strains fail to form gas The flagellar antigens comprise also  $z_4$

*Salm kaapstad* A.F. IV, XII e, h  $\longleftrightarrow$  1, 7

*Salm essen* A.F. IV, XII g, m —

Isolated from the stools of an infant who had been suffering for 3 days from enteritis (Hohn and Herrmann 1936a) Not to be confused with the *essen* fermentative sub type of *Salm enteritidis* (see p. 736)

*Salm budapest* A.F. I, IV, XII g, t —

Isolated in Hungary from the stools of 3 persons who had been suffering for a week from enteritis and from 3 healthy carriers (Rauss 1939a) The presence of the I antigen was pointed out by Kauffmann (1940b) According to Rauss (1939a) the flagellar antigen consists of g, t,  $z_4$  and  $z_5$ , this may be confused with that of *Salm senftenberg* which consists of g, s, t,  $z_4$  and  $z_5$  though, of course, the O antigens of the two organisms are quite different The t antigen of *budapest* is said to be different from that of *oramenburg* (Edwards, Bruner and Hinshaw 1940) Non pathogenic to mice by the mouth

*Salm californica* A.F. IV, XII g, m, t —

Isolated from young turkeys suffering from paratyphoid fever Described by Edwards, Bruner and Hinshaw (1940) Later isolated from fowls (Mallmann *et al.* 1942) and from American spray dried egg in Great Britain Both the g and the t antigens are complex Non pathogenic to monkeys by the mouth

*Salm brandenburg* A.F. IV, XII I  $\gamma \longleftrightarrow$  e, n,  $z_{15}$

Isolated from a case of acute gastro enteritis coming on after a meal of raw ham (Kauffmann and Mitsui 1930) Since isolated from the faeces of other cases of gastro enteritis and from the blood of patients suffering from a pyrexial disease (Černozubov *et al.* 1936-37) Not known to be a natural pathogen of animals

*Salm bispebjerg* A.F. I, IV, XII a  $\longleftrightarrow$  e, n, x

Isolated from the stools of a 71 year-old man suffering from acute gastro enteritis and bronchopneumonia (Kauffmann 1936b)

*Salm. abortus-egui* A.F. IV, XII . . .  $c \rightarrow e, n, x$  . . .

Isolated by Kilborne (1933) from a case of abortion in a mare. Has since been isolated or studied by Smith (1933), de Jong (1933), Kewer and Boerner (1933), Gold and Gruber (1933), van Heekeren (1934), Munster (1934), and Fisch and Billings (1934). For antigenic structure see White (1937) and Kaufmann (1931, 1934a). Phase 2 contains antigens  $x_1$  and  $x_2$  in addition to  $e, n, x$ . Edwards and Boerner (1933) have been able to grow *Salm. abortus-egui* in the presence of suitable immune sera, to induce two artificial phase variations: one, referred to as Phase 1, containing the antigen of *Salm. paratyphi A*, the other, referred to as Phase 3, containing an antigen  $x_3$  which was similar to, though not identical with, a similar antigen in the antigenically mixed phase of *Salm. schleslesheim*. It should be pointed out, however, that neither Phase 1 nor 3 has been met with under natural conditions. Occasional strains fail to form gas. *Salm. abortus-egui* is a natural pathogen of the horse, causing abortion in mares. It is not known to have caused infection in man.

*Salm. abortus-egui* A.F. IV, XII . . .  $c \rightarrow 1, 6$  . . .

Isolated by Schinner and Elisch (1921) from cases of abortion in sheep. For antigenic structure see Lovell (1931) and Kaufmann (1931, 1934a). Compared with most other organisms of the *Salmonella* group, this organism grows poorly. Many of the usual serum reactions are negative, late, or irregular. Not known to infect man or any animal other than the sheep.

*Salm. arechevalis* A.F. IV [V], XII . . .  $c \rightarrow 1, 7$  . . .

Described by Huemach and Peluffo (see Kaufmann 1931). Has been found in the Panama Canal zone in patients suffering from gastro-enteritis. Found once more in a case of gastro-enteritis (Edwards and Boerner 1933).

*Salm. attendi* A.F. IV, XII . . .  $c \rightarrow 1, 7$  . . .

Isolated by Hohn (1941) from the faeces of a woman suffering from severe fibrile diarrhea.

able to give rise to acute gastro enteritis in man, since about 40 persons were taken ill after eating a meat dish containing *Salmonella schleissheim*, and the organism was isolated from their faeces (see Kauffmann 1941)

### GROUP C

*Salmonella paratyphi* C A.F. VI<sub>1</sub>, VI<sub>2</sub>, VII {VI<sub>1</sub>}, c ↔ 1, 5

Isolated from cases of enteric fever in man mainly in Eastern Europe, more recently in British Guiana. It has received many other names and, in particular, is often referred to as "Hirschfeld's Bacillus," or the 'Eastern European type of *Bacterium paratyphosum* C' (see Weil 1917, Neukirch 1918, MacAdam 1919, Mackie and Bowen 1919, Hirschfeld 1919, Schultze 1920, Dudgeon and Urquhart 1920, Andrews and Neave 1921, Weigmann 1925a, b, Iwaschenzoff 1926, White 1926, Kauffmann 1931, 1934a, Giglioli 1930). Kauffmann (1935a) demonstrated the presence of a Vi antigen in some strains. Its apparent identity with the Vi antigen of *Salmonella typhi* was shown by Kauffmann (1936a) and Rouché (1938). The two antigens can, however, be distinguished by the bacteriophage technique (Scholtens 1937). The Vi antigen seems to play no part in determining the high pathogenicity of *Salmonella paratyphi* C for the mouse. This organism is an important pathogen of man, giving rise to enteric fever that is often associated with septic lesions. It is not known to be a natural pathogen of animals.

*Salmonella cholerae-suis* A.F. VI<sub>1</sub>, VII c ↔ 1, 5 or  
VI<sub>2</sub>, VII c ↔ 1, 5

The American hog cholera bacillus isolated by Salmon and Smith (1885-1886) from the former of whom the name *Salmonella* is derived. Though hog cholera is now known to be a virus disease (see p. 1963), *Salmonella cholerae-suis* is a common secondary invader in this disease. It is probably the commonest salmonella found in pigs. Either in the diphasic form or in the non-specific phase (*kunzendorf* variety) it has been isolated from the mesenteric lymph nodes of apparently normal pigs in Great Britain by Scott (1940) in South America by Hormaeche and Salsamendi (1939), in Mexico by Varela and Zozaya (1942), and in the United States by Rubin, Scherago and Weaver (1942) and Edwards and Bruner (1943). It is also commonly found in silver foxes (Benedict *et al.* 1941, Edwards and Bruner 1943) and has been isolated from cattle (Lutje 1939). Occasionally present in imported American dried egg. In the past it has been usual to refer to the diphasic H<sub>2</sub>S-negative variety as the American type, and to the H<sub>2</sub>S-positive variety, which exists in the group phase only, as the European or *kunzendorf* type. More extensive observations, however, have rendered it doubtful whether this distinction should be maintained. It is clear that their geographical relationship is subject to numerous exceptions, so that the names European and American had better be dropped. Whether the two organisms should be awarded separate varietal names is less clear. Kauffmann (1941) is in favour of abolishing the term *kunzendorf* altogether on the ground that numerous workers have been able to demonstrate Phase 1 in *kunzendorf* strains by growing them in immune serum. On the other hand, the two organisms usually differ in their H<sub>2</sub>S production, and—what is probably more important—their natural pathogenicity. As Edwards and Bruner (1943) have shown, *Salmonella cholerae-suis* is far more invasive than the *kunzendorf* variety; it can often be isolated from the heart blood of infected animals, whereas the *kunzendorf* variety remains confined to the intestine or mesenteric lymph nodes. Both organisms are closely related to *Salmonella paratyphi* C; they are distinguished from it by their failure to ferment arabinose and trehalose and to some extent by their lack of the Vi antigen. Of their pathogenicity to human beings there is no doubt. Numerous outbreaks of food poisoning have been ascribed to them (Clauberg 1971, Boecker and Silberstein 1932, Kauffmann 1934a, 1941). A typhoid-like fever may result from infection sometimes complicated by pneumonia, arthritis, purulent meningitis, or bacterial endocarditis (Boycott and McNee 1936, Harvey 1937, Goulder *et al.* 1942, Schwabacher, Taylor and White 1943).

*Salm. typhi-suis* A.F. VI, VI<sub>r</sub>, VII     c ↔ 1, 5

This organism was isolated from young pigs suffering from a typhoid like disease by Glässer (1909 1910). The monophasic type, which exists in the group phase only and is sometimes known as the *colldages* variety, was isolated from a similar disease in pigs by Dammann and Stedefeder (1910). These organisms represent the "Ferkeltypus" bacillus of German literature (see also Neukirch 1918, Andrews and Neave 1921 White 1926, Kauffmann 1931, 1934a, Bartel 1935). *Salm. typhi-suis* is almost antizoonally identical with *Salm. cholera-suis*. It differs, however, in growing poorly on ordinary media and in its fermentation reactions. It forms gas very slowly and sparsely, it usually fails to ferment mannitol or does so late, it does not attack citrate, mucate, or the tartrates, and it produces a slight permanent acidity in litmus milk. It also ferments trehalose, which *Salm. cholera-suis* does not. The specific phase can be demonstrated in the *colldages* variety by growth in presence of immune serum. So far as is known, *Salm. typhi-suis* does not naturally infect animals other than the pig nor does it give rise to disease in man. It seems to be very uncommon in the United States (Bruner and Edwards 1940).

*Salm. thompson* A.F. VI, VI<sub>r</sub>, VII     k ↔ 1, 5

Isolated from cases of food poisoning in man by Scott (1926), and since by numerous other workers. The *berlin* variety, which exists in the group phase, was isolated from cases of food poisoning by Kauffmann (1929a b, see also Kauffmann 1931, 1934a Boecker and Kauffmann 1930 Kauffmann and Mitsu 1930 Claiberg 1931, Sehmman and Claiberg 1932, Boecker and Silberstein 1932). *Salm. thompson* is very common in Great Britain, where it is about the third most frequent *Salmonella* type met with in outbreaks of food poisoning. It was demonstrated in the mesenteric lymph nodes of normal pigs by Scott (1940), and was found once in a rat by Khalil (1935) at Liverpool. Knorr (1931) isolated it from Chinese egg yolk. In the United States it appears to be uncommon (Edwards and Bruner 1943). Has been isolated from imported American dried egg

*Salm. enteritidis* A.F. VI, VII     k ↔ 1, 10

Isolated in South Wales from a case of acute gastro-enteritis, and recognized as a new type by Taylor at Oxford and Edwards in Kentucky (Taylor, Edward, and Edwards 1945).

*Salm. virchow* A.F. VI, VI<sub>r</sub>, VII     r ↔ 1, 2, 3

Isolated from a case of acute gastro-enteritis in Germany. Antigenic structure determined by Kauffmann (1930b 1931, 1934a). May give rise to a prolonged febrile disease.

*Salm. infantis* A.F. VI, VI<sub>r</sub>, VII     r ↔ 1, 5

Isolated from the blood of an infant in New Haven Hospital suffering from diarrhoea and mild fever. Described by Wheeler and Borman (1943). Biochemically indistinguishable from *Salm. virchow*.

*Salm. oranienburg* A.F. VI, VI<sub>r</sub>, VII     m 1 —

Isolated from faeces of normal persons and of patients suffering from gastro-enteritis (see Kauffmann, 1930b, 1931, 1934a, Claiberg 1932, Sehmman and Claiberg 1932, Boecker and Silberstein 1932). Its first demonstration in animals was by Edwards (1936), who found it responsible for a disease of baby quail in the United States. It has been found in chickens in the United States (Edwards 1939), judging from the frequency with which it has been isolated from American dried egg imported into Great Britain, it must be a common parasite of fowls. In human beings it gives rise to infantile diarrhoea (Hormaeche Peluffo and Alepyo 1940), and to gastro-enteritis in adults. Since the introduction of American dried egg in 1941, numerous sporadic cases of infection with this organism and at least one outbreak have been reported in England.

*Salm. potsdam* A.F. VI, VI<sub>r</sub>, VII     l r ↔ e, n z<sub>11</sub>

Isolated from cases of food poisoning. Examined by Kauffmann and Mitsu (1930) see also Kauffmann 1931 1934a Claiberg 1931 Sehmman and Claiberg 1932. Contains the z<sub>11</sub> and z<sub>12</sub> antigens in Phase 2 in addition to z<sub>11</sub>. Has been found in imported American spray-dried egg.

*Salm bareilly* A F VI<sub>1</sub>, VI<sub>2</sub>, VII . . y ↔ 1, 5 .

Isolated in India from mild enteric infections (Bridges and Scott 1931) Since found in infantile diarrhoea in the United States (Bornstein, Saphra and Strauss 1941), and in typhoid like cases in Hungary (Rauss 1941) and in Canada (Wyllie 1943) Demonstrated in chickens and turkeys in the United States (Edwards 1939), and in the mesenteric lymph nodes of normal pigs (Edwards, Bruner and Rubin 1940, Rubin *et al* 1942) Has been isolated from imported American dried egg, and from cases of food poisoning in Great Britain

*Salm hartford* A F VI<sub>1</sub>, VI<sub>2</sub>, VII y ↔ e, n, x

Isolated from the faeces of a 71 year-old man who suffered from abdominal cramps, followed by diarrhoea for a week (Edwards and Bruner 1941a) Phase 2 contains z<sub>11</sub> and z<sub>12</sub> in addition to e, n, x

*Salm mikawasima* A F VI<sub>1</sub>, VI<sub>2</sub>, VII y ↔ e, n, z<sub>11</sub>

Isolated from rat's faeces in Japan Described by Hatta (1938) and studied by Hormaeche and by Kauffmann (see Kauffmann 1941) Is closely related to *Salm hartford*, from which it differs in the possession of the z<sub>11</sub> instead of the x antigen in Phase 2, and in the fermentation of inositol Phase 2 also contains z<sub>12</sub> Non pathogenic to mice by the mouth

*Salm montevidео* A F VI<sub>1</sub>, VI<sub>2</sub>, VII g, m, s — or  
VI<sub>1</sub>, VII g, m, s —

Isolated by Hormaeche and Peluffo (1936) in Uruguay from the faeces of a monkey suffering from chronic enterocolitis, from an infant with chronic enterocolitis, from a pulmonary abscess in a child with enteritis, and from the mesenteric lymph nodes of normal pigs Since found in Uruguay in cases of infantile diarrhoea (Hormaeche, Peluffo and Aleppo 1940) and in the mesenteric lymph nodes of pigs (Hormaeche and Salsamendi 1936, 1939), in the United States in two gall bladder carriers (Edwards and Bruner 1943) in a patient with an attack of mild febrile diarrhoea (Schiff and Saphra 1940) in chickens (Jungherr and Clancy 1939) and in turkeys (Edwards 1939), in cases and outbreaks of food poisoning and in imported American spray-dried egg in Great Britain, in Denmark and in Palestine (see Kauffmann 1941) The Danish strain lacked the VI<sub>2</sub> antigen and differed in some cultural respects

*Salm oslo* A F VI<sub>1</sub>, VI<sub>2</sub>, VII a ↔ e, n, x

Isolated by Tesdal (1937) from faeces of patients with gastro enteritis in five different places in Norway Phase 2 also contains z<sub>12</sub> and z<sub>13</sub>

*Salm amersfoort* A F VI<sub>1</sub>, VI<sub>2</sub>, VII d ↔ e, n, x

Isolated from the heart blood of a 7-day old chick in the Transvaal, the chickens at the time were suffering from a severe infectious disease accompanied by a high case mortality Described by Henning (1937) Also isolated, together with *Salm typhimurium*, by Schiff and Strauss (1939b) in the United States from a patient suffering from a typhoid like fever The d antigen in Phase I consists of the partial antigens d<sub>1</sub> and d<sub>2</sub> Phase 2 also contains z<sub>12</sub> and z<sub>13</sub> Pathogenic for chickens and mice on intraperitoneal injection

*Salm braenderup* A F VI<sub>1</sub>, VI<sub>2</sub>, VII e, h ↔ e, n, z<sub>11</sub>

Isolated by Kauffmann and Henningsen (1938) in Denmark from the faeces of a 54 year old man suffering from enteritis and from his cat, which had died of diarrhoea Since found in South Africa (see Kauffmann 1941) Phase 2 also contains z<sub>17</sub> and z<sub>18</sub>

*Salm georgia* A F VI, VII . b ↔ e, n, z<sub>11</sub>

Isolated from healthy carrier (Morris, Brim and Sellers 1945)

*Salm concord* A F VI, VII . 1, v ↔ 1, 2, 3

Isolated in Massachusetts, also in England by Taylor at Oxford from a case of acute gastro enteritis in a child living on a farm, and from her cat

*Salm. tennessee* A.F. VI, VI<sub>2</sub>, VII  $x_2$  —

Isolated from the faeces of a healthy food handler employed in a fraternity house a number of whose occupants were suffering from food poisoning (Bruner and Edwards 1942a). Isolated from cases of food poisoning in Great Britain and from imported American spray-dried egg. Apparently monophase. A trace of  $x_1$  may be present.

*Salm. puertus* A.F. VI, VIII  $e, h \leftrightarrow 1, 2$

Isolated from the faeces of a boy at New Haven Hospital suffering from gastro-enteritis. Described by Wheeler and Borman (1943). Biochemically indistinguishable from *S. m. newport*.

*Salm. newport* A.F. VI, VIII  $e, h \leftrightarrow 1, 2, 3$

Isolated from cases of food poisoning in man (Schutze 1930). Corresponds to Paratyphus  $\beta$  of Weil and Saxl (1911). (See also White 1936, Kauffmann 1930a, 1931, 1934a, Schutze 1930, Claiberg 1931, Seligmann and Claiberg 1932, Boecker and Silberstein 1932.) One of the commonest types met with in outbreaks of food poisoning in Great Britain. Isolated by Khalil (1935) from rats at Liverpool from cases of infantile diarrhoea in Uruguay (Hormaeche Peluffo and Aleppo 1930, 1940), from chickens and turkeys in the United States (Edwards 1939) from retail meat in the United States (Cherry Scherago and Weaver 1943) and from the mesenteric lymph nodes of normal pigs in England (Scott 1940) and in Uruguay (Hormaeche and Salsamendi 1930, 1939). The *puerto-rico* variety (Jordan 1934, Kauffmann 1934a) exists only in the non-specific phase but by cultivation in the presence of an immune serum the specific phase may be obtained from it (Bruner and Edwards 1939). Has been isolated from imported American spray-dried egg.

*Salm. kotlibus* A.F. VI, VIII  $e, h \leftrightarrow 1, 3$

Isolated from cases of acute gastro-enteritis (Kauffmann 1934a). Differs from *S. m. newport* of which it used to be considered a variety in the non-specific phase.

*Salm. bovis-morbificans* A.F. VI, VIII  $r \leftrightarrow 1, 5$

Isolated from an infected cow by Baseman (1894). Has also been isolated from gastro-enteritis in man (see White 1936, Cladden and Scott 1927, Kauffmann and Miron 1930, Kauffmann 1931, 1934a, 1941, Claiberg 1931, Seligmann and Claiberg 1932, Boecker and Silberstein 1932). Not very common in cattle (Bartel 1935, Lutje 1939). Cultured from imported American spray-dried egg in Great Britain.

*Salm. muenchen* A.F. VI, VIII  $d \leftrightarrow 1, 2$

Isolated by Mandelbaum (1932) from a fatal case of gastro-enteritis (see also Silberstein 1932, Kauffmann 1934a, 1941). Found by Hormaeche Peluffo and Aleppo (1940) in infantile diarrhoea in Uruguay. Isolated from mesenteric lymph nodes of normal pigs in Uruguay by Hormaeche and Salsamendi (1939), and from chickens in the United States (Edwards 1939). Sometimes present in imported American spray-dried egg. Appeared to be responsible for an epidemic among guinea pigs in New York City (Bornstein, Saphra and Strauss 1941). The d antigen contains the partial antibodies d<sub>1</sub>, d<sub>2</sub> and d<sub>3</sub>.

*Salm. oregon* A.F. VI, VIII  $d \leftrightarrow 1, 2, 3$

Isolated from a turkey and from the mesenteric lymph nodes of normal pigs in the United States. Also found in imported American spray-dried egg. Described by Putan (1940) as a variant of *Salm. muenchen* but regarded by Edwards and Bruner (1941b) as a new type. The d antigen in the specific phase is identical with that of *muenchen*, *gim sara* and *shangri* but differs to some extent from that of *muenchen*, *typhi* and *starkey*.

*Salm. mexicana* A.F. VI, VIII  $d \leftrightarrow 1, 2, 4$

Isolated in Mexico from the mesenteric glands of a subject post mortem, and from the faeces of a child with diarrhoea (Varela, Zetava and Oarte 1943).

*Salm. manhattan* A.F. VI, VIII  $d \leftrightarrow 1, 5$

Isolated from a chicken and from a turkey in the United States. Found also in imported American spray-dried egg. Described by Edwards and Bruner (1941b). Isolated also

from a human source in Massachusetts (Bornstein and Saphra 1942) For the nature of the d antigen see *Salm oregon*

*Salm narashino* A F VI<sub>1</sub> VIII a ↔ e n x

According to Kauffmann (1941) this organism was isolated in 1937 from the blood and faeces of a patient with a typhoid like disease in Japan and its antigenic structure was determined by Nakaguro and Yamashita Phase 2 contains also z<sub>10</sub> and z<sub>11</sub>

*Salm glostrup* A F VI<sub>1</sub> VIII z<sub>10</sub> ↔ e n z<sub>15</sub>

Isolated by Kauffmann and Henningsen (1939) from the faeces of patients in a family outbreak of gastro enteritis and from the faeces and blood of their dog which was ill at the same time Phase 2 contains also z<sub>11</sub> and z<sub>13</sub> Non pathogenic to mice by the mouth

*Salm Hitchfield* A F VI<sub>1</sub> VIII 1 v ↔ 1 2 3

Isolated from the liver of a turkey poult in the United States Described by Edwards and Bruner (1940b) A strain isolated many years previously from a case of food poisoning in man was later found to belong to this type Also isolated from American spray dried egg in Great Britain

*Salm duesseldorf* A F VI<sub>1</sub> VIII z<sub>4</sub> z<sub>24</sub> —

Isolated in Germany from two patients in hospital—a boy of 3 years and a man of 40 years who died (Hohn 1940)

*Salm bonariensis* A F VI<sub>1</sub> VIII 1 ↔ e n x

Isolated from the mesenteric gland of a normal pig (Monteverde 1942) Since met with in a normal human carrier (Edwards and Bruner 1943) Antigenic structure not yet completely worked out

*Salm virginia* A F VIII d —

Studied by Selgmann and Saphra (see Edwards 1945)

*Salm amherstiana* A F (VIII) 1 (v) ↔ 1 6

Isolated from the liver of one of a group of poult affected with a fatal disease This is the only known salmonella to contain the VIII antigen alone and even this is incomplete It is perhaps doubtful whether it should be included in Group C Described by Edwards and Bruner (1942b)

## GROUP D

*Salm typhi* A F IX XII [Vi] d —

The cause of typhoid fever in man Its general characters have been referred to in previous sections The great majority of freshly isolated strains contain the Vi antigen The d antigen contains the partial antigens d and d Dwarf colonies containing the O but not the H or Vi antigens may occur Artificial phase variation may follow growth in the presence of a d antiserum the d antigen being replaced by j (Kauffmann 1936c) the new phase is referred to as Phase 3 Though many strains of *Salm typhi-suis* *Salm sendai* and *Salm gallinarum* fail to produce gas from glucose *Salm typhi* is unique among the *Salmonella* group in never forming gas Sub types based on the presence or absence of fermental on of xylose arabiose d tartrate and sodium citrate have been described but for epidemiological purposes the method of bacteriophage typing is of greater usefulness Does not appear to infect animals under natural conditions (For properties of Vi antigen see p 152c and a short review by Almon 1943)

*Salm enteritidis* A F [I] IX XII g m —

Referred to sometimes as the *gartner* or *jena* variety Isolated from the spleen of a fatal case of food poisoning which affected 58 persons at Frankenhausem who had eaten the flesh of an emergency slaughtered cow (Gaertner 1888) Since isolated from numerous sporadic cases and outbreaks of food poisoning in different parts of the world (see White 1926 1930 Kauffmann 1930b 1931 1934a Warren and Scott 1930 Boecker and Silberstein 1932 Boecker 1936) The full formula of the flagellar antigen is g o, m z<sub>1</sub> z<sub>4</sub>



*Salm enteritidis* has four fermentative varieties (see Table 46). the original, *gartner*, or *jena* variety, just described, the *danzysz* variety, sometimes known as the *Ratin bacillus*, the *essen* variety, and the *chaco* variety. The *danzysz* variety was isolated by Danysz (1900) from an epidemic of mouse typhoid in field mice. The *Ratin* strain of this variety was cultured from the urine of a sick child by Neumann in 1902 (see Report 1934 Kauffmann 1934a). Both the *Ratin* and the *Liverpool* "viruses" are prepared with this variety (Leslie 1942). The *essen* variety was isolated from cases of gastro-enteritis in man, from ducks, and from ducks' eggs by Hohn and Herrmann (1935a, b) and described as *Salm moskau*, it is not to be confused with *Salm essen*, which belongs to the B group of *Salmonella* (see p 729), nor with *Salm moscov* (see below). The *chaco* variety was isolated by Savino and Menéndez (1934) from cases of continued fever in the Chaco war.

Apart from the *chaco* variety, which has so far been met with only in man, the natural habitat and frequency distribution of the other three varieties of *Salm enteritidis* is subject to doubt. So many workers who have isolated *Salm enteritidis* from cattle, horses, pigs, chickens, ducklings, rats, and other sources have failed to differentiate between, or make no mention of the different fermentative varieties that much of the available information is difficult to interpret. The *jena* variety appears to be responsible for most outbreaks of food poisoning due to *Salm enteritidis* that follow the consumption of meat. It does not, however, appear to be very common and it constituted only 1.06 per cent of 1 690 *Salmonella* strains isolated from domestic animals commonly used for food (Barthel 1938). It seems probable, also, that it may give rise at times to purulent meningitis (Guthrie and Montgomery 1933). The *danzysz* variety appears to be predominantly a parasite of rodents: it has been isolated from silver foxes suffering from distemper or paratyphoid (Benedict *et al* 1941). Its pathogenicity for man has been disputed, but a number of cases and outbreaks of gastro enteritis in man have now been described in which the causative role of this organism and its origin from rat "virus" are well established (see Leslie 1942): some of the cases have proved fatal. The *essen* variety appears to be characteristically a parasite of ducks (Jansen 1937) but is capable of giving rise to acute gastro enteritis in human beings. (For further information on the varieties of *Salm enteritidis*, see Kauffmann 1941.)

*Salm dublin* A F I, IX, XII G, P —

Sometimes known as the *kyl* variety of *Salm enteritidis*. Isolated on several occasions from calf diarrhoea and described under the name "paracolon bacillus," but not adequately differentiated till examined by White (1930). (See also Warren and Scott 1930, Smith and Scott 1930, Stroman and Örn 1932, Kauffmann 1930b, 1931, 1934a, Bosworth and Lovell 1931, Boecker and Silberstein 1932, Hohn and Herrmann 1935a). In Europe its principal habitat appears to be calves, less frequently older cattle. It is very common: for instance, it constituted 394 out of 456 *Salmonella* strains from cattle studied by Lütje (1939), 314 of the 394 strains were isolated from calves or occasionally foetuses and only 72 from adult cattle. It formed 78.37 per cent of Bartels (1938) series of 1,690 *Salmonella* strains isolated from domestic animals commonly used for food. In the United States it seems to be found principally in silver foxes, and only seldom in cattle (Edwards and Bruner 1943). It has been isolated from fowls suffering from enteritis (Lutje 1937), and from the mesenteric lymph nodes of normal pigs (Scott 1940). In man it may give rise to a continued fever of the enteric type (Smith and Scott 1930), and sometimes to meningitis and cholecystitis (Guthrie and Montgomery 1939). As a cause of acute gastro enteritis in man it is less common than the *jena* variety of *Salm enteritidis* but it has been responsible for more than one milk borne outbreak of the acute food poisoning type (Conybeare and Thornton 1938, Tulloch 1939). In calves it is responsible for enteritis (Bosworth and Lovell 1931)—the so-called calf-diarrhoea—but in adult cattle it affects more often the udder, lungs or heart giving rise to inflammation. Chronic carriers are not unknown in cattle. In the past many strains of this organism have probably been described as *Salm enteritidis*. Two fermentative varieties have been described *accra* and *koeln*. Occasional variants fail to form gas.

*Salm rostock* A.F. I IV XII g p u —

Isolated from cattle. First differentiated on basis of fermentation reactions (Bahr 1930; b) Antigenic structure studied by Kauffmann (1930b 1931 1934a, 1935b) (See also Stroman and Orn 1932 Hohn and Herrmann 1935a) It appears to be relatively uncommon in cattle in Germany (Bartel 1938 Iltje 1939) and has not yet been isolated from human beings.

*Salm moscow* A.F. IX XII g q —

Isolated mainly in Russia from febrile infections in man (Weigmann 1925a b, Iwaschenko 1926) It has since been isolated from cases of gastro-enteritis in man (Hohn and Herrmann 1935a Kauffmann 1935b) and from a cow and a horse (Kauffmann 1935b) Its structure was first determined by Hicks (1930) (see also Kauffmann 1930b 1931 1934a 1935b) The full formula for its flagellar antigen is g o, q x<sub>1</sub> It is doubtful whether it can give rise primarily to a typhoid like disease in man most of the Russian strains appear to have been cases of relapsing fever in which *Salm moscow* was a secondary invader Not to be confused with *Salm enteridis* var *essen* which was first described as *Salm moscow* (Hohn and Herrmann 1935a)

*Salm blegdam* A.F. IX XII g m q —

Isolated from the blood of a patient in Denmark suffering from pneumonia described by Kauffmann (1935b) Does not appear to have been met with again

*Salm pensacola* A.F. IX XII g m t —

Studied by Edwards (1945)

*Salm berta* A.F. IX XII f g t —

Isolated from the pooled mesenteric lymph nodes of normal pigs in Uruguay and reported to a local society in 1937 described by Hormaeche Peluffo and Salsamendi in 1938 Studied also by Kauffmann (1937b) H antigen contains also x<sub>2</sub> and x<sub>3</sub> H<sub>2</sub>S production irregular Pathogenic to rats by mouth and by subcutaneous and intraperitoneal injection when given in large doses

*Salm eastbourne* A.F. [1] IX, XII c h ↔ 1, 5

Isolated from a case of enteric fever (Leslie and Shera 1931) (See also Kauffmann 1934a) Subsequently met with in the United States and in Norway Bornstein and Saphra (1942) examined a strain that had been isolated from the spinal fluid of a child who died of meningitis Also found in animals Some strains are peculiar in producing indole Inoculation fermentation is irregular and occasional variants fail to form gas

*Salm sendai* A.F. [1] IX, XII a ↔ 1, 5

Isolated from cases of enteric fever in Japan (Aoki and Sakai 1925) Antigenic structure determined by White (1926) and Kauffmann (1931 1934a) Since isolated from a patient with a typhoid like fever in Georgia (Bornstein and Saphra 1942) Closely related to *Salm paratyphi A* from which it differs in containing the ix antigen in possessing a group j base (which can however be artificially induced in *Salm paratyphi A* by growth in the presence of immune serum) in fermenting xylose and in forming little or no gas from dextrose Resembles *Salm paratyphi A* in growing more poorly than most other *Salmonella* strains

*Salm loma linda* A.F. IX, XII a ↔ e n x

*Salm onarimon* A.F. I IV XII b ↔ 1, 2

Isolated from the stools of a healthy woman in Japan Described by Kusida (1940) Closely resembles *Salm paratyphi B* and like that organism it forms a mucoid wall Was later isolated by Anzai and Tsurumi (1940) from the blood of a patient with a typhoid like disease which began with dysenteric symptoms

*Salm dar-es-salaam* A.F. I IV XII l w ↔ e n

Isolated from the urine of a febrile patient in Dar es Salaam Antigenic structure determined by White (1926) (See also Kauffmann and Mitsui 1930 Kauffmann 1931 1934a 1940e) Peculiar in that it liquefies gelatin Variable in its ability to grow on an ammonium base medium Full formula of Phase 2 is e n x<sub>12</sub> x<sub>13</sub>

*Salm. goettingen* A.F. IX, XII . . . 1,  $v \leftrightarrow e$ , n,  $z_{15}$  . . .

Described by Hohn (1940) in Germany; origin not stated, except that it was isolated locally. Studied by Kauffmann (1940c).

*Salm. durban* A.F. IX, XII . . .  $s \leftrightarrow e$ , n,  $z_{15}$  . . .

*Salm. panama* A.F. I, IX, XII . . . 1,  $v \leftrightarrow 1$ , 5 . . .

Isolated by Jordan (1934) from a food poisoning outbreak. Studied by Kauffmann (1934a, 1937a). Found by Schiff (1938) in New York City in 4 cases of infantile diarrhoea, 2 of which proved fatal, and by Schiff and Strauss (1939b) in three further infections, one of which was in an adult. Isolated from cases of infantile diarrhoea in Uruguay by Hormaeche, Peluffo and Aleppo (1940). Was responsible for 6.1 per cent of 2,285 *Salmonella* infections in the United States and Panama in which the causative organism was studied by Edwards and Bruner (1943). It is found in animals, but its natural habitat is not known. Strains vary in their ability to ferment *d* tartrate, and in the rate at which they attack dulcitol. Occasional strains are said to form indole (Seligmann and Saphra 1943).

*Salm. italiana* A.F. IX, XII . . . 1,  $v \leftrightarrow 1$ , 11 . . .

Studied by Bruner (see Edwards 1945).

*Salm. clabornei* A.F. I, IX, XII . . .  $k \leftrightarrow 1$ , 5 . . .

Isolated from a patient suffering from gastro-enteritis at Camp Claiborne, Louisiana, and described by Wilcox and Lennox (1944).

*Salm. gallinarum* A.F. IX, XII . . . — . —

Isolated from fowls suffering from fowl typhoid (Klein 1889). (See also Moore 1893, Pfeiler and Rehse 1913, Pfeiler and Roepke 1917, White 1926, Kauffmann 1934a, b). Differs from other members of *Salmonella* group in being non flagellated and possessing therefore only an O antigen. Forms little or no gas from dextrose, fermentation of other sugars is relatively slow and feeble. *Salm. gallinarum* appears to be seldom pathogenic to man, but a fermentative variant, known as *dunburg*, which fails to attack *d* tartrate or to produce  $H_2S$ , may be responsible for cases of acute gastro-enteritis (see Muller 1933, Kauffmann 1934b). A more important fermentative variant, often regarded as a separate type and called *Salm. pullorum*, is responsible for the widespread disease of chickens known as bacillary white diarrhoea. Whether this organism is antigenically identical with *Salm. gallinarum* is still under discussion. It was isolated by Rettger (1900), and has been studied by numerous workers (see Rettger and Harvey 1908, Rettger 1909, Smith and Tenbroeck 1915, Gage and Martin 1916, Krumwiede and Kohn 1917, Rettger and Hoser 1917, Hadley *et al.* 1917, Mulsow 1919, Winslow *et al.* 1919, St. John Brooks and Rhodes 1923, White 1926, Kauffmann 1931, 1934a, b). *Salm. pullorum* differs from *Salm. gallinarum* in producing gas in dextrose, and in failing to ferment maltose, dulcitol, dextrin or *d* tartrate, it also grows more poorly. Hinshaw, Browne and Taylor (1943), however, who studied 300 strains of *Salm. pullorum*, found that 43 strains produced acidity in maltose in 24 hours, and that a further 158 produced some degree of acidity within 4 weeks. Of the total 300 strains, 40 failed to produce gas in dextrose. It is mainly a parasite of chickens, but has been isolated from sparrows (Dalling, Mason and Gordon 1928), silver foxes (Benedict *et al.* 1941), pigs, mink and man (Edwards and Bruner 1943), and also from imported American spray-dried egg.

*Salm. napoli* A.F. IX, XII . . . 1,  $z_{13} \leftrightarrow e$ , n, x . . .

*Salm. javiana* A.F. [I], IX, XII . . . 1,  $z_{13} \leftrightarrow 1$ , 5 . . .

Isolated from faeces of a child suffering from gastro-enteritis in Batavia, and from human carriers in Panama. Described by Edwards and Bruner (1942b). Closely related to *Salm. panama*, but differs in the specific phase. The  $z_{13}$  antigen contains part of the  $z_{13}$  factor of *uganda*. Not to be confused with the *java* variety of *Salm. paratyphi* B.

*Salm. canstel* A.F. IX, XII . . .  $z_{13} \leftrightarrow 1$ , 5 . . .

Studied by Bruner and Randall (see Edwards 1945).

## GROUP E

*Salm london* A F III, X, XXVI, l, v  $\longleftrightarrow$  1, 6

Isolated by White (1926) from a patient at Reading with gastro enteritis and described as Type L. Later called *london* by Kauffmann (See also Kauffmann 1930b 1931, 1934a, 1939a Kauffmann and Silberstein 1934) Cultivated by Černozubov, Filipović and Staval (1936-37) in Yugoslavia from the faeces of three patients who developed febrile diarrhoea, accompanied by the passage of blood and mucus, after eating sausage. Found by Hormaeche, Peluffo and Aleppo (1936) in infantile diarrhoea in Uruguay. Isolated from mesenteric glands of normal pigs in Uruguay (Hormaeche and Salsamendi 1939) and in Great Britain (Scott 1940). Also isolated from chickens in the United States (Edwards 1939), and from imported American spray dried egg in Great Britain.

*Salm give* A F III, X, XXVI, l, v  $\longleftrightarrow$  1, 7

Isolated from a patient in Spain with long standing diarrhoea. Described by Kauffmann (1937b). Since isolated from gastro enteritis in a child in the United States (see Bornstein, Saphra and Straus 1941), from mesenteric lymph nodes of healthy pigs in the United States (Rubin, Scherago and Weaver 1942) and from chickens in the United States (see Mallmann *et al* 1942). Found also in imported American spray-dried egg.

*Salm uganda* A F III, X, XXVI, l, z<sub>13</sub>  $\longleftrightarrow$  1, 5

Isolated from the spleen of a fatal case of pyrexia of unknown origin in Uganda. Described by Kauffmann (1940c). The l antigen is not identical with that of *london* and *dar es-salaam*, nor are the 1, 5 antigens identical with the 1, 5 antigens of *Salm thompson*. Phase l contains small amounts of antigens v and w. Distinguished biochemically from *london* by its fermentation of : tartrate, its more rapid fermentation of l tartrate, and its failure to ferment inositol.

*Salm anatum* A F III, X, XXVI, e, h  $\longleftrightarrow$  1, 6

Isolated from an epidemic intestinal infection of ducklings, known as keel disease (Rettger and Scoville 1919, 1920) (See also Edwards and Rettger 1927 Lovell 1932b, Kauffmann 1934a, Kauffmann and Silberstein 1934). Many early strains described under this name were really *Salm typhi murum*. Has been isolated by Hormaeche, Peluffo and Aleppo (1936, 1940) from infantile diarrhoea in Uruguay, by Rauss (1941) in Hungary from the faeces of healthy persons, by Edwards (1939) in the United States from chickens and turkeys, by workers in Great Britain from cases of food poisoning and from imported American spray dried egg, from mesenteric lymph nodes of normal pigs in the United States by Rubin, Scherago and Weaver (1942) and in Mexico by Varela and Zozaya (1942) and from silver foxes in the United States (Benedict *et al* 1941).

*Salm muenster* A F III, X, XXVI, e, h  $\longleftrightarrow$  1, 5

Isolated from a food poisoning outbreak due to raw horse flesh. Described by Kauffmann and Silberstein (1934). Also met with in cases of infantile diarrhoea in Uruguay (Hormaeche, Peluffo and Aleppo 1940). Differs biochemically from *Salm anatum* in fermenting inositol and in failing to ferment : tartrate.

*Salm vejle* A F III, X, XXVI, e, h  $\longleftrightarrow$  1, 2, 3

According to Kauffmann (1941) this organism was isolated by Møller at Copenhagen from acute gastro enteritis, and described by Harhoff.

*Salm amager* A F III, X, XXVI, y  $\longleftrightarrow$  1, 2, 3

Isolated in Denmark from the faeces of a patient suffering from enteritis. Described by Kauffmann (1939a). Non pathogenic to mice by the mouth.

*Salm shangani* A F III, X, XXVI, d  $\longleftrightarrow$  1, 5

Isolated from faeces of a patient with febrile diarrhoea and a commencing miscarriage at Zanzibar. Described by Kauffmann (1939a). The d antigen is not identical with that of *typhi*. Non pathogenic to mice by the mouth.

*Salm. meleagridis* A.F. III, X, XXVI, e, h  $\leftrightarrow$  1, w

(Name derived from *meleagris*—a kind of guinea fowl, pronounced *mèlèagris*). Isolated in Minnesota from outbreaks of infection in turkey poults, later found in Massachusetts, Michigan and California. Described by Bruner and Edwards (1911b). Isolated also from a patient with a typhoid like fever in Venezuela and from New York sewage (Bornstein and Saphra 1942), from the mesenteric lymph nodes of normal pigs in Mexico (Varela and Zozava 1942), from German soldiers in Norway (see Kauffmann 1941), and from American dried egg in England. As in *Salm. worthington*, the H antigens in Phase 2 are common in Phase 1 of other *Salmonella* types

*Salm. nyborg* A.F. III, X, XXVI, e, h  $\leftrightarrow$  1, 7 . . .

Isolated in Denmark from a case of acute enteritis in a child of 6 years. Described by Kristensen and Bojlen (1936) and studied by Kauffmann (1937b). The O antigen is not completely identical with that of *Salm. anatum*, as an *anatum* serum after absorption with *nyborg* still contains agglutinins for *anatum* and for other organisms containing the III, X, XXVI combination.

*Salm. zanzibar* A.F. III, X, XXVI, k  $\leftrightarrow$  1, 5 . . .

Isolated from faeces of a healthy carrier in Zanzibar. Described by Kauffmann (1939a). The k antigen is identical with that of *thompson*. The O antigen is slightly aberrant. Non pathogenic to mice by the mouth.

*Salm. lexington* A.F. III, X, XXVI,  $\alpha_{12}$   $\leftrightarrow$  1, 5 . . .

Isolated from mesenteric lymph nodes of normal pigs. Described by Edwards, Bruner and Rubin (1940). (See also Rubin *et al.* 1942). According to Kauffmann (1941) an organism having the same antigenic formula was found independently by Erber in the Dutch East Indies and described as *Salm. bataria*. The O antigen of *Salm. lexington* is slightly aberrant and contains a special factor in addition to the III, X, XXVI factors.

*Salm. weltevreden* A.F. III, X, XXVI, r  $\leftrightarrow$   $\alpha_4$  . . .

*Salm. newington* A.F. III, XV, e, h  $\leftrightarrow$  1, 6 . . .

Isolated by Rettger from ducklings in Connecticut, and described by Edwards (1937). Also isolated from 3 cases of gastro-enteritis in man in the United States and from sewage (Bornstein and Saphra 1942), from chickens and turkeys in the United States by Edwards (1939), from the mesenteric lymph nodes of normal pigs in Uruguay (Hormaeche and Salsamendi 1939) and in the United States (Rubin *et al.* 1942), from silver foxes in the United States (Benedict *et al.* 1941), and from imported spray-dried egg in Great Britain. Phase 2 contains 4 in addition to the I and 6 factors. Under the name of *Salm. tim*, Kauffmann (1937b) described a variant of *newington* isolated from two patients suffering from gastro-enteritis in Denmark. It is differentiated from *newington* by the greater complexity of the e, h antigen, and by its late or inconstant fermentation of maltose and dextrin.

*Salm. selandia* A.F. III, XV, e, h  $\leftrightarrow$  1, 7 . . .

Isolated from the faeces of a young sailor, belonging to S.S. *Selandia*, who on a long voyage to Asia and Australia had suffered from repeated diarrhoea. On his return to Denmark he developed fever, lung symptoms, and diarrhoea following constipation. Described by Kauffmann (1937b). Also found in Havana (Seligmann, Saphra and Wassermann 1944).

*Salm. new-brunswick* A.F. III, XV, I, r  $\leftrightarrow$  1, 7

Isolated from a baby chick. Described by Edwards (1937). Found also in mesenteric lymph nodes of normal pigs in the United States (Rubin *et al.* 1942). Two strains have been isolated in Denmark, one from a woman with acute gastro-enteritis, the other from a patient who had returned from the tropics and was suspected of having malaria (Kauffmann 1941).

*Salm cambridge* III XV  $1 \leftrightarrow e h$

Isolated from a soldier suffering from Sonne dysentery Studied by Taylor (unpublished)

*Salm illinois* A.F. III XV  $z_{10} \leftrightarrow 1 5$

Isolated from pgs in Illinois Hungarian partridges in Michigan and turkeys in Minnesota Described by Edwards and Bruner (1941c) Some doubt about the constitution of the O antigen which is given as (III) (XV) XXXIV by Edwards and Bruner but as III XV by Kauffmann (1941)

*Salm taksony* A.F. I III XV  $i \leftrightarrow z_4$

Described by Rauss (1943) O antigen is identical with that of *Salm senftenberg*

*Salm senftenberg* A.F. I III XV  $g s t$  —

Isolated by Kauffmann (1939c) though not under the name of *Salm senftenberg* from an 8 year old boy suffering from acute gastro-enteritis (See also Kauffmann 1939b 1934a 1941 Kauffmann and Ntsui 1939 Boecker and Silberstein 1939) Found also in the United States in young turkeys suffering from an epidemic disease (Edwards 1937) and in chickens (Edwards 1939) in a human carrier in the United States and in Chinese egg (Bornstein and Saphra 1942) in retail meat in the United States (Cherry Scherago and Weaver 1943) and in the mesenteric lymph nodes of normal pgs in Mexico (Varela and Zozaya 1942) Isolated from imported American spray-dried egg in Great Britain Included in the flagellar antigens are  $z_4$  and  $z_5$  *Salm senftenberg* var *newcastle* was isolated from the faeces of a healthy woman under conditions that precluded any opinion as to its pathogenic role (Warren and Scott 1930) It differs from *senftenberg* in failing to produce  $H_2S$  and to ferment glycerol and tartrate

*Salm niloese* A.F. I III XV  $d \leftrightarrow z_4$

Isolated from the faeces of a patient suffering from acute gastro-enteritis in Denmark and later found repeatedly in similar cases Described by Kauffmann (1939a) The d antigen is not identical with that of *Salm typh* but the  $z_4$  antigen is identical with that of *Salm kentucky* Non pathogenic to mice by the mouth

*Salm simsbury* A.F. I III XV  $z_{11}$  —

Isolated from normal human faeces in the United States (Bruner and Edwards 1942a)

## OTHER GROUPS

*Salm aberdeen* A.F. VI  $i \leftrightarrow 1 2 3$

Isolated from a case of acute gastro-enteritis in an infant (Smith 1934) Since found in chickens in the United States (Kauffmann *et al* 1942)

*Salm rubislaw* A.F. VI  $r \leftrightarrow e n x$

Isolated from the faeces of a boy in Scotland who was suffering from enteritis (Smith and Kauffmann 1940) The r antigen is identical with that of *tychov* and the  $e n x$  antigens with those of *abortus equi* Also found in American sewage (Bornstein and Saphra 1942)

*Salm pretoria* A.F. VI  $k \leftrightarrow 1 2 3$

*Salm salt* A.F. XI  $y \leftrightarrow 1 5$

Isolated from faeces of a normal person Described by Rauss (1943) O antigen is identical with that of *Salm aberdeen*

*Salm grumpensis* A.F. XIII XXII  $d \leftrightarrow 1 7$

*Salm poona* A.F. XIII XXII  $z \leftrightarrow 1 6$

Isolated from a case of acute gastro-enteritis in an infant (Bridges and Scott 1935)

*Salm borbeck* A.F. XIII XXII  $i v \leftrightarrow 1 6$

Isolated from the faeces of a child suffering from typhoid fever in Germany (Hohn and Herrmann 1940)

*Salm. worthington* A.F. I, XIII, XXIII, 1, w  $\longleftrightarrow$  z . . .

Isolated from a young turkey and a chick in Minnesota (Edwards and Bruner 1933). Found also in the United States in human faeces (Bornstein and Saphra 1942, Borman *et al.* 1943) and in the mesenteric lymph nodes of normal pigs (Rubin *et al.* 1942). Isolated from imported American spray-dried egg in Great Britain. The z antigen is not quite identical with that of *poona*.

*Salm. wichita* A.F. I, XIII, XXIII, d . . . —

Isolated from the faeces of infants in Kansas who were suffering from neonatal diarrhoea (McKinlay 1937), and studied by Schiff and Strauss (1939a).

*Salm. habana* A.F. I, XIII, XXIII, f, g . . . —

Isolated from cerebrospinal fluid, blood and feces during the course of a hospital outbreak of purulent meningitis among new born babies in Havana, all the infants admitted to hospital, 21 in number, died (Schiff and Saphra 1941). Since found in imported American spray-dried egg. Like all *Salmonella* strains containing the g antigen, it is monophasic. The flagellar antigen is not quite identical with that of *derby*, there appears to be an extra minor antigen in both organisms.

*Salm. mississippi* A.F. I, XIII, XXIII, b  $\longleftrightarrow$  1, 5 . . .

Isolated from the faeces of a normal human carrier (Edwards and Bruner 1943).

*Salm. heves* A.F. (I), VI, XIV, XXIV . d  $\longleftrightarrow$  1, 5 . . .

Described by Rauss (1943).

*Salm. carrau* A.F. VI, XIV, XXIV, y  $\longleftrightarrow$  1, 7 . . .

Isolated from the mesenteric lymph node of a normal pig in Uruguay (Hormaeche, Peluffo and Salsamendi 1938). Also found in infantile diarrhoea (Hormaeche, Peluffo and Aleppo 1940). It is probable that the XIV antigen of *onderstepoort* (see below) consists of two fractions, one of which is specific and the other common to *carrau*. Hormaeche, Peluffo and de Pereyra (1944) find that *Salmonella* strains having factor 7 in Phase 2 must be divided into two sub-groups, sub group 1 possesses a factor that is lacking in sub group 2. Sub group 1 contains *arecharaleia*, *allendorf*, *florida*, *gaminara*, *kaapstad* and *pomona*. Sub-group 2 contains *bredeney*, *carrau*, *grumpensis*, *selandia*, *give*, *new-bruns wick*, *nyborg* and *madela*.

*Salm. onderstepoort* A.F. (I), VI, XIV, XXV, e, (h)  $\longleftrightarrow$  1, 5 . . .

Isolated from 2 sheep in South Africa. Described by Henning (1935). (For the constitution of the XIV antigen, see *Salm. carrau*). Is pathogenic to mice inoculated intra peritoneally.

*Salm. florida* A.F. (I), VI, XIV, XXV, d  $\longleftrightarrow$  1, 7 . . .

Isolated from faeces of a patient suffering from febrile diarrhoea. Described by Cherry, Edwards and Bruner (1943). The d antigen is not quite identical with that of *Salm. typhi*, *Salm. oregon* or *Salm. muenchen*.

*Salm. madella* A.F. (I), VI, XIV, XXV, y  $\longleftrightarrow$  1, 7 . . .

Isolated from the liver of a poult. Described by Cherry, Edwards and Bruner (1943). The y antigen is identical with that of *Salm. bareilly*.

*Salm. sundsvall* A.F. (I), VI, XIV, XXV . . . z  $\longleftrightarrow$  e, n, x, z<sub>11</sub> . . .

Isolated in Scandinavia. Responsible in England for an outbreak of gastro-enteritis. Also found in imported American spray-dried egg.

*Salm. horsham* A.F. (I), VI, XIV, XXV . 1, v  $\longleftrightarrow$  e, n, x . . .

Isolated in England from imported spray-dried egg powder. Studied by Taylor (unpublished).

*Salm. hvittingfloss* A.F. XVI, b  $\longleftrightarrow$  e, n, x . . .

Isolated in Norway from the faeces of patients who were suffering from acute gastro-enteritis following the consumption of soft cheese, and from the cheese itself, one patient had a typhoid like disease that lasted for a week (Teedal 1936, 1938). Also found in

fowls in the United States (Mallmann *et al* 1942) The O antigen was given originally as VIII Non pathogenic to mice by the mouth

*Salm gaminara* A F XVI d  $\leftrightarrow$  1 7

Isolated from a child in Uruguay suffering from dysenteriform enteritis (Hormaeche and Peluffo 1939) Hormaeche and Peluffo who analysed the d antigen of this and other salmonellae gave the following distribution —*Salm gam nara* d d<sub>1</sub> d<sub>2</sub> *Salm stanley* d d<sub>2</sub> *Salm amersfoort* d d<sub>2</sub> d<sub>3</sub> *Salm munichen* d d<sub>2</sub> d<sub>4</sub> *Salm typhi* d d<sub>1</sub> The non specific phase contains an antigen that has not been met with in other species of *Salmonella* Pathogenic for rats by the mouth and subcutaneously if given in large doses and for rabbits intravenously and subcutaneously The non specific phase is said to be more toxic than the specific phase

*Salm szentes* A F XVI k  $\leftrightarrow$  1 2 3

Described by Rauss (1943) O antigen appears to be identical with that of *Salm huttnigfoss*

*Salm kirkee* A F XVII b  $\leftrightarrow$  1 2

Isolated from the stools of an infant in India suffering from diarrhoea of the dysenteric type (Bridges and Dunbar 1936) The O antigen was given originally as XIV

*Salm cerro* A F XVIII z<sub>1</sub> z<sub>23</sub> z<sub>25</sub> —

Isolated in 1936 from the pooled mesenteric lymph nodes of normal pigs in Uruguay Later isolated from infants suffering from enteritis and sometimes rhinopneumonitis (Hormaeche Peluffo and Aleppo 1941) Found in imported American spray-dried egg Shares the z<sub>4</sub> antigen with *duesseldorf* and *arizona*

*Salm kentucky* A F (VIII) XX 1  $\leftrightarrow$  z<sub>6</sub>

Isolated from a chicken in the United States suffering from enteritis (Edwards 1938) and from pheasants (Edwards 1939) Found also in a human case of gastroenteritis in the United States (Bornstein and Saphra 1942) in a mild case of febrile diarrhoea in Palestine and in camels in Palestine (Oltzki 1942) and in imported American spray-dried egg in Great Britain The Palestine strains differed from the American *kentucky* strains in minor fermentative respects

*Salm minnesota* A F XXI XXVI b  $\leftrightarrow$  e n x

Isolated from a turkey poult (Edwards and Bruner 1938) (See also Kauffmann 1939a) Found by Hormaeche Peluffo and Aleppo (1940) in infantile diarrhoea in Uruguay Isolated from American spray-dried egg The b antigen is not completely identical with that of *Salmonella paratyphi B*

*Salm tel-aviv* A F XXVIII y  $\leftrightarrow$  e n z<sub>15</sub>

Isolated from a sick cow and from an epidemic disease of chickens accompanied by a 50 per cent case mortality in Palestine Studied by Kauffmann (1940a)

*Salm pomona* A F XXVIII y  $\leftrightarrow$  1 7

*Salm ballerup* A F XXIX (vi) z<sub>14</sub> —

Isolated from the feces of a woman in Denmark who had a history of gastro-enteritis lasting for several weeks (Kauffmann and Moller 1940) Somatic antigen contains also a small amount of the XIX fraction related to *senftenberg* *Salm ballerup* forms two types of colony The relatively stable V form which is smooth and opaque contains a vi antigen the unstable W form which is smooth and translucent does not The vi antigen is the same as that in *Salm typhi* and *Salm paratyphi C* and will immunize mice against the V form of *Salm typhi* (Kauffmann and Moller 1940) Similarly rabbit serum containing vi antibodies to *Salm ballerup* affords passive protection to mice against inoculation with *Salm typhi* (Longfellow and Luppold 1943) Both the V and the W forms of *Salm ballerup* are relatively non pathogenic to mice by the mouth and intraperitoneally An extra flagellar antigen z<sub>21</sub> is said to be present (Monteverde and Leiguarda 1944)



*Salm. hormæchel* A.F. XXIX, [Vi],  $z_{30}$  —

Isolated from sewage at Buenos Aires and described by Monteverde and Leiguarda (1944). Resembles *Salm. ballerup* in the variability of its Vi antigen, which may be present or absent. Contains in addition to  $z_{30}$  the flagellar antigen  $z_{31}$ , but this is subject to quantitative fluctuation. Non pathogenic for guinea pigs on intraperitoneal inoculation, but large doses inoculated intraperitoneally kill mice in 24 hours, and the organism can be recovered from the heart blood.

*Salm. urbana* A.F. XXX,  $b \leftrightarrow c, n, x \dots$ 

Isolated in the United States from the colon of a pig dying of hæmorrhagic enteritis, and from the gut of a dead chicken (Edwards and Bruner 1941a). Found also in patients suffering from enteritis in Massachusetts (Bornstein and Saphra 1942). Contains some XVI antigen, which it shares with *kruttingsfoss*. The b antigen is incomplete, so that absorption with *Salm. urbana* of a serum prepared against *Salm. paratyphi B* leaves a considerable part of the b antibody behind. Phase 1 probably contains an antigen, as yet unidentified, in addition to b. Phase 2 contains the  $z_{11}$  antigen, the c, n, x,  $z_{11}$  complex is shared with *abortus-boris* and *minnesota*.

*Salm. arizona* A.F. XXXIII,  $z_1, z_{11}, z_{11} \dots$  —

Isolated in the United States from certain reptiles by Caldwell and Rveron (1939). Studied by Kauffmann (see Kauffmann 1941). Resembles *Salm. dar-es-salaam* in slowly liquefying gelatin. Is peculiar in sometimes fermenting lactose, though not for 2 weeks or so. Cultures appear to be partly rough, and the antigenic structure is therefore still in doubt. Appears to be pathogenic for certain reptiles, and is highly pathogenic for guinea pigs and rabbits. Has since been isolated from a woman suffering from high fever, diarrhoea and vomiting (Seligmann, Saphra and Wassermann 1944).

*Salm. adelaide* A.F. XXXV, f, g —

Isolated from faeces during life, and from liver and spleen post mortem, of a man suffering from enteritis in Australia. Described by Atkinson (1943). Isolated on at least four occasions in England from cases of gastro-enteritis—one of them in a German prisoner-of-war. Original strain said not to ferment sorbitol, but English strains all ferment this sugar.

*Salm. inverness* A.F. XXXVIII,  $k \leftrightarrow 1, 6$ 

Isolated from a normal food handler in Florida and described by Edwards and Hughes (1944). Possesses a somatic antigen not previously described. The k antigen is the same as that in *Salm. thompson*.

Other organisms containing antigens of the *Salmonella* group

Since Habs and Arjona (1935) described a paracolon bacillus containing part of one of the *Salmonella* O antigens, and Gard (1937) and Gard and Eriksson (1939) described coliform bacilli containing *Salmonella* H antigens, several workers have reported the occurrence of O less often of H, antigens in paracolon, coliform and Flexner dysentery bacilli (Bornstein, Saphra and Daniels 1941, Schuff, Bornstein and Saphra 1941, Kauffmann 1941, Saphra and Silberberg 1942, Peluffo, Edwards and Bruner 1942). These workers alone have met with about 50 strains. Some of these strains contain only part of one O antigen—others contain a complete complement of O antigens and are indistinguishable from such *Salmonella* types as *onderstepoort*, *worthington*, or *carrau*. At one time only H antigens common to the non specific phase of diphasic *Salmonella* types were found, but Peluffo, Edwards and Bruner (1942) have now met with paracolon bacilli containing antigens, such as  $z_6$ , that are present in monophasic types. Four strains at least have been described that contain a Vi antigen apparently identical with that present in *Salm. typhi*, *Salm. paratyphi C*, and *Salm. ballerup*.

The fermentative behaviour of these coliform and paracolon bacilli varies considerably. Some of them form acid and gas in lactose within 24 hours, others not till after some days, some form acid only, and some fail to attack lactose altogether. Most of them fail to ferment dulcitol and tartrates, many attack sucrose or salicin or both; and a

few of them form indole or liquefy gelatin. None of them has yet been shown to be pathogenic to man but there is strong reason to believe that some are pathogenic to animals (Edwards, Cherry and Bruner 1943)

Of the Flexner dysentery bacilli, two strains were shown by agglutination, absorption, and production of antibody tests to contain O antigens VI and XIII, and 14 other strains were agglutinated by sera containing VI and XIII antibodies (Bornstein, Saphra and Daniels 1941)

Though Kauffmann (1941) has included five types of coliform and paracoliform bacilli in his diagnostic table under the name of *Salmonella coli* 1, 2, 3, 4, 5, there seems little justification for regarding these organisms as members of the *Salmonella* group. Our present knowledge suggests that *Salmonella* antigens are by no means uncommon in the *Bacterium* group. In our own laboratory we have met several such strains, and it may well be that, when they are looked for, they will be found to be widely distributed. As we have already pointed out it is doing violence to the principles of taxonomy to include every organism in the *Salmonella* group merely because it contains some antigenic fraction that has hitherto been regarded as peculiar to this group. The main interest of these organisms is that they serve as pitfalls for the unwary, and that unless a proper study of the cultural and biochemical behaviour of every strain suspected of being a *Salmonella* is made, in addition to a serological examination, they are liable to give rise to false positive results from time to time. Admittedly, there are intermediate strains such as those described by Edwards, Cherry and Bruner (1943), some of which appear to be pathogenic, which are at present very difficult to classify, but it will be wise for the moment to maintain a conservative attitude, and to include in the *Salmonella* group only those organisms possessing the general, as well as the antigenic characters, of accepted members of this group.

## REFERENCES

- ACHARD, C and BESSAUDE, R. (1896) *Bull. Mém. Soc. méd. Hôp.* 13, 820  
 ALNOV, L. (1943) *Bact. Rev.* 7, 43  
 ANDREWS, F. W. (1927) *J. Path. Bact.* 25, 505. (1925) *Ibid.* 28, 345  
 ANDREWS, F. W. and LEAVE, S. (1921) *Brit. J. exp. Path.* 2, 157  
 ANZAI, H. and TSUBUMI, H. (1940) *Kiassato Arch.* 17, 106  
 AOKI, K. and SAKAI, K. (1925) *Zbl. Bakt.* 95, 152  
 ARIMA, R. (1912) *Zbl. Bakt.* 63, 424  
 ATKINSON, N. (1943) *Aust. J. exp. Biol. med. Sci.* 21, 171  
 BAHR, L. (1930a) *Dietsch tierarztl. Wschr.* 38, 145. (1930b) *Ibid.* 38, 165  
 BARTEL, H. (1938) *Tierarztl. Rdsch.* 44, 601  
 BASENAU, F. (1894) *Arch. Hyg., Berl.* 20, 242  
 BASILEWSKY, B. G. and REMGILD, W. I. (1935) *Z. Immunforsch.* 85, 10  
 BEAUDETTE, F. R. (1926) *J. Amer. vet. med. Ass.* 68, 642  
 BENEDICT, R. G., MCCOY, E., and WISNICKY, W. (1941) *J. infect. Dis.* 69, 167  
 BERNARD, H. (1935-36) *Z. Hyg. Infekthkr.* 117, 352  
 BIRCH HIRSCHFELD, L. (1935) *Z. Hyg. Infekthkr.* 117, 626  
 BITTER, L., WEIGMANN, F., and HABS, H. (1926) *Munch. med. Wschr.* 73, 940  
 BOECKER, E. (1936) *Zbl. Bakt.* 135, 501  
 BOECKER, E. and KAUFFMANN, F. (1930) *Zbl. Bakt.* 116, 458  
 BOECKER, E. and SILBERSTEIN, W. (1932) *Zbl. Bakt.* 125, 237  
 BOVIN, A. and MESROBEANU, L. (1933) *C. R. Soc. Biol.* 112, 76. (1934a) *Ibid.* 115, 304, 309. (1934b) *Ibid.* 117, 273. (1934c) *C. R. Acad. Sci.* 198, 2211. (1935a) *C. R. Soc. Biol.* 118, 612. (1935b) *C. R. Acad. Sci.* 201, 168  
 BOVIN, A., MESROBEANU, I., and MESROBEANU, L. (1933a) *C. R. Soc. Biol.* 113, 490. (1933b) *Ibid.* 114, 307. (1934a) *Ibid.* 115, 306. (1934b) *Ibid.* 117, 271. (1935c) *Arch. roum. Path. exp. Microbiol.* 8, 45  
 BORMAN, E. K., WHEELER, K. M., WEST, D. E., and MICKLE, F. L. (1943) *Amer. J. publ. Hlth* 33, 127  
 BORNSTEIN, S. (1943) *J. Immunol.* 46, 439  
 BORNSTEIN, S. and SAPHRA, I. (1942) *J. infect. Dis.* 71, 55  
 BORNSTEIN, S., SAPHRA, I., and DANIELS, J. B. (1941) *J. Immunol.* 42, 401  
 BORNSTEIN, S., SAPHRA, I., and STRAUSS, L. (1941) *J. infect. Dis.* 69, 69  
 BOSWORTH, T. J. and LOVELL, R. (1931) *Univ. Camb. Inst. Animal Path.*, 2nd Ann. Rep.

- BOYCOTT J and McNEIL, J. W. (1925) *Lancet*, ii, 741.
- BRANHAM, S. E. (1925) *J. infect. Dis.*, 37, 291.
- BRIDGES, R. F. and D'WEAR, L. (1907) *J. R. Army med. Corps*, 67, 249.
- BRIDGES, R. F. and BOOTH, V. M. (1931) *J. R. Army med. Corps*, 53, 241, (1932) *Ibid.* 65, 221.
- BRIDGES, L. (1912) *Disch. med. Week.*, 23, 477.
- BROWN, A. and HAYES, H. (1912) *Week. med. Week.*, 43, 611.
- BROUEN, L. (1927) *C. R. Soc. Biol.*, 97, 140.
- BROWN, H. C., DUFFAN, J. T., and HENRY, T. A. (1924) *J. Hyg., Camb.*, 23, 1, (1925) *Lancet*, i, 117.
- BROWNING, C. H., GILMORE, W., and MACKIE, T. J. (1913) *J. Hyg., Camb.*, 12, 225.
- BRUYER, D. W. and EDWARDS, I. R. (1930) *J. Bact.*, 27, 365, (1930) *Agree. Exp. Biol.*, 1, 494, (1931a) *J. Bact.*, 42, 477, (1931b) *Amer. J. Hyg.*, 24, 52, (1932a) *Proc. Soc. exp. Biol.*, 5 Y., 50, 174, (1932) *J. Immunol.*, 44, 219.
- BRUYER, H. and GANTER, (1929) *Z. Hyg. Inf. Bakt.*, 90, 252.
- CALDWELL, M. E. and IERSON, D. L. (1929) *J. infect. Dis.*, 65, 242.
- CARFANO, M. (1913) *Zbl. Bakt.*, 70, 42.
- CAUFER, W. (1928-29) *Z. Hyg. Inf. Bakt.*, 109, 170.
- CERVALANT, C. and PROPERT, L. (1933) *C. R. Soc. Biol.*, 112, 579.
- CHENCHOURY, N., FILIPPOVIC, D., and STAVEL, J. (1933-37) *Zbl. Bakt.*, 123, 45.
- CHANTENET, A. (1907) *C. R. Soc. Biol.*, 43, 57, 191.
- CHEERY, W. B., EDWARDS, P. R., and BRUYER, D. W. (1942) *Proc. Soc. exp. Biol.*, 5 Y., 52, 125.
- CHEERY, W. B., SCHERAGO, M., and WEAVER, R. H. (1942) *Amer. J. Hyg.*, 37, 211.
- CLATCHEY, K. V. (1931) *Elm. Week.*, 10, 549, (1932) *Zbl. Bakt.*, 124, 22.
- COMBES, D., STAMATESCO, S., and SORE, E. (1930) *Arch. roum. Path. exp. Microbiol.*, 2, 189.
- CONRAD, H. (1903) *Disch. med. Week.*, 23, 26, (1905) *Ibid.*, 22, 58.
- COSTEY, E. T. and THORNTON, L. H. D. (1932) *Exp. publ. Hlth. med. Sci., Min. Hlth., Lond.*, No. 82.
- CRAIGIE, J. (1912) *Canad. publ. Hlth. J.*, 23, 41.
- CRAIGIE, J. and BRADY, K. P. (1906a) *J. Path. Bact.*, 43, 223, (1906b) *Ibid.*, 42, 241.
- CRAIGIE, J. and VAN C. H. (1926) *Canad. publ. Hlth. J.*, 23, 449, 454.
- CRICKER, J. C. (1929) *J. Hyg., Camb.*, 23, 224.
- DALLING, T., MASON, J. H., and GORDON, W. E. (1928) *Vol. Rev.*, 2, 229.
- DALLING, T. and WARRACK, G. K. (1922) *J. Path. Bact.*, 25, 655.
- DANMANN, S. and STEDEBERG, (1910) *Arch. roum. publ. Microbiol.*, 2, 432.
- DANIEL, J. (1900) *Ann. Inst. Pasteur*, 14, 183.
- DELAFIELD, M. E. (1931) *J. Path. Bact.*, 34, 177, (1932) *Ibid.*, 25, 53, (1934) *Brit. J. exp. Path.*, 15, 139.
- DOEREN, C. A. VAN (1927) *Disch. med. Week.*, 45, 147.
- DOUGLAS, S. R. (1921) *Brit. J. exp. Path.*, 2, 175.
- DOYLE, T. M. (1927) *J. comp. Path.*, 40, 71.
- DUDGEON, L. S. and CROCHANT, A. L. (1929) *Lancet*, ii, 15.
- ECKER, E. E. (1917) *J. infect. Dis.*, 21, 541.
- ECKER, E. E. and RICHARDSON, M. L. (1925) *J. infect. Dis.*, 37, 523.
- ECKER, E. E. and RICHARDSON, C. (1928) *J. Hyg., Camb.*, 27, 44.
- EDWARDS, P. R. (1929) *J. infect. Dis.*, 45, 191, (1930) *J. Bact.*, 30, 465, (1931) *Ibid.*, 22, 259, (1932) *J. Hyg., Camb.*, 27, 261, (1933) *J. Hyg., Camb.*, 28, 306, (1937) *Proc. 7th World's Poultry Congr.*, p. 271, (1947) *Proc. comm.*
- EDWARDS, P. R. and BRUYER, D. W. (1931) *J. Hyg., Camb.*, 23, 716, (1932) *J. Bact.*, 23, 63, (1933a) *Agree. Exp. Biol.*, 1, 494, (1933b) *J. infect. Dis.*, 63, 218, (1934a) *J. infect. Dis.*, 63, 229, (1934b) *Amer. J. Hyg.*, 34, 50, 121, (1935) *Proc. Soc. exp. Biol.*, 5 Y., 43, 240, (1936a) *J. Bact.*, 44, 249, (1936b) *J. Immunol.*, 44, 219, (1937a) *Agree. exp. Biol.*, 1, 494, (1937b) *Canad. J. Zool.*, 15, 121, (1938) *J. infect. Dis.*, 72, 58.
- EDWARDS, P. R., BRUYER, D. W., and HINCHAW, W. R. (1941) *J. infect. Dis.*, 69, 17.
- EDWARDS, P. R., BRUYER, D. W., and BUTCH, H. L. (1947) *Proc. Soc. exp. Biol.*, 5 Y., 44, 230.
- EDWARDS, P. R., CHEERY, W. B., and BRUYER, D. W. (1943) *J. infect. Dis.*, 72, 229.
- EDWARDS, P. R. and HUGHES, H. (1944) *Proc. Soc. exp. Biol.*, 5 Y., 53, 23.
- EDWARDS, P. R. and PETTIGREW, L. F. (1927) *J. Bact.*, 12, 73.
- FELIX, A. (1943) *Brit. med. J.*, i, 435.
- FELIX, A. and CALLOW, E. R. (1943) *Brit. med. J.*, ii, 157.
- FELIX, A. and PITT, R. M. (1934a) *J. Path. Bact.*, 28, 479, (1934b) *Lancet*, ii, 159.
- FERRACOLA, R., MONTVERDE, J. J. and LINDGARD, R. H. (1943) *Bol. Ofras. sanit. Narda.*, Arg. No. 74, p. 103.
- FITCH, C. P. and BILLINGS, W. A. (1920) *J. Bact.*, 5, 479.

- FLETCHER, W (1918) *Lancet*, ii 102  
 FREEMAN, G G (1943) *Biochem. J.* 37, 601  
 FREEMAN, G G, CHALLINOR, S W, and WILSON, J (1940) *Biochem. J.* 34, 307  
 FURTH, J. and LANDSTEINER, K (1929) *J exp Med.* 47, 171, (1929) *Ibid.* 49, 727  
 GAERTNER, (1888) *KorrespBl arztl Ver Thüringen* 17, 573  
 GAGE, G E and MARTIN, J F. (1916) *J med Res.* 34, 149  
 GARD, S (1937) *Z Hyg InfektKr.* 120, 59, (1938) *Ibid.* 121, 139  
 GARD, S and ERIKSSON, E J (1939) *Z Hyg InfektKr.* 122, 54  
 GAT, F P. and CLAYPOLE, E J (1913) *Arch intern Med.* 12, 621  
 GIGLIOLI, G (1930) *J Hyg. Camb.* 29, 273  
 GLÄSSER, K (1909) *Deutsch. tierarztl Wochr.* 18, 513, (1910) *Zbl Bakt Ref.* 45, 612  
 GOOD, E S and CORBETT, L S (1913) *J. infect Dis.* 13, 53  
 GOULDER, N E, KINGSLAND, M P, and JANEWAY, C. A. (1912) *New Engl J Med.* 226, 127  
 GRAY, I D A (1931) *J Path Bact.* 34, 330  
 GUTH, F (1916) *Zbl Bakt.* 77, 487  
 GUTHRIE, h J and MONTGOMERY, G L. (1939) *J Path Bact.* 49, 393  
 GWYN, N B (1898) *Johns Hopk Hosp Bull* 9, 54  
 HABS, H (1933) *Zbl Bakt.* 130, 367  
 HABS, H and ARJONA, E (1935) *Zbl Bakt.* 133, 204  
 HADLEY, P B, CALDWELL, D W, FLEKINS, M W, and LAMBERT, D J (1917) *R I agric Exp Sta Bull.* No 172  
 HARVEY, A M (1937) *Arch intern Med.* 59, 118  
 HATTA, S (1938) *Jap J exp med.* 18, 201  
 HAVENS, L C. and MAYFIELD, C R (1933) *J infect Dis.* 52, 157  
 HEELSBERGEN, T VAN (1914) *Zbl Bakt.* 72, 35  
 HENNINO, M W (1936) *J Hyg. Camb.* 36, 525, (1937) *Ibid* 37, 581  
 HICKS, E P (1930) *J Hyg. Camb.* 29, 446  
 HINSHAW, W. R., BROWNE, A S, and TAYLOR, T J (1943) *J infect Dis.* 72, 197  
 HIRSCHFELD, L. (1919) *Lancet*, i 296  
 HOFFMAN, W. and PICKER, M (1904) *Hyg Rundsch.* 14, 1  
 HOHN, J (1940) *Zbl Bakt.* 146, 215  
 HOHN, J and HERRMANN, W. (1935a) *Zbl Bakt.* 133, 183, (1935b) *Ibid.*, 134, 277, (1936a) *Zbl Bakt.* 135, 605, (1936b) *Z Hyg InfektKr.* 117, 722, (1936c) *Ibid.* 118, 656, (1940) *Zbl Bakt.* 145, 209  
 HORMAECHER, E and PELUFFO, C A (1936) *Arch urug Med.* 9, 673, (1939) *Ibid.* 14, 217  
 HORMAECHER, E, PELUFFO, C. A, and ALEFFO, P L (1936) *Arch. urug Med* 9, 113, (1940) *Arch Pediat Urug.* 11, 8, (1941) *Arch urug Med.* 19, 125  
 HORMAECHER, E, PELUFFO, C A and PEREYRA, V R. DE (1944) *J Bact.* 47, 323  
 HORMAECHER, E, PELUFFO, C A, and SALSAMENDI, R (1938) *Arch urug Med.* 12, 377  
 HORMAECHER, E and SALSAMENDI, R (1936) *Arch urug Med.* 9, 665, (1939) *Ibid.* 14, 375  
 HYNES, M (1942) *J Path Bact.* 54, 193  
 IWASCHETZOFF, G (1926) *Arch Schiffs u Tropenhyg.* 30, 1  
 JACOBSEN, K A (1910) *Zbl Bakt.* 58, 208  
 JANSEN, J (1937) *Bull. Hyg., Lond* 12, 139  
 JONES, E R (1938) *J Path Bact.* 42, 455  
 JONG, D A DE (1913) *Zbl Bakt.* 67, 148  
 JORDAN, E O (1934) *Amer J trop Med.* 14, 27  
 JUNGHERR, E and CLANCY, C F (1939) *J infect Dis.* 64, 1  
 KAUFFMANN, F (1929a) *Zbl Bakt. Ref.* 94, 282, (1929b) *Z Hyg InfektKr.* 110, 537, (1929c) *Ibid.* 109, 427, 491, (1930a) *Ibid.* 111, 210, (1930b) *Ibid.* 111, 221, 233 247, (1931) *Zbl ges Hyg.* 25, 273 (1934a) *Ergebn Hyg Bakt.* 15, 219, (1934b) *Zbl Bakt.* 132, 337, (1934c) *Z Hyg InfektKr.* 128, 368, (1935a) *Ibid* 128, 617, (1935b) *Ibid.* 127, 431, (1936a) *Ibid.* 117, 778, (1936b) *Ibid.* 118, 540, (1936c) *Ibid.* 119, 103, (1937a) *Z Hyg InfektKr.* 119, 356, (1937b) *Ibid.* 120, 177, (1939a) *Acta path microbiol scand.* 16, 347, (1939b) *Ibid.* 18, 278, (1940a) *Ibid.* 17, 1, (1940b) *Ibid.* 17, 135, (1940c) *Ibid.* 17, 189, (1940d) *Ibid.* 17, 429, (1941) *Die Bakteriologie der Salmonella Gruppe* "Einar Munksgaard, Copenhagen.  
 KAUFFMANN, F and BURÓN, F A (1935) *Z Hyg InfektKr.* 117, 650  
 KAUFFMANN, F and HENNINGSSEN, E J (1938) *Z Hyg InfektKr.* 120, 640, (1939) *Acta path microbiol scand.* 18, 99  
 KAUFFMANN, F and MITSUI, C (1930) *Z Hyg InfektKr.* 111, 749  
 KAUFFMANN, F and MÖLLER, E (1940) *J Hyg. Camb.* 40, 246  
 KAUFFMANN, F. and SILBERSTEIN, W (1934) *Zbl Bakt.* 132, 431.  
 KAUFFMANN, F and TESDAL, M (1937-38) *Z Hyg InfektKr.* 120, 168  
 KHALIL, A M. (1939) *J Hyg. Camb.* 33, 75  
 KILBORN, F L (1893) *Misc Infect. parasit Dis.* 8<sup>o</sup> Washington, 49

- KJSDA, S. (1940) *Kitasato Arch.* 17, 1.
- KLEIN, E. (1889) *Zbl. Bakt.*, 5, 689.
- KNOX, M. (1931) *Arch. Hyg., Brit.*, 105, 237.
- KRISTENSEN, M. and BOJLEN, K. (1929) *Zbl. Bakt.*, 114, 86; (1936) *Z. Hyg. InfektKr.*, 120, 177.
- KRISTENSEN, M. and KAUFFMANN, F. (1937) *Z. Hyg. InfektKr.*, 120, 149.
- KRUMWIEDE, C. and KOHN, L. A. (1917) *J. med. Res.*, 38, 509.
- KRUMWIEDE, C. and PRATT, J. S. (1914) *J. exp. Med.*, 19, 501.
- KÜHNEMANN, G. (1911) *Zbl. Bakt.*, 57, 497.
- LEITSON, E. (1933) *J. Path. Bact.*, 40, 531; (1936) *Amer. J. Hyg.*, 24, 423.
- LEYL, O. and TIETZ, J. (1903) *Müsch. med. Wochr.*, 50, 2139; (1905) *Klin. Wb.*, 14, 493.
- LEON, A. P. (1942) *Rev. Inst. Salub. Enferm. trop.*, 3, 273.
- LESBOUYRIES and VERGE. (1932) *Bull. Acad. Vét. France*, 5, 294.
- LESLIE, P. H. (1942) *J. Hyg., Camb.*, 42, 552.
- LESLIE, P. H. and SHYRA, A. G. (1931) *J. Path. Bact.*, 34, 533.
- LOEFFLER, F. (1892) *Zbl. Bakt.*, 11, 129; (1903) *Discr. med. Wochr.*, 29, 36; (1906) *Ibid.*, 32, 269.
- LONGFELLOW, D. and LUFFFOLD, G. F. (1943) *Amer. J. Hyg.*, 37, 206.
- LOVELL, R. (1931) *J. Path. Bact.*, 34, 13; (1932a) *Bull. Hyg.*, 7, 403; (1932b) *Vet. Rec.*, 12, 1032.
- LÜTJE (1937) *Disch. tierärztl. Wochr.*, 45, 242; (1939) *Ibid.*, 47, 227, 246, 257.
- MACADAM, W. (1919) *Lancet*, ii, 189.
- MACFADYEN, A. and ROWLAND, S. (1901) *Zbl. Bakt.*, 30, 753; (1903) *Proc. roy. Soc.*, 71, 77.
- MACKE, F. P. and BOWEN, C. J. (1919) *J. R. Army med. Cps.*, 33, 154.
- McKINLAY, B. (1937) *Amer. J. Dis. Child.*, 54, 1252.
- MALLMANN, W. L., RIFF, J. F., and MATTHEWS, E. (1942) *J. infect. Dis.*, 70, 253.
- MANDELBAUM. (1932) *Müsch. med. Wochr.*, 79, 1566.
- MARRASSINI, A. (1913) *Zbl. Bakt.*, 71, 113.
- MARTIN, A. R. (1934) *Brit. J. exp. Path.*, 15, 137.
- MELTON, R. R. and JOST, E. L. (1926) *J. Immunol.* 12, 331.
- MENTEN, M. L. and KING, C. G. (1930) *J. infect. Dis.*, 48, 275.
- METCHNIKOFF, E. and BESREDEA, A. (1911) *Ann. Inst. Pasteur*, 25, 193.
- MEYER, K. and BERGELL. (1907) *Berl. Klin. Wochr.*, 54, 568.
- MEYER, K. F. and BOERNER, F. (1913) *J. med. Res.*, 29, 325.
- MEYER, K. F. and MATSUMURA, K. (1927) *J. infect. Dis.*, 41, 395.
- MONTVERDE, J. J. (1942) *Nature*, 149, 472.
- MONTVERDE, J. J. and LKIGIARDA, R. H. (1944) *Bol. Obr. sanit. Nac.*, 8, 168.
- MOORE, V. A. (1895) *U.S. Dept. Agri. Bur. Animal Ind.*, 8, 71.
- MOROS, W. T. J. and PARTRIDGE, S. M. (1940) *Biochem. J.*, 34, 169; (1942) *Brit. J. exp. Path.*, 23, 151.
- MORRIS, J. F., BARNES, C. G., and SELLERS, T. F. (1943) *Amer. J. publ. Hlth.*, 33, 246.
- MORRIS, J. F., BRIM, A. and SELLERS, T. F. (1944) *Amer. J. publ. Hlth.*, 34, 1277.
- MORRIS, J. F., SELLERS, T. F., and BROWN, A. W. (1941) *J. infect. Dis.*, 63, 117.
- MÜLLER, L. (1923) *C. R. Soc. Biol.*, 89, 434.
- MÜLLER, M. (1912) *Zbl. Bakt.*, 62, 335.
- MÜLLER, R. (1910) *Disch. med. Wochr.*, 36, 2357; (1933) *Müsch. med. Wochr.*, 80, 1771.
- MULSOW, F. W. (1919) *J. infect. Dis.*, 25, 135.
- MURRAY, C. (1919) *J. infect. Dis.*, 25, 341.
- NEUKIRCH, P. (1916) *Z. Hyg. InfektKr.*, 85, 103.
- NOBLE, DE. (1898) *Ann. Soc. Méd. Gand*, 72, 281.
- NOCARD. (1893) *Cons. Hyg. Publ. Sal. Dept. Scine. Séance. Mar. 24.*
- OLITSKI, L. (1942) *J. Hyg., Camb.*, 42, 547.
- ORSKOV, J., JENSEN, K., and KOBAYASHI, K. (1928) *Z. Immunforsch.*, 55, 31.
- ORSKOV, J. and LASSEN, H. C. A. (1930) *Z. Immunforsch.*, 67, 137.
- ORSKOV, J. and MOLTRE, O. (1928) *Z. Immunforsch.*, 59, 357.
- PECKHAM, C. F. (1923) *J. Hyg., Camb.*, 22, 69.
- PELUFFO, C. A., EDWARDS, P. R., and BRUNER, D. W. (1942) *J. infect. Dis.*, 70, 155.
- PFLEIFER, R. (1894) *Disch. med. Wochr.*, 20, 898.
- PFLEIFER, W. and REHSE, A. (1913) *Zbl. Bakt.*, 63, 174.
- PFLEIFER, W. and ROEPKE, E. (1917) *Zbl. Bakt.*, 79, 123.
- PIDDER, A. (1938) *J. Path. Bact.*, 47, 1; (1940) *J. biol. photogr. Ass.*, 8, 153.
- PIROSKY, I. (1935) *C. R. Soc. Biol.*, 128, 347.
- POLLOCK, M. R., KNOX, R., and GRILL, P. G. H. (1942) *Nature*, 150, 94.
- RAISTRICK, H. and TOPLEY, W. W. C. (1934) *Brit. J. exp. Path.*, 15, 113.
- RAUSS, K. (1939a) *Z. Immunforsch.*, 95, 489; (1939b) *Ibid.*, 97, 281; (1941) *Zbl. Bakt.*, 147, 253; (1943) *Z. Immunforsch.*, 103, 220.
- Report (1934) Salmonella Subcommittee of Nomenclature Comm. int. Soc. Microbiol. *J. Hyg., Camb.*, 34, 333.

- RETTGER, L. F. (1900) *N. Y. med. J.*, 71, 803. (1909) *J. med. Res.*, 21, 115  
 RETTGER, L. F. and HARVEY, S. G. (1908) *J. med. Res.*, 18, 277  
 RETTGER, L. F. and KOSER, S. A. (1917) *J. med. Res.*, 35, 443  
 RETTGER, L. F., PLASTRIDGE, W. N., and CAMERON, R. (1933) *J. infect. Dis.*, 53, 272  
 RETTGER, L. F. and SCOVILLE, M. M. (1919) *Abstr. Bact.*, 3, 8. (1920) *J. infect. Dis.*, 28, 217  
 ROTH, E. (1903) *Hyg. Rundsch.*, 13, 489  
 ROUCHDI, M. (1938) *C. R. Soc. Biol.*, 128, 1022  
 RUBIN, H. L. (1940) *J. Bact.*, 40, 463  
 RUBIN, H. L., SCHERAGO, M., and WEAVER, R. H. (1942) *Amer. J. Hyg.*, 36, 43  
 SALMON, E. and SMITH, T. (1880) *Ann. Rep. Bureau Animal Industry*, (1886) *Amer. mon. micr. J.*, 7, 204  
 SANARELLI, J. (1894) *Ann. Inst. Pasteur*, 8, 193, 303  
 SAPHRA, I. and SILBERBERG, M. (1942) *J. Immunol.*, 44, 129  
 SAVAGE, W. G. and WHITE, P. B. (1920) *Spec. Rep. Ser. med. Res. Coun., Lond.*, No. 91  
 SAVINO, E. and MENÉNDEZ, P. E. (1934) *Rev. Inst. Bact.*, 6, 347  
 SCHÄFER, W. (1934-35) *Zbl. Bakt.*, 133, 458  
 SCHERMER and EHRLICH (1921) *Berl. tierarztl. Wochr.*, 37, 469  
 SCHIFF, F. (1938) *J. Amer. med. Ass.*, 111, 2458  
 SCHIFF, F., BORNSTEIN, S., and SAPHRA, I. (1941) *J. Immunol.*, 40, 360  
 SCHIFF, F. and SAPHRA, I. (1940) *J. infect. Dis.*, 66, 97. (1941) *Ibid.*, 68, 123  
 SCHIFF, F. and STRAUSS, L. (1939a) *J. infect. Dis.*, 65, 120. (1939b) *Ibid.*, 65, 160  
 SCHOLTENS, R. T. (1937) *J. Hyg., Camb.*, 37, 315  
 SCHOTTMÜLLER, H. (1900) *Dtsch. med. Wochr.*, 26, 511. (1901) *Z. Hyg. Infektkr.*, 36, 368  
 SCHÜTTE, H. (1920) *Lancet*, i, 93. (1928) *Arch. Hyg., Berl.*, 100, 181. (1930) *Brit. J. exp. Path.*, 11, 34  
 SCHWABACHER, H., TAYLOR, J. and WHITE, M. H. G. (1943) *Brit. med. J.*, ii, 308  
 SCOTT, W. M. (1926) *J. Hyg., Camb.*, 25, 398. (1932) *J. Path. Bact.*, 35, 655. (1933) *Bull. Office int. Hyg.*, 25, fasc. 5. (1940) *Proc. R. Soc. Med.*, 33, 366  
 SEIFFERT, G., JAHNCKE, A., and ARNOLD, A. (1928) *Zbl. Bakt.*, 109, 193  
 SELIGMANN, E. and CLAUBERG, K. W. (1932) *Zbl. Bakt.*, 125, 266  
 SELIGMANN, E. and SAPHRA, I. (1943) *Amer. J. Hyg.*, 38, 223  
 SELIGMANN, E., SAPHRA, I., and WASSERMANN, M. (1944) *Amer. J. Hyg.*, 40, 227  
 SHIMDSU, A. (1913) *Zbl. Bakt.*, 71, 338  
 SILBERSTEIN, W. (1931) *Zbl. Bakt.*, 122, Beiheft 131. (1932) *Z. Hyg. Infektkr.*, 114, 124  
 SIDMONS, J. S. (1926) *J. infect. Dis.*, 39, 209  
 SLADEN, A. F. and SCOTT, W. M. (1917) *J. Hyg., Camb.*, 26, 111  
 SMITH, J. (1934) *J. Hyg., Camb.*, 34, 301  
 SMITH, J. and KAUFFMANN, F. (1940) *J. Hyg., Camb.*, 40, 122  
 SMITH, J. and SCOTT, W. M. (1930) *J. Hyg., Camb.*, 30, 32  
 SMITH, T. (1893) *Misc. Invest. infect. parant. Dis. Dom. Animals*, 8<sup>th</sup> Washington, 53  
 SMITH, T. and TENNENBERG, C. (1915) *J. med. Res.*, 31, 503  
 ST. JOHN BROOKS, R. and RHODES, M. (1923) *J. Path. Bact.*, 26, 433  
 STERN, W. (1916) *Zbl. Bakt.*, 78, 481  
 STRÖMAN, R. and ÖRN, G. (1932) *Zbl. Bakt.*, 126, 340  
 TAYLOR, J., EDWARD, D. G. H., and EDWARDS, P. R. (1945) *Brit. med. J.*, i, 368  
 TESDAI, M. (1936) *Z. Hyg. Infektkr.*, 118, 533. (1937) *Ibid.*, 119, 28. (1938) *Die Salmonellagruppe mit besonderer Berücksichtigung der Salmonellinfektionen in Norwegen.* Olaf Norli, Oslo  
 TOPLEY, W. W. C. and AYRTON, J. (1924) *J. Hyg., Camb.*, 23, 198  
 TULLOCH, W. J. (1939) *J. Hyg., Camb.*, 39, 324  
 VARELA, G. and ZOLAYA, J. (1942) *Bull. Hyg., Lond.*, 17, 721  
 VARELA, G., ZOLAYA, J. and OLARTE, J. (1943) *Bull. Hyg., Lond.*, 18, 411  
 WARREN, S. H. and SCOTT, W. M. (1930) *J. Hyg., Camb.*, 29, 415  
 WEIGMANN, F. (1925a) *Zbl. Bakt.*, 95, 396. (1925b) *Ibid.*, 97, Beiheft. 299  
 WEIL, E. (1917) *Wien klin. Wochr.*, 30, 1001  
 WEIL, E. and SAXL, P. (1917) *Wien klin. Wochr.*, 30, 619  
 WHEELER, K. M. and BORMAN, E. K. (1943) *J. Bact.*, 46, 481  
 WHEELER, K. M., STUART, C. A., RUSTIGIAN, R., and BORMAN, E. K. (1943) *J. Immunol.*, 47, 59  
 WHEAT, W. B. (1908) *J. infect. Dis.*, 5, 519  
 WHITE, P. B. (1926) *Spec. Rep. Ser. med. Res. Coun., Lond.*, No. 103. (1929a) *J. Path. Bact.*, 32, 85. (1929b) *Med. Res. Coun., 'System of Bacteriology.'*, 4, 80. (1930) *J. Hyg., Camb.*, 29, 443. (1931) *J. Path. Bact.*, 34, 325. (1932) *Ibid.*, 35, 77  
 WILCOX, K. S. and LENOX, E. H. (1944) *J. Immunol.*, 49, 71  
 WILSON, W. J. (1938) *J. Hyg., Camb.*, 38, 507  
 WINSLOW, C. E. A., KLIGLER, I. J., and ROTHBERG, W. (1919) *J. Bact.*, 4, 429  
 WYLLIE, J. (1943) *Canad. publ. Hlth J.*, 34, 82  
 YAMANOUCHI, T. (1909) *C.R. Soc. Biol.*, 86, 1050

## CHAPTER 31

### LACTOBACILLUS

#### DEFINITION — *Lactobacillus*

Rods often long and slender, sometimes pleomorphic. Gram positive. Non motile and non sporing. Growth on surface media poor. Some members grow best at 40°–44° C. Usually produce acid from carbohydrates, as a rule lactic, some members form gas in addition. Little or no proteolytic activity. Most members are microaerophilic and facultative anaerobes, some members are strict anaerobes. Readily destroyed by heat, but unusually resistant to acid. Not known to be pathogenic to man or animals. Widely distributed in fermenting vegetable and animal products.

Type species is *Lactobacillus caucasicus*, Beijerinck

HISTORY — The first organism of this group was isolated by Kern in 1881 from the fermented milk of the Caucasus, known as *Kefir*. The name he gave it was *Dispora Kaulasica* but later it was called *Bacillus Kaukasicus*, and is now known as *Lactobacillus caucasicus*. As Kern did not give a complete description of the organism, and as it is impossible to be certain of its identity, it is somewhat unfortunate that it has been adopted as the type species. A similar bacillus was observed by Döderlein in 1892 in the acid vaginal secretion of pregnant women; this is usually known as Döderlein's bacillus, but is also known as *B. vaginalis* and *B. crassus*. Slender Gram positive bacilli were observed microscopically in the stomach contents of patients with gastric carcinoma by Oppler in 1895, working in the clinic of Dr. Boas at Berlin; this organism, which was not cultivated, is generally known as the Boas-Oppler bacillus. In 1900 Moro (1900a, b) cultivated a similar bacillus from the faeces of breast fed infants; this organism he called *B. acidophilus*, his findings were confirmed in the same year by Finkelstein (1900). Tissier, also in 1900, isolated two new organisms of the same group from the faeces of infants, to which he gave the names *B. bifidus* and *B. exilis*. In 1905 Grigoroff, working in Massol's laboratory, isolated from *Kisselo-mleko*, better known as *Ioghurt*, the fermented milk of Bulgaria, three organisms to which he gave the names A, B, and C; the first of these is now known as *Lactobacillus bulgaricus* or Massol's bacillus. Similar bacilli have been found by other workers in a number of fermented milks, chiefly the Armenian *Marun*, the Sardinian *Gioddu*, and the Egyptian *Leben raib* (Cohendy 1906, White and Avery 1910); they have also been isolated from ordinary market milk and from human milk (Moro 1900a; Heinemann and Hefferan 1909, Sherman and Stark 1927). Bacilli of this group have been cultivated by Mereshkowsky and his pupils (Mereshkowsky 1905, 1906, Petrow 1907) from the faeces of a large series of invertebrates, fishes and mammals, by Heinemann and Hefferan (1909) from human saliva and gastric juice, from soil, and from a number of different foods, such as

bran, silage, cornmeal, and olive-juice; by McIntosh, James and Lazarus Barlow (1922, 1924) from carious teeth, and by Kendall (1910) from sewage. Eggerth and Gagnon (1933) and Eggerth (1935) drew attention to the presence in normal adult faeces of Gram positive and Gram negative anaerobic bacilli, which they described under the generic name of *Bacteroides*. There seems little doubt that the Gram positive members are closely related to the lactobacilli and they will therefore be described in this chapter.

**Morphology.**—The members of this group are in general fairly large, non motile, non sporing, Gram positive bacilli. They are arranged singly, in pairs end to end, in chains, and sometimes in palisades (Fig 151). Some members are markedly pleomorphic, especially in old cultures, forming clubbed, knobbed, curled, spiral, candle-flame, vacuolated, whorled, and filamentous forms, and frequently showing irregular, granular, or beaded staining. In some species the bacilli tend to be arranged at angles to each other, giving rise to Y forms, which may simulate true branching. Another characteristic of some members is the formation of lateral offshoots or buds, either directly adherent to the parent cell, or connected with it by a short stem, these buds may themselves be bifid.

**Cultivation.**—These bacteria do not as a rule grow well on the usual laboratory media, their growth is much improved by the addition of whey or glucose. Surface colonies show a good deal of variation, but on the whole conform to one or other of the two types described by Vereshkowsky (1905, 1906) (1) round or navicular, pinhead in size, opaque, whitish, and surrounded by an areola of turbid agar, (2) round or irregularly round, less than pinhead in size, greyish, translucent colonies with a finely erose edge, and with no areola around them, microscopically these colonies are of typically rhizoid structure. Deep colonies in glucose agar likewise tend to be either compact, with an entire edge or sometimes a single lateral knob (Rettger and Horton's (1914) Y type), or curled, rhizoid, and feathery, looking like a tuft of hair or moss (Rettger and Horton's X type). Intermediate types of colony are not uncommon. The compact and feathery types of colony are referred to by some writers as "smooth" and "rough" respectively, but since the particular type of colony formed seems to depend largely on environmental conditions, and since there seems to be little relationship between the colonial type and any other important characteristic of the organisms, it is probably wiser to refrain from the use of terms that have now come to possess a wider connotation. A very characteristic appearance is the turbidity or milkiness of the agar produced in shake plates or tubes, it is a variable characteristic, however,



FIG 151.—Döderlein's bacillus  
From an agar culture, 48 hours, 37° C ( $\times 1000$ )



and has little or no differentiating value amongst the members of the group. Growth in gelatin is either poor or absent. Liquefaction never occurs.

**Resistance and Methods of Isolation from Natural Sources.**—The organisms are not particularly resistant to heat and are generally destroyed by an exposure to 60° C for half an hour. One of their most striking features which gives to them the names acidophilic, acid resisting or aciduric is their ability to survive in concentrations of acid that usually prove fatal to other non-sporing bacteria. It is this characteristic that is generally made use of in their isolation.

One of the most successful methods is to incubate the material for 1 to 3 days in 0.5 per cent acetic acid broth and subsequently to plate on 2 per cent glucose agar. McIntosh and his co-workers (1922) recommend incubating the material in broth of pH 3.5 for 24 hours, then subculturing into a series of broth tubes varying in pH from 3.0 to 4.5 and plating after a further 24 hours. Kendall (1910) made three consecutive subcultures in acid broth and plated the last on dextrose agar containing 0.2 per cent sodium oleate which is said to improve the growth. The acid may be added to the tubes directly or the organisms may be seeded into a medium containing a fermentable carbohydrate; the acid produced in this medium is usually sufficient to kill off most other micro-organisms. Cruickshank (1926) recommends for the isolation of *L. bifidus* inoculating the faeces into a deep tube containing 20 ml. of 1 per cent glucose or lactose broth together with a small piece of fresh sterile rabbit kidney; the kidney is added to promote anaerobiosis but does not appear to be essential. The medium is covered with a vaseline seal. The culture is incubated for about a week at 37° C and then plated on to 1 per cent glucose agar or Loeffler's serum which is incubated aerobically and anaerobically. After 48 hours on the anaerobic plates greyish pinhead colonies appear. The only other organism that is likely to develop under these conditions is the enterococcus, which forms larger whitish colonies. Another method that may be employed for the isolation of the acid-resistant bacteria is the use of Veillon tubes (Veillon and Zuber 1893; Rettger and Cheplin 1921) containing 2 per cent glucose agar or whey agar; the different organisms adapt themselves to the varying oxygen pressure in the medium and form characteristic colonies which may be picked off with ease. The tomato broth medium described by Hulp (1927) has given very favourable results. If the primary cultures are plated on tomato agar single colony isolations are frequently successful.

In our experience (Crowley *et al.* 1941) direct plating of consecutive dilutions on the glucose liver blood agar medium recommended by Eggerth and Gagnon (1933) has proved satisfactory for the isolation of lactobacilli from faeces. Lewis, Bedell and Rettger (1940) have described a glucose cysteine agar medium which likewise seems well suited for this purpose. The presence of added CO<sub>2</sub> is beneficial for growth (Hulp 1926; Cruickshank 1934; Lewis *et al.* 1940).

**Metabolism.**—Most of the members are microaerophilic or facultative anaerobes. For their isolation fairly strict anaerobic conditions are often though not always necessary. After a few subcultures many of them can be brought to grow aerobically but others remain persistently obligate anaerobes. Peroxide is produced without or with only very small quantities of catalase. This probably accounts for the poor viability of the organisms in media not maintaining a low oxidation-reduction potential. Gillespie and Rettger (1933) noted a difference between different species in the intensity of their reducing activity. Oral lactobacilli for example were found in a particular medium to reduce the Eh to about -240 mv whereas *L. acidophilus* of intestinal origin, *L. bifidus*, Döderlein's bacillus and *L. bulgaricus* reduced it to only about -140 mv. According to Curran, Rogers and Whittier (1933) the optimum growth temperature is 37°-40° C; the range

of growth with most strains is 25°–46° C, but certain strains particularly those of the cusei type, may grow even at 10° C. Though some of the earlier workers (Rodella 1901) stated that an alkaline was preferable to an acid medium later workers (Morishita 1929, Weiss and Rettger 1931, Longworth and MacInnes 1935) have found that both growth and acid production occur best in the neighbourhood of pH 5.0, the optimal range being about pH 5.4–6.8. Pantothenic acid, riboflavin and pyridoxin appear to be important accessory growth factors (Snell, Strong and Peterson 1939). No pigment is produced but deep colonies in glucose agar often develop a brownish centre, and the agar itself is frequently clouded. Haemolysin production is variable and has not been studied fully. No toxins are formed. The organisms have very little effect on proteins and growth on protein media without carbohydrates is very poor. Peterson, Pruess and Fred (1928) have found that they do possess some proteolytic action as judged by the quantitative estimation of non protein and amino-acid nitrogen but according to Kendall and Haner (1924a, b) this is very slight, no indole, scatole or histamine is formed.

On the other hand they are very active in fermenting carbohydrates. The acid produced from lactose is partly fixed, consisting of *laevo* or *dextro* rotatory or in active lactic acid, and partly volatile, consisting of formic, acetic, and butyric acids in the ratio of 6 : 3 : 1 (Curran, Rogers and Whittier 1933). The proportion of volatile to fixed acids varies with different strains from about 4 : 20 per cent. According to Barker and Haas (1944) however, the intestinal members of the group do not produce lactic acid from lactate but volatile fatty acids. This type of butyric acid fermentation differs from others in that no appreciable amount of molecular hydrogen and very little CO<sub>2</sub> are formed. Malic acid is said to be produced by *L. odontolyticus* in greater quantity than lactic acid (McIntosh *et al* 1924). Gas production is not detectable by the ordinary Durham fermentation tube, except with *L. acidophil-aerogenes* (Torrey and Rahe 1915) which produces 4–6 volumes of H<sub>2</sub> to 1 of CO<sub>2</sub>. Curran, Rogers and Whittier (1933) however, have shown that most strains produce small quantities of gas from fermentable carbohydrates. The formation of lactic acid from glucose does not require the presence of oxygen or lead to the production of CO<sub>2</sub>. Consequently CO<sub>2</sub> is not a major product of fermentation with most strains. An exception to this rule is furnished by *L. pentoaceticus*, which is able to oxidize lactic to acetic acid (Hunt 1933). In this process one molecule of O<sub>2</sub> is used and one molecule of CO<sub>2</sub> produced. Hence CO<sub>2</sub> constitutes a more important product of fermentation with this organism than with the other members of the group. The usual products of fermentation, such as alcohol, acetone, acetylmethylcarbinol, and butylene glycol, are not formed (Bertrand and Duchacek 1909). For all practical purposes the organisms may be considered as of the obligatory saccharolytic type.

**Biochemical**—There is considerable variation in the sugars fermented. Glucose and lactose are fermented by practically all strains, maltose and sucrose by a high proportion, mannitol, salicin, and raffinose by a small proportion while dextrin, inulin, dulcitol, and starch are rarely fermented. Strains of *L. bifidus* are said, however, to ferment inulin (Weiss and Rettger 1934) and strains isolated from soil and grain are said not to ferment lactose (Hunt and Rettger 1930). Both these statements await confirmation. *L. pentoaceticus*, the organism described by Fred, Peterson, and Davenport (1919) from silage, sauerkraut, and manure, is peculiar in its ability to ferment xylose. According to Weinstein and Rettger (1932), it is

further distinguished from most of the other members of the group by its failure to ferment lactose or to curdle milk. Cruickshank (1934) says that Döderlein's bacillus ferments glycogen with the production of lactic acid in a few days, while strains of *acidophilus*, *odontolyticus*, and *bifidus* take 7-10 days to ferment it, *bulgaricus* does not ferment it at all. It is very doubtful whether the reactions of individual species and strains are constant, according to some authors they are not (McIntosh *et al* 1924, Day and Gibbs 1928). Most, but not all, the members produce acid in milk, often in sufficient quantity to precipitate the casein in the form of a loose clot, which does not contract and express whey, the litmus is frequently decolorized, especially in the lower part of the tube. The rate at which clotting is produced is of some slight differential significance. White and Avery (1910) divided the acid resistant organisms obtained from milk into two types, their Type A produced a large quantity of lactic acid in milk—27 to 37 per cent, their Type B produced a smaller quantity—12 to 16 per cent. The most active acid producer is *L. bulgaricus*, the least active *L. bifidus*, *L. acidophilus* occupies an intermediate position.

**Antigenic Structure**—The serological reactions of these organisms have been incompletely studied, and so far no satisfactory classification of the group has been possible by agglutination or absorption. Generally speaking, the members of a single species show a considerable amount of heterogeneity (Kendall and Haner 1924b, Lash and Kaplan 1926, Thomas 1928, Howitt 1930). McIntosh and his co workers (McIntosh *et al* 1924) however, observed a marked group reaction between members of the *acidophilus* and *acidophilus-odontolyticus* types, and Cruickshank (1925) and Weiss and Rettger (1934) found a close relationship between *L. bifidus* and *L. acidophilus*. Thomas (1928) found that Döderlein's bacillus had some relation to *L. acidophilus*, but none to *L. bulgaricus*. Working with oral strains many of which had been isolated several months previously, Harrison, Zidek and Hemmens (1939) extracted a carbohydrate-containing substance from lactobacilli and by means of precipitating antisera prepared by the inoculation of rabbits with heat killed bacilli were able to divide the strains into four serological types and a heterogeneous group.

**Pathogenicity**—None of the members appears to be pathogenic to man or animals. It is true that, in fermentative diarrhoea acid resisting bacteria may be present in large numbers in the stools, but whether they are responsible for initiating the diarrhoea, or whether they merely take advantage of the abnormal conditions prevailing in the intestine to multiply abundantly, is not clear. Their numbers increase in the intestine when lactose or dextrin are given in considerable quantities in the diet (Rettger and Cheplin 1921, Cannon and McNease 1923), and because, under these conditions, they tend to replace the proteolytic flora, their administration along with these sugars has been advocated for therapeutic purposes.

There is reason to believe that the oral lactobacilli play some part in the development of dental caries. The opinion that this disease is due to the action of acids formed in the mouth from retained food was put forward a long time ago by Robertson (1835), Tomes (1873), and others. It was not, however, till the appearance of Miller's (1889) monograph, in which it was maintained that lactic acid resulting from the bacterial fermentation of starch and sugar was primarily responsible for decalcification and solution of the enamel, that serious suspicion was thrown on the acid forming flora of the mouth. Though the streptococci have received attention from a number of workers, (see Snyder 1939), it

seems more probable that the lactobacilli, on account of their greater acid production and their ability to grow at a low pH, are implicated. Kligler (1915) and Snyder (1933) noted the increased numbers of *L. acidophilus* in the mouths of people with caries. McIntosh James and Lazarus Barlow (1922, 1924) isolated two varieties of lactobacilli from carious teeth, which they named *L. odontolyticus* I and II. In glucose broth cultures these organisms produced a final pH of 2.2 to 3.4, and it was found that teeth left in these cultures gradually became decalcified, the change being evident in 7 weeks. There is evidence to suggest that refined sugar in the diet is a more potent predisposing factor to caries than other forms of carbohydrate, but the exact mechanism by which it acts is still unknown. The effect of the acid is limited, of course, by the structural integrity and proper calcification of the enamel, which in their turn depend on nutritional factors (see McCollum 1911).

The repeated inoculation intravenously of very large doses of lactobacilli into rabbits is said to be followed by the development of joint lesions. A mucopurulent exudate is found in the joints, and cultures can be obtained for a week after the last inoculation (Howitt and van Meter 1930). It is doubtful, however, whether the organisms actually multiply within the tissues under these conditions.

### CLASSIFICATION

Classification is at present unsatisfactory, partly because of some difference of opinion as to which organisms should be included in the *Lactobacillus* group and partly by the absence of suitable criteria to serve as a basis of sub-division. There seems to us good reason for including the Gram positive members of the non-sporing anaerobic bacilli isolated by Eggerth and Gagnon (1933) from the human intestine in the *Lactobacillus* rather than in the *Bacteroides* group and in this we are supported by King and Rettger (1942). When we look for differential characters for use in classification we are in serious difficulties. Morphological and colonial appearances vary considerably, and metabolic properties and fermentation reactions tend to show continuous as opposed to discontinuous variation so that it is difficult to do more than select modal points around which strains can be grouped. Whether it is justifiable to assign specific rank to these type organisms is doubtful, but provided not too many types are recognized and provided it is realized that many of them are tentative convenience alone affords a strong plea for their characterization by name.

The most serious attempt to classify the lactobacilli is undoubtedly that made by Curran, Rogers, and Whittier (1933). As the result of a very careful study, particularly of their metabolic characteristics, these workers were able to divide 103 strains of varied origin into three groups. Group A strains produced inactive lactic acid, i.e., equal quantities of dextro and laevo acid from whey failed to grow above 13°–46° C or as low as 20° C, fermented raffinose but not mannitol, and on agar plates formed either the fuzzy (X) type of colony, or a mixture of fuzzy and compact ( $\Delta$ ) types. As a rule they gave rise to more CO<sub>2</sub> and a larger proportion of volatile to fixed acids, and grew in higher concentrations of phenol and indole (see Kulp 1929), than the members of the second group. Group B strains produced an excess of dextro lactic acid, grew as high as 43°–50° C and as low as 10°–15° C, fermented mannitol but not raffinose, and formed either the compact (Y) type of colony, or a mixture of compact and fuzzy ( $\Delta$ ) types. They were less active than

Group A strains in the production of volatile acids and  $\text{CO}_2$ , and were inhibited by relatively dilute solutions of phenol and indole. Group C strains differed from Group A in growing at  $20^\circ\text{C}$  and in a number of minor particulars. Of the 103 strains, Group A comprised 58, Group B 30, and Group C 15. With regard to their source of origin, about three-quarters of the intestinal strains belonged to Group A, while over half of the dental strains belonged to Group B. Group A probably represented the typical *L. acidophilus*, Group B probably included *L. bulgaricus* and *L. casei*, while Group C comprised a heterogeneous collection of strains whose identity was doubtful, and whose classification into one group was largely a matter of temporary convenience. Lewis and Rettger (1940) suggest a classification of the anaerobic members into three groups, A, B and C—which do not, of course, correspond to the similarly named groups of Curran, Rogers and Whittier—based on morphology, minimum growth temperature, resistance to heat, and the production of gas from fermentable carbohydrates.

The identity of *L. bulgaricus* has often been under discussion. Sherman and Hodge (1940) state that it differs from *L. acidophilus* in the following respects. Freshly isolated strains of *L. bulgaricus* grow at  $50^\circ\text{C}$ , but are unable to grow in serial culture in a simple medium containing 1 per cent. each of lactose, peptone and yeast extract or in broth of pH 7.8, or in broth containing 2.5 per cent. sodium chloride. *L. acidophilus*, on the other hand, will not grow above a temperature of  $48^\circ\text{C}$ , but will grow under the other conditions described. It need hardly be mentioned that minor physiological differences of this type, however useful in practice, can afford no satisfactory basis for permanent classification.

A classification of the gas-producing strains has been suggested by Pedersen (1935) based primarily on arabinose fermentation, and secondarily on the fermentation of lactose, sucrose and raffinose. *L. brevis* (Synonym *L. pentaoeticus*, *B. acidophil-aerogenes*) ferments arabinose, and varies in its effect on the other three sugars, its growth range is  $10^\circ$  to  $45^\circ\text{C}$ , and its optimum  $30^\circ$  to  $35^\circ\text{C}$ . *L. fermenti* does not ferment arabinose, but usually ferments the three other sugars, its growth range is  $15^\circ$  to  $50^\circ\text{C}$ , and its optimum  $35^\circ$  to  $40^\circ\text{C}$ . *L. buchneri* (Synonym *B. uehleri*) usually ferments arabinose, lactose, sucrose and raffinose, its growth range is  $10^\circ$  to  $48^\circ\text{C}$ , and its optimum  $44^\circ$  to  $48^\circ\text{C}$ . *L. pastorianus* (Synonym *L. berolinensis*) usually ferments arabinose, lactose, sucrose and raffinose, its optimum temperature for growth is  $27^\circ$  to  $32^\circ\text{C}$ . No satisfactory description, however, appears to have been given of this organism.

Barker and Haas (1944) find that the Gram positive, anaerobic, non-sporing intestinal bacteria break down lactate with the production of volatile fatty acids, and they therefore suggest that they should be classified in a separate genus, *Butyrobacterium*. Further work will clearly be required to find out whether there is, in fact, a sharp cleavage in their type of metabolism among the organisms hitherto classified as lactobacilli. In the meantime, we may point out the confusion that is likely to be caused by using the termination "bacterium" for anaerobic bacilli, it would be much better to reserve this term for non-sporing aerobic rods. Much the same criticism applies to the termination "bacillus," and it is unfortunate that the generic name *Lactobacillus* should have been chosen for organisms which are mainly anaerobic or microaerophilic. It would be better if the name could be replaced by one more suitable, such, for instance, as *Lactiformans*.

As has frequently happened with other groups, several members have been accredited with specific names without an adequate description being given of them. Moreover

it seems probable that some species that have been called by different names are in reality identical. It seems likely that Döderlein's bacillus, for example, is the same as *L. acidophilus* (Heinemann and Ecker 1916, Thomas 1928), and for that reason we have not given a separate description of it. Similarly, according to Schlot (1926) Goadby's *B. necrodentilis* is identical with *L. acidophilus*. It is doubtful whether the Boas Oppler bacillus is a separate species; it may quite well be identical with, or a variety of, *L. acidophilus* (Heinemann and Ecker 1916). *L. bulgaricus* is quite possibly the same as the type species *L. caucasicus*, of which no adequate description has ever been given (White and Avery 1910). Cruickshank (1925) suggests that *L. exilis* is the aerobic phase of *L. bifidus*, but from Tissier's (1900) original description this seems doubtful. Weiss and Rettger (1934) were unable to detect any greater difference between strains of *L. bifidus* and *L. acidophilus* than existed between the individual strains themselves, and they would therefore regard *L. bifidus* as a variant of the species of which *L. acidophilus* is the central type. The organisms described by McIntosh and his colleagues as *L. odontolyticus* I and II have received considerable study. Morishita (1929), Rosbury, Linton and Buchinder (1929), Howitt (1930), and Hadley, Bunting, and Delves (1930) have failed to find any clear distinction between oral and intestinal strains of *L. acidophilus*. Curran, Rogers and Whittier (1933), on the other hand, conclude that the lactobacilli in carious teeth do not all belong to one species and are not usually of the *acidophilus* type. Gillespie and Rettger (1938) also have noted differences between the oral and the intestinal strains of lactobacilli. For this reason we shall describe the *odontolyticus* strains separately from *L. acidophilus*.

A number of lactic acid forming bacilli were isolated from cheese by von Freudenreich and Thöni (1903), and named *Bacillus casei* by Orla Jensen (1904). These organisms have not been fully described, and we do not propose to deal with them further here. They are frequently found in milk (Sherman and Stark 1927). Another organism, described by Pederson (1936) as *Lactobacillus plantarum*, is widely distributed in fermenting plant and animal products. The differentiation of this organism from *Lactobacillus casei* is not very clear.

With regard to the nomenclature of this group, the term "acid resisting" is frequently employed, and, though correct, it is open to the objection that it may cause confusion with the acid fast bacilli. The term "acidophilic" is justifiable, but is unfortunately a hybrid. Probably Kendall's (1910) term 'aciduric' (able to endure acid), which has the advantages of not being a hybrid, of not being hyphenated, and of being technically correct, is the best one to employ.

Workers studying this group may consult the annotated bibliography on *L. acidophilus* drawn up by Frost and Hankinson (1931). Those interested in the chemical constitution of this organism are referred to a series of papers by Crowder and Anderson (1932, 1934a, b).

While emphasizing again the impossibility of classifying these organisms satisfactorily at present, we give a differential table of some of the main species, pointing out, however, that it is to be used only as a very rough indication of the characteristics of these organisms, Table 49, p. 758.

#### *Lactobacillus acidophilus*

*Synonyms*—Probably identical with Döderlein's bacillus, which is sometimes called *B. vaginalis* or *B. crustus*.

*Isolation*—Isolated by Moro (1900b) in 1900 from the faeces of breast fed infants.

*Habitat*—Found in milk, the faeces of bottle-fed infants, and often of adults, the faeces of nearly all mammals, and of many fish and invertebrates, in saliva and carious teeth.

TABLE 49

	Morphology in Culture	Surface Colonies on Tryptic Agar	Growth at 20° C	Acid production in		Optical activity of Lactic Acid produced	Clotting of Mux at 37° C	Growth in presence of 1/50, 1/100, 1/200 and 1/1000 of dose
				Marshall	Raffinose			
<i>L. acidophilus</i> Group A <sup>1</sup>	Large stumpy bacillus, often in chains and palisades, pleomorphic	Fuzzy, or fuzzy and compact	—	—	+	Inactive	1-3 days with firm curd. Latexes reduced	+
<i>L. acidophilus</i> Group B probably includes <i>L. bifidus</i> and <i>L. casei</i>	Variable size, often stout frequently in chains pleomorphic	Compact or compact and fuzzy	+	+	—	Inactive and lextro rotatory	1-3 days with firm curd. Latexes reduced	—
<i>L. bifidus</i>	More delicate bacillus with slightly jointed ends rarely in chains pleomorphic	Compact or intermediate	±	—	+	Type I in active Type II lextro rotatory	Usually several days. Loose or irregular curd	?
<i>L. brevis</i> probably same as <i>L. acidophilus arvensis</i> and <i>L. lactis arvensis</i>	Rather long bacillus slenderer than <i>L. bifidus</i> often arranged in long curved chains	Compact or fuzzy	?	—	+	?	Little or no growth, some strains produce late clotting	?

<sup>1</sup> Groups A and B refer to Curran, Rogers and Whittier's classification

<sup>2</sup> *L. bifidus* is generally stated not to grow at 20° C but it is nevertheless included by Curran, Rogers and Whittier (1933) in their Group B

**Morphology**—In faeces it is a large stumpy bacillus of variable length and fairly constant breadth, generally straight, with parallel sides and rounded ends. On agar plate cultures, after 48 hours at 37° C, it forms fairly thick rods, 1-3  $\mu$  long by 1.0  $\mu$  broad, straight or slightly curved, with parallel sides and rounded or slightly truncated ends, arranged singly, in pairs end to end, in short chains, and in palisades. On glucose agar longer chains and filamentous forms are common. In broth and litmus milk cultures the bacilli are thinner, 0.6-0.8  $\mu$  broad. Considerable variation in morphology on artificial media, forms with markedly curved extremities, curled forms, forms with bulbous extremities, clubbed forms, filamentous forms, and large swollen oval forms in pairs are not uncommon. Under anaerobic conditions long curved filamentous forms, often with pointed, swollen, or spatulate ends are seen. Non motile. Gram positive. Staining is uniform in young cultures, but irregularly stained, bipolar stained, and beaded forms are met with in old cultures. Non acid fast.



FIG 152—*Lactobacillus acidophilus*  
From an agar culture, 48 hours, 37° C ( $\times 1000$ )



FIG 153—*Lactobacillus acidophilus*

Surface colony on agar, 4 days  
37° C, showing differentiation  
( $\times 8$ )

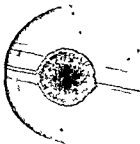


FIG 154—*Lactobacillus acidophilus*

Surface colony on agar, 4 days,  
37° C ( $\times 8$ )

**Agar Plates**—Great variability in colonial appearance. After 48 hours at 37° C one of the commonest is a small, irregularly round, raised, colourless, transparent colony, about 0.5 mm in diameter, of coarsely frosted glass structure, with a dull uneven relief map-like surface and a fimbriate or curled edge. Later, differentiation occurs into a thicker darker centre, and a thinner periphery consisting of delicately curled and branched hair like streamers—not unlike a colony of *C. tetani*. Other forms are feathery colonies, rosette-like colonies, smooth colonies with entire or slightly fimbriate edges, and opaque striated colonies.



- Agar Slope.**—24 hours 37° C. Poor filament growth of small transparent granular colonies, 0.25 to 0.5 mm. in diameter. Sometimes after a few days a thick greyish-yellow secondary growth occurs, having a contoured surface, and an edge from which thin translucent branching tufts, looking like sea-anemones, project.
- 2 per cent. Glucose Agar Slab.**—24 hours, 37° C. Good growth throughout tube of small, spherical, lenticular, or lobulated colonies, 0.25 mm. in diameter, having an entire edge (Y type), or of small irregular spherical colonies with a furry filamentous border (X type). The agar is clouded and has an almost milky look. After a week the colonies are 0.5 mm. in diameter, are porcelain-white in colour, and look like colonies of mould.
- Gelatin Slab.**—7 days 22° C. Very poor greenish-white filament growth, no surface growth, no liquefaction.
- Broth.**—48 hours, 37° C. Poor to moderate growth with a very slight, only just perceptible turbidity and a moderate flocculo-granular deposit disintegrating to some extent on shaking, sometimes the deposit sticks to the walls of the tube.
- 2 per cent. Glucose Broth.**—Growth is better than in plain broth. Sometimes after a week an enormous loose flocculent deposit forms, filling the lower 1 cm. of the tube, it disintegrates partly on shaking, producing a dense turbidity.
- Potato.**—No growth.
- Horse Blood Agar Plates.**—48 hours 37° C. Small discrete colonies, similar to those on agar. No haemolysis.
- Glucose Blood Liver Agar Plates.**—48 hours, 37° C. Flat, dingy colonies with a serrated edge.
- MacConkey Plate.**—Results discrepant. Crumshank (1931) says that all strains grow on this medium, but this has not been our own experience.
- Resistance.**—Not particularly resistant. Killed by moist heat at 56° C. in 30 minutes. Very resistant to acids, living for 1 to 3 days in broth containing 0.5-1.0 per cent. acetic or lactic acid. Glucose broth cultures at 37° C. remain viable for about a fortnight.
- Metabolism.**—Often microaerophilic on first isolation. Grows better under aerobic than anaerobic conditions. Grows slightly or not at all at 20° C., optimum temperature for growth 37° C. Forms no pigment and no toxin. Does not live on horse blood. Growth is improved by glucose and by whey, but not by blood serum. Grows best at pH 6.0 but will grow even at pH 5.0.
- Biochemical.**—Sugar reactions variable. Produces acid in glucose and in lactose, often in maltose and sucrose, sometimes in mannitol, salicin, and raffinose, less frequently in other sugars. L.M. acid and clot in 24 to 48 hours, the clot is really an acid precipitate, and does not contract, on shaking it breaks up into flocculent masses, the litmus is at first reduced at the bottom of the tube only, but later the decolorization spreads upwards. Indole negative. M.R. positive. V.P. negative. Nitrates reduced slightly or not at all. Catalase very weak positive. H<sub>2</sub>S negative. NH<sub>3</sub> negative.
- Antigenic Structure.**—Not studied fully. By agglutination numerous groups can be made out, having little affinity with each other. Some group relationship to *L. lactis*.
- Pathogenicity.**—Non-pathogenic to man or to laboratory animals.

### *Lactobacillus odontolyticus* I

- Isolation.**—Isolated by McIntosh, James, and Lazarus-Barlow in 1922 from carious teeth and from saliva, and called *B. acidophilus odontolyticus* I.
- Morphology.**—Thin bacillus, 2-3  $\mu$  long by 0.75  $\mu$  broad, occurring singly, in pairs, or chains, and in palisades. Non-motile. Gram-positive.
- Agar Plates.**—48 hours 37° C. Small, round, greenish, opaque colonies, 0.6-1.0 mm. in diameter with a finely granular appearance and an entire edge. On serum agar the colonies are larger up to 2 mm. in diameter.

*Gelatin Slab*—Grows to the bottom. No liquefaction.

*Gelatin Agar Shake*.—Deep colonies are roughly biconvex or tam-o'-shanter-shaped.

*Broth*—Uniform turbidity, sometimes the growth settles to the bottom.

*Resistance*.—Not particularly resistant. Killed by 56° C. in 25 minutes, and by 2 per cent phenol in 7½ minutes. Highly resistant to acids, will withstand incubation for 24 hours at 37° C. in broth of pH 3.5. No growth above pH 9.1-9.6.

*Metabolism*.—Aerobe and facultative anaerobe.

*Biochemical*—Sugar reactions variable. Usually produces acid in glucose, maltose, salicin and lactose, but not in sucrose, dextrin, dulcitol, or raffinose. L.M. acid and clot in 2 to 3 days, lower ⅔ of tube decolorized. Indole negative. Final pH in glucose broth cultures is about pH 2.75. Chief acid formed is malic acid, lactic acid is formed in only a very small amount. Methyl red positive. Voges-Proskauer negative. Nitrates not reduced. Catalase very slight positive. NH<sub>4</sub> negative.

*Antigenic Structure*—Appears to be fairly homogeneous, and to be closely related to *L. odontolyticus* II, and to *L. acidophilus*.

*Pathogenicity*—Suspected of being responsible for production of dental caries. Non-pathogenic to laboratory animals.

### *Lactobacillus odontolyticus* II

*Isolation*—Isolated by McIntosh, James, and Lazarus Barlow in 1922 from carious teeth and from saliva, and called *B. acidophilus odontolyticus* II.

*Morphology*—Rather short bacillus, 1.2 µ long by 0.5 µ broad, usually arranged in short chains. Often very pleomorphic, coccil forms being mixed with bacillary forms in the same chain, may closely resemble a streptococcus. Non motile. Gram positive.

*Cultural Reactions*—Similar to *odontolyticus* I.

*Resistance and Metabolism*—Similar to *odontolyticus* I.

*Biochemical*—Variable sugar reactions. 7 out of 18 strains produced acid in glucose, lactose, and sucrose. L.M. Some strains produce acid and clot, others have no action on it. Indole negative.

*Antigenic Structure*.—Closely allied to *odontolyticus* I and to *L. acidophilus*.

*Pathogenicity*—Like *odontolyticus* I.

### *Lactobacillus brevis*

*Synonyms*—Probably *L. pentoceticus*, *B. acidophilus-aerogenes*.

*Isolation*.—By Torrey and Rabe in 1915 from the faeces of human beings, sheep and hens.

*Habitat*—Common in fermenting plant and animal products.

*Morphology*—Variable morphology. Size given by some authors as 1.5-11.5 µ long by 0.8 µ broad, and by others as 1-4 µ long by 0.6-0.8 µ broad, often arranged in long curved strings. Non motile. Gram positive. Stain uniformly with Loeffler's methylene blue.

*Cultural Reactions*—On glucose oleate agar it forms either (1) small, round or navicular, opaque, whitish colonies surrounded by an aureole of turbid agar, or (2) tiny, round, translucent, greyish colonies with a finely erose edge, on microscopic examination these appear typically rhizoid. In glucose broth a growth forms adherent to the bottom and sides of the tube, on shaking this gives rise to a dense turbidity.

*Resistance*.—Highly resistant to acids, will remain alive in a glucose broth culture at 37° C. for 1 week.

*Metabolism*—Microaerophilic. Range of growth 10°-45° C., optimum temperature 30°-35° C.

**Biochemical.**—Produces acid and gas in glucose, maltose, arabinose and xylose. Fermentation variable of lactose, sucrose, mannose, raffinose and mannitol.

The gas ratio is  $4\text{H}_2/\text{CO}_2$  or  $6\text{H}_2/\text{CO}_2$ . L.M. grows poorly or not at all, some strains produce partial clotting in 2 to 3 weeks.

**Antigenic Structure**—Appears to be more homogeneous than the non-gas-forming group.

### *Lactobacillus bifidus* I

**Isolation.**—Isolated by Tieszer in 1900 from the faeces of breast fed infants.

**Habitat.**—Common in the faeces of breast fed and much less common in those of bottle-fed infants. Sometimes present in the faeces of adults and of animals. In breast fed infants during the first few weeks of life it may form 99 per cent. of the faecal flora.

**Morphology.**—In faeces it is a delicate bacillus, about  $4\mu$  long and  $0.7\mu$  broad, with tapering pointed ends, arranged in pairs end-to-end, with the distal ends pointed and the proximal ends swollen, they generally lie parallel to one



FIG 1-5.—*Lactobacillus bifidus*

From a glucose agar culture 7 days,  $37^\circ\text{C}$ . ( $\times 1000$ )

another rarely intertwined. Two or three bacilli often radiate from a single point forming a Y-shaped structure simulating branching, clubbed forms and forms ending in knobs are not uncommon. Often arranged in palisades or Chinese letters. General appearance is not unlike a diphtheroid bacillus. In young cultures bacilli with slightly pointed ends of varying length, arranged singly or in pairs end-to-end, are usual. In older cultures longer clubbed forms, geniculate forms, bifid forms showing false branching, forms ending in knobs, forms with lateral buds, bladder forms, candle-flame forms, and filamentous forms may appear. Both in faeces and in culture there is a striking pleomorphism. The absence of chain formation is noteworthy. Non-motile. In young cultures staining is fairly uniform but in older cultures and in faeces irregular, granular, and beaded staining are common. Gram-positive in young cultures, later Gram-negative forms appear. Non-acid-fast.

**Agar Plate.**—No growth.

**Glucose Agar Plate.**—48 hours  $37^\circ\text{C}$ . Small round low convex colonies, 0.5 mm. in diameter showing under the microscope a delicately granular structure a brownish opaque centre, a thinner translucent periphery and a finely crenated edge (Fig. 1-6).

**Glucose Agar Shake Tubes.**—3 days  $37^\circ\text{C}$ . Small, greyish-brown, lenticular or ovoid colonies, 1.2 mm. in diameter extending up to within about 3 cm. of the surface, where they may form a ring. At first the edge of the colony is entire, but after 4 or 5 days a lateral projection or bud often develops from one of the faces.



FIG 1-6.—*Lactobacillus bifidus*

Surface colonies on glucose agar 6 days,  $37^\circ\text{C}$ . ( $\times 8$ )

- Gelatin Shake**—If incubated at 37° C. for 24 hours and then left at room temperature for some days fine discrete colonies appear, no liquefaction.
- Broth**—24 hours 37° C Granular turbidity
- Glucose Broth**—24 hours, 37° C More abundant growth, in 3 or 4 days the organisms fall to the bottom of the tube, producing an abundant loose flocculo-granular deposit, easily disintegrated on shaking
- Blood Agar**—White colonies surrounded by a greenish halo of a hæmolytic
- Glucose Blood Liver Agar Plates**—48 hours 37° C Raised globular opaque colonies 1-3 mm in diameter buff to reddish brown in colour
- Resistance**—Cultures live for about a month at room temperature Broth cultures are killed by heat at 55° C. in half an hour, and at 70° C. in 5 minutes Is resistant to acids, will withstand 0.5-1.0 per cent acetic or lactic acid in broth for 2 to 3 days
- Metabolism**—Strict anaerobe on first isolation, later it may be grown in air Very slight or no growth at 20° C, optimum temperature 37° C No pigment or toxin formed a hæmolytic on blood agar Growth improved by glucose serum and blood
- Biochemical**—Sugar reactions variable Produces acid in glucose mannose maltose inulin and generally in lactose sucrose salicin and raffinose sometimes in mannitol and dextrin, occasionally in dulcitol, not in arabinose xylose or melezitose Produces mainly lactic acid, of the inactive variety, and some acetic acid L.M. grows well and produces an acid clot Indole negative Methyl red positive Voges-Proskauer negative Nitrates very slight reduction or none at all Catalase very slight positive. NH<sub>3</sub> slight production or none at all
- Antigenic Structure**—Not fully worked out By agglutination they appear to fall into more than one group Some relationship to *L. acidophilus*
- Pathogenicity**—Non pathogenic to man and laboratory animals

### Lactobacillus bifidus II

**SYNONYMS** *Bacteroides bifidus* (Eggerth Group II 1935), *Bacterium bifidum* (Orla-Jensen see Orla-Jensen et al 1936)

Differs from *L. bifidus I* mainly in the following characters It is found in the intestine of the adult living on a mixed diet in which it may form quite a high proportion of the total flora It branches more readily and continues to branch on subculture It is an obligate anaerobe and rarely becomes accustomed to aerobic conditions Deep colonies in tomato agar have an entire not a filamentous edge (Weiss and Rettger 1938) Mannose is not usually fermented but most strains ferment arabinose xylose and melezitose It produces lactic acid of the dextro variety and about 50 per cent of acetic acid It appears to be antigenically more strain-specific

### Lactobacillus bulgaricus

**Synonyms**—*Massalia bacillus*. Probably identical with *L. caucasicus*

**Isolation**—Isolated by Gligoroff in 1905 from kasselo-mleko the fermented milk of Bulgarians, described originally as *Bacillus A*.

**Habitat**—Found in milk particularly the fermented milks of Bulgaria Turkey Egypt and Sardinia.

**Morphology**—Large rods 2-20  $\mu$  long and about 1  $\mu$  broad with parallel sides and slightly rounded ends, arranged singly or in short chains. Non motile Gram positive Two morphological types are described by White and Avery (1910) in whey Type A consists of chains of short bacilli with oval or reniform nodules extruding from the cell substance, the bacilli stain uniformly. Type B forms long bacilli arranged singly having spherical bodies attached to the cell wall not stemmed nodules as in A, the bacilli show intense granular staining with Loeffler's methylene blue or Neisser's stain.

*Agar Plate*.—No growth.

*Whey Agar Plate*.—Usual type of colony is irregularly round, greyish white, 0.5-1.5 mm. in diameter, of loose curled structure with a streaming filamentous edge, microscopically these colonies are typically rhizoid. Sometimes, especially in old laboratory cultures, a rounder more regular colony is formed, with a smooth or slightly fissured surface and an entire edge.

*Whey Agar Shake Tubes*.—Deep colonies are lenticular or umbilicated, whitish in colour, and 1 mm. in diameter.

*Whey Agar Slab*.—Filiform growth, beaded, later with horizontal ramifications, no surface growth, medium is clouded.

*Gelatin*.—Not liquefied.

*2 per cent. Glucose Broth*.—Heavy uniform turbidity.

*Potato*.—No growth.

*Resistance*.—Killed by moist heat at 60° C. in 1 hour. Is very resistant to acids, glucose broth cultures at 37° C. remain viable for about 6 days.

*Metabolism*.—Facultative anaerobe, is said by some authors to prefer anaerobic conditions. No growth at 15° C. grows very slightly at 20° C., growth is poor under 35° C., optimum temperature for growth is 44-45° C., usually grows at 50° C. No pigment, toxin, or haemolysin formed. Is resistant to acids, but grows best in a neutral or slightly alkaline medium. Difficult to cultivate, growth in most media is feeble, and when freshly isolated it will grow only on media containing whey or malt, or in milk.

*Biochemical*.—Sugar reactions described differently by different authors. Is generally considered to produce acid in glucose, lactose, and sometimes levulose but not in mallose, sucrose, mannitol, or raffinose. Lactic acid is the chief acid formed from fermentable carbohydrates. L.M. acid and coagulation in 15 hours at 37° C. the clot does not contract. Indole negative. White and Avery's Type A in milk produces 2-3.7 per cent. of lactic acid of the inactive variety, Type B produces only 1-1.6 per cent. of lactic acid, of the laro-rotatory variety.

*Antigenic Structure*.—Nothing known.

*Pathogenicity*.—Non-pathogenic to man and animals.

*Grigoroff's Bacillus C*.—Isolated from kusselo-milko, the fermented milk of Bulgarians by Grigoroff in 1903. Is a streptobacillus forming short rods arranged in chains of four to twenty members. Gram positive. Culturally it resembles *L. bulgaricus*, but is more heat resistant, requiring an exposure of 1 hour at 70° C. to kill it. Produces acid in glucose, levulose, lactose, sucrose, and glycerol, but not in mallose, mannitol, or dulcitol. Produces acid and clot in milk, the acid being inactive lactic acid.

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*The Boas-Oppler Bacillus*.—Observed microscopically in the stomach contents of patients with gastric carcinoma by Oppler in 1893, working in Boas' clinic in Berlin. He did not succeed in cultivating it. In stomach contents it occurs as a rather slender bacillus,

arranged in long threads and zigzags, the bacilli may be so numerous as to fill every space between the other elements in the field. Has since been isolated by Heinemann and Ecker (1916) from the stomach contents of patients with gastritis, gastric ulcer, carcinoma, and pernicious anemia. It probably occurs in moderate numbers in normal gastric juice. In culture the organism is a large, rather slender bacillus, showing granular staining. Gram positive. Forms compact colonies as a rule, but colonies with woolly edges have been described. Does not ferment maltose. Produces acid in milk. Is of the low acid producing type—White and Avery's Type B.

## REFERENCES

- BARKER, H. A. and HAAS, V. (1914) *J. Bact.*, 47, 301.  
 BERTHARD, G. and DUCHACER, F. (1909) *Ann. Inst. Pasteur*, 23, 402.  
 CANTON, P. R. and McNEASE, B. W. (1923) *J. infect. Dis.*, 32, 175.  
 CONNEDY, M. (1900) *C. R. Soc. Biol.*, 52, 558.  
 CROWDER, J. A. and ANDERSON, R. J. (1932) *J. biol. Chem.*, 97, 393. (1934a) *Ibid.* 104, 399. (1934b) *Ibid.* 104, 487.  
 CROWLEY, N., DOWD, A. W., FULTON, F., and WILSON, G. S. (1941) *Lancet*, ii, 590.  
 CRICKSHANK, R. (1925) *J. Hyg., Camb.*, 24, 241. (1931) *J. Hyg., Camb.*, 31, 375. (1934) *J. Path. Bact.*, 39, 213.  
 CURRAN, H. R., ROGERS, L. A., and WHITTIER, E. O. (1933) *J. Bact.*, 25, 595.  
 DAY, A. A. and GIBBS, W. M. (1928) *J. infect. Dis.*, 43, 97.  
 DÖDERLEIN (1892) "Das Scheidensekret und seine Bedeutung für das Puerperalfieber." Leipzig.  
 EGGERTH, A. H. (1935) *J. Bact.*, 30, 277.  
 EGGERTH, A. H. and GAGNON, B. H. (1933) *J. Bact.*, 25, 389.  
 FINKELSTEIN, H. (1900) *Dtsch. med. Wschr.*, 26, 263.  
 FRED, E. B., PETERSON, W. H., and DAVENPORT, A. (1919) *J. biol. Chem.* 39, 347.  
 FREUDENREICH, E. von and THÖNI, J. (1903) *Zbl. Bakt., IIte Abt.* 10, 305-340.  
 FROST, W. D. and HANKINSON, H. (1931) "Lactobacillus acidophilus." Davis Greene Corp., Milton, Wis.  
 GILLESPIE, R. W. H. and RETTGER, L. F. (1938) *J. Bact.*, 36, 621.  
 GRIGOROFF, S. (1905) *Rev. med. Suisse rom.*, 25, 714.  
 HADLEY, F. P., BUNTING, R. W., and DELVES, E. A. (1930) *J. Amer. dent. Ass.* 17, 2041.  
 HARRISON, R. W., ZIDEN, Z. C., and HEMMENS, E. S. (1939) *J. infect. Dis.* 65, 255.  
 HEINEMANN, P. G. and ECKER, E. E. (1916) *J. Bact.*, 1, 435.  
 HEINEMANN, P. G. and HEPPEL, M. (1909) *J. infect. Dis.*, 6, 304.  
 HOWITT, B. (1930) *J. infect. Dis.*, 46, 351.  
 HOWITT, B. and MEYER, M. VAN (1930) *J. infect. Dis.*, 46, 368.  
 HUNT, G. A. (1933) *J. Bact.*, 26, 341.  
 HUNT, G. A. and RETTGER, L. F. (1930) *J. Bact.*, 20, 61.  
 KENDALL, A. I. (1910) *J. med. Res.*, 22, 153.  
 KENDALL, A. I. and HANER, R. C. (1924a) *J. infect. Dis.*, 35, 77. (1924b) *Ibid.* 35, 89.  
 KERN (1881) *Bull. Soc. Nat. Moscou* No. 3.  
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 KLIGLER, I. J. (1915) *J. allied dent. Soc.*, 10, 141, 282, 445.  
 KULP, W. L. (1926) *Science*, 64, 304. (1927) *Ibid.*, 66, 512. (1929) *J. Bact.*, 17, 350.  
 LASH, A. F. and KAUFMAN, B. (1926) *J. infect. Dis.* 38, 333.  
 LEWIS, R. H., BEDZELL, M., and RETTGER, L. F. (1940) *J. Bact.*, 40, 309.  
 LEWIS, R. H. and RETTGER, L. F. (1940) *J. Bact.*, 40, 287.  
 LONGSWORTH, L. G. and MACINTOSH, D. A. (1935) *J. Bact.*, 29, 595.  
 MCCOLLUM, E. V. (1941) *Nature*, 147, 104.  
 MCINTOSH, J., JAMES, W. W., and LAZARUS BARLOW, P. (1922) *Brit. J. exp. Path.*, 3, 138. (1924) *Ibid.*, 5, 175.  
 MEZENSKOWSKY, S. S. (1905) *Zbl. Bakt.* 39, 390, 584-696. (1906) *Ibid.*, 40, 118.  
 MILLER, W. D. (1889) "Die Mikroorganismen der Mundhöhle." G. Thieme, Leipzig.  
 MORISHITA, T. (1929) *J. Bact.*, 18, 181.  
 MORO, E. (1900a) *Jb. Kinderheilk.*, 52, 38. (1900b) *Wien. klin. Wschr.*, 13, 114.  
 OPFLER, B. (1935) *Dtsch. med. Wschr.*, 21, 73.  
 ORLA-JENSEN (1904) *Zbl. Bakt., IIte Abt.*, 13, 161, 291, 428, 514, 604, 687, 753.  
 ORLA-JENSEN, S., ORLA-JENSEN, A. D., and WINTNER, O. (1936) *Zbl. Bakt., IIte Abt.*, 93, 321.  
 PEDERSON, C. S. (1936) *J. Bact.*, 31, 217. (1938) *Ibid.*, 35, 95.  
 PETERSON, W. H., PRUESS, L. M., and FAXD, E. B. (1928) *J. Bact.*, 15, 165.

*Agar Plate*.—No growth.

*Whey Agar Plate*.—Usual type of colony is irregularly round, greyish white, 0.5–1.5 mm. in diameter, of loose curled structure with a streaming filamentous edge, microscopically these colonies are typically rhizoid. Sometimes, especially in old laboratory cultures, a rounder, more regular colony is formed, with a smooth or slightly fissured surface and an entire edge.

*Whey Agar Shake Tubes*.—Deep colonies are lenticular or umblicated, whitish in colour, and 1 mm. in diameter.

*Whey Agar Slab*.—Filiform growth, beaded, later with horizontal ramifications, no surface growth, medium is clouded.

*Gelatin*.—Not liquefied.

*2 per cent. Glucose Broth*.—Heavy uniform turbidity.

*Potato*.—No growth.

*Resistance*.—Killed by moist heat at 60° C. in 1 hour. Is very resistant to acids, glucose broth cultures at 37° C. remain viable for about 6 days.

*Metabolism*.—Facultative anaerobe, is said by some authors to prefer anaerobic conditions. No growth at 15° C., grows very slightly at 25° C., growth is poor under 35° C., optimum temperature for growth is 44–45° C., usually grows at 50° C. No pigment, toxin, or haemolysin formed. Is resistant to acids, but grows best in a neutral or slightly alkaline medium. Difficult to cultivate, growth in most media is feeble, and when freshly isolated it will grow only on media containing whey or malt, or in milk.

*Biochemical*.—Sugar reactions described differently by different authors. Is generally considered to produce acid in glucose, lactose, and sometimes levulose, but not in maltose, sucrose, mannitol, or raffinose. Lactic acid is the chief acid formed from fermentable carbohydrates. L.M. acid and coagulation in 18 hours at 37° C., the clot does not contract. Indole negative. White and Avery's Type A in milk produces 2.7–3.7 per cent. of lactic acid of the inactive variety, Type B produces only 1.2–1.6 per cent. of lactic acid, of the levo-rotatory variety.

*Antigenic Structure*.—Nothing known.

*Pathogenicity*.—Non pathogenic to man and animals.

*Grigoroff's Bacillus C*.—Isolated from *kiselo-mleko*, the fermented milk of Bulgaria by Grigoroff in 1905. Is a streptobacillus forming short rods arranged in chains of four to twenty members. Gram positive. Culturally it resembles *L. bulgaricus*, but is more heat resistant requiring an exposure of 1 hour at 70° C. to kill it. Produces acid in glucose, levulose, lactose, sucrose and glycerol but not in maltose, mannitol, or dulcitol. Produces acid and clot in milk, the acid being inactive lactic acid.

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## REFERENCES

- BARKER, H. A. and HAAS, V. (1944) *J. Bact.* 47, 301.  
 BERTRAND, G. and DUCHACEK, F. (1909) *Ann. Inst. Pasteur* 23, 402.  
 CANNON, P. R. and McNEASE, B. W. (1923) *J. infect. Dis.* 32, 175.  
 CONENDY, M. (1906) *C. R. Soc. Biol.* 58, 658.  
 CROWDER, J. A. and ANDERSON, R. J. (1932) *J. biol. Chem.* 97, 393. (1934a) *Ibid.* 104, 399. (1934b) *Ibid.* 104, 487.  
 CROWLEY, N., DOWNIE, A. W., FULTON, T., and WILSON, G. S. (1941) *Lancet*, ii 590.  
 CRICKSHANK, R. (1925) *J. Hyg., Camb.* 24, 241. (1931) *J. Hyg., Camb.* 31, 375. (1934) *J. Path. Bact.* 39, 213.  
 CURRAN, H. R., ROGERS, L. A., and WHITTIER, E. O. (1933) *J. Bact.* 25, 595.  
 DAY, A. A. and GIBBS, W. M. (1928) *J. infect. Dis.* 43, 97.  
 DÖDERLEIN (1892) "Das Scheidensekret und seine Bedeutung für das Puerperalfieber." Leipzig.  
 EGGERTH, A. H. (1935) *J. Bact.* 30, 277.  
 EGGERTH, A. H. and GAGNON, B. H. (1933) *J. Bact.* 25, 389.  
 FINKELSTEIN, H. (1900) *Dtsch. med. Wochschr.* 26, 203.  
 FRED, E. B., PETERSON, W. H., and DAVENPORT, A. (1919) *J. biol. Chem.* 39, 347.  
 FREUDENREICH, E. von and THÖNY, J. (1903) *Zbl. Bakt., IIte Abt.* 10, 305-340.  
 FROST, W. D. and HARRISON, H. (1931) 'Lactobacillus acidophilus' Davis Greene Corp., Milton, Wis.  
 GILLESPIE, R. W. H. and RETTOER, L. F. (1938) *J. Bact.* 36, 621.  
 GRIGOROFF, S. (1905) *Rev. med. Suisse rom.* 25, 714.  
 HADLEY, F. P., BUNTING, R. W., and DELVES, E. A. (1930) *J. Amer. dent. Ass.* 17, 2041.  
 HARRISON, R. W., ZIDEK, Z. C., and HEMMENS, E. S. (1939) *J. infect. Dis.* 65, 255.  
 HEINEMANN, P. G. and ECKER, E. E. (1916) *J. Bact.* 1, 435.  
 HEINEMANN, P. G. and HEFFERAN, M. (1909) *J. infect. Dis.* 6, 304.  
 HOWITT, B. (1930) *J. infect. Dis.* 46, 351.  
 HOWITT, B. and METER, M. VAN (1930) *J. infect. Dis.* 46, 368.  
 HUNT, G. A. (1933) *J. Bact.* 26, 341.  
 HUNT, G. A. and RETTOER, L. F. (1930) *J. Bact.* 20, 61.  
 KENDALL, A. I. (1910) *J. med. Res.* 22, 153.  
 KENDALL, A. I. and HANER, R. C. (1924a) *J. infect. Dis.* 35, 77. (1924b) *Ibid.* 35, 89.  
 KERN (1881) *Bull. Soc. Nat. Moscou* No. 3.  
 KING, J. W. and RETTOER, L. F. (1942) *J. Bact.* 44, 301.  
 KLIGLER, I. J. (1915) *J. allied dent. Soc.* 10, 141, 282, 445.  
 KULP, W. L. (1926) *Science*, 64, 304. (1927) *Ibid.* 66, 612. (1929) *J. Bact.* 17, 355.  
 LASH, A. F. and KAPLAN, B. (1926) *J. infect. Dis.* 38, 333.  
 LEWIS, R. H., BEDELL, M., and RETTOER, L. F. (1940) *J. Bact.* 40, 309.  
 LEWIS, R. H. and RETTOER, L. F. (1940) *J. Bact.* 40, 267.  
 LONGSWORTH, L. G. and MACINNIS, D. A. (1935) *J. Bact.* 29, 595.  
 MCCOLLUM, E. V. (1941) *Nature*, 147, 104.  
 MCINTOSH, J., JAMES, W. W., and LAZARUS BARLOW, P. (1922) *Brit. J. exp. Path.* 3, 138. (1924) *Ibid.* 5, 175.  
 MERESKOWSKY, S. B. (1905) *Zbl. Bakt.* 39, 380, 584, 696. (1906) *Ibid.* 40, 118.  
 MILLER, W. D. (1899) "Die Mikroorganismen der Mundhöhle." G. Thieme, Leipzig.  
 MORISHITA, T. (1929) *J. Bact.* 18, 181.  
 MORO, E. (1900a) *Jb. Kinderheilk.* 52, 38. (1900b) *Wien. klin. Wochschr.* 13, 114.  
 OPPLER, B. (1895) *Dtsch. med. Wochschr.* 21, 73.  
 ORLA-JENSEN (1904) *Zbl. Bakt., IIte Abt.* 13, 161, 291, 428, 514, 604, 687, 753.  
 ORLA-JENSEN, S., ORLA-JENSEN, A. D. and WINTHER, O. (1936) *Zbl. Bakt., IIte Abt.* 93, 321.  
 PEDERSON, C. S. (1936) *J. Bact.* 31, 217. (1938) *Ibid.* 35, 95.  
 PETERSON, W. H., PRUESS, L. M., and FRED, E. B. (1928) *J. Bact.* 15, 165.



- PETROW, N. P. (1907) *Zbl. Bakt.*, 43, 349
- REITGER, L. F. and CHEPLIN, H. A. (1931) 'A Treatise on the Transformation of the Intestinal Flora with Special Reference to the Implantation of *Acidophalus*.' New Haven.
- REITGER, L. F. and HORTON, G. D. (1914) *Zbl. Bakt.*, 73, 362.
- ROBERTSON, W. (1835) 'A practical treatise on the human teeth etc.' Quoted from Miller (1889)
- RODELLA, A. (1901) *Zbl. Bakt.*, 29, 717
- ROSEBURY, T., LINTON, R. W., and RECHBINDER, L. (1929) *J. Bact.*, 18, 395
- SCHLIEF, K. (1936) *Zbl. Bakt.*, 97, 104
- SHERMAN, J. M. and HODGE, H. M. (1940) *J. Bact.*, 40, 11
- SHERMAN, J. M. and STARK, C. V. (1927) *J. Bact.*, 13, 60.
- SKELL, E. E., STRONG, F. M., and PETERSON, W. H. (1939) *J. Bact.*, 38, 203.
- SNYDER, M. L. (1939) *J. dent. Res.*, 18, 497
- THOMAS, S. (1928) *J. infect. Dis.*, 43, 218.
- TOSSIER, H. (1900) "Recherches sur la flore intestinale des nourrissons." Paris.
- TOMES, J. (1873) *Dent. Surg.*, p. 734 Quoted from Miller (1889)
- TORREY, J. C. and RAHZE, A. H. (1915) *J. infect. Dis.*, 17, 437
- VEILLOU, A. and ZUCKER, A. (1898) *Arch. Med. exp.*, 10, 517
- WEINSTEIN, L. and REITGER, L. F. (1932) *J. Bact.*, 24, 1
- WEISS, J. E. and REITGER, L. F. (1934) *J. Bact.*, 28, 501 (1938) *J. infect. Dis.*, 62, 115.
- WHITE, B. and AVERY, O. T. (1910) *Zbl. Bakt. IIte Abt.*, 25, 161

## CHAPTER 32

### PASTEURELLA

#### DEFINITION — *Pasteurella*

Small, Gram negative, ovoid bacilli, showing bipolar staining. Aerobic and facultatively anaerobic. Powers of carbohydrate fermentation relatively slight, no gas produced. Gelatin not liquefied. Parasites in man and animals producing characteristic infections.

The type species is *Pasteurella aviseptica*.

**Isolation**—The first member of this group was isolated by Kitt in 1878 from an epidemic disease affecting wild hogs and deer. Similar organisms have been isolated from several species of animals and birds suffering from a disease known as hæmorrhagic septicæmia. It has become customary to give a specific name to each organism, corresponding to the animal from which it was derived, thus we have *Past. aviseptica* from fowls, *Past. leipseptica* from rabbits, *Past. suissepica* from pigs, *Past. vituliseptica* from calves, *Past. oviseptica* from sheep, *Past. bovissepica* from cattle, and *Past. muriseptica* from mice (not to be confused with *Erysipelothrix muriseptica*). Such a nomenclature is purely arbitrary, and is clearly unjustifiable from the systematic point of view. The differentiation of species within this group, as in all others, must depend on the detailed study of an adequate sample of strains. Such data as are available do not suggest that the strains from the various animal species, which are liable to natural pasteurellosis, are themselves specifically distinct. In the description which follows we have taken *Past. aviseptica* as a type of the hæmorrhagic septicæmia group, but, as we point out in a later section, it is doubtful whether this name will survive as a designation for a distinct species.

Malassez and Vignal (1883) were apparently the first to describe pseudotuberculosis in the guinea pig. Several workers recorded the finding of a bacillus in this disease (see Chapter 73), chief amongst whom was Pfeiffer (1890), who named it *B. pseudotuberculosis*. It is not to be confused with *Corynebacterium pseudotuberculosis ovis*, described by Preisz and Nocard as the cause of pseudo tuberculosis in sheep (see Chapter 17), or with *Corynebacterium pseudotuberculosis murium*, described by Kutscher (1894) and Bongert (1901) as the cause of pseudo tuberculosis in mice (see Chapter 17).

The plague bacillus, *Past. pestis*, was isolated almost simultaneously by Kitasato (1894) and by Yersin (1894), from human patients suffering from plague.

These three organisms resemble each other in so many characters, and appear to be so closely related, that they may well be considered as falling within a single genus.

**Morphology and Staining**—All the members of the group are small, ovoid bacilli, with convex sides and rounded ends, there is no characteristic arrange-

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- TORREY, J. C. and RAHE, A. H. (1915) *J. infect. Dis.*, 17, 437
- VEILLOU, A. and ZUCKER, A. (1895) *Arch. Med. exp.*, 10, 51
- WEINSTEIN, L. and REITZGER, L. F. (1932) *J. Bact.*, 24, 1
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These three organisms resemble each other in so many characters and appear to be so closely related that they may well be considered as falling within a single genus.

**Morphology and Staining**—All the members of the group are small ovoid bacilli with convex sides and rounded ends, there is no characteristic arrange-

ment, they are generally disposed singly, in pairs, in short chains, or in small groups. The most striking feature is their pleomorphism, which is most noticeable with *Past pestis*, least with the hæmorrhagic septicæmia bacilli. Though well marked on ordinary media, it is best brought out by cultivation on nutrient agar containing 3 per cent sodium chloride. The growth on this medium is poor, microscopically, as well as the usual ovoid or short bacillary forms, there will be found shadow forms, filamentous snake like forms, club forms, large yeast-like globules, and other irregular forms. *Past pestis* and *Past pseudotuberculosis* are larger and more ovoid than *Past aviseptica*, but their shape varies considerably with the medium on which they are grown. Generally speaking, the bacilli of all three species tend to be ovoid and to show bipolar staining when taken from smooth colonies, and to be more bacillary, filamentous, or pleomorphic, without bipolar staining, when taken from rough colonies, but many exceptions occur.



FIG 157—*Pasteurella pestis*

From an agar culture, 3 days, 37° C ( $\times 1000$ )

*Past pestis* and *Past aviseptica* are non motile. *Past pseudotuberculosis*, on the other hand, though usually non motile in cultures incubated at 37° C, is often motile in broth cultures grown for 18 hours at 20–22° C. The possession of motility by this organism is of considerable value in differentiating it from *Past pestis*, with which it may easily be confused (Arkwright 1927). The value of this test, however, is limited. Weitzenberg (1935), for example, made observations on 25 strains of *pseudotuberculosis*, and found that, though all were motile at room temperature, some were very poorly so, and had to be examined repeatedly before their motility

could be definitely established. A negative result, therefore, on a single examination cannot be regarded as conclusive. Both Levinthal (1930) and Weitzenberg (1935) have demonstrated the presence of flagella on *Past pseudotuberculosis*. The usual number appears to be 1–2, arranged at one or both poles.

In the animal body *Past pestis* may form a true capsule with a definite edge (Kitasato 1894). More often it is surrounded by a gelatinous envelope of ill-defined capsular material, which is soluble in weak alkalis (Rowland 1914a). The same envelope may be formed in artificial media, especially in 10 per cent inactivated horse serum broth incubated at 36° C. According to Schütze (1932a), it develops best at 37° C, poorly at 26° C, and not at all at 20° C. By serological methods it can be shown to contain a special antigen distinct from that present in the body of the organism. *Past aviseptica* may show an indefinite capsule in the animal body. From this capsule Hoffenreich (1928) has extracted a polysaccharide, which, though unable to give rise to precipitins on injection into rabbits, yet reacts to a high titre with a specific precipitating serum. Priestley (1936a, c) has shown that an envelope substance, apparently similar to that formed by

the plague bacillus, can be demonstrated in cultures of virulent, but not of avirulent strains of *Pasteurella septica*. It reaches its maximum development after 24 hours at 37° C, and then gradually disappears. Temperatures above and below 37° C tend to inhibit its formation. Antigenically it is distinct from the somatic substances.

Bipolar staining is very common and gives to the ovoid bacilli a characteristic appearance. The rod forms often stain irregularly, appearing as granular or barred forms. The reaction to Gram's stain is uniformly negative.

**Cultural Characteristics.**—Moderate growth occurs on the ordinary media. *Past pseudotuberculosis* and *Past atiseptica* grow fairly rapidly, giving a confluent growth on agar after 24 hours, *Past pestis*, on the other hand, develops more slowly, and gives a less abundant growth, often barely noticeable after this time. The agar colonies of *Past pestis* and *Past pseudotuberculosis* resemble each other in many respects, and are characterized by the effuse, clear or slightly granular, peripheral extension that occurs after 2 to 4 days' growth (Figs 158, 159), the



FIG 158—*Pasteurella pestis*

Surface colonies on agar 3 days, 37° C., showing differentiation and effuse edge ( $\times 8$ )



FIG 159—*Pasteurella pseudotuberculosis*

Surface colonies on agar, 24 hours 37° C., showing irregular granular surface and effuse edge ( $\times 8$ )

colonies of *Past pseudotuberculosis*, however, develop more rapidly, and are larger and more granular. With further incubation the central raised part of the colony may assume a ringed or draughtsman like appearance. On moist agar the colonies especially of *Past pestis*, are of viscous consistency and tend to adhere to the medium (Eastwood and Griffith 1914).

Colonial variants have been described for each member. From *Past pestis* Gotschlich (1912) obtained round, slimy, undifferentiated colonies which were poorly agglutinable and proved avirulent for rats, they subsequently reverted to the normal virulent type. Variant types, including smooth compact, small fringed, large irregular, and "sunflower" types, have been described by a number of other workers (see Bessonowa, Sémikoz, and Kotelnikow 1927, Pirie 1929, Burgess 1930, Bessonowa and Lenskaja 1931, Bhatnagar 1940a), but the exact form produced seems to be so influenced by environmental factors that it is probably better not to refer to these types by the terms rough and smooth. Kakehi (1915-16) described two variants of *Past pseudotuberculosis*, A was almost transparent and had a bluish, glimmering appearance, B was greyish white and opaque. A rendered broth turbid, B did not. Zlatogoroff and Moghilewskaja (1928a, b)

have similarly encountered two variants. On first isolation from the animal body, the organisms formed smooth colonies, consisting of short bacilli, which often showed well marked bipolar staining, on plates seeded from old broth cultures rough colonies developed, with a dull wrinkled surface and a lobate or crenated edge, morphologically these colonies consisted of larger, often longer, bacilli, which did not exhibit bipolar staining. These two variants likewise differed in their biochemical and serological characters (see below). In cultures of *Past lepreptica* De Kruijff (1921, 1922a, b, 1923) found two different types, Type D grew diffusely in broth, formed rather opaque, fluorescent colonies on serum agar, and was highly virulent for rabbits, Type G gave a granular deposit in broth, formed translucent bluish colonies with little fluorescence, and was completely avirulent. The D type gave rise to G variants, but the G type did not revert to D. A mucoid variant of intermediate virulence has been described for *Past lepreptica* by Webster and Burn (1926), while colonies similar in many respects to the D and G types have been recorded for *Past arseptica* by Anderson, Coombes, and Mallick (1929) Morch and Krogh Lund (1930), and Hughes (1930).

In broth *Past. pestis* causes little or no turbidity, but gives rise to a deposit of fine flocculi, with *Past pseudotuberculosis* there is no turbidity but a deposit of coarse flocculi, with *Past arseptica* there is a uniform turbidity with a powdery deposit (Zlatogoroff 1904). These differences are not entirely constant. A noteworthy feature is that whereas growth of all three organisms on an agar slope reaches its maximum in 2 to 4 days in broth growth continues for 7 to 10 days, or at room temperature, for several weeks. An old broth culture is almost clear, there is a heavy deposit which is difficult or impossible to disintegrate, and there may be a surface pellicle and ring, in cultures of *Past. arseptica* the deposit is viscous, in cultures of the other two it is usually floccular or membranous.



FIG 160—  
*Pasteurella*  
*muriseptica*

Surface colony of  
smooth type on  
agar 24 hours  
3 ° C (× 8)

If *Past pestis* is seeded into broth covered with melted butter or oil, and the flask is allowed to remain undisturbed in the incubator, growth occurs in the form of stalactites depending from the under surface of the droplets. This property is not peculiar to the plague bacillus, nor is it possessed by all strains of that species. *Past pseudotuberculosis* may likewise give a stalactite growth in broth.

None of the members liquefies gelatin, in a stab culture there is a surface layer, and a filiform growth extending to the bottom of the tube. In the case of *Past pestis* little feathery projections sometimes occur from the stab into the surrounding gelatin.

Potato is not a suitable medium. *Past. pestis* and *Past arseptica* give little or no growth (Kitasato 1894, Magnusson 1914, Tanaka 1926). *Past pseudotuberculosis* either gives no growth at all or else forms a thin, yellowish layer, which may later turn brown (Preisz 1894).

In bile salt media—MacConkey's liquid or solid medium—*Past pestis* and *Past pseudotuberculosis* give a slight but definite growth, which disappears in the course of 2 or 3 days, owing presumably to autolysis of the bacilli, *Past arseptica* fails entirely to grow.

Resistance—None of the members is highly resistant to inimical agencies. Broth cultures are killed by heat at 55° C., and by 0.5 per cent. phenol, within 15

minutes. Agar plate cultures exposed to sunlight are sterilized in 3 or 4 hours (Ogata 1897). Dried on threads and kept at room temperature in a desiccator the bacilli survive for not more than a few days. In bubo juice dried on a cover slip *Past. pestis* dies in under 4 days (Kitasato 1894), but in dry flea faeces kept at room temperature (66° F) it may survive for 5 weeks (Eskey and Haas 1940). The blood of animals dying from hæmorrhagic septicæmia remains virulent in the dried state for about 3 weeks, blood which is allowed to putrefy in a glass tube may remain virulent for 100 days (Ostertag 1908). Bacilli in cultures or infected organs kept in the ice chest may survive for months. According to Francis (1932) *Past. pestis* in an infected guinea pig's spleen kept in pure neutral glycerol at -15° C, may retain its virulence for years.

**Metabolism**—The bacilli have a wide range of growth. *Past. pestis* and *Past. pseudotuberculosis* can grow to some extent at very low temperatures—according to Tumansky and his colleagues (1935) even at 0° C. Their upper limit of growth is about 43° C. Different observers disagree about their optimum temperature for growth, but 30° C is generally considered to be the most favourable. Growth however, at both 24° C and 37° C is often nearly as good as at 30° C. According to Sokhey and Habbu (1943) *Past. pestis* grows in nutrient broth between -2° and +45° C, at 27 to 28° C—its optimum temperature—growth is five times as rapid as at 37° C. *Pasteurella septica* has a rather narrower range of growth and develops best at 37° C.

All the members are aerobes and facultative anaerobes. Working with *Past. leptoseptica*, Webster (1924a, 192b) and Webster and Baudisch (1925) found that the D or smooth variant would not grow in plain broth unless large numbers of organisms were introduced—about 100 000 per ml, whereas the G or rough variant grew if only a few organisms were introduced. But if a trace of rabbit blood or an iron compound with strongly catalytic properties was added to the medium or the partial pressure of oxygen was lowered mechanically, growth of the D variant occurred with the smallest inoculum. Schutze and Hassanain (1929) made similar observations on *Past. pestis*. They found that the difficulty of obtaining growth from small inocula on agar plates could be overcome by the addition of a small amount of blood or of 0.025 per cent sodium sulphite to the medium or by incubation under anaerobic conditions. Their conclusion that the organisms were sensitive to oxygen was confirmed by Wright (1934), who found that plague bacilli were destroyed fairly rapidly on the surface of agar plates if exposed at 37° C to a partial pressure of oxygen exceeding 1 per cent. Aerobic surface cultivation was however, successful if 0.1 per cent of blood, 10 per cent of serum or 0.05 per cent of sodium sulphite was added to the agar. Under these conditions growth was more profuse aerobically than anaerobically. For the growth of *Past. pestis* Rao (1939) found that three amino-acids, proline, phenylalanine, and cystine were required; glucose and lactate were the two best sources of carbon and energy with which to supplement media. Growth was stimulated by glycine, hæmatin, cozymase, thiamin and nicotinic acid. Hæmatin appeared to be essential for the aerobic growth of plague bacilli when these were present in only small numbers.

On horse blood agar plates there is no hæmolysis around the colonies but the whole plate is slightly cleared and browned. When a suspension of bacilli is incubated with sheep's red cells there is again no hæmolysis, but the oxyhæmoglobin is reduced to hæmoglobin. In this respect all members behave alike.

No true exotoxin is formed (Hadley 1918). Old broth cultures are, however,



very toxic to animals suggesting that endotoxins are liberated by the autolysis of the bacilli

**Biochemical Reactions**—(MacConkey 1908 Vourloud 1908 Magnusson 1914 Kakehi 1915-16 Besemer 1917 Brooks and Rhodes 1923 Pons 1925 Colas Belcour 1926 Csontos 1926 Tanaka 1926 Morch and Krogh Lund 1931)

*Past pestis* and *Past pseudotuberculosis* produce acid without gas in glucose maltose mannitol and salicin within 14 days salicin is not usually fermented for about 10 days Some strains of *Past pestis* fail to attack maltose and some strains of *Past pseudotuberculosis* ferment sucrose *Past pseudotuberculosis* is stated (Colas-Belcour 1926 Zlatogoroff and Moghilewskaja 1928a) to form acid from glycerol but so also do some strains of *Past pestis* (Bessonowa 1933) Bessonowa (1930) and Russo (1939) say that *Past pseudotuberculosis* ferments rhamnose while *Past pestis* does not Kauffmann (1932) however says that *Past pestis* may ferment rhamnose though its reaction in this sugar is inconstant *Past aviseptica* produces acid in glucose mannitol and sucrose some types also produce acid in maltose Tanaka (1926) found that *Past bubaliseptica* (from buffaloes) formed acid in lactose and clotted milk the same observation was made by Magnusson (1914) for a reindeer strain and by Besemer (1917) for a calf strain of *Pasteurella* We have encountered a strain of *Past lepiseptica* that fermented lactose and to a less extent salicin Newsom and Cross (1932) found differences in the fermentation of arabinose dulcitol and glycerol and Rosenbusch and Merchant (1939) were able on the basis of fermentation of xylose arabinose or both to divide 114 strains of *Past septica* of diverse origin into three sub-groups each of which showed a considerable degree of antigenic homogeneity In practice great care is required in determining the fermentation reactions since growth in sugar peptone water media is very poor

In plain broth *Past pestis* produces alkali the maximum production is not reached for 6 to 8 weeks (Bannerman 1908) in a litre flask of medium In peptone water containing 0.5 per cent glucose the hydrogen ion concentration reached in 7 days is pH 4.6-4.9 for *Past pestis* pH 4.6-4.8 for *Past pseudotuberculosis* and pH 5.6-6.1 for *Past aviseptica* (Otten 1926) That is to say the first two give a positive the last a negative methyl red reaction. But in peptone water containing 0.05 per cent glucose the final hydrogen ion concentration is pH 5.1-5.5 for *Past pestis* 7.0-7.3 for *Past pseudotuberculosis* and pH 5.8-6.1 for *Past aviseptica* After exhausting the sugar *Past pseudotuberculosis* would therefore appear to produce alkali more rapidly than the other two According to Zlatogoroff and Moghilewskaja (1928a) cultures of the rough variant form of *Past pseudotuberculosis* return to neutral more rapidly than those of the smooth variant form. It is not yet certain whether the different end reactions in glucose broth can be used for differentiating between *Past pestis* and *Past pseudotuberculosis* according to d'Aunoy (1923) the results obtained depend to some extent on the initial H ion concentration of the medium.

*Past aviseptica* forms indole and gives a negative M.R. reaction *Past pestis* and *Past pseudotuberculosis* form no indole and give a positive M.R. reaction *Past pestis* and *Past aviseptica* have no action on litmus milk *Past pseudotuberculosis* turns it slightly alkaline All members reduce nitrates form ammonia and a small amount of  $H_2S$  and give a positive catalase test According to our observations *Past pestis* does not reduce methylene blue whereas *Past aviseptica* and *Past pseudotuberculosis* are able to do so In confirmation of this Iwanowsky and Sassy

kina (1930) have found that saline suspensions of *Past pseudotuberculosis* reduce Schardinger's reagent (formolized M B solution) much more rapidly than those of *Past pestis*. *Past aviseptica* is stated to form phenol in peptone water (Bunzl-Federn 1891)

**Antigenic Structure**—Antigenically all the members of the group are closely related. An agglutinating serum prepared against any one of them is said to act not only on its homologous strain but to a less extent on the heterologous strains. It is stated that the relationship between *Past pestis* and *Past pseudotuberculosis* is very close, and that it is often impossible to distinguish between them by direct agglutination. *Past aviseptica* is not so closely allied, but nevertheless it may give a definite group reaction with sera prepared against either of the other members. This, however, has not been our experience, using low titre sera we have had no difficulty in distinguishing between the three species by direct agglutination.

According to Schütze (1932a), there are only two antigens in the plague bacillus, one corresponding to the envelope and the other to the somatic substance. The envelope antigen is developed best in cultures grown at 37° C and is heat labile, the somatic antigen is formed as well at 20° C as at 37° C and is heat stable. Both antigens may be present in avirulent as well as in virulent forms, though variants lacking the power to form the envelope substance have been described (Schütze 1939). Wats, Wagle, and Puduval (1939) find that organisms provided with an envelope agglutinate more rapidly than those containing only the somatic antigen, they differ also from these in forming larger flakes of varying size, and a voluminous deposit, which is easily homogenized by shaking. The antigenic structure of the flagellated *Past pseudotuberculosis* is more complex. Arkwright (1927) has shown that agar cultures incubated for 24 hours at 18 to 22° C contain a heat-labile H, and a heat-stable O antigen. The H antigen is apparently associated with the flagella and is destroyed by boiling for half an hour, though not by exposure to 56° C for a similar length of time, it agglutinates in the form of loose flocculi, the O antigen is apparently associated with the bacterial bodies is not destroyed by boiling for 1 hour, and agglutinates in the form of granules. Further work by Schütze (1928, 1932b), Zlatogoroff and Moghilewskaja (1928a), Kauffmann (1933), and Bhatnagar (1940b) seems to show that the flagellar antigen which is formed only in cultures grown at 26° C or lower, is common to all strains within the species, but that the somatic antigen is more complex. One somatic antigen is shared with the plague bacillus, another is closely related to the O antigen present in *Salmonella paratyphi B* and related organisms of the *Salmonella* group. In addition, there appear to be one or more type specific antigens that characterize individual strains or groups of strains. According to Bhatnagar (1940b) a serum prepared against the plague bacillus will agglutinate *Past pseudotuberculosis* by virtue of the antibody to their common somatic antigen, but a serum prepared against *Past pseudotuberculosis* will not agglutinate plague bacilli grown at 37° C because the somatic antigen is protected by the envelope substance.

The antigenic structure of the members of the *Pasteurella septica* group of organisms is still very uncertain. With regard to *Past muriseptica* we have found two distinct types, distinguishable either by direct agglutination or by absorption of agglutinins. The two types were further distinguished by the fact that one type fermented maltose and the other did not. Using the complement fixation reaction, Lal (1927) found that there was a cross reaction between different

members of the hæmorrhagic septicæmia group but that differences in the degree of fixation were usually sufficient to enable strains coming from one animal source to be separated from those from another. Cornelius (1929) by means of the agglutinin-absorption test, classified 17 out of 70 strains of *Pasteurella* of diverse origin into 4 groups—the remaining 9 strains defied classification. Very similar results have been recorded by Yusuf (1930) using the precipitation test. Pineslev's (1936a c) observations suggest that *Past. septica* resembles *Past. pestis* in forming a heat labile envelope antigen and a heat-stable somatic antigen. It differs from the plague bacillus in that the envelope antigen appears to be present only in virulent strains, though this requires confirmation—moreover the somatic antigen is not the same in all members of the species. Hoffenreich (1921) reported the isolation of a polysaccharide hapten that reacted to high titre with a specific precipitating serum. By means of the trichloroacetic acid technique Pirotsky (1930a c d) has now extracted from smooth and rough variants four different glycolipid antigens—judged by cross-precipitation tests one of these antigens was found to be related to the Vi antigen of the typhoid bacillus and another to the O antigen shared by *Salmonella typhi* and *Salmonella enteridis*. Unfortunately the relation between Pirotsky's glycolipid antigens and the envelope and capsular antigens is still far from clear.

Immune sera can be prepared for all the members by injection of rabbits or horses with living or dead organisms (Haffkine 1905). If living organisms are used, a weakly virulent culture should be chosen for the first few injections. The sera have prophylactic and to a less extent curative, properties for laboratory animals (Yersin *et al* 1890 Chamberland and Jouan 1906). Schultze's (1932c 1934) observations suggest that the protective power of a plague vaccine is largely dependent on the presence of the envelope antigen. Working with rats he found that a plague culture grown at 3 ° C—a temperature at which the envelope antigen is well developed—was considerably more potent for purposes of immunization than one grown at 26 ° C—a temperature which is less favourable for the formation of the capsular material. Moreover heating of a culture to 100 ° C for 15 minutes destroyed the envelope substance and rendered the vaccine useless while exposure to a temperature of 56 ° C for 30 minutes had no such deleterious effect. Sokolov and Maurice (1930) however working with mice found that cultures grown at 25 ° C were as effective as those grown at 3 ° C. According to Schultze (1939) the explanation of this discrepancy lies in the relative importance of the two antigens for the type of animal under study. In the protection of rats and guinea pigs a large part is played by the envelope antigen, but in mice this antigen appears to be of less importance.

It has been stated that it is possible, by the use of a *pseudotuberculosis* vaccine, to immunize rats and guinea pigs against infection with virulent plague bacilli (MacConker 1906 McCoy 1911 Report 1915) Zlatogoroff (1904) on the other hand was unable to immunize animals against plague with a *pseudotuberculosis* antigen, or against *pseudotuberculosis* with a *pestis* antigen. He also found that a specific anti-plague serum would protect guinea pigs against plague but not against *pseudotuberculosis*. Boquet (1931) was able by the use of avirulent bacilli to protect guinea pigs against infection with virulent *Past. pseudotuberculosis*.

As regards the hæmorrhagic septicæmic group most workers agree that a strain from one animal can be used to vaccinate against infection with strains from other animals (Chamberland and Jouan 1906 Magnusson 1914). A fowl-cholera

strain, *Past aviseptica*, for example, will protect mice against infection with a strain from swine plague or pleuropneumonia of calves. Likewise an immune serum prepared against one strain is said to protect against all other strains. Schriop (1908), however, obtained evidence of varietal differences in the species, according to him protection can be realized with certainty only by the use of monovalent sera (see also Priestley 1936b).

**Pathogenicity**—*Past pestis* and *Past pseudotuberculosis* cause disease in rodents, *Past pestis* is also pathogenic for man. The hæmorrhagic septicæmia group is pathogenic for a large number of animals and birds, but only very exceptionally for man (see Chapter 73).

The virulence of all three organisms is subject to considerable variation, and appears to be determined, at least in part, by the particular variant that has gained the ascendancy. The D variant of *Past leipseptica* is highly virulent the G variant comparatively avirulent. There is evidence that the smooth type of *Past pseudotuberculosis* is more virulent for guinea pigs than the rough type (Zlatogoroff and Moghilewskaja 1928a) though, according to Boquet (1937) this relationship is fortuitous. There are several reports on variations in virulence of the plague bacillus occurring under natural and experimental conditions (See Yersin, Calmette, and Borrel 1895, Report 1906, McCoy 1911, Rowland 1914b, Eberson 1917, Pirie 1929, Burgess 1930). Many of the statements are conflicting rendering it impossible at the moment to draw any definite conclusions on the relation of virulence to colonial appearance or antigenic structure. Further study of this problem by careful quantitative methods is required. For maintaining the virulence of the plague bacillus, Sokhey (1939b) recommends growing the organisms on 5 per cent rabbit blood agar for 4 days at 26°–32° C, sealing the tubes in the flame, and storing them at 4° C.

### Experimental Reproduction of Plague in Animals

Bubonic plague can be reproduced in rodents and monkeys by experimental inoculation. Dogs, cats, pigs, cattle, sheep, goats, and horses are difficult to infect; birds, with the exception of sparrows, are completely resistant. The disease is said to occur naturally in camels, but these animals are refractory to experimental inoculation (Zabolotny 1923). Even the rodents show great variation in susceptibility to infection. The less resistant members succumb rapidly, whereas the more resistant ones either fail to develop the disease, or else develop a subacute or chronic type. Spencer (1921) found in America that about 30 per cent of the rats from a plague free district were resistant to subcutaneous inoculation of *Past pestis*. According to Sokhey (1939a), the white mouse is the most susceptible laboratory animal, but Otten (1938) on the contrary maintains that the guinea pig, in spite of its greater weight, is more susceptible than the mouse.

Pneumonic plague has been produced in rats and also in marmots (*Spermophilus citellus*) by causing them to inhale cultures of *Past pestis* (Eberson and Wu Lien Teh 1917, Wu Lien Teh and Eberson 1917).

**RATS**—*Subcutaneous injection* of a very small number of virulent plague bacilli leads to death in 2 to 8 days. *Post mortem* there is necrosis and œdema at the site of inoculation, the regional lymph glands are swollen and surrounded by a hæmorrhagic infiltration of the subcutaneous tissue. Glands in other parts of the body are often congested and swollen, the spleen may be enlarged and dark red, the liver and lungs are hyperæmic, and sometimes a pleural exudate is seen. Bacilli are found in large numbers in the local

lesion and the nearest glands; they are usually irregular in size and shape, exhibit bipolar staining, and may show involution forms. They are often present in the spleen and in the blood. If the animal lives for a week, small, irregular, necrotic foci may be observed in the spleen and liver.

The German Plague Commission (Report 1899) had apparently no difficulty in infecting rats by the mouth, but other workers have not been so successful. They fed the animals with a drop of plague culture or with the cadaver of a rat that had died of plague. Death followed uniformly in 2 to 3 days. Post mortem, three types of lesion were found. (a) Most commonly enlargement and congestion of the submaxillary and suprahyoid glands, with the general picture of septicæmia. (b) Less often primary infection of the stomach and intestine, numerous punctiform hæmorrhages round the pylorus, swelling and hæmorrhagic infiltration of the lymph follicles and mesenteric glands, which were often quite large, and contained innumerable plague bacilli. (c) Quite frequently an aspiration pneumonia; the lungs showed inflammatory foci of varying size, containing large numbers of plague bacilli; the spleen was enlarged, and the liver hyperæmic.

The English Plague Commission had less success with feeding. Of the wild rats of Bombay only 38 per cent. were found to be susceptible when fed on the carcasses of dead plague rats. In the Punjab the proportion was nearly 70 per cent. The lesions found in rats infected by feeding were of a similar type to those in rats infected naturally. Two striking differences, however, were present. (a) In naturally infected plague rats the bubo is in the neck; a mesenteric bubo was not encountered in 5,000 examinations; in the case of fed rats the bubo is generally in the mesentery. (b) In naturally infected rats the stomach and intestines show no marked pathological changes: in rats infected by feeding well-marked lesions are found in the intestines—hæmorrhages in the stomach wall 3 per cent., congestion of intestines 27 per cent., enlargement of Peyer's patches 31 per cent.

The nasal mucosa and conjunctiva are favourable spots for inoculation in rats (Report 1899). A trace of infective material smeared over the conjunctiva proves fatal in 3 to 4 days. Post mortem, there is swelling of the cervical lymph glands, enlargement of the spleen, and frequently numerous hæmorrhages in the stomach and jejunum. The appearances are in fact similar to those of an animal dying after oral infection. Contamination of the nasal mucosa is frequently followed by an inhalation pneumonia.

The English Plague Commission (Report 1912, p. 287) were able to reproduce the lesions of chronic or—as they prefer to call it—of resolving plague in rats by inoculating large numbers of animals with small doses, and retaining the survivors for 3 weeks after inoculation. The chief lesions found were chronic buboes, necrotic areas in the spleen, and chronic abscesses in the spleen or more rarely the liver.

MICE.—The mouse reacts to inoculation in much the same way as the rat. After subcutaneous inoculation the septicæmia is very marked; the blood and internal organs swarm with bacilli. Infection can be accomplished by feeding, if a sufficiently large dose of a virulent culture is used.

GUINEA-PIGS.—Guinea-pigs are highly susceptible to plague, dying in 2 to 5 days after subcutaneous injection of a pure culture. Post mortem, there is a necrotic focus at the site of injection surrounded by intense congestion and œdema; the regional lymph glands are swollen and embedded in a bloody œdema; their interior is soft and necrotic. There is enlargement and congestion of the spleen, which is often studded with milium, soft, grey nodules up to 1 mm. in diameter, sometimes projecting above its surface, and containing large numbers of bacilli. The suprarenals may be congested (Yersin 1894). The liver may be peppered with tiny necrotic foci, and occasional small necrotic nodules are visible in the lungs. Sometimes there is a pleural effusion. Guinea-pigs can be infected by the cutaneous route. If the plague material is rubbed on the shaven skin of the abdomen, an inflammatory reaction appears in the neighbourhood, marked by a slight reddening and the formation of umbilicated pustules in which plague bacilli are present (Theodorou and Otto 1912). After a few days the regional glands swell and death occurs in 4 to 5 days. The post-mortem signs are similar to those after subcutaneous inoculation.

*Intraperitoneal* injection is fatal in 24 to 36 hours. Post mortem there is a rich fibrinous exudate containing enormous quantities of plague bacilli. Infection by the mouth, nasal mucosa and vaginal mucosa is not constant.

The animals are very sensitive to *conjunctival* infection. By *inhalation* or *intratracheal* inoculation it is possible to set up an acute primary pneumonia. Symptoms appear in 48 hours and death occurs in about 72 hours. At necropsy there is a confluent broncho-pneumonia with œdema or commencing necrosis of the lung tissue (Bessonowa, Kotelnikow and Semikoz 1927, Bablet and Girard 1933). However infection occurs the organisms sooner or later gain access to the blood stream. At post mortem they can be isolated from the blood, spleen, liver, lung and bone marrow. After bacteremia has developed the organisms may often be found in the bile, urine and less frequently in other excretions (Sémikoz, Bessonowa and Kotelnikow 1927).

RABBITS are less susceptible to plague than rats and guinea pigs but they can generally be infected by subcutaneous inoculation (Dreudonné and Otto 1912).

MONKEYS vary in susceptibility. The German Commission (Report 1899) infected *Macaca s. r. latus* by subcutaneous and intraperitoneal inoculation and by feeding. This species was not nearly so susceptible however as *Presbytis entell. s. (Semnopithecus entell. s.)* which succumbed after minute quantities of plague culture subcutaneously.

### Experimental Reproduction of Pasteurellosis in Animals

Most animals are susceptible to experimental infection with *Pasteurella*. On primary isolation a strain from one species of animal may prove of low infectivity for other species (Karlinski 1890) but this is by no means always true. Magnusson for example found that his reindeer organism was pathogenic to mice, guinea pigs, rabbits, sheep, dogs and pigeons. The most suitable animals for inoculation are the mouse, rabbit and pigeon.

MICE—Subcutaneous inoculation of a small quantity of a 24 hours broth culture proves fatal in 24 to 48 hours. Post mortem there may be local œdema and congestion with practically no other signs. Microscopically the bacilli are found in large numbers in the blood and viscera. If very few organisms are injected or a culture of relatively low virulence is used, the mouse does not die for 2 to 8 days or even longer. At necropsy there is a fibrino-purulent pericarditis, a layer of fibrin over the pleura, partial consolidation of the lungs and not infrequently a purulent exudate in the peritoneum. Bacilli are plentiful in the blood and organs. Intraperitoneal inoculation is more rapidly fatal.

RABBITS can be infected by subcutaneous, intraperitoneal, intravenous, intratracheal or intranasal inoculation. Death occurs in 2 to 5 days as a rule after intraperitoneal injection with lesions similar to those in mice. In addition there may be hæmorrhagic tracheitis (Magnusson 1914) and hyperemia of the kidneys and intestine (Poels 1886). Intranasal insufflation of the bacilli is often followed by snuffles or pleuro-pneumonia (Beck 1893, Webster 1924b, 1926) and sometimes by purulent otitis media (Smith and Webster 1925).

PIGEONS are very susceptible to intravenous or intraperitoneal, less so to intramuscular injection. Death occurs in 24 to 48 hours. Bacilli are abundant in the blood.

### Experimental Reproduction of Pseudotuberculosis in Animals

The term pseudotuberculosis is used to refer to a number of diseases that are caused by different organisms (see Chapter 73). We shall restrict ourselves here to the lesions due to *Past. pseudotuberculosis*. The experimental disease can be produced in guinea pigs, rabbits, rats, mice, and according to Pallaske (1933) to whom reference should be made for a detailed account of the lesions produced in cats, pigeons, canaries and turkeys. For laboratory purposes the guinea pig is the most suitable animal to study.

*Subcutaneous* or *intramuscular* inoculation of the guinea pig is followed by a disease which, depending on the dose and the virulence of the strain, may be acute subacute or chronic. The acute disease resembles plague and is fatal in a few days. The differential diagnosis can be made only by cultivation and a thorough study of the organism responsible. Macroscopically, the focal lymphatic glands tend to be more affected in plague than in pseudotuberculosis. The subacute disease proves fatal in about 2 weeks, and the chronic in 3 weeks or longer. Post mortem there is a caseous local swelling, the regional lymphatic glands are enlarged, and there are nodules varying in number size and degree of caseation, in the spleen liver and lungs. If the animal lives for 3 weeks or so, the nodules are usually very conspicuous. They are more or less spherical, greyish white in colour and 0.2 to 3.0 mm. in diameter, they project above the surface of the organ, and in the liver they show no particular localization for the free border as do the necrotic areas in rodent typhoid infection. Microscopically the bacilli are generally present in considerable numbers at the site of inoculation and in the regional glands, and they can be readily cultivated from all the lesions. The disease can also be reproduced by *feeding* which is the natural method of infection. Death occurs in 1 to 3 weeks. Nodules of varying size are found in the intestinal wall, the mesenteric glands are enlarged and often caseous, and nodules may be present in the spleen, liver and lungs.

**Classification and Identification**—The members of this group resemble each other very closely, between *Past pestis* and *Past pseudotuberculosis* the similarity is so great that the identification of a given strain is not always easy. Agglutination and precipitation may be of assistance especially if supplemented by the absorption test. The production of alkali in litmus milk by *Past pseudotuberculosis*, and its comparative harmlessness for white rats (Peport 1912 p 300) are two differential tests that are sometimes recommended. *Past pseudotuberculosis* is often motile in broth cultures incubated for 18 hours at 20–22° C whereas *Past pestis* is uniformly non motile. The greater rapidity and luxuriance of growth of *Past pseudotuberculosis* is usually very striking particularly if studied on nutrient agar plates incubated aerobically for 24 hours. Under these conditions *Past pestis* shows very slight confluent growth restricted to the first line of inoculation. Single colonies are rarely seen unless a strong reducing substance is present in the inoculum. On the other hand colonies of *Past pseudotuberculosis* reaching 0.5 or 1 mm. in diameter are usually evident over the whole plate. The haemorrhagic septicæmia bacilli can be differentiated from *Past pestis* and *Past pseudotuberculosis* by their fermentation of sucrose their production of indole and their negative methyl red reaction. It must be noted however that sucrose is fermented by certain strains of *Past pseudotuberculosis*. According to Brigham and Rettger (1930) *Past pestis* and *Past pseudotuberculosis* grow on potato at 20° C while *Past septicæ* does not. Between the haemorrhagic septicæmia strains of different animal origin, there appears to be no constant characteristic of diagnostic value. Most workers therefore agree that these bacilli form a single group though it appears possible using a careful technique, to make out antigenic differences between them. At present each member is given a specific name probably it would be better to call them all by one specific name such as *Pasteurella septicæ* and to indicate the animal origin where necessary.

A group of strains in many respects resembling *Past septicæ* but differing from it in others have been described by Jones (1921) Tweed and Edington (1930) Newsom and Cross (1932) and Rosenbusch and Merchant (1933). These strains are characterized by the production of  $\beta$  hemolysis on horse or rabbit blood agar failure to produce indole and non pathogenicity to mice and rabbits, most of

them appear to ferment maltose lactose, dextrin and inositol in addition to glucose mannitol and sucrose. It is very doubtful whether these strains form a true soluble hæmolysin, the clearing on a blood agar plate is insufficient evidence of this. If it should be proved that a true hæmolysin is formed, it may be necessary to divide the hæmorrhagic septicæmia group of organisms into a non hæmolytic and a hæmolytic sub group, and award each sub group a specific name.

We may note here that Dungal (1931) in Iceland, studying acute contagious pneumonia in sheep cultivated an organism which resembled the hæmorrhagic septicæmia group but differed from it in the following respects. It was Gram positive in young broth cultures and tended to be definitely rod shaped  $1-3 \mu$  in length, it failed to grow at  $22^{\circ} \text{C}$ , in gelatin or broth, it grew on MacConkey agar, it fermented maltose, it did not produce indole. It died out in 2 to 5 days in culture at room temperature and it was pathogenic to mice but not to rabbits or guinea pigs. The organism was said to be non hæmolytic differing in this respect from the hæmolytic group of organisms described by Newsom and Cross some of which were likewise isolated from sheep suffering from pneumonia. In Australia, Beveridge (1937) isolated a bacillus very similar to that described by Dungal, from sheep suffering from lesions of the digestive organs and lungs.

TABLE 50

	<i>Past pestis</i>	<i>Past pseudotuberculosis</i>	<i>Past septica</i>
Motility in 18 hour cultures at $22^{\circ} \text{C}$	—	+	—
Litmus Milk	— or slight acid	Alkaline	—
Sugars	Acid in glucose maltose mannitol and salicin	Acid in glucose maltose, mannitol and salicin some times in sucrose	Acid in glucose mannitol and sucrose sometimes in maltose
Indole	—	—	+
M R	+	+	—
<sup>1</sup> Methylene Blue Reduction	—	+	+
Growth on MacConkey	+	+	—
Pathogenicity to white rats	+	—	+

<sup>1</sup> Personal observations on a relatively few strains

Fedorova and Lalazarow (1935) described a *Pasteurella* like organism which they isolated from a spontaneous epidemic among mice in the outskirts of Astrakhan. The organisms were  $1-2 \mu$  long by  $0.5 \mu$  wide, Gram negative non motile, and capsulated. Acid was formed in lactose, mannitol and dextrin and acid plus gas in glucose. Sucrose was unchanged. No indole was formed. The organism was highly pathogenic for mice but had no effect on rabbits pigeons or rats injected by various routes. It is doubtful whether this organism should be placed in the *Pasteurella* group.



*Pasteurella pestis*

*Isolation*—Independently by Kitasato (1894) and by Yersin (1894).

*Habitat*—Parasite of rodents and man.

*Morphology*—Small, straight, ovoid bacillus,  $1.5 \mu \times 0.7 \mu$ , with rounded ends and convex sides, arranged singly, in short chains, or in small groups. Shows high degree of pleomorphism, especially in buboes and on 3 per cent. salt agar, there is every degree of variation in depth of staining, and clubs, shadow forms, snake-like filaments, coccoid forms, yeast like forms, and numerous others may be seen. Non-motile. Non-sporing. In the animal body a true capsule may be formed, in culture media there is often a slimy envelope around each bacillus. Shows bipolar staining, is Gram negative, and non-acid fast.

*Agar Plate*.—24 hours at  $37^{\circ} \text{C}$  Very small, 0.1–0.2 mm. in diameter, round, glistening, transparent, colourless, finely granular, umbonate colonies, with smooth or finely granular surface and an entire or delicately notched edge, differentiated into a raised centre and a flat periphery.

5 days at  $37^{\circ} \text{C}$  Larger, up to 4 mm. in diameter, with a raised, sometimes ringed, nearly opaque, greyish yellow centre and a flat or shelving, finely granular, translucent, greyish white periphery, consistency is butyrous or viscous, emulsifiability easy, sometimes a secondary ring of growth is seen. Variant colonies occur.

*Deep Agar Shake*.—5 days at  $37^{\circ} \text{C}$  Maximum growth at the surface, numerous round, transparent, colourless, punctiform colonies, visible with a hand lens, scattered throughout the medium.

*Agar Stroke*.—24 hours at  $37^{\circ} \text{C}$  Poor, slightly raised, translucent, greyish yellow, glistening growth, with a wavy or frosted-glass surface, and an irregularly lobate edge. Growth increases very little with subsequent incubation.

*Gelatin Slab*.—7 days at  $22^{\circ} \text{C}$  Good, filiform growth, confluent at top discrete below, extending to bottom of tube, and sometimes sending out little feathery projections into medium. Surface growth is raised, 5 mm. in diameter, with a slightly lobate edge. No liquefaction.

*Broth*.—24 hours at  $37^{\circ} \text{C}$  Moderate growth, little or no turbidity, a floccular or powdery deposit not disintegrating completely on shaking. Later the flaky deposit increases and may crawl up the sides of the tube, a delicate surface pellicle often forms. If butter or oil is floated on the medium, stalactites grow down from the under-surface of the droplets.

*Loeffler's Serum*.—24 hours at  $37^{\circ} \text{C}$  Fairly good, confluent growth, better than that on agar.

*Horse Blood Agar Plate*.—2 days at  $37^{\circ} \text{C}$  Colonies are similar to those on agar but show less tendency to differentiation and peripheral spread. No hæmolytic, whole plate is slightly cleared and browned.

*Potato*.—7 days at  $22^{\circ} \text{C}$  Usually a thin layer of growth.

*MacConkey Plate*.—24 hours at  $37^{\circ} \text{C}$  Very slight, effuse, confluent growth, just visible to the naked eye. Colonies disappear after 2 or 3 days owing presumably to autolysis.

*Resistance*—Fairly susceptible to chemical agencies. Killed by drying in a day or two, by heat at  $55^{\circ} \text{C}$ . in 5 minutes, by 5 per cent. phenol immediately, and by 0.5 per cent. phenol in 15 minutes. Agar plate cultures exposed to sun are sterilized in 1 to 5 hours. Cultures in the ice-chest may survive for months.

*Metabolism*—Aerobic, facultative anaerobe. Requires low O<sub>2</sub> potential for initiation of growth. Opt temp  $27\text{--}28^{\circ} \text{C}$ ., limits  $-2^{\circ}$  to  $+45^{\circ} \text{C}$ . Opt. pH 7.2, limits pH 5.0–9.6. Forms alkali in broth. Growth favoured slightly by serum, uninfluenced by glucose, partly inhibited by glycerol. No hæmolytic.

**Biochemical**—Acid, no gas, in glucose, maltose, mannitol, salicin, arabinose, xylose, and sometimes rhamnose and glycerol within 14 days L M unaltered or turned slightly acid, Indole —, M R +, V P —, Nitrates +,  $\text{NH}_3$  +, Methylene blue reduction —,  $\text{H}_2\text{S}$  very slight +, Catalase ++

**Antigenic Structure**—Appears to possess a heat labile envelope antigen and a heat stable somatic antigen The somatic antigen is similar to one of the somatic antigens in *Past pseudotuberculosis* Immune sera with protective and curative properties for animals can be prepared by injection of horses with living or dead bacilli

**Pathogenicity**—No true exotoxin formed. Virulence subject to considerable variation Causes plague in man and rodents. Experimental inoculation reproduces disease in mice, rats, guinea pigs, rabbits, marmots, ground squirrels, and other rodents, also in monkeys Dogs, cats, pigs, cattle, sheep, goats, and horses are difficult to infect Birds, with exception of sparrows, are completely resistant

*Subcutaneous inoculation of a 24 hours' broth culture into a mouse or guinea pig is fatal in 2 to 5 days* P M necrotic local lesion surrounded by congestion and oedema. Regional glands enlarged and embedded in bloody oedema, they are soft and necrotic on section Spleen firm, slightly enlarged and congested, may contain milky, soft, grey nodules, liver peppered with tiny necrotic foci Microscopically, bacilli found in abundance in local lesion and bubo, smaller numbers in spleen and heart's blood

### Pasteurella septica

**Isolation**.—First member of the *Pasteurella* group isolated by Kitt in 1878 *Past aviseptica* isolated by Pasteur in 1880

**Habitat**—Parasites of domestic and wild animals and birds.

**Morphology**—Very small,  $0.7-2 \mu \times 0.3-0.6 \mu$ , ovoid bacilli, with straight axis slightly convex sides, and rounded ends, arranged singly, in pairs, or in small bundles In smears from the animal body the organisms are regular ovoid, and evenly distributed, on agar cultures they are more rod shaped and often show pleo morphism Non motile, non-sporing May form a capsule in animal body, and an envelope in artificial media. Shows bipolar staining Gram negative and non acid fast

**Agar Plate**.—24 hours at  $37^\circ \text{C}$  Round, 0.5-1.0 mm in diameter, low convex, amorphous, grayish yellow, translucent colonies, with smooth, glistening surface and entire edge, consistency butyrous, emulsifiability easy

5 days at  $37^\circ \text{C}$  Up to 6 mm in diameter, differentiated into a brownish, finely granular, sometimes ringed or striated, nearly opaque centre and a clearer smooth, homogeneous, grayish yellow translucent periphery

**Deep Agar Shake**.—5 days at  $37^\circ \text{C}$  Thick surface growth, numerous punctiform, undifferentiated colonies scattered throughout medium

**Agar Stroke**.—24 hours at  $37^\circ \text{C}$  Moderate, confluent, raised grayish yellow, translucent growth, with glistening wavy or beaten copper surface and finely lobate edge.

**Gelatin Stab**.—7 days at  $22^\circ \text{C}$  Good, filiform growth, confluent at top discrete below, extending to bottom, raised surface growth 5 mm. in diameter, with crenated edge, no liquefaction.

**Broth**.—24 hours at  $37^\circ \text{C}$  Moderate growth with slight turbidity, and a slight powdery or viscous deposit Later the turbidity increases, and a heavy, viscous deposit forms, disintegrating partly on shaking but leaving irregularly sized wisp-like masses of growth in suspension. An incomplete surface pellicle forms with an inconspicuous ring growth.

**Loeffler's Serum**.—24 hours at  $37^\circ \text{C}$  Good confluent growth, similar to that on agar

*Horse Blood Agar Plate*.—2 days at 37° C. Good growth similar to that on agar, no hæmoly-sis but blood plate is slightly cleared and browned.

*Potato*.—7 days at 22° C. No visible growth.

*MucConkey Plate*.—5 days at 37° C. No visible growth.

*Resistance*.—Very susceptible to inimical agencies, killed by heat at 60° C. in a few minutes, by 0.5 per cent. phenol in 15 minutes.

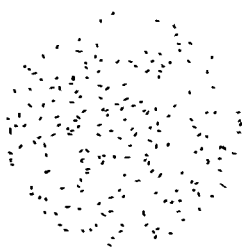


FIG 161.—*Pasteurella optica*.

From an agar culture 24 hours, 37° C. (× 1000)

*Metabolism*.—Aerobe, facultative anaerobe. May require low O-R potential on first isolation. Opt. temp. 37° C., limits 12°–43° C. Growth improved slightly by serum, uninfluenced by glucose, slightly inhibited by glycerol. No hæmoly-sis, but see p. 772.

*Biochemical*.—Acid, no gas, in glucose, mannitol, sucrose, and sorbitol within 14 days, one type forms acid in maltose. Some strains produce acid in arabinose, xylose, and glycerol. I.M. unchanged, Indole +, M.R. —, V.P. —, Nitrates reduced, NH<sub>4</sub> very slightly +, M.B. reduction —+, H<sub>2</sub>S +, Catalase +.

*Antigenic Structure*.—At least two types distinguishable by agglutination. Probably one envelope antigen, but more than one somatic antigen. Specific sera agglutinate *Past. pestis* and *Past. pseudotuberculosis* to a certain extent. Immune sera with protective and curative properties for animal can be prepared by injection of horses with living or dead bacilli.

*Pathogenicity*.—No true exotoxin produced. Virulence subject to alteration. Causes fowl cholera in birds. Other members of this group produce hæmorrhagic septic æmia in pigs, cattle, sheep, rabbits, mice, rats, reindeer, buffaloes, and other animals. Experimental inoculation reproduces the disease in these animals. Subcutaneous inoculation of a 24-hours' broth culture into a mouse proves fatal in 18 to 72 hours. P.M. local œdema and congestion, often no other signs, microscopically bacilli present in enormous numbers in blood and viscera. If a small dose is given and the animal does not die for 4 to 7 days, there is often a fibrinopurulent pericarditis, a layer of fibrin over the pleura, and partial consolidation of the lungs. Bacilli are numerous in blood and organs.

### *Pasteurella pseudotuberculosis*

*Synonym*.—*B. pseudotuberculosis rodentium*.

*Isolation*.—First observed by Malassez and Vignal in 1853, named *B. pseudotuberculosis rodentium* by Pfeiffer (1890) in 1889.

*Habitat*.—Parasite of rodents, particularly guinea pigs.

*Morphology*.—Small, pleomorphic cocco-bacillus varying greatly in length and shape. Some strains consist of regular ovoid or coccoid organisms, 0.8–2.0  $\mu$  × 0.8  $\mu$ , with convex sides, rounded ends, and straight axis, arranged singly. Other strains consist of rod-shaped organisms, 1.5–5.0  $\mu$  × 0.6  $\mu$ , with parallel sides, rounded ends, and straight or curved axis, arranged singly in groups, or in short chains. Long curved filaments are not uncommon (Fig. 162). Motile in broth cultures at 22° C. Non-sporing. Non-capsulated. Ovoid forms show bipolar staining, rod

forms show great irregularity of staining, the barred and granular type of staining is very common. Gram negative and non acid fast.

**Agar Plate**—24 hours at 37° C. Round 0.5–1.0 mm in diameter umbonate, granular, translucent greyish yellow colonies with dull finely granular or beaten copper surface and entire edge, butyrous consistency, easily emulsifiable, differentiated into a raised more opaque centre and a flat, clearer periphery with radial striation. A rough variant with an irregular surface and a crenated edge also occurs.

**Deep Agar Shale**—5 days at 37° C. Heavy surface growth. No colonies beneath surface.

**Agar Stroke**—24 hours at 37° C. Moderate, confluent raised greyish yellow translucent growth, with glistening wavy or beaten-copper surface and an irregularly lobate edge.

**Gelatin Slab**—7 days at 22° C. Good filiform growth, confluent at top, discrete below, extending to bottom of tube. Raised surface growth, 5 mm in diameter, with finely lobate edge. No liquefaction.

**Broth**—24 hours at 37° C. Moderate growth without turbidity and a viscous deposit disintegrating on shaking. Later the broth clears and a heavy flocculo membranous deposit forms partly disintegrating on shaking. Incomplete surface and ring growth.

**Loeffler's Serum**—24 hours at 37° C. Confluent growth not so good as on agar.

**Horse Blood Agar Plate**—2 days at 37° C. Good growth but colonies are more compact and less differentiated than on agar. No hæmolyysis. Whole plate is slightly cleared and browned.

**Potato**—7 days at 22° C. A thin yellowish membrane which subsequently turns brown.

**MacConkey Plate**—24 hours at 37° C. Very slight effuse confluent growth. Colonies disappear after a few days owing presumably to autolysis.

**Resistance**—Fairly susceptible to chemical agencies. Killed by moist heat at 60° C. in 10 minutes.

**Metabolism**—Aerobe, facultative anaerobe. Opt temp 30° C., limits 5–43° C. Growth uninfluenced by serum and glucose slightly inhibited by glycerol. No hæmolyysis.

**Biochemical**—Acid no gas in glucose maltose mannitol, salicin arabinose xylose rhamnose and glycerol within 14 days sometimes in sucrose. I.M. usually slight alkali formation, Indole —, M.R. +, V.P. —, Nitrates +, NH<sub>3</sub> + M.B. reduction ++, H<sub>2</sub>S + Catalase ++.

**Antigenic Structure**—Apparently contains (1) a heat-labile flagellar antigen which is formed only in cultures grown below about 25° C., (2) a heat stable somatic antigen similar to that of the plague bacillus (3) a heat stable somatic antigen similar to that in *Salmonella paratyphi B* (4) possibly other relatively strain specific somatic antigens. A plague serum does not protect animals against infection with *Pasteurella pseudotuberculosis*.



FIG 16'—*Pasteurella pseudotuberculosis*  
From an agar culture 24 hours 37° C. (× 1000)

*Pathogen city*—No true exotoxin formed. Virulence subject to alteration. Causes pseudo-tuberculosis in rodents especially guinea pigs. Experimental inoculation reproduces the disease in rodents. Subcutaneous injection of a 24-hour broth culture into a guinea pig is fatal in 1 to 3 weeks. P.V. caseous local swelling enlargement of regional glands and nodules in spleen liver and lungs. Microscopically the bacilli are numerous in the local lesion and gland.

## REFERENCES

- ANDERSON L. A. P., COOMBS M. G. and MALLICK, S. M. K. (1929) *Indian J med Res* 17 611  
 ARKWRIGHT J. A. (1927) *Lancet*, 13  
 BAUNOY R. (1923) *J infect Dis* 33, 291  
 BAELET J and GIRARD G. (1933) *C P Soc. Ed* 114, 41  
 BANNERMAN W. B. (1908) *Sci Mem med sanit Dep. India New Ser* No 33.  
 BECK, M. (1933) *Z Hyg InfektKr* 15 363  
 BESMER, A. M. (191) *J Bact* 2, 17  
 BESSONOWA A. (1929) *Re Microbiol Sarator* 336 (1930) *Zbl Bakt.* 119 32  
 BESSONOWA A. KOTELNIKOW G. and SEMIKOV, F. (1929) *C R Int Congr anthrop.* U.R.S.S p. 485  
 BESSONOWA A. and LE SKAJA G. (1931) *Zbl Bakt* 119 430.  
 BESSONOWA A., SEMIKOV, T. and KOTELNIKOW G. (1929) *Per Microbiol Sarator* 6, 4  
 BEVERIDGE, W. L. B. (193) *Am J vet. J* 13, 155  
 BHATTNAGAR, S. S. (1940a) *Indian J med R* 28, 1 (1940b) *Ibid* 28, 17  
 BONGERT (1901) *Z Hyg InfektKr* 3 449  
 BOQUET P. (1937) *Ann Inst Pasteur* 59 341  
 BRIGHAM G. D. and RETTGER, L. F. (1935) *J infect. Dis.* 56, 225.  
 BROOKS P. ST J. and PHOENES, M. (1923) *J Path. Bact* 26, 433  
 BUNZEL-FEDERN E. (1891) *Zbl Bakt* 9 87  
 BURGESS, A. S. (1920) *J Hyg Camb* 30 165  
 CHAMBERLAND and JOUAN (1906) *Ann Inst Pasteur* 20 81  
 COLAS BELCOUR, J. (1926) *C R Soc Biol* 94, 233  
 CORNELIUS, J. T. (1929) *J Path. Bact.* 22, 355  
 CSONTOS, J. (1926) *Zbl Bakt* 9 1 S.  
 DIEUDONNÉ A. and OTTO R. (1919) See Kolle and Wassermanns "Hdb. path. Mikroorg. IIte Abt. 1919 13, 4, 155.  
 DUNGAL, V. (1931) *J comp Path* 44, 196  
 EASTWOOD A. and GRIFFITH F. (1914) *J Hyg Camb* 14, 285.  
 EBERSON F. (1917) *J infect. Dis* 20 180  
 EBERSON F. and WU LIEN TEH (191) *J infect. Dis* 20, 10.  
 E. KEY C. R. and HAAS V. H. (1940) *Publ Hlth Bull. Wash.* No. 254  
 FEDOROVA T. and LALAZAROW G. (1935) *Per Microbiol Sarator* 14, 55  
 FRANCIS, E. (193) *Publ Hlth Rep Wash.* 4 1257  
 GIRARD G. (1936) *Ann Med Pharm* 34, 235  
 GOTTSCHLICH, E. (1919) See Kolle and Wassermann, Hdb. path. Mikroorg., IIte Abt., 1919, 13, 1, 16  
 HADLEY P. (1918) *J Bact* 3, 227  
 HAPPEINE, W. M. (1905) *Sci Mem med sanit Dep India New Ser* No 20.  
 HOFFENREICH, F. (1928) *Zbl Bakt.* 108, 8  
 HUGHES, T. P. (1930) *J exp Med* 51, 225.  
 IWANOWSKY V. and KASYKINA T. (1930) *Zbl. Bakt.* 11 535.  
 JONES F. S. (1921) *J exp Med* 34, 361  
 KAKEHI, S. (1915-16) *J Path. Bact* 20 269  
 KARLINSKI, J. (1890) *Zbl. Bakt.* 335  
 KATSEMAN, F. (193) *Z Hyg InfektKr.* 114, 97  
 KITASATO S. (1894) *Lancet*, ii, 48  
 KRIEF P. H. DE (1921) *J exp Med.* 33, 773 (1922) *Ibid* 35 561 (1925) *Ibid* 38, 309 (1923) *Ibid* 37 647  
 KUTSCHER (1894) *Z Hyg InfektKr* 18, 327  
 LAL, R. B. (1929) *Amer J Hyg* 561  
 LEVINTHAL, W. (1930) *Z Hyg InfektKr* 111 140  
 MCCONKEY A. T. (1908) *J Hyg Camb* 8, 335  
 MCCOY G. W. (1911) *Publ Hlth Bull Wash.* No. 43.  
 MAGNUSOV H. (1914) *Z InfektKr Haustiere* 15, 61  
 MALASSEZ, L. C. and VIGIAT, W. (1883) *Arch. Physiol norm. path.*, 3rd Series 2 369

- MORCH, J R and KROGH LUND, G (1930) *C R Soc Biol*, 105, 319, (1931) *Z Hyg Infektkr.*, 112, 471
- NEWSON I E and CROSS, I (1932) *J Amer vet med Ass*, 80, 711
- OGATA, M (1937) *Zbl Bakt*, 21, 769
- OSTERTAG, R (1908) *Z Infektkr Haustiere* 4, 1
- OTTEV, L (1936) *Zbl Bakt*, 98, 484, (1935a) *P I Seances Comite permanent Off int Hyg publ, Session d'Octobre, 1934* p 93, (1935b) *Bull Off int Hyg publ*, 29, 1542, (1938) *Meded Dienst Volksgezondh Ned Ind*, 27, 111
- PALLASKE, G (1933) *Z Infektkr Haustiere*, 44, 43
- PREIFFER, A (1890) *Zbl Bakt* 7, 219
- PIRIE, J H H (1929) *Publ S Afr Inst med Res*, 4, 203
- PIROSKY, I (1938a) *C R Soc Biol*, 127, 98, (1938b) *Ibid.*, 127, 966, (1938c) *Ibid*, 128, 346, (1938d) *Ibid.*, 128, 347
- POELS J (1896) *Fortschr Med*, 4, 388
- POYS R (1925) *Ann Inst Pasteur*, 39, 884
- PREISZ, H (1894) *Ann Inst Pasteur* 8, 231
- PRIESTLEY, F W (1936a) *Brit J exp Path*, 17, 374, (1936b) *J comp Path*, 49, 340 (1936c) *Ibid*, 49, 348
- RAO, M S (1939) *Indian J med Res*, 27, 75, 617, 833
- Reports (1899) Germ Plague Comm, Arb Reichsgesundhdsamt 16, 1 (1906) Eng Plague Comm *J Hyg Camb*, 6, 421, (1912) Engl Plague Comm, *J Hyg, Camb* 12, Suppl, p 287, (1915) *Ibid*, 14, Suppl p 754
- ROSENBUSCH C T and MERCHANT, I A. (1939) *J Bact*, 37, 69
- ROWLAND, S (1914a) *J Hyg, Camb.*, Engl Plague Comm, 13, Suppl, 418, (1914b) *Ibid* 440
- RUSSO, E (1939) *O Hospital* 16, 57
- SCHIEF, H (1908) *Zbl Bakt*, 47, 307
- SCHÜTZE, H (1928) *Arch. Hyg*, 100, 181 (1932a) *Brit J exp Path*, 13, 284 (1932b) *Ibid*, 13, 289, (1932c) *Ibid*, 13, 293, (1934) *Ibid*, 15, 200 (1939) *Ibid*, 20, 235
- SCHÜTZE, H and HASSAINEIN, M A (1929) *Brit J exp Path*, 10, 204
- SÉMÉROZ, F, BESSONOWA, A, and KOTELNIKOW, G (1927) *C R 1st Congr antipest U.R.S.S.*, p. 488
- SMITH, D T and WEBSTER, L T (1925) *J exp Med*, 41, 275
- SORHEY, S S (1939a) *Indian J med. Res.*, 27, 341, (1939b) *Ibid*, 27, 363
- SORHEY, S S and HABBU M K (1943) *J Bact*, 48, 25
- SORHEY, S S and MAURICE H (1935) *Bull Off int Hyg publ*, 27, 1534
- SPENCER, R R (1921) *Publ Hlth Rep, Wash*, No 36, p 2836
- TANAKA, A (1926) *J infect Dis*, 38, 421
- TUMANSKY, W, MÜLLER, M, BOKALO, A, WEDISTSCHEW, S, and SABININ A (1935) *Rev Microbiol Sarator*, 14, 128
- TWEED, W and EDINGTON, J W (1930) *J comp Path*, 43, 234
- VOITLOUP (1908) *Zbl. Bakt.*, 45, 97, 193
- WATS, R C, WAGLE P M, and PUDUVAL T K. (1939) *Indian J med Res* 27, 373
- WEBSTER, L T (1924a) *Proc Soc. exp Biol*, 22, 139, (1924b) *J exp Med*, 40, 109 117 (1925) *Ibid* 41, 571, (1926) *Ibid*, 43, 555
- WEBSTER, L T and DAUDISCH, O (1925) *J exp Med* 40, 473
- WEBSTER, L T and BURN, C G (1926) *J exp Med*, 44, 343 359
- WEITENBERG R (1935) *Zbl Bakt* 133, 343
- WRIGHT, H D (1934) *J Path. Bact*, 39, 381
- WU LIEH TEH, and EBERSON, F (1917) *J Hyg, Camb*, 16, 1
- YERSIN (1894) *Ann Inst Pasteur*, 8, 662
- YERSIN CALMETTE, and BORREL. (1895) *Ann Inst Pasteur*, 9, 589
- YUSEF, H S (1935) *J Path Bact*, 41, 203
- ZABOLOTVY, D (1923) *Ann Inst Pasteur*, 37, 618
- ZLATOGOROFF, S J (1904) *Zbl Bakt*, 37, 345, 513 654
- ZLATOGOROFF, S I and MOCHILEWSKAJA, B I (1928a) *Ann Inst Pasteur*, 42, 1615 (1928b) *C R Soc Biol.*, 99, 506

## CHAPTER 33

### HÆMOPHILUS

#### DEFINITION —*Hæmophilus*

Minute rods, sometimes almost coccid, sometimes thread like, may be highly pleomorphic. Usually non motile. Non sporing. Gram negative, non-acid fast. On first isolation dependent for growth on some factor, or factors contained in blood or in plant tissues. Some species retain this dependence after prolonged cultivation on laboratory media. Some species are obligatory aerobes, or grow very poorly under anaerobic conditions. All known species appear to be obligatory parasites, inhabiting particularly the upper respiratory tract, and most of the described species or types are pathogenic.

Type species. *H. influenza*.

SINCE the isolation and description by Pfeiffer (1892, 1893) of the bacillus which, though not the primary causal organism of influenza, is closely associated with that disease, several other small Gram negative bacilli have been described, which share with it certain characteristic growth requirements. In their final report on classification and nomenclature, the American Committee (see Winslow *et al* 1920) grouped these species together under the generic name of *Hæmophilus*. The generic definition suggested in the Committee's report opens the door more widely than insistence on a close similarity in behaviour to the type species would allow, and such species as the Bordet-Gengou bacillus of whooping cough, the Morax-Axenfeld bacillus of angular conjunctivitis, and Ducrey's bacillus of soft sore, have been included, by certain writers within this generic group. Such an extension of the term "hæmophilic bacilli," whether in the form of a generic name or as a convenient appellation for a characteristic bacterial group, has been opposed by Kristensen (1922) and by Fildes (1923). It seems clear that their objection is valid if the character from which the name is derived is to retain a decisive differential significance. To include all the species referred to, it would be necessary to define the genus on some other basis, with a sub-group characterized by the particular growth requirements that *H. influenza* displays. There is no reason why we should not do this.

The two most important growth factors in blood which determine the hæmophilic nature of Pfeiffer's bacillus have been identified, the one as a co-enzyme, the other as an iron-containing pigment which presumably supplies the tetra pyrrole compounds necessary for the synthesis of cytochrome and related substances (see Chapter 3). But if we confine the term hæmophilic bacilli to strains that need one or both of these substances for growth, we may be excluding organisms which can synthesize the growth factors, but which are nevertheless closely related. Moreover the existence of organisms which synthesize small, sub-optimal amounts of these growth factors provides a link between the definitely hæmophilic bacilli

and those, like the bacillus of whooping cough, for which the growth factors are not essential as nutrients

Ducrey's bacillus in any event is *hemophilic* on the narrower definition, but it is doubtful whether the Morax Axenfeld bacillus would be included in this genus by any such redefinition, and we have therefore described it in Chapter 37, together with others that cannot at present be assigned to any named bacterial genus. The Bordet Gengou bacillus cannot, we think, be dealt with in the same way. It resembles *H. influenzae* so closely in morphology, in habitat and in many other ways, that it would certainly be placed in close association with it by any systematic definition that did not rely exclusively on a narrow nutritional criterion.

In 1911, Ferry in the United States described a short bacillus that he had isolated from the respiratory tract of dogs in the early stage of distemper, to this organism he gave the name of *Bacillus bronchicanis*. A similar organism had been described as early as 1896 by Galli Valerio, by Tartakowsky (1897-98) in 1898, by Strada and Traina in 1900 as *B. pneumoniae caninarum* by Martin in 1900 as *B. pulmonum glutinosus*, and by Selter in 1906 as *B. causepticus mobilis*. Ferry, however, was the first to study it fully. Later Ferry (1912, 1913) found the same bacillus in a guinea pig epizootic, and in monkeys and rabbits. He therefore changed its name to *B. bronchisepticus*. This organism has in recent years been included in the *Brucella* group, whose type species *Br. melitensis*, it resembles in both individual and colonial morphology, and in its inability to ferment carbohydrates. However, the conspicuous degree of antigenic similarity between *B. bronchisepticus* and *H. pertussis* that has become evident in the past seven years demands a reconsideration of its classification. Antigenic relationships alone do not establish a taxonomic relationship, but in this case other resemblances are sufficiently good to warrant provisional inclusion of *B. bronchisepticus* in the group containing *H. pertussis*. Thus, *B. bronchisepticus* produces in guinea pigs lesions similar to those produced by *H. pertussis* in rabbits and puppies (Smith 1913, Mallory and Horner 1912, Mallory, Horner and Henderson 1912). Both *H. pertussis* and *B. bronchisepticus* are natural pathogens in the upper respiratory tract, and, according to Rhea (1915) *B. bronchisepticus* lesions in rabbit lungs are similar to the lung lesions of human whooping cough. And finally the general cultural resemblances between *H. pertussis* and *B. bronchisepticus* are as close as those between *B. bronchisepticus* and *Br. melitensis* (Chapter 34). We have therefore provisionally included both the Bordet Gengou bacillus and *B. bronchisepticus* in the genus, and have emended the generic definition accordingly (see Lwoff 1939). It should, however, be pointed out that *H. bronchisepticus* grows very much more readily than any other member of the *Hæmophilus* group—so much so that it has been classified by some workers with the paracolon bacilli. Such a difference, taken together with the free growth on plain nutrient agar and the possession of motility, renders the classification of this organism in the *Hæmophilus* group necessarily tentative.

**Morphology**—*H. influenzae*, as originally described by Pfeiffer (1893) and as most commonly seen in strains recently isolated from cases of influenza, is a short rod, so short as to be almost coccoid. It is very small, 1-1.5  $\mu$  by 0.3-0.4  $\mu$ , with rounded, sometimes rather pointed ends. In some cultures these coccoid bacillary forms are the only forms seen. More usually, among these predominating short forms, are found a proportion of longer bacilli, and a few long thread forms. In other cultures, the coccoid bacilli may be relatively scanty, or altogether absent,



and longer and somewhat stouter rods may predominate. Other strains, again, may present an entirely different picture, the bacilli being thin, long, wavy or curved and sometimes lying together in tangled masses. In films prepared from strains showing any of these diverse morphological types, but especially those which show some proportion of thread forms, it is not unusual to encounter large, spherical swollen bodies, often attaining a diameter of  $2-3\mu$ , or even more. These are sometimes attached to the end of a thread, sometimes laterally, and sometimes apparently at the end of a short lateral stalk (Wade and Manalang 1920, Kristensen 1922). Another form which is occasionally met with consists of a long thread with an enormous fusiform swelling, situated centrally or towards one end.

This morphological diversity raises a problem of considerable difficulty from the point of view of classification. Any one of the types we have referred to may predominate in a single strain, and two strains may yield a microscopical picture so different that it is difficult to believe that we are dealing with a single bacterial

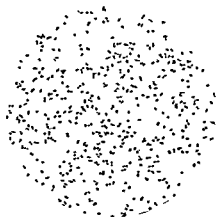


FIG 163.—*H. influenzae*

From 24-hours' culture on Fildes agar, showing typical cocco-bacillary forms ( $\times 1000$ )



FIG 164.—*H. influenzae*

From 24 hours culture on Fildes agar, showing short and long bacillary forms ( $\times 1000$ )

species. The only test of the real significance of such morphological differences is their constancy, and this test is not so easy to apply as might at first sight appear. It is quite certain that many strains maintain their morphological individuality over long periods of artificial cultivation, involving numerous successive subcultures, and Dible (1924) regards these morphological characters as sufficiently well differentiated and sufficiently stable, to justify the recognition of several different varieties, or types. Many of those who have studied this group, on the other hand, have rejected morphological criteria as a basis of classification within the group, on the following grounds. If a large series of strains is carefully examined the variation in form is not found to be discontinuous. In a small sample of strains it is easy to obtain an appearance of discontinuity; but, unless one classes all strains which depart from the typical cocco-bacillary form in a single heterogeneous group, it is not possible to define a limited number of morphological categories, to which all strains can be definitely assigned. Between the minute, short bacilli, and the long tangled threads, there exists a long range of intermediate forms. Moreover, while some strains are morphologically homogeneous, others display, in the

same culture, a heterogeneous mixture of coccobacilli, rods, and long threads. Finally, though many strains retain their morphological characters unchanged for weeks, months, or longer periods, others show quite definite changes in form after a few cultures. Wollstein (1915) has noted the frequency with which the coccobacillary type acquires the power of forming threads in artificial culture, and there has been a very general tendency to discard the old conception of a para influenza bacillus, differentiated on purely morphological grounds (Pfeiffer 1893), in favour of the tentative view that such differences, unless associated with other well marked characters, must be disregarded by the systematist (Kristensen 1922). Smith (1931) records a careful study of a series of strains isolated from the human nasopharynx. In almost every instance morphological variation occurred after a varying period of cultivation in artificial media, the most usual change being from the short coccobacillary form to longer bacilli, or to curved or jointed filaments. This change was in general associated with a change in colony form (see below).



FIG 165—*H. influenzae*

From 24 hours' culture on Fildes' agar, atypical form showing long curved bacillary forms ( $\times 1000$ )



FIG 166—*H. influenzae*

From 24 hours' culture on Fildes' agar showing thread forms, and large spherical bodies ( $\times 1000$ )

There is general agreement that it is possible to make a rough classification of strains into two groups, "typical" and "atypical" on the basis of morphology alone, and there are indications that, apart from certain strains isolated from cases of meningitis (see Cohen 1909, Ritchie 1910, Henry 1912, Wollstein 1915) the typical morphology predominates among strains isolated from pathological conditions. The studies of Pittman (1931) which will be discussed more fully in relation to the problem of antigenic structure, have emphasized the possible importance of this distinction. But the evidence at present available suggests that typical and atypical morphology are associated with a change presenting many analogies to the smooth  $\rightarrow$  rough type of variation, rather than permanent characters serving to differentiate stable varieties or types.

The organisms of this genus form no spores and are, with the exception of *H. bronchisepticus* non flagellated. They are usually described as non-capsulated, but since Pittman's (1931) description of a "smooth" capsulated form of virulent *H. influenzae* it is now clear that capsulation is not an uncommon feature within the group.

*H. influenza* stains with some difficulty, and many of the ordinary bacteriological dyes are unsuitable for this purpose. Dilute carbol-fuchsin applied for 5 to 15 minutes usually gives satisfactory results. All species within this genus are frankly Gram negative and non acid-fast.

It may be added that it is impossible to differentiate any given strain of *H. pertussis* from *H. influenza* on grounds of morphology alone. If, however, large samples of strains are compared, certain modal differences may be observed. *H. pertussis* displays a more constant morphology than *H. influenza*, and there is a marked tendency towards the predominance of the short oval form of cell. Longer bacillary or thread forms may occur, but they are relatively uncommon.

**Cultural Characters. Growth Requirements.**—Since certain nutritional requirements provide the criteria by which this genus has been defined, and also the basis on which many of the species included within it are differentiated from one another, it will be convenient to discuss this aspect of their behaviour before dealing with their type of growth, enzymic activities, antizenic structure or pathogenicity. The most characteristic feature of the hemophilic bacilli is their failure to grow in the absence of certain factors which are present in blood. The ability of the influenza bacillus to grow on blood agar, and its inability to grow on agar, with or without the addition of serum, or other native protein, has been noted by all workers from Pfeiffer onwards. Grassberger (1897) described another phenomenon—that of satellitism—which is highly characteristic of *H. influenza*. In cultures on blood agar plates, streaked with sputum or with bronchial secretion, he noted the appearance of relatively large colonies of the influenza bacillus, with a slightly granular central portion. These large colonies (1 mm. or more in diameter) always developed in the immediate vicinity of a colony of staphylococcus. Studying this phenomenon in greater detail, Grassberger streaked agar plates with a suspension of *H. influenza* mixed with a small quantity of blood, and then inoculated the central portion of the plate with a trace of a pure culture of *Staph. aureus*. After 24 hours' incubation such plates showed a well-defined zone of colonies of *H. influenza*, surrounding each colony of staphylococcus. A similar result was obtained with *Staph. albus*, *Staph. citreus*, and certain other chromogenic micrococci. These observations have been repeatedly confirmed (see Davis 1921, Kristensen 1922, and Fig. 167).

The inability of *H. influenza* to grow on serum agar indicates that some constituent of the red cells is essential for growth; and it was at first assumed that this constituent was hæmoglobin itself. More detailed study, however, showed that the addition to an agar medium, prepared with water or with peptone solution, of pure crystallized hæmoglobin does not suffice to ensure growth (Ghon and von Preyß 1904, Thalmer 1914, Davis 1917, Olsen 1920, Fildes 1921). It would appear (Fildes 1921) that the growth promoting substance derived from the blood pigment is methæmoglobin, or hæmatin, rather than hæmoglobin itself. Hæmatin is more active in this respect than the other derivatives which have been tested; while hæmatoporphyrin is inactive (Fildes 1921). There is, however, another factor which comes into play, besides the presence of some suitable iron-containing pigment. Davis (1917) showed that *H. influenza* required for its growth the presence of two distinct substances: one contained in, or derived from, hæmoglobin; the other present in the tissues of various plants and animals, and synthesized by most bacterial species other than *H. influenza*. This second factor he likened to a vitamin. This substance, as compared with the factor provided by blood pigment,

is relatively thermolabile; it is inactivated by heating to 120° C for 30 minutes. Both substances are present in blood. The label "X factor" is generally applied to the substance present in blood pigments, the label "V factor" to the relatively thermolabile substance provided by animal or vegetable tissues or by most bacterial cells (see also Thjotta 1921, Thjotta and Avery 1921, Davis 1921, Fildes 1922, 1923, 1924, Kristensen 1922, Valentine and Rivers 1927).

Pittman (1935) brought evidence to show that V factor was closely concerned in oxidation reduction processes of the growing cell. With regard to X factor, early observations (Olsen 1920, Fildes 1921) suggested that its growth promoting activity was correlated with its ability to act as a peroxidase, but not all peroxides promote the growth of *H. influenzae*, and certain iron compounds without peroxidase activity will act as X factor (Baudisch 1932). All such iron compounds that have been tested in this respect have shown catalase activity (see Davis 1921, Webster and Baudisch 1925, Bourn 1927, Baudisch 1932, Knight 1936). *H. influenzae* grows poorly under anaerobic conditions, but Kopp (1927-28), Eirund (1929) and Anderson (1931) have recorded the anaerobic growth of certain strains. In these conditions, the organism grows in the absence of X factor. These facts together suggest that X factor is closely associated with the aerobic respiration of the bacillus.

It is now apparent from the work of the Lwoffs (Lwoff and Lwoff 1937a, b, c) that V factor is one of two co dehydrogenases, di- and tri phosphopyridine nucleotide (see Chapter 3), and that the X factor is haem, which, they conclude, is normally utilized by *H. influenzae* for synthesis of cytochrome, cytochrome oxidase, catalase and peroxidase.

These observations clearly provide an explanation of the satellitism described by Grassberger. The staphylococci, and many other organisms of greater synthetic ability than the hemophilic bacilli, synthesize the co dehydrogenase, which diffuses into the medium and stimulates the growth of bacilli that require it. They explain, too, the anaerobic growth of *H. influenzae* without the help of X factor, since in the absence of molecular oxygen there is no need for that part of the cytochrome system which protects the organism against the inhibitory effect of oxygen (see Chapter 3).

It should be noted that Hoagland and his colleagues (1942) have found in blood an unidentified factor which stimulates the growth of *H. influenzae* in the presence of optimum amounts of X and V factors.

Among the species included within this genus, besides *H. influenzae* itself, are (1) the bacillus associated with conjunctivitis, described by Koch (1887) and Weeks (1887), and commonly known as the "Koch Weeks bacillus", (2) the bacillus isolated by Friedberger (1903) from the prepuce of dogs, named by him *B. haemoglobinophilus canis* and now known as *H. canis*, (3) the organism isolated by Shope (1931) (see also Lewis and Shope 1931) from swine influenza and named by him *H. influenzae suis*, (4) the causative organism of whooping cough, described by Bordet and Gengou (1906) and named by them *B. pertussis*, now known as *H. pertussis*, (5) the causative organism of soft sore described by Ducrey as the probable cause of soft chancre (see Chapter 79) and named *H. ducreyi* by Lwoff and his colleagues (Lwoff and Pirofsky 1937, Lwoff 1939), (6) the bacilli described by Rivers (1922a) as *H. para influenzae*, associated with acute pharyngitis and bacterial endocarditis in man, and (7) the causative organism of broncho pneumonia in rodents and dogs, described by Ferry (1912-13) as *B. bronchisepticus* and now

named *H. bronchisepticus*. The nutritional requirements of most of these species have been studied in some detail by various observers with results that may be summarized as follows.

With regard to the Koch Weeks bacillus the statements in the literature are somewhat contradictory (see Kristensen 1929) thus, some authors have stated that this organism grows on ascitic agar and hydrocele fluid agar. Later studies by Fildes (1923) and by Knorr (1924) however show quite clearly that the strains isolated by them from cases of muco-purulent conjunctivitis, behave in exactly the same way as *H. influenza* and require both the X and the V factor.

*H. canis* is unable to synthesize the X factor and is therefore dependent on hæmatin or some similarly acting substance. It can however synthesize the V factor and hence it does not show the phenomenon of satellitism but can itself

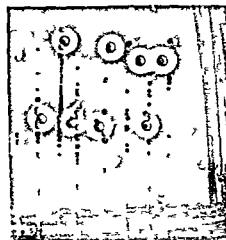


FIG. 167

Showing satellite growth of *H. influenza* in the neighbourhood of hemolytic colonies of *Staph. aureus* on blood agar

induce the formation of satellite colonies of *H. influenza* (see Friedberger 1903, Krage 1910, Odaira 1911, Rivers 1922, Kristensen 1922, Fildes 1923, Valentine and Rivers 1927). *H. dyscrea* resembles *H. canis* in its growth requirements (Lwoff and Pirotsky 1937). Khairat (1940) has described a hæmophilic bacillus from a human endocardial lesion having the same X and V factor requirements as *H. canis*.

*H. influenzae-suis* requires both the X and V factors for its growth resembling in this as in almost all other respects the human influenza bacillus (Lewis and Shope 1931).

*H. para-influenzae* requires V but not X factor for growth. It shows the phenomenon of satellitism. *H. pertussis* and *H. bronchisepticus* differ from the

other species within this genus in being capable of growth in the absence of both the X and V factors. For primary isolation of *H. pertussis* Bordet and Gengou (1906) employed an agar medium containing blood, glycerine and potato extract and this medium with various minor modifications, is still used for this purpose. Even on first isolation *H. pertussis* and *H. influenza* show differences in their nutritional requirements. The former grows well on media containing large amounts of blood and vegetable extract poorly on media containing the X and V factors in the absence of other blood or tissue constituents. The latter grows better on Fildes or Levinthal's medium than on media of the Bordet-Gengou type (see Gundel and Schluter 1933). On subculture in the laboratory these differences become more marked. *H. pertussis* can readily be trained to grow on serum agar and with slightly more difficulty on ordinary agar. There is however general agreement that on such media the organism rapidly loses its natural virulence (see below). These relations are clearly shown in Table 51.

TABLE 51

SHOWING GROWTH OF CERTAIN BACTERIAL SPECIES IN PEPTONE WATER, WITH AND WITHOUT X AND Y FACTORS

	Peptone Water			
	Alone	With X factor	With Y factor	With X and Y factors
<i>H. influenzae</i>	0	0	0	++
Koch Weeks bacillus	0	0	0	++
<i>H. canis</i>	0	++	0	++
<i>H. d. icereys</i>	0	++	0	++
<i>H. para influenzae</i>	0	0	++	++
<i>H. pertussis</i>	+	++	++	++
<i>H. bronchisepticus</i>	+	++	++	++

It may be convenient at this point to indicate the various media that are, at the present time employed in the study of the hæmophilic bacilli

Ordinary blood agar is by no means a satisfactory medium from this point of view far better results are obtained with media in which the red cells have been broken up and their modified contents distributed throughout the medium. The well known 'chocolate' agar, prepared by adding blood to melted agar raising the mixture to the boiling point for 3 minutes and then preparing slopes or plates from the chocolate coloured mass is a considerable improvement on the ordinary blood agar plate, but it shares the disadvantage that the medium is opaque.

The medium devised by Levinthal (1918) has the great advantage of being colourless and transparent. It is prepared by adding 5 per cent of defibrinated rabbit or human blood to melted agar in a flask and raising it to the boiling point over a flame with several shakings. The precipitate of coagulated blood and serum protein is allowed to settle, and the clear supernatant fluid is carefully decanted, or may be filtered through sterile glass-wool. The medium may be for safety sterilized by a further short heating but must not be subjected to prolonged sterilization in the steamer.

Fildes (1920) has introduced a peptic digest of blood which is preserved with chloroform and may be added to broth, or melted agar as required. This medium which is transparent and has the colour of ordinary broth or agar, gives copious growths of *H. influenzae* and inhibits the growth of many other organisms. It is admirably suited for the primary isolation of the influenza bacillus. The ability to support the optimal growth of hæmophilic bacilli is not a property of blood from all species of animals. For example on horse blood media, *H. pertussis* loses its smooth characters more readily than on media made with human or sheep blood (Toomey and Takacs 1938). The influenza bacillus may be even more susceptible to species variations in blood. Thus Arumwede and Kuttner (1938) describe thermolabile substances inhibiting the growth of *H. influenzae* and *H. para influenzae* in sheep goat, bovine and human blood but not in the blood of the rat, rabbit or guinea-pig.

For primary culture from such a source as the nasopharynx advantage may be taken of the selective action of penicillin which inhibits the growth of Gram positive cocci and of diphtheroid bacilli but has almost no action on *H. influenzae* or *H. pertussis* (see Chapter 77).

With regard to other conditions of growth the optimum temperature for *H. influenzae* is in the neighbourhood of 37° C. The minimal temperature for growth lies between 20° and 25° C. The same conditions hold for other hæmophilic bacilli. There is general agreement that *H. influenzae* grows far better under aerobic than

under anaerobic conditions. Statements with regard to its ability to develop under strictly anaerobic conditions are somewhat contradictory (see Kristensen 1922). Fildes (1921) states that *H. influenza* gives good initial anaerobic growth on a suitable medium but quickly dies out.

**Cultural Reactions** **Type of Growth.**—The type of colony given by *H. influenza* on solid media varies widely with the kind of medium employed. On blood agar it forms tiny transparent pin point colonies sometimes flat and tending to become confluent sometimes more convex and with less tendency to confluence. On a more favourable medium such as Levinthal's agar and especially Fildes agar the colonies are far larger. After 24 hours' incubation they attain a diameter of 0.5–0.8 mm. They are circular in outline, raised and dome-shaped with a slightly splayed-out entire edge. The surface is usually smooth, the colony is translucent, there is little differentiation and the growth emulsifies easily (Fig. 168).

On further incubation, and in many cases during the first 24 hours of growth the colony becomes differentiated into a central portion with a granular or contoured surface, an intermediate flattened portion, and a sharply levelled periphery.



FIG. 168—*H. influenza*

Colonies on Fildes agar after 24 hours ( $\times 8$ ).

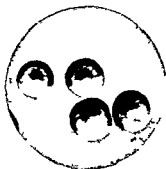


FIG. 169—*H. influenza*

Colonies on Fildes agar after 48 hours ( $\times 8$ ).

with a narrow splayed-out edge. Between the 24th and 48th hours there is usually a considerable enlargement of the colonies which may attain a diameter of 1–1.5 mm. This increase in size results in the formation of a flatter colony retaining a raised central boss, sometimes smooth, sometimes granular or contoured (Fig. 169). Some colonies may be from the start flatter and more granular, others may remain raised, conical and smooth with little central differentiation. Kristensen (1922) lays considerable stress on such colonial differences especially those which develop on blood agar and apparently regards them as more important than cellular differences in morphology in distinguishing typical from atypical strains. Smith (1931) in the study referred to above noted a definite but not absolute correlation between morphology and colony form. The morphologically typical strains tended to give on Fildes agar smooth colonies with little differentiation. The morphologically atypical strains or variants tended to produce a more granular colony, with earlier and more considerable differentiation.

The smooth strains described by Pittman (1931) have distinctive colonial characters. Levinthal's agar gives rather better differentiation than Fildes. Smooth strains give relatively large colonies, sometimes attaining 3 mm. in diameter. They

are slightly opaque, and viewed by obliquely transmitted light they are iridescent. This iridescence is, perhaps, their most characteristic property. The surface is smooth and slightly mucoid in appearance. The edge is entire.

These "smooth" strains are described by Pittman as readily giving rise to "rough" variants producing colonies of varying granularity and differentiation, the organisms composing them being in the form either of short bacilli, or of longer rods or threads. The rough variants were never capsulated.

The relation of Pittman's "smooth" strains to the strains isolated by other workers from infective conditions, or from the normal respiratory tract, raises a problem in terminology which is at the moment difficult to solve. Her observations have been confirmed, in whole or in part, by several subsequent workers, and there can be little doubt that the strains she has differentiated correspond to a form of *H. influenza* that is frequently associated with acute infections in man. If, however, we accept Pittman's colonial differentiation as a criterion separating "smooth" from "rough" strains, we must include among our "roughs" many, probably the majority, of those strains that give, on Fildes' or Levinthal's agar, colonies of the type that earlier observers had generally regarded as "smooth," and that consist of morphologically typical cocco bacillary organisms. These difficulties may be resolved if we accept three forms of *H. influenza*, the mucoid capsulated M form with a characteristic specific soluble substance, an S form without a capsule, and R forms (Chandler, Fothergill and Dingle 1937, 1939) (see p. 799). The balance of evidence is, we think, in favour of accepting Pittman's nomenclature, but this question will be discussed more fully in relation to antigenic structure.

*H. canis* forms colonies that are at first indistinguishable from those of *H. influenza*, but as they grow older they become larger and distinctly more opaque.

*H. pertussis*, when grown on Bordet Gengou medium, also forms colonies that, during the early stages of growth, may resemble those of *H. influenza*. But when incubation is prolonged beyond 24 hours the colonies become larger, more opaque, and greyish in colour, a form that is never assumed by *H. influenza*. They are also smoother, more shining and more distinctly dome shaped. The combination of slight opacity, greyiness of hue, and shining surface, gives them an appearance that has not inaptly been compared to that of a bisected pearl. They have also been compared to drops of mercury, but this overstates their metallic appearance. A confluent row of colonies has been compared to an "aluminum streak," and this simile again is not inapt. As already noted, *H. bronchisepticus* grows much more readily than other members of the group, forming quite well developed colonies on plain nutrient agar within 24 hours.

In liquid media, such as Fildes' broth, the majority of strains of *H. influenza* give rise to a uniform turbidity, with or without a slight powdery deposit. Some, on the other hand, give a flocculent deposit, with a varying degree of turbidity of the supernatant fluid. There is, as would be expected, a correlation between morphology and type of growth in a fluid medium. Cocco bacillary strains give a uniform turbidity. Many of those showing long bacilli, or twisted and convoluted threads, give flocculent growths.

*H. canis* gives a diffuse growth with a slight deposit. So does *H. pertussis*.

**Resistance.**—*H. influenza* is killed by exposure to a temperature of 50–55° C for 30 minutes. *H. canis* and *H. pertussis* behave similarly.

**Hæmolysin Production.**—Before considering the fermentation of carbohydrates, or other substrates, it will be convenient to discuss the hæmolytic activity of the



influenza bacillus, since this character serves to divide the species into two distinct types.

Pritchett and Stillman (1919) noted the occurrence, among a large sample of cultures of hæmophilic bacilli isolated from cases of influenza and from normal persons, of a small proportion of strains which produced a well-defined zone of hæmolysis on blood agar. These strains were morphologically of the bacillary or thread type. They were studied in greater detail by Stillman and Bourn (1920), and their occurrence has been noted by many subsequent workers. Kristensen (1922) studied several strains of these hæmolytic bacilli, and noted that some of them seemed less dependent on the presence of hæmoglobin than *H. influenza*. On the other hand, the majority of his strains showed well marked satellitism, thus demonstrating their dependence on the V factor. Fildes (1924) found that 13 of 14 hæmolytic strains grew in the presence of the V factor alone. In morphology these strains are, for the most part, definitely atypical, showing numerous threads, and coarser bacillary forms than are commonly encountered in *H. influenza* itself. The colonies produced by these strains tend to assume the characters which Kristensen regards as atypical, being more opaque and more friable than the typical form. Another striking characteristic of these hæmolytic strains is their tendency to die out in subculture—a character which has been noted by subsequent observers (Dible 1924). Kristensen notes that the power to cause hæmolysis is maintained unaltered by those strains which survive artificial cultivation for considerable periods, and that there is no tendency for other strains to acquire this property. Dible (1924) studied 67 strains of hæmophilic bacilli isolated from the nasopharynx of normal persons, and found 14 of them to be hæmolytic. Five of these strains resembled the coccobacillary form of *H. influenza*, with the single exception that the bacilli were a little larger and rather more definitely bacillary—the other 9 were atypical, in forming larger colonies, giving a flocculent growth in broth, or departing widely from the coccobacillary form. Valentine and Rivers (1927) report that the majority of these hæmolytic strains require the V factor only for their growth, while a minority require both the X and V factors, a proportion of non hæmolytic strains of hæmophilic bacilli require the V factor only. Valentine and Rivers proposed the name *H. parainfluenza* for the hæmophilic organisms requiring only V factor, irrespective of hæmolysin production. It is clear from their descriptions that organisms resembling Pfeiffer's bacillus in cultural characters and growth requirements may or may not form hæmolysin. We propose to adopt their nomenclature, and distinguish *H. parainfluenza* from *H. influenza* by X and V factor requirements and sub-divide each if necessary into hæmolytic and non hæmolytic varieties (see Miles and Gray 1935).

**Biochemical Activities.**—The study of the fermentation reactions of *H. influenza*, and of other hæmophilic bacilli, has been retarded by the difficulty experienced in preparing a medium which allows of copious growth, and has, at the same time, the transparency and absence of colour which are essential, if changes in hydrogen ion concentration are to be detected by the usual methods. Some of the media which have been devised within recent years are, however, well adapted for this purpose.

Levinthal (1916) added various carbohydrates to the agar medium which he devised, tinted it with litmus, and tested the fermentative ability of several strains of *H. influenza*. He noted acid production from glucose, but not from levulose, lactose, mannitol or maltose. Messerschmidt, Hundeshagen and Scheer (1919), using a similar technique, noted slight acid production in glucose but not in mannitol, lactose, or saccharose.

Stillman and Bourn (1920) employed a liquid medium prepared by adding an extract of boiled rabbit blood to peptone water, and carried out a careful series of tests on 119 strains of *H. influenza* and 29 hæmolytic strains. More than 90 per cent. of the 119 non hæmolytic strains produced acid from dextrose and galactose, and reduced nitrates, 73 per cent. produced acid from levulose, and about 50 per cent. from maltose, saccharose

and dextrin; no strain fermented mannitol or lactose; 53 per cent produced indole. Of the 29 hæmolytic strains, all fermented dextrose and reduced nitrates, the majority fermented maltose and saccharose, 3 fermented galactose, 10 levulose, and 15 dextrin, none fermented mannitol or lactose, 3 produced indole, and 4 formed gas. It would appear that the hæmolytic, as compared with the non hæmolytic strains, ferment maltose and saccharose more frequently, galactose and levulose less frequently, and seldom produce indole; but the number of hæmolytic strains examined was small. Stillman and Bourn specifically note that they obtained irregular results when they carried out repeated tests on the same strains. Fildes (1924) notes that the hæmolytic strains studied by him fermented glucose, saccharose, and maltose; but not lactose, dulcitol, or mannitol. The non-hæmolytic strains of *H. influenzae* which he examined did not ferment any of these sugars.

Kristensen (1922) carried out a considerable number of fermentation tests, but obtained almost entirely negative results. It seems probable that these were due to an unsatisfactory technique.

Dible (1924), using a technique which did not differ essentially from that employed by Stillman and Bourn, obtained results which he regarded as sufficiently sharp and constant to afford a basis for a tentative grouping of his strains, though he notes that, of 25 strains which were retested after 8 months, 9 showed changes in their fermentation reactions. In 8 cases this change involved a loss of the power to ferment one or more carbohydrates, in the remaining instance a strain, previously inactive, was found to ferment glucose. It may be noted that, of 14 hæmolytic strains, 9 fermented glucose and levulose, none galactose, 8 saccharose, and 6 maltose, while none formed indole. Of 6 non hæmolytic strains, which Dible excludes from the species *H. influenzae* on account of their bacillary or thread like morphology, 4 fermented glucose, 4 levulose, 5 galactose, 4 saccharose and none maltose, while none produced indole. Of 45 strains which showed the typical minute bacilli and coccobacilli, 38 fermented glucose, levulose and galactose, none fermented saccharose or maltose, while 16 produced indole. Dible's results thus tend to confirm those of Stillman and Bourn with regard to the frequency of saccharose fermentation, and infrequency of indole formation, among the hæmolytic as compared with the non hæmolytic strains.

In regard to the relation between morphology and fermentation reactions among the non hæmolytic strains, Smith (1931) records observations on 143 strains isolated from the nasopharynx of normal persons. There was no clear cut fermentative separation between morphologically typical and atypical strains, but, in conformity with the results recorded by other workers, it was found that the typical strains showed a more restricted enzymic activity than the atypical. Thus, 22.6 per cent of the atypicals fermented saccharose, as compared with 4.3 per cent of the typicals. The correlation between typical morphology and ability to form indole was further confirmed, 63.9 per cent of the coccobacillary strains were indole producers, as against 18.0 per cent of the morphologically atypical strains.

Later work with para-influenzal strains makes it clear that the hæmolytic influenzal strains referred to above and the strains isolated in an epidemic of pharyngitis by Lamont (1928) had the biochemical reactions of *H. para-influenzae*. Miles and Gray (1938) found that all of 12 hæmolytic strains of this organism were alike in fermenting dextrose, levulose, maltose and sucrose, and in producing no indole. The reaction of 9 non hæmolytic strains of *H. para-influenzae* were variable, though all fermented dextrose and sucrose and two were indole positive.

On the basis of hæmolysin production, indole production and other fermentative reactions, taken in conjunction with morphology and dependence on the X and V factors, we may, then, recognize several different types of *H. influenzae*.

(1) Typical *H. influenzae*—requiring both X and V factors, showing a predominantly coccobacillary morphology, not producing hæmolysin, usually showing a restricted range of enzymic activities, particularly in failing to ferment saccharose, and usually producing indole.

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### The Antigenic Relationships of the Hæmophilic Bacilli.

Taking first *H. influenza*, as a species, the peculiarity that has emerged from most of the recorded attempts at serological analysis is its extreme antigenic heterogeneity

By direct agglutination with 20 antisera, followed by absorption tests where necessary, Park, Williams and Cooper (1918) could find only four identical pairs among 160 strains Valentine and Cooper (1919) record a similar experience Among 10 strains isolated at autopsy, tested against the 10 homologous antisera, no two were identical Among 73 miscellaneous strains tested against 18 antisera no two were identical Among 54 strains isolated from a group of marines, and tested against 18 antisera, 2 strains from different individuals were identical It was noted in this group that strains isolated from the same individual on different days were usually, but not always, identical Among 28 strains isolated from the inmates of an orphan asylum, there was one pair of identical strains Of 6 strains isolated from the members of a single family, all of whom had contracted influenza at about the same time, no two strains were identical

This extreme heterogeneity, as judged by agglutination tests, has been amply confirmed by numerous workers (Rivers and Kohn 1921, Yabe 1921, Anderson and Schultz 1921, Cooper *et al* 1921, Povitsky and Denny 1921, Kristensen 1922, Knorr 1924, Izuka 1938, and others) Izuka, for example, records more than 50 different agglutinating types among 249 strains isolated from sick and healthy persons

The actual significance of these earlier observations has been rendered very doubtful by the observations of Pittman (1931)

Among 97 strains of influenza bacilli isolated from various sources, she noted 15 that produced colonies of a characteristic "smooth" type (see above) All these 15 strains were isolated from sources, or under conditions, which indicated that they were playing a pathogenic role In addition to forming a characteristic colony, the bacilli of these "smooth" strains were found to be capsulated When tested by agglutination at 37° C these 15 strains were found to fall into two antigenic types A and B, one containing 12 strains, the other 3 This specificity was not apparent if the reactions were carried out at a higher temperature, a possible reason being the loss of the bacterial capsules It was also found possible to separate from these 15 strains a soluble specific substance, apparently carbohydrate in nature, and presumably associated with the capsule Precipitin tests carried out with this material gave the same antigenic grouping as the agglutination tests carried out at 37° C These smooth strains, in artificial culture, readily gave rise to non capsulated rough variants, usually with a bacillary or filamentous morphology The rough variants no longer produced the soluble specific substance, nor did they conform to the antigenic grouping of the smooth parent strains These findings were confirmed in their essential points at least, by several subsequent workers (see Dochez *et al* 1932, Wright and Ward 1932, Platt 1937) Later work has shown that characteristic "smooth" strains are commonly found among bacilli isolated from infections of the meninges Among respiratory strains, either from infected or healthy persons, the "smooth" types are less common Mulder (1937) for example, found 7 in 90 sputum strains, and Platt (1937) 16 in 86 nasopharyngeal strains Respiratory strains tend to be serologically heterogeneous, whereas "smooth" meningeal strains tend to be homogeneous (Fothergill and Chandler 1936, Wilkes Weiss 1937, Platt 1937) The homogeneity is not complete Pittman has divided these capsulated smooth strains into six serological types, a, b, c, d, e and f Type b occurs with the greatest frequency, but all are rare in the normal upper respiratory tract (see, e.g., Silverthorne *et al* 1943)

As in the pneumococci, type specificity depends on a polysaccharide component in the capsule of the organism (Platt 1937, Alexander and Heidelberger 1940) If we follow Chandler, Fothergill and Dingle (1939) and designate capsulated forms as M or mucoid forms, then both the S and R forms, which have no type specific capsular substances, display extreme antigenic heterogeneity According to Platt (1939) individual non

capsulated strains appear to possess a relatively specific protein component P, and a protein component M which is common to most strains of *H influenzae*. It may be noted that several earlier workers found that complement-fixation tests gave evidence of the presence of an antigenic relationship that was not revealed by agglutination (Wollstein 1919, Biesing and Weichbrodt 1920, Kristensen 1922).

It is clear that the change that Pittman describes as occurring in her smooth strains falls within the definition that we adopted for the  $S \rightarrow R$ , or the  $M \rightarrow S \rightarrow R$  variation in Chapter 8 the loss of the surface antigen which determines type specificity in the normal, virulent form of a bacillus. Pittman's capsulated forms are apparently the most virulent pathogenic type of *H influenzae*, they are isolated commonly from the meninges in one of the severest of human infections caused by the organism, and Gordon, Woodcock and Zinnemann (1944) report that meningeal infections with strains not among Pittman's types are less severe. They are also the most mouse virulent of the observed varieties (Fothergill and Chandler 1936, Chandler *et al* 1937, 1939, Raettig 1940). The implication that most of the strains isolated from the nose and throat of normal persons are in an intermediate or a "rough" phase, must, therefore, we think, be accepted. We must also, it would appear, accept the view that the antigens that dominate the rough forms in this bacterial species are more heterogeneous than those present in the normal smooth phase, a finding that differs from that recorded for most other groups.

It will be noted that, under this definition, all the strains referred to in preceding sections as "atypical" and many, probably the great majority, of those referred to as "typical," would be classed as rough variants. The antigenic structure of the para-influenza bacilli has not yet been submitted to any special study, but it seems very unlikely that any of them would fall into Pittman's "smooth" category. Whether they are in any way antigenically different from the non-hæmolytic rough forms we do not know. Miles and Gray (1938) found an antigenic relationship between a proportion of the strains of non-hæmolytic *H para-influenzae* they studied.

The Koch Weeks bacillus, in its serological relationships as in all its other characters, appears to be indistinguishable from *H influenzae*. Knorr (1924) has found that different strains of this organism show marked antigenic heterogeneity, while some strains are identical with certain strains of the influenza bacillus.

A small sample of strains of *H influenzae-suis* examined by Lewis and Shope (1931) showed the same type of antigenic heterogeneity that is encountered among the ordinary strains of human influenza bacilli. Comparison with a few strains of *H influenzae* of human origin did not reveal any example of antigenic identity, though there was some overlapping in cross-agglutination tests. Similar findings are recorded by Harchenbauer (1934). These observations were made before the publication of Pittman's findings, so that there was no differentiation between smooth and rough strains.

We have as yet no information in regard to the antigenic relationships of *H canis*.

*H pertussis* differs from *H influenzae* in that all recently isolated smooth strains appear to belong to a single antigenic type. Moreover, it would seem that all strains, when first isolated from the body on an optimal medium, are in the smooth state. The behaviour of these strains on artificial culture raises points of considerable interest.

Bordet and Slesawyk (1910) noted that recently isolated strains of *H pertussis*, grown on the Bordet-Gengou medium, all agglutinated with a serum prepared against any one

of them. Strains that had been trained to grow on agar, however, failed to agglutinate with the sera prepared against the recently isolated strains, and sera prepared against the agar strains failed to agglutinate the strains grown on the Bordet-Gengou medium. These observations were confirmed and extended by Bordet (1912). The change in antigenic structure was ascribed to the medium, but it was noted that a similar change might occur on a blood containing medium after repeated subculture. Most observers have confirmed Bordet's findings that all recently isolated strains belong to a single serological type (see Kristensen 1922, 1927). A few have recorded the existence of two different types among strains maintained permanently on a blood containing medium (Krumwiede *et al.* 1923), but there is little doubt that such findings have been due to the slow occurrence of an antigenic variation that takes place more rapidly when smooth strains are grown on an unsuitable medium. Leslie and Gardner (1931) made a careful study of 32 strains of *H. pertussis*, none of which had been regarded as rough variants. They found that these strains fell into four different antigenic groups, to which they refer as Phases I, II, III and IV. Of 20 recently isolated strains 18 fell into Phase I, and 2 were intermediate between Phase I and Phase II. Of 7 laboratory strains that had been maintained on an egg medium, 3 were in Phase III and 4 in Phase IV. Of 5 other laboratory strains, one was intermediate between Phases II and III, 3 were in Phase III and 1 in Phase IV. Studies by later workers (Shibley and Hoelscher 1934, Toomey *et al.* 1935) have been in general agreement with Leslie and Gardner's findings, though they regarded the various phases as arbitrary stages in the course of an S  $\rightarrow$  R variation, depending on the amount of Phase I antigen on the bacillary surface (see also Toomey, Takacs and Ranta 1936, Toomey and Takacs 1937). Flosdorf, Dozons and Kimball (1941) on the other hand, find, like Leslie and Gardner, that the phases differ qualitatively, and suggest the following antigenic structure for three of the four phases they studied.

Phase	Major antigens	Minor antigens
I .	a	b c
III . .	b	c or d
IV . . .	c, d	

They also described a new Phase X, related to III and IV. Taking these studies as a whole it seems safe to conclude that *H. pertussis*, in the form in which it exists in the tissues or in recent cultures on Bordet-Gengou medium, belongs to a single, homogeneous antigenic type, but that an S  $\rightarrow$  R variation occurs somewhat readily, even in cultures kept on a blood containing medium, and very readily on less favourable media. This variation is apparently step like, so that intermediate stages exist between the normal smooth form, which corresponds to Leslie and Gardner's Phase I, and the fully developed rough form which corresponds to their Phase IV. It was noted by Leslie and Gardner that there is no very obvious and striking colonial difference between the rough and the smooth forms, nor any constant and measurable difference in salt sensitivity, though strains in their Phase III and Phase IV are, on the average, rougher in colonial appearance and less stable in saline suspensions than strains in Phase I or II. The S form is capsulated (Lawson 1933). The capsular substance, which determines the agglutination of the S form by smooth antisera, is readily removed by washing (Miller 1937). Flosdorf, Kimball and Chambers (1939) and Flosdorf and Kimball (1940a, b) have studied this soluble agglutinin extensively. It removes agglutinating antibodies from smooth antisera. It may be liberated by sonic vibrations. It is non-toxic and induces agglutinins in the rabbit and stimulates pertussis immunity. By tryptic digestion of *H. pertussis*, Cruickshank and Freeman (1937) obtained a carbohydrate-containing water soluble fraction capable of inducing active immunity to experimental infection in mice. It is probable that this fraction is the same as the "agglutinin" of later workers, mixed perhaps with some toxin. Smolens and Mudd (1943) obtained a large yield of agglutinin by acid extraction of the bacilli.

Eldering and Kendrick (1937, 1938) and Bradford and Slavin (1937) isolated an organism from cases of whooping cough which differed from *H. pertussis* in producing definite



hemolytic on Bordet-Gengou medium, in growing more profusely and developing a brownish pigment on nutrient agar and in producing a large amount of catalase. The strains isolated were antigenically homogeneous, and cross-agglutinated with Phase I *H. pertussis* and *H. bronchisepticus*. Like *H. pertussis* the organism was serologically characterized by a readily extractable non-toxic and stable agglutinin, having a minor antigenic component in common with *H. pertussis* (Floedorf Bondi and Dozois 1941 Bondi and Floedorf 1943). This organism is now generally known as *H. parapertussis*.

The antigenic relationship of the agglutinogens of these three organisms is paralleled to some extent by an antigenic relationship of their toxins (see below). There are cultural resemblances between the three and all are associated with infections of the lung in the higher mammals. These facts justify the provisional inclusion of *Br. bronchiseptica* in the group containing *H. pertussis* and we have accordingly implemented the suggestion of Eldering and Kendrick and of Evans and Matland (1939) and renamed *Br. bronchiseptica* *H. bronchisepticus* (see also Watanabe 1935).

It is clear that, as far as whooping cough is concerned, any bacillary material used for whooping cough inoculation must be derived from organisms in Phase I so treated that the very soluble agglutinin is not removed during the preparation of the vaccine, it may also be necessary to include in the vaccine the major antigens of *H. parapertussis*.

Finally, we may note that such comparative tests as have been performed show little if any antigenic relationship between *H. pertussis* and *H. influenza* (Odaira 1911, Shiga *et al.* 1913 Winholt 1915, Olmstead and Povitzky 1916 Kristensen 1922, Schluter 1936).

#### Pathogenicity and Toxin Production.

The probable role of *H. influenza* in human influenza, which is now known to be a virus disease is considered in Chapter 74. It is a common cause of sinusitis, alone or in association with the pneumococcus, an occasional cause of meningitis, almost always in children, and a rare cause of ulcerative endocarditis. The Koch Weeks bacillus has been isolated from epidemics of conjunctivitis in many parts of the world, children being mainly infected. As, however, there is no known method by which this organism can be distinguished from *H. influenza*, it seems unnecessary to regard it as a different species.

*H. para-influenza* is occasionally associated with acute pharyngitis, it is a rare cause of ulcerative endocarditis, though probably less rare than *H. influenza* itself (Miles and Gray 1935) and it is occasionally found in infected wounds and sinuses.

There appears to be little doubt that *H. ducreyi* is responsible for soft chancre, and for the buboes which are sometimes associated with the primary lesion.

*H. bronchisepticus* is essentially parasitic giving rise to lesions in the respiratory tract of dogs, monkeys, guinea pigs and other laboratory animals. It is occasionally found in the nasopharynx of man. It appears to be a secondary invader in dogs suffering from distemper, being frequently responsible for the pulmonary complications of the disease (McGowan 1911, Laudlaw and Dunkin 1926). Spooner (1933) found it playing a similar role in a spontaneous distemper like disease of ferrets.

*H. pertussis* is the cause of whooping cough, and as such is one of the more important human pathogens. *H. parapertussis* appears to be responsible for a minority of cases of whooping cough. Unlike *H. influenza*, *H. pertussis* seems seldom to play a harmless parasitic role.

Both *H. pertussis* and *H. bronchisepticus* elaborate at least one toxin. *H. pertussis* toxin was first described by Evans and Matland (1937) who extracted it

from ground up bacilli. It was lethal on intravenous injection into guinea pigs and produced areas of necrosis on intradermal injection in the rabbit. Evans and Maitland's preparations also contained the agglutinin; this proved to be distinct from the toxin since antisera to the extract were protective against experimental infection, agglutinated bacillary suspensions but had no antitoxic activity as judged by their ability to modify skin necrosis induced by the toxin. The toxin was easily destroyed by formalin, was unstable at 37° C. and was rapidly destroyed at 55° C. A similar toxin with very similar properties was obtained from *H. bronchisepticus* (Evans and Maitland 1939) and from *H. parapertussis* (Brueckner and Evans 1939). The toxin presumably owing to its marked instability did not at first appear to be antigenic. Evans (1940, 1942) later found that formalized toxin was antigenic and that antitoxin prepared against it neutralized the toxins of *H. pertussis*, *H. parapertussis* and *H. bronchisepticus*. It is of practical interest that he was unable to induce antitoxin formation in the rabbit by the injection of whole bacilli but found that toxic extracts of the bacilli were antitoxinogenic (see also Katsampes, Brooks and Bradford 1942). These observations have been confirmed and extended (Flosdorf, Bondi and Dozois 1941; Ehrlich, Bondi, Mudd and Flosdorf 1942; Eldering 1941, 1943). Flosdorf and his colleagues distinguish in *H. pertussis* a feebly antigenic thermostable toxin in addition to the thermolabile toxin of Evans and Maitland. Toxin is produced by all phases of *H. pertussis* though most abundantly by Phase I (see also Wood 1940; Roberts and Ospeck 1942). Eldering obtained toxic fractions from *H. pertussis*, *H. parapertussis* and *H. bronchisepticus* and demonstrated varying degrees of cross protection against infection by living bacteria in animals actively immunized by the fractions. It is probable that her results reflect the antigenic similarity of both toxins and agglutinogens present in the extracts.

The mode of action of the toxin is at present obscure. Ehrlich and his colleagues (1942) describe generalized degenerative changes in the viscera of intoxicated rabbits, particularly in the lymphoid tissue. Given intratracheally, the heat labile toxin produces in the lungs of rabbits a severe oedematous reaction, followed by a characteristic accumulation of macrophages in the alveoli of lymphocytes round the blood vessels and severe necrosis in scattered areas—a histological picture not unlike that found in the lung in whooping cough. Antitoxin protected rabbits against this effect (Sprunt and Martin 1943). Though the precise role of toxin and antitoxin in infection with *H. pertussis* is not yet fully understood, the protective action of antitoxin in experimental infection appears to be limited to neutralizing the toxin contained in the infecting dose and thus reducing the likelihood of the organisms establishing a foothold in the tissues. Thus Anderson and North (1943) protected mice against an intraperitoneal injection of *H. pertussis* with antitoxin but not with antiserum. Systemically administered antitoxin had no effect in animals infected by the nasal route but antibacterial serum was effective. In their hands toxin had no aggressive action (see Chapter 48) in nasal infections. It should be noted, however, that North and his colleagues (1939) found that in intranasally infected mice the protective effect of human sera from pertussis convalescents could not be accounted for simply in terms of agglutinin or toxin neutralizing power though the effect as a whole appeared to be antibacterial. The labile toxin, however, may have immunizing properties. In a later study (1941) of active immunization they found that whole bacilli treated with the minimum of heat (58° C. for 8 min.) or with phenolic preservatives induced a higher degree of active immunity than bacilli preserved with formalin or heated to 60° C. for one hour. Evans (1944) confirmed the inability of intravenous antitoxin to protect mice against intranasal infections but found that antitoxin mixed with the bacteria before instillation into the nose lowered their infectivity.

suggesting that in these circumstances antitoxin had an anti-aggressive effect. Ospeck and Roberts (1944) were able to protect mice and rabbits against live bacilli or toxin by previously administered antitoxin. (For active immunization of laboratory animals by *H. pertussis* and various fractions of the organism, see also Cruckshank and Freeman 1937, Müller and Silverberg 1939, Mishulow *et al* 1939, Silverthorne 1940, Silverthorne and Cameron 1942, Holm and Bunney 1942, Lapan 1942, Streat *et al* 1941)

*H. influenzae suis* plays an important part in swine influenza (see Chapter 74) *H. canis* was isolated by Friedberger (1903) from 19 of 20 dogs suffering from a suppurative inflammation of the prepuce, but he was unable to reproduce the disease with it, and concluded that it was a harmless parasite of the preputial sac. It has also been isolated from normal dogs by Krage (1910), Kristensen (1922), Rivers (1922b), and Kirchenbauer (1934)

#### Experimental Infections.

*H. influenza*—Attempts to produce an infection resembling influenza in man by experiments on human volunteers or on the higher apes are considered in Chapter 74

As regards the usual laboratory animals, the injection of large doses of living culture (the growth from  $\frac{1}{4}$  to 1 blood-agar slope suspended in saline) into the peritoneum of rabbits, guinea pigs, or mice, often results in death within 24–48 hours. At necropsy petechial hæmorrhages may be found, scattered over the peritoneum, and sometimes over the pleura. The suprarenals may be congested or hæmorrhagic. The organisms can be recovered from the peritoneal cavity, but not often from the heart's blood. The cause of death seems to be a toxæmia, rather than an invasive infection (see Pfeiffer 1893, Delius and Kofle 1897, McIntosh 1922). Similar results may be obtained with filtrates of cultures in liquid media, and these may produce death on intravenous injection into rabbits or guinea pigs, though relatively large doses (0.5–5 ml.) are usually required (see Parker 1919, Ferry and Houghton 1919, Wollstein 1919, McIntosh 1922). There is no evidence that these filtrates contain an exotoxin in the usually accepted sense. In view of Pittman's observations and of her reports that her smooth strains are more virulent than the usual rough strains, it is of interest to note that many observers have recorded wide variations in virulence when a number of strains are tested by the intraperitoneal injection of living cultures. McIntosh (1922), for instance, found that only a small minority of recently isolated strains proved to be of high virulence when tested in this way. It would seem also that strains of *H. influenza* isolated from cases of meningitis are usually far more virulent for laboratory animals than strains isolated from the respiratory tract, and that some of these meningeal strains have definite invasive powers (Cohen 1909, Henry 1912, Wollstein 1915). The incorporation of the intraperitoneal infecting dose in mucin—a technique which has been successfully applied to enhancing the virulence of meningococci (see Chapter 23)—increases the virulence of *H. influenza*, small doses produce a fatal septicæmia in mice against which anti-influenzal horse serum is protective (Fothergill, Dingle and Chandler 1937) (see also Silverthorne 1940). Certain strains of *H. influenza* of human origin give rise to a fatal infection after intracerebral injection of about 2,000 organisms into mice, other strains, and strains of *H. para-influenza* are non virulent by this route (de Torregrósa and Francis 1941).

*H. pertussis*—The effect of the intraperitoneal injection of this organism into rabbits or guinea pigs is very similar to that of *H. influenza*. Here again large doses are required to produce death, and the infection seems to be toxæmic rather than invasive (Bordet and Gengou 1907, 1909, Wollstein 1909). Leabe and Gardner (1931) carried out a careful series of experiments in which they determined the relative toxicity of suspensions of strains of *H. pertussis*, antigenically in Phase I, II, III or IV, by intraperitoneal injections in guinea-pigs. They found that the minimal lethal dose of strains in Phase III or IV (rough strains) was twenty to thirty times greater than the minimal lethal dose of strains in Phase I or II (smooth, or relatively smooth strains).

The intranasal instillation of *H. pertussis* into anesthetized mice produces a patchy or diffuse interstitial pneumonia leucocytic infiltration round vessels and bronchioles proliferation of the bronchiolar epithelium and mucous secretion in the bronchioles containing masses of bacteria (Burnet and Timmins 1937, Bradford 1938). The histological picture in many respects resembles that of the lung in human pertussis and clearly offers a near approach to the natural disease for immunological study. For intraperitoneal infection, the normally low virulence of *H. pertussis* by the intraperitoneal route may be enhanced by starch (Powell and Jameson 1937) or mucin (Silverthorne 1938). Witebsky and Salm (1937) using rabbits injected intradermally, produced inflammatory lesions followed in 2-3 days by necrosis.

After intratracheal inoculation Culotta, Harvey and Gordon (1935) produced in three monkeys a disease with a 10-day incubation period, a catarrhal stage and a febrile coughing stage not unlike human pertussis. By similar means Sprunt, Martin and McDermann (1938) produced an interstitial pneumonia in the monkey characterized by a mononuclear cell reaction, and accompanied by a lymphocytosis, and North and his colleagues (1940) induced in *Macacus* monkeys an infection which by the seventh day resulted in a sticky tracheal and bronchiolar exudate full of *H. pertussis* and pulmonary congestion with conspicuous fibrinous and cellular infiltration both interstitially and in the alveoli. None of the monkeys developed a cough.

*H. influenzae suis*—In association with a filtrable virus (see Chapter 74) this organism is an important natural pathogen of swine, and the disease can be experimentally produced in these animals. In relation to the small animals of the laboratory this organism appears to behave much in the same way as *H. influenzae*. Large intravenous injections may be fatal for rabbits and large intraperitoneal injections for guinea pigs or mice, but the results are very irregular and there appear to be great differences in the virulence or toxicity of different strains (see Lewis and Shope 1931, Kirchenbauer 1934).

*H. canis*—The data with regard to the pathogenicity of this species for laboratory animals are extremely scanty. Rivers (1922b) notes that the intraperitoneal injection of 1 ml of a 24 hours culture in blood broth failed to kill a mouse, 2 ml intraperitoneally did not kill a small guinea pig, 1 ml intravenously did not kill a small rabbit.

*H. ducreyi*—Tomasczewski (1903) was successful in reproducing the disease in human subjects with pure cultures. In man progressive purulent lesions follow the intradermal injection of cultures, and it is apparently a common practice to separate *H. ducreyi* from contaminating saprophytes in genital material by injecting it intradermally into the patient (see Cunha 1939). Ulcerative lesions have followed the inoculation of monkeys and rabbits with cultures several generations removed from primary isolation (Reknskierna 1921, Nicolle 1923).

*H. para influenzae*—This organism appears to be non pathogenic for laboratory animals.

Variation.

The available data with regard to variation in the genus *Haemophilus* have already been referred to in the discussion of antigenic structure and of pathogenicity.

Both *H. influenzae* and *H. pertussis* give rise in artificial culture to variants that are essentially of the rough type. It would, indeed, seem that these species are peculiarly liable to undergo this change. The evidence suggests that rough strains of *H. influenzae* occur very commonly in the normal nasopharynx, so that the smooth  $\rightarrow$  rough variation must be supposed in this case to be of frequent occurrence when the organism is living in its normal habitat. In the case of *H. pertussis* we have, at present, no evidence that rough variations occur among recently isolated parasitic strains, but there is much evidence to suggest that the production of rough variants is readily induced by growing the organism on a relatively unfavourable medium.

We have, as yet, no evidence with regard to variation in the other species of this group

### *H. influenzae*

*Isolation*—Isolated by Pfeiffer (1892) from cases of influenza in man.

*Habitat*.—Strict parasite, living particularly in the upper respiratory tract of man.

*Morphology*.—In its typical form *H. influenzae* is a tiny coccus-bacillus (1-1.5 by 0.3-0.4  $\mu$ ). According to Pittman (1931), the bacillus in its virulent smooth form is capsulated. Most strains, even when first isolated from the tissues are non-capsulated, but it is possible that these should be regarded as rough variants. Among any large sample of strains, or in any one strain during prolonged subculture in the laboratory wide departures from the typical morphology will usually be found. Longer bacillary forms and definitely filamentous forms often occur and the latter may show angular bendings or sinuous curves. In the filamentous forms globular or ovoid swellings are not uncommon. The organism is non-flagellated and forms no spores. It stains feebly with many of the ordinary bacteriological dyes, more readily with dilute carbol fuchsin. It is Gram negative and not acid fast.

*Growth requirements*.—*H. influenzae* requires both the X factor and V factor for its growth. It grows far more readily under aerobic than under anaerobic conditions, and it would appear that some strains are incapable of prolonged anaerobic subcultivation. The optimal temperature for growth is in the neighbourhood of 37° C.

*Growth on Solid Media*.—On Fildes or Levinthal's medium the usual type of colony produced by *H. influenzae* is transparent, or slightly opaque, circular and dome-shaped, or slightly conical, with a slightly everted-out entire edge. At the end of 24 hours growth at 37° C. these colonies usually attain a diameter of 0.5-0.8 mm. On further incubation, and in some cases during the first 24 hours, the colony becomes differentiated into a central portion with a granular or contoured surface, an intermediate flattened portion, and a sharply bevelled periphery with a narrow everted-out edge. During the second 24 hours of growth the colony usually enlarges to a diameter of 1-1.5 mm. There is a tendency, which is not absolute, for differentiation to occur earlier and to be more pronounced in strains that have an atypical morphology. The growth is butyrous and emulsifies easily.

Some strains, described by Pittman (1931) as smooth, and by Chandler, Fothergill and Dingle (1933) as mucoid, give colonies that differ from those described above in having a smooth, undifferentiated, slightly mucoid surface. They have an entire edge. They tend to attain a larger size (1-3 mm. in diameter). They are slightly opaque, and, when viewed by obliquely transmitted light, they are indolent. Strains that give this type of colony show antigenic characters, and differences in pathogenicity, which are in accord with the view that they represent the "smooth" phase of the organism while the more frequently encountered strains, having the colonial appearances previously described, are in the non-mucoid or the rough state and are not indolent.

*Growth in Liquid Media*.—In a suitable liquid medium most strains of *H. influenzae* give rise to a uniform turbidity with or without a slight powdery deposit. Some give a more flocculent deposit. The latter usually show an atypical morphology, and the colonial appearances associated with the more advanced stage of rough variation.

*Resistance*.—*H. influenzae* is killed by an exposure to a temperature of 50-55° C. for 30 minutes.

*Biochemical Activities*.—*H. influenzae* usually ferments dextrose though no vigorously producing acid without gas. Lactose and mannitol are never fermented. The action on maltose, saccharose and dextrin, varies. Smooth and morphologically typical strains tend not to attack these substrates. The rougher morphologically atypical strains ferment them rather more frequently. The production of indole shows a high correlation with other characters that differentiate between relatively

smooth and relatively rough strains, a high proportion of the former, including both Pittman's smooth strains and the more 'typical' strains isolated from the normal nasopharynx, produce indole, but only a small proportion of the latter. All strains reduce nitrates.

*H. influenzae*, as that species is here defined, does not as a rule produce hæmolysis, which has been recorded only in a very few strains.

**Antigenic Structure**—The majority of smooth strains, as defined by Pittman (1931), fall into 6 well-differentiated antigenic types which appear to be characterized by specific polysaccharide surface antigens, sometimes occurring in a capsular form. The more common rough, or partially rough, strains are antigenically heterogeneous.

**Pathogenicity**—Pathogenic for man, particularly in association with virus infections, or with other bacterial diseases. Produces toxic death when injected in large doses into laboratory animals, and infective death when injected in small doses together with mucin.

### The Koch-Weeks Bacillus

There is no known way in which this organism can be distinguished from *H. influenzae*. The fact that strains so labelled have been isolated from the conjunctiva does not seem to warrant the allotment of a separate specific name.

### *H. influenzae-suis*

Isolated from cases of swine influenza in which it is associated with a filtrable virus. The characters of this species as recorded by Lewis and Shopo (1931) differ from the human strains of *H. influenzae* only in that no carbohydrates are fermented, and no indole is produced. The number of strains as yet examined does not, however, justify any definite generalized statement on this point. (See also Kirchenbauer 1934.)

### *H. para-influenzae*

Strains of this organism differ from *H. influenzae* in requiring the V factor but not the X factor for their growth, and in fermenting maltose, saccharose and often dextrin. In this last respect they resemble the rougher strains of *H. influenzae* except that their action on maltose is far more consistent. Their individual and colonial morphology often resembles that of "atypical" *H. influenzae* strains. Some are hæmolytic, others are not, and a few strains produce indole. In man hæmolytic strains tend to be associated with acute pharyngitis and both the hæmolytic and non hæmolytic strains with ulcerative endocarditis (Russell and Fildes 1928, Fox 1935, Stuart Harris *et al.* 1935, Miles and Gray 1938). The organism survives only 2-4 days on solid culture media.

### *H. canis*

Isolated by Friedberger (1903) from the prepuce of dogs. It is apparently parasitic, but not pathogenic. It differs from *H. influenzae* in the following ways. It requires the X factor, but not the V factor, for its growth. On solid media it forms colonies that are at first indistinguishable from those of *H. influenzae*, but later become larger and more opaque. As regards its fermentation reactions it ferments dextrose, saccharose and mannitol, produces indole and reduces nitrates. In its fermentation of mannitol it differs from both typical and atypical strains of *H. influenzae*.

### *H. ducreyi*

**Morphologically** in the purulent discharge from the ulcerated surface of the lesion the organisms appear as small ovoid rods arranged in pairs, in groups, or in chains lying parallel to one another. Several forms may, however, be assumed. Thus, it may appear as a short rod with parallel sides and rounded ends, staining evenly, or it may be ovoid or navicular in shape with marked bipolar staining,

or it may occur in pairs end-to-end having a dumb bell appearance. In size the bacillus is about  $1.1-1.5 \mu$  long by  $0.6 \mu$  broad (Stein 1938). It may be intra or extracellular in position. It does not form spores. It is Gram negative and non acid fast. In cultures on solid media the organisms appear as isolated individuals in groups and in short chains. In fluid media very long chains are frequently formed and in certain media it produces a pellicle with dependent stalactites of growth (Cunha 1939, 1943).

The organisms may be cultivated by inoculating scrapings from the floor of the ulcer on to a medium consisting of 3 per cent agar containing 20-33 per cent defibrinated rabbit's blood. The medium should be prepared on the day of inoculation and should be distributed into wide tubes having a large surface exposed to the air (Nicolle 1923, Reenstierna 1923, Nicolle and Durand 1924). Several tubes should be inoculated and incubated at  $35^{\circ}\text{C}$ . Colonies appear in 24 hours, and may be picked off for purification. On blood agar after 24 hours the colonies are circular 0.5-1.0 mm in diameter, low convex, greyish white and glistening with a smooth surface and entire edge. After 2 to 3 days they may reach a diameter of 2 mm and the surface may show a crateriform depression. According to Hunt (1935) growth occurs best in sealed tubes suggesting that it is favoured by an increased partial pressure of  $\text{CO}_2$ .

The necessity for blood in the medium and a low partial oxygen pressure is stressed by Sanderson and Greenblatt (1937). Watanabe (1939) confirmed the necessity for blood. Rabbit blood was best, followed by that of the goat, sheep, ox or man. He observed no growth stimulation by  $\text{CO}_2$ .

According to Lwoff and Pirofsky (1937) *H. ducreyi* requires X but not V factor for growth. Only small quantities of hæmin are required. The growth of some strains in the absence of blood or serum in the medium (Hababou-Sala 1925, de Assis 1926) may be attributed to the presence of small but sufficient quantities of hæmin in nutrient broth.

Another medium that is recommended for primary isolation consists of 1 part of 5 per cent glycerine agar and 4 parts of Beeredka's egg medium. On this medium the colonies are said to be round, transparent and of a rose mother-of-pearl colour (Hababou-Sala 1925). After preliminary incubation at  $35^{\circ}\text{C}$  cultures are said to remain viable at room temperature for about a month.

In Martins' broth, to which 20 per cent of defibrinated rabbit's blood has been added, the organism develops rapidly forming granules which are suspended in the liquid or become attached to the walls of the tube. After a few days an incomplete film may form on the surface. Cultures in this medium remain viable in the incubator for at least 10 days.

For preserving the organism it should be inoculated into a medium consisting of 0.25 per cent of nutrient agar, 1 per cent starch and 20 per cent of defibrinated rabbit's blood. Cultures on this medium remain alive for a month at incubator temperature and for a similar period at room temperature provided they are previously incubated for 5 days.

*H. ducreyi* is not specially resistant. It is killed by moist heat at  $55^{\circ}\text{C}$  within an hour and by 0.5 per cent phenol in a comparatively short time.

The fermentation reactions of *H. ducreyi* do not appear to have received much attention. Serologically suspensions from blood-agar cultures are agglutinated by a specific antiserum; this reaction may be used for identification.

*H. ducreyi* is naturally pathogenic for man. Monkeys have been successfully infected. The organism has a low pathogenicity for chick embryos (Anderson and Snow 1940).

A hæmophilic organism resembling *H. canis* and *H. ducreyi* in requiring only X factor but requiring an excess of  $\text{CO}_2$  in the atmosphere for its growth has been isolated from an ulcerative endocardial lesion in man (Kharat 1940).

## H pertussis

**Isolation**—Isolated by Bordet and Gengou (1906) from cases of whooping cough, and now recognized as the causal organism of that disease

**Morphology**—*H. pertussis* bears a general resemblance to *H. influenzae* in its morphology. The cell form is more constant, being usually of the short bacillary type. Longer bacillary or thread forms may occur, but they are relatively uncommon.

**Growth Requirements**—*H. pertussis* is not dependent on either the V factor or the X factor for growth. On first isolation it requires a complex medium, the most suitable being that devised by Bordet and Gengou, containing blood, potato extract and glycerol. It can, however, be trained to grow on agar. The optimal temperature for growth is in the near neighbourhood of 37° C.

**Growth on Solid Media**—On the Bordet Gengou medium *H. pertussis* gives smooth, dome-shaped, glistening colonies, with an entire edge. They are more opaque than those of *H. influenzae*, and are greyish as well as glistening. They have been likened not inappropriately to a bisected pearl, less appropriately to a small drop of mercury. When fully developed they tend to be rather larger than the colonies of *H. influenzae*, but they develop more slowly and the characteristic appearances described above are often not obvious in less than 48–72 hours' incubation.

**Growth in Liquid Media**—In serum *H. pertussis* gives a uniform turbidity with a slight deposit, which is sometimes slightly flocculent.

**Resistance**—*H. pertussis* is killed by exposure to a temperature of 55° C for 30 minutes.

**Biochemical Activities**—*H. pertussis* does not ferment any sugar. It does not form indole, or reduce nitrates. It produces a hazy zone of hæmolysis.

**Antigenic Structure**—*H. pertussis* in the normal smooth phase constitutes a single antigenic type. In artificial culture, particularly on a relatively unfavourable medium, it gives rise to rough or partially rough, variants, with a different antigenic structure. The antigen characterizing the S form, and the endotoxin of *H. pertussis* are serologically related to the corresponding substances in *H. parapertussis* and *H. bronchisepticus*.

**Pathogenicity**—*H. pertussis* is the cause of whooping cough in man. Injected in large doses into laboratory animals it gives rise to a fatal toxæmic infection very similar to that produced by *H. influenzae*. Introduced intranasally or intratracheally, it produces a fatal broncho-pneumonic infection.

## H parapertussis

An organism isolated by Eldering and Kendrick (1937) from cases of whooping cough in the United States which differs from *H. pertussis* (a) in producing a brown pigment on certain blood media, (b) in producing catalase, and (c) in having an S somatic antigen and an endotoxin which are apparently distinct from those related to, those in *H. pertussis*.

## H bronchisepticus

**Synonyms**—*B. bronchisepticus*, *Br. bronchiseptica*

**Isolation**—By Ferry (1911) in the United States, and by M Gowan (1911) in Edinburgh from dogs affected with distemper.

**Habitat**—Strict parasite, occurring in several different species of animals and sometimes in man.

**Morphology**—Similar to *H. pertussis* but is motile by peritrichate flagella.

**Cultural Characteristics**—Grows fairly well on nutrient agar media producing small, round, convex amorphous colonies, with smooth glistening surface, of butyrous consistency. Grows best under aerobic conditions, no growth under strictly anaerobic conditions. In agar shake cultures growth is almost entirely on the surface. Some strains are hæmolytic.



or it may occur in pairs end to end, having a dumb bell appearance. In size the bacillus is about  $1.1-1.5 \mu$  long by  $0.6 \mu$  broad (Stein 1923). It may be intra or extracellular in position. It does not form spores, it is Gram negative, and non acid fast. In cultures on solid media the organisms appear as isolated individuals in groups and in short chains, in fluid media very long chains are frequently formed and in certain media it produces a pellicle with dependent "stalactites" of growth (Cunha 1939 1943).

The organisms may be cultivated by inoculating scrapings from the floor of the ulcer on to a medium consisting of 3 per cent agar containing 20-33 per cent defibrinated rabbit's blood. The medium should be prepared on the day of inoculation and should be distributed into wide tubes having a large surface exposed to the air (Nicolle 1923 Reenstierna 1923, Nicolle and Durand 1924). Several tubes should be inoculated and incubated at  $35^{\circ}\text{C}$ . Colonies appear in 24 hours, and may be picked off for purification. On blood agar after 24 hours the colonies are circular  $0.5-1.0 \text{ mm}$  in diameter, low convex, greyish white and glistening, with a smooth surface and entire edge, after 2 to 3 days, they may reach a diameter of 2 mm and the surface may show a crateriform depression. According to Hunt (1935) growth occurs best in sealed tubes, suggesting that it is favoured by an increased partial pressure of  $\text{CO}_2$ .

The necessity for blood in the medium, and a low partial oxygen pressure is stressed by Sanderson and Greenblatt (1937). Watanabe (1939) confirmed the necessity for blood. rabbit blood was best, followed by that of the goat, sheep or man. He observed no growth stimulation by  $\text{CO}_2$ .

According to Lwoff and Pirosky (1937) *H. ducreyi* requires  $\lambda$  but not  $\gamma$  factor for growth. Only small quantities of hæmin are required. The growth of some strains in the absence of blood or serum in the medium (Hababou-Sala 1925 de Aze 1926) may be attributed to the presence of small but sufficient quantities of hæmin in nutrient broth.

Another medium that is recommended for primary isolation consists of 1 part of 5 per cent glycerine agar and 4 parts of Besredka's egg medium. On this medium the colonies are said to be round, transparent, and of a rose mother-of-pearl colour (Hababou-Sala 1925). After preliminary incubation at  $35^{\circ}\text{C}$ , cultures are said to remain viable at room temperature for about a month.

In Martin's broth, to which 20 per cent of defibrinated rabbit's blood has been added the organism develops rapidly, forming granules, which are suspended in the liquid or become attached to the walls of the tube. After a few days an incomplete film may form on the surface. Cultures in this medium remain viable in the incubator for at least 10 days.

For preserving the organism it should be inoculated into a medium consisting of 0.25 per cent of nutrient agar, 1 per cent starch and 20 per cent of defibrinated rabbit's blood. Cultures on this medium remain alive for a month at incubator temperature and for a similar period at room temperature, provided they are previously incubated for 5 days.

*H. ducreyi* is not specially resistant. It is killed by moist heat at  $55^{\circ}\text{C}$  within an hour and by 0.5 per cent phenol in a comparatively short time.

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A hæmophilic organism resembling *H. canis* and *H. ducreyi* in requiring only X factor but requiring an excess of  $\text{CO}_2$  in the atmosphere for its growth, has been isolated from an ulcerative endocardial lesion in man (Khairst 1940).

**H pertussis**

**Isolation**—Isolated by Bordet and Gengou (1906) from cases of whooping cough and now recognized as the causal organism of that disease

**Morphology**—*H. pertussis* bears a general resemblance to *H. influenzae* in its morphology. The cell form is more constant being usually of the short bacillary type. Longer bacillary or thread forms may occur but they are relatively uncommon.

**Growth Requirements**—*H. pertussis* is not dependent on either the V factor or the X factor for growth. On first isolation it requires a complex medium, the most suitable being that devised by Bordet and Gengou containing blood, potato extract and glycerol. It can however be trained to grow on agar. The optimal temperature for growth is in the near neighbourhood of 37° C.

**Growth on Solid Media**—On the Bordet-Gengou medium *H. pertussis* gives smooth, dome-shaped, glistening colonies with an entire edge. They are more opaque than those of *H. influenzae* and are greyish as well as glistening. They have been likened not inappropriately to a bisected pearl, less appropriately to a small drop of mercury. When fully developed they tend to be rather larger than the colonies of *H. influenzae* but they develop more slowly and the characteristic appearances described above are often not obvious in less than 48–72 hours incubation.

**Growth in Liquid Media**—In serum *H. pertussis* gives a uniform turbidity with a slight deposit which is sometimes slightly flocculent.

**Resistance**—*H. pertussis* is killed by exposure to a temperature of 50° C for 30 minutes.

**Biochemical Activities**—*H. pertussis* does not ferment any sugar. It does not form indole or reduce nitrates. It produces a hazy zone of haemolysis.

**Antigenic Structure**—*H. pertussis* in the normal smooth phase constitutes a single antigenic type. In artificial culture, particularly on a relatively unfavourable medium, it gives rise to rough or partially rough variants with a different antigenic structure. The antigen characterizing the S form and the endotoxin of *H. pertussis* are serologically related to the corresponding substances in *H. parapertussis* and *H. bronchisepticus*.

**Pathogenicity**—*H. pertussis* is the cause of whooping cough in man. Injected in large doses into laboratory animals it gives rise to a fatal toxæmic infection very similar to that produced by *H. influenzae*. Introduced intranasally or intratracheally it produces a fatal broncho-pneumonic infection.

**H parapertussis**

An organism isolated by Eldering and Kendrick (1937) from cases of whooping cough in the United States which differs from *H. pertussis* (a) in producing a brown pigment on certain blood media, (b) in producing catalase and (c) in having an S somatic antigen and an endotoxin which are apparently distinct from though related to those in *H. pertussis*.

**H bronchisepticus**

**Synonyms**—*B. bronchisepticus*, *Br. bronchiseptica*.

**Isolation**—By Ferry (1911) in the United States and by M. Gowan (1911) in Edinburgh from dogs affected with distemper.

**Habitat**—Strict parasite occurring in several different species of animals and sometimes in man.

**Morphology**—Similar to *H. pertussis* but is motile by peritrichate flagella.

**Cultural Characteristics**—Grows fairly well on nutrient agar media producing small, round, convex, amorphous colonies with smooth glistening surface of butyrous consistency. Grows best under aerobic conditions, no growth under strictly anaerobic conditions. In agar shake cultures growth is almost entirely on the surface. Some strains are hæmolytic.



FIG 170—*Hemophilus bronchisepticus*  
From an agar culture, 24 hours, 37° C.  
( $\times 1000$ )

*Persistence*—Similar to *H. pertussis*

*Biochemical Activities*—Haemolysin produced, active on red corpuscles of rabbit, dog and guinea pig. Grows freely on MacConkey. No carbohydrates fermented. Produces marked alkalinity in litmus milk. Nitrates often reduced.  $H_2S$ —,  $NH_3$  very slight production or none at all. Catalase—++ Grows in Koser's citrate

*Antigenic Structure*—The surface antigen of the S form, and its endotoxin, are serologically homogeneous, and related to the corresponding substances in *H. pertussis* and *H. paraptus*

*Pathogenicity*—Frequent cause of bronchopneumonia in rodents, and of bronchopneumonia complicating distemper in dogs. Experimentally, intraperitoneal

inoculation of guinea pigs with 0.5 ml of a 24 hours' broth culture causes death in 24 to 48 hours. Post mortem, there are small haemorrhages on the peritoneum, and a viscid translucent exudate forming pseudo-membranes on the liver, spleen and the less mobile parts of the intestine. The bacilli are easily recovered from the peritoneal cavity, but with difficulty from the blood, liver and lungs. Subcutaneous inoculation produces only a local lesion. Feeding and inhalation are without effect. The organism is non pathogenic to mice. It rapidly loses its virulence in culture.

## REFERENCES

- ALEXANDER, H. E. and HEIDELBERGER, M. (1940) *J. exp. Med.*, 71, 1.  
 ANDERSON, G. and NORTH, E. A. (1943) *Aust. J. exp. Biol. med. Sci.*, 21, 1.  
 ANDERSON, K. and SNOW, J. S. (1940) *Amer. J. Path.*, 16, 263.  
 ANDERSON, L. R. (1931) *Amer. J. Hyg.*, 13, 161.  
 ANDERSON, R. A. and SCHULTZ, O. T. (1921) *J. exp. Med.*, 33, 653.  
 ASSIE, A. DE. (1926) *C. P. Soc. Biol.*, 95, 1003.  
 BAUDISCH, O. (1932) *Biochem. Z.*, 245, 265.  
 BIELING, R. and WEICHERD, R. (1920) *Dtsch. med. Wochs.*, 46, 1183.  
 BONDI, A. and FLOSDORF, E. W. (1943) *J. Immunol.*, 47, 315.  
 BORDET, J. (1912) *Zbl. Bakt.*, 68, 276.  
 BORDET, J. and GENGOU, O. (1906) *Ann. Inst. Pasteur*, 20, 731. (1907) *Ibid.*, 21, 720. (1909) *Ibid.*, 23, 415.  
 BORDET, J. and SLEESWYK, (1910) *Ann. Inst. Pasteur*, 24, 476.  
 BOURN, J. M. (1927) *J. infect. Dis.*, 41, 294.  
 BRADFORD, W. L. (1933) *Amer. J. Path.*, 14, 377.  
 BRADFORD, W. L. and SLAVIN, B. (1937) *Amer. J. publ. Hlth*, 27, 1277.  
 BRUCKNER, I. E. and EVANS, D. G. (1939) *J. Path. Bact.*, 49, 563.  
 BUEYER, F. M. and TIMMINS, C. (1937) *Brit. J. exp. Path.*, 18, 83.  
 CHANDLER, C. A., FOTHERGILL, L. D., and DINGLE, J. H. (1937) *J. exp. Med.*, 66, 709. (1939) *J. Bact.*, 37, 415.  
 COHEN, C. (1909) *Ann. Inst. Pasteur*, 23, 273.  
 COOPER, G. M., MISHLOW, L., and BLANC, N. E. (1921) *J. Immunol.*, 6, 25.  
 CRICKSHANK, J. C. and FREEMAN, G. G. (1937) *Lancet*, ii, 567.  
 CULOTTA, C. S., HARVEY, D. F., and GORDON, E. F. (1935) *J. Pediat.*, 6, 743.  
 CUNHA, R. (1939) *Zbl. Bakt.*, 144, 503. (1943) *O. Hospital*, 23, 393.  
 DAVIS, J. D. (1917) *J. infect. Dis.*, 21, 392. (1921) *Ibid.*, 29, 178, 187.  
 DELIUS, W. and KOLLE, W. (1897) *Z. Hyg. Infektkr.*, 24, 327.  
 DIBLE, J. H. (1924) *J. Path. Bact.*, 27, 151.

- DOCHEZ, A R, MILLS, K C, and KNEELAND, Y JR (1932) *Proc Soc exp Biol, N Y*, 30, 314
- EHRRICH, W. E, BONDY, A, MUDD, S, and FLOSDORF, E W (1942) *Amer J med Sci*, 204, 530
- EHRUND, A (1929) *Zbl Bakt*, 111, 195
- ELDERING, G (1941) *Amer J Hyg*, 34, B 1, (1941) *Ibid*, 36, 294
- ELDERING, G and KENDRICK P (1937) *J Bact*, 33, 71, (1938) *Ibid*, 35, 561
- EVANS, D G (1940) *J Path Bact*, 51, 49, (1942) *Lancet*, i 529, (1944) *J Path Bact*, 56, 49
- EVANS, D G and MAITLAND, H B (1937) *J Path Bact*, 45, 715, (1939) *Ibid*, 49, 67
- FERRY, N S (1911) *J infect Dis*, 8, 399, (1912) *Vet J*, 68, 376 (1912-13) *Amer vet Rev*, 41, 77
- FERRY, N S and HOUGHTON, E M (1919) *J Immunol*, 4, 233
- FILDES, P (1920) *Brit J exp Path*, 1, 129, (1921) *Ibid*, 2, 16, (1922) *Ibid*, 3, 210 (1923) *Ibid*, 4, 265, (1924) *Ibid*, 5, 69
- FLEMING, A (1929) *Brit J exp Path*, 10, 226
- FLEMING, A and MACLEAN, I H (1930) *Brit J exp Path* 11, 127
- FLOSDORF, E W, BONDY, A, and DOZOIS T F (1941) *J Immunol* 42, 133
- FLOSDORF, E W, BONDY, A, FELTON, H and MCGUINNERS, A C (1942) *J Pediat*, 21, 625
- FLOSDORF, E W, DOZOIS, T F, and KIMBALL, A C (1941) *J Bact* 41, 457
- FLOSDORF, E W and KIMBALL A C (1940a) *J Immunol* 39 287 (1940b) *Ibid* 39, 475
- FLOSDORF, E W, KIMBALL, A C, and CHAMBERS, L A (1939) *Proc Soc exp Biol N Y*, 41, 122
- FOTHERGILL, L D and CHANDLER, C A (1936) *J Immunol*, 31, 401
- FOTHERGILL, L D, DINGLE, J H, and CHANDLER, C A (1937) *J exp Med*, 65, 721
- FOX, W W (1933) *J Amer med Ass*, 105, 876
- FRIEDBERGER, E (1903) *Zbl Bakt*, 33, 401
- GALLI VALERIO, B (1896) *Zbl Bakt*, 19, 694
- GHOV, A and PREYSS, W VON (1904) *Zbl Bakt*, 35, 531
- GORDON, J, WOODCOCK, H F DE C, and ZINNEMANN, K (1944) *Brit med J* i 779
- GRASSBERGER, R (1897) *Z Hyg InfektKr*, 25, 453
- GUNDEL, M and SCHLUTER, W (1933) *Zbl Bakt*, 129, 461
- HABABOU SALA J (1925) *C R Soc Biol*, 92, 498
- HEVEY, H (1912) *J Path Bact*, 17, 174
- HOAGLAND, C L, WARD, S M, GILDER H, and SHANK, R E (1942) *J exp Med*, 76, 241
- HOLM, A and BUNNEY, W E (1942) *J Immunol*, 44, 33
- HUNT, G A (1935) *Proc Soc exp Biol, N Y*, 33, 293
- IIZUKA, A (1938) *Z ImmunForsch*, 94, 312-318
- KATSMANES, C P, BROOKS A M, and BRADFORD, W L. (1942) *Proc Soc exp Biol, N Y*, 49, 615
- KHAIRAT, O (1940) *J Path Bact*, 50, 497
- KIRCHENBAUER, H (1934) *Z InfektKr Haustiere*, 45, 273
- KNIGHT, B C, J G (1936) *Spec Rep Ser med Res Coun, Lond*, No 210
- KNORR, M (1924) *Zbl Bakt*, 92, 371, 385
- KOCH, R (1887) *Arb ReichsgesundhAmt*, 3, 62
- KOPP, H (1927-8) *Zbl Bakt*, 105, 54
- KRAGE, P (1910) *Z InfektKr Haustiere*, 7, 380
- KRISTENSEN, M (1922) *Hæmoglobulinophile Bacteria* ' Copenhagen, (1927) *C R Soc Biol*, 96, 355
- KRUNWIEDE, C, MISHULOW, L, and OLDENBUSCH, C (1923) *J infect Dis* 32, 22
- KRUNWIEDE, E and KUTTNER A G (1938) *J exp Med* 67, 429
- LAIDLAW, P P and DUCKIN G W (1926) *J comp Path*, 39, 222
- LAMONT, J A (1926) *Canad med Ass J*, 16, 1447
- LAPIN, J H (1942) *J Pediat*, 20, 161
- LAWSON, G M (1933) *Amer J Dis Child*, 47, 1454
- LESLIE, P H and GARDNER, A D (1931) *J Hyg, Camb*, 31, 423
- LEVINTHAL, W (1918) *Z Hyg InfektKr*, 86, 1
- LEWIS, P A and SHOPE, R E (1931) *J exp Med*, 54, 361
- LWOFF, A (1939) *Ann Inst Pasteur*, 62, 165
- LWOFF, A and LWOFF M (1937a) *Proc roy Soc, B* 122, 352-360, (1937b) *Ann Inst Pasteur*, 59, 129, (1937c) *C R Acad Sci*, 204, 1510
- LWOFF, A and PIROSKY, I (1937) *C R Soc Biol*, 124, 1169
- M GOWAN, J P (1911) *J Path Bact* 15, 372
- MCINTOSH, J (1922) *Spec Rep Ser med Res Coun, Lond*, No 63
- MALLORY, F B and HORNOR, A A (1912) *J med Res*, 27, 115
- MALLORY, F B, HORNOR A A, and HEYDERSON, F F (1912) *Ibid*, 27, 391

- MARTINI, E (1900) *Arch Hyg, Berl*, 38, 114
- MESSERSCHMIDT, T, HÜNDESHAGEN, K., and SCHEER, K. (1919) *Z Hyg InfektKr*, 83, 552
- MILES, A A and GRAY, J (1938) *J Path Bact*, 47, 257
- MILLER, J J (1937) *Proc Soc exp Biol N Y*, 37, 45
- MILLER, J J and SILVERBERG, R J (1939) *J infect Dis*, 85, 16.
- MISHTLOW, L., ALKIN I F, LISS M M., and LEIFER, L. (1939) *J Immunol*, 37, 17
- MULDER, J (1937) *Acta med scand.*, 91, 390
- NICOLLE, C (1923) *C R Soc. Biol*, 83, 871
- NICOLLE and DURAND (1924) *Arch. Inst Pasteur Tunis* 13, 243
- NORTH, E. A., ANDERSON, G., and GRAYDON, J J (1941) *Med J Aust*, 21, 589
- NORTH E A, KEOGH, E V, ANDERSON, G., and WILLIAMS, S (1939) *Aust J exp Biol med Sci*, 17, 275
- NORTH E A KEOGH, E V, CHRISTIE R., and ANDERSON, G (1940) *Aust J exp Biol med Sci*, 18, 125
- ODDIAI, (1911) *Zbl Bakt*, 61, 289
- OLMSTEAD, M and POVITZKY, O R (1916) *J med Pes*, 33, 379
- OLSEN, O (1920) *Zbl. Bakt*, 85, 12
- OSPECK A G and ROBERTS, M E (1944) *J infect. Dis*, 74, 22
- PARK W H, WILLIAMS A. W., and COOPER, G (1918) *Proc Soc. exp Biol, N Y*, 16, 120
- PARKER, J T (1919) *J Amer med Ass.*, 72, 476
- PFEIFFER, B (1892) *Dietsch med Wochr*, 18, 28. (1933) *Z Hyg InfektKr*, 13, 357
- PITTMAN, M (1931) *J exp Med.*, 53, 471. (1935) *J Bact*, 30, 149
- PLATT A E (1937) *J Hyg, Camb*, 37, 98. (1939) *Aust J exp Biol med Sci*, 17, 19
- POVITZKY, O R and DENNY, H T (1921) *J Immunol*, 6, 60
- POWELL, H. M and JAMIESON, W A (1937) *J Immunol.*, 32, 153
- PRITCHETT I W and STILLMAN, E. G (1919) *J exp Med* 29, 259
- RAETIG H. (1940) *Zbl Bakt*, 145, 386
- REENSTIERNA, J (1921) *Acta derm venerol., Stockh.*, 2, 1. (1923) *Arch. Inst. Pasteur, Tunis* 12, 273
- REEL, L. J (1915) *J med Pes* 32, 471
- RITCHIE, J (1910) *J Path Bact*, 14, 615
- RIVERS T M. (1922a) *Johns Hopk Hosp Bull*, 33, 429. (1922b) *J Bact.*, 7, 579
- RIVERS T M and KOHN, L A (1921) *J exp Med*, 34, 477
- ROBERTS M E and OSPECK A G (1942) *J infect Dis*, 71, 264.
- RUSSELL, D S and FIDLER, P (1928) *J Path. Bact*, 31, 601
- SANDERSON E S and GREENBLATT R B (1937) *Stb med J*, 30, 147
- SCHLUTER, W (1936) *Zbl Bakt*, 136, 362.
- SELTZER, H. (1906) *Z Hyg InfektKr*, 54, 347
- SHIRLEY, G S and HOELSCHER, H (1934) *J exp Med.*, 60, 403
- SHIGA, K., IMAI, N., and EGUCHI, C (1913) *Zbl Bakt.*, 69, 164
- SHOPE, R E (1931) *J exp Med*, 54, 349
- SILVERTHORNE, N (1935) *Canad J publ Hlth*, 29, 233. (1940) *Ibid.*, 31, 560
- SILVERTHORNE, N and CAMERON, C (1942) *J Pediat.*, 20, 1
- SILVERTHORNE, N, CAMERON, C., and PATERSON, M. (1943) *Canad J publ Hlth*, 34, 175
- SILVERTHORNE, N and PATERSON, M. (1943) *Canad. J publ Hlth*, 34, 178
- SMITH, M M. (1931) *J Hyg, Camb*, 31, 321
- SMITH, T (1913) *J med Res* 29, 291
- SMOLENS, J and MUDD, S (1943) *J Immunol.*, 47, 155
- STILLMAN, E. G and BOURN, J M. (1920) *J exp Med*, 32, 665
- SPOONER, E. T C. (1935) *J Hyg, Camb*, 33, 79
- SPRUNT D H. and MARTIN D S (1943) *Amer J Path*, 19, 255.
- STEIN, R O (1924) See Kolle and Wasserman, 'Hdb path Mikroorg "Hite Aufl., 1923-9, 6, 185
- STRADA F and TRAINA, R (1900) *Zbl. Bakt*, 23, 630
- STREAN L. P., LAPOINTE, D and DECHENE, E. (1941) *Canad med J Ass*, 45, 326.
- STUART HARRIS, C H, WELLS A Q, ROSHER, A. B, MACKIE F P., and WILSON, G S. (1935) *J Path. Bact.*, 41, 407
- TARTAKOWSKY, M. G (1897-8) *Arch. Sci biol., St Petersb*, 6, 203
- THALMER, W (1914) *Zbl. Bakt*, 74, 189
- THUTTA, T (1921) *J exp Med.*, 33, 763
- THUTTA T and AVERY, O T (1921) *J exp Med*, 34, 97, 455
- TOMASZCZEWSKI E (1903) *Z Hyg InfektKr.*, 42, 327
- TOOMEY J A., RANTA, K., POSEY, L., and MCCLELLAND, J E. (1930) *J infect. Dis.*, 57, 49
- TOOMEY J H. and TARACS, W S (1937) *J infect. Dis*, 60, 41. (1939) *Ibid.*, 62, 297
- TOOMEY, J A., TARACS, W S., and RANTA, K (1936) *J infect. Dis.*, 59, 326.

- VALENTINE E and COOPER G M (1919) *J Immunol* 4, 329  
 VALENTINE, F C O and RIVERS, T M (1927) *J exp Med*, 45, 993  
 WADE H W and MANALANG, C (1920) *J exp Med*, 31, 95  
 WATANABE, S (1939) *Kitasato Arch* 16, 1  
 WATANABE I (1938) *Jap J exp Med* 16, 529  
 WEBSTER, L T and BAUDISCH, O (1925) *J exp Med*, 42, 473  
 WEEKS, J E (1887) *Arch Augenheilk*, 17, 318  
 WILKES WEISS D (1936) *Proc Soc exp Biol A 1*, 35, 289, (1937) *J infect Dis*, 60, 213  
 WINHOLT, W (1915) *J infect Dis* 16, 389  
 WINSLOW, C E A, BROADHURST J, BUCHANAN R E KREMWIEDE C, ROGERS, L A  
 and SMITH G H (1920) *J Bact*, 5, 101  
 WITERSKY, E and SALM H (1937) *J exp Med* 65, 43  
 WOLLSTEIN, M (1909) *J exp Med* 11, 41, (1915) *Ibid.*, 22, 445 (1919) *Ibid*, 30, 553  
 HOOD, M L (1940) *J Immunol* 39, 25  
 WRIGHT, J and WARD H K (1932) *J exp Med* 55 235  
 YABE, S (1921) *Brit J exp Path* 2, 197

## CHAPTER 34

### BRUCELLA

#### DEFINITION—*Brucella*.

Small, non motile non-sporing Gram negative coccus-bacilli. Grow rather poorly on ordinary media, or may require special media. Aerobic no growth under strict anaerobic conditions. Growth often improved by CO<sub>2</sub>. Little or no fermentative action on carbohydrates. Usually tend to produce alkali in litmus milk, and a brown pigmentation on potato. Strict parasites, occurring in man and animals, and producing characteristic infections.

Type species. *Brucella melitensis*.

HISTORY—The first member of the group *Br melitensis* was isolated in 188 by Bruce from the spleen of patients who had died of Malta fever. At that time and for a long time afterwards the bacillary nature of the organism was not recognized in all the older textbooks it is therefore described as a micrococcus. The organism finds its natural habitat in the goat and the sheep. It may however infect other animals. In man it gives rise to undulant fever. It is fairly widely distributed throughout the world.

The discovery of the second member *Br abortus* was made by Bang of Copenhagen in 1897. Working in conjunction with Seibert he isolated the organism from cows suffering from infectious abortion and by a series of experiments demonstrated its specific role in this disease. The organism is parasitic in cattle. To a less extent it infects certain other animals. In man it gives rise to undulant fever. It is perhaps even more widespread than *Br melitensis* having been found in practically every country of the world.

The third member of the group *Brucella tularensis* was isolated by McCoy and Chapin in 1912 from a plague-like disease among rodents in California and was called by them *Bacterium tularense*. It infects ground-squirrels jack rabbits and other rodents and occasionally gives rise to a disease in man called tularemia.

The fourth member *Br suis* was isolated by Traum (1914) from the foetus of a sow. It is a natural parasite of pigs in which it gives rise to a disease frequently characterized by inflammatory lesions in the reproductive organs. It may occasionally infect other animals. In man it shares with *Br melitensis* and *Br abortus* the ability to produce undulant fever. It appears to be very much less widespread than these two organisms its chief home being in the large hog raising districts of the middle western states of North America. In Denmark *Br suis* strains have been isolated by Thomsen (1931-1934) which differ in certain respects from those found in the United States they will be referred to as the Danish porcine type. The American type has been found occasionally in Europe (see Thomsen 1934) and has been reported from Brazil (Neiva 1934) the Argentine and Australia (King 1934).

**Nomenclature.**—We have no space to discuss the early confusion of terminology that existed over members of this group. The whole position was altered when Evans in 1918 drew attention to the essential similarity of the organisms which at that time were described as *Micrococcus melitensis* and *Bacillus abortus*. Nor do we propose to discuss the validity of the generic name *Brucella* suggested for them by Meyer and Shaw (1920) and by Feusier and Meyer (1920) in honour of Sir David Bruce. This is so appropriate and has met with such universal approval that no other term seems likely to enter into serious competition with it. There are, however, certain points that require discussion. The three organisms isolated from goats, cattle, and pigs respectively are so closely allied that their differentiation can be accomplished only with difficulty. The question is, therefore, whether they should be regarded as varieties of one species, or should be ranked as separate species. Both proposals have their advocates. On the whole we favour the latter course, mainly for the sake of convenience. We shall therefore refer to these three

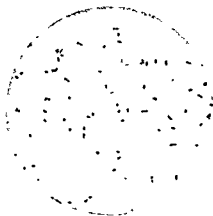


FIG. 171.—*Brucella abortus*.

From an agar culture, 24 hours, 37° C., showing very short bacillary forms ( $\times 1000$ ).

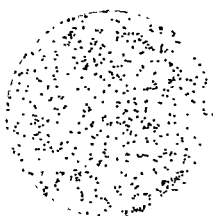


FIG. 172.—*Brucella melitensis*.

From an agar culture, 2 days, 37° C., showing mainly coccial forms ( $\times 1000$ ).

organisms as *Br. melitensis*, *Br. abortus* and *Br. suis*. It must be pointed out, however, that the differences between the American and Danish porcine types are almost as great as those between the porcine and bovine types, and the decision to treat them as varieties of *Br. suis* is purely arbitrary.

The names *paramelitensis*, *para-abortus*, and *parasuis* are frequently used to refer to inagglutinable strains of *Brucella*, corresponding most closely to the *melitensis*, *abortus*, and *suis* types. So long as the so-called para-strains were regarded as distinct species, no objection could be raised to this terminology; but now that they are known to be merely rough variants of the original smooth forms, this practice is no longer justifiable and merely serves to confuse the nomenclature. We shall refer to these, therefore, as rough *melitensis*, *abortus*, or *suis* strains, as the case may be.

The inclusion of *Br. tularensis* in this group is largely tentative. Reimann (1932), along with several other workers, would assign it to the *Pasteurella* group on account of its bipolar staining, its solubility in 1/800 sodium ricinoleate, and its transmission by insect vectors. On the other hand its general morphology, the beneficial effect



of  $\text{CO}_2$  on its growth, its cytotrophism (see Buddingh and Womack 1911), its failure to develop anaerobically, its production of  $\text{H}_2\text{S}$ , its very weak fermentative ability, its antigenic affinity to *Br melitensis* and *Br abortus* and its high pathogenicity for man in the laboratory qualify it perhaps even better for inclusion in the *Brucella* group. Since, however, it has not been studied with the same thoroughness as the other members, we shall exclude it from the following description of the group characteristics, and describe it separately at the end of the chapter.

**Morphology and Staining**—The bacilli are short and slender, the axis is straight, the ends are rounded, the sides may be parallel or convex outwards. In length they vary from about 0.6 to 1.5  $\mu$ , and in breadth from 0.5 to 0.7  $\mu$ . The short forms may appear as oval cocci, or, if they are about to divide, as diplococci. As a rule they are arranged singly, in pairs end-to-end or in small groups; sometimes short chains of 4-6 members may be seen, especially in liquid media. Owing to the frequent coccoid appearance, their bacillary nature may be in doubt, but it may be noted that in size they are smaller than any of the Gram-negative cocci. Moreover, when arranged in pairs, their long diameter is in the same axis as that in which they are lying in distinction to the Gram-negative diplococci, whose long axis is generally at right angles to that in which they are lying.

*Br melitensis* is generally considered to be more coccoid in form than *Br abortus*, and for this there is some justification. The difference in size and shape, however, is so slight as to render it impossible to distinguish with certainty between individual strains. Duncan (1925) has pointed out that these organisms, when grown on agar or glucose agar, show no marked morphological differences, but if they are cultivated on a relatively rich medium, such as Fildes' peptic digest blood agar, the *Br abortus* strains frequently develop long bacillary forms, reaching 2.0 or 3.0  $\mu$  in length, whereas *Br melitensis* strains usually retain their coccoid shape and rarely exceed 1.0  $\mu$  in length.

The organisms stain fairly well with the ordinary dyes. Bipolar staining is not uncommon, and occasionally irregularity in the depth of colour is seen. In old cultures irregular forms may be noted. They are Gram-negative, non-acid fast, non-motile and non-sporing. The presence of capsules in freshly isolated smooth strains has been described (Huddleston 1940, Mickle 1940).

**Cultural Reactions**—Apart from their different  $\text{CO}_2$  requirements, the members of this group resemble each other closely in their cultural characteristics. None of them is difficult to grow, none grows profusely. On agar the colonies are small, translucent and undifferentiated. In broth there is a moderate turbidity with a slight powdery or viscous sediment, which disintegrates completely on shaking, after about 2 weeks in the incubator or at room temperature the deposit becomes extremely viscous, and can be disintegrated only with difficulty. According to Thomsen (1933) if the organisms are grown in flasks of broth instead of in tubes the  $\text{H}_2\text{S}$ - and  $\text{CO}_2$ -sensitive *abortus* types give rise in 1 to 3 weeks to a mealy or scaly surface pellicle and a heavy deposit that is difficult to disintegrate by shaking. Aerobic *abortus* strains form no pellicle, but produce a uniform turbidity and a slight deposit that is easily disintegrated. Strains of *Br melitensis* give rise to a fairly dense turbidity, a moderately heavy deposit, and a granular, usually incomplete, surface growth. Growth in gelatin is poor, and is unaccompanied by liquefaction. Perhaps the most striking peculiarity is the yellowish colour that develops on potato

in 2 to 3 days, deepening to a café au lait or chocolate tint in the course of a fortnight. Individual strains vary in the depth of colour they produce, some giving a darker brown than others. The pigment is not confined to the layer of growth, it spreads throughout the potato. It will be recalled that a brown growth on potato is also given by *Pf mallei*, *Pf whitmorei*, *V cholerae*, and *Ps pyocyanea*, and certain other organisms. A similar but less intense brownish colour is sometimes noticeable in old agar slope cultures, particularly of *Br melitensis* (Kristensen 1931), it is not sufficiently constant, however, to be of differential value.

Huddleson, Hasley, and Torrey (1927) and Huddleson and Winter (1927) have observed the development of crystals of ammonium magnesium phosphate in cultures of *Brucella* on liver agar incubated aerobically, but not in 5-10 per cent  $\text{CO}_2$ . They regard their formation as being due to the production of ammonia by the growing organisms and to its combination with the magnesium phosphate in the medium. In their experience crystals are formed much more rapidly by *melitensis* and *paramelitensis* than by *abortus* strains. Our own experience (Wilson 1933) bears out their observations to some extent but does not suggest that the differences between different types are sufficient to be of value in the identification of individual strains. Incidentally American *suis* strains appear to be most active in the production of ammonia.

Attention has been drawn by de Santis (1933) to the different appearances presented on *Petragnani's* egg medium. Strains of *Br melitensis* are said to grow on this medium, and usually to change the colour from light yellowish green to dark green. *Br abortus* strains on the other hand, generally fail to develop. The behaviour of *Br suis* strains is doubtful, but in our limited experience growth is not infrequent. The reliability of this test for differential purposes is still under discussion (see Menzani 1934, Tosatti 1934, Messeri 1935, Vittone 1935, Pagnani 1935, Foresti 1935). The general opinion seems to be that it is less satisfactory than the  $\text{H}_2\text{S}$  and dye tests (see later). According to Schwarzmaier (1936) *Br melitensis* grows on *Petragnani's* medium both with and without malachite green, *Br suis* grows only without malachite green, and *Br abortus* fails to grow under either condition. The mode of preparation of the medium appears to be of importance. If it is heated to a temperature above  $80^\circ\text{C}$  for too long the normal inhibitory action of the egg albumin or the bovine and porcine strains is destroyed thus allowing some development of these organisms to occur (de Santis 1935). According to Martini (1935) the inhibitory effect of the egg albumin on *abortus* strains can be removed by the addition of sufficient  $\text{HCl}$  to lower the reaction of the finished medium from pH 8.4 to pH 6.8. If this is true, it suggests that the effect may be related to the greater necessity of *abortus* strains for  $\text{CO}_2$ .

Growth is rather slow, and unless a fairly heavy inoculum is made, colonies are not usually visible for 2 days or even longer. In broth the maximum turbidity is not reached for a week or more. On the whole the American porcine strains probably give the best, and the Danish porcine strains the poorest growth; the *melitensis* and *abortus* strains occupying an intermediate position. The behaviour, however, of different strains of the same type is subject to so much variation that no reliance can be placed on this character for differential purposes.

*Br abortus*, *Br melitensis* and *Br suis*, when inoculated on to the chorio-allantoic membrane of the developing chick embryo, are able to multiply and to bring about death of the embryo in a few days with lesions in the spleen and liver. All three organisms grow intracellularly—*Br melitensis* in the ectodermal epithelium, *Br abortus* and *Br suis* in cells of mesodermal origin and in the vascular endothelium. Rough strains are non-invasive (Goodpasture and Anderson 1937, Buddingh and Womack 1941, de Ropp 1944).

The colonial appearance of *Brucella* depends on the smoothness or roughness of the strain. The difference between smooth and rough colonies is not great, and is best brought out by examination under a binocular plate microscope using obliquely transmitted light. Colonies of antigenically smooth strains of *Br. abortus* on potato agar are small, bluish and translucent with regular margins and a smooth glistening surface. The individual cells are uniformly short rods arranged singly. Colonies of antigenically rough strains are of much the same size as the smooth colonies, but are less convex, more opaque, and have a dull granular appearance. The individual cells are usually somewhat larger than those of the smooth type, and occasional long slender rods may be observed. Intermediate colonial and morphological forms have also been described (Mingle and Mante 1941). The colonial differences are intensified on glycerol glucose agar. On this medium S and R colonies, if examined by oblique transmitted light against a dark background, produce the appearance of an irregular mosaic of light and dark, curved and angular areas (Henry 1933).

**Growth Requirements**—Growth is generally improved by the addition of natural animal protein to the medium. The most satisfactory media, particularly for the growth of *Br. abortus* and *Br. melitensis*, are liver extract agar—first described by Holth (1911), subsequently by Stafseth (1920), and frequently referred to as Huddleson's medium—2 per cent glycerol agar (Zeller and Stockmayer 1933), Fleming's (1919) chocolate agar (Henry *et al.* 1932), 5 per cent serum agar and Bacto-tryptose agar. Zobell and Meyer (1932) have described a synthetic medium in which the metabolism of *Brucella* strains may be studied. *Br. melitensis* and *Br. suis* are said not to grow in an amino-acid glucose inorganic salt medium unless nicotinamide, thiamin and pantothenic acid are added. *Br. abortus* requires biotin as well (Kerby 1939, Koser *et al.* 1941, Koser and Wright 1942).

The range of temperature consistent with growth is 20°–40° C, the optimum being about 37° C. At 20° C growth is very slow. The effect of H ion concentration is rather difficult to dissociate from that of CO<sub>2</sub>. Many strains of *Br. abortus* require for their optimum development a concentration of 5–10 per cent CO<sub>2</sub> in the atmosphere. This has the effect of turning an alkaline medium acid. For the growth of these organisms an initial H ion concentration of pH 6.6 is desirable. The other members of the group usually grow as well on an alkaline as on a slightly acid medium, but since, as will be pointed out directly, even *melitensis* strains are often benefited by a small amount of extra CO<sub>2</sub>, it is advisable for practical purposes to adjust media to pH 6.6–6.8.

**CO<sub>2</sub> Requirements**—One of the most interesting features of the *Brucella* group is their peculiar respiratory behaviour. Ever since its original isolation by Bang (1897) *Br. abortus* has presented certain difficulties in cultivation. No growth occurs on a solid medium under aerobic conditions. If, however, the tube is suitably sealed (Preis 1903) (see Fig. 173), or if it is attached by rubber tubing to another tube inoculated with an organism such as *B. subtilis* (Nowak 1906), growth occurs after a delay of a few days. These observations were generally interpreted as showing that *Br. abortus* was microaerophilic, and could not grow till the partial pressure of oxygen over the culture had been lowered to a suitable extent. A similar interpretation was also placed on the fact that, when inoculated into a serum agar gelatin shake medium, it grew in the form of a band situated about  $\frac{1}{2}$  cm. below the surface (Fig. 174).

Credit is due to Huddleson (1921) for showing that this organism requires for its development a partial pressure of CO<sub>2</sub> higher than that normally present in the

atmosphere (0.03-0.01 per cent). He found that if slopes of *Br. abortus* were incubated in a glass jar containing 10 per cent  $\text{CO}_2$ , good growth occurred in 24 hours while under aerobic conditions there was no growth at all. Analysis of the gas over a culture of *B. subtilis* revealed the presence of  $\text{CO}_2$ , and it was therefore concluded that the success of Nowak's method depended on the evolution of this gas rather than on a decrease in the partial pressure of oxygen. Further work by Smith (1921) and McAlpine and Slanetz (1928b) confirmed the importance of  $\text{CO}_2$ . Smith showed that development was much better in an atmosphere of 10 per cent  $\text{CO}_2$ , than in sealed tubes and that in agar shake cultures either sealed or incubated in an atmosphere of 10 per cent  $\text{CO}_2$ , growth occurred not in a band below the surface as Bang (1897) and his co-worker Stribolt had found, but on the surface itself.

In spite of these observations it was not easy to understand why growth should occur in sealed tubes or why in the absence of added  $\text{CO}_2$ , growth in shake tubes should occur in a band below the surface.

A fuller study of the gaseous requirements of *Br. abortus* (Wilson 1931a) showed (1) that the organism would not grow anaerobically even in the presence of added  $\text{CO}_2$ , nor aerobically in its absence. (2) that growth would occur in partial pressures of oxygen varying from 0.5-99.0 per cent, provided a minimum of 0.5 per cent  $\text{CO}_2$  was added, and in partial pressures of  $\text{CO}_2$  varying from 0.5-99.0 per cent, provided a minimum of 0.5 per cent oxygen was added. (3) that the optimum partial pressure of oxygen for development was about 21 per cent, i.e. that normally present in the atmosphere, and of  $\text{CO}_2$  about 10 per cent. No evidence was obtained to suggest that a partial pressure of oxygen lower than that normally present in the atmosphere was beneficial to growth. It seemed clear therefore that neither growth in sealed tubes nor the band phenomenon in shake tubes could be due to a preference of the organism for microaerophilic conditions. Further observations (Wilson 1930) showed that  $\text{CO}_2$  was given off by burning cotton wool plugs and to a less extent by heated paraffin wax rubber stoppers and sealing wax. Analysis of the gas inside sterile sealed tubes revealed the presence of  $\text{CO}_2$  in amounts varying from about 1-3 per cent—a proportion ample to initiate growth in inoculated tubes. The larger the number of organisms inoculated the less need was there for additional  $\text{CO}_2$ , since the organisms themselves produced a certain amount of this gas. But with inocula of any size, growth was always most rapid and luxuriant when the partial pressures of oxygen and  $\text{CO}_2$  most nearly approached the optimum.

Similarly, evidence was brought (Wilson 1931b) to suggest that the band phenomenon in shake tubes was due to the necessity of an adequate concentration of  $\text{CO}_2$  (Fig. 174). It was found that this gas was given off to a certain extent by the organisms themselves and to a still greater extent by certain media, particularly those containing serum. Growth could not occur at the surface because the  $\text{CO}_2$  was given off into the atmosphere, nor could it occur in the depths of the medium because the conditions were anaerobic. It therefore commenced in a zone as near the surface as was consistent with the maintenance of an adequate partial pressure of  $\text{CO}_2$ . If the tube was sealed or was incubated in an atmosphere of 10 per cent  $\text{CO}_2$ , then growth occurred at the surface where the optimum partial pressure of oxygen existed. This explanation, when slightly amplified, was found



FIG. 173.—*Brucella abortus*

(glycerine agar slope culture 3 days 37°C. in corked tube showing character of growth on direct isolation from the tissues)

to fit the numerous observations on variation in the distance of the band from the surface, and on the so-called double-zone phenomenon, in which two bands of growth separated from each other by apparently unaltered medium, are visible.



FIG 174.—*Br abortus*

Growth in form of band situated 6-8 mm. below the surface. Bang's gelatin agar serum medium, pH 7.0, incubated aerobically.

Richardson (1935) have shown,  $\text{CO}_2$  seems to be required in a greater or less degree by practically all bacteria, and presumably plays an important part in their metabolism (see Chapter 3)

**Cultivation in the Presence of Dyes**—To Huddleson and Abell (1925) and Huddleson (1929, 1931) we owe a valuable method of distinguishing between the *melitensis abortus*, and *rus* types, depending on their ability to grow in the presence of certain aniline dyes. Without entering into the detailed technique of the method we may say that the general procedure is to prepare plates of liver agar, pH 6.6 containing 1/30 000 and 1/60 000 thionin, 1/25 000 and 1/50 000 basic fuchsin, 1/50 000 and 1/100 000 methyl violet, and 1/100 000 and 1/200 000 pyronin. The dyes used must be obtained from the National Aniline Chemical Company of New York, or standardized against these dyes. The organisms are inoculated rather heavily on to the plates, which are then incubated for 3 days aerobically, or in 10 per cent.  $\text{CO}_2$ , according to the probable nature of the strains under examination. Strains of *Br melitensis* usually grow to some extent in the presence of all four dyes, *Br abortus* strains are inhibited by thionin but grow freely in the presence of the other three, *Br rus* strains grow well in the presence of thionin, but are inhibited by basic fuchsin, methyl violet, and pyronin. Though this is the general behaviour of the three types, there is considerable variation between different strains of the same type, especially those coming from different localities (Meyer and Zobell 1932, Wilson 1933). Some strains of *melitensis* for example, may grow very poorly on the thionin, methyl violet, or pyronin plates. Southern Rhodesian strains

of *Br abortus* often have a rather greater resistance to thionin than *abortus* strains from other sources. The Danish *suis* strains are more susceptible to all dyes than the American *suis* strains, though their differential susceptibility is the same, for this reason they must be tested on plates containing only half the dye concentrations just given. If reliance is placed exclusively on this method of differentiation, confusion will not infrequently result between strains of different types. If on the other hand, it is used, as we believe it should be used, in conjunction with other methods, it will be found of considerable value. No other method, it may be noted, is so useful in distinguishing between bovine and porcine strains. The method has been subjected to some criticism (Saitta 1929, Meyer and Eddie 1930, Marshall and Jared 1930, Cerruti 1932, Maggiora Vergano 1932), but most workers have reported on it very favourably (Kristensen and Holm 1929, Kristensen 1931, Taylor, Lisbonne and Roman 1932, Grumbach and Grillicchess 1932, Meyer and Zobell 1932, Wilson 1933, Olin and Lindstrom 1934, Pagnini 1934, di Mino 1935).

**Resistance.**—The members of this group exhibit the usual susceptibility of vegetative bacteria to heat and disinfectants. In aqueous suspensions of moderate density they are destroyed by heating for about 10 minutes at 60° C, and by exposure for about 15 minutes to 10 per cent phenol. In milk they are readily destroyed by holder pasteurization. In agar cultures kept sealed at 0° C they generally live for at least 1 month, and often for considerably longer. Considerable attention has been paid to their resistance under natural conditions, and much information on this subject will be found in the Report of the Mediterranean Fever Commission (1905-07). So many factors determine the exact outcome of any given observation under natural conditions that it is dangerous to draw general conclusions from the data so collected. In favourable circumstances, however, *Br melitensis* may remain alive for 6 days in urine, 6 weeks in dust, and 10 weeks in water or soil. *Br abortus* may survive for 7 months in infected uterine exudate kept at about freezing point (Bang 1897). In raw milk at room temperature it seems to die out fairly rapidly with the production of acid. Acid production also seems to be the cause of its rapid death in butter and cheese, the organisms can rarely be found in these articles for more than a few days (Smith 1934, Pullinger 1935). It may live for a month in ice cream (Thompson 1933). *Br suis* may live on sacking for 4 weeks and in sterile faeces for 100 days in the dark (Cameron 1932, 1933).

**Metabolism and Biochemical Properties.**—The effect of temperature and H ion conditions on growth has already been considered. All the members require the presence of oxygen, most strains of *Br abortus* require in addition a partial pressure of CO<sub>2</sub> considerably higher than that found in atmospheric air. Under ordinary aerobic conditions of incubation broth cultures become markedly alkaline, owing to the production of ammonia. Litmus milk is turned weakly alkaline. Occasional haemolytic strains of *Br melitensis* have been described (Form 1927) but usually neither the *melitensis*, *abortus*, nor *suis* strains have any lytic action on blood. The effect of bile salt on growth has not been studied fully, on MacConkey's medium strains of *Br abortus*, *Br melitensis*, and *Br suis* generally give rise to small non lactose fermenting colonies after 3 or 4 days.

In ordinary sugar media no fermentation is observable. Unlike most pathogenic organisms, the members of this group are unable to produce obvious acid even from glucose. However, quantitative observations have shown that some *melitensis* and American *suis* strains, if grown in 1 per cent glucose peptone water, may utilize 5-20 per cent of the glucose within a week, while *abortus* strains are unable to use

more than 2 per cent. The acid produced is more than neutralized by the alkali formed as the result of protein breakdown, so that it is not detected by the usual indicators. McAlpine and Slanetz (1928a) have recommended the glucose utilization test as a means of differentiating between the *abortus*, *melitensis*, and *suis* types but most workers have found it unreliable, and it has now been generally discarded. There is evidence that arabinose and xylose are fermented by members of the *Brucella* group (Mallardo 1930, Coleman et al. 1930, McNutt and Purwin 1931, Silberstein 1932), but the reaction is of no differential significance.

The methyl red and Voges Proskauer tests are negative. No indole is formed. According to Zobell and Meyer (1932), all types reduce nitrates to nitrites. Nitrites are also rapidly reduced, so that the Griess-Hosvay test on nitrate broth cultures may be negative. American *suis* strains are more active than the other types in reducing nitrites. Ammonia is produced to a variable extent from peptone, urea, and asparagin. Catalase is formed, being strongest with *Br. suis* and weakest with *Br. abortus*. According to Huddleson and Stahl (1943), the degree of catalase activity is closely associated with virulence. The reducing action of these organisms is comparatively weak (Habs 1930), and in broth cultures methylene blue is often not decolorized. Tuttle and Huddleson (1934) found that liver extract broth cultures showed a negative drift to a limiting potential after 8 days of +0.15 to +0.09 volt. *Br. suis* appeared to be slightly more active than the *abortus* or *melitensis* types, but the difference was insufficient to be of value in species identification (see also Bau and Wang 1935-36). It may be noted that some strains of *Br. abortus* reduce basic fuchsin. Huddleson (1931) thought that this was a property of non-pathogenic strains but our observations do not bear this out.

**H<sub>2</sub>S Production.**—One of the most important differential criteria, to which attention was first drawn by Huddleson and Abell (1927), and Huddleson (1929), is the production of H<sub>2</sub>S. This test should be carried out on liver agar using lead acetate papers (see p. 369). Freshly isolated strains of *Br. abortus* and *Br. suis* (American variety) give off H<sub>2</sub>S for at least the first 4 days, while strains of *melitensis* produce either none at all, or only during the first 24 hours of incubation. The Danish variety of *Br. suis* forms no H<sub>2</sub>S. In the laboratory, *abortus* and American *suis* strains sometimes lose their ability to produce H<sub>2</sub>S, the test should therefore be made as soon after isolation as possible. The interpretation of this test can be summed up by saying that, while failure of a given strain to produce H<sub>2</sub>S beyond the first day does not exclude its being of *abortus* or American *suis* type, the continued production of H<sub>2</sub>S after the first day affords a strong presumption that it is not of *melitensis* or Danish *suis* type. This test has now been widely used, and to those who have realized its limitations it has given satisfaction (Favilli 1930, Kristensen 1931, Taylor, Lasbonne, and Roman 1932, Zobell and Meyer 1932, Zeller and Stockmayer 1933, Wilson 1933, Olin and Lindstrom 1934, Pagnani 1934, di Mino 1935).

**Antigenic Structure.**—It would be idle to recapitulate here the confusion that reigned for so long over the antigenic relationship of members of this group. Previous to 1918 when Evans demonstrated an antigenic affinity between *Br. melitensis* and *Br. abortus*, most workers had concerned themselves with comparison of *melitensis* and so-called *paramelitensis* strains, while for many years subsequently progress was hindered by a failure to realize the difference in antigenic structure between strains in the smooth and rough phases.

The early work of Sergeant, Gillot, and Lemaire (1908), and Negre and Ravaud (1912a, b), demonstrated the existence of strains morphologically and culturally resembling *Br melitensis* but failing to agglutinate to more than a fraction of the titre with an anti *melitensis* serum. These irregular strains were given the name of *paramelitensis*. Later on, so-called *para abortus* strains were encountered, and these were believed to represent merely a special antigenic type of *Br abortus*. Further study, however, by such workers as Favilli (1926a, b), Ross (1927a), Valenti (1927), Vidal and Abella (1928), de Antoni (1929), Zdrodowski *et al* (1930), Pampana (1931) and Pandit and Wilson (1932) showed that *paramelitensis* and *para-abortus* strains were agglutinable by non specific agents particularly acid salt peptone, and certain aniline dyes whereas freshly isolated strains of *Br melitensis* and *Br abortus* were unaffected by these agents under similar conditions. Moreover it was found that continued cultivation in broth, or better still in broth containing immune serum led to a transition of *melitensis* and *abortus* strains into *paramelitensis* and *para abortus* respectively. The transition, it may be noted, occurs much more readily with *melitensis* than with *abortus* strains, and evidence of its commencement is often noticeable within a very short time of isolation, even when the organisms are kept on solid media. There seems to be little doubt that this change, which may be accompanied by alterations in colonial appearance and by a decrease in virulence for laboratory animals, is essentially a manifestation of the S  $\rightarrow$  R variation. The antigenic change concerned is not yet clearly understood, but there is evidence that it involves a loss of the specific smooth antigen. Strains of different degrees of roughness are encountered varying from those that agglutinate to titre with a smooth serum but are slightly susceptible to non specific agglutination to those that are unaffected by a smooth serum and are incapable of remaining homogeneously distributed even in cold saline. Moreover the degree of roughness of a given strain appears to vary from one culture to another. Once roughness has appeared, it persists or recurs after an intervening period of apparent smoothness. Roughness may be tested for by boiling in saline for 2 hours (thermo agglutination test) by incubation at 37° C with 1/500 or 1/1,000 acriflavine (Alessandrini and Sabatucci 1931, Pampana 1931), by agglutination with an antiserum prepared against a completely rough strain of the corresponding type or by the ease of phagocytosis by normal leucocytes (Munger and Huddleson 1933). Little work has so far been done on the antigenic structure of the rough types. Our own incomplete observations suggest that, though there may be a common antigen to *paramelitensis*, *para-abortus*, and *parasuis* types, there are certain differences between them. One important practical point is that partly rough strains are liable to be agglutinated non specifically by the sera of normal persons and particularly of those suffering from certain febrile diseases (see Mohr 1930), and are therefore liable to lead to an erroneous diagnosis of undulant fever in routine serological work.

Turning now to the differentiation of *Br melitensis*, *Br abortus* and *Br suis* on the basis of antigenic structure, we are faced with a mass of conflicting reports most of which may be summarized by saying either that no difference was found between the three organisms or that they fall serologically into a number of different types (see Feusser and Meyer 1920, Burnet 1925, Evans 1925a, b, Ross 1927b, Cerruti 1927, Kristensen and Holm 1929, Biehing 1930, Kristensen 1931, Francis 1931, Plastringe and McAlpine 1932). The reason for this confusion is probably due to the failure of most of these workers to realize the disturbance caused by



antigenic variation of the S  $\rightarrow$  R type. If, as Wilson and Miles (1932) showed, care is taken to exclude all but absolutely smooth strains, then it is possible by means of quantitative agglutinin absorption tests to differentiate between *Br melitensis* on the one hand and *Br abortus* and *Br suis* on the other. The antigenic picture so obtained is represented in Fig. 175. It will be seen that all three types contain the same two antigens, but with a different quantitative distribution, the M antigen being in excess in the *melitensis*, the A antigen in the *abortus* and *suis* types. By carefully adjusting the absorbing dose to the titre of the serum, it is generally possible to absorb out all the minor agglutinins without removing more than a fraction of the major agglutinins. The resulting serum is therefore monospecific,

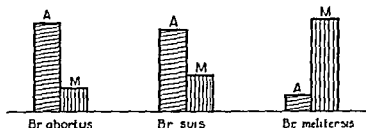


FIG. 175.—SCHEMATIC REPRESENTATION OF ANTIGENIC STRUCTURE OF *Brucella* STRAINS

and will agglutinate only those strains in which the corresponding antigen is predominant. Usually an absorbing dose standardized by opacity to match  $3\,000 \times 10^4$  col. per ml. is satisfactory for absorbing a serum diluted to 1/32–1/64 of its titre, but preliminary adjustment may be necessary before a monospecific serum can be obtained. With such a serum direct agglutination tests can be put up against strains of unknown type, and their antigenic identity established.

Why it is that a serum, from which the whole of the minor and part of the major agglutinins have been absorbed, will agglutinate only those organisms in which the corresponding antigen predominates, is rather puzzling. Why is it, for example, that a *melitensis* serum, from which the minor A agglutinin has been absorbed by an *abortus* or *suis* strain, will agglutinate only *melitensis* strains? On general grounds, *abortus* and *suis* strains, each of which contains a certain amount of M antigen, might be expected to agglutinate with such a serum to at least a quarter or half titre, since there must be ample M agglutinins in the serum to satisfy the limited number of M receptors on the organisms. The fact that no agglutination does occur suggests that the quantitative and spatial distribution of the satisfied M receptors is such that any considerable degree of adhesion between adjacent organisms is improbable. This explanation is borne out by the observations of Miles (1933), who finds that the antigenic A:M ratio in *Br abortus* is about 20:1, and in *Br melitensis* about 1:20. If, therefore, the minor antigenic surface constitutes only about 5 per cent. of the total, it may be supposed on the two stage hypothesis (see Chapter 7) that its sensitization is insufficient to decrease the salt stability of the suspension beyond the critical point, or on the lattice hypothesis that the linkages between the minor antigenic groups on the heterologous bacteria are insufficient to hold the individual bacteria in clumps.

The conclusions reached on the antigenic structure of these organisms have received confirmation from the work of Miles (1933), who has shown that the *melitensis* may be separated from the *abortus suis* types by the optimal proportion agglutination method, and from that of Habs (1933), Habs and Sievert (1935),

Sievert (1936), Olin and Lindström (1934), Olin (1935), and Veazie and Meyer (1936), using the method of quantitative absorption

As a means of typing unknown strains, direct agglutination with monospecific serum is one of the simplest and most rapid methods. Like other tests, however, for differentiation of this group, it cannot be relied upon entirely, because, as Wilson (1933) has shown, certain strains, particularly from the South East of France, may possess the antigenic structure of *abortus*, while having the biochemical and pathogenic characteristics of *melitensis*.

Several workers claim to have established an antigenic affinity between members of the *Brucella* group and those of the *Pasteurella*, *Proteus*, or *Pfeifferella* groups. Most of these conclusions have been based on the observation that a given serum agglutinated strains of both *Brucella* and some other group. By itself this is, of course, quite insufficient evidence on which to base any such conclusion, since it takes no account of non specific agglutination or of the presence of specific agglutinins to two different organisms co existing in the same serum. Absorption and other experiments have entirely failed to confirm the conclusions of these workers (for references see Priestley 1933, Wilson 1934).

**Chemical Fractionation**—The reported results of chemical fractionation of *Brucella* strains are to some extent conflicting especially with regard to the fractions exhibiting antigenic activity. The chemical characterization of many of the protein, nucleoprotein and polysaccharide fractions isolated varies greatly, and it is hard to judge the degree of purification, both chemical and serological, that has been achieved in each fraction. The more recent work has shown that a precipitin titre, by a constant antibody titration, of the order of 1:5 million represents the limit of purification so far achieved in the main antigenic fractions of this group.

The fractions isolated by the earlier workers had precipitin titres of 1:2000 to 1:50 000, and were clearly impure (Favilli and Biancalani 1932-1934, Topping 1934, Gwatkin 1935, Reiter 1936, Schapira 1936, Huggenbotham and Heathman 1936). These substances appeared to be polysaccharide in nature, and when fractions from different species were compared serologically they proved to be mainly group specific and displayed only minor degrees of species specificity. Huddleson and his colleagues (Huston, Huddleson and Hershey 1934, Hershey, Huddleson and Pennell 1935) isolated inactive proteins polysaccharides and lipoids from the three *Brucella* species, whose proportions varied with the species studied. They also found a substance "S" free from protein and polysaccharide, precipitating with antisera at 1:2,000,000 which was readily extractable from *Br. melitensis*, but was apparently bound to protein in *Br. abortus* and *Br. suis*. The "S" substance appeared to be related to the specific soluble substances isolated by Favilli and Biancalani (1932-1934). The technique of treatment of the bacterial cell with trichloroacetic acid developed by Borvin for the extraction of complete antigens from salmonellae (see Chapter 8) was applied to *Br. melitensis* by Lasbonne and Monner (1936) and to all three species of *Brucella* by Pop and his colleagues (Pop *et al.* 1938, Damboviceanu *et al.* 1938, Vangelovici *et al.* 1938). (See also Davoli 1938, Stahl and Hamann 1941). The solutions obtained by dialysing the trichloroacetic acid extracts were antigenic, toxic and precipitated in titres up to 1:100 000. The reported nitrogen content varied from 0.4 to 8 per cent, lipins 10-17 per cent, and reducing sugars on hydrolysis 4-20 per cent. Pennell and Huddleson (1937) by digestion of acetone-dried organisms extracted toxic and antigenic "endo antigens" with a precipitin titre of 1:5 million which in many respects resembled the "complete" antigen obtained by the Borvin technique. They found certain chemical differences between the endo-antigens

from the three species, and considerable cross-precipitation reactions between the three endo-antigens and their respective antisera. *Br. abortus* and *Br. suis* endo-antigens reacted similarly, though not identically, and *Br. melitensis* endo-antigen was more sharply distinguished from the other two (Pennell and Huddleson 1938). Huddleson (1943) has more recently reported that neither the "complete" antigen nor the "endo-antigen," which are both antigenic, will induce a significant degree of active immunity to experimental infection in the guinea pig but that a watery extract of crushed living *Br. abortus* or *Br. suis* is highly effective against infection with *Br. abortus*.

Miles and Pine (1933a, b) attempted to separate from *Br. melitensis* the A and M antigens postulated by Wilson and Miles (1932). By progressive degradation of antigenic material obtained by the gentlest possible treatment of the bacterium, they prepared a series of substances. The most degraded product was a formyl amino-polyhydroxy compound containing carbohydrates with a probable molecular weight of about 3,300. This substance inhibited agglutination of *Br. melitensis* by homologous antibody, but did not itself react with antisera, i.e. it was an inhibiting hapten. Next in complexity was a substance analogous with the "endo-antigens" described above. It was antigenic, toxic, and precipitated with antisera in a dilution of 1:5 million. Its molecular weight was about 1,000,000. On hydrolysis it yielded a phospholipin, phosphate, and the formyl amino compound. This antigen was in turn derived from a more polymerized material combined with a protein-like material, and finally, this last substance, combined with one third to one quarter its weight of a mixture of lipins and phospholipins, constituted the relatively unstable native antigen. The native antigen, which comprised about 10 per cent. of the dry weight of the cell, was in a state of greater aggregation than the simple antigen, and weight for weight was more antigenic and less toxic. The native antigen consequently appears to be a large complex of a protein-like substance with two sets of phospholipins differing in their readiness of separation from the complex, the second of which, together with a polymerized formyl amino-polyhydroxy compound, determines the specificity and antigenicity of the main antigen of *Br. melitensis*. In the light of these findings it seems probable that the antigenic fractions of brucellae prepared by trichloroacetic acid extraction or tryptic digestion of the bacilli consisted of mixtures in varying proportions of native antigen and its various degradation products. Miles and Pine failed to separate the A and M antigens but, as in previous work, they obtained evidence that the antigens of the S forms of *Br. abortus* and *Br. melitensis* showed a qualitative similarity, but a quantitative difference in distribution.

The apparent efficacy of live as compared with dead vaccines, led Priestley (1935) to search—without success—for a labile antigen in *Br. abortus*, similar, perhaps, to the Vi antigen of *Salmonella typhi*. Topping (1934) isolated substances of a nucleoprotein nature from *Br. abortus* and *Br. melitensis*, which were feebly reactive with heterologous and homologous sera. The reactivity may have been due to contamination with the main soluble antigen. Stahl, Pennell and Huddleson (1939) extracted non-toxic "protein nucleates" from all three species, those from S strains were serologically reactive, but only group-specific and constituted about 14 per cent. of the dry weight of the cell, those from R strains constituted about 18 per cent. of the weight of the cell, and were serologically inactive. The serological reactivity of the S protein nucleates was associated with the protein, and not with the nucleic acid part of the fractions. Stahl (1941) found 5-6 per cent. of lipins in the dry cells. They were non-toxic and serologically inactive.

**Pathogenicity**—*Br. melitensis*, *Br. abortus*, and *Br. suis* are all infective for man and animals. Though undulant fever is the most characteristic result of infection in man, numerous other clinical manifestations occur. Often the infection remains latent, causing no recognizable symptoms of disease. Since we cannot carry out large-scale experiments on man under similar conditions, it is impossible to make any definite statement on the comparative virulence of these three organisms, but the limited observations on human volunteers of Morales Otero (1929,

1930 1933) a careful study of the available epidemiological data and the frequency with which laboratory infections occur suggest that *Br melitensis* is the most pathogenic and *Br abortus* the least pathogenic of the three types. The American *suis* type appears to occupy an intermediate position. The Danish *suis* type on the other hand is probably even less pathogenic than *Br abortus* since there is no record of its ever having been responsible for disease in man.

Under natural conditions *Br melitensis* is pathogenic for goats and sheep giving rise to an infection which may be acute and accompanied by abortion but is more frequently chronic and detectable only by bacteriological examination of the milk blood or urine or by allergic skin tests. In areas where infected goats or sheep are numerous cows may also become infected (Taylor and Roman 1934). No symptoms of disease are manifest the animals do not abort when pregnant but the organisms are often excreted in the milk.

Both varieties of *Br suis* are pathogenic for pigs in which they give rise to a disease sometimes accompanied by abortion. Like *Br melitensis* the American type of *Br suis* may infect cows and be excreted in the milk (Huddleson 1931 Beattie and Rice 1934). Horses dogs and fowls are occasionally infected.

*Br abortus* probably has the widest range of pathogenicity. Besides being responsible for the almost universal and economically important disease of contagious abortion in cattle it occasionally infects other animals such as horses and dogs and less often sheep and goats. In the United States it is said to give rise to fairly extensive infection of fowls and other birds (Emmel and Huddleson 1929 1930 Emmel 1930) though the evidence for this is not wholly satisfactory. Guinea pigs have also been found infected under natural conditions (Manzullo 1935).

All three species are infective to a variable extent for laboratory animals. On the whole the guinea pig appears to be the most susceptible but rabbits rats and mice can often be infected. In the guinea pig the disease produced by parenteral inoculation with moderate doses is usually chronic and retrogressive. The brunt of the infection is borne by the reticulo-endothelial system. The resulting lesions are relatively inconspicuous and consist mainly of a non hyperæmic enlargement of the lymphatic glands some degree of enlargement of the spleen and the presence of a variable number of circular necrotic foci in the spleen and liver. In male guinea pigs abscess formation is not uncommon in the testicle or epididymis and intraperitoneal inoculation is sometimes followed by a Straus reaction. Occasionally the bones joints or other organs may be affected. The lesions are extremely variable in size and number and may be completely absent on naked-eye inspection. In infections with *Br suis* (American variety) the necrotic lesions tend to be few in number large in size and purulent in consistency. The lesions in *melitensis* and *abortus* infections on the other hand are smaller more numerous and generally non purulent except in the testicle. Numerous statements have been made about the relative virulence of the three main types for guinea pigs but workers who have had the widest experience are the most cautious in drawing conclusions. At the moment it is probably safe to conclude that there is no satisfactory method of distinguishing between them on the basis of pathogenicity to laboratory animals. Kristensen (1931) and Bang (1931) regard the Danish *suis* type as probably the least virulent while Thomsen (1934) regards it as slightly more virulent than *Br abortus*. Its differentiation from the other members is not practicable by guinea pig inoculation. The rough so-called *paramelitensis para-abortus* and *parasuis* varieties are comparatively avirulent to guinea pigs. Kritschewski and Halperin (1934) state

that a suspension of *Br. abortus* has a powerful stimulating effect on the uterine muscle of the virgin guinea pig. According to Jadassohn, Ruedmüller and Schaaf (1931) the Schultz Dale technique may be used to differentiate between the different types of *Bruceella*, the reaction in sensitized guinea pigs being type specific.

Little work has been done on monkeys but the observations of Huddleson and Hallman (1929), Weigmann (1931) and Zeller, Beller and Stockmayer (1934) suggest that *melitensis* and American swiss strains are more virulent than *abortus* strains. (For reproduction of disease in the larger animals see Chapter 75.)

No exotoxin is formed but the intraperitoneal inoculation of mice with very large numbers of virulent *Br. abortus* brings about death within a few days. The toxicity of the organisms is said to be destroyed by heating to 55° C. for 20 minutes and to be related to the virulence of the strain (Priestley and McEwen 1934).

#### Pathogenicity of *Br. melitensis* for Small Animals

**GUINEA PIGS.**—Though death within a few days may follow intracerebral inoculation (Durham 1893, Eyre 1900) or intraperitoneal inoculation with large doses, the disease set up by intramuscular or cutaneous inoculation is chronic, retrogressive and rarely proves fatal. Small numbers of organisms cannot be relied on to cause infection. Most of the animals continue to gain weight. If they are killed 6 weeks after inoculation, the following lesions may be found. Occasionally there is a local abscess containing creamy pus. The regional and the more distal lymphatic glands often show a certain amount of hyperplasia of the pale bloodless variety. The spleen may show a variable degree of enlargement, and may contain a number of circular greyish yellow necrotic foci, 0.1–0.5 mm in diameter rarely projecting above the surface. Similar foci, usually few in number, may be present in the liver. Sometimes abscesses are found in connection with the joints or bones. The organisms can be cultivated most readily from the glands, spleen, and bone-marrow. The blood usually contains agglutinins in fairly high titre. The lesions are very variable and may not be detectable by naked-eye examination. The diagnosis must always be made by testing the blood serum for agglutinins and by cultivation of the causative organism from the tissues. A titre of 1/25 or over is strongly suggestive of infection. The intradermal test with a nucleo-protein extract may be used during life for diagnostic purposes but reliance must never be placed on it alone. After 6 weeks the diagnosis becomes less easy because the infection tends to regress. Guinea pigs may also be infected by feeding by conjunctival or nasal instillation, and by inoculation of the scarified skin. Sometimes they contract the disease naturally from their fellows. (For a description of the histopathology of the disease, see Fabyan 1912.)

**RABBITS** appear to be rather less susceptible but otherwise the infection runs much the same course as in guinea pigs. The lymphatic system is less affected, and the organisms can rarely be demonstrated in the blood stream. Agglutinin formation is common, but the intradermal reaction is negative.

**RATS AND MICE.**—There is little information about the effect of inoculation of rats or mice with *Br. melitensis* but there is reason to believe that these animals are slightly susceptible to infection (see Singer, Brooks 1937).

**MONKEYS** may be infected either by feeding or by subcutaneous inoculation. Frequently an intermittent fever is set up, simulating in many respects undulant fever. If the animals are killed after a few weeks, there may be some enlargement of the lymph glands and spleen, occasionally necrotic lesions are found in the lungs or liver. Agglutinins are demonstrable in the serum, and the organisms can often be recovered from the tissues (see Bruce 1893, Hughes 1893, Horrocks and Kennedy 1906, Huddleson and Hallman 1929, Weigmann 1931, Zeller, Beller and Stockmayer 1934).

(References to pathogenicity of *Br. melitensis* for small animals: Durham 1893, Eyre 1900, 1908–09, Nicolle and Conseil 1909, Burnet 1922, Zdrodowski *et al.* 1930, Rainsford 1933.) For reproduction of disease in larger animals see Chapter 75.

### Pathogenicity of *Br. abortus* for Small Animals.

IN GUINEA PIGS a disease is set up closely resembling that caused by *Br. melitensis*. After intramuscular inoculation a local suppurating lesion is rare, but abscess formation in the testis or epididymis is not uncommon. A mild infection can be produced in RABBITS, RATS and MICE. The morbid anatomy and histopathology of the disease have been described by Fabyan (1912). In MICE inoculated subcutaneously with 10-1000 million organisms a retrogressive disease is set up. The organisms can be demonstrated in the regional lymphatic glands and the spleen for a month or so, and agglutinins are present in the blood serum. Mice may also be infected by feeding with large doses. RATS may be infected by feeding as well as by intraperitoneal inoculation, and the organisms may be excreted for a time in the urine and faeces, but unless very large doses are used the infection retrogresses (Ber 1936, Sandholm 1938, Bosworth 1938). According to Emmel and Huddleson (1929, 1930), FOWLS can be infected by feeding or parenteral inoculation. The birds stop laying and develop severe diarrhoea. There is a gradually increasing pallor of the head, comb, and wattles, emaciation, and often paralysis and death. The course of the disease ranges from about 2 to 14 weeks. Post mortem, the main lesions consist of a necrotic enteritis and degenerative changes in the liver and kidneys. The majority of other workers, however, who have studied this question, have found that, on the whole, fowls are resistant to infection except with large doses administered parenterally (McNutt and Purwin 1930, 1932, van Roekel *et al.* 1932, Beller and Stockmayer 1933). With smaller doses the organisms can rarely be recovered from the tissues. In the absence of direct cultural experiments, a rise in the agglutinin titre cannot be interpreted as necessarily indicative of infection. The conclusions of Emmel and Huddleson require confirmation before being accepted.

MONKEYS may be infected with *Br. abortus*, but they are less susceptible to it than to infection with *Br. melitensis*. (References to pathogenicity of *Br. abortus* for small animals: Schroeder and Cotton 1911, Smith and Fabyan 1912, Emmel and Huddleson 1929, 1930, Morales-Otero 1930, McNutt and Purwin 1930, 1932, Bang 1931, Pagnini 1932, Henry, Traub, and Haring 1932, Henriksen 1932, Helms, Holm, and Orskov 1932, Rainsford 1933, Ber 1933, Olin and Landstrom 1934, Huddleson 1934, Thomsen 1934, Feldman and Olson 1935, Singer Brooks 1937, Scorgie 1938). For reproduction of disease in larger animals, see Chapter 75.

### Pathogenicity of *Br. suis* for Small Animals.

Experimentally, this organism gives rise in GUINEA PIGS to a disease closely resembling that caused by *Br. melitensis*. Local abscess formation is rare, but in infections by the American type large suppurating lesions, few in number, are not uncommon in the spleen, liver, lymph glands, testicles, and joints. The Danish type appears to be less virulent than the American type. *Br. suis* appears to resemble *Br. abortus* in its infectivity for MICE, RABBITS, and FOWLS, but there is little exact information available. For MICE *Br. suis* is said to be more virulent than *Br. abortus* (Singer Brooks 1937). For MONKEYS it appears to be perhaps even more virulent than *Br. melitensis*. (References to pathogenicity of *Br. suis* for small animals: Smith 1926a, Hardy *et al.* 1930, Cotton 1932, Thomsen 1934, Huddleson 1934, Feldman and Olson 1935). For reproduction of disease in larger animals, see Chapter 75.

**Variation.**—It has already been mentioned that under artificial conditions of cultivation *Brucella* strains, particularly of the *melitensis* type, tend to undergo a change which is characterized by a gradual loss of specific, and gradual increase of non specific, agglutinability, together with a decrease in virulence to animals. This change appears to be an example of the smooth → rough variation. Whether the change is accompanied by any corresponding alteration in the morphological and colonial appearances of the organisms is still a little doubtful, though there is reason to believe that some change does occur. The descriptions of various workers,

however, many of whom have used different media, are so difficult to summarize, that we shall content ourselves with giving references to some of the more important papers on this subject (Henry 1928 1933, Plastringe and McAlpine 1930, Marshall and Jared 1930, 1931, Morales Otero 1931, Grumbach and Grillichess 1932). The growth of smooth and rough forms in the presence of dyes appears to be very much the same, but there is some evidence that the  $S \rightarrow R$  variation may be accompanied by a decrease in biochemical activity, particularly in the production of  $H_2S$ . Though it is true that some rough *abortus* and American *suis* strains still produce  $H_2S$ , it is equally true that many do not. Since practically all freshly isolated smooth strains produce  $H_2S$ , it seems not improbable that the loss of this property on continued subcultivation is a manifestation of the  $S \rightarrow R$  variation.

**Classification and Identification**—As has already been pointed out, the inclusion of *Br. tularensis* in the *Brucella* group is largely tentative. This organism may be distinguished from the other three members on the basis of morphological, cultural, biochemical, antigenic, and pathogenic properties.

The main difficulty lies in distinguishing between *Br. melitensis*, *Br. abortus*, and *Br. suis*. This difficulty is accentuated by the fact that within each species there are a number of sub-types differing from one another in minor respects and approaching closely to the sub-types of adjacent species (see Meyer and Zobell 1932, Wilson 1933). These sub-types are often associated with some special topographical distribution. *Melitensis* strains, for example, from Malta, may differ from those from Palestine or from the South of France. Southern Rhodesian *abortus* strains differ from European or American strains. The American *suis* strains differ from the Danish strains, and so on. For purposes of identification, therefore, the fullest possible examination is required, and no strain should be definitely allocated to a particular species without a careful study by all available bacteriological methods including  $CO_2$  sensitivity, growth in the presence of dyes,  $H_2S$  formation, antigenic analysis, and if possible virulence. Once the infecting type has been firmly established, help is often afforded by a knowledge of the animal source and country of origin. For instance, in this country *Br. suis* has never been found, and *Br. melitensis* has been observed only once among cattle on a small number of farms in the Midlands (see Duke 1940, Wilson 1940), so that any indigenous strain of *Brucella* isolated from man, horses, dogs, or cattle is probably of the *abortus* type. In any country, strains isolated from sheep or goats are usually of the *melitensis* type, from pigs of the *suis* type, from cattle, horses, and dogs of the *abortus* type, while strains isolated from man usually belong to that type which is most prevalent in the neighbouring animal population. Exceptions, however, are not uncommon, so that too much reliance should not be placed on this particular aid to identification.

Though there are numerous sub-types of *Brucella* with particular geographical locations, indicating that the members of the group are relatively labile and responsive to environmental changes, no one has yet succeeded in converting one type into another. Even prolonged residence of the *melitensis* and *suis* types in cows, and of the *abortus* type in sheep, seems to have no effect on the type of organism introduced. For practical purposes, therefore, the main types can be regarded as constant. Table 53 summarizes the chief differential features of members of this group. (Useful reviews of the *Brucella* group will be found in papers by Kristensen 1931, Taylor, Lisbonne and Roman 1932, Zeller 1933, Habs 1933, Wilson 1933, Huddleson 1934, Thomsen 1934, Olin and Landström 1934.)

TABLE 53  
CLASSIFICATION OF *Brucella* GROUP

Type	Usual Habitat	Growth in absence of extra CO <sub>2</sub>	Growth in presence of				H <sub>2</sub> S formation	Antigenically
			Thionin	Basic Fuchsin	Methyl Violet	Pyronin		
<i>melitensis</i>	Goats sheep	+	+	+	+	+	—	<i>melitensis</i>
<i>abortus</i>	Cows, horses	—	—	+	+	+	+	<i>abortus</i>
<i>American suis</i>	Pigs	+	+	—	—	—	+	<i>abortus</i>
<i>Danish suis</i>	Pigs	+	+	—	—	—	—	<i>abortus</i>

### *Brucella melitensis*

*Synonyms*—*M. melitensis*, *Allkaligenes melitensis*.

*Isolation*—Isolated by Bruce (1887) from the spleen of patients dying of Malta fever

*Habitat*—Strict parasite living in goats, sheep and man

*Morphology*—Small bacilli, 0.6–1.2  $\mu$  long  $\times$  0.5–0.7  $\mu$  broad coccoid forms abundant. Axis straight, ends rounded sides bulging or parallel. Arranged singly, in pairs end to end in small groups, or—especially in liquid media—in short chains of four to six members. Non motile. Bipolar staining not uncommon. Gram negative.

*Agar Plate*—48 hours at 37° C. Small, round, convex, amorphous colonies about 0.5 mm in diameter. Smooth, glistening surface, entire edge, translucent, greyish white by reflected light, almost colourless by transmitted light, consistency butyrous, emulsification easy. 6-day colonies slightly larger and greyish yellow. No differentiation.

*Agar Stroke*—48 hours at 37° C. Poor to moderate, partly confluent, slightly raised, translucent growth, with pitted surface, and edge formed of single colonies. After a week the agar is turned brownish and crystals may appear.

*Gelatin Stab*—10 days at 22° C. Poor to moderate, filiform, greyish white growth, consisting of very small colonies closely packed, extends to bottom of tube. No surface growth and no liquefaction.

*Broth*—24 hours at 37° C. Poor growth with slight turbidity, no surface growth and no deposit. After 10 days there is an abundant growth with moderate turbidity, and a moderate powdery deposit disintegrating completely on shaking. Later the deposit becomes very viscous, and almost impossible to disintegrate.

*Loeffler's Serum*—48 hours at 37° C. Moderate, slightly raised chiefly confluent growth of yellowish colour. No liquefaction.

*Potato*—6 days at 37° C. Thin, mostly confluent growth of greyish brown colour. After 14 days the growth has a café-au-lait or chocolate colour.

*Shale Agar*—4 days at 37° C. Growth of tiny, discrete colonies situated at the surface, or some distance below the surface, exact position depends on the CO<sub>2</sub> sensitivity of the strain.

*Liver Agar Plates containing Dyes*—3 days at 37° C. Usually some growth in presence of thionin, basic fuchsin, methyl violet and pyronin, but reaction varies considerably according to source of origin of strain.

*MacConkey Agar Plate*—7 days at 37° C. Small circular, convex, amorphous, yellowish colonies, 0.1–1.0 mm. in diameter, with smooth surface and entire edge. May appear slightly mucoid.



**Resistance.**—Not specially resistant. Killed by moist heat at 60° C. in 10 minutes and by 1.0 per cent. phenol in about 15 minutes. In the dried, powdered condition they may survive for 3 months. Sealed agar slope cultures at room temperature may remain alive for 1-6 months.

**Metabolism.**—Aerobic, no growth under strictly anaerobic conditions. Growth is often improved by 10 per cent. CO<sub>2</sub>. Opt. temp 37° C., limits 20-40° C. Opt. H ion concentration pH 6.6-7.4. Growth slightly improved by glucose, glycenne liver extract, blood and serum. Brown pigment formed on potato and sometimes in old agar cultures. Broth turned alkaline—to pH 8.0 or even higher. Growth in all media is relatively slow. Some growth on MacConkey's medium. Does not hemolyse blood.

**Biochemical.**—No carbohydrates fermented. L.M. turned slightly alkaline. Indole —, M.R. —, V.P. —, Nitrates and nitrites reduced. NH<sub>3</sub> sometimes +, H<sub>2</sub>S —, M.B. reduced, catalase +.

**Antigenic Structure.**—Only one serological type known. Appears to contain the same antigens as *Br. abortus* and *Br. suis*, but in different quantitative proportions. Provided absolutely smooth strains are used, it may be differentiated from *Br. abortus* and *Br. suis* by quantitative absorption of agglutinins. The rough variant incorrectly called *Br. paramelitensis* is agglutinable by non specific agents, but not by a serum prepared against the smooth form.

**Pathogenicity.**—Causes undulant fever in man, and a septicæmic infection of goats and sheep, sometimes accompanied by abortion. May infect cows and be excreted in the milk. Experimentally, it is pathogenic to a variable degree for man, goats, sheep, monkeys, and the small laboratory animals. The rough variant is avirulent.

### *Brucella abortus*

**Isolation.**—By Bang (1897) from cows with infectious abortion.

**Habitat.**—Strict parasite occurring in cattle, horses, dogs, and man.

**Morphology.**—Similar to that of type species but usually more bacillary, rods reach 1.5  $\mu$  in length or on special media even 3.0  $\mu$ . May be capsulated.

**Cultural Characters.**—Similar to those of type species, except that growth of the bovine type whether isolated from cattle or from man usually occurs only in the presence of added CO<sub>2</sub>, preferably 5-10 per cent. In shake agar cultures growth occurs 1-2 mm or more below the surface, and extends downwards for 1 cm. Old laboratory cultures grow freely under aerobic conditions, no growth under strictly anaerobic conditions.

**Persistence.**—Similar to type species. In uterine exudate kept in the ice-chest it survives for 9 months. May live in sterile water for 3 or 4 months. Readily killed in milk by holder pasteurization. May survive in ice-cream for a month.

**Metabolism.**—Similar to type species, but as most strains on isolation require CO<sub>2</sub>, the optimum H ion concentration of media is about pH 6.6. Brown coloration in old agar cultures less common than with *Br. melitensis*.

**Biochemical.**—Similar to type species, but nearly all strains on isolation produce H<sub>2</sub>S in liver agar for at least 4 days.

**Antigenic Structure.**—Appears to possess the same antigens as *Br. melitensis* but distributed in different quantitative proportions. Provided absolutely smooth strains are used it may be differentiated from *Br. melitensis* by quantitative absorption of agglutinins. The rough variant, incorrectly referred to as *Br. para-abortus*, is agglutinable by non specific agents.

**Pathogenicity.**—Causes epizootic abortion in cattle, fistulous withers in horses, and a mild septicæmic infection in dogs. Is said to infect rats. Gives rise to undulant fever in man. Experimentally, it is pathogenic to a variable degree for man, cattle, horses, dogs, fowls, monkeys, and the small laboratory animals. The rough variant is avirulent.

*Brucella suis*

*Isolation*—American type by Traum (1914) from the foetus of a sow and Danish type by Thomsen (1931)

*Habitat*—Strict parasite, occurring in pigs and man

*Morphology*—Similar to *Br. abortus*

*Cultural Characters*—Similar to those of type species except that growth is never improved by addition of CO<sub>2</sub>. In shake agar cultures growth occurs on the surface. No growth under strictly anaerobic conditions. The American type grows rather more freely than the Danish type. On liver agar plates both types grow in the presence of thionin, but are inhibited by basic fuchsin, methyl violet, and pyronin. The Danish type has the same differential susceptibility as the American type but is rather more susceptible to all dyes; consequently half the usual concentrations of dye should be employed when testing it.

*Resistance*—Similar to that of type species and *Br. abortus*

*Metabolism*—Similar to type species, but growth is not improved by CO<sub>2</sub>. Brown coloration in old agar cultures less common than with *Br. melitensis*.

*Biochemical*.—Similar to type species but American type produces H<sub>2</sub>S in liver agar for at least 4 days, Danish type produces no H<sub>2</sub>S.

*Antigenic Structure*—Appears to possess the same antigens as *Br. melitensis*, but distributed in quantitative proportions nearer those of *Br. abortus* than *Br. melitensis*. Provided absolutely smooth strains are used it may be differentiated from *Br. melitensis*, but not from *Br. abortus*, by quantitative absorption of agglutinins. The rough variant, incorrectly referred to as *Br. parvus* is agglutinable by non specific agents.

*Pathogenicity*—Gives rise to a disease of pigs, which may be accompanied by abortion and to undulant fever in man. May infect cows and be excreted in the milk. Experimentally, it is pathogenic to a variable degree for man, pigs, cows, monkeys, and the small laboratory animals. Possibly pathogenic to some degree for horses and dogs. The rough variant is avirulent.

*Brucella tularensis*

This organism is a tiny, non motile, Gram negative bacillus which was isolated by McCoy and Chapin in 1912 from rodents suffering from tularemia (see Chapter 75). In the animal body it occurs as a coccoid or rod shaped organism surrounded by a clear area which probably represents a capsule. The diameter of the organism is 0.3–0.7  $\mu$  long by 0.2  $\mu$  wide, the diameter with the capsule is 0.4–1.0  $\mu$  by 0.3–0.5  $\mu$ . The organisms stain best with carbol fuchsin or aniline gentian violet, with methylene blue they stain very poorly and show no capsule. In culture, coccoid forms alone are seen (Wherry and Lamb 1914) a capsule is visible if the organisms are mixed with serum. No growth occurs in the usual media. It was first cultivated on Dorset's egg but later it was found that coagulated egg yolk was more satisfactory (McCoy 1912). On this medium the maximum growth is reached in 2 days; it is pale, translucent slightly mucoid and pearly in appearance, not easily distinguishable from the medium, it is readily emulsifiable. Growth occurs also on glucose blood agar, glucose serum agar and blood agar slopes provided that a piece of rabbit's spleen is rubbed over the surface and then left in the condensation water (Francis and Lake 1922), and on agar to which 0.02 per cent. of cystine is added (Francis 1922, 1923). On these media the organism should be subcultured every other day but on egg yolk it may remain viable for 3 months (Wherry and Lamb 1914). Shaw and Hunnicutt (1930) recommend a medium composed of brain veal infusion agar pH 7.6 containing 5 per cent rabbit serum, 1 per cent dextrose, and 0.05 per cent cystine while Kudo

(1934) prepares a mixture of 60 per cent egg yolk and 40 per cent rabbit serum sterilized at 70°-75° C on 3 successive days. Liquid media can also be used (Tamura and Cibby 1943). Steinhaus, Parker and McHoe (1944) recommend for this purpose a medium containing 1 per cent dextrose, 0.15 per cent cystine and 0.5 per cent hæmoglobin. *Br. tularensis* grows well in the developing chick embryo multiplying particularly in the ectodermal epithelial cells. When inoculated on to the chorio-allantoic membrane it leads to the death of the embryo in 3-4 days. In both these respects it resembles *Br. melitensis* (Buddingh and Womack 1941; Ransmeier 1943). Under suitable conditions the organism is said to produce acid in glucose and glycerol and usually in maltose, mannose and levulose (Downs and Bond 1935; Francis 1947) but the acid produced seems to be very slight and may possibly be of the order of that formed by *melitensis* and American *suis* strains.

Moist heat at 50°-60° C is fatal in 10 minutes. Antigenically *Br. tularensis* is allied to the other members of the *Bruceella* group. Francis and Evans (1936) found that a serum prepared against *Br. tularensis* agglutinated *Br. melitensis* and *Br. abortus* to about  $\frac{1}{2}$  or  $\frac{1}{4}$  of the titre. Neither organism was able, however, to absorb the homologous agglutinins from a *tularensis* serum. *Br. tularensis* was agglutinated to a low titre by anti-*melitensis* and anti-*abortus* sera but was unable to absorb the homologous agglutinins from these sera. The 3 strains of *Br. tularensis* examined appeared to be antigenically homogeneous.

Under natural conditions it gives rise to tularemia in rodents—especially ground squirrels and jack rabbits—and occasionally in man. Sheep are sometimes affected (Parker and Dade 1939). Experimentally the disease can be reproduced in ground squirrels, gophers, guinea pigs, rabbits, mice and monkeys, rats are more resistant (Dieter and Rhodes 1936), cats, dogs and pigeons appear to be immune. Feeding-nasal instillation, cutaneous, subcutaneous, intraperitoneal and conjunctival infection are all successful. After subcutaneous infection of the guinea pig death occurs in 5 to 8 days. Post mortem there is a whitish membrane-like area at the site of inoculation, the regional lymphatic glands may be enlarged and caseous, the spleen is enlarged, very dark in colour and contains discrete yellowish white caseous granules up to 1 mm. in diameter projecting slightly above the surface, there are numerous granules in the liver, focal necrotic areas are sometimes present in the bone marrow (Lille and Francis 1933), the lungs are rarely involved. The bacilli are present in large numbers in the blood and organs, as little as 0.000 000 1 ml. of the heart's blood may prove infective for fresh animals. The virulence of the organism may decline in culture so that instead of causing an acute or subacute disease in guinea pigs it gives rise to a chronic disease from which the animal often recovers (McCoy 1912; Foshay 1932). Strains of lowered virulence have also been isolated directly from ticks (Davis *et al.* 1934). The organism is extremely dangerous to handle in the laboratory and large numbers of workers have contracted the infection.

#### REFERENCES

- ALESSANDRINI, A. and SARATUCCI, M. (1931) *Ann. Igiene (Sper.)* **41**, 29-83.  
 ANTONI, V. DE (1929) *Boll. Ist. sieroter. Milano*, **8**, 631.  
 BANG, B. (1897) *Z. Tiermed.* **1**, 241.  
 BANG, O. (1931) *2me Congr. int. Path. comp.* **2**, 95.  
 BAU, K. H. and WANG, H. (1935-36) *Z. Hyg. Infektkr.* **117**, 399.  
 BEATTIE, C. P. and RICE, R. M. (1934) *J. Amer. med. Ass.*, **102**, 1670.  
 BELLEZ, H. and STOCKMAYER, W. (1933) *Dtsch. tierärztl. Wschr.*, **41**, 551.  
 BER, A. (1933) *Z. Infektkr. Haust.* **44**, 179. (1936) *C. P. Soc. Biol.*, **122**, 845.  
 BEVAN, L. E. W. (1930) *Brit. med. J.* **ii**, 967.  
 BILLING, R. (1930) *Z. Hyg. Infektkr.* **111**, 728.  
 BOSWORTH, T. J. (1938) *J. comp. Path.* **50**, 345.  
 BRUCE, D. (1887) *Practitioner* **39**, 161. (1893) *Ann. Inst. Pasteur* **7**, 289.

- BUDDINGH, G J and WOMACK, F C (1941) *J exp Med*, 74, 213
- BURNET, E (1922) *Arch Inst Pasteur Afrique nord*, 2, 165, (1925) *Arch Inst Pasteur Tunis*, 14, 247
- CAMERON, H S (1932) *Cornell Veterinarian*, 22, 212, (1933) *Rep N Y St vet Coll* 1931-32, No 18
- CERRUTI, C F (1927) *Boll Ist sieroter Milano*, 6, 425, (1932) *Ibid*, 11, 400
- COLEMAN, M R., OWEN, H H, and DACEY, H G (1930) *J lab clin Med*, 15, 641
- COTTON, W E (1932) *J agric Res*, 45, 705
- DAMBOVICEANU, A, BARBER, C, POP, A, and MARINOV, I (1938) *C R Soc Biol* 127, 736
- DAVIS, G E, PHILIP, C B, and PARKER, R R (1934) *Amer J Hyg*, 19, 449
- DAVOLI, R. (1938) *Boll Sez ital Soc. int. Microbiol*, 10, 124
- DIETER, L V and RHODES, B (1926) *J infect Dis*, 38, 541
- DOWNS, C M and BOND, G C (1935) *J Bact*, 30, 485
- DUKE, F W (1940) *Lancet*, ii 517
- DUNCAN, J T (1928) *Trans roy Soc. trop Med Hyg*, 22, 269
- DURHAM, H E (1898) *J Path Bact*, 5, 377
- EMMEL, M W. (1930) *J Amer vet med Ass*, 76, 452, 564
- EMMEL, M W and HUDDLESON, I F (1929) *J Amer vet med Ass*, 75, 578 (1930) *Ibid*, 76, 449
- EVANS, A C (1918) *J infect Dis*, 22, 580, (1925a) *Amer J trop Med*, 5, 419 (1925b) *Bull U S Hyg Lab*, No 143
- EYRE, J W H (1905) *Rep Comm Medit Fev, Lond*, Part II, p 67 (1908-9) *Proc roy Soc Edin*, 29, 537
- FABYAN, M (1912) *J med Res*, 26, 441
- FAVILLI, G (1926a) *Sperimentale*, 80, 41, (1926b) *Ibid*, 80, 396, (1930) *Ibid* 84, 287
- FAVILLI, G and BIANCALANI, G (1932) *Sperimentale*, 86, 357 (1934) *Ibid*, 88, 337
- FELDMAN, W H and OLSON, C (1935) *J infect Dis*, 57, 212
- FEUSHER, M L and MEYER, K F (1920) *J infect Dis*, 27, 185
- FLEMING, A (1919) *Lancet* i 133
- FORESTI C (1935) *Nuova Vet.*, 13, 11
- FORNI, G (1927) *G Batt Immun*, 2, 823
- FOSHAY, L (1933) *J infect Dis*, 51, 280
- FRANCIS, E (1922) *Publ Hlth Rep, Wash*, No 17, 37, 937, (1923) *Ibid*, No 25, 38, 1991, (1931) *Ibid*, 46, 2416, (1942) *J Bact*, 43, 343
- FRANCIS, E and EVANS, A C (1926) *Publ Hlth Rep, Wash*, No 26 41, 1273
- FRANCIS, E and LAKE, G C (1922) *Publ Hlth Rep, Wash*, No 3 37, 63
- GLADSTONE, G P, FILDES, P, and RICHARDSON, G M (1935) *Brit J exp Path* 16, 335
- GOODPASTURE, E W and ANDERSON, K. (1937) *Amer J Path*, 13, 149
- GRUMBACH, A and GRILLICHESS, R K. (1932) *Zbl Bakt*, 126, 321
- GWATHIN, R (1935) *Canad J Res*, 12, 115, 133
- HAES, H (1930) *Zbl Bakt*, 118, 89, (1933) *Zbl ges Hyg*, 28, 431
- HAES, H and SIEVERT, L (1935) *Dtsch med Wschr*, 61, 1398
- HARDY, A V, JORDAN, C F, BORTS, I H, and HARDY, G C (1930) *Nat Inst Hlth Bull*, No 168
- HELMS, T, HOLM, P, and ØRSKOV, J (1932) *Z ImmunForsch*, 75, 55
- HENRICSSON, E (1932) "Epizootischer Abortus und Undulantfieber" Isaac Marcus Boktryckeri Aktiebolag Stockholm
- HENRY, B S (1928) *Proc Soc exp Biol, N Y*, 26, 101, (1929) *Ibid*, 27, 8 (1933) *J infect Dis*, 52, 374 403
- HENRY, B S, TRAUM, J, and HARING, C M (1932) *Hilgardia*, 6, 355
- HERSHEY, A D, HUDDLESON, I F, and PENNELL, R B (1935) *J infect Dis*, 57, 183
- HIGGINSBOTHAM M and HEATHEMAN, L S (1936) *J infect Dis*, 59, 30
- HOLTH, H (1911) *Z InfektKr Haustiere*, 10, 207
- HORROCKS, W H and KENNEDY, J C (1906) *Rep Comm Medit Fev, Lond*, Part IV, p 37
- HUDDLESON, I F (1921) *Cornell veterinarian*, 11, 210, (1929) *Mich State College agric Exp Sta, Tec Bull*, No 100, (1931) *Amer J publ Hlth*, 21, 491, (1934) *Brucella Infections in Animals and Man* Commonwealth Fund New York, (1940) *J Amer. vet. med Ass*, 96, 708, (1943) *Tech Bull Mich. agric Exp Sta*, No 182 45
- HUDDLESON, I F and ABELL, E (1927) *J Bact*, 13, 13, (1928) *J infect Dis*, 43, 81
- HUDDLESON, I F and HALLMAN, E T (1929) *J infect Dis*, 45, 293
- HUDDLESON, I F, HASLEY, D E, and TORREY, J P (1927) *J infect Dis*, 40, 352
- HUDDLESON, I F and STAHL, W H (1943) *Tech Bull Mich agric Exp Sta*, No 182, p 57
- HUDDLESON, I F and WINTER, O B (1927) *J infect Dis*, 40, 476
- HUGHES, M L (1933) *Ann Inst Pasteur*, 7, 628

- HUSTON, R. C., HUDDLESON, I. F., and HERSHEY, A. D. (1934) *Tech. Bull., Mich. agric. Exp. Sta.*, No. 137.
- JAPASSOHN, W., FIEDMULLER, L. and SCHAAF, F. (1934) *Klin. Wochschr.*, 13, 879.
- KIRBY, G. P. (1933) *J. Bact.* 37, 493.
- KING, R. O. C. (1934) *Aust. vet. J.*, 10, 93.
- KOSER, S. A., BRESLOVE, B. B., and DORFMAN, A. (1941) *J. infect. Dis.*, 69, 114.
- KOSER, S. A. and WRIGHT, M. H. (1947) *J. infect. Dis.*, 71, 56.
- KRISTENSEN, M. (1931) *Zbl. Bakt.*, 120, 179.
- KRISTENSEN, M. and HOLM, P. (1929) *Zbl. Bakt.*, 112, 281.
- KRITSCHESKI, I. L. and HALPERIN, E. P. (1934) *Z. Immunforsch.*, 82, 421.
- KUDO, M. (1934) *Jap. J. exp. Med.*, 12, 371.
- LILLIE, R. D. and FRANCIS, E. (1933) *Publ. Hlth Rep., Wash.*, 48, 1127.
- LISBONNE, H. and MONIER, P. (1936) *C. R. Soc. Biol.*, 123, 1114.
- MCALPINE, J. G. and SLANETZ, C. A. (1928a) *J. infect. Dis.*, 42, 66, 73. (1928b) *Ibid.*, 43, 232.
- MCCOY, G. W. (1912) *Publ. Hlth Bull., Wash.* No. 53, p. 17.
- MCCOY, G. W. and CHAPIN, C. W. (1912) *J. infect. Dis.*, 10, 61.
- MCCUTT, S. H. and PURVIS, P. (1930) *J. Amer. vet. med. Ass.*, 20, 330. (1931) *J. infect. Dis.*, 48, 292. (1932) *J. Amer. vet. med. Ass.*, 81, 641.
- MAGGIORA VERGANO, A. (1932) *Boll. Ist. sieroter. Milano*, 11, 400.
- MAILLARD, C. A. (1930) *J. trop. Med. Hyg.*, 33, 125.
- MANZULLO, A. (1935) *Fed. Biol.*, No. 47, 211.
- MARSHALL, M. S. and JARVIS, D. (1930) *Proc. Soc. exp. Biol.*, N. Y., 27, 525. (1931) *J. infect. Dis.*, 49, 318.
- MARTINI, G. (1933) *Boll. Ist. sieroter. Milano* 14, 431.
- MEVIANI, C. (1934) *Nuova Fed.*, 12, 83.
- MESSIERI, A. (1935) *Nuova Fed.*, 13, 1, 13.
- MEYER, A. F. and EDDIE, B. (1930) *J. Lab. clin. Med.*, 15, 447.
- MEYER, K. F. and SHAW, E. B. (1930) *J. infect. Dis.*, 27, 173.
- MEYER, K. F. and ZOBELL, C. E. (1932) *J. infect. Dis.*, 51, 72.
- MICKLE, W. A. (1940) *J. infect. Dis.*, 66, 271.
- MILES, A. A. (1933) *Brit. J. exp. Path.*, 14, 43. (1939) *Ibid.*, 20, 63.
- MILES, A. A. and PIRIE, N. W. (1939a) *Brit. J. exp. Path.*, 20, 83, 109, 278. (1939b) *Biochem. J.* 33, 1709, 1716.
- MINGLE, C. K. and MANTHEI, C. A. (1941) *Amer. J. vet. Res.*, 2, 181.
- MINGO, G. DI. (1935) *Boll. Ist. sieroter. Milanese*, 14, 123.
- MOHR, W. (1935) *Z. Immunforsch.*, 86, 235.
- MORALES OTERO, P. (1939) *Porto Rico J. publ. Hlth trop. Med.*, 5, 144. (1930) *Ibid.*, 6, 3. (1931) *Porto Rico J. publ. Hlth trop. Med.*, 7, 233. (1933) *J. infect. Dis.*, 52, 54.
- MUNGER, M. and HUDDLESON, I. F. (1935) *J. Bact.*, 35, 255.
- NÉGEZ, L. and RAYNAUD, M. (1912a) *C. R. Soc. Biol.*, 72, 791. (1912b) *Ibid.*, 72, 1052.
- NEIVA, C. (1934) *Brasil-Médico* 48, 421.
- NICOLE, C. and COVSEIL, E. (1909) *C. P. Soc. Biol.*, 67, 267.
- NOWAK, J. (1908) *Ann. Inst. Pasteur* 22, 541.
- OLIN, G. (1935) "Studien über das Undulantfieber in Schweden." Isaac Marcus Boktryckeri Aktiebolag Stockholm.
- OLIN, G. and LINDSTROM, B. (1934) *Zbl. Bakt.*, 131, 257.
- PAONINI, U. (1932) *G. Batt. Immun.*, 9, 1004. (1934) *Boll. Ist. sieroter. Milano* 13, 145. (1935) *G. Batt. Immun.* 15, 847.
- PAMPANA, E. J. (1931) *Ann. Igien. (Sper.)*, 41, 537.
- PANDIT, S. R. and WILSON, J. S. (1932) *J. Hyg., Camb.*, 32, 45.
- PARKER, R. R. and DADE, J. S. (1939) *Publ. Hlth Rep., Wash.*, 44, 126.
- PEVNEY, R. B. and HUDDLESON, I. F. (1932) *Tech. Bull. Mich. agric. exp. Sta.* No. 156, 1. (1933) *J. exp. Med.*, 63, 73, 83.
- PLASTRIDGE, W. V. and MCALPINE, J. G. (1930) *J. infect. Dis.*, 48, 315. (1932) *J. infect. Dis.*, 50, 555.
- POP, A., DAMBOVICANU, A., BARBER, C. and MARINOV, I. (1935) *C. R. Soc. Biol.*, 127, 733.
- PREISS, H. (1903) *Zbl. Bakt.*, 33, 190.
- PRIESTLEY, F. W. (1933) *J. comp. Path.*, 46, 39. (1935) *Fet. Rec.* 50, 137.
- PRIESTLEY, F. W. and MCEWEN, A. D. (1935) *J. comp. Path.*, 51, 285.
- PULLINGER, E. J. (1935) *Lancet*, i, 1312.
- RAINSFORD, S. G. (1933) *Irish J. med. Sci.*, April, p. 150.
- RANSMEIER, J. C. (1943) *J. infect. Dis.*, 72, 86.
- REIMANN, H. A. (1932) *Amer. J. Hyg.*, 16, 206.
- REITER, D. O. (1936) *J. infect. Dis.*, 58, 45.
- Report. (1930-7) *Comm. Mediterranean Fever*, Parts I, II, III, IV. Harrison and Sons, London.

- ROEDEL, H VAN, BULLIS, K L, FLINT, O S, and CLARKE, M K (1932) *J Amer vet med Ass*, 80, 641
- KOPP, R. S DE (1944) *J. comp Path*, 54, 53
- ROSS, G R (1927a) *J. Hyg, Camb*, 26, 279, (1927b) *Ibid*, 26, 403
- SAITTA, S (1929) *G Batt Immun*, 4, 307
- SANDHOLM, A (1938) *Z InfektKr Haust*, 53, 201
- SANTIS, M DE (1933) *Boll Ist sieroter Milano*, 12, 846, (1935) *Ibid*, 14, 113
- SCHAFIRA, G B (1936) *Sperimentale*, 90, 450
- SCHROEDER, E C and COTTON, W E (1911) *Bur Animal Ind*, 28th Ann Rep, p 139
- SCHWARZMAIER, E (1936) *Z InfektKr Haust*, 49, 309
- SCORGIE, N J (1938) *J Path Bact*, 46, 165
- SERGEANT, E, GILLOT, V, and LEMAIRE, G (1908) *Ann Inst Pasteur*, 22, 209
- SHAW, F W and HUNNICUTT, T (1930) *J Lab. clin. Med*, 18, 46
- SIEVERT, L. (1936) *Z ImmunForsch*, 89, 249
- SILBERSTEIN, W (1932) *Z Hyg InfektKr*, 114, 177
- SINGER BROOKS, C H (1937) *J infect Dis*, 60, 265
- SMITH, J (1934) *J Hyg, Camb*, 34, 242
- SMITH, T (1924) *J exp Med*, 40, 219, (1926a) *Ibid*, 43, 207, (1926b) *Ibid*, 43, 317
- SMITH, T and FABYAN, M. (1912) *Zbl Bakt*, 61, 549
- STAPSETH, H J (1920) *Misch agric exp Sta, Tec Bull*, No 49
- STAHL, W H (1941) *Tech Bull Misch agric Exp Sta*, No 177, 29
- STAHL, W H and HAMANN, E E (1941) *Ibid*, No 177, 17
- STAHL, W H., PENNELL, R. B and HUDDLESON, I F (1939) *Ibid*, No 168, 1
- STEINHAUS, E A, PARKER, R R, and MCKEE, M T (1944) *Publ. Hlth Rep Wash* 59, 78
- TAMURA, J T and GIBBY, I W (1943) *J Bact*, 45, 361
- TAYLOR, R M., LISBOVNE, M., and ROMAN, G (1932) *Ann Inst Pasteur*, 49, 281
- TAYLOR, R M., VIDAL, L. F, and ROMAN, G (1934) *C R Soc Biol* 116, 132
- THOMSEY, A. (1931) *Rev gen Méd vet*, 40, 457, (1933) *Zbl Bakt*, 130, 257, (1934) "Brucella Infection in Swine" *Acta path microbiol. Scand, Suppl* No 21
- THOMPSON, R (1933) *Canad med Ass J*, 29, 9
- TOPPING, L. E (1934) *J Path Bact*, 39, 665
- TOSATTI, E (1934) *Pathologica*, 26, 247
- TRAUM, J E (1914) *Rep Chief Bur Anim Industry* p 30
- TUTTLE, C D and HUDDLESON, I. F (1934) *J infect Dis*, 54, 209
- VALENTE, E (1927) *Biochim. Terap exper*, 14, 77
- VANGHELOVICI, M., DAMBOVICEANU, A, BARBER, C, and POP, A (1938) *C R Soc Biol* 127, 739
- VEALIE, L. and MEYER, K F (1936) *J infect. Dis*, 58, 280
- VIDAL, J and ABELLA, R (1928) *C R Soc Biol*, 99, 1271
- VITTORE, R (1935) *Boll Sez ital, Soc int Microbiol*, 7, 277
- WEIGMANN, F (1931) *Zbl Bakt*, 121, 318
- WHEBBY, W B and LAMB, B H (1914) *J infect Dis*, 15, 331
- WILSON, G S (1930) *Brit J exp Path*, 11, 157, (1931a) *Ibid*, 12, 89, (1931b) *Ibid*, 12, 162, (1933) *J Hyg, Camb*, 33, 516, (1934) *Ibid*, 34, 361, (1940) *Vet Rec* 52, 737
- WILSON, G S and MILES, A. A (1932) *Brit J exp Path*, 13, 1
- ZDRODOWSKI, P, BREYER, H., and VOSKRESSENSKI, B (1930) *Ann Inst Pasteur*, 45, 769
- ZELLER, H (1933) *Munch tierarztl Wochr*, 84, 337, 349, 361, 373, 389
- ZELLER, H, BEILER, K., and STOCKMAYER, W (1934) *Munch tierarztl Wochr*, 85, 143
- ZELLER, H and STOCKMAYER, W (1933) *Z InfektKr Haustiere*, 44, 67
- ZOBELL, C E and MEYER, K. F (1932) *J infect Dis*, 51, 91, 99, 109, 341, 361

## CHAPTER 35

### BACILLUS

#### AEROBIC SPORE BEARING BACILLI

##### DEFINITION—*Bacillus*

Aerobic spore-bearing rods, usually Gram positive. Often occur in long threads and form rhizoid colonies. Form of rod as a rule not greatly changed at sporulation. Liquefy gelatin. Mostly saprophytes.

Type species. *Bacillus subtilis*

##### INTRODUCTORY

The aerobic spore bearing *Bacillus* forms one division of the family *Bacillaceae* while the anaerobic spore-bearing *Clostridium* forms the other. As many of the organisms in the former division are widely distributed being found in air, soil, water, milk, dust, fish meal, wool, faeces, and other situations, it is not unnatural that they were among the first micro-organisms to be studied, but as there are large numbers of different species, almost all of which are devoid of pathogenic action it follows from the way in which bacteriological investigation has been directed mainly along medical, veterinary, or agricultural lines, that our knowledge of these organisms is far from complete.

Some authors have separated the group into two divisions, the one containing *B. anthracis* and the closely allied pseudoanthrax bacillus the other containing *B. subtilis* and other saprophytic forms. As, however *B. subtilis* may itself be confused with *B. anthracis* this division is of no real value. We shall treat all the members as belonging to one single group—the group *Bacillus*—reserving the term ‘pseudoanthrax bacillus’ as an inexact but convenient designation for any organism of the aerobic spore-bearing group that is liable to be confused with the true *B. anthracis*.

There is a large group of thermophilic bacilli found in milk, manure, and other situations which grow best at temperatures round about 60° C. We do not propose to describe these organisms but references to them will be found in a paper by Wilson and his colleagues (1935). Not all of them belong to the *Bacillus* group, many species appear to be streptococci or non-sporing rods.

##### GROUP CHARACTERISTICS

**Morphology and Staining**—Members of this group are rod shaped organisms varying in size from about  $3\mu \times 0.4\mu$  to  $9\mu \times 2\mu$ . The sides are parallel the axis straight or slightly curved the ends either truncated as in *B. anthracis*, or more usually convex. Their arrangement varies considerably, though single and diplobacillary forms predominate, they may be arranged in chains, often of con-

considerable length, or in groups. Long, unjointed filaments are characteristic of some species, notably of the anthrax bacillus. Irregular forms, consisting mainly of poorly-stained thin bacilli, or of club- or bottle-shaped bacilli, are not uncommon. With a few exceptions, of which *B anthracis* is the most important, all the members are motile by about 4-12 peritrichous flagella. Spores are present in all, and are formed only in the presence of oxygen, they vary in shape from spherical to ellipsoidal, and may appear at the equator, subterminally, or at the very end of the bacillus. In some members their diameter does not exceed that of the bacillus, but in others the rod is swollen to resemble a clostridium. Capsules are met with in only one member, *B anthracis*, and then only when it is growing in the animal body, or in media rich in animal protein. The organisms are usually Gram positive, but considerable variation may be shown, some are strongly positive, others weakly positive, and a few frankly negative. When stained with various dyes, it is generally possible to distinguish areas of uneven staining, in large bacilli a number of small particles are seen, quite distinct from spores. Some of these particles appear to consist of fat (see Burdon *et al* 1942), others of volutin or glycogen. By a few authors the volutin granules have been regarded as nuclear material scattered diffusely through the cell, the evidence against this contention has already been given in Chapter 2. None of the vegetative bacilli is acid fast, though, in the sporing condition they resist decolorization for a short time with weak acids and alcohol.

**Cultural Reactions**—Growth is free on all the ordinary media. Single colonies on agar are generally large, varying from 2 to several millimetres in diameter. Some have a finely granular, mealy appearance, others are membranous and thrown into wrinkles. In broth there is a tendency towards the formation of a surface scum, with or without turbidity, or of a heavy flocculent or membranous deposit. Gelatin is usually liquefied rapidly. In a stroke agar culture the growth is raised and confluent, and generally of membranous consistency, rendering emulsification difficult. Growth is not improved by the addition of blood, serum, or glucose. *Variant colonial types have been described for several members of the group.* Some members form motile colonies (see *B. rotans* p. 855).

**Resistance**—In the vegetative condition the bacilli are killed by moist heat at a temperature of 55° C in 1 hour. The spores vary greatly in resistance, some, like those of *B anthracis*, are destroyed by boiling for about 10 minutes, others, like those of *B subtilis*, may withstand boiling for hours. All are killed by steam under pressure at 120° C in 40 minutes. Similarly with disinfectants the resistance varies, HgCl<sub>2</sub> even in a 1/1000 solution may fail to kill anthrax spores in less than 70 hours (Poppe 1922). Potassium permanganate on the other hand, in a 4 per cent solution kills them in 15 minutes, and a 3 per cent solution of hydrogen peroxide in 1 hour. Generally speaking, the spores are



FIG 176 — *B. megatherium*  
From an agar slope culture ( $\times 1000$ )



extremely resistant to chemical disinfectants, with the exception of those substances which act by oxidation.

**Metabolic and Biochemical Reactions**—Some members form a pigment generally brownish yellow in colour occasionally pink. On the whole, pigment formation is not a striking characteristic and tends to appear late. The optimum temperature for growth varies from 20° C. to 37° C., few grow below 12° C., and excluding the thermophilic bacilli which grow at 60° C., none grows above 55° C. According to Lamanna (1940a) the small-celled species, like *B. subtilis*, *B. vulgaris*, and *B. mesentericus* have an average minimum and maximum temperature about 5° C. higher than the large-celled species like *B. cereus*. As regards oxygen pressure, they are aerobic or facultatively anaerobic. A high partial pressure of CO<sub>2</sub> tends to inhibit growth of *B. subtilis* (Levine 1936) and to favour capsulation but to restrict sporulation of *B. anthracis* (Sterne 1937).

On carbohydrate media the majority of members form acid only, but a few, like *B. asteroides*, *B. polymyxa*, and *B. acetosilicicus* produce gas. Most of the members ferment glucose, maltose and sucrose, some are able to attack mannitol and salicin. Lactose is rarely fermented. A diastatic ferment capable of inverting starch is secreted by some. A proteolytic ferment for gelatin is produced by nearly all and by a few for blood serum. A true rennet like clot is often formed in litmus milk, and is subsequently digested, the litmus is reduced. Some strains peptonize milk without actually clotting it. The reaction becomes alkaline. Both the catalase test and the oxidase reaction described by Gordon and McLeod (1923) are usually positive. Methylene blue is reduced in broth. Some members are able to produce H<sub>2</sub>S, and some to reduce nitrates to nitrites. Indole is not produced. A powerful filtrable haemolysin is formed by one member of the group *B. megaterium*—but many species are said to be haemolytic (Poppe 1922). Highly active bactericidal substances, called tyrocidin and gramicidin (see Chapter 6) appear to be formed by several different members of the group (Dubos 1939, Dubos and Cattaneo 1939, Dubos and Hotchkiss 1941).

**Antigenic Structure**—Most of the serological work has been carried out with a view to separating *B. anthracis* from the other members of the group. A precipitating serum prepared against the anthrax bacillus will react not only with its homologous antigen, but also with the pseudoanthrax bacilli, though in a lower titre, conversely the anthrax bacillus will react in a low titre with a serum prepared against some of the pseudoanthrax bacilli. A similar group reaction is noticeable in the complement-fixation test (Poppe 1922). From this we gather that the aerobic spore-bearing bacilli form a group the members of which are closely related antigenically, the differentiation of *B. anthracis*, at least, can be carried out on a quantitative basis.

An attempt has been made by Sievers and Zetterberg (1940) with some slight success to classify other members of the aerobic spore-bearing group by precipitation and complement-fixation tests. The most important antigenic study, however of recent years is that of Howie and Cruickshank (1940) who, working with *B. cereus*, *B. mesentericus*, and certain other members of the group, were able to show that the vegetative bacilli and the spores contained different antigens thus confirming the previous general conclusions of Defalle (1902) and Mellon and Anderson (1919). Pure anti-spore sera could be obtained by injecting rabbits with a suspension of spores practically free from bacilli or with a suspension of organisms in which the bacillary antigen had been destroyed by autoclaving for

20 minutes at 15 lb., or by absorbing out the anti bacillary agglutinins from a mixed serum. No attempt was made in this work to study the relationship of different members of the group to each other, but in a paper published shortly after Howie and Cruickshank's, Lamanna (1940b) brought evidence to show that *B. subtilis*, *B. vulgaris*, and *B. mesentericus* along with *B. agri*, belonging to the small-celled species, could be separated serologically from each other by means of their spore antigens (see also Lamanna 1942). Separation of members of the large celled species was less successful, but a broad distinction could be drawn between *B. cereus* and *B. megatherium*.

**Pathogenicity.**—Speaking generally, we may say that the anthrax bacillus is highly pathogenic for most animals, and that most other members are non pathogenic for all animals. This statement, however, must be qualified. Under natural conditions the anthrax bacillus gives rise to disease in man, cattle, sheep, and certain other of the domesticated animals, under experimental conditions it is pathogenic for the laboratory animals. Under natural conditions other species of *Bacillus* rarely give rise to disease, but an exception must be made for *B. subtilis*, which may cause severe eye lesions, notably iridocyclitis and panophthalmitis (Axenfeld 1908), and which may occasionally invade the blood stream of patients whose powers of resistance are lowered by the attack of some fatal disease (Sweany and Pinner 1925). Occasionally too, infections such as meningitis and pneumonia may be due to pseudoanthrax bacilli (Senge 1913, Wilamowski 1912). There is evidence that some members, when allowed to grow excessively in food, may produce toxic substances capable of giving rise on ingestion by man to gastro enteritis (see Chapter 72). Under experimental conditions, the pseudo anthrax bacilli are non pathogenic for all laboratory animals except mice, and for these animals only when injected intraperitoneally in a large dose—1–3 loopfuls of an agar culture. *B. megatherium* is, however, definitely toxic, and is able to kill guinea-pigs injected intraperitoneally in less than 24 hours. This is due to the formation of a haemolyxin. The fact that some pseudoanthrax bacilli may on occasion prove pathogenic to man and animals, and that after long subculture *B. anthracis* may lose its virulence for laboratory animals, suggests that there may be a gradual transition from the non pathogenic to the pathogenic state (but see p. 845). A pseudoanthrax bacillus, described as *B. tropicus*, has been isolated from mice inoculated with the blood of patients suffering from "coastal fever" in Queensland, but its relation to the disease is doubtful (see Heaslip 1941).

**Classification.**—This is very difficult, and any classification adopted is bound to be arbitrary. Some authors divide the group on the basis of motility, others on the character and situation of the spore, others on cultural characteristics, and others on several properties taken together. To each of these methods there are objections, and agreement is still far from being reached. In two of the most recent studies, for example, the criteria used for classification differ widely, with necessarily contradictory results. De Soriano (1935) who made a systematic study of 206 strains, proposes a classification based primarily on whether or not the spore causes deformation of the bacillus, and secondarily on morphological, cultural and biochemical characteristics. Lamanna (1940a), on the other hand, who studied 105 strains, suggests a classification based primarily on the size of the bacterial cell, and secondarily on the mode of germination of the spore. Thus he distinguishes first of all between small-celled and large-celled species. The small

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celled species are subdivided into those that germinate (1) by shedding their spore coat equatorially or at the pole or (2) by comma-shaped expansion. The large-celled species all germinate by absorption of the spore coat. Unfortunately this brings many of Lamanna's large-celled species like *B. mycoides*, *B. cereus*, *B. megatherium* and many of his small-celled species like *B. subtilis*, *B. vulgaris* and *B. mesentericus* into the same primary subdivision suggested by de Soriano of species whose spores do not cause bulging of the vegetative portion of the cell. Some hope is held out by the serological studies of Howie and Cruickshank (1940) and Lamanna (1940b) who have shown that the spore antigen is distinct from the bacillary antigen that a classification of the non pathogenic members of the group may ultimately be based on antigenic characteristics.

To the medical and veterinary student the chief interest is in the differentiation of *B. anthracis* from the non pathogenic species this will be discussed under the description of *B. anthracis* which is given below.

### *Bacillus anthracis*.

Named *B. anthracis* by Cohn (1875b) and *Bacteridium* by Davaine (1864). This bacillus is non motile, forms capsules in the animal body and sometimes on artificial media and grows on agar in characteristic long segmented, parallel or interwoven chains. The spores are ellipsoidal or oval in shape are found equatorially and germinate by polar rupture. It is



FIG. 1—*B. anthracis*.

Edge of colony on agar to show curled hair lock appearance ( $\times 15$ ).

It is interesting to note that the anthrax bacillus was the first micro-organism in which the presence of resistant spores was demonstrated. Spores are never found in the animal body during life and in culture appear more slowly than those of the other members of the group. They seem to be formed under conditions unfavourable to continued growth of the vegetative bacilli. Their appearance can be hastened by the addition of distilled water 2 per cent sodium chloride, and other salts (Bongert 1903). According to Bordet and Renaux (1930) sporulation is inhibited by the presence of calcium chloride and favoured by its absence. Cultures grown on oxalated agar often come to consist mainly of spores while those grown on agar to which  $\text{CaCl}_2$  has been added may lose their spore-forming power completely.

The curled hair lock appearance of single colonies on agar or gelatin is characteristic but may be closely simulated by *B. subtilis*. Microscopically this is seen to be due to the growth of the bacilli in long interwoven chains. Growth particularly in broth at a temperature of  $40-50^\circ\text{C}$  for some days causes the appearance of several different variants: some have tough well-defined capsules give rise on agar to the typical curled colonies and are highly virulent; some have soft poorly-defined capsules form thin shining colonies on agar and are slightly virulent; others are non-capsulated give rise to smooth round convex glistening mucoid colonies on agar and are entirely avirulent (Preis 1911; Bongester 1929a, b; Bordet and Renaux 1930). Capsulation is favoured by growth in air containing

10-25 per cent of  $\text{CO}_2$  (Ivánovics 1937) Some variants are asporogenous, it was these which attracted Pasteur's attention and which he considered to be avirulent Preisz (1911) has however shown that though there is a definite correlation between capsule formation and virulence there is none between spore formation and



FIG 178—*B. anthracis*  
Smooth type of colony Agar  
24 hours  $37^\circ\text{C}$  ( $\times 8$ )



FIG 179—*B. anthracis*  
Rough type of colony Agar  
24 hours  $37^\circ\text{C}$  ( $\times 8$ )

virulence Virulent strains may be either sporogenous or asporogenous similarly with avirulent strains Asporogenous varieties may appear spontaneously in cultures incubated at the usual temperature (Behring 1889) or in cultures containing weak antiseptics such as 1/2 000 potassium dichromate or 1/1 000 phenol (Roux 1890) Such varieties when arising from a virulent strain are themselves fully virulent, though prolonged contact with weak antiseptics may eventually lower their virulence The normal highly virulent bacillus forms a large rough colony of frosted glass appearance with a curled edge morphologically it consists of bacilli arranged in chains (Figs 179 181) The avirulent bacillus forms a smaller smoother type of colony with a slightly crenated edge morphologically it consists of bacilli arranged singly in pairs end to end or in small bundles (Figs 178 180)



FIG 180—*B. anthracis*  
From smooth colony on agar 24 hours  
 $37^\circ\text{C}$  ( $\times 1000$ )



FIG 181—*B. anthracis*  
From rough colony on agar 24 hours  
 $37^\circ\text{C}$  ( $\times 1000$ )

In the dry state the spores may remain alive for 12 years or more (Pasteur 1851a). According to Murray (1931) a saline suspension containing 1 000 000 spores per ml. is sterilized by moist heat at 90° C in 15 to 45 minutes at 95° C in 10 to 25 minutes and at 100° C in 5 to 10 minutes.

The antigenic structure of the anthrax bacillus has received considerable attention. As early as 1921 Kramar obtained evidence that the capsule contained a glycoprotein probably belonging to the class of pseudomucins. A soluble specific carbohydrate incapable of giving rise to antibodies on injection into animals but reacting to a high titre with anti-anthrax serum was extracted from anthrax bacilli by Combesco Soru and Stamatesco (1929). A similar substance was prepared by Schockaert (1929). It is however mainly to the work of Tomcsik (1930) Tomcsik and Szongott (1932-1933) Tomcsik and Bodon (1934-1935) Bodon and Tomcsik (1934) Sordelli and Deulofeu (1930-1933) and Sordelli Deulofeu and Ferrari (1939) that our knowledge of the antigenic structure of this organism is due. There appear to be two main antigens: one a protein-like substance present in the capsule, the other a polysaccharide substance present in the body of the organism. Specific precipitins for both the protein-like and polysaccharide constituents are found in serum prepared by the injection of animals with capsulated bacilli and each can be differentially absorbed in the usual way. In animals infected with anthrax both antigens can be demonstrated in the tissues by the use of precipitating sera. Later observations by Ivanovics and Erdos (1934) and Ivanovics and Bruckner (1934a & b 1938) have shown that the capsular substance contains a group-specific hapten common to *B. anthracis* and certain other members of the *Bacillus* group. It appears to consist of a polypeptide of high molecular weight containing  $d(-)$ -glutamic acid. It reacts in high titre with a precipitating serum prepared by the inoculation of rabbits with heat-killed capsulated anthrax bacilli but cannot by itself stimulate the formation of antibodies (Tomcsik and Ivanovics 1938). The somatic polysaccharide is said by Ivanovics (1940) to consist of glucosamine and galactose with acetic acid attached to the molecule.

The anthrax bacillus is naturally *pathogenic* mainly to the herbivora and to man but occasionally it attacks other animals. Experimentally, it proves fatal to the mouse, guinea pig, and rabbit less often to the rat. The larger the dose the shorter is the time to death. Subcutaneous injection of 1 loopful of an 18 hours agar culture kills mice, guinea pigs and rabbits in about 12 to 30 hours; a smaller dose 1/100 of a loopful kills them in 30 to 40 hours; a still smaller dose 1/2 000 000 of a loopful kills mice in 96 hours, guinea pigs in 56 hours and rabbits in 104 hours (Sobernheim 1897). Other members of this group never kill in such small doses.

Post mortem, there is a gelatinous hæmorrhagic local oedema; the viscera are congested, the blood is dark red and coagulates less firmly than usual; the spleen is enlarged, dark red, and very friable. Microscopically the bacilli are found in large numbers in the local lesion, in the blood, and in the thoracic and abdominal viscera; they are confined almost entirely to the interior of the capillaries, where their numbers may be so great as to cause obstruction to the blood flow; the tissues themselves are rarely penetrated. Though the disease terminates in a septicæmia it is not till 4 or 5 hours before death as a rule that the bacilli actually gain access to the blood stream.

Infection may also be successfully achieved by cutaneous, intracutaneous, intramuscular, intraperitoneal or intravenous injection or by feeding. The most certain route is the intramuscular; the least certain the oral (Sobernheim and

Murata 1924) In general it requires a large dose to produce a fatal infection by the mouth (Giovannardi 1931) Post mortem in cases of oral infection, in addition to the enlargement of the spleen and the occurrence of septicaemia, the intestinal mucosa is seen to be covered with small furuncular swellings, through which the bacilli have gained access to the blood Of the three animals the mouse is the most susceptible and the rabbit the least, the guinea pig occupying an intermediate position This difference in susceptibility is scarcely noticeable, except with a strain of weakened virulence (see Chapter 66) Rats are more difficult to infect than other rodents, but are said to succumb easily if fatigued by continuous exercise on a revolving drum (Charrin and Roger 1890) They may develop a chronic disease after subcutaneous injection, which does not prove fatal for 4 or 5 weeks Dogs may be infected by subcutaneous injection, though not uniformly Birds, with the exception of sparrows and young doves and cold blooded animals are resistant, likewise Algerian sheep (Chauveau 1880a, b) (See Davaine 1863a, b, 1864, Koch 1877, Frank and Lubarsch 1892, Sobernheim 1897, Oppermann 1906, Balteano 1922, Poppe 1922, Basset 1925, Katzu 1925, Muller 1925, Sanarelli 1925) The experimental reproduction of the disease in larger animals is considered on p. 1736

Pasteur (1881b, c) found that by growing the anthrax bacillus at 42.5° C for about a month he was able to lower its virulence to such an extent that it proved harmless to all animals except new born guinea pigs. By successive passage through these animals the bacillus gradually regained its virulence till it was able to kill 2, 3 and 4 day and later fully grown guinea pigs, eventually its virulence was entirely restored From the work of Preuz (1911) it would appear that this resumption of virulence is due not to a gradually increasing virulence of the individual organisms, but to an alteration in the proportions of virulent and avirulent bacilli in the culture He found that the effect of incubating a virulent culture at 42.5° C was to cause the appearance of variants that were no longer virulent to animals, so that in one and the same culture both virulent and avirulent bacilli were found side by side The longer the incubation, the higher was the proportion of avirulent bacilli After a month or more the culture consisted almost entirely of avirulent variants, and on injection into mice proved to be harmless, the virulent bacilli that were still present were too few to cause death But if such a culture is injected into a new born guinea pig these few virulent bacilli may be just sufficient to overcome the very low resistance of the animal, in consequence they proliferate and during the course of successive passages increase in proportion relatively to the avirulent variants till eventually the culture consists almost entirely of the virulent type The modern conception of the essential heterogeneity of single strains, i.e. the presence in one and the same strain of organisms showing sharp discontinuous variations in virulence not only has more evidence in its favour than the older conception of the simultaneous and equal raising or lowering of all the bacilli in the strain, but explains more easily the variations in virulence that are noted consequent on altered environmental conditions

Several observers (Hankin 1889, Martin 1890, Marmier 1895, Standfusz and Schnauder 1925) have shown that



FIG 182—*B. anthracis*

In gelatin stab culture 3 days 22° C showing inverted fir tree growth with commencing liquefaction



when *B anthracis* is grown in a broth culture toxic albumoses are formed, which prove fatal on injection into animals, Martin also found a toxic alkaloid. There is no evidence that a true exotoxin is formed, the toxic substances appear to be formed largely by the disintegration of the proteins in the medium. Aoki and Yamamoto (1939) bring evidence to suggest that *B anthracis* forms an endotoxin which is capable of activating spores *in vivo*. The formation of a hæmolysin has been asserted by some authors, and denied by others, it is possible that different strains may vary in this respect. There is certainly no hæmolysin produced for cattle or horse blood, but there is evidence to suggest that one may be formed for sheep, goat and rabbit blood (Poppe 1922).

The main criteria of value in the differentiation of *B anthracis* from those bacilli which may be confused with it may be given in tabular form

<i>B anthracis</i>	<i>Anthrax like or so-called pseudoanthrax bacilli</i>
1 Non motile	Generally motile
2 Capsulated	Non-capsulated
3 Grows in long chains	Grow in short chains
4 No turbidity in broth	Frequent turbidity in broth.
5 <i>Inverted fir tree growth in gelatin</i>	<i>Fir tree growth absent or atypical</i>
6 Polysaccharide precipitin reaction strongly positive	Polysaccharide precipitin reaction weakly positive
7 Pathogenic to laboratory animals	Non pathogenic to laboratory animals
8 Liquefaction of gelatin slow	Liquefaction of gelatin rapid

An inverted fir tree growth in gelatin is given by some strains of pseudoanthrax bacilli, but the branches are thick and interlaced, quite different from the regular, delicate, lateral outgrowths of *B anthracis*. It is sometimes stated that the anthrax bacillus does not show hæmolysis on blood agar plates, whereas the pseudoanthrax bacilli form colonies surrounded by a zone of hæmolysis. This depends on the type of blood used, and is at best an uncertain criterion for differentiation.

When freshly isolated from the animal body, the anthrax bacillus rarely causes difficulty in identification, but after prolonged subculture in the laboratory it may lose several of its important characteristics, such as capsule formation, the inverted fir tree growth in stab gelatin, and its pathogenicity for laboratory animals, and may then be very difficult to classify. Nevertheless bacilli have been described which have given rise to an anthrax like disease in man and other animals, yet which have not conformed to the usual criteria of *B anthracis* (Schulz 1901, Wilamowski 1912, Senge 1913). Such bacilli have usually been classed as pseudoanthrax bacilli, but it is probable that some at least have been variants of the real *B anthracis*, similar to those described by Preisz (1911).

The characters of certain species of *Bacillus* are summarized below

*Bacillus anthracis*  
Carbon, Milzbrandbacillus  
sheep and other animals.

*Synonyms* —Bacterium  
*Habitat*.—Parasitic in  
*Morphology* —Rods,  
agar plates  
woven cloth  
malls mostly  
polar

Straight or slightly curved, ends truncate; occasionally in very long, segmented, parallel, or interlaced not infrequent in cultures. In blood of animals or 4 Spores equatorial, ellipsoidal, not bulging, animal body. Non-motile. Capsule found in

animal body, and on serum media, lost on agar, surrounds entire chain of bacilli Gram positive. Non acid fast

*Agar Plate*—Irregularly round, 2-3 mm in diameter, raised dull opaque, greyish white, plumose colonies, with a tessellated or reticular structure, an uneven surface and a curled edge. Membranous consistency, emulsifiability difficult, colony consists of parallel interlacing chains of bacilli, and is characteristic. After about a week irregular round scales appear on the surface of the colony. Several colonial variants have been described.

*Agar Slope*—Thick, raised, spreading, greyish yellow growth, with an uneven surface and an undulate edge showing little curled projections, moist and slightly glistening. Looks as if there were innumerable tiny air bubbles beneath the surface. After about a week irregular round scales appear on the surface of the growth.

*Gelatin Stab*—Poor filiform growth followed by outgrowth of delicate lateral extensions, longest at the upper part of the culture, giving an inverted fir-tree or lamp brush effect. Liquefaction crateriform, occurs very slowly.

*Broth*—No turbidity, or very fine floccular turbidity, moderate floccular deposit, consisting of interwoven threads, and disintegrating partly on shaking. No surface growth.

*Blood Serum*—Abundant, creamy yellow, confluent, curled growth with uneven surface. No liquefaction.

*Potato*—Raised, dry, greyish white, slightly spreading growth, with undulate or serrated edge.

*Resistance*—Spores killed by boiling in 10 minutes. In dry state remain alive for years.

*Metabolic*—Acrobic, facultative anaerobe. Opt temp 37° C. Limits 12° to 44° C. Pigment none. Haemolysis some strains are stated to haemolyse sheep's red cells. Nutritional, grows well on ordinary media, growth not improved by blood serum or glucose.

*Biochemical*—Acid, no gas, in glucose, maltose, sucrose, and later in salicin, final pH 5.5-5.9. Indole—, MR ±, VP ±, nitrates reduced to nitrites. H<sub>2</sub>S—NH<sub>4</sub>++, methylene blue reduced, catalase +. Litmus milk coagulated and decolorized, later peptonized.

*Antigenic Structure*—There is a capsular polypeptide containing antigen and a somatic polysaccharide-containing antigen, the former has some group relationship to capsular antigens found in certain other members of the *Bacillus* group. Both antigens react specifically with precipitating antisera.

*Pathogenicity*—Naturally pathogenic to man, cattle, sheep (not Algerian), goats, pigs and camels; rarely to carnivores. Experimentally mice, guinea pigs and rabbits, injected sc or sm, die in 12 to 40 hours, p.m., haemorrhagic local exudate enlarged spleen, and bacilli in blood. Rats less susceptible. Birds, except sparrows and young pigeons, cold blooded animals, and fish are resistant.

### "*Bacillus anthracoides*," or "*Bacillus pseudoanthracis*"

Bacilli more or less closely resembling the anthrax bacillus have been isolated by numerous workers from such substances as soil, water, meat, fish and bone meal, wool, dust, oil cake, and less frequently from animals and man. These organisms have frequently been termed *B. pseudoanthracis* or *B. anthracoides*. Reference to the available papers renders it evident that more than one species has been described. We do not propose to consider these organisms in more detail, since their differential diagnosis from the anthrax bacillus has already been dealt with. It is sufficient to point out that most of them are motile, non-capsulated, form spores abundantly within 24 hours on agar, produce an even turbidity or a surface pellicle in broth, give rise to colonies on agar which are less curled and have fewer and less regular outgrowths at the edges, are more

resistant to heat, form alkali in litmus milk, and are generally non pathogenic, though sometimes they may be fatal to mice and even guinea pigs on intraperitoneal inoculation in fairly large doses. The classification of these organisms is at present impossible, they seem to range from avirulent variants of *B anthracis*, on the one hand, to virulent variants of *B subtilis* on the other. For some of the strains which have been described, see Hueppe and Wood (1889) Hartleb and Stutzer (1897) Schulz (1901) Bambridge (1903), Wilamowski (1912) and Grierson (1928), and for two useful reviews see Pokschschewsky (1914) and Poppe (1922)

### *B subtilis*

Great confusion has prevailed, and in fact still prevails, over this organism. It was described by Ehrenberg in 1838 who found it in hay infusion, as *Libro subtilis* and by Cohn (1875a) as *Bacillus subtilis*. Neither of the descriptions was sufficiently full to enable the organism to be distinguished from others that simulate it closely, and in consequence organisms that are almost certainly different from Cohn's original bacillus have been identified with this organism. In an extensive investigation of the *Bacillus* group, Lawrence and Ford (Ford 1916) in America gave this name to an organism that differs in several important particulars from that given by Cohn. The bacillus described by the German workers is fairly large 3-4  $\mu$  long by 1  $\mu$  thick, may form threads, is actively motile, forms anthrax like colonies on agar, gives rise to a thick wrinkled surface membrane in broth, liquefies blood serum, and gives a thick, yellowish white creamy growth on potato, later appearing as if strewn with dry, white granules. The bacillus described by the American bacteriologists is one of the smallest of the aerobic spore-bearing bacilli, is 2  $\mu$  long by 0.4  $\mu$  broad, does not usually form threads, is sluggishly motile, forms dry, hard, glassy colonies on agar adherent to the medium, gives rise to a thin branching scum in broth, later becoming more dense, fails to liquefy blood serum, and on potato gives a luxuriant, warty, pink growth.

According to Conn (1930), there are two different types of bacilli commonly called *B subtilis* one forming small spores which germinate equatorially (Marburg type) the other forming larger spores showing polar germination (Michigan type). Conn advances reasons to prove that the Marburg type is the original and genuine type, while Soule (1932) maintains that the classical *B subtilis* is represented by the Michigan type. At the second international Microbiological Congress the Marburg type was officially accepted as the type strain (St John Brooks and Breed 1937). Unfortunately however, evidence on serological and other grounds has since been adduced by Lamanna (1940b) to show that the Marburg type strain is really a strain not of *B subtilis* but of *B vulgatus* (see also Lamanna 1942, Knaysi and Gonsalus 1944).

In the following summary of the properties of *B subtilis* we have described the small-celled type of organism showing equatorial germination of the spore, which was in the mind of the bacteriological nomenclature committee of the Congress. For some of its properties we have drawn on the description given by de Soriano (1930) Lamanna (1940a, b) and on our own personal observations.

Variant colonies with different bacillary morphology have been described by a number of workers. According to Soule (1928), there is a rough and a smooth type closely simulating the corresponding type of *B anthracis* (see p 843). Graham (1930) who like Soule probably worked mainly with strains of the Michigan type, described four variants two of which were motile and two usually non motile.

Variants I and III formed smooth, circular, shiny colonies with regular margins, II formed "medusa head" colonies, while IV formed slightly irregular colonies with an uneven surface and a rather granular texture. All four variants had the same heat stable somatic antigen. Variants I and II had in addition a common heat labile flagellar antigen.

**Bacillus subtilis** Cohn emendavit Prazmowski

**Synonyms**—Hay bacillus

**Habitat**—Hay, dust, milk, soil, water

**Morphology**—Slender rods  $1.5-3 \mu \times 0.5-0.8 \mu$ , straight or curved, rounded ends, occurring singly or in short chains. Actively motile by 8-12 peritrichate flagella. Non capsulated. Spores are oval  $1.5 \mu \times 0.6 \mu$  formed sub terminally, do not cause bulging of the cell and germinate equatorially without splitting along the transverse axis, appear on agar in 18 hours. Gram positive. Non acid fast.

**Agar Plate**—Irregularly circular colonies, 4-6 mm. in diameter, slightly raised, greyish yellow, and having a darker crumbly centre surrounded by a lighter periphery with a curled edge. Surface is finely granular, membranous or friable consistency, adherent to medium, emulsification rather difficult. Resemble anthrax colonies.

**Agar Slope**—Abundant, confluent, greyish white, raised, opaque, sometimes wrinkled growth, with an undulate and finely serrated edge and a mealy surface. Membranous consistency, adherent to medium emulsifying fairly easily.



FIG 184—*B. subtilis*

In gelatin stab culture 4 days  $20^{\circ}\text{C}$ , showing infundibuliform liquefaction.

**Gelatin Stab**—Filiform growth with rapid infundibuliform or saccate liquefaction, thick white membrane on surface adhering to the sides of the tube.

**Broth**—Moderate turbidity, slight deposit, with formation of a thick wrinkled surface membrane adhering to the walls of the tube, pellicle often sinks to the bottom.

**Blood Serum**—Thick folded membrane, liquefaction.

**Potato**—Thick, yellowish white, raised, dull, creamy growth, later sprinkled with dry white granules giving a mealy appearance.

**Biochemical**—Acid, no gas, in glucose, maltose and sucrose.

Indole — MR —, VP  $\mp$ , nitrates reduced to nitrites  $\text{H}_2\text{S}$  —,  $\text{NH}_3$  +, methylene blue reduced, catalase +. Latmus milk partially clotted, peptonized, and decolorized from above downwards. Starch is hydrolyzed.

**Metabolic**—Aerobe, facultative anaerobe. Opt temp.  $37^{\circ}\text{C}$ . Limits  $15-55^{\circ}\text{C}$ .

**Pigment**—Cream to chestnut brown.

**Nutritional**—Grows freely on ordinary media, growth not improved by blood, serum, or glucose.

**Hemolysis**— $\beta$ -type on horse blood agar plates given by some strains.

**Resistance**—Spores withstand boiling for hours.

**Antigenic Structure**—Little exact knowledge available. Motile strains appear to have a heat labile flagellar and a heat stable somatic antigen. Spore antigen is different from bacillary antigen.

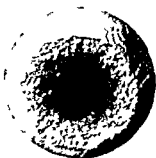


FIG 183—*B. subtilis*  
Smooth type of colony Agar  
24 hours,  $37^{\circ}\text{C}$  ( $\times 8$ )

**Pathogenicity**—May give rise to conjunctivitis, iridochoroiditis, and panophthalmitis in man. Occasionally invades the blood stream in cachectic diseases. 1 ml. 24-hours' broth culture sometimes proves fatal to mice injected intraperitoneally, subcutaneous inoculation into rats occasionally gives rise to a local infiltration, and to submiliary abscesses in the lungs. Most strains are non pathogenic.

### **B mesentericus and B vulgaris**

Flügge (1886) first described an organism, generally known as the potato bacillus, under the name *B mesentericus vulgaris*. There is a tendency now to regard *B mesentericus* and *B vulgaris* as two distinct species though Flinn and Rettger (1934) think that they are variants of a single species. Even the most recent descriptions, such as those of de Soriano (1935) and Lamanna (1940c), are not in entire harmony, and it is difficult in the absence of international agreement to do more than reproduce the characters that have been assigned to these organisms.

A variety, by some regarded as a distinct species, of *B mesentericus* is *B mesentericus fuscus* (Flügge) or the brown potato bacillus. It resembles *B mesentericus*, but differs from it in being slightly smaller, in having less tendency to thread formation and in forming on agar and potato a thinner, greyish brown layer of growth. A red potato bacillus was described by Globig (1888) as *B mesentericus ruber*. It is sometimes known as *B globigii* Migula. On potato it forms a reddish wrinkled growth of a tough viscous consistency. There is also a black potato bacillus, first described as *B mesentericus niger*, and sometimes known as *B aterrimus* Lehman and Neumann, on solid media, especially on potato, it gives a characteristic thick, wrinkled, black growth. A bacillus, *B mesentericus panis viscosi*, sometimes known as *B panis*, is responsible for ropy bread.

### **Bacillus vulgaris (Flügge) Trevisan**

**Synonyms**—*B mesentericus vulgaris*, potato bacillus

**Habitat**—Found in dust, soil water, milk.

**Morphology**—Rather slender about  $2-4 \mu \times 0.75 \mu$ , slightly rounded ends occurring singly and in short chains. Motile by peritrichate flagella. Spores ovoid  $1.5 \mu \times 0.6 \mu$  formed subterminally, do not cause bulging of the cell and germinate equatorially with splitting along the transverse axis. Non capsulated. Gram positive. Non acid fast.



FIG 185—*B vulgaris*  
Smooth type of colony. Agar, 24 hours,  
37°C ( $\times 8$ )



FIG 186—*B vulgaris*  
Same colony as in Fig 185 after 48 hours  
incubation showing marked irregularity  
of surface and edge and raised per-  
ipheral ring ( $\times 8$ )

**Agar Plate**—Round or oval, 4 mm in diameter, raised greyish yellow, opaque colonies with entire edge. Surface is smooth in centre and tends to be wrinkled towards periphery. Consistency membranous or friable, not adherent to medium, emulsification difficult.

**Agar Slope**—Profuse, confluent, raised, greyish yellow, dull opaque, wrinkled growth, with finely granular surface, and smooth or undulate edge, membranous and difficult to emulsify. Membrane on water of condensation.

**Gelatin Stab**—Fibriform growth, rapid liquefaction, infundibulo saecate, with heavy surface membrane and deposit.

**Broth**—Forms thick surface scum, which falls to the bottom, but re-forms, turbidity moderate at first, later clears, with heavy tough membranous deposit.

**Blood Serum**—Moderate, confluent greyish yellow growth with uneven or nodular surface, slight liquefaction.

**Potato**—Thick, creamy yellow, coarsely wrinkled, viscous growth. Surface mealy and yellowish.

**Metabolism**—Aerobe, facultative anaerobe. Opt temp 37° C. Limits 12–55° C.

**Biochemical**—Acid in glucose and sucrose, but not lactose or mannitol. Indole —, MR —, VP ±, nitrates reduced to nitrites.  $H_2S$  —,  $NH_3$  +, methylene blue not reduced, catalase +. Litmus milk decolorized, and peptonized from above downwards. Starch is hydrolysed.

**Pigment**—Slight yellowish or brownish red.

**Hæmolysis**— $\beta$ -hæmolysis on horse blood agar plate by some strains.

**Nutritional**—Grows freely on ordinary media. Growth not improved by blood serum, or glucose.

**Antigenic Structure**—Spore antigen differs from that of *B. subtilis*. According to Sievers (1942) the vegetative bacilli may be sub divided into at least four groups by means of precipitation tests.

**Pathogenicity**—1 ml. 24-hours' broth culture may kill mice injected intraperitoneally. Otherwise non pathogenic.



FIG 187—*B. vulgatus*  
In stab gelatin 4 days 20° C showing napiform liquefaction

### **Bacillus mesentericus (Flügge) Migula**

**Synonyms**—*B. mesentericus vulgatus*, *B. mesentericus fuscus* potato bacillus.

**Habitat**—Found in dust soil water, milk.

**Morphology**—Slender rods 1–3  $\mu \times 0.5$ –0.7  $\mu$ , rounded ends occurring singly or in pairs. Motile. Spores ovoid, 1.15  $\mu \times 0.6 \mu$  formed sub terminally, do not cause bulging of the cell, and germinate by comma shaped expansion (Lamanna 1940). Non capsulated. Gram positive. Non acid fast.

**Agar Plate**—Raised, whitish, irregular colonies of moderate size, with smooth glistening surface and entire or undulate edge.

**Agar Slope**—Abundant, glistening, smooth, whitish growth with undulate edge.

**Gelatin Stab**—Fibriform growth with slow stratiform liquefaction.

**Broth**—Marked turbidity without formation of pellicle and with cottony sediment followed gradually by sedimentation and clearing.

**Potato**—Abundant glistening whitish growth.

**Metabolism**—Aerobe. Opt temp 37° C. Limits 12–55° C.

**Biochemical**—Acid in glucose, maltose, mannitol and sucrose. Indole —, MR —, VP ±, nitrates not reduced to nitrites.  $H_2S$  —, catalase +, Litmus milk decolorized, coagulated, and slowly peptonized. Starch not hydrolysed.

*Pigment*—None

*Antigenic Structure*—Spore antigen is different from that of *B. subtilis* or *B. vulgaris*

### *B. megatherium* de Bary

First described by de Bary in 1884, who found it on cooked cabbage leaves. It is one of the largest members of the *Bacillus* group and occurs in dust, soil, air, milk, and water. Morphological and colonial variants have been described by Knaysi (1933). According to Rettger and Gillespie (1935), the well known morphological pleomorphism of this organism is governed largely by environmental conditions particularly oxygen starvation. It forms a powerful hæmolysin (Todd 1901, 1902; Warden *et al.* 1921), most active towards the red corpuscles of man, monkey, and the guinea pig; it appears in broth cultures at 37° C. on the 2nd or 3rd day, increases to a maximum on the 6th or 7th day, and then diminishes slowly. Oxygen is essential for its production. The hæmolysin, which can be filtered through a Pasteur Chamberland candle, deteriorates rapidly on keeping and like many other true toxins is destroyed by heating at 56° C. for half an hour. Subcutaneous injection into guinea pigs gives rise to a large local swelling with subsequent necrosis. On intravenous injection into guinea pigs it gives rise to hæmoglobinuria but is not fatal except in large doses—about 10 ml. Anti-hæmolysin can be prepared by injection of the hæmolysin into goats. Warden, Connell and Holly (1921) found that when 2 ml. of centrifuged broth culture were injected intraperitoneally into guinea pigs the animals died in less than 12 hours. Post mortem the abdomen was distended, the peritoneum congested, there was hæmolyzed blood in the peritoneal cavity and bloody fluid in the lumen of the gut, in the pleural cavities and over the thighs. They bring evidence to suggest that the toxin and the hæmolysin are one and the same body.

### *Bacillus megatherium* de Bary

*Synonyms*—Probably represents some strains known as *B. anthracoides* or *B. pseudoanthracis*

*Habitat*—Found in dust, soil, water, milk.

*Morphology*—Large, rod-shaped  $3.9 \mu \times 1.2 \mu$ . Long unsegmented forms are common and shadow forms appear early. Ends slightly rounded, axis curved, occurs singly in pairs and in chains. Cells contain fat globules. Motile by 4–8 peritrichate flagella. Spores equatorial, oval or ellipsoidal, not bulging, germination by absorption of spore coat. Non-capsulated. Gram positive. Non-acid fast.

*Agar Plate*.—Round 3–5 mm in diameter, raised, dull greyish white, opaque colonies with entire edge and finely granular surface, sometimes radially striated, may show differentiation into raised opaque centre and thin translucent periphery, membranous consistency, emulsifiability fairly easy. After about a week irregular round scales

appear on the surface of the colony similar to those on anthrax colonies.

*Agar Slope*.—Profuse moist raised glistening greyish yellow creamy growth with smooth surface and entire edge, butyrous consistency, sometimes may show parallel raised ridges like contour lines, emulsifies easily. After about a week irregular round scales appear on the surface of the growth similar to those of anthrax.



FIG 188.—*B. megatherium*  
Surface colony on agar 36  
hours 37° C. ( $\times 8$ )

- Gelatin Slab**—Abundant filiform growth with infundibuliform or saccate liquefaction, no surface membrana.
- Broth**—Moderate, finely floccular, turbidity, with slight ring growth and a powdery deposit, later becoming heavy and viscous
- Blood Serum**—Abundant, moist, creamy, yellowish growth, with granular structure and finely contoured surface. No liquefaction
- Potato**—Thick, greyish yellow, mealy growth.
- Resistance**—Spores are said to withstand 18 lbs steam pressure for 1 hour, killed by 20 lbs for 1 hour
- Metabolism**—Aerobe, facultative anaerobe Opt. temp 35° C Limits 10–45° C
- Pigment**—None
- Hæmolysis**—Powerful hæmolysin produced, acting especially on the red cells of man monkeys, and guinea pigs.
- Toxin**—The hæmolysin is fatal to laboratory animals.
- Nutritional**—Grows well on ordinary media, not improved by blood, serum, or glucose
- Biochemical**—Acid, no gas, in glucose, maltose, and sucrose. Indole—, MR—, VP— Nitrates reduced to nitrites, slight, NH<sub>3</sub>+, H<sub>2</sub>S—, methylene blue reduced, catalase+ Litmus milk sometimes clot, followed by peptonization and decolorization Starch is hydrolysed
- Pathogenicity**—Non pathogenic under natural conditions The hæmolysin is fatal in 1–2 ml. doses to mice and guinea-pigs injected intraperitoneally P.M. hæmorrhagic exudate in peritoneum and pleura

### *B. mycoides* Flugge

First described by Flugge (1886), common in milk, water, soil and dust. Is easily distinguishable from other members by its typical rhizoid growth on agar. It is a highly proteolytic organism, which is said to convert half the protein nitrogen of the medium into ammonia, when growing in the soil it therefore plays an important part in the process of denitrification. Some strains are said to secrete a highly active proteolytic ferment capable of lysing cultures of certain bacteria (Schubert 1928). Variant morphological and colonial types have been described by Lewis (1932, 1933) and den Dooren de Jong (1933).

### *Bacillus mycoides*

- Synonyms**—*B. ramosus* Eisenberg Root bacillus
- Habitat**—Found in milk, water, soil and dust
- Morphology**—Rod shaped 3–5  $\mu$   $\times$  10  $\mu$ , truncated or slightly rounded ends occurring singly, in pairs small groups, and chains, long unjointed threads not uncommon. Motile by peritrichate flagella. Spores are large, equatorial and ellipsoidal measuring 18  $\mu$   $\times$  0.8  $\mu$  not bulging, germinate by absorption of spore coat. Non capsulated. Cells store fat as reserve material. Gram positive. Non acid fast.
- Agar Plate**—Large, spreading raised, greyish white, dull, opaque and rhizoid colonies, with finely granular surface, denser nuclei, dark in colour, are visible from which the peripheral shoots arise, membranous consistency, emulsification fairly easy.
- Agar Slope**—Abundant, confluent, spreading rhizoid, opaque growth, greyish white and slightly glistening, surface honeycombed, due to the presence of arborescent ridges forming a raised network. Growth penetrates the medium and is hence firmly adherent to it.
- Gelatin Slab**—Arborescent, filamentous growth, saccate liquefaction clearing of gelatin with formation of a deposit and a surface membrane.
- Broth**—No turbidity, firm, sometimes wrinkled, surface membrane, depositing later.
- Blood Serum**—Luxuriant, rhizoid growth, no digestion.



*Pigment*—None

*Antigenic Structure*—Spore antigen is different from that of *B. subtilis* or *B. vulgaris*

### *B. megatherium* de Bary

First described by de Bary in 1884, who found it on cooked cabbage leaves. It is one of the largest members of the *Bacillus* group and occurs in dust, soil, air, milk, and water. Morphological and colonial variants have been described by Knaysi (1933). According to Rettger and Gillespie (1935), the well known morphological pleomorphism of this organism is governed largely by environmental conditions, particularly oxygen starvation. It forms a powerful hæmolyxin (Todd 1901, 1902, Warden *et al.* 1921), most active towards the red corpuscles of man, monkey, and the guinea pig; it appears in broth cultures at 37° C. on the 2nd or 3rd day, increases to a maximum on the 6th or 7th day, and then diminishes slowly. Oxygen is essential for its production. The hæmolyxin, which can be filtered through a Pasteur Chamberland candle, deteriorates rapidly on keeping, and like many other true toxins is destroyed by heating at 56° C. for half an hour. Subcutaneous injection into guinea pigs gives rise to a large local swelling with subsequent necrosis. On intravenous injection into guinea pigs it gives rise to hæmoglobinuria, but is not fatal except in large doses—about 10 ml. Anti-hæmolyxin can be prepared by injection of the hæmolyxin into goats. Warden, Connell and Holly (1921) found that when 2 ml. of centrifuged broth culture were injected intraperitoneally into guinea pigs, the animals died in less than 12 hours. Post mortem the abdomen was distended, the peritoneum congested, there was hæmolyzed blood in the peritoneal cavity and bloody fluid in the lumen of the gut, in the pleural cavities and over the thighs. They bring evidence to suggest that the toxin and the hæmolyxin are one and the same body.

### *Bacillus megatherium* de Bary

*Synonyms*—Probably represents some strains known as *B. anthracoides* or *B. pseudo-anthraxis*

*Habitat*—Found in dust, soil, water, milk.



FIG 188—*B. megatherium*

Surface colony on agar 36 hours 37° C. ( $\times 8$ )

*Morphology*—Large rod-shaped  $3.9 \mu \times 1.2 \mu$ . Long unsegmented forms are common and shadow forms appear early. Ends slightly rounded, axis curved, occurs singly, in pairs and in chains. Cells contain fat globules. Motile by 4-6 peritrichate flagella. Spores equatorial, oval, or ellipsoidal, not bulging, germination by absorption of spore coat. Non capsulated. Gram positive. Non acid fast.

*Agar Plate*.—Round, 3-5 mm. in diameter, raised, dull, greyish white opaque colonies with entire edge and finely granular surface. Sometimes radially striated, may show differentiation into raised opaque centre and thin translucent periphery, membranous consistency, emulsifiability fairly easy. After about a week irregular round scales

appear on the surface of the colony, similar to those on anthrax colonies.

*Agar Slope*.—Profuse moist raised glistening greyish yellow, creamy growth with smooth surface and entire edge, butyrous consistency, sometimes may show parallel raised ridges like contour lines, emulsifies easily. After about a week irregular round scales appear on the surface of the growth similar to those of anthrax.

- Gelatin Slab*—Abundant filiform growth with infundibuliform or saccate liquefaction, no surface membrane.
- Broth*—Moderate, finely floccular, turbidity, with slight ring growth and a powdery deposit, later becoming heavy and viscous.
- Blood Serum*—Abundant, moist, creamy, yellowish growth, with granular structure and finely contoured surface. No liquefaction.
- Potato*—Thick, greyish yellow, mealy growth.
- Resistance*—Spores are said to withstand 18 lbs steam pressure for 1 hour, killed by 20 lbs. for 1 hour.
- Metabolism*—Aerobe, facultative anaerobe. Opt temp 35° C. Limits 10–45° C.
- Pigment*—None.
- Hæmolysins*—Powerful hæmolysin produced, acting especially on the red cells of man, monkeys, and guinea pigs.
- Toxin*—The hæmolysin is fatal to laboratory animals.
- Nutritional*—Grows well on ordinary media, not improved by blood, serum, or glucose.
- Biochemical*—Acid, no gas, in glucose, maltose, and sucrose. Indole—, M.R.—, V.P. ±. Nitrates reduced to nitrites, slight.  $\text{NH}_4^+$ ,  $\text{H}_2\text{S}$ —, methylene blue reduced, catalase +. Litmus milk sometimes clot, followed by peptonization and decolorization. Starch is hydrolysed.
- Pathogenicity*—Non pathogenic under natural conditions. The hæmolysin is fatal in 1–2 ml doses to mice and guinea pigs injected intraperitoneally. P.M. hæmorrhagic exudate in peritoneum and pleura.

### B. mycoides Flügge

First described by Flügge (1886), common in milk, water, soil and dust. Is easily distinguishable from other members by its typical rhizoid growth on agar. It is a highly proteolytic organism, which is said to convert half the protein nitrogen of the medium into ammonia, when growing in the soil it therefore plays an important part in the process of denitrification. Some strains are said to secrete a highly active proteolytic ferment capable of lysing cultures of certain bacteria (Schubert 1923). Variant morphological and colonial types have been described by Lewis (1932, 1933) and den Dooren de Jong (1933).

### Bacillus mycoides

- Synonyms*—*B. ramosus* Eisenberg. Root bacillus.
- Habitat*—Found in milk, water, soil and dust.
- Morphology*—Rod-shaped  $3\text{--}5\ \mu \times 1\text{--}0\ \mu$ , truncated or slightly rounded ends occurring singly, in pairs, small groups, and chains, long unjointed threads not uncommon. Motile by peritrichate flagella. Spores are large, equatorial and ellipsoidal, measuring  $18\ \mu \times 0\cdot8\ \mu$ , not bulging, germinate by absorption of spore coat. Non capsulated. Cells store fat as reserve material. Gram positive. Non acid fast.
- Agar Plate*—Large spreading, raised, greyish white, dull, opaque and rhizoid colonies, with finely granular surface, denser nuclei, dark in colour, are visible from which the peripheral shoots arise. membranous consistency, emulsification fairly easy.
- Agar Slope*—Abundant confluent spreading rhizoid, opaque growth, greyish white and slightly glistening, surface honeycombed, due to the presence of arborescent ridges forming a raised network. Growth penetrates the medium and is hence firmly adherent to it.
- Gelatin Slab*—Arborescent, filamentous growth, saccate liquefaction, clearing of gelatin with formation of a deposit and a surface membrane.
- Broth*—No turbidity, firm sometimes wrinkled surface membrane, depositing later.
- Blood Serum*—Luxuriant, rhizoid growth, no digestion.

*Potato*.—Abundant, mealy greyish brown growth of viscous consistency, surface granular

*Persistence*.—Spores are said to withstand 15 lbs. steam pressure for 1 hour Killed by 20 lbs. in half an hour



FIG 189 — *B. mycoides*

Surface growth of rhizoid type on agar 3 days, 30° C. (1/3 natural size)

*Metabolism* —Aerobe facultative anaerobe Opt. temp. 30 C. limits 10–40° C. Pigment none Some strains secrete a lysin capable of dissolving certain bacteria.

*Hemolysis* —None on horse blood agar plates.

*Nutritional*.—Grows fairly well on ordinary media growth not improved by blood, serum, or glucose but augmented by nitrates.



FIG 190 — *B. mycoides*

From an agar slope culture 2 days, 30° C. ( $\times 1000$ ).



FIG 191 — *B. mycoides*

Central part of a surface colony on agar 3 days, 30 C., showing rhizoid structure ( $\times 5$ )

**Biochemical**—Acid, no gas, in glucose, maltose and sucrose Indole —, M.R. —, V.P. —, nitrates +,  $\text{NH}_3$  +,  $\text{H}_2\text{S}$  slight +, methylene blue reduced, catalase +  
 Litmus milk, slow peptonization and decolorization Starch is hydrolysed  
**Pathogenicity**—Non pathogenic to man and animals.

### *B. cereus* Frankland

This organism is one of the large celled species Its exact identity is doubtful Lamanna (1940c) who studied 31 strains found three distinct physiological groups, and concluded that *B. cereus* represents a group of organisms rather than a single species It is generally described as an organism  $3-7 \mu \times 1-1.2 \mu$ , forming ovoid non bulging, sub terminal spores Large, smooth, irregular, very finely granular colonies on agar with a rhizoid periphery Abundant smooth, glistening, whitish growth on agar slope Very marked turbidity in broth with formation of a surface pellicle and a cottony deposit Filiform growth in gelatin with fine ramified offshoots, infundibuliform liquefaction Range of growth  $10^\circ-40^\circ \text{C}$  Forms acid in glucose and maltose, Indole —,  $\text{H}_2\text{S}$  —, V.P. —, nitrates usually reduced to nitrites, litmus milk decolorized coagulated, and slowly peptonized, starch hydrolysis variable The spore antigen seems to differ from that of *B. megatherium*.

### *B. rotans*.

The interest of this organism, which was described by Roberts (1935), lies in the fact that its colonies are motile Two sorts of co ordinated motility are displayed rotation and migration Rotation may occur either clockwise, or anti clockwise and is common in the early stages of colony formation Later the whole colony migrates, pursuing an involved and sometimes spiral course, leaving behind it a few cells to mark its snail like track Unlike *Proteus*, this organism moves freely even on a dry surface Other organisms showing colonial migration have been described by Smith and Clark (1938) Russ Munzer (1938) Shinn (1938), and Turner and Eales (1941) under such names as *B. circulans* and *B. alvei* The organism studied by Turner and Eales formed colonies that migrated at  $37^\circ \text{C}$ . at the rate of 2.5 mm per minute

As well as the more common representatives, there are large numbers of other organisms that have been described, such as *B. polymyxa* Migula, *B. albolactis* Migula, *B. fusiformis* Gottheil, *B. cohaerens* Gottheil, *B. terminalis* Migula, *B. pelastus* Gottheil, *B. tumescens* Zopf, and *B. graveolens* Gottheil For their description the reader is referred to the publications of Loeffler 1887, Globig 1888, Hueppe and Wood 1889, Flügge 1896, Lehmann and Neumann 1896, Hartleb and Stutzer 1897, Chester 1901, Gottheil 1901, Schulz 1901, Bainbridge 1903, Neide 1904, Neufeld 1913, Poppe 1913, 1922, Senge 1913, Pokschischewsky 1914, Ford 1916, Laubach 1916, Bergey 1939, and to the monograph of de Soriano, 1935

### REFERENCES

- AXENFELD, T. (1908) "The Bacteriology of the Eye" London  
 AOKI K. and YAMAMOTO K. (1939) *Z. Immunforsch.*, **95**, 374  
 BAINBRIDGE, F. A. (1903) *J. Path. Bact.*, **8**, 117  
 BALTEANO, L. (1922) *Ann. Inst. Pasteur*, **38**, 805  
 BARY, H. A. DE (1884) "Vergleichende Morphologie und Biologie der Pilze Mycetozoen, und Bakterien" Leipzig  
 BASSET, J. (1925) *C. R. Soc. Biol.* **93**, 1513, 1515, 1517  
 BEHRING (1889) *Z. Hyg. Infekthkr.*, **7**, 171  
 BERGEY, D. H. (1939) "Manual of Determinative Bacteriology" 5th ed. William Wilkins Baltimore.  
 BODON, G. and TOMCSIK, J. (1934) *Proc. Soc. exp. Biol.* **N 1**, **32**, 122  
 BONGERT, J. (1903) *Zbl. Bakt.*, **34**, 497, 623, 772  
 BORDET, J. and RENAUX, E. (1930) *Ann. Inst. Pasteur*, **45**, 1,

- BURDON H. L. STOKES, J. C. and HAMBROUGH, C. E. (1942) *J. Bact.* 43, 717
- CHABRIER A. and ROGER, G. H. (1890) *Arch. Physiol. norm. path.*, 22, 273
- CHAUVEAU, A. (1880a) *C. R. Acad. Sci.*, 81, 33, (1880b) *Ibid.*, 91, 649
- CHRYSTER. (1901) 'A Manual of Determinative Bacteriology' New York.
- COHN F. (1875a) *Cohn's Beitr. Biol. Pflanz.*, 1, Heft 2, p. 175, (1875b) *Ibid.*, 2, Heft 3, p. 141
- COMBESCO D., SOREU, E., and STAMATESCO, S. (1929) *C. R. Soc. Biol.*, 102, 124
- CONN, H. J. (1930) *J. infect. Dis.*, 46, 341
- DAVAINE, C. (1863a) *C. R. Acad. Sci.*, 57, 220, (1863b) *Ibid.*, 57, 301, (1864) *Ibid.*, 59, 333
- DEPAILLE, W. (1902) *Ann. Inst. Pasteur*, 18, 756.
- DOOREY DE JONG, L. E. DE V. (1933) *Arch. Mikrobiol.*, 4, 36
- DUBOS, R. J. (1939) *J. exp. Med.* 70, 1, 11
- DUBOS, R. J. and CATTANEO C. (1939) *J. exp. Med.*, 70, 249
- DUBOS P. J. and HOTCHKISS, R. D. (1941) *J. exp. Med.*, 73, 679
- EHRENBERG (1838) 'Infusionsthierehen als vollkommene Organismen' Leipzig
- FLÖGGE, C. G. F. W. (1886) "Die Mikroorganismen." Leipzig (1896) *Ibid.*, 3te Aufl. Vol. 2 Leipzig
- FLYNN, C. S. and RETTGER, L. F. (1934) *J. Bact.*, 28, 1
- FORD, W. W. (1916) *J. Bact.*, 1, 273
- FRANK G. and LUBARSCH, O. (1892) *Z. Hyg. Infektkr.*, 11, 259
- GIOVANARDI, A. (1931) *Krankheitsforschung*, 9, 13
- GLOBIG (1888) *Z. Hyg. Infektkr.*, 3, 322
- GORDON, J. and McLEOD, J. W. (1928) *J. Path. Bact.*, 31, 185
- GOTTFRED, O. (1901) *Zbl. Bakt., IIte Abt.* 7, 430, 449, 529, 627, 680, 717
- GRAHAM, A. C. (1930) *J. Path. Bact.* 33, 663
- GRIERSON, A. M. M. (1908) *J. Hyg., Camb.*, 27, 300
- HANKIN, E. H. (1889) *Brit. med. J.*, n. 810
- HARTLER, R. and STUTZER, A. (1897) *Zbl. Bakt., IIte Abt.*, 3, 81, 129, 179
- HEASLIP W. G. (1941) *Med. J. Aust.*, n. 536
- HOWIE, J. W. and CRICKSHANK, J. (1940) *J. Path. Bact.*, 50, 233
- HUEFFZ, F. and WOOD, G. C. (1889) *Berl. klin. Woch.*, 26, 347
- IVÁNOVICS, G. (1937) *Zbl. Bakt.*, 133, 449, (1940) *Z. Immunforsch.*, 97, 402
- IVÁNOVICS, G. and BRUCKNER, V. (1937a) *Z. Immunforsch.*, 90, 304, (1937b) *Ibid.*, 91, 175, (1938) *Ibid.*, 93, 119
- IVÁNOVICS, G. and ERDOS, L. (1937) *Z. Immunforsch.*, 90, 5
- KATZ, S. (1925) *Zbl. Bakt.*, 94, 165
- KAYATSI, G. (1933) *J. Bact.*, 26, 623
- KAYATSI, G. and GUNSALES, L. C. (1944) *J. Bact.*, 47, 381
- KOCH, R. (1877) *Cohn's Beitr. Biol. Pflanz.*, 2, 277
- KRAMÁR, E. (1911) *Zbl. Bakt.*, 87, 401
- LAMANN, C. (1940a) *J. Bact.* 39, 593, (1940b) *Ibid.*, 40, 347, (1940c) *J. infect. Dis.*, 67, 193, 205, (1942) *J. Bact.*, 44, 611
- LAUBACH, C. H. (1916) *J. Bact.*, 1, 493
- LEHMANN, K. and NEUMANN, R. (1896) "Atlas und Grundriss der Bakteriologie, etc." J. F. Lehmann, Munich.
- LEVINE P. P. (1936) *J. Bact.*, 31, 151
- LEWIS, L. M. (1937) *J. Bact.*, 24, 381, (1938) *Ibid.*, 25, 359
- LOEFFLER, F. (1887) *Berl. klin. Woch.*, 24, 607, 629
- MARMIER, L. (1895) *Ann. Inst. Pasteur*, 9, 533
- MARTIN, S. (1890) *20th Rep. Loc. Govt. Bd. pub. Hlth., Suppl.*, p. 200
- MELLON R. R. and ANDERSON, L. M. (1919) *J. Immunol.*, 4, 203
- MULLER, L. (1925) *C. R. Soc. Biol.*, 93, 1243
- MURRAY, T. J. (1931) *J. infect. Dis.*, 43, 457
- NEIDE, E. (1904) *Zbl. Bakt., IIte Abt.*, 12, 1, 161, 337, 539
- NEUFELD (1913) *Zbl. Bakt., Fes. Beih.*, 57, 279
- NEUGESTER, W. J. (1909a) *J. infect. Dis.*, 44, 73, (1909b) *Ibid.*, 45, 214
- OPFERMANN (1906) *J. comp. Path.*, 19, 264
- PASTEUR, L. (1881a) *C. R. Acad. Sci.*, 92, 209, (1881b) *Ibid.*, 92, 429, (1881c) *Ibid.*, 92, 668
- POKOSCHIEWSKY, N. (1914) *Arch. Peichgesundh. Amt.*, 47, 541
- POPFZ, K. (1913) *Zbl. Bakt., Ref. Ser.*, (1927) *Ergebn. Hyg. Bakt.*, 5, 597
- PREISZ, H. (1911) *Zbl. Bakt.* 53, 510
- RETTGER, L. F. and GILLESPIE, H. B. (1935) *J. Lact.*, 30, 213
- ROBERTS, J. L. (1935) *J. Bact.* 29, 229
- ROUX E. (1890) *Ann. Inst. Pasteur*, 4, 25
- RUSS MÜTZER, A. (1938) *Zbl. Bakt.*, 142, 175
- ST. JOHN BROOKS, R. and BREED, P. S. (1937) *J. Bact.*, 33, 415.

- SANARELLI, G (1925) *Ann Inst Pasteur*, 39, 209  
 SCHOCKAERT, J (1929) *Arch int. Med exp.*, 5, 165  
 SCHUBERT, J (1928) *Zbl. Bakt.*, 108, 151  
 SCHULZ, R. (1901) *Zbl. Bakt.*, 30, 582  
 SENGE, J (1913) *Zbl. Bakt.*, 70, 353  
 SHINN, L. E. (1938) *J. Bact.*, 36, 419  
 SIEYERS, O (1942) *J. Bact.*, 43, 300  
 SIEYERS, O and ZETTERBERG, B (1940) *J. Bact.*, 40, 45  
 SMITH, N. R. and CLARK, F. E. (1938) *J. Bact.*, 35, 59  
 SOBERNHEIM, G (1897) *Z. Hyg. Infektkr.*, 25, 301  
 SOBERNHEIM, G and MURATA, H (1924) *Z. Hyg. Infektkr.*, 103, 691  
 SORDELLI, A and DEULOFEU, V (1930) *C. R. Soc. Biol.*, 105, 721, (1933) *Folia biol.*, No 26-27, p 121  
 SORDELLI, A, DEULOFEU, V, and FERRARI, J (1932) *Folia biol.*, No 11, p 45, No 20, p 93 and 94  
 SORIANO, M. de (1935) *Rev. Inst. bact., B. Aires*, 6, 507  
 SOULE, M. H. (1928) *J. infect. Dis.*, 42, 93, (1932) *Ibid.*, 51, 191  
 STANDFUSZ, R. and SCHNAUDER, F (1925) *Zbl. Bakt.*, 95, 61  
 STERN, M (1937) *Onderstepoort J. vet. Sci.*, 8, 271, (1939) *Ibid.*, 13, 307  
 SWEANY, H. C. and PINNER, M (1925) *J. infect. Dis.*, 37, 340  
 TODD, C (1901) *Lancet* ii 1663, (1902) *Trans. path. Soc., Lond.*, 53, 196  
 TOMCSIK, J (1930) *Z. Hyg. Infektkr.*, 111, 119  
 TOMCSIK, J and BODOV, G (1934) *Z. Immunforsch.*, 83, 426, (1935) *Ibid.*, 84, 308  
 TOMCSIK, J and IVANOVICS, G (1938) *Z. Immunforsch.*, 93, 196  
 TOMCSIK, J and SZOGOTT, H (1932) *Z. Immunforsch.*, 78, 214, (1933) *Ibid.*, 78, 86  
 TURNER, A. W. and EALE, C. F. (1941) *Aust. J. exp. Biol.*, 19, 161  
 WARDEN, C. C., CONNELL, J. T., and HOLLY, L. E. (1921) *J. Bact.*, 6, 103  
 WILANOWSKI, B. I. (1912) *Zbl. Bakt.*, 66, 39  
 WILSON, G. S., TWIDG, R. S., WRIGHT, R. C., HENDEY, C. B., COWELL, M. P., and MAIER, I (1935) *Spec. Rep. Ser. med. Res. Coun., Lond.*, No 206, p 376

## CHAPTER 36

### CLOSTRIDIUM

#### THE SPORE BEARING ANAEROBES

##### DEFINITION—*Clostridium*

Anaerobic or microaerophilic rods producing endospores, which are usually wider than the vegetative organisms in which they arise—so-called clostridium forms. Generally Gram positive. Often decompose protein media and often ferment carbohydrates. Many species form exotoxins and many are pathogenic. Type species is *Clostridium butyricum* Prazmowski.

Before the war of 1914-18, the study of the spore bearing anaerobes had been undertaken fitfully and by imperfect methods, much attention had been paid to their pathogenicity, but little to their general biological characters. One and the same organism had received many different names, and many organisms with the same name undoubtedly belonged to different species. The only two organisms about which no doubt existed were the two that formed a highly potent toxin, recognizable by the specific effects they produced on injection into animals—namely *Cl tetani* and *Cl botulinum*. It was not till the exigencies of war rendered an intensive study of the anaerobes necessary, and till the introduction of McIntosh and Fildes jar made it feasible to obtain pure cultures with relative ease, that the obscurity surrounding this group was dispersed.

Most of the older workers had failed to realize the difficulty inherent in obtaining pure cultures of the anaerobic bacilli. The new technique, especially by enabling plate cultures to be made, revealed at once the impurity of many of the classical strains and provided a means for the preparation of single-colony cultures. For the first time a distinctive account was provided of the main species, which made possible their identification, and which disposed of many spurious characters that had been attributed to them. Incidentally fresh species were discovered (For references on the production of anaerobiosis see Liborius 1886, Frankland 1889, Smith 1890, Tarozzi 1900, Smith *et al* 1900-06, Laidlaw 1915, McIntosh and Fildes 1916, Report 1917, Holker 1918-19, Rockwell 1924, Varney 1926, Wilson 1923, Hall 1929a, Dickens 1934, McClung *et al* 1935).

**Habitat**—The anaerobes are widely distributed in nature, but their main habitat is undoubtedly the soil. Some of them appear to be common inhabitants of the intestinal canal of man and animals. *Cl welchii*, for example, is uniformly present in human faeces, *Cl tetani* has been found in about 10-40 per cent. of faecal specimens of domestic animals, *Cl sporogenes* is frequently, and *Cl histolyticum* occasionally present. It has been held by some that the intestinal canal is the main habitat of certain of the anaerobes, particularly *Cl tetani*, and that their presence in the soil can be explained by faecal contamination. The fact that this

organism is found in virgin soil taken far from human or animal habitations renders this view improbable. It would seem more likely that the primary habitat of the majority of the anaerobes is the soil, that they are ingested frequently with vegetables and fruit, and that some of them are able to adapt themselves temporarily or permanently to a life in the intestinal canal.

Their presence in soil and faeces accounts for their frequent appearance in dust, milk and sewage. In spite of the fact that they lead a saprophytic existence several of these species are causally related to well recognized diseases in man and animals.

**Morphology**—The anaerobes are endowed with a pleomorphism that renders their identification on a morphological basis very difficult and often frankly impossible. Not only may an organism assume different shapes under changing environmental conditions but under one and the same set of conditions it may present very different forms.

Like the aerobic spore bearing bacilli they are large rod shaped organisms. In length they vary from about  $3\ \mu$  to  $7$  or  $8\ \mu$  but long filamentous forms are quite common. Their breadth varies from about  $0.4$  to  $1.2\ \mu$ . The vegetative bacilli are straight or curved, their sides are parallel and their ends rounded or somewhat truncated. Most are arranged singly but some occur in pairs or in chains, others in bundles the members of which are arranged parallel to each other. Irregular forms include navicular or boat shaped organisms, citron forms shaped like a lemon with a small knob at each end, large swollen non sporing rods or orgonts, snake like filaments, deeply stained bulb like types, and a great variety of so-called involution forms varying both in shape and in depth of staining. Autolysis frequently sets in with the commencement of sporulation so that shadow forms are numerous particularly in certain species.

Sporulation is common to all members but there is considerable variation in the readiness with which it occurs. *Cl. sporogenes*, for example, spores readily on all media, *Cl. welchii* only in media free from a fermentable carbohydrate and then inconstantly. All the pathogenic members are able to form spores in the animal body though *Cl. welchii* does so rarely.

It has been customary to classify the anaerobes according to the shape of the spore and the position in the rod at which it appears. Thus we have (1) those with an equatorial or subterminal spore, (2) those with an oval terminal spore, and (3) those with a spherical terminal spore. This division is useful for certain purposes but it must not be used too rigidly. It is common, for instance, to find organisms that usually form subterminal spores giving rise to spores that are strictly terminal. The distinction between a spherical and an oval terminal spore may also be a matter of the utmost nicety.

The spores of most members are wider than the vegetative bacilli, they therefore confer on the organism a distinctive appearance according to the position in which they arise. If they are formed at the equator the clostridium is spindle shaped, if subterminally club-shaped. With an oval terminal spore the organism may look like a tennis racket, with a spherical terminal spore like a drum stick.

With the exception of *Cl. welchii* all the members are motile by peritrichate flagella. Motility however is often difficult to demonstrate especially in artificial cultures and in strains that have been subcultured for some time. Young cultures in broth or cooked meat medium not more than 6 to 24 hours old are the most suitable for examination. If these are negative the organisms should be examined



in the tissue fluids of injected animals. The usual coverslip method is satisfactory in most instances, but if this fails, examination should be conducted in a closed capillary tube that has been inoculated with a young broth culture and kept at 37° C for about half an hour. As is customary with large organisms, motility is rarely well marked, and is usually of the slow and stately variety, in contrast to the rapid, darting movements of smaller organisms such as *Salmon typhi*.

*Cl. butyricum* and *Cl. welchii* are the only members possessing a capsule; the capsule of *Cl. welchii* is noticeable in the animal body, and sometimes in cultures containing serum.

**Staining Reactions.**—All members stain readily with the usual dyes. Great irregularity is noticeable in the depth of staining, especially in cultures more than a day or two old. Sometimes metachromatic granules are noticeable, or points of more intense coloration. Provided young cultures are examined, the bacilli are all Gram positive. Some species rapidly lose this property, and some can be decolorized if the alcohol is applied for too long. In the early stages of spore formation, the position of the spore is often marked by an area of intense staining, as it matures, however, the spore presents a colourless centre surrounded by a peripherally stained ring.

**Cultural Reactions**—On solid media growth is relatively slow, and takes the form of a thin, effuse, often spreading film, which may be difficult to distinguish from the underlying medium.

The tendency to film formation is promoted by moisture. On first isolation *Cl. septicum*, and particularly *Cl. tetani*, tend to spread rapidly over a moist surface. If *Cl. tetani* is inoculated into the condensation water of an agar slope, it will in the course of a day spread over the whole medium, the film is so thin that, were it not for the dentate edge presented at the upper end of the slope, where the medium is drier, it might easily escape detection. Advantage may be taken of this fact in the isolation of this organism (Fildes 1925a). The spreading of clostridia can be inhibited by the incorporation of certain chemicals in the solid media. The majority of these, however, are to some extent bacteriostatic, inhibition of spreading without bacteriostasis, as Hayward (Miles and Hayward 1943) has shown, may be achieved by increasing the concentration of agar up to about 6 per cent. Certain clostridia also produce on agar motile daughter-colonies, which rotate and wander over the surface of the medium (Turner and Eales 1941). Concentrated agar is less effective as an inhibitor of this type of spreading.

**AGAR PLATES**—Single colonies are rounded, generally effuse, and present crenated, fimbriate, or rhizoid edges. *Cl. welchii*, which is one of the less strict anaerobes, forms low convex colonies with an entire edge, *Cl. sporogenes* and *Cl. histolyticum* may form umbonate colonies with a raised centre and a flat periphery. The colonial appearances are often characteristic, but some species give rise to variants which not only are unlike the typical colony, but which strongly suggest the occurrence of contamination. Several different types of colony may be formed, for example, by *Cl. sporogenes*.

**GLUCOSE AGAR SHAKE CULTURES**—These are commonly employed for studying the form of deep colonies, and by many workers for the preparation of pure cultures. Except near the surface, growth occurs throughout the medium, this is frequently disrupted and blown upwards by the development of gas. Single colonies are rounded or lenticular in shape, and lenticular forms may later develop irregular tufts of growth, sprouting from the edge or the poles of the lenses, sometimes

they are differentiated into an opaque centre and a translucent periphery their edge may be entire but is more often woolly or presents that curious reticular filamentous appearance of a cigarette thrown into water. There is a general correspondence between the form of surface and deep colonies. Thus round entire edged and raised surface colonies usually correspond to opaque and lenticular colonies in deep agar irregular or coarsely rhizoidal to opaque and lumpy, delicately rhizoidal to fluffy, and spreading colonies to deep colonies like a snow flake. Again, surface colonies with central papillæ usually correspond to deep colonies with a marked central opacity.

**BLOOD AGAR PLATES**—On these not only is the colonial form characteristic, but the degree and type of hæmolysis afford a useful differentiating feature between the members of the group. Hæmolysis is well marked after 3-days incubation at 37° C, if the plates are then stored in a dark cupboard at room temperature it often continues to increase. With a thick seeding the whole plate may be completely decolorized.

Many organisms give hæmolysis of the  $\alpha$  prime type after 3 days incubation (see Chapter 24), after a further 3 days this passes into the fully developed  $\beta$  variety. In some cases it is possible to specify the hæmolytic factors concerned. For instance the relatively wide zone of hæmolysis produced by toxigenic strains of *Cl. welchii* on routine horse blood agar is usually due to the  $\theta$  toxin (p. 866) if the action of  $\theta$  toxin is suppressed by  $\theta$  antitoxin a narrower ill defined zone of partial hæmolysis is revealed due to  $\alpha$  toxin. The action of the  $\alpha$  lysin is greatly enhanced by calcium ions. Some strains of *Cl. welchii* produce only the  $\alpha$  type of hæmolysis (Evans 1915).

**COOKED MEAT MEDIUM**—Most of the members grow well in this medium. All render the fluid turbid to some extent and most produce gas. The proteolytic members turn the meat black and may obviously digest it the saccharolytic members do not digest the meat and frequently turn it pink. Varying reactions are recorded in this medium, depending on the strain used the batch of medium and the length of incubation. Both in this medium and in other media the proteolytic members form characteristic foul and pervasive odours while in cultures of the saccharolytic members there is no odour or if there is it is not foul.

**COAGULATED SERUM AND COAGULATED EGG**—These media are used for testing the proteolytic powers. None of the saccharolytic organisms is able to liquefy them.

**GELATIN**—At 23° C most members grow poorly. In stab culture *Cl. tetani* gives a characteristic fir tree growth followed later by liquefaction. At 37° C growth is improved and is generally accompanied by permanent liquefaction.

**Resistance**—In the sporing stage all the members present a marked but variable resistance to heat drying and disinfectants. Thus the spores of *Cl. botulinum* withstand boiling for 3 or 4 hours and even at 105° C are not killed completely in less than 100 minutes. *Cl. adamsiensis* is a little less resistant than *Cl. botulinum* (Hoyt Chaney and Cavell 1938). On the other hand spores of *Cl. welchii* are said to be destroyed by boiling in less than 5 minutes (Headlee 1931). *Cl. sporogenes* can survive exposure for 8 days to a 5 per cent phenol solution. In dried earth or dust *Cl. tetani* may live for years. Stock cultures of most members in cooked meat medium remain viable for months some such as *Cl. fallax* and *Cl. cochlearium* are more delicate and require transferring frequently.

**Metabolism.**—Up till within recent years it was generally believed that members of this group were unable to grow except when oxygen was rigidly excluded from the medium. Though free oxygen does inhibit their growth and may actually destroy organisms in the non sporing state it is quite possible to obtain growth of anaerobic bacteria in the presence of air provided a sufficiently low oxidation reduction potential is established in the medium. This can be done by including substances in the medium which will take up molecular oxygen and bring about a fall in the Eh below that necessary for the initiation of growth. Many such substances are available some of which act mainly by absorbing oxygen others of which are chiefly responsible for the establishment of a low Eh after the molecular oxygen has been nearly used up or removed by mechanical means. Sulphites reduced iron compounds unsaturated fatty acids activated glucose, cysteine glutathione ascorbic acid, thioglycolic acid, and metallic iron are examples of some of the substances commonly added to media to bring about the requisite anaerobic conditions. Cooked meat is an example of a medium that affords excellent conditions for anaerobic growth even when incubated aerobically. Its virtue lies in its containing (1) unsaturated fatty acids which take up oxygen, the reaction being catalysed by the haematin of the muscle and (2) glutathione which brings about a negative O R potential corresponding to an Eh of about  $-0.2$  volt (Lepper and Martin 1929 1930). Fildes (1929) has shown that for the germination of tetanus spores an Eh in the medium approximating to  $+0.01$  volt at pH 7.0 is required, this corresponds to the zone of complete reduction of thionin. It is probable that similar conditions determine the growth of most other anaerobes. *Cl. histolyticum* *Cl. tertium* and *Cl. carnis* however are exceptions. These organisms are microaerophilic rather than anaerobic and can grow to a limited extent aerobically though they are said to be incapable of forming spores under these conditions (Hall and Duffett 1933). Once growth has started most anaerobic organisms appear to bring about a rapid fall in the O R potential of the medium, probably owing to the production of a more active reducing system than that present in the medium itself. The Eh frequently falls to below  $-0.4$  volt. According to Gillespie and Rettger (1933) the final Eh reached by the various clostridia in a given medium may be useful in species characterization. As has just been pointed out in the presence of powerful reducing systems growth may continue even though considerable quantities of oxygen are gaining access to the medium.

We have discussed the exact nutritive requirements of certain clostridia and the problems of anaerobiosis at some length in Chapter 3. The earlier work of Fildes and his colleagues (see Fildes 1933 Fildes and Knight 1933 Knight and Fildes 1933 Fildes and Richardson 1933 Pappenheimer 1933 Knight 1933) and of Stickland (1934 1935) on essential nutrients of clostridia and the modes of their utilization has been developed to the point where it is clear that the majority of pathogenic clostridia are heterotrophs, requiring a battery of amino-acids, carbohydrates and vitamins for growth in artificial media. Moreover, the energy producing mechanisms especially of those clostridia that depend mainly on amino-acid breakdown for their energy have been studied in some detail (see for example Gale 1940 Woods and Trim 1942 Clifton 1942 Guggenheim 1944). A small concentration of  $\text{CO}_2$  seems to be as essential for the growth of the anaerobic as it is for so many of the aerobic bacteria (Gladstone Fildes and Richardson 1935). In addition, the growth of some clostridia is greatly improved by a con-

centration of  $\text{CO}_2$  of the order of 2-10 per cent (Rockwell 1924 Dack *et al* 1927 Atken *et al* 1936)

On ordinary media growth of the anaerobes is poor compared with that of the aerobic spore bearers. Some strains grow better than others—*Cl welchii*, *Cl bifermentans*, *Cl botulinum*; some give poorer growths—*Cl chauvæi*, *Cl cochlearium*.

Glucose favours the saccharolytic species. blood or serum improves the growth of all. The optimum  $\text{H}^+$  ion concentration for growth is about pH 7.0 to 7.4 (Reddish and Rettger 1924)

On media containing bile salts such as MacConkey's medium growth of *Cl sporogenes*, *Cl botulinum*, *Cl histolyticum*, *Cl welchii*, *Cl tetani* and *Cl septicum* is accompanied by a greenish fluorescence. In our experience *Cl chauvæi* and *Cl adematensis* have failed to grow on this medium.

Most of the members with which we are dealing here grow best at about 37° C though many of them are capable of growing at temperatures of 20° C and even lower. There is a group of thermophilic clostridia which have an optimum temperature about 50°-60° C and which sometimes do not grow at all below 30° C.

**HÆMOLYSIN PRODUCTION**—Apart from their action on blood agar plates many of the anaerobes such as *Cl tetani*, *Cl welchii*, *Cl septicum*, *Cl adematensis* and *Cl chauvæi* produce filtrable hæmolysins capable of dissolving sheep's red blood corpuscles. Kerrin (1930) states that atoxic strains of *Cl tetani* produce as powerful a hæmolysin as do toxic strains and that normal rabbit horse and human serum have a very strong antihæmolytic effect. For the detection of hæmolysins care must be taken to buffer the hæmolytic systems at the pH of optimum activity (Walburn 1938). Fibrinolysins are formed by some species (Carlen 1939 Reed Orr and Brown 1943) and leucocidins by others.

**Biochemical Reactions**—The action on sugars is of some value in differentiating the anaerobes and constitutes one basis of classification. Great care must be exercised in carrying out the tests since even with known stock strains the results are often irregular and must be repeated two or three times before they can be relied on. Some clostridia decolorize indicators irreversibly so that the formation of acid in a fermentation tube should always be tested by the addition of fresh indicator to a sample of the culture.

Reed (1942) points out that both indole formation from tryptophan and the reduction of nitrates to nitrites depend on the relative rates of breakdown of the original substrates and of substances formed from them. Thus most clostridia reduce both nitrates and nitrites and only if the reduction of nitrates is the quicker of the two processes will a positive test for nitrites be obtained.

One of the striking features of the anaerobic bacteria is the large amount of gas that they are able to produce even in media free from fermentable carbohydrates. Thus Wolf and Harris (1917) found that *Cl welchii* in casein water produced 90 ml of gas per litre of medium and in peptone water 186 ml. *Cl sporogenes* formed 1.044 ml of gas per litre of casein water in 157 hours and in peptone water 360 ml. in 24 hours. The gas consists of  $\text{CO}_2$  and  $\text{H}_2$  in different proportions according to the species of anaerobe. The addition of a fermentable carbohydrate to the medium increases the gas production. Acids are formed as the result of the fermentation with *Cl welchii* rather more than 50 per cent are volatile—mostly butyric acid. Ammonia appears to be formed in large quantities by the proteolytic, and in much smaller quantities by the saccharolytic anaerobes.

An attempt has been made (Anderson 1924) to classify the anaerobes on the

basis of their gaseous metabolism. Growth in plain peptone water results in the production of  $\text{CO}_2$  and  $\text{H}_2$  in different proportions, the  $\text{CO}_2/\text{H}_2$  ratio is said to be high with the proteolytic and low with the saccharolytic members. Thus for *Cl. histolyticum* it is over 91, for *Cl. sporogenes* 36.9, for *Cl. botulinum* 18.3, for *Cl. tetani* 1.17, for *Cl. septicum* 0.98, and for *Cl. welchii* 0.4.

Litmus milk is a useful medium for differentiation (see Wolf and Harris 1917, Weinberg and Seguin 1918, Wolf 1918-19, 1919-20, Report 1919, Anderson 1924, Wagner *et al.* 1924). Spray (1936) introduced iron litmus milk and iron-gelatin as differential media, in which reactions with the iron, notably blackening of the medium, provide several useful distinctive characters among clostridia (see Table 57).

It has been stated that none of the anaerobes is able to form catalase (Adamson 1919-20). This statement probably needs modification, we have obtained evidence of its production by *Cl. sporogenes* and *Cl. histolyticum*, though only in small amounts.

**Antigenic Structure.**—Antisera have been prepared against a number of species, and agglutination and complement fixation reactions have been carried out. Difficulty has often been experienced in the preparation of stable suspensions, there is a great tendency for auto-agglutination to occur. The work of Felix and Robertson (1923) showed that the motile species of anaerobes contained thermolabile H and thermostable O antigens, similar to those described for so many of the aerobic bacteria. It was thought that type specificity, as determined by agglutination, was dependent on the H antigen, and group specificity on the O antigen.

The more recent work of Henderson and others, however, seems to show that the position is rather more complex. With *Cl. septicum*, for example, Henderson (1934) finds that the most convenient subdivision is made on the basis of the O antigen. Though there is considerable overlapping in different strains, there appear to be four specific O receptors. These four primary groups can be further subdivided according to the type of the H antigen. Henderson (1932) states that there is an O antigen common to the ovine and the bovine strains of *Cl. chauvoei*, but that the H antigen is complex, differing to some extent according to the animal source and the country of origin of the strain (see also McEwen 1926, Roberts 1931). The relation between *Cl. septicum* and *Cl. chauvoei* is not very clear, but the work of Weinberg, Davesne, Mihailesco and Sanchez (1929) and Kreuzer (1939) suggests that the two organisms are closely related antigenically. *Cl. tetani* is divisible into at least 10 types, all of which possess a common O antigen and a second O antigen is present in Types I, III, VI, VII, VIII and X. Type specificity depends on a flagellar antigen. *Cl. tetanomorphum* has a minor somatic antigen in common with *Cl. tetani* (Wilson 1931, Gunnison 1937, MacLennan 1939). Attempts to form serological groups of *Cl. welchii* have met with varying success. Test antisera usually react fully with the homologous strain, and only with a few heterologous strains (Henriksen 1937, Duffett 1938, Orr and Reed 1940). There is some evidence that the four toxigenic types (see below) differ in their bacterial antigens (Kreuzer 1939) but each type is antigenically heterogeneous. Thus Henderson (1940) distinguished two kinds of somatic antigen, a heat-stable O, and a heat labile (L), antigen. He found strain specific O antigens in Type A, but no L antigen, 13 strains of Type B fell into two O-antigenic groups, and into 7 L-antigenic groups, all his Type C strains had a common O antigen but no L antigen, and there were various O and L antigenic groups in Type D. Rodwell (1941) found a similar variety of O antigens in the four types. In *Cl. welchii* the specificity of agglutination among S forms appears to depend on the nature of the capsular substance, which contains polysaccharides. On the whole, the serological reactions of antigenic extracts of capsular substance are as heterogeneous as the agglutination reactions of the bacilli.

(Weisel 1938 Orr and Reed 1940 Svec and McCoy 1944) The  $S \rightarrow R$  variation in *Cl welchii* is accompanied by a loss of specific O antigen (Henderson 1940)

*Cl botulinum* is divisible into seven groups according to the flagellar antigens and the proteolytic strains appear to possess a common O antigen (Schoenholz and Meyer 1925 McClung 1937)

Among the proteolytic species we may note that *Cl sporogenes* can be divided into at least two groups. Serological tests have been useful in resolving some problems of identity of various species. Thus it appears from the work of Clark and Hall (1937) and Stewart (1938) that *Cl sordellii* may be considered as a variety of *Cl bisfermentans* (For other examples see the review of McCoy and McClung 1938). One of the more interesting results of serological study is the confirmation of a cultural and biochemical relationship between R variants of *Cl histolyticum* and *Cl sporogenes* (Smith 1937 Hoogerbeide 1937), its significance is not clear.

**Toxin Formation**—It is remarkable that with the exception of the diphtheria bacillus, the organisms forming powerful exotoxins belong almost entirely to the group of anaerobic spore bearing bacilli. Two of them—*Cl botulinum* and *Cl tetani*—give rise to toxins more poisonous than any other substances with which we are acquainted. It has been calculated that the most powerful toxin of *Cl tetani* would kill a man in a dose of 0.25 mgm, and of *Cl botulinum* in a dose of 0.0084 mgm.

The formation of a powerful exotoxin does not appear to be associated with the proteolytic activity of the organism. Soluble diffusible toxins have been described in only two proteolytic species, *Cl bisfermentans* and *Cl histolyticum*. Whether the toxins are formed intra- or extra-cellularly is still unknown. Stark, Sherman, and Stark (1923) have found that if bacteria free filtrates of *Cl botulinum* are added to sterilized skim milk in suitable proportions and incubated for 4 days at 37° C, a considerable increase in toxicity occurs, suggesting that enzymes present in the filtrate have formed fresh toxin from some constituent of the milk. A number of clostridial toxins resemble the diphtheria toxin in that they can be detoxified by formaldehyde with the formation of an antigenic toxoid that can be used for active immunization (see Chapters 77, 78). The preparation of these toxins requires attention to a number of factors with which we have no space to deal. But their properties are important, and must be considered briefly. It should be emphasized that many of the properties described are those of toxic culture filtrates, not of isolated substances. Filtrates from cultures of *Cl welchii* for example have been resolved into a number of components, and it is probable that other "toxins" hitherto referred to for convenience as single substances will prove to be mixtures.

**Tetanus Toxin**—This varies in potency, a good filtrate will kill a mouse in a dose of 0.00001 ml. It is destroyed by heat at 85° C for 5 minutes, but if dried it will resist 120° C for 1 hour. Exposure to 55° C for 1 hour is said to destroy the greater part of its toxicity, while having little effect on its antitoxin combining power (Tschertkow 1929). It is destroyed by direct sunlight in about 15 hours at 40° C, exposure to diffuse daylight results in a gradual weakening of the toxin. If precipitated with ammonium sulphate, dried over sulphuric acid, ground to powder, and preserved in the dark at 5° C in vacuum tubes under phosphorus pentoxide the toxin will remain unchanged for 2 years or more. 0.55 per cent HCl, 0.3 per cent NaOH and 70 per cent alcohol each destroy the toxin in 1 hour. The toxicity can be modified by iodine trichloride, by formol and (Velluz 1936) by carbon disulphide, these reagents are used in serum institutes for weakening

the toxin prior to injection of animals. Tetanus toxin is not absorbed from the intact alimentary canal; there is evidence that it is destroyed by the digestive juices. It combines with and is neutralized by specific antitoxin. There is evidence, based on a lack of parallelism in the toxicity of culture filtrates of *Cl. tetani* for different species of laboratory animals, that there is more than one component in tetanus toxin. These hypothetical components, however, appear to have a similar antigenic specificity (Ipsen 1940-41, Smith, M. L., 1942-43). Petrie (1942-43) found corresponding variations in the activity of antitoxins, and concluded that crude tetanus toxin contained varying proportions of a "primary" toxin molecule, and an antigenic variant of it. From somewhat similar experiences Friedemann and Hollander (1943) postulated qualitative differences between tetanus toxins from various sources.

The toxin of *Cl. botulinum* resembles tetanus toxin in many respects, but is more resistant to heat and to acids. Thus it requires for its destruction a temperature of 80° C. for half an hour. Normal hydrochloric acid fails to destroy it even in 24 hours, but normal soda destroys it rapidly. It is non-dialysable. The potency of the toxin varies; it has been possible to obtain filtrates with a M.L.D. for a guinea-pig of 0·000001 ml., but this is exceptionally strong. It is often said to be the only exotoxin that can be absorbed from the alimentary canal, but the recent work on enterotoxaemic diseases of sheep (see Chapter 78) suggests that the  $\epsilon$ -toxin of *Cl. welchii* shares this property.

The general properties of the toxins of *Cl. welchii*, *Cl. septicum*, and *Cl. oedematiens* may be considered together. They are all moderately thermolabile, being destroyed by heating to 70° C. for 30 to 60 minutes. They are likewise destroyed by weak concentrations of acids. When toxic filtrates are injected into guinea-pigs or mice they give rise to a gelatinous oedema and a varying amount of necrosis. Weinberg and Combiesco (1930) state that *welchii* toxin lyses the red blood corpuscles, producing hæmoglobinuria, causes focal areas of necrosis in the kidney and liver, and leads to an increase in blood pressure, which may in its turn be responsible for hæmorrhages in various parts of the body.

Before discussing the toxins of *Cl. welchii* in particular, we must refer briefly to the four main varieties of the species, each associated with a different disease (see Chapter 78); these are the classical *Cl. welchii* of human gas gangrene, "*Cl. agni*," causing lamb dysentery, "*Cl. paludis*," causing the sheep-disease "Struck," and "*Cl. oritoxicum*," causing an infectious enterotoxaemia in sheep. The claim of the last three to specific status has not yet been established. Each variety, however, was shown by Wilsdon (1931, 1933) to produce toxic filtrates that could be distinguished by their content of a number of toxic components.

Glenny and his colleagues (1933) identified five separate toxic components in culture-filtrates of Wilsdon's types,  $\alpha$ ,  $\beta$ ,  $\gamma$ ,  $\delta$ , and  $\epsilon$ . The existence of these components was confirmed by a number of workers (Bosworth and Glover 1935, Borthwick 1935, Mason 1935, Weinberg and Guillaumie 1936, Dalling and Ross 1938, Duffett 1933, Stewart 1940, Taylor and Stewart 1941). Of these five components, *Cl. welchii* Type A was at first thought to contain only  $\alpha$ . Prigge (1936, 1937) and Ipsen and his colleagues (Ipsen 1939, Ipsen *et al.* 1939a, b) found two components,  $\alpha$  and  $\zeta$ , and in one Type A strain, a third component, which they designated  $\eta$  (see also Nagler 1940). It is now clear that Prigge's  $\zeta$  toxin is equivalent to Glenny's  $\alpha$ , and British workers (see Dalling and Stephenson 1942) have recently adopted a convention whereby Prigge's  $\alpha$  and  $\zeta$  are designated  $\theta$  and  $\alpha$  respectively. The  $\eta$  toxin of Ipsen retains its original designation. There are, therefore, seven toxic components to consider,  $\alpha$ ,  $\beta$ ,  $\gamma$ ,  $\delta$ ,  $\epsilon$ ,  $\theta$  and  $\eta$ . Space forbids more than a

brief outline of the properties and relationships of the *Cl. welchii* toxins. For a full discussion the reader is referred to the exhaustive review of Oakley (1943) from which with certain modifications we reproduce Table 54 showing the toxins present in the different filtrates, and Table 55 showing the properties of the toxins.

TABLE 54

GIVING THE DISTRIBUTION OF TOXIC COMPONENTS IN CULTURE FILTRATES OF *Cl. welchii* (Oakley 1943)

Variety of <i>Cl. welchii</i>	Wilson's Type	Toxin						
		$\alpha$	$\beta$	$\gamma$	$\delta$	$\epsilon$	$\eta$	$\theta$
<i>Cl. welchii</i>	A	+++	—	—	—	—	(+)	+
<i>Cl. agnis</i>	B	+	+++	+	+	++	?	?+
<i>Cl. paludis</i>	C	+	+++	+	++	—	?	?+
<i>Cl. ordalii</i>	D	+	—	—	—	+++	?	?+

It will be seen that the  $\alpha$  toxin predominates in Type A  $\beta$  and  $\epsilon$  in Type B  $\beta$  and  $\delta$  in Type C and  $\epsilon$  in Type D filtrates. The distinctions both qualitative and quantitative between the types are not absolute. Mason (1935) for instance records the loss of ability to produce  $\epsilon$  toxin in Type B strains. Borthwick (1937) a similar loss in Type D strains and Taylor (1940) a loss of  $\beta$  toxigenicity in a Type B strain. The properties of the toxins are summarized in Table 55.

TABLE 55

GIVING THE PROPERTIES OF THE TOXINS OF *Cl. welchii* (Oakley 1943)

Toxin	Hæmolytic	Lethal	Necrotizing	Lecithinase	Effect of heat
$\alpha$	+	+	+	+	Thermostable
$\beta$	+	+	+	—	Thermolabile
$\eta$	—	+	—	—	
$\beta$	—	+	+	—	Thermolabile
$\gamma$	—	+	—	—	
$\delta$	+	+	—	—	
$\epsilon$	—	+	+	—	Culture filtrate thermostable Intestinal fluid thermolabile

The  $\alpha$  toxin is a thermostable substance lethal for mice guinea pigs rabbits pigeons and sheep and when given intradermally produces a necrotic lesion. It is hæmolytic for the red cells of most laboratory animals excepting the horse and the goat and is a powerful lecithinase. The lecithinase activity of  $\alpha$  toxin is of great interest since it is the first known instance of an exotoxin dependent upon a demonstrable enzyme for its activity. Its discovery dates from the demonstration by Nagler (1930) that toxic filtrates of all four types of *Cl. welchii* produced an opalescence in human sera and that the reaction was specifically inhibited by antisera to Type A filtrates (see also Seiffert 1939). The reactivity ran parallel to the toxin content of filtrates. Macfarlane, Oakley and Anderson (1941) demonstrated a similar action of  $\alpha$  toxin on extract of egg yolk and suggested that both phenomena were due to an enzymic splitting of lipo-protein complexes in serum and egg yolk respectively, and that hæmolysis by  $\alpha$  toxin might also be due to its action on lipo proteins in the red cell envelope. The identity of  $\alpha$  toxin and lecithinase was further established by Macfarlane and Knight (1941) who demonstrated a quantitative splitting of lecithin by  $\alpha$  toxin into phosphocholine and a glyceride and the necessity for Ca or Mg ions in the reaction. The lecithinase activity of a filtrate may be used as



a measure of its  $\alpha$  toxin content and for comparisons of the neutralizing power of  $\alpha$  anti-toxins (for details see Nagler 1939 Oakley and Warrack 1941 van Heyningen 1941a Crook 1942). Lecithinase production in fluid and on agar cultures of *Cl. welchii* may be detected by incorporating human serum or egg yolk extract in media containing a sufficiency of free calcium. Other bacteria, mainly spore bearing bacilli, produce lecithinase-like substances (Nagler 1939 Hayward 1941 Crook 1942) but with one exception, *Cl. bisfermentans* (Hayward 1943) none is fully neutralized by  $\alpha$ -antitoxin.

The  $\theta$  toxin has been found only in Type A filtrates. It is a strong haemolysin, and is also lethal and necrotizing. Todd (1941) has shown that it is thermolabile and oxygen-labile. The activity of oxidized  $\theta$  toxin is restored by reducing agents containing sulphhydryl groups. This property of reversible oxidation is shared by the O-streptolysin of *Streptococcus* (Chapter 24). There is also an antigenic relationship between the two substances, each being to a large extent neutralized by antisera prepared against the other (Todd 1941).  $\theta$  toxin may be removed from mixtures with  $\alpha$  toxin by adsorption on to susceptible red cell stromata (van Heyningen 1941b see also Gale and van Heyningen 1942).

The  $\beta$  toxin is not haemolytic: it is lethal to mice producing on intravenous injection a spasmodic twitching rapidly followed by death given intradermally in guinea pigs and rabbits it produces necrotic lesions. The  $\gamma$  toxin is neither necrotizing nor haemolytic. Its existence in filtrates can be proved only by its lethal activity in mice after the other toxins have been neutralized by appropriate antitoxins. The  $\delta$ -toxin is haemolytic and lethal and, like  $\gamma$  toxin, is detected after neutralization of other components by anti-toxins. The  $\epsilon$  toxin, which predominates in Type D filtrates is not haemolytic, but is both lethal and necrotizing. Large doses in mice produce spasmodic twitching similar to those following injections of  $\beta$ -toxin. Smaller doses after a latent period of several days, produce paralysis. The  $\epsilon$  toxin is produced by Type D strains as a thermostable relatively non-toxic substance which is activated by trypsin and other proteolytic enzymes with the formation of the thermolabile toxin (Gill 1933, Boeworth and Glover 1935). In disease produced by Type D strains (see Chapter 3) the activating enzyme is apparently supplied by the infected animal, though according to Turner and Rodwell (1943) in favourable conditions extracellular proteinases of the bacillus itself will activate the toxin. It should be noted that culture filtrates of Type A strains may also contain large amounts of a hyaluronidase (McClellan 1936 see Chapters 44, 53).

The toxin of *Cl. ordalii* is the most potent of the gas-gangrene toxins: the average M.L.D. for a mouse is about 0.0005 ml. of *welchii* toxin the M.L.D. is about 0.25 ml. and of *Cl. septicum* toxin about 0.005 ml.

Toxic filtrates of *Cl. septicum* produce a marked liquefactive necrosis of muscle: lethal doses when given intravenously produce intense capillary engorgement and interstitial haemorrhages in the heart with hyaline degeneration of the muscle-fibres, and a toxic nephrosis in the kidney of experimental animals (Pasternack and Bengtson 1936). Filtrates may also contain a haemolysin and a hyaluronidase. The haemolysin has usually been regarded as distinct from the lethal toxin but Bernheimer (1944) has produced filtrates in which the lethal and the haemolytic activities are substantially parallel.

Toxic filtrates of *Cl. ordalii* contain a potent toxin that produces intense gelatinous oedema in muscle. There are few gross changes in the organs following a lethal intravenous dose of toxin, but degenerative changes, particularly in the spleen and kidney have been observed (Pasternack and Bengtson 1940). Traces of lecithinase, a hyaluronidase and a haemolysin are sometimes present.

*Cl. histolyticum* usually produces a weakly toxic filtrate that contains an active proteolytic enzyme. Toxicogenic strains of *Cl. bisfermentans* have been described under the name of *Cl. sordellii*: the filtrates are moderately toxic. All the toxins

of these five organisms associated with gas gangrene give rise to specific antitoxins on injection into suitable animals.

*Cl chauvei* under suitable conditions forms a weak toxin that is very heat-labile, being destroyed in 5 minutes by exposure to a temperature of 52° C. Injected intravenously into mice in a dose of 0.025-0.5 ml., it causes respiratory embarrassment and death within a few minutes. It is also toxic to guinea pigs, though not rabbits, on intravenous inoculation. Subcutaneous inoculation into mice and guinea pigs is not fatal, but produces a local blood-stained oedema (Kerrin 1934).

It is interesting to note that all the different groups of *Cl tetani* give rise to identical toxins, the antitoxin prepared against any one type will neutralize the toxins of all types. With *Cl botulinum* it is otherwise. Type A toxin is different from Type B toxin. By agglutination Type A strains can be divided into 4, and Type B into 3 groups (Starin and Dack 1923), but the divisions do not appear to be clear-cut. Three further types have been described, C, D, and E, which appear to differ in the type of toxin produced.

**Pathogenicity**—The pathogenicity of the anaerobes appears to depend almost entirely on their toxin production. *Cl tetani*, for example, multiplies locally, and does not invade the body. *Cl botulinum* is not even a parasite, it is apparently unable to grow in the tissues, and its pathogenic effects are determined by the formation of toxin in food-substances prior to their ingestion. *Cl oedematiens* remains almost confined to the site of inoculation. *Cl welchii* and *Cl septicum* become generalized in the final stages of an infection, but they multiply only locally before the death of the animal. Tetanus, botulism, and to a large extent gas gangrene are intoxications.

#### Pathogenicity of *Cl. botulinum* for Laboratory Animals

**MONKEYS.**—Van Ermengem (1897) fed a *Macacus rhesus* with 5 ml. of a preparation of macerated ham which was known to be toxic. Symptoms developed in 12 hours, and consisted of restlessness, crying, coughing and sneezing. Later there was a secretion of viscid mucus in the nose and mouth, leading to transient suffocation. The pupils were dilated, reacting weakly to light. The animal became motionless, its head drooped, its eyes were fixed and half covered by the lids. Death occurred after 24 hours from the time of feeding. At necropsy the stomach, the bases of the lungs, and the meninges were congested, and petechial hæmorrhages were noticed on the arachnoid and throughout the brain and medulla.

**CATS.**—The typical toxæmia may be reproduced in cats by feeding, but more certainly by subcutaneous injection of cultures or of toxin. After a latent period of about 24 hours the animal becomes quiet, loses its interest in external objects, and may refuse food. In 2 or 3 days the characteristic paralysis appears, giving a peculiar facies to the animal. Its general aspect is stupid, the lids remain open, the eyes fixed in a glassy stare, the pupils dilated and sluggish in their reaction to light. The animal sits in a dark corner, moves little, and when disturbed takes a few uncoordinated steps across the cage and drops down as if exhausted. Its head droops and its tongue protrudes. Thick, viscous secretion fills the throat and nose, and causes severe paroxysmal attacks of suffocation relieved by a hoarse croup-like cough. The mew takes on a dull tone and is succeeded by complete aphonia. For the first 2 or 3 days milk is accepted, but later owing to the dysphagia or complete aphagia it is left untouched, when delivered by a pipette into the mouth it is not swallowed, but trickles down the trachea and causes choking. No urine or faeces are voided. The animal remains susceptible to sensory impressions till the end but is unable to express its emotions in any way. Death occurs after a week or more, according to the dose, and is apparently due as much to starvation as to the lethal effect of the

toxin. Occasionally life may be prolonged for 3 or 4 weeks, and recovery may eventually take place. At necropsy no local lesion is visible at the site of injection; the mucosa of the small and large intestine is hyperemic. The kidneys are congested and the liver may show areas of degeneration. Clear urine distends the bladder. In the lungs, which are very congested, there may be infarcts or areas of hepatization. Sometimes oedema or hemorrhages of the central nervous system may be observed, especially round the fourth ventricle. Cultures of the organs are usually sterile.

Dogs are very much less susceptible than cats, but they may succumb to the disease after subcutaneous injection of toxin, or occasionally after feeding with large doses. Mice and guinea pigs are highly susceptible and succumb in 1-4 days.

### Pathogenicity of CL tetani for Laboratory Animals

Tetanus can be reproduced by the inoculation of pure cultures or of the toxin into mice, rats, guinea pigs, rabbits, goats, horses and monkeys. Cats and dogs are more resistant; birds and cold blooded animals are highly resistant. The most susceptible animal, calculated on the amount of toxin per gram of body weight necessary to prove fatal on injection, is the horse. This is about 12 times as susceptible as the mouse; the guinea pig is 6 times, and the monkey 4 times, as susceptible as the mouse (von Lingelheim 1912; Sherrington 1917). On the other hand, the rabbit is twice, the dog 50 times, the cat 600 and the hen 30 000 times as resistant as the mouse (Kitasato 1891; von Lingelheim 1912).

**MICE.**—After the *subcutaneous* injection of a small quantity of toxin or of pure culture into the mouse near the root of the tail, symptoms develop in about 1<sup>st</sup> to 2<sup>nd</sup> hours. The spasms start near the site of injection, and spread to the rest of the body till the animal dies in a state of general tonic contraction. The first symptom noticed is a stiffening of the tail, which becomes erect and is turned towards the side of inoculation; the hinder extremity of the side becomes stiff followed later by rigidity of the opposite leg. The contractions pass to the muscles of the trunk, and the mouse develops kyphosis or pleurothotonos. Next, the fore legs become involved, and finally trismus and opisthotonos set in. The contractions occur spasmodically and are succeeded by intervals of rest during which the animal becomes exhausted; in this phase they can be readily excited by the slightest touch or a breath of air. Death follows in about 2<sup>nd</sup> to 4<sup>th</sup> hours. Post mortem there is little to be seen. There may be slight congestion and oedema round the site of inoculation, and the spleen may be somewhat enlarged. An exudate of fluid, sometimes blood-stained, may be seen in the pleura or peritoneum. After injection of a pure culture, the bacilli can generally be cultivated from the local site, but are difficult to find under the microscope. The heart's blood and viscera are sterile.

**GUINEA PIGS.**—The experimental disease in guinea pigs follows much the same course in about the same time as in mice.

**RABBITS.**—After subcutaneous or intramuscular injection the incubation period in rabbits is at least 24 and generally 36 hours; death does not occur for 3 or 4 days. The general tetanic spasms are more marked than in mice or guinea-pigs (Rosenbach 1896).

### Pathogenicity of CL welchii for Laboratory Animals.

Intramuscular injection of about 0.2 ml. of an 18-hours glucose broth culture into the thigh of a guinea-pig usually results in gas gangrene with death in 1<sup>st</sup> to 48 hours. Post mortem there is a large brawny crepitant swelling at the site of inoculation covered with a dark red, tense layer of skin. The muscle is pale and is undergoing liquefactive necrosis. In the subcutaneous tissue around the local lesion and spreading up to the abdomen, reaching sometimes to the sternum and over to the opposite thigh, is a collection of slightly blood-stained fluid and gas smelling of hydrogen sulphide. The suprarenal glands are often congested, so that the normally sharp differentiation of cortex from medulla

becomes obscured. Microscopically the organisms are present in large numbers in the local effusion and in much smaller numbers in the blood stream. Sporing forms are absent. An even more typical picture of gas gangrene can be obtained by the injection of *Cl. welchii* intramuscularly into pigs (Bull and Pritchett 1917a). Mice are less susceptible than guinea pigs.

#### Pathogenicity of *Cl. septicum* for Laboratory Animals

Intramuscular injection of about 0.1 ml. of a 36 hours glucose broth culture into a guinea pig causes death in 12 to 24 hours. Post mortem there is a blood stained gaseous oedema at the site of inoculation spreading up over the abdominal wall with collections of gas in the groins and axillae. The thigh and abdominal muscles are soft and deep red in colour. In the pericardial and peritoneal cavities there may be some fluid, the suprarenals are congested but not so markedly as in animals infected with *Cl. welchii*. Microscopically the exudate shows large numbers of motile rods and usually the characteristic navicular or citron forms. Most characteristic are the long curved filaments found on the peritoneal surface of the liver. A similar picture can be reproduced by the inoculation of mice.

#### Pathogenicity of *Cl. oedematiens* for Laboratory Animals

Intramuscular injection of about 1 ml. of a 24 hours glucose broth culture into a guinea pig or mouse produces death in 1 to 2 days. Post mortem the muscles at the site of inoculation are very congested purplish red in colour, and infiltrated with small bubbles of gas. There is a spreading gelatinous oedema sometimes slightly blood tinged extending over the thigh. The abdominal muscles are unaltered. Microscopically bacilli are found in small numbers in the oedema fluid and on the peritoneal surface of the liver. Cultures from the heart's blood may or may not be positive.

It will be seen that the action of these last three organisms varies in certain particulars. *Cl. welchii* gives rise to a large amount of gas, *Cl. oedematiens* to very little. The oedema fluid of *Cl. oedematiens* infections is practically clear, of *Cl. welchii* infections slightly blood tinged and of *Cl. septicum* infections strongly blood tinged. With *Cl. welchii* the muscles are pale pink, with *Cl. oedematiens* purplish red and with *Cl. septicum* intensely and deeply red. Human cases of gas gangrene differ too in certain respects, as a rule either oedema or more rarely gas production is dominant, occasionally both are apparent. The particular form in any individual case is determined by the nature of the organisms present.

### CLASSIFICATION

Although it is clear that the time is not yet ripe for any rigid classification of the anaerobic bacilli, we can recognize certain well-differentiated types which should clearly be accorded specific rank. Table 56 presents a rough classification of the organisms chiefly associated with gas gangrene and similar infections in man which were considered by the Anaerobic Committee of the Medical Research Council (Report 1919) to be separate species. We may note however that the characters differentiating *Cl. parasporogenes* from *Cl. sporogenes* are hardly of sufficient importance to entitle it to classification as a separate species; it may prove on further investigation to be merely a variant of the latter organism. Moreover the identity of *Cl. butyricum* with the organism originally described by Pasteur seems to be very doubtful; it is unfortunate that this organism has been selected as the type species. In many respects it resembles *Cl. fallax* and *Cl. multi-fermentans*. A few organisms have since been studied in sufficient detail to provide an adequate description of their biological characters and to differentiate them

clearly and unmistakably from other forms which have been described and named. Some of these we have included in Table 57 and others are described at the end of the chapter. Nevertheless, a large number of named forms remain, whose place in the *Clostridium* group is still a matter of conjecture, as reference to the monograph of Weinberg, Natrville and Prevot (1937) will show. Prevot (1932) rejects the genus *Clostridium* as incapable of covering all the anaerobic spore-bearing bacilli, and proposes to create four families and nine genera, of which *Clostridium* is one, this classification in our opinion places too much weight on morphological distinctions. The recognition of variation of the S → R type and detailed serological studies are helping to resolve some taxonomic difficulties, but it is clear that a large number of clostridia so far studied are antigenically heterogeneous, and that the variety of antigens, H, thermostable O, labile O, etc., is probably as great as that displayed by the salmonellas.

TABLE 56  
GIVING A CLASSIFICATION OF THE *Clostridia*.

Spores.	Both Proteolytic and Saccharolytic Properties.		Slight Proteolytic but no Saccharolytic Properties.	Saccharolytic but no Proteolytic Properties.	Neither Proteolytic nor Saccharolytic Properties.
	Proteolytic Predominating.	Saccharolytic Predominating.			
Equatorial or Subterminal	<i>Cl. sporogenes</i> <i>Cl. parasporegenes</i> <i>Cl. histolyticum</i> <i>Cl. acetatidum</i> <i>Cl. bisfermentans</i> <i>Cl. botulinum</i>	<i>Cl. welchii</i> <i>Cl. septicum</i> <i>Cl. charovii</i> <i>Cl. oedematis</i>	—	<i>Cl. fallax</i> <i>Cl. butyricum</i> <i>Cl. malisfermentans</i>	—
Oval and Terminal	—	—	—	<i>Cl. tertium</i>	<i>Cl. cockleorum</i>
Spherical and Terminal	—	—	<i>Cl. tetani</i>	<i>Cl. tetanomorphum</i> <i>Cl. sphenoides</i>	—

(For classification see Weinberg and Segum 1918, Report 1919, Heller 1921, and Hall 1922.)

The grouping of the species within the genus *Clostridium* presents even greater difficulties. Whether the primary division should be made on morphological grounds—mainly on the shape and position of the spore—or on physiological grounds—mainly on the relative activity of proteolytic and saccharolytic fermentation—must at the moment remain a matter of choice. We give in Table 57 (pp. 874, 875) the more important characters of the recognized species. We also append a summarized description of each of a number of recognized species. It should be noted that we have omitted to discuss clostridia like *Cl. acetobutylicum* (see Chapters 3 and 6), which are of interest to the biochemist, and have made only a brief reference to the clostridia associated with the spoilage of canned foods. Of particular interest is *Cl. thermosaccharolyticum* (McClung 1935), which is an apparently non pathogenic thermophilic organism having an optimum temperature

of growth between 50° and 60° C, associated with the type of spoilage of non acid canned foods known in the United States as "hard swell" (see also Paine 1931, McCoy 1937) In addition to the common proteolytic clostridia of the *Cl sporogenes* type associated with food spoilage, we may note an organism resembling *Cl welchii* (McClung and Wheaton 1936), and an organism resembling *Cl oedematiens* which Haines and Scott (1940) found associated with "bone taint" of cattle carcasses (For practical keys to the separation of the clostridia see Spray 1936, Reed and Orr 1941)

### *Clostridium butyricum*

*Synonyms* — *Clos pasteurianum*, *B amylobacter*, *Granulobacter saccharobutyricum* Pasteur & *Vibron butyrique*

*Isolation* — Apparently first described by Prazmowski (see Report 1919) Possibly identical with Pasteur's *Vibron butyrique*, described fully by Winogradsky in 1902

*Habitat* — Soil.

*Morphology* — Rods, 3-4  $\mu$   $\times$  0.7  $\mu$ , parallel sides, flattened ends, axis straight or slightly curved, arranged singly or in pairs end to end, considerable variation in length. Motile by peritrichate flagella Spores oval, subterminal, measuring 1.6  $\mu$   $\times$  1.3  $\mu$ ; rod becomes spindle-shaped Germination polar Capsule formed on agar Cells store glycogen; stain yellow with iodine Gram positive.

*Agar Plate*.—2 days, 30° C Circular colonies, 0.5-1.0 mm in diameter, low convex, amorphous, faintly translucent or opaque, greyish white, with smooth *glistening surface and entire edge*, butyrous consistency and easily emulsifiable After 6 days the colonies are slightly larger

*Agar Stroke*—4 days at 30° C Very poor growth of discrete, irregular colonies, slightly raised, and water clear On 0.5 per cent mannitol agar there is a moderate,

raised, greyish white, opaque growth, consisting chiefly of discrete colonies with a moist, glistening, smooth surface

*Gelatin Stab*—No growth

*Glucose Agar Shake*—2 days at 30° C Good growth, the medium is disrupted by large bubbles of gas, and blown up the tube Colonies are yellowish grey, opaque, biconvex with clear cut edges, and about 1 mm in diameter

*Broth*—2 days at 30° C Poor to moderate growth, with slight turbidity, and slight, very finely granular deposit, after 6 days, moderate turbidity, and moderate powdery deposit, disintegrating completely on shaking

*Loeffler's Blood Serum*—2 days at 30° C Moderate, raised, confluent, colourless growth with irregularly contoured surface. No digestion

*Cooked Meat Medium*.—5 days, 37° C. Marked turbidity, no digestion.

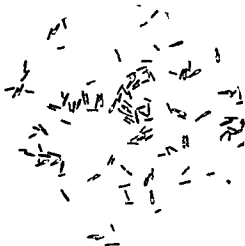


FIG 192 — *Clostridium butyricum*  
From a surface agar culture, anaerobically, 5 days,  
37° C ( $\times$  1000)

TABLE

Name	St. area	Liquefaction of Gelatin	Digestion of Starch	Cooked Meat Medium	Iron Litmus Milk	Dextrose	Maltose
<i>Cl butyricum</i>	O, S	—	—	G	A, C, G	AG	AG
<i>Cl sporogenes</i>	O, S	+	+	G, O, D, b	a, g, → ppt, D, B	AG	AG
<i>Cl histolyticum</i>	O, S	+	+	O, D, B xtls	a, g, ppt., → D, B	(A)	(A)
<i>Cl aerofaciendum</i>	O, S, rare	+	—	G, O, D, b	a, → ppt, D, B	AG	AG
<i>Cl bisfermentans</i>	O, S	+	+	G, O, d, B, xtls	a, g, → ppt, D, B	AG	AG
<i>Cl botulinum</i>	O, S	+	(+)	G, O, (D, b)	a, → ppt, D, B	AG	AG
<i>Cl adematensis</i>	O, S	+	—	G, pink	a, g, → C	AG	AG
<i>Cl fallax</i>	O, S, rare	—	—	G, pink	A, C, g	AG	AG
<i>Cl multifementans</i>	O, S	—	—	G	A, C, G	AG	AG
<i>Cl chauvoei</i>	O, S	+	—	G, pink	A, C, g	AG	AG
<i>Cl septicum</i>	O, S	+	—	G, pink	A, (C), g	AG	AG
<i>Cl welchii</i>	O, S, rare	+	—	G, pink	A, (C, G)	AG	AG
<i>Cl difficile</i>	O, S	(+)	—	—	g	AG	—
<i>Cl carnis</i>	O, S	—	—	G	G	AG	AG
<i>Cl haemolyticum</i>	O, S	+	—	G	A, C	AG	—
<i>Cl hastiforme</i>	O, S	+	—	(o)	Ppt, → D, b	—	—
<i>Cl tertium</i>	O, T	—	—	G	A, g, → C	AG	AG
<i>Cl cochlearium</i>	O, T rare	—	—	—	—	—	—
<i>Cl capitovale</i>	O, T	+	—	G, O, B	ag, → (ppt, d)	AG	—
<i>Cl paraputrificum</i>	O, T	—	—	G	A, C, g	AG	AG
<i>Cl tetani</i>	R, T	+	(+)	g (b, o)	a, → (ppt)	—	—
<i>Cl putrificum</i>	R, T, rare	+	+	O, D, b	ppt., → d B	—	—
<i>Cl tetanomorphum</i>	R, T	—	—	G	(a)	AG	AG
<i>Cl sphenoides</i>	R, T	—	—	G	A, G	AG	AG

A or a = acid, B or b = blackening, C or c = clot, D or d = digestion, G or g = gas, ppt = precipitate, R = round (spherical), S = subterminal (or equatorial), T = terminal. Brackets indicate that the activity is not a characteristic of all members of the species. Rare = rarely observed microscopically in cultures.

Mannitol	Lactose	Sucrose	Saltin	Glycerol	Exotoxin	Pathogenicity to Guinea pigs	Remarks
?	AG	AG	AG	(AG)	-	-	Possesses a capsule
-	-	-	(AG)	(AG)	-	-	Vanillin violet test +
-	-	-	-	-	+	(+)	Feeble aerobic growth
-	AG	-	(AG)	-	-	-	
-	-	-	(AG)	AG	(+)	(+)	Only toxigenic strains are pathogenic Indole +
-	(AG)	-	(AG)	(AG)	++++	+	5 types A B C D and F each with a different toxin Some strains not proteolytic
-	-	-	-	(AG)	+++	+	
(AG)	AG	AG	AG	-	+	(+)	Pathogenic only when freshly isolated
-	AG	AG	AG	AG	-	-	
-	AG	AG	-	-	++	+	
-	AG	-	AG	-	++	+	
-	AG	AG	(AG)	(AG)	++	(+)	Possesses a capsule Non motile Four types each with a different combination of toxins
AG	-	-	AG	-	+	+	
-	AG	AG	AG	-	+	+	Feeble aerobic growth
-	-	-	-	AG	++	+	
-	-	-	-	-	-	-	
AG	AG	AG	AG	-	-	-	Grows aerobically
-	-	-	-	-	-	-	
-	-	-	-	-	-	-	Indole (+)
-	AG	AG	AG	-	-	-	
-	-	-	-	-	++++	(+)	Indole +
-	-	-	-	-	-	-	
-	-	-	-	-	-	-	
AG	AG	-	AG	-	-	-	Indole (+)

O (under Spores) = oval O or o (under Cooked Meat Medium) = foul odour  
 xils = late deposit of white crystals Small letter symbols indicate a slighter degree of change  
 Under exotoxin +, ++, +++, +++++ indicate the relative potency of toxic filtrates



**Potato**—14 days at 30° C No definite growth visible on the potato itself but there is a considerable evolution of gas from the liquid in which the potato is soaked the liquid is turbid.

**Metabolic**—Obligate anaerobe. Opt temp 30–40° C Pigment none. *Nutritional* grows best in sugar solutions Growth on agar improved by 1 per cent mannitol Non proteolytic

**Biochemical**—Forms acid and gas in dextrose laevulose maltose galactose lactose sucrose inulin dextrin and starch Fermentation of mannitol variously reported Indole —, MR + VP — nitrates reduced VLB reduct on — catalase — NH<sub>3</sub> slight + LM acid and clot Can fix atmospheric nitrogen in presence of a fermentable sugar the sugar is broken down with the formation of butyric and acetic acids CO<sub>2</sub> and H<sub>2</sub> Can utilize NH<sub>3</sub> peptone and asparagin as its source of nitrogen

**Pathogenicity**—N L

### *Clostridium sporogenes*

**Isolat on**—Described by Metchnikoff in 1908

**Habitat**—Found in soil and in faeces of man and animals

**Morphology**—Rod shaped 3–6  $\mu \times 0.5$   $\mu$  parallel sides rounded ends axis straight or slightly curved arranged singly in pairs and small groups long filaments occasionally formed. Spores

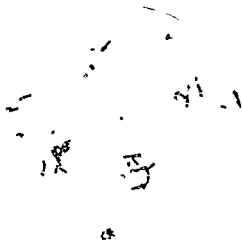


FIG 193—*Clostridium sporogenes*

From a surface agar culture anaerobically 2 days 37° C ( $\times 1000$ )

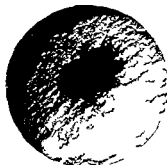


FIG 194—*Clostridium sporogenes*

Surface colony on agar anaerobically 4 days 37° C ( $\times 8$ )

freely spores are oval subterminal and wider than bacillus free spores numerous Motile No capsule Strongly Gram positive except in old cultures.

**Agar Plate**—4 days at 37° C Irregularly round colonies growing from a central focus like *B. mycoides* 2–6 mm. in diameter effuse or slightly umbonate and rhizoid, surface covered by arborescent ridges, edge rhizoid rather dull greyish yellow by reflected bluish grey by transmitted light butyrous and easily emulsifiable differentiated into brownish opaque centre and bluish translucent periphery

**Deep Glucose Agar**—4 days at 37° C Abundant gas formation, medium driven to plug and disrupted. Colonies throughout medium rounded, 0.5–2 mm in diameter with opaque brown centre and a woolly translucent periphery

**Horse Blood Agar**—3 days at 37° C Colonies 3 mm in diameter consisting of tangled rhizoids growing from a raised glistening centre 0.5 mm in diameter on moist

media smooth lobate outgrowths may arise from a rhizoidal centre. Occasionally round dew drop colonies are formed. Hemolysis for 2-4 mm around colony.

*Agar Slope*.—4 days at 37° C. Moderate, confluent, effuse, glistening greyish yellow, translucent growth, with beaten copper surface and cigarette-in water edge.

*Gelatin*.—2 days at 37° C. Liquefied.

*Broth*.—4 days at 37° C. Good growth with moderate turbidity, and moderate powdery or irregularly granular sediment, not disintegrating completely, strong putrid odour.

*Loeffler's Serum*.—15 days at 37° C. Serum digested and rendered turbid, medium dark blue.

*Coagulated Egg*.—15 days at 37° C. Poor, effuse growth, slight digestion. In alkaline egg broth the white coagulum is digested.

*Cooked Meat Medium*.—15 days at 37° C. Heavy growth with dense turbidity, gas production, meat digested and blackened, putrid odour.

*Persistence*.—Withstands moist heat at 100° C. for 10 to 150 minutes, at 105° C. for 4 to 45 minutes, and at 110° C. for 1 to 12 minutes.

*Metabolic*.—Anaerobic, but not strictly so. Opt temp 37° C. Hemolysis on horse blood agar plates. Hemolyzes human but not sheep's red cells. *Nutritional* grows well on ordinary media, and in media containing very little nutrient material, such as tap water containing fragments of coagulated egg white. Certain amino acids, such as tryptophan, leucine, phenylalanine, tyrosine, and arginine, as well as the *sporogenes* vitamin, are essential. Growth not improved by glucose. Green fluorescent colonies on MacConkey plate.

*Biochemical*.—Acid and gas in glucose and maltose. No action on mannitol, lactose, or sucrose. Some strains ferment salicin. Indole —, vanillin violet + (Spray 1936), MR —, VP —, nitrites not produced in nitrate broth,  $\text{NH}_4^+$  ±,  $\text{H}_2\text{S}$  + + +, MB reduction —, catalase weak +. Litmus milk casein precipitated and almost completely digested in 15 days, reduction and marked alkaline reaction, acid in young cultures.

*Antigenic Structure*.—Can be divided by agglutination into at least two groups.

*Pathogenicity*.—Not naturally pathogenic. Experimentally is non pathogenic to laboratory animals but enhances the pathogenicity of other anaerobes, such as *Cl. welchii*, in mixed cultures. No exotoxin formed but a broth filtrate is toxic to guinea pigs in a dose of 1 ml., this is due apparently to a volatile substance possibly an ammonium base. Forms a fibrinolysin.

(See von Hibler 1908, Wolf and Harris 1917, Weinberg and Séguin 1918, Report 1919, Hall 1922, de Smidt 1924, Weinberg and Ginsbourg 1927, Knight and Fildes 1933, Fildes and Richardson 1935, Stickland 1934-1935, Pappenheimer 1935, Spray 1936.)

### *Clostridium histolyticum*

*Isolation*.—Described by Weinberg and Séguin in 1916 (1916, 1918).

*Habitat*.—Soil, possibly intestinal canal of man and animals.

*Morphology*.—Rod-shaped 3-5  $\mu \times$  0.5-0.8  $\mu$ , parallel sides, rounded ends, axis generally straight, occur singly and as diplobacilli. In cultures more than a day old irregular forms appear—long curved filaments, and irregularly stained forms. Spores are readily formed in all media, they are oval, subterminal, and wider than the bacillus, become free in old cultures. Motile by about 20 peritrichate flagella. Gram positive in young cultures. No capsule.

*Agar Plate*.—4 days at 37° C. Variable. Colonies may be delicate and flat with crenated edges, or may be cuttle-fish like, umbonate, amorphous and glistening, with very finely granular surface and a fimbriate edge, greyish white by reflected light, bluish grey by transmitted light, differentiated into opaque yellowish centre and greyish translucent periphery, butyrous and easily emulsifiable.

- Deep Glucose Agar Shake**—4 days at 37° C No gas. Abundant growth throughout medium. Colonies are 1 mm in diameter irregularly round opaque brown with blunt coral like projections with very fine woolly ends.
- Horse Blood Agar Plate**—4 days at 37° C Irregularly round colonies slightly raised, 2-3 mm in diameter with irregularly lobate edge, and smooth or pitted surface. No hæmolytic.
- Agar Slope**—4 days at 37° C Moderate partly confluent effuse, glistening translucent, greyish yellow growth with beaten-copper surface and delicate fimbriate edge.
- Gelatin**—Liquefied in 3 days at 37° C.
- Broth**—4 days at 37° C Moderate growth with moderate turbidity and a granulo-powdery deposit partly disintegrating slight foetid odour.
- Loeffler's Serum**—15 days at 37° C Almost completely digested, the fluid is almost clear.
- Coagulated Egg**—15 days at 37° C Partly digested butt turned bluish green.
- Cooked Meat Medium**—15 days at 37° C Abundant growth meat digested and slightly blackened, long column of slightly turbid fluid, gas produced, a deposit of white tyrosine crystals occurs increasing with age. Slightly foetid odour.



FIG 195—*Clostridium histolyticum*

From a surface culture on agar anaerobically  
6 days 37° C (× 1000)



FIG 196—*Clostridium histolyticum*

Surface colony on agar anaerobically,  
2 days 37° C (× 8)

- Persistence**—Killed in 6 minutes at 105° C. (moist heat)
- Metabolism**—Microaerophilic Opt temp 37° C. No hæmolytic on horse blood agar plates. Hæmolytic human but not sheep's red cells. Nutritional grows well in ordinary media, growth not improved by glucose. Green fluorescent colonies on MacConkey plate. Forms a fibrinolysin.
- Biochemical**—Some strains produce acid, but no gas in glucose and occasionally in maltose. No fermentation of mannitol, lactose, sucrose or salicin, sometimes no acid produced at all. Indole —, MR —, VP —, nitrates not reduced, NH<sub>3</sub> —, H<sub>2</sub>S +++ ALB reduction —, catalase + weak. Litmus milk casein precipitated and digested reduction after 8 to 10 days it is transformed into a clear amber coloured fluid.
- Antigenic Structure**—Agglutinating sera seem to act chiefly on homologous strains. Anti-toxin can be prepared by injection of toxin into horses.
- Pathogenicity**—Exotoxin is said to be formed in very young cultures. Natural pathogenicity doubtful, appears often in gangrenous processes in man. Experimentally strains vary in pathogenicity, susceptible animals are guinea pig rabbit and mouse. Bacillus is actively proteolytic and digests living tissue. 1 ml. of a young

broth culture injected intramuscularly into a guinea pig causes digestion of skin and muscles and a hæmorrhagic liquefaction of the softer parts of the limb. This digestion may spread over the abdomen and death occur during the next 12 to 24 hours or recovery may follow with more or less complete necrosis of the limb. The fluid contains no gas and is not putrid.

(See Weinberg and Séguin 1918 Report 1919 Hall 1922 Torrey 1925 Weinberg *et al* 1926.)

### *Clostridium botulinum*

**Isolation**—By van Ermengem from ham in 1896 (1896 1897). Several other organisms have since been isolated from botulism like diseases in animals. These are sometimes referred to as *Cl. paratubulinum* but it is probably better to give them letters using A and B to refer to the two main toxigenic types. Type C<sub>1</sub> was isolated by Bengtson (1922a b 1923) and by Graham and Boughton (1930a b) in the United States from chickens and ducks. Type C<sub>2</sub> by Seddon (1922) in Australia from cattle, Type D by Theiler and his colleagues (1926) in South Africa from cattle and Type E by Theiler (1928) in South Africa from horses. The relation between Types C, D and E is not yet entirely clear but Types C and E are said to be identical. Following description refers chiefly to Types A and B.

**Habitat**—Widely distributed in soil both virgin and cultivated. Not infrequently present in intestinal tract of domestic animals.

**Morphology**—4-day agar slope at 37° C. Rather large stout rods 4.6  $\mu$   $\times$  0.9  $\mu$ , axis straight parallel sides and slightly rounded ends arranged singly or sometimes in pairs or chains. Variations in depth of staining. Spores are oval wider than the bacillus, thick walled and situated at or near the end. Free spores are numerous. Some strains spore readily others hardly at all. Spores formed best in sugar-free media. Sluggishly motile by 4-8 peritrichate flagella. No capsule. Gram positive in young cultures.

**Agar Plate**—4 days at 37° C. Irregularly round 5-10 mm in diameter glistening translucent effuse filamentous colonies with an alternately smooth and granular surface (due to crossing of fila-



FIG 197—*Clostridium botulinum*  
from a surface agar culture anaerobically  
2 days 37° C ( $\times$  1000)

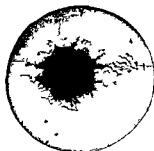


FIG 198—*Clostridium botulinum*  
Surface colony on agar anaerobically  
2 days 37° C ( $\times$  8)

ments) and an indefinite fimbriate reticular edge. Greyish yellow by reflected bluish grey by transmitted light. Butyrous consistency easily emulsifiable. Differentiated into thicker browner centre and thinner more translucent peri-

phery Single colonies often difficult to obtain owing to tendency to spread. Variant types have been described on blood agar (Schoenholz 1928)

**Deep Glucose Agar Slab.**—4 days at 37° C Abundant gas formation, medium disrupted and driven up to plug. Colonies throughout medium, varying in size. Large colonies are 1.2 mm. in diameter, having an opaque, brown, spherical or biconvex centre and a large clear more translucent cigarette-in-water edge. Another type consists of a thin translucent disc with an eccentric opaque nucleus, the edge of the disc being indented at the point nearest the nucleus, the disc may contain gas bubbles. Types C, D and E form woolly colonies without a central nucleus.

**Agar Slope.**—4 days at 37° C Moderate confluent effuse greenish-yellow translucent growth with a very finely granular surface, and an irregular villous edge

**Gelatin.**—7 days at 37° C Completely liquefied

**Broth.**—4 days at 37° C Abundant growth with dense turbidity and a moderate, powdery and granular deposit mostly disintegrating. Rancid odour. Types C, D and E cause little turbidity, but form a flaky deposit, sticking to the sides of the tube.

**Loeffler's Serum.**—15 days at 37° C Moderate growth of small, discrete colonies, digestion by A and B, but not by C and D types.

**Horse Blood Agar Plates.**—3 days at 37° C Irregularly round, 2-3 mm. in diameter umbonate colonies with smooth centre and curled or fimbriate periphery, zone of hæmolysis, sometimes co-extensive with the colony, sometimes larger around the colony, blood is clear transparent and brown.

**Coagulated Egg.**—21 days at 37° C Very poor growth. Butt turned bluish-green, and egg partly digested by A and B but not by C and D types.

**Cooked Meat Medium.**—15 days at 37° C Abundant growth, gas produced, long column of slightly turbid fluid with digested meat beneath, blackening. Putrid odour. No digestion by C, D or E types.

**Resistance.**—Spores are destroyed by dry heat at 180° C. in 5 to 15 minutes. Moist heat at 100° C. destroys them in 5 hours, at 105° C. in 100 minutes, and at 190° C. in 5 minutes (but see p. 1616). Gelatin cultures remain viable for a year or more.

**Metabolic.**—Strict anaerobe. Opt temp. 35° C. Grows well at 20° C.  $\alpha$ -prime hæmolysis on horse blood agar plates. Hæmolysis of human, but not of sheep's red cells. Types A and B are generally proteolytic, digesting gelatin, serum, egg and meat. Types C and D digest gelatin only. Nutritional grows fairly well on ordinary media, growth not improved by glucose, tryptophan and the sporogenes factor are both required. Green fluorescent colonies on MacConkey plates. Powerful exotoxin produced, specific to each type.

**Biochemical.**—Type A gives acid and gas in glucose, maltose and salicin, Types B and C do not ferment salicin. Types A and B ferment glycerol, Type C does not. Indole —, M.R. —, V.P. —, nitrate reduction —,  $\text{NH}_4^+$  —,  $\text{H}_2\text{S}$  —, methylene blue reduction —, catalase —. Litmus milk fine precipitate of casein with almost complete digestion in a fortnight, litmus reduced, reaction alkaline.

**Antigenic Structure.**—Two main types A and B distinguished by their toxin production. Antitoxin to A does not neutralize toxin of B nor vice versa. By agglutination and complement fixation the two types can also be distinguished, and are found to contain 3 or 4 sub-types each. Three other types, C, D and E, have been described forming separate specific toxins.

**Pathogenicity.**—Types A, B and E cause botulism in man. Type C<sub>1</sub> causes limberneck in chickens and ducks, C<sub>2</sub> causes one type of forage poisoning in horses in Australia and U.S.A. C also causes botulism in equines in South Africa, D causes lambsiekte in cattle in South Africa. (For fuller description see Chapter 72.) The organism itself is a saprophyte and does not multiply in the body, it acts entirely by its toxin. Injected subcutaneously a broth culture of Type A or B is fatal to guinea pigs, mice, rabbits, cats, monkeys, and often chickens in 1 to 4 days, symptoms are

muscular paralysis, dilatation of the pupils, shallow breathing, intense salivation, prostration and death. The toxin is the most powerful known, and may kill a mouse in a dose of 0.00001 ml.

(See Kempner 1897, v Hibler 1908, Leuchs 1910, Dickson 1918, Weinberg and Seguin 1918, Burke 1919a, b, Graham and Brueckner 1919, Report 1919, Shuppen 1919, Edmonson *et al* 1920, Orr 1920, 1922, Bengtson 1921, 1922a, b, 1923, 1924a, Nevin 1921, Weiss 1921, Coleman 1922, 1923, Coleman and Meyer 1922, Dubovsky and Meyer 1922a, b, Esty and Meyer 1922, Hall 1922, Meyer and Dubovsky 1922a, b, c, Schoenholz and Meyer 1922, 1924, Seddon 1922, Tanner and Dack 1922, Graham and Boughton 1923a, b, Hall and Davis 1923, Starin and Dack 1923, 1924, 1925, Dozier 1924, Easton and Meyer 1924, Pfenninger 1924, Starin 1924, Wagner 1924, Wheeler and Humphreys 1924, Dickson *et al* 1925, Tanner and Twohey 1926, Theiler *et al* 1926, Weinberg and Ginsbourg 1927, Theiler 1928, Schoenholz 1928, Stark, Sherman and Stark 1928, Robinson 1929, Graham and Thorp 1929, Lommel and Gunnison 1929, Kerrin 1930, Gunnison and Meyer 1930, Sommer and Sommer 1932, Gunnison and Coleman 1932, Fildes 1935, Gunnison, Cummings and Meyer 1936-37, Hazen 1937, 1942)

### *Clostridium oedematiens*

*Isolation*—Described by Weinberg and Seguin in 1915

*Synonyms*—Probably identical with Novy's *B. oedematis malignus* II, or *Cl. novyi* I (Novy 1894), Zeisler and Rasfeld's (1929) *B. gigas*, and Kraneveld's (1930) bacillus of osteomyelitis bacillosa bubalorum. Scott, Turner and Vawter (1933) propose the terms Type A, B and C for the classical *Cl. oedematiens*, *B. gigas* and Kranefeld's bacillus respectively

*Habitat*.—Soil.

*Morphology*.—Rod shaped,  $3.10\ \mu \times 0.8.10\ \mu$ , not unlike *Cl. welchii*, but longer, Types B and C strains may be as large as  $4-20\ \mu \times 1.2\ \mu$ , sides parallel, ends rounded, axis straight or curved, arranged singly, in pairs or chains, jointed filaments not uncommon. Spores formed freely in all media, they are large, oval and subterminal, generally free. Motile by 20 or more peritrichate flagella, but motility is observed only under strictly anaerobic conditions. No capsule. Gram positive in young cultures.



FIG. 199.—*Clostridium oedematiens*

Surface colony on agar anaerobically, 2 days,  $37^{\circ}\text{C}$  ( $\times 8$ )



FIG. 200.—*Cl. oedematiens*

From a surface agar culture anaerobically 2 days,  $37^{\circ}\text{C}$  ( $\times 1000$ )

*Agar Plate*.—4 days at  $37^{\circ}\text{C}$ . Irregularly round colonies, 2-3 mm in diameter, effuse, filamentous or curled, glistening, translucent with finely sponge-like surface and

irregularly lobate edge with very fine dentations, greenish-yellow by reflected, greenish-blue by transmitted light, butyrous and easily emulsifiable.

*Deep Glucose Agar Shake*—4 days at 37° C. Good growth, gas produced, and agar disrupted. Colonies throughout medium, varying in appearance, usually resemble snowflakes—some have an opaque brownish centre with a finely filamentous or fluffy periphery, some have the appearance of a conventional bursting grenade.

*Horse Blood Agar Plates*—3 days at 37° C. Large spreading colonies, as on nutrient agar, and round, slightly umbonate colonies, with entire or undulate edge and a finely granular surface, about 3 mm. in diameter. Zone of  $\beta$ -hemolysis coincident with colony. Centre of colony is more opaque, periphery more translucent.

*Agar Slope*—4 days at 37° C. Thin spreading film of growth, of poor to moderate, partly confluent, slightly raised, glutinous greenish yellow growth with finely granular surface, and an edge made up of single colonies.

*Gelatin*—3 days at 37° C. Liquefaction.

*Broth*—4 days at 37° C. Poor growth with no turbidity and a granulo-powdery deposit, partly disintegrating, slight rancid odour. In glucose broth there is an early turbidity, which clears after a day or two with the deposit of the organisms in a flocculent mass.

*Loeffler's Serum*—15 days at 37° C. Moderate confluent, slightly raised growth, no digestion.

*Coagulated Egg*—15 days at 37° C. No digestion.

*Cooked Meat Medium*—15 days at 37° C. Moderate growth, fluid turbid, gas produced, meat turned slightly pink or bleached, no digestion, rancid odour.

*Persistence*—Destroyed by moist heat at 105° C. in 6 minutes.

*Metabolic*—Strict anaerobe. Opt. temp. 37° C.  $\beta$ -hemolysis on horse blood agar plates. Hemolyses human and sheep red cells. Nutritional grows fairly well in ordinary media, growth improved by glucose. No growth in bile-salt media. Toxin produced. Forms a fibrinolysin.

*Biochemical*—Acid and gas in glucose, and malose, not in mannitol, lactose, sucrose, or salicin. Glycerol fermented by Type A but not by Type B or C strains. Indole —, M.R. —, V.P. —, nitrites not produced in nitrate broth.  $\text{NH}_3$  —,  $\text{H}_2\text{S}$  ±, M.B. reduction —, catalase —. Litmus milk and production in 1 to 5 days with gas formation, after 10 to 30 days a clot appears in the form of fine flocculi. No digestion.

*Antigenic Structure*—Agglutinins act on homologous strains only, auto-agglutination frequent. Antitoxin can be produced by injection of horses.

*Pathogenicity*—Produces a potent exotoxin. One agent in causation of gas gangrene in man. Responsible for one type of braxy in Europe, for black disease in Australia, and for a non-fatal osteomyelitis of the humerus and femur of Dutch East Indian buffaloes. Experimentally it is pathogenic for guinea-pigs, mice and rabbits. 0.2–1 ml. of a 24-hour broth culture injected intramuscularly into a guinea-pig causes death in 24 to 48 hours. P.M. muscles are red and softened, little gas production, but a spreading gelatinous oedema. Bacilli found at site of inoculation, and occasionally on surface of liver. Blood cultures may or may not be positive. (See p. 871.)

(See Weinberg and Seguin 1913, Report 1919, Wolf 1919–20, Hall 1922, Turner 1933, Miesner, Mew and Schoop 1931, Kraneveld 1930, Zessler and Kraneveld 1929, Kraneveld and Djaenodim 1933, Scott, Turner and Vawter 1933, Djaenodim and Kraneveld 1934, Turner and Eales 1941.)

### *Clostridium chauvoei*

*Isolation*—First distinctive description by Arloing, Cornet and Thomas in 1879 (Arloing et al. 1887).

*Habitat*—Lives in soil.

**Morphology**—4-day agar plate at 37° C Rod-shaped,  $3.8 \mu \times 0.6 \mu$ , with parallel sides and rounded ends. Short filaments are not uncommon. On serum or in meat medium navicular and swollen forms are seen. Axis straight or slightly curved, arranged singly, in small groups, or in short chains. Variation in depth of staining, some organisms show chromatic granules near the poles. Spores are elongated oval, subterminal and wider than the bacillus. Clostridial forms are lemon or pear shaped. Motile. Gram positive, but weakly so after 4 days. No capsule. On surface of liver of infected animals it is found singly or in pairs not in chains or filaments like *Cl. septicum*.

**Agar Plate**.—4 days at 37° C Irregularly round, 4-8 mm in diameter granulo-filamentous, effuse, transparent colonies, difficult to see, surface is glistening and very finely granular, edge is fern like and irregularly dentate, greyish by reflected and bluish grey by transmitted light, consistency butyrous easily emulsifiable no differentiation

**Deep Glucose Agar Shake**—4 days at 37° C Abundant growth of discrete colonies throughout medium, except for 3 mm below surface. Colonies are irregularly round 0.5-1 mm in diameter, with an opaque brownish centre and a lighter translucent plumose periphery, with an irregularly erose edge. Moderate gas formation, medium split slightly in 4 or 5 places.

**Horse Blood Agar Plate**—3 days at 37° C Colonies irregularly round 3-6 mm in diameter, effuse, transparent, with granulo-filamentous structure, and entire or rhizoid edge. No definite hæmolytic zone, but plate is cleared slightly

**Agar Slope**—4 days at 37° C Poor, confluent, effuse, transparent growth difficult to see. Surface smooth or very finely granular, edge fern like and irregularly dentate

**Gelatin**—Liquefaction complete in 14 days at 37° C

**Broth**—4 days at 37° C Poor growth, no turbidity, slight powdery deposit disintegrating on shaking, weakly rancid odour. In young cultures a turbidity is noticeable but this clears as the bacilli sediment

**Loeffler's Serum**.—15 days at 37° C Effuse confluent growth, no digestion

**Coagulated Egg**—15 days at 37° C Poor, effuse, confluent growth, no digestion

**Cooked Meat Medium**—15 days at 37° C Very slight or no turbidity, some gas production, meat turned pink. Rancid odour

**Resistance**—Dried on silk threads spores are destroyed by steam in 38 to 48 minutes

**Metabolic**.—Strict anaerobe. On the whole grows poorly. Opt temp 37° C. Very slight hæmolysis on horse blood agar plates. Hæmolyses human and sheep's red cells. Forms a fibrinolysin

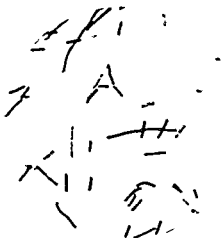


FIG 201—*Clostridium chauvæi*  
From a surface agar culture anaerobically,  
2 days 37° C ( $\times 1000$ )



FIG 202—*Clostridium chauvæi*  
Surface colonies on agar anaerobi-  
cally, 6 days, 37° C ( $\times 8$ )



**Nutritional**—Grows poorly in ordinary media, growth improved by glucose, and by heart extract, very poor growth in casein digest broth. No growth in bile-salt media. Weak toxin produced.

**Biochemical**—Acid and gas in glucose, maltose, lactose, and sucrose, not in mannitol or salicin. Indole—, M.R.—, V.P.—, nitrate reduction +;  $\text{NH}_3$ —,  $\text{H}_2\text{S}$  +, M.B. reduction—, catalase— Litmus milk variable, sometimes no change, sometimes slight acid production, and partial precipitation of casein.

**Antigenic Structure**—Appears by agglutination to be serologically homogeneous. Agglutinating sera prepared against *Cl. chauvoei* agglutinate this organism, but do not agglutinate *Cl. septicum* except to very low titre. By complement fixation *Cl. chauvoei* and *Cl. septicum* appear to be closely related. Antitoxin is specific, protecting against *Cl. chauvoei* but not against *Cl. septicum*.

**Pathogenicity**—Exotoxin produced. Causes blackleg in cattle, and less often in sheep. Non-pathogenic to man. Experimentally it is fatal to guinea pigs and less often to mice, rabbits and pigeons are fairly resistant. 0.25 ml. of a 24-hour culture in Hibler's medium injected intramuscularly kills a guinea pig in 24 to 48 hours, p.m. slightly blood-stained serous exudate at site of injection, abdominal muscles are deep red and contain numerous small gas bubbles. *Cl. chauvoei* can be recovered from local lesion, peritoneal cavity, and heart blood.

(See Kitt 1887, Nocard and Roux 1887, Roux 1888, Kitasato 1890a, 1890, Leclainche and Vallee 1900, Eisenberg 1907, Markoff 1911, Landau 1917, Weinberg and Seguin 1918, Haslam and Lumb 1919, Report 1919, Heller 1920, Goss *et al.* 1921, Gaiger 1922, 1924, Hall 1922, Weinberg and Ginsbourg 1927, Weinberg and Mihaulesco 1929, Roberts 1931, Henderson 1932, Kerrin 1934.)

### *Clostridium septicum*

**Isolation**—Described by Pasteur and Joubert in 1877.

**Synonyms**—*B. adematous malignus*, Koch (1881) *Vibrio septicus*, Pasteur.

**Habitat**—Found chiefly in soil.

**Morphology**—Rod-shaped, of variable length and thickness, on agar cultures,  $2-6 \mu \times 0.4-0.6 \mu$ , sides parallel, ends rounded, axis straight or curved, arranged singly, in pairs and in short chains. On peritoneal surface of dead guinea pig it forms long jointed filaments. In tissue exudates and in fluid media containing fresh tissue there are navicular or citron forms with pale swollen bodies and deeper-staining pointed extremities. In agar cultures there is marked pleomorphism, organisms vary in size, shape, and depth of staining, large numbers of shadow forms are seen. Spores readily formed, and are oval, subterminal, and slightly wider than bacilli, often found free. Motile by 4-16 peritrichate flagella. No capsule. Gram positive in young cultures, but often frankly Gram negative in 4 to 5 days.

**Agar Plate**—4 days at  $37^\circ \text{C}$ . Irregularly round, having a general cigarette-in-water appearance, 10 mm. in diameter, effuse, filamentous, translucent colonies, with finely honeycombed surface due to crossing of numerous filaments, and fimbriate edge, greyish by reflected, bluish-grey by transmitted light, butyrus and easily emulsifiable. No definite differentiation but filaments are less dense at periphery. Recently isolated strains tend to form continuous spreading films.

**Deep Glucose Agar Shake**—4 days at  $37^\circ \text{C}$ . Abundant gas formation, medium disrupted and driven up nearly to plug. Numerous colonies throughout medium, varying in appearance, most usual type is delicate, arborescent, and flocculent, sometimes opaque with an irregularly dentate, well-defined edge, from which later woolly filamentous outshoots appear.

**Horse Blood Agar Plates**—3 days at  $37^\circ \text{C}$ . a prime hæmolysis, after 6 days hæmolysis is of  $\beta$ -type.

FIG 203—*Cl septicum*

From a surface agar culture anaerobically,  
2 days, 37° C. (× 1000)

FIG 204—*Clostridium septicum*

Surface colony on agar anaerobi-  
cally, 2 days, 37° C (× 8)

**Agar Slope.**—4 days at 37° C Scanty to moderate, effuse, translucent, glistening, greyish yellow growth, forming little islands, each with a coarsely erose edge, surface smooth or very finely granular

**Gelatin.**—7 days at 37° C Liquefied.

**Broth.**—4 days at 37° C Poor to moderate growth with slight turbidity and a moderate, powdery deposit disintegrating completely, slight rancid odour

**Loeffler's Serum.**—15 days at 37° C Fairly good confluent growth, no liquefaction.

**Coagulated Egg.**—15 days at 37° C Fairly good, partly confluent growth, with a moderately granular surface, no digestion.

**Cooked Meat Medium.**—15 days at 37° C Moderate growth with slight turbidity, gas production, meat turned pink, no digestion, rancid odour

**Resistance.**—Not recorded.

**Metabolic.**—Strict anaerobe. Opt temp 37° C.  $\alpha$ -prime, and later  $\beta$ -haemolysis on horse blood agar plates. Haemolyses human and sheep's red cells. Nutritional grows fairly well on ordinary media, growth improved by glucose. Green fluorescent colonies on MacConkey plate. Toxin produced. Forms a fibrinolysin.

**Biochemical.**—Acid and gas in glucose, maltose, lactose and salicin, not in mannitol or sucrose. Indole—, M.R.—, V.P.—, nitrates reduced,  $\text{NH}_4$  slight +,  $\text{H}_2\text{S}$  +, M.B. reduction—, catalase—, Litmus milk acid and clot and some gas, the clot does not form for 3 to 6 days

**Antigenic Structure.**—By agglutination four groups can be distinguished on basis of O antigen, further subdivision is possible on basis of H antigen. Some cross-agglutination and much cross-complement-fixation with *Cl chauvoei* strains. Antitoxin appears to be specific

**Pathogenicity.**—Exotoxin produced. One agent in production of gas gangrene in man. Causes blackleg and braxy in sheep, and sometimes blackleg in cattle. Experimentally, it is pathogenic to guinea pigs, mice, rabbits, and pigeons. Pathogenicity is retained for years in subculture. 0.01–0.5 ml. of a 24-hour glucose broth culture injected intramuscularly into guinea pigs causes death in 12 to 24 hours. P.M. blood-stained oedema and gas production, muscles intense deep red in colour and softened, sometimes fluid in peritoneum and pericardium. Motile rods and navicular forms at site of injection, and long jointed snake-like filaments on peritoneal surface of liver (See p 871)

(See von Hübner 1908, Meyer 1915, Weinberg and Soggin 1918, Wolf 1918-19, Report 1919, Heller 1920, Gaiger 1922, 1924, Hall 1922, Weinberg and Gunsbourg 1927, Henderson 1934)

### *Clostridium welchii*

**Synonyms**—*B. aerogenes capsulatus*, *B. phlegmonis emphysematosae*, *B. perfringens*, *B. saccharobutylicus immobilis*, *B. enteritidis sporogenes*, *Granulobacillus butyricus*, *B. cadaveris butyricus*, *B. vaginae emphysematosae*, Achalmé's bacillus.

**Isolation**—First complete description by Welch and Nuttall in 1892, who isolated it from a cadaver. Various organisms closely resembling *Cl. welchii*, but differing from it in type of toxin production, have been isolated from diseased sheep, and called the lamb dysentery bacillus (Dalling 1926), *Cl. paludis* (McEwen 1930), and *Cl. oritoxicum* (Bennett 1932). Exact relation of these organisms to *Cl. welchii* and to one another is doubtful, but Wilsdon's (1931) classification into A, B, C, and D types corresponding respectively to the *welchii*, lamb dysentery, *paludis*, and *oritoxicum* types may be accepted provisionally.

**Habitat**.—Found in soil, water, milk, dust, sewage, and intestinal canal of man and animals.

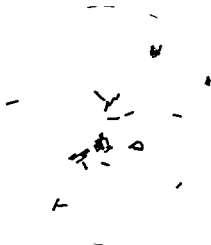


FIG 205.—*Clostridium welchii*

From a surface agar culture, anaerobically, 2 days, 37° C. (× 1000)



FIG 206.—*Clostridium welchii*

Surface colony on agar anaerobically, 5 days, 37° C. (× 8).

**Morphology**—Rather stout rods, varying considerably in length, 4-8  $\mu$  × 0.8-1.0  $\mu$ , sometimes shorter and more slender, filaments not uncommon. Parallel sides, ends truncated or slightly rounded, axis straight. Arranged singly, often side by side forming small bundles. Variation in depth of staining, involution forms—clubs, filaments, tadpoles, granular forms—frequent in old cultures. Spores large, oval, and subterminal. Sporulation occurs more readily with some strains than with others, and is favoured by an alkaline reaction, does not occur below pH 6.6, and hence is unusual in media containing a fermentable carbohydrate. Spores are seldom seen in cultures, but are commonly formed under natural conditions. Non motile. Capsules formed in animal body.

**Agar Plate**.—4 days at 37° C. 2 main types of colony formed. One round, 2-4 mm. in diameter low convex, amorphous, greyish yellow, opaque, with smooth surface and entire edge, butyrous and easily emulsifiable. Other is umbonate, and is differentiated into an opaque brownish centre and a lighter, more translucent,

radially striated periphery with a crenated edge (Fig 206). Other variant forms have been described, differing in morphology, colonial appearance, and sometimes toxicity (Sordell, Prado, and Ferrari 1932, McGaughey 1933, Livesay 1933, Stevens 1935).

**Deep Glucose Agar Shake**—4 days at 37° C. Abundant gas, medium ruptured and driven nearly to plug. Numerous colonies throughout medium, they are biconvex, 1 mm long opaque, with an entire edge.

**Horse Blood Agar**—3 days at 37° C. Round colonies, 2-5 mm in diameter, umbonate greyish white, with opaque raised centre, and a translucent flattened periphery, surface smooth, entire edge. Zone of  $\beta$  haemolysis for 1-3 mm around colony in most strains (see p 861).

**Agar Slope**—4 days at 37° C. Good growth consisting of discrete colonies.

**Gelatin**—2 days at 37° C. Complete liquefaction.

**Broth**—4 days at 37° C. Good growth with moderate turbidity, and moderate powdery deposit, disintegrating completely. Slight sour odour.

**Loeffler's Serum**—15 days at 37° C. Good confluent slightly raised growth with crenated edge, no digestion.

**Congulated Egg**—15 days at 37° C. Fairly good confluent, slightly raised growth, no digestion.

**Cooked Meat Medium**—15 days at 37° C. Good growth, fluid slightly turbid, gas evolved, meat turned pink, no digestion, acid reaction, sour odour.

**Resistance**—Cultures in fermentable carbohydrate media die in a few days owing to the effect of the acid produced. In sugar free protein media in which spores have formed, the organisms may live for months. A suspension containing a million spores per ml. is sterilized in 30 minutes at 90° C. and in 5 minutes or less at 100° C. (Headlee 1931).

**Metabolic**—Fairly strict anaerobe. Opt temp 37° C. Haemolysins and leucocidins are formed by some but not by all strains. Usually gives  $\beta$  haemolysis on horse blood agar plates, haemolyses human and sheep's red cells. **Nutritional** grows fairly well on ordinary media; growth greatly improved by 1 per cent glucose. Green fluorescence on MacConkey plate. Toxin produced. Forms a fibrinolysin.

**Biochemical**—Acid and gas in glucose, maltose, lactose, sucrose and occasionally salicin, not in mannitol, some strains ferment inulin, some glycerol. Indole —, MR +, VP —, nitrates slight reduction,  $\text{NH}_3$  slight +,  $\text{H}_2\text{S}$  ++, MB reduction —, catalase —. Litmus milk: acid, gas, clot—stormy fermentation—occurring in 12 to 48 hours in a proportion of strains. Type A, C, and D strains form acrolein from glycerol.

**Antigenic Structure**—By agglutination no clear cut grouping is apparent, considerable overlapping of antigens. No obvious relation between agglutination results and typing of strains by toxin, antitoxin or biochemical methods. Antitoxic serum can be readily prepared in horses.

**Pathogenicity**—Apparently 7 different exotoxin substances produced, of which up to 5 may be produced by one strain. Chief agent in causation of gas gangrene in man. May play a part in causation of enteritis, appendicitis, and puerperal fever. Causes gas gangrene in animals, especially sheep. Experimentally great



FIG 207—*Clostridium welchii*

Culture in litmus milk anaerobically 24 hours, 37° C., showing stormy fermentation.

variation in pathogenicity of different strains. Washed bacilli or spores are non pathogenic. 0.1-1.0 ml broth culture injected intramuscularly into guinea pig causes local tumefaction, spreading oedema, and death in 24 to 48 hours. P.M. suprarenal glands congested. Also pathogenic to mice, pigeons, and less so to rabbits (see p. 870). B type is responsible for lamb dysentery, C type for "struck"—an enteritis of sheep—and D type for an enterotoxaemic disease and for pulpy kidney disease of sheep (see Chapter 78).

(See Kamen 1904, Sumonds 1915a, b, c, Robertson 1916, Bull 1917, Bull and Pritchett 1917a, b, De Kruij and Bollman 1917, De Kruij *et al.* 1917, Weinberg and Segum 1918, Report 1919, Bengtson 1920, Caulfield 1920, Hall 1922, Humphreys 1924, Dalling 1926, Weinberg and Ginsbourg 1927, Howard 1928, Weinberg 1929, Weinberg and Combresco 1930, McEwen 1930, Torrey, Kahn and Salinger 1930, Headlee 1931, Mason, Ross and Dalling 1931, Wilsdon 1931, 1933, Bennetts 1932, Glenny *et al.* 1933, Walburn and Reymann 1933, McGaughey 1933, Lavesay 1933, Weinberg and Guillaume 1936.)

### *Clostridium tetani*

*Isolation*—Described by Nicolaier in 1834, isolated by Kitasato in 1889 (1889b).

*Habitat*.—Found in soil—especially cultivated soil—and in the intestine of man and animals.

*Morphology*—4-day agar slope at 37° C. Rods, 2-5  $\mu \times 0.5 \mu$ , considerable variation in length, long curved, filamentous forms are not uncommon. Axis straight sides parallel, ends rounded, arranged singly and occasionally in chains. Variation in depth of staining. Spores spherical, terminal, and wider than the bacillus giving characteristic drum-stick appearance. Sluggishly motile, peritrichate flagella. No capsule. Strongly Gram-positive in young cultures. In early stages spores stain solidly, later only the thin wall stains, spores rarely become free. Bizarre involution forms appear in old cultures.

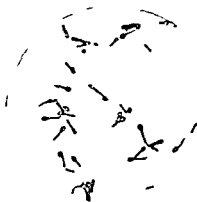


FIG 208—*Clostridium tetani*.

From a surface agar culture anaerobically, 7 days, 37° C., showing ring form of stain ( $\times 1000$ ).



FIG 209—*Clostridium tetani*.

From a broth culture anaerobically 7 days, 37° C., showing the spores stained solidly ( $\times 1000$ ).

*Agar Plate*.—4 days at 37° C. Irregularly round, 2-3 mm. in diameter, effuse, glistening translucent, greyish yellow colonies with irregularly granular surface, and ill-defined edge, showing filamentous, curled projections, structure very finely granular or filamentous, butyrous consistency, emulsifying easily. Some strains form colonies differentiated into thicker translucent, yellowish brown centre, and thinner, transparent, almost colourless periphery. Whole colony has

a fuzzy appearance. Isolated colonies of the normal motile type of *Cl. tetani* are extremely difficult to obtain owing to the tendency of the organisms to spread in a proteus like or a delicately rhizoidal film over the surface of the agar but it is stated that non motile variants give rise to separate discrete colonies even on moist media.

**Deep Glucose Agar Slake**—4 days at 37° C. Slight gas formation, medium disrupted in 2 or 3 places. Colonies scattered throughout medium except at surface, they are rounded 1-2 mm in diameter with sometimes a brownish opaque centre and a lighter more translucent periphery, fuzzy filamentous branching edge.

**Agar Slope**—4 days at 37° C. Scanty to moderate greyish yellow almost transparent effuse growth stretching to sides of tube with very finely granular gl. staining surface. If inoculated into condensation water spreads rapidly over the whole surface of the agar and presents a filamentous edge at the top.

**Gelatin Slab**—10 days at 23° C. Poor growth of fir tree appearance. At 37° C. liquefies gelatin in a week.

**Broth**—4 days at 37° C. Poor to moderate growth with slight turbidity and a finely granular deposit not disintegrating on shaking. Odour of manure.

**Loeffler's Serum**—4 days at 37° C. Poor effuse mostly discrete growth, serum may be softened or disintegrated after several days but no true liquefaction occurs.

**Horse Blood Agar Plates**—3 days at 37° C. Rounded, 2-5 mm. in diameter effuse, greyish translucent colonies with finely granular surface and lobate or fimbriate edge. Sometimes zone of a prime hæmolytic diam. 1-5 mm., later passing into the  $\beta$  type.

**Congl. Egg**—4 days at 37° C. Poor effuse partly confluent growth, no digestion.

**Cooked Meat Medium**—4 days at 37° C. Good growth, fluid shows moderate turbidity, some gas production, no blackening and no digestion of meat. Slight blackening is said to occur after some weeks.

**Resistance**.—Spores resist boiling for 15 to 90 minutes. Killed at 105° C. in 3 to 25 minutes, killed by 5 per cent phenol in 15 hours.

**Metabolic**.—Strict anaerobe. Opt temp 37° C. Grows poorly at 20° C. a prime or  $\beta$ -hæmolytic on blood agar plates. Hæmolytic acts on R.B.C. of rabbit, goat, sheep, horse and many other animals. *Nutritional* grows fairly well on ordinary media, growth improved by blood and serum, not by glucose. Tuller's exhausted medium is particularly suitable. Produces green fluorescence on MacConkey's agar. Possesses slight proteolytic properties.

**Biochemical**.—Ferments no sugars. Indole + M.R. — V.P. — nitrites not produced in nitrate broth.  $\text{NH}_3$  slight +  $\text{H}_2\text{S}$  — M.B. reduction — catalase — Litmus milk, reduction of litmus usually slight precipitation of casein, sometimes no change. Said to form phenol.

**Antigenic Structure**.—Differentiated by agglutination into at least 10 types of which Types I and III appear to be the commonest in this country. Toxin formed by all types is identical and is neutralized by antitoxin prepared against any one type. Anti-bacterial sera contain agglutinins and opsonins specific to each type.

**Pathogenicity**.—Potent exotoxin formed. Naturally pathogenic to man and horses in particular. Experimentally mice, guinea pigs and rabbits are susceptible, dying of tetanus in 1 to 4 days after subcutaneous injection of a broth culture. Washed spores or bacilli are innocuous. The toxin is very potent and will kill a mouse in a dose of 0.0001 ml. Birds are resistant. (See Chapter 77.)



FIG. 210.—*Clostridium tetani*. Surface colony on agar anaerobically 2 days 37° C. showing parent colony surrounded by an effuse outgrowth ( $\times 8$ ).

(See Rosenbach 1886, Behring and Kitasato 1890, Kitasato and Weir 1890, Kitasato 1891, Toledo and Veillon 1891, Behring 1892, Vaillard and Rouget 1892, Ushinsky 1893, Pizzini 1898, Madsen 1899, Ritchie 1901, Rosenau and Anderson 1908, von Hübner 1903, Metchnikoff 1908, Smith 1908, Noble 1915, Corbitt 1918, Weinberg and Seguin 1918, Report 1919, Tulloch 1919, Hall 1922, Heller 1922a, b, Anderson 1924, Bengtson 1924b, Descombes 1924, Fildes 1925a, b, Weinberg and Ginsbourg 1927, Tschertkow 1929, Fildes 1929, Knight and Fildes 1930, Kerrin 1930, 1934, Belm 1931, Mutermilch et al 1933.)

Notes on Less Important Strains (see Table 57).

*Cl. aerofortidum*—Described by Weinberg and Seguin (1918). Small slender bacillus, 3–5  $\mu$  long, slightly motile, and weakly Gram positive. Spores are subterminal, and are not readily formed. Surface colonies are round and transparent, deep colonies are small and irregular. Non pathogenic to guinea pigs.

*Cl. bifermentans*—Isolated in 1902 by Tissier and Martelly from putrefying meat, and named *B. bifermentans sporogenes*. So called from its being the first anaerobe shown to decompose both sugars and proteins. Probably identical with *Cl. centrosporogenes* described by Hall (1922), McCoy and McClung (1936) and Clark and Hall (1937), and, according to Stewart (1933), with *Cl. sordellii* (synonym *B. oedematis sporogenes*) isolated by Sordelli (1922, 1923) in Buenos Aires from a human case of gas gangrene (see also Weinberg, Davesne and Lefranc 1931, Hall and Scott 1937). Gram positive rod 3.6–6.0  $\mu \times 1.2$ –1.5  $\mu$ , motile by peritrichate flagella, readily forming central or subterminal oval spores. Uniform turbidity in broth with filament production, and viscous deposit, which is easily disintegrated on shaking, nauseous odour. Surface colonies on horse blood agar are round, crenated, or irregular, and occasionally hemolytic, deep colonies are round or irregular and opaque with a woolly periphery. Gelatin liquefied rapidly. Coagulated serum disintegrated or liquefied. Cooked meat medium digested, blackened, with a late deposit of small rounded masses of white crystals. Ferments glucose and maltose but not lactose, saccharose, or mannitol. Forms a fibrinolysin. Pathogenic strains (equivalent to *Cl. sordellii*) produce a fairly powerful toxin which is destroyed by 30° C in 60 minutes. Both toxigenic and non toxigenic strains produce a lecithinase, specifically neutralized by the  $\alpha$  antitoxin of *Cl. welchii* (Harward 1943). 0.01 ml. of a 48-hour glucose broth culture of a pathogenic strain injected intramuscularly kills a guinea-pig in 2 days with lesions similar to those caused by *Cl. oedematis*. Appears to be morphologically, culturally and antigenically more or less identical with *Cl. oedematis*, an organism isolated from a wound infection by Meloney, Humphreys and Carp (1926–27, 1927). (See Humphreys and Meloney 1927–28, Hall, Rymer and Jungherr 1929, Hall and Scott 1931, Levenson 1933.)

*Cl. capitorale*—Isolated by Snyder and Hall (1935) from various situations including the pleural fluid of a sheep that had died of gas gangrene, the heart blood and peritoneal fluid at post mortem of cases of septic infection in human beings, and the faeces of normal infants. It is a slender, motile, Gram positive rod with rounded ends, measuring 2.0–2.5  $\mu \times 0.5$ –0.8  $\mu$ , and forming terminal oval spores. It gives rise on blood agar to minute circular or irregularly circular, transparent, non hemolytic surface colonies. It produces acid and gas in glucose, but not in maltose, mannitol, lactose, sucrose, or salicin. It liquefies gelatin, is mildly proteolytic, clots milk irregularly and sometimes digests the clot, is non pathogenic to guinea-pigs and rabbits inoculated subcutaneously, and appears to be antigenically homogeneous. For its differentiation from *Cl. paraputrificum*, *Cl. calorolerans* and *Cl. cochlearium*, see Snyder and Hall (1935). Serologically distinct from *Cl. paraputrificum* (Snyder 1936).

*Cl. carnis*—Described by Klein (1903). A similar bacillus, pathogenic for rabbits "von Hübner vi," was isolated from soil by von Hübner (1903). Hall and Duffett (1933) consider the two to be identical. Gram positive rod, sluggishly motile by peritrichate flagella, 1.5 to 4.5  $\mu \times 0.5$ –0.7  $\mu$ . Readily forms subterminal elongated spores slightly wider than the bacillus, which later appear to be terminal. Microaerophilic. Surface

colonies round and transparent, later flat and lobate, deep colonies lenticular. Uniform turbidity in broth, and later, granular, then mucoid deposit. Ferments glucose, maltose, lactose, saccharose and salicin, but not mannitol. Litmus milk unchanged. Coagulated serum and gelatin not liquefied. Indole —. Produces a soluble exotoxin. Pathogenic to mice, white rats and rabbits, with production of an oedematous and congested local lesion.

*Cl cochlearium* — Described by McIntosh (1917) as Type III C, and named *B cochlearis* by Douglas, Fleming and Colebrook (1920) on account of its likeness to a spoon. Its claim to the rank of a species confirmed by MacLennan (1939). Slender rod, Gram positive only in young cultures,  $3.5 \mu \times 0.5-0.6 \mu$ , actively motile by peritrichate flagella. Spores formed late and in small numbers, are oval, terminal and twice the width of the bacillus. Surface colonies are translucent, round, with delicately crenated edge, deep colonies are lenticular, and may develop polar tufts of growth. Non-haemolytic, non-saccharolytic, non-proteolytic. Coagulated serum and gelatin not liquefied.  $\text{NH}_3$  —, indole —, VP —, MR —, MB reductase —, catalase —,  $\text{H}_2\text{S}$  +. Antigenically homogeneous. Non-pathogenic to guinea pigs.

*Cl difficile* — Described by Hall and O'Toole (1935) and Snyder (1937). Isolated from the faeces of normal infants. Strictly anaerobic large Gram positive rod with subterminal elongated spores of about the same width as the bacillus. Colonies irregular, flat, rough and non-haemolytic. Ferments glucose, levulose, xylose, salicin, mannose and mannitol. Does not ferment galactose, lactose, saccharose, raffinose, inulin or glycerol.  $\text{H}_2\text{S}$  ±, Indole —, coagulated serum not liquefied, late liquefaction of gelatin by a few strains. Some strains produce a filtrable, thermolabile antitoxinogenic toxin which induces local oedema and convulsions in guinea pigs. The toxin is lethal on injection into the cat, dog, rat, guinea pig, rabbit and pigeon but has no effect by mouth in the rat, guinea pig and dog. The bacteria are antigenically heterogeneous, the toxin apparently antigenically homogeneous.

*Cl fallax* was found by Weinberg and Séguin (1918) in infected wounds and gas gangrene. It is a motile, Gram positive bacillus,  $1.2-5 \mu \times 0.6 \mu$  in diameter, with rounded ends, straight axis arranged singly. Spores, which are rarely formed, are subterminal or central oval and wider than the bacillus. Surface colonies round and transparent, later raised, irregular and more opaque, deep colonies lenticular, irregular or bean-shaped. Uniform turbidity in broth, later granular, then mucoid deposit. Ferments glucose, maltose, lactose, saccharose and salicin, mannitol fermentation variously reported. Litmus milk, coagulation in 7 days. Coagulated serum and gelatin not liquefied. Indole —, (see Duffett 1935). Produces a soluble toxin. When freshly isolated is pathogenic for mice and guinea pigs.

*Cl hastiforme* isolated by Cunningham (1930-31, 1931) and by MacLennan (1939), who accorded it the rank of a species. A Gram positive rod,  $2-6 \mu \times 0.3-0.6 \mu$ , sluggishly motile by peritrichate flagella. Readily produces oval subterminal spores which together with a minute terminal tip of bacillary protoplasm, constitute the spear head shape from which the organism is named. Surface colonies are non-hemolytic, minute, transparent and round, later becoming irregular, deep colonies round and coarsely filamentous. Gelatin liquefied 7-10 days, coagulated serum not liquefied. Non-saccharolytic.  $\text{H}_2\text{S}$  —, Indole —,  $\text{NH}_3$  —. Antigenically homogeneous. Hayward (personal communication) has met with strains resembling *Cl hastiforme* which are haemolytic on horse blood agar.

*Cl haemolyticum* — Described by Vawter and Records (1926, 1931) as the cause of red water disease or bacillary haemoglobinuria of cattle in the United States and by Sordelli, Ferrari, and Prado (1930) in South America. Named *B haemolyticus* by Hall (1929b). It is a fairly large bacillus,  $3.0-5.6 \mu \times 1.0-1.3 \mu$ , occurring individually, in pairs, and occasionally in short chains. Spores are oval, terminal or subterminal. Sluggishly motile by 6-16 peritrichate flagella. Gram positive in young cultures. Deep colonies in agar are at first lenticular, later fluffy. Gelatin is liquefied, but otherwise no



proteolytic action. Acid and gas produced in glucose, acid and clot in litmus milk. Complete hemolysis on blood agar plates in 24 hours. A toxin is produced. By agglutination most strains appear to be antigenically homogeneous. Intramuscular inoculation of guinea-pigs with a toxic culture gives rise to an extensive bloody edema sometimes accompanied by hemoglobinuria. Rabbits also susceptible. A disease simulating in many respects the natural disease can be reproduced in cattle by injection of toxic cultures.

*Cl. multifermentans* (tenalium).—Described by Stoddard (1919a), isolated from a case of gas gangrene. Resembles *Cl. septicum* morphologically, especially in its formation of citrons, but is non pathogenic. Gram positive, motile bacillus with subterminal spores. Surface colonies are large, round with slightly irregular edges, after several days they become white and opaque, and rise up from the surface. Deep colonies are white and opaque, irregular or biconvex, with projecting outgrowths. Acid and gas in glucose, maltose, lactose, sucrose, and salicin, acid and clot in L.M. Non proteolytic. Non pathogenic to guinea pigs.

*Cl. putrificum*.—Described by Bienstock (1884, 1899, 1901), who isolated it from faeces. Appears to have been a slender Gram positive rod with spherical or oval terminal spores, which digested proteins but had no action on carbohydrates. Its identity has been in doubt (for critical discussion see Hall and Snyder 1934, Hartsell and Rettger 1934, Morgan and Wright 1934). Many workers regard it as identical with *Cl. cochlearium*, but this is disputed by Hartsell and Rettger and by MacLennan (1939). According to MacLennan, *Cl. putrificum* is a slender bacillus, Gram positive only in young cultures, 0.3–0.5  $\mu$  in diameter, forming long tangled threads in old cultures, with slowly developing, large round terminal spores. Sluggishly motile by peritrichous flagella. Colonies are irregularly round, transparent, with filamentous or delicately fimbriate edge, non hemolytic. Deep colonies are minute, spherical and hairy. Non-saccharolytic,  $H_2S$  +, M.R. —, M.B. reduction —, catalase —, liquefies coagulated serum and gelatin in 7–20 days and blackens cooked meat medium slightly. Three serological groups, which are distinct from *Cl. cochlearium*.

*Cl. paraputrificum*.—According to Hall and his colleagues (Hall and Snyder 1934, Snyder 1936, Hall and Ridgeway 1937) this organism, which was described by Bienstock (1906) is probably identical with Escherich's "Kopfenbakterien," von Hübner's ix bacillus, Rodella's ix bacillus, and Kleinschmidt's (1934) *B. munitus*. It is found in the faeces particularly of infants, both normal and ill nourished, and is a slender, motile, Gram variable bacillus with terminal oval spores. It is non proteolytic, it ferments glucose, maltose, lactose, sucrose, and salicin, but not mannitol or xylose, with the production of acid and gas. It is non pathogenic for guinea-pigs and rabbits.

*Cl. parasporogenes*.—Described by McIntosh (1917). Resembles *Cl. sporogenes*, but deep colonies in agar shake cultures are biconvex or irregular in shape. Also forms specific agglutinins which do not act on *Cl. sporogenes*. Non pathogenic to guinea pigs.

*Cl. sphenoides*.—Isolated by Douglas, Fleming and Colebrook (1920) from wounds. So called from the wedge-shape of the sporing bacillus. Small, motile, weakly Gram positive, vegetative bacilli are fusiform in shape and arranged in pairs end to end. Spores are large and round, appear subterminally, but soon become strictly terminal. Surface colonies are round with entire edges. Pathogenicity not examined.

*Cl. tertium*.—Described by Henry (1917). Resembles, but is probably different from, *Cl. paraputrificum* (see above). Thin, slightly curved bacillus, 3–5  $\mu$  long, sluggishly motile. Gram positive, often showing granular staining. Spores freely, giving rise to large, oval, elongated terminal spores. Surface colonies are rounded, delicate, indescant, and almost transparent, with entire or slightly crenated edge. Deep colonies are small, biconvex or irregular in shape. Ferments mannitol and xylose. Non pathogenic to guinea pigs. (See von Hübner 1908, Hall and Matsumura 1924). According to Hall and Duffett (1935) both *Cl. tertium* and *Cl. histolyticum* are microaerophilic rather than strictly anaerobic, but spores are formed only under anaerobic conditions.

*Cl. tetanomorphum*—Described by McIntosh (1917) as Type IX. Morphologically resembles *Cl. tetani*, but differs in cultural and biochemical characteristics, fails to form a specific toxin. Surface colonies are small, flat irregularly round, and almost transparent, deep colonies are small and irregular in shape, but are not woolly or branched. Non pathogenic to guinea pigs.

## REFERENCES

- ADAMSON R S (1919-20) *J Path Bact*, **23**, 241  
 AITKEN, R. S. BARLING, B. and MILES A A (1936) *Lancet*, ii 780  
 ANDERSON B G (1924) *J infect Dis*, **35**, 213, 244  
 ARLOING, S., CORNETT, C E. and THOMAS, O (1887) "Le charbon symptomatique du bœuf, 2nd edition Paris  
 BEHRING (1892) *Z Hyg InfektKr*, **12**, 45  
 BEHRING and KITASATO (1890) *Dtsch med Wschr*, **16**, 1113  
 BELIN, M (1931) *C R Soc Biol*, **105**, 840  
 BENNETTSON, I A (1920) *Bull U S Hyg Lab*, No 122, (1921) *Publ Hlth Rep Wash* No 29, p 1665, (1922a) *Publ Hlth Rep, Wash*, **37**, 164, (1922b) *Ibid* **37**, 2252, (1923) *Ibid*, **38**, 340; (1924a) *Bull U S Hyg Lab*, No 136 (1924b) *Ibid* No 139  
 BENNETTS, H W (1932) *Aust Couns sci industr Res*, Bull No 57  
 BEENHEIMER A W (1944) *J exp Med*, **80**, 309, 321, 333  
 BIENSTOCK, B (1884) *Z klin Med*, **8**, 1, (1899) *Arch Hyg*, **36**, 335, (1901) *Ibid*, **39**, 390, (1906) *Ann Inst Pasteur* **20**, 407  
 BORTHWICK, G R (1935) *Zbl Bakt*, **124**, 289, (1937) *Brit J exp Path* **18**, 475  
 BOSWORTH T J and GLOVER, R E (1935) *Proc R Soc Med*, **28**, 1004  
 BULL, C G (1917) *J exp Med*, **26**, 603  
 BULL, C G and PRITCHETT, I W (1917a) *J exp Med*, **26**, 119, (1917b) *Ibid*, **26**, 867  
 BURKE, G S (1919a) *J Bact*, **4**, 555 (1919b) *J Amer med Ass*, **72**, 88  
 CARLEN S A (1939) *Proc Soc exp Biol N Y*, **40**, 39  
 CAULFIELD, A H W (1920) *J infect Dis*, **27**, 151  
 CLARK, F L and HALL, I C (1937) *J Bact*, **33**, 23  
 CLIFTON, C E (1942) *J Bact*, **44**, 179  
 COLEMAN, G E (1922) *J infect Dis*, **31**, 556, (1923) *Ibid*, **33**, 384  
 COLEMAN G E and MEYER, K F (1922) *J infect Dis*, **31**, 622  
 CORBITT H B (1918) *Bull U S Hyg Lab*, No 112  
 CROOK, E M (1942) *Brit J exp Path*, **23**, 37  
 CONNINGHAM A (1930-31) *Zbl Bakt IIte Abt* **82**, 25 481, (1931) *Ibid* **83**, 1 22 219  
 DACK G M, STARIN W A. and WERNER M (1927) *J infect Dis*, **40**, 525  
 DALLING, T (1906) *J comp Path* **39**, 148  
 DALLING, T and ROSS H E. (1934) *J comp Path*, **51**, 235  
 DALLING T and STEPHENSON, M (1942) *Nature Lond*, **149**, 56  
 DESCOMBEY, P (1924) *C R Soc Biol*, **91**, 239  
 DICKENS P F (1934) *U S nav med Bull* **32**, 267  
 DICKSON, E C (1918) *Monogr Rockefeller Inst med Res*, No. 8  
 DICKSON, E C, BURKE G S BECK D, and JOHNSTON, J (1925) *J infect Dis*, **36**, 472  
 DIJONHOEDIN R and KRANEVELD, F C (1936) *Ned Ind Bl Diergeneesk*, **48**, 290  
 DOUGLAS, S R, FLEMING A, and COLBROOK, L (1920) *Spec Rep Ser med Res Coun*, Lond No 57  
 DOZIER, C C (1924) *J infect Dis*, **35**, 105  
 DUBOVSKY B J and MEYER K F (1922a) *J infect Dis*, **31**, 501, (1922b) *Ibid*, **31**, 595  
 DUFFETT N D (1935) *J Bact*, **29**, 573 (1938) *Univ Colo Stud*, **28**, 46  
 EASTON, L J and MEYER, K F (1924) *J infect Dis*, **35**, 207  
 EDMONDSON, R B, GILTNER, L T, and THOM C (1920) *Arch intern Med*, **26**, 357  
 EISENBERG, P (1907) *C R Soc Biol*, **62**, 613  
 LEMNGEM, E VAN (1895) *Zbl Bakt* **19**, 442, (1897) *Z Hyg InfektKr*, **26**, 1  
 ESTY, J R and MEYER K F (1922) *J infect Dis*, **31**, 650  
 EVANS, D G (1945) *J Path Bact*, **57**, 75  
 FELIX, A and ROBERTSON, M (1928) *Brit J exp Path*, **9**, 6  
 FIELDS P (1925a) *Brit J exp Path*, **6**, 62, (1925b) *Ibid*, **6**, 91, (1929) *Brit J exp Path*, **10**, 151, (1935) *Brit J exp Path*, **16**, 309  
 FIELDS P and KNIGHT B C J G (1933) *Brit J exp Path*, **14**, 343  
 FIELDS P and RICHARDSON G M (1935) *Brit J exp Path*, **16**, 326  
 FRANKLAND, P F (1889) *Z Hyg InfektKr* **8**, 13  
 FRIEDMANN, U and HOLLANDER A (1943) *J Immunol* **47**, 23, 29  
 GAIGER, S H (1922) *J comp Path*, **35**, 191, 235, (1924) *Ibid*, **37**, 163

- GALE, E F (1940) *Bact Rev.* 4, 135  
 GALE, E F and HEYNINGEN W E VAN (1942) *Biochem J.* 36, 624  
 GILL, D A (1933) *I et J.* 89, 399  
 GILLESPIE R W II and RETTOER, L F (1939) *J Bact.* 36, 605  
 GLADSTONE, G P, FILDES, P. and RICHARDSON G M (1935) *Brit J exp Path.* 16, 335  
 GLENNY, A T, BARR M., LLEWELLYN-JONES, M., DALLING, T., and ROSS H E (1933) *J Path Bact.* 37, 53  
 GOSS, L. W., BARBARIN, R E., and HAINES, A. W (1921) *J infect Dis* 29, 615  
 GRAHAM, R. and BOUGHTON, T B (1923a) *Abstr Bact.* 1, 29, (1923b) *Ibid.* 7, 30  
 GRAHAM, R. and BRUECKNER, A. L (1919) *J Bact.* 4, 1  
 GRAHAM, R. and THORP, F (1929) *J Immunol.* 18, 391  
 GUGGENHEIM, K. (1944) *J Bact.* 47, 313  
 GUNNISON, J B (1937) *J Immunol.* 32, 63  
 GUNNISON, J B and COLEMAN, G E (1932) *J infect Dis* 51, 542  
 GUNNISON, J B, CUMMINGS J R., and MEYER, K. F (1936-37) *Proc. Soc exp Biol. N Y.* 35, 278  
 GUNNISON, J B and MEYER K. F (1930) *J infect Dis* 46, 335  
 HAINES, R B. and SCOTT, W J (1940) *J Hyg., Camb.* 40, 154  
 HALL, I C (1922) *J infect Dis* 30, 445, (1929a) *J Bact.* 17, 255, (1929b) *J infect Dis* 45, 156  
 HALL, I C. and DAVIS, N C (1923) *J exp Med.* 37, 585  
 HALL, I C and DUFFETT, N D (1935) *J Bact.* 29, 269  
 HALL, I C and MATSUMA, K. (1924) *J infect Dis.* 35, 502  
 HALL, I C and RIDGWAY, D (1937) *J Bact.* 34, 631  
 HALL, I C, RYMER, M R., and JUNGHER, E (1929) *J infect Dis.* 45, 42  
 HALL, I C. and SCOTT, A. L. (1927) *J infect Dis.* 41, 329, (1931) *J Bact.* 22, 375  
 HALL, I C. and SYDER, M. L. (1934) *J Bact.* 23, 181  
 HALL, I C and OTOOLE, E (1935) *Amer J Dis Child.* 49, 390  
 HARTSELL, S E and RETTOER, L F (1934) *J Bact.* 27, 497  
 HASLAM T P and LUMB, J W (1919) *J infect Dis.* 24, 363  
 HAYWARD N J (1941) *Brit med J.* 1, 811, (1943) *J Path. Bact.* 55, 235  
 HAZEN, E. L. (1931) *J infect Dis.* 60, 260, (1942) *Proc. Soc. exp Biol. N Y.* 50, 112  
 HEADLEE, M. R (1931) *J infect Dis.* 48, 468  
 HELLER, H H (1920) *J infect Dis* 27, 385, (1921) *J Bact.* 6, 521, (1922a) *J infect Dis.* 30, 18, (1922b) *Ibid.* 30, 33  
 HENDERSON, D W (1932) *Brit. J exp Path.* 13, 412, (1934) *Ibid* 15, 166, (1940) *J Hyg., Camb.* 40, 501  
 HENRIKSEN, S D (1937) *Acta. path. microbiol. Scand.* 14, 570  
 HENRY, H (1917) *J Path. Bact.* 21, 344  
 HEYNINGEN, W E VAN (1941a) *Biochem J* 35, 1246 (1941b) *Ibid.* 35, 1257  
 HIEBER, E VON (1908) "Untersuchungen über die pathogenen Anaeroben." Jena.  
 HOLKER, J (1918-19) *J Path. Bact.* 22, 28  
 HOOGERHEIDE J C. (1937) *J Bact.* 34, 397  
 HOWARD A. (1928) *Ann Inst Pasteur.* 42, 1403  
 HOYT, A, CHANEY, A L., and CAVELL, K. (1938) *J Bact.* 36, 639  
 HUMPHREYS F and MELENEY, F L (1927-8) *Proc Soc exp Biol. N Y.* 25, 611  
 HUMPHREYS F B (1924) *J infect Dis.* 35, 282  
 IPSEN, J (1939) *Bull Hlth. Org., L.N.* 8, 825, (1940-41) *Ibid.* 9, 447, 452  
 IPSEN J and DAVOLL, R (1939a) *Bull Hlth Org L.N.* 8, 833  
 IPSEN J, SMITH, M L., and SORDELLI, A (1939b) *Ibid.* 8, 797  
 KAMEY, L. (1904) *Zbl Bakt.* 35, 686  
 KEMPNER, W (1897) *Z Hyg InfektKr.* 26, 481  
 KERRIN, J C (1930) *Brit J exp Path.* 11, 153, (1934) *J Path Bact.* 38, 219  
 KITASATO, S (1889a) *Z Hyg InfektKr.* 6, 105, (1889b) *Ibid.* 7, 225, (1890) *Ibid.* 8, 55, (1891) *Ibid.* 10, 267  
 KITASATO, S and WEYL, TH. (1890) *Z Hyg InfektKr* 8, 41, 404  
 KITT T (1887) *Zbl Bakt.* 1, 684 716 741  
 KLEIN E. (1903) *Zbl Bakt* 35, 459  
 KLEINSCHMIDT H (1934) *Möchr Kinderheilk.* 62, 14  
 KNIGHT, B C J G (1936) *Spec. Rep Ser med Res Coun., Lond.* No. 210  
 KNIGHT, B C J G and FILDES, P (1930) *Biochem J.* 24, 1496, (1933) *Brit J exp. Path.* 14, 112  
 KOCH R (1881) *Mitt Reichsgesundh.Amt.* 1, 49  
 KRANEVELD, F C. (1930) *Ned Ind Bl. Diergeneesk.* 42, 564  
 KRANEVELD F C. and DUAENOEDIN P (1933) *Ned Ind Bl. Diergeneesk.* 45, 80  
 KREUZER, E (1939) *Z Immunforsch.* 95, 345  
 KRUUF, P H DE, ADAMS, T W., and IRELAND, P M (1917) *J infect Dis.* 21, 580  
 KRUUF, P H DE and BOLLMAN, J L. (1917) *J infect. Dis.* 21, 683.

- LAIDLAW, P P (1915) *Brit med J*, 1, 497.
- LANDAU, H (1917) *Zbl Bakt*, 79, 417
- LECLAINCHE, E and VALLÉE, H (1900) *Ann Inst Pasteur*, 14, 202
- LEPPER, E and MARTIN, C J (1929) *Brit J exp Path*, 10, 327, (1930) *Ibid*, 11, 137, 140
- LEUCHS, J (1910) *Z Hyg InfektKr*, 65, 55
- LEVENSON, S (1936) *C R Soc Biol*, 121, 221
- LIEBOWITZ, P (1886) *Z Hyg InfektKr*, 1, 115
- LINGKISHEIM, von. (1912) See Kollé and Wassermann's "Hdb path Mikroorg" 11te Abt (1912-13), 4, 737
- LIVESAY, H R (1933) *J infect Dis*, 53, 125
- LOMMEL, J and GUNNISON, J B (1929) *J Immunol*, 16, 403
- MCCLEAN, D (1936) *J Path. Bact*, 42, 477
- MCCLEUNG, L S (1935) *J Bact*, 29, 189, (1937) *J infect Dis*, 60, 122
- MCCLEUNG, L S., MCCOY, E., and FRED, E B (1935) *Zbl Bakt*, 11te Abt, 91, 225
- MCCLEUNG, L S and WHEATON, E (1936) *Food Research*, 1, 307
- MCCOY, E (1937) *J Bact*, 34, 321
- MCCOY, E and McCLEUNG, L S (1936) *J Bact*, 31, 557, (1938) *Bact Rev*, 2, 47
- McEWEN, A. D (1926) *J comp Path*, 39, 253, (1930) *Ibid*, 43, 1
- MACFARLANE, M G and KNIGHT, B C J G (1941) *Biochem J*, 35, 884
- MACFARLANE, R G., OAKLEY, C L., and ANDERSON, C G (1941) *J Path. Bact*, 52, 99
- McGaughey, C A (1933) *J Path Bact*, 36, 263
- McINTOSH, J (1917) *Spec. Rep Ser med Res Coun., Lond*, No 12
- McINTOSH, J and FIELDS, P (1916) *Lancet*, i, 768
- MACLENNAN, J D (1939) *Brit J exp Path*, 20, 371
- MADSEN, T (1899) *Z Hyg InfektKr*, 32, 214
- MARKOFF, W N (1911) *Zbl Bakt*, 60, 183
- MASON, J H (1935) *Onderstepoort J vet Sci*, 5, 363
- MASON, J H., ROSS, H E., and DALLING, T (1931) *J comp Path*, 44, 258
- MEISEL, H (1938) *Z ImmunForsch*, 82, 79
- MELNEY, F L., HUMPHREYS, F B., and CARP, L. (1926-7) *Proc Soc exp Biol, N Y*, 24, 675, (1927) *Surg Gynec Obstet*, 45, 775
- METCHNIKOFF, E (1908) *Ann Inst Pasteur*, 22, 929
- MEYER, K. F (1915) *J infect Dis*, 17, 458
- MEYER, K. F and DUBOVSKY, B J (1922a) *J infect Dis*, 31, 541, (1922b) *Ibid*, 31, 559, (1922c) *Ibid*, 31, 600
- MIESZNER, H., MEYER, A., and SCHOOF, G (1931) *Zbl Bakt*, 120, 258
- MILES, A A and HAYWARD, N J (1943) *Lancet*, ii 116
- MORGAN, E L and WRIGHT, H D (1934) *J Path Bact*, 39, 457
- MUTERMILCH, S., BELIN, M., and SALAMON, E (1933) *C R Soc Biol*, 114, 1005
- NAGLER, F P O (1936) *Z ImmunForsch*, 89, 477, (1937) *Ibid*, 91, 457, (1939) *Brit J exp Path*, 20, 473, (1940) *Z ImmunForsch*, 97, 273
- NEVIN, M (1921) *J infect Dis*, 28, 226
- NICOLAÏER (1884) *Dtsch med Wschr*, 10, 842
- NOBLE, W (1915) *J infect Dis*, 16, 132
- NOCARD and ROUX (1887) *Ann Inst Pasteur*, 1, 257
- NOVY, F G (1894) *Z Hyg InfektKr*, 17, 209
- OAKLEY, C L (1943) *Bull Hyg Lond*, 18, 781
- OAKLEY, C L and WARRACK, G H (1941) *J Path Bact*, 53, 335
- ORR, J H and REED, G B (1940) *J Bact*, 40, 441
- ORR, P F (1920) *Abstr Bact*, 4, 10, (1922) *J infect Dis*, 30, 118
- PAINE, F S (1931) *Zbl Bakt*, 11te Abt, 85, 122
- PAPPENHEIMER, A M (1935) *Biochem J*, 29, 2057
- PASTERNAK, J G and BENATSON, I A (1936) *Nat Inst Hlth Bull*, No 168, (1940) *Publ Hlth Rep, Wash*, 55, 775
- PASTEUR and JOUBERT (1877) *Bull Acad Méd*, 6, 781
- PETRIE, G F (1942-43) *Bull Hlth Org, Lo N*, 10, 113
- PFENNINGER, W (1924) *J infect Dis*, 35, 347
- PIZZINI, L (1898) *Zbl Bakt*, 24, 890
- PRÉVOT, A R (1938) *Ann Inst. Pasteur*, 61, 72
- PRIDGE, R (1936) *Z ImmunForsch*, 89, 477, (1937) *Ibid*, 91, 457
- REDDISH, G F and RETTORI, L F (1924) *J Bact*, 9, 13
- REED, G B and ORR, J H (1941) *War Med*, 1, 493
- REED, G B., ORR, J H., and BROWN, H J (1943) *J Bact*, 46, 475
- REED, R W (1942) *J Bact*, 44, 425
- Reports (1917) *Spec. Rep Ser med Res Coun., Lond*, No 12, (1919) *Ibid*, No 39
- RITCHIE, J (1901) *J Hyg, Camb*, 1, 125
- ROBERTS, R S (1931) *J comp Path*, 44, 245

- ROBERTSON, M. (1916) *J Path Bact*, 20, 327
- ROBINSON, E. M. (1923) 15th Ann Rep Dir vet Serv Union S Afr, 1, 111
- ROCKWELL, G. E. (1924) *J infect Dis.*, 35, 581
- RODWELL, A. W. (1941) *Aust Vet. J.*, 17, 58
- ROSENAU, M. J. and ANDERSON, J. F. (1908) *Bull U.S. Hyg Lab*, No 43.
- ROSENBAUM, (1886) *Arch lin Chir.*, 34, 306
- ROUX, E. (1888) *Ann Inst. Pasteur*, 2, 49
- SACQUÉPÉ, E. (1915) *Pr méd.*, 23, 183
- SCHOENHOLZ, P. (1928) *J infect Dis.*, 42, 40
- SCHOENHOLZ, P. and MEYER, H. F. (1922) *J infect Dis.*, 31, 610, (1924) *Ibid*, 35, 361, (1925) *J Immunol.*, 10, 1
- SCOTT, J. P., TURNER, A. W., and VAWTER, L. R. (1933) *Proc 12th int vet Congr N.Y.*, p 168
- SEDDON, H. R. (1922) *J. comp Path.*, 35, 147
- SEIFFERT, G. (1939) *Z Immunforsch.*, 96, 515
- SHERINGTON, C. S. (1917) *Lancet*, II, 964
- SHIFFEN, L. P. (1919) *Arch intern Med.*, 23, 348
- SIMONDS, J. P. (1915a) *J infect Dis.*, 16, 31, (1915b) *Ibid*, 16, 30, (1915c) *Monogr Rockefeller Inst.*, No 5
- DE SMIDT, F. P. G. (1924) *J Hyg, Camb.*, 22, 314
- SMITH, L. (1937) *J Bact.*, 34, 409
- SMITH, M. L. (1941-42) *Bull Hlth Org., L.O.N.*, 10, 104
- SMITH, T. (1890) *Zbl Bakt.*, 7, 502, (1909) *Trans Chicago path Soc.*, 7, 1
- SMITH, T., BROWN, T. H. R., and WALKER, E. L. (1905-6) *J med. Res.*, 14, 193
- SNYDER, M. L. (1936) *J Bact.*, 32, 401, (1937) *J infect Dis.*, 60, 223
- SNYDER, M. L. and HALL, I. C. (1935) *Zbl Bakt.*, 135, 290
- SOMMER, H. and SOMMER, E. W. (1932) *J infect Dis.*, 51, 243
- SORDELLI, A. (1922) *C. R. Soc. Biol.*, 87, 838, (1923) *Ibid*, 89, 53
- SORDELLI, A., FERRARI, J., and PRADO, M. (1930) *Per Inst. bact., B Aires*, 5, 797
- SORDELLI, A., PRADO, M., and FERRARI, J. (1932) *Folia biol., B Aires*, Nos. 14 and 15, pp 58-63
- SPRAY, R. S. (1936) *J Bact*, 32, 135
- STARIN, W. A. (1924) *J infect. Dis.*, 34, 148
- STARIN, W. A. and DACK, G. M. (1923) *J infect. Dis.*, 33, 169, (1924) *Ibid*, 34, 137, (1925) *Ibid*, 36, 383
- STARK, C. N., SHERMAN, J. M., and STARK, P. (1928) *J infect Dis.*, 43, 560
- STEVENS, F. A. (1930) *J infect Dis.*, 57, 270
- STEWART, S. E. (1938) *J Bact.*, 35, 13, (1940) *Publ Hlth Rep, Wash.*, 55, 703
- STICKLAND, L. H. (1934) *Biochem. J.*, 28, 1746, (1935) *Ibid.*, 29, 288, 889
- STODDARD, J. L. (1919a) *Lancet* L 12, (1919b) *J exp med.*, 29, 187.
- SVEC, M. H. and MCCOY, E. (1944) *J Bact*, 48, 31
- TANVER, F. W. and DACK, G. M. (1922) *J infect Dis.*, 31, 92
- TANVER, F. W. and TWOHEY, H. B. (1926) *Zbl. Bakt.*, 98, 136
- TAROZZI, G. (1905) *Zbl. Bakt.*, 37, 619
- TAYLOR, A. W. (1940) *J comp Path.*, 53, 50
- TAYLOR, A. W. and STEWART, J. (1941) *J Path Bact.*, 53, 87
- THEILER, A. (1928) 13th and 14th Rep, Director vet Educat. Pes, S Africa, p. 47
- THEILER, A., VILJOEN, P. R., GREY, H. H., DU TOIT, P. J., MEYER, H., and ROBINSON, E. M. (1926) 11th and 12th Rep, Director vet. Educat. Res, S Africa Part II, p. 821
- TISSIER, H. and MARTELLE, (1902) *Ann. Inst Pasteur*, 16, 865
- TODD, E. W. (1941) *Brit J exp Path.*, 22, 172
- TOLDO, S. and VEILLOX, (1891) *Zbl Bakt.*, 9, 18
- TORREY, J. C. (1925) *J infect Dis.*, 36, 517
- TORREY, J. C., KAHN, M. C., and SALINGER, M. H. (1930) *J Bact.*, 20, 80.
- TSCHEKROW, L. (1929) *Z Immunforsch.*, 63, 262.
- TULLOCH, W. J. (1919) *J Hyg, Camb.*, 18, 103
- TURNER, A. W. (1930) *Aust Coun. sci industr Res, Bull.*, No 46.
- TURNER, A. W. and EALES, C. E. (1941) *Aust J exp Biol med. Sci.*, 19, 167
- TURNER, A. W. and RODWELL, A. W. (1943) *Aust J exp Biol med Sci.*, 21, 17, 27
- USCHINSKY, N. (1893) *Zbl. Bakt.*, 14, 316
- VAILLARD, L. and ROUGET, J. (1892) *Ann Inst Pasteur*, 6, 380
- VARNY, P. L. (1926) *J Lab clin. Med.*, 11, 1
- VAWTER, L. R. and RECORDS, E. (1926) *J Amer vet. med. Ass.*, 68, 494, (1931) *J infect. Dis.*, 48, 581
- VELLIZ, L. (1936) *C. R. Acad Sci*, 203, 471, 498
- WAGNER, E. (1924) *J infect. Dis.*, 35, 353
- WAGNER, E., DOZIER, C. C., and MEYER, H. F. (1924) *J infect Dis.*, 34, 63
- WALBURN, L. E. (1938) *J. Path. Bact.*, 46, 85

- WALBURN L E and REYMAN, C G (1933) *J Path Bact*, 36, 469
- WEINBERG, M. (1929) *Bull Inst Pasteur*, 27, 529, 577
- WEINBERG, M and COMBESCO, N (1930) *Ann Inst Pasteur*, 45, 547
- WEINBERG, M, DAVESNE, J, and LEFRANC, M (1931) *C R Soc Biol*, 107, 506
- WEINBERG, M, DAVESNE, J, MIHAILESCO, M, and SANCHEZ, C (1929) *C R Soc Biol*, 101, 907
- WEINBERG, M and GINSBOURG, B (1927) 'Données récentes sur les microbes anaérobies et leur rôle en pathologie' Masson et Cie, Paris
- WEINBERG, M and GUILLAUMIE, M (1936) *C R Soc Biol*, 121, 127.
- WEINBERG, M and MIHAILESCO, M (1929) *Ann Inst Pasteur*, 43, 1408
- WEINBERG, M, NATIVELLE, R, and PRÉVOT, A R (1937) *Les Microbes Anaérobies* 'Masson et Cie, Paris
- WEINBERG, RENARD, C, and DAVESNE, J (1926) *C R Soc Biol*, 94, 813
- WEINBERG and SÉGUIN, P (1915) *C R Soc Biol*, 78, 274, (1916) *C R Acad Sci*, 163, 449, (1918) "La gangrène gazeuse" Paris
- WEISS, H (1921) *J infect Dis*, 28, 70
- WELCH, W H and NUTTALL, G H F (1892) *Johns Hopk Hosp Bull*, 3, 81
- WHEELER, M W and HUMPHREYS, E M (1924) *Johns Hopk Hosp Bull*, 35, 305
- WILSDON, A J (1931) *2nd Rep Director, Inst Anim Path*, Camb, p 53 (1933) *Ibid* 3rd Rep, p 46
- WILSON, G S (1928) *J Path Bact*, 31, 113
- WINOGRADSKY, S (1902) *Zbl Bakt*, 11te Abt, 9, 43, 107
- WOODS D D and TRIM A R (1942) *Biochem J*, 36, 501
- WOLF, C G L (1918-19) *J Path Bact*, 22, 115, (1919-20) *Ibid*, 23, 254.
- WOLF C G L and HARRIS, J E (1917) *J Path Bact*, 21, 386
- ZEISSLER, J and KRANEVELD, F C (1929) *Arch wiss prakt Tierheilk* 60, 441
- ZEISSLER, J and RASZELD, L (1929) *Arch wiss prakt Tierheilk*, 59, 419.

## CHAPTER 37

### MISCELLANEOUS BACTERIA

We include in this chapter a number of organisms which, for one reason or another, cannot justifiably be allotted to any of the named groups<sup>1</sup>

#### The Morax-Axenfeld Bacillus.

This organism, which is responsible for subacute or angular conjunctivitis in human beings, was described independently by Morax (1896) and Axenfeld (1897). The name *B. lacunatus* was suggested for it by Eyre (1900).

**MORPHOLOGY**—In films of the conjunctival secretion it occurs in the form of rods 2-3  $\mu$  long and 1  $\mu$  broad, with parallel or slightly convex sides and rounded ends. The bacilli occur in pairs, placed end to end, and sometimes in short chains. They are found free in the secretion, or within the polymorphonuclear and desquamated epithelial cells. They are non motile, non sporing, Gram negative, and except for the absence of a capsule they closely resemble Friedlander's pneumobacillus. In old cultures pleomorphic forms are numerous, ranging in size and shape from short stunted diplococcal forms to long jointed or filamentous, sometimes fusiform threads (Eyre 1900).



FIG 211.—MORAX AXENFELD BACILLUS

Surface colonies on Fildes' agar plate, 24 hours, 37° C ( $\times 8$ )

**CULTIVATION**—Growth occurs only in the presence of some natural animal protein, such as serum, blood, or ascitic fluid, there is no development on ordinary nutrient agar or potato nor in broth, milk, or gelatin. Development is best on Fildes' agar and on serum or egg medium, it is poor on blood agar and very poor on chocolate agar. The organism is said to grow only between 30° and 40° C., and to be strictly aerobic. Our own limited experience suggests that some growth may occur below 30° C., and that under anaerobic conditions on favourable media slight but definite growth may be evident. On serum agar plates after 24 hours at 37° C. the colonies are round, up to 1 mm. in diameter, raised, greyish, and translucent, during the next few days they increase in size, reaching 2-5 mm. in diameter, and become differentiated into a slightly raised opaque whitish centre, and a thin, translucent periphery with a lobate edge, the medium becomes pitted owing to liquefaction of the serum. On Loeffler's

serum no actual colonies are visible, but the whole surface is covered with pits of liquefaction—hence the term *lacunatus*. On Fildes' agar after 24 hours at 37° C. the colonies are water clear, amorphous, low convex, 0.4 mm. or so in diameter, with a smooth glistening surface and entire edge, after 4 days they are larger, and are differentiated into a smooth, raised, central papilla and a wide, effuse, granular, dull, transparent, peripheral extension with an irregularly undulate margin. The central papilla may be so small and the peripheral portion so transparent that the colonies are difficult to see. By transmitted light they have a frosted glass appearance. On horse blood

<sup>1</sup> The terms *Bact. pneumoninles* and *Bact. granulosis* are used for convenience but it must be pointed out that neither of these organisms belongs to the *Bacterium* group as defined on p. 654.

agar the colonies of some strains are surrounded by a fairly wide zone of incomplete hemolysis, though no soluble haemolysin is formed in fluid media (Oag 1942). On Dorset egg low convex colonies are formed, which lead in a few days to slight pitting of the surface. There is no growth on MacConkey agar. In serum broth after 24 hours at 37° C., there is a uniform turbidity; later a greyish white deposit appears, and increases for 6 to 10 days, after which the medium clears, the whole of the growth sinking to the bottom of the tube.

**RESISTANCE, BIOCHEMICAL REACTIONS, AND PATHOGENICITY**—Cultures in serum broth live for weeks at 35° C., but die in 48 hours if left at room temperature. The bacillus is killed by moist heat at 58° C. in 15 minutes, and is apparently very susceptible to zinc salts, which have an almost specific action in the treatment of angular conjunctivitis. In our experience no sugars are fermented. Gelatin may be liquefied and litmus milk becomes very slowly alkaline, though according to Oag (1942) acid is produced. Nitrates are reduced. Catalase is negative. No indole is formed in serum broth cultures even after 7 days. According to Oag (1942) antigenic differences can be demonstrated by agglutination between haemolytic and non haemolytic strains. The organism is non-pathogenic to laboratory animals, whether inoculated into the conjunctival sac or directly into the tissues, but a drop of culture instilled into the conjunctival sac of a healthy human volunteer gives rise in about 5 days to a typical attack of angular conjunctivitis.

A similar organism *B. duplex* was described by Petit in 1900 who isolated it from cases of conjunctivitis associated with corneal ulceration. It differs from the Morax-Axenfeld bacillus mainly in its ability to grow on media without the addition of natural animal protein (though no growth may occur in ordinary broth) in its development at room temperature in its liquefaction of gelatin, and in its more active digestion of Loeffler's serum, which it liquefies in 3 or 4 days. It is also said to be antigenically distinct (Audureau 1940). On ascitic agar it gives rise to convex not umbonate colonies which are greyish in colour, viscous and not so translucent as those of the Morax-Axenfeld bacillus.

Another organism which is closely related to if not identical with the Morax-Axenfeld bacillus, was isolated by Jones and Little (1923) from cattle suffering from acute conjunctivitis sometimes complicated by corneal ulceration. It is said to liquefy gelatin, and to produce alkali in litmus milk.

**CLASSIFICATION**—The classification of these organisms is still in doubt (see Chapter 33). Iloff (1939) suggested that they should be grouped in a new genus called *Moraxella*. Audureau (1940) accepts this suggestion and recognizes three main species *lacunata*, *duplex* and *licoffi*. *Moraxella lacunata* grows only in the presence of serum. *Moraxella duplex* grows in the absence of serum but not in synthetic media, and *Moraxella licoffi* which was isolated by Audureau from patients affected with conjunctivitis will grow in a synthetic medium free from protein and amino acids provided a little carbohydrate is added to provide energy.

### *Bacterium pneumosintes*

Isolated by Olitsky and Gates (1921*a, b, c*, 1922*a, b*) from the nasopharyngeal washings of patients in the early stage of influenza and since observed by certain other workers (Hall 1926). It is a very small organism and is capable of passing through Berkefeld N and V candles.

**MORPHOLOGY**—In Smith-Noguchi medium on first isolation, it is described as consisting of minute bodies, arranged singly, in pairs, or short chains, its length is given as 0.15–0.3  $\mu$ , and its breadth as two or three times less. After subculture for some time in the laboratory, the organisms appear in dextrose peptone broth as plump rods with rather pointed ends arranged in pairs end-to-end, and in fairly long chains. It may be 0.5–1.0  $\mu$  long, it stains more deeply in the middle than at the ends. Our own observations on one of Olitsky and Gates's original strains, grown anaerobically for 12



days on blood agar, show that it is a very small straight rod-shaped organism, 0.5-1.0  $\mu$  long by 0.2-0.3  $\mu$  broad, with parallel sides and rounded ends, it is arranged singly, in pairs end-to-end, and in small dense groups. No irregular forms are seen, it stains regularly with the usual aniline dyes, is Gram-negative, and non motile (Fig. 212).

CULTURALLY, the organism can be isolated only in Smith-Noguchi medium (human ascitic fluid containing a piece of sterile rabbit kidney, and covered with a vaseline seal); but after three or four subcultures in this medium it can be brought to grow anaerobically on blood agar, chocolate agar, Fildes' agar, Bordet's medium, and certain other media.

*Smith-Noguchi Medium at 37° C*—A faint haze becomes visible around the kidney about the 5th day, and reaches a maximum about the 8th day, when it is 3 cm. deep. These appearances are not characteristic; they often appear in uninoculated tubes.

*Horse Blood Agar*—7 days, 37° C Round, convex, milky white opaque colonies, 0.5 mm. in diameter, of amorphous structure, and with a smooth glistening surface and an entire edge; butyrous in consistency and easily emulsified, no hemolysis.

*Horse Blood Agar or Chocolate Agar Slope*—12 days, 37° C Slightly raised, partly confluent, glistening growth with an irregular surface, due to imperfect fusion, and an edge made up of very tiny discrete colonies.

*Bordet's Medium*—12 days, 37° C Slightly raised, confluent, glistening growth with a finely pitted surface.

*Glycerol Egg*—7 days, 37° C Good, slightly raised, glistening, viscous growth with a smooth surface and an entire edge.

*Leifert's Serum*—7 days, 37° C Similar to growth on glycerol egg, but not so abundant.

*Trypsinated Heart Agar Slope, and Glycerol Agar Slope*—7 days, 37° C Rather poor, slightly raised, confluent, nearly colourless, translucent growth with an irregular surface due to imperfect fusion, edge made up of single colonies.

*Trypsinated Heart Broth*—7 days, 37° C Poor to moderate, uniform turbidity, with a moderate, highly viscous deposit, which, on shaking, coheres in a ropy mass and is difficult to disintegrate. No surface growth, slightly aromatic odour.

*Fildes Broth*—7 days, 37° C No turbidity, but slight powdery deposit disintegrating on shaking. After 14 days the deposit is viscous, coherent, and difficult to disintegrate.

*Astrate Broth*—7 days, 37° C Slight turbidity, moderate, viscous deposit, disintegrating completely, no surface growth.

*Serum Broth*—Growth similar to that in heart broth.

*Coli Broth*—5 days, 37° C (Prepared by growing a strain of *Bact. coli* in 1 per cent. dextrose broth till the first sign of turbidity appears, the culture is then steamed for half an hour) Moderate to dense turbidity, moderate viscous deposit, very difficult to disintegrate.

RESISTANCE—Cultures in Smith-Noguchi medium, after 5 to 7 days' incubation at 37° C., remain viable at room temperature for 2½ years. Cultures in coli-broth become sterile in about a week. The organisms withstand freezing, and drying *in vacuo*, and appear to remain alive for a long time when dried. Infected lungs of rabbits, kept in 50 per cent. glycerol at 4° C., remain virulent for 9 months. The organism is destroyed by moist heat at 56° C. in 30 minutes, and by chloroform vapour in 1 to 1½ hours.



FIG. 212.—*Bacterium pneumonales*

From a surface growth on blood agar anaerobically 12 days 37° C ( $\times 1000$ )

Indole is not formed; nitrates are reduced to nitrites; and gelatin is not liquefied. It is destroyed by heat at 57° C. in 10 minutes (Olitsky 1930). The organism is non-pathogenic for laboratory animals, but when inoculated subconjunctivally into *rhesus* monkeys it gives rise to a granular conjunctivitis simulating trachoma (see Chapter 85). (For a fuller description of this organism see Noguchi 1923, Tilden and Tyler 1930.)

A Gram-negative bacillus simulating *Bact. granulosis* in many respects has been described under the name of *Bact. simia* by Olitsky, Syverton, and Tyler (1933), who isolated it from monkeys suffering from spontaneous conjunctival folliculosis. Like *Bact. granulosis*, which it resembles in size, it is motile by a single polar flagellum, produces a nebulous opacity in leptospiral medium, fails to grow anaerobically, has an optimum growth temperature of 30° C., and is non pathogenic for laboratory animals. It differs, however, from this organism in having a capsule, in growing on plain nutrient agar, in fermenting different carbohydrates, in failing to reduce nitrates, and in showing no agglutination with *granulosis* antisera. What relation *Bact. granulosis* and *Bact. simia* bear to Ducey's bacillus, and to the less exacting members of the *Hamophilus* group, has still to be determined.

#### *Bact. alkaligenes* and *Vibrio alkaligenes*.

Petruschky (1896) isolated from human faeces an organism that produced an alkaline reaction in certain media, and was named by him *Bact. faecalis alkaligenes*. It was described as a Gram-negative bacillus, and has long been included by most authorities in the genus *Bacterium* under the name of *Bact. alkaligenes*. Though it appears to be an almost constant inhabitant of the intestinal tract of man, this organism may give rise to infections of the enteric type (Petruschky 1896, Hirst 1917, Khaled 1923).

The characters usually ascribed to *Bact. alkaligenes* are as follows: The cells are very variable in size, but are usually longer and thinner than those of *Bact. coli* ( $5-7 \mu \times 0.4 \mu$ ), and the ends are less definitely rounded. The bacilli are motile, with peritrichate flagella. The colonies on agar are flatter than those of *Bact. coli*, and more contoured, with a raised central portion and a spreading undulate edge. Neither acid nor gas is produced in any of the usual test substrates. Litmus milk is rendered strongly alkaline. A characteristic brown colour is produced on potato.

Many of these characters are very unlike those of the genus *Bacterium*, and doubts have been frequently expressed as to its real systematic affinities. A recent study by Nyberg (1935) raises even stronger doubts as to whether the strains that have been described under this name can be regarded as forming a bacterial species. Nyberg examined with great care 134 strains labelled *Bact. alkaligenes*, and was able to distinguish among them two quite distinct forms, and a number of less well-differentiated types. The form for which he would reserve the name *Bact. alkaligenes* is a short, thick bacillus, usually non-motile or very feebly motile, but possessing in most cases poorly formed peritrichate flagella. It fails to ferment dextrose, levulose, lactose, maltose, saccharose, rhamnose, xylose, arabinose, mannitol, sorbitol, dulcitol, inositol or salicin. It does not form indole, and it produces no change in milk. Of the 134 strains examined, 71 were of this type. Nyberg notes that this description does not agree with that given of *Bact. alkaligenes* in many books and papers, and that it is impossible to be certain that his strains correspond to the organism originally isolated by Petruschky. He considers, however, that they probably belong to the same species, and that the name must certainly be given to them and not to the type to which his other strains belong.

This second type is a long, thin, slightly curved rod, actively motile by lophotrichate flagella. It is therefore not a bacillus, but a vibrio. Unlike the former type, it renders a dextrose medium slightly but definitely alkaline. Nyberg regards this organism as identical with the *Vibrio alkaligenes* of Lehmann and Neumann (1896); and there would seem no doubt that this is its proper name. It also seems very probable, as Nyberg

emphasizes, that many of the strains that have been isolated by various workers and have been given the description summarized earlier were *Vibrio alkaligenes* not *Bacterium alkaligenes*

It is worth noting that as pointed out by Conn (1942) failure to produce acid in a glucose medium may be due either (1) to non fermentation or (2) to utilization of the glucose so completely that there are no by products capable of giving an acid reaction except  $\text{CO}_2$  which in a buffered medium will not be detected

### Bartonella, Eperythrozoon, and Grahamella

Various bodies have been described by different workers in close association with the red blood corpuscles of man and animals suffering usually though not always from certain types of anaemia. The evidence that *Bartonella* and *Eperythrozoon* are living reproducible micro-organisms capable of giving rise under favourable conditions to disease is now very strong but considerably less is known about the *Grahamella* though there is increasing reason to believe that these bodies also are definite bacteria. The interest that *Bartonella muris* particularly has stimulated of recent years is due to the remarkable part played by the spleen in the normal defence mechanism of the host (see Chapter 79)

Some authors regard the organism causing human Oroya fever as generically distinct from the organisms responsible for infective anaemia of animals. They would classify the first as *Bartonella* the remainder as *Hanobartonella*. The distinctions between them are summarized as follows. *Bartonella* besides invading red blood corpuscles, is able to develop in fixed tissue cells and to cause a skin eruption. It is moreover insusceptible to arsenic preparations. *Hanobartonella* on the other hand is said not to grow outside the blood. It rarely produces disease without removal of the spleen and the disease it causes is influenced by arsenotherapy (see Weinman 1944). Whether these differences are sufficient to justify the establishment of a separate genus for the animal bartonellæ is very doubtful and for the present we shall group them all under the one genus *Bartonella*. Though we shall refer to only three species of *Bartonella* it may be mentioned that numerous other species have been described infecting guinea pigs, voles, squirrels, hamsters, opossums, cattle, buffaloes and other animals. For a review of *Bartonella* and *Eperythrozoon* the reader is referred to the monograph by Weinman (1944).

### *Bartonella bacilliformis*

This organism is a small bacillus, which invades the red blood cells, and is responsible for Peruvian Oroya fever and for verruga peruana (see Chapter 79). It was called *Bartonella bacilliformis* by Strong and his colleagues (1915) in honour of Barton who was one of the first to observe the bacillus in the red cells. Noguchi and Battistini in 1906 cultivated the organism and reproduced a disease in monkeys bearing a close resemblance to the natural disease in human beings. The organism was recovered in pure culture from the blood of the injected monkeys.

**Morphologically in culture** it is a small pleomorphic bacillus, varying in length from 0.3-2.5  $\mu$  and in breadth from less than 0.2 to as much as 0.5  $\mu$ . Dumb-bell forms predominate and coccoid forms are common. It is arranged singly and in dense masses. It is motile, Gram negative and stains reddish violet with Giemsa. Cultivation can be effected on a variety of media such as the semi-solid serum haemoglobin agar medium used for leptospiræ or a blood glucose cystine agar (Jimenez 1940) or a 2 per cent proteose agar to which 20 per cent of fresh defibrinated blood or serum from the rabbit or sheep is added together with 0.2 per cent of an ascorbic acid glutathione solution (German 1941). Though a high proportion of natural animal protein favours growth it does not

appear to be essential. Jimenez (1940) for example states that good growth was obtained on 1 per cent glycerol infusion agar provided the X though not the V factor (see Chapter 33) was added. On solid media growth may occur in 4-5 days either as minute circular, clear mucoid colonies, or as an opaque finely granular mucoid film that has a tendency to outgrow the original boundaries of the inoculum (Jimenez 1940). The organism is aerobic grows well at 25-37° C., though best at about 25-25° C., prefers a pH of 7-8, and survives in cultures for 1 to 4 months. Pinkerton and Weinman (1937) found that in tissue cultures the organisms, unlike *Picketia*, grew extracellularly as well as intracellularly. Growth occurs also in the allantoic fluid of the developing chick embryo incubated at 25-25° C., but this medium is unsuitable for serial cultivation (Jimenez and Boddinck 1940).

Injected intravenously into young rhesus monkeys the organism gives rise to a peculiar irregularly remittent type of fever sometimes accompanied by severe anemia, injected intradermally into the eyelid it gives rise to a nodule rich in cellular elements and capillary formation. (For further description see Noguchi 1925, Kikuth 1931, Pittaluga 1935.)

#### *Bartonella muris*

This organism was first described by Mayer (1921) who found it in the blood cells of rats experimentally infected with trypanosomes. Morphologically it closely resembles *Bartonella bacilliformis*. It is actively motile in culture media, and flagella have been demonstrated. On blood agar minute colonies appear in 48 hours, and gradually increase in size till after a few days they coalesce to form a thin, film-like tenacious growth on the surface of the

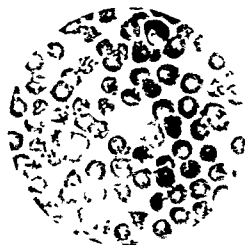


FIG. 213.—*Bartonella muris*.

Organisms in red blood cells of rat ( $\times 1000$ ).  
[From specimen kindly supplied by the late Professor J G Thomson.]

medium. The blood is not hemolyzed. Cultures on solid media have a sweet odour resembling canned pineapple. The optimum temperature for growth is 25° C. In infected blood the organisms are destroyed by exposure to a temperature of 5° C. for 30 minutes (Ford and Ebot 1925) but remain virulent in the frozen state for at least 11 weeks (Kessler 1942). Pure cultures inoculated into splenectomized rats give rise to anemia (see Chapter 9). The organism is fairly common in rats, causing an infection which normally remains latent but which can be activated by splenectomy by poisons such as toluylenediamine and by certain infections (for general description see Lauda and Marcus 1925, Marmorston-Gottesman and Perl 1932, Kikuth 1931, 1934).

#### *Bartonella canis*

A similar organism has been described under this name by Kikuth (1928), who is responsible for infectious anemia of dogs. In the red cells the organisms are very pleomorphic, large and small, coccoid and rod shaped forms being seen. Kikuth was unsuccessful in cultivating them on artificial media. (See Kikuth 1929, Perard 1929, Lwoff and Provost 1929, Reverdanz and Reichenow 1932, and Chapter 9.)

Besides the species already mentioned *Bartonella* has been found infecting various rodents in different parts of the world monkeys and some other mammals in America, and cattle in Algeria and Palestine (see Pittaluga 1935). Whether these organisms deserve specific rank or not must await further study.

*Eperythrozoon coccoides*.

This is a parasite of mice which was discovered independently by Dinger (1928, 1929), and Schilling (1928). A considerable proportion of normal mice appear to be infected, but the organisms are rarely found unless the spleen is removed. Two to four days after splenectomy the organisms appear in the blood in the form of rings or discs stuck on to the external surface of the red blood corpuscles, and staining a bluish purple colour with Giemsa. Unlike *Bartonella muris*, they have a preference for polychromatic red cells. They may persist in the blood for weeks or months, their numbers varying from time to time. Beyond producing a slight degree of anaemia, they seem to be without any very definite effect on the animal. Mice infected with *Eperythrozoon coccoides* are susceptible to *Bartonella muris* showing that the two organisms are distinct (see also McCluskie and Niven 1934, Schwetz 1934, Marmorston 1935). In contrast to *Bartonella* *Eperythrozoon* is round in shape with numerous annular and disc like elements, rods are rare and are seldom in chain formation. The organisms have not yet been cultivated. Infection appears to be spread by lice. Numerous other animals, besides mice, are liable to infection with *Eperythrozoon* (see Weinman 1944).

*Grahamella*

This parasite was first described by Graham Smith (1905) at Cambridge who observed it in the red blood cells of 10 per cent. of moles that were being examined for *Piroplasma*. Since then it has been found in the blood of several other animals. The organisms appear as longer or shorter rods of irregular contour lying within the red corpuscles. Though resembling *Bartonella*, they are much coarser, and more like ordinary bacteria. With Giemsa they take on a blue rather than a reddish tint. Only occasional red cells are affected. According to Jettmar (1932) they can be cultivated on serum blood agar. They appear to be non pathogenic and to have no effect on the health of the host. In the rat Vassiliadis (1935) has been able to transmit infection from one animal to another in series. In most animals splenectomy has little or no influence on infection though the rat is said to constitute an exception (Vassiliadis 1935). Ectoparasites, such as lice are probably responsible for the natural transmission of infection. Unlike *Bartonella muris*, *Grahamella* seems to be resistant to arsenic (see also Kikuth 1934).

## REFERENCES

- AUDREAU, A. (1940) *Ann. Inst. Pasteur* 64, 126.  
 AXENFELD, T. (1897) *Zbl. Bakt.* 21, 340.  
 COCK, H. J. (1942) *J. Bact.* 44, 353.  
 DINGER, J. E. (1928) *Ned. Tijdschr. Geneesl.* No. 48 72, 5903, (1929) *Zbl. Bakt.* 113, 503.  
 EYRE, J. W. (1900) *J. Path. Bact.* 6, 1.  
 FORD, W. W. and ELIOT, C. P. (1928) *J. exp. Med.* 48, 475.  
 GARROD, L. P. (1918) *Brit. J. exp. Path.* 9, 155.  
 GATES, F. L. (1926) *J. exp. Med.* 44, 787.  
 GEINAN, Q. M. (1941) *Proc. Soc. exp. Biol.* A 1 47, 329.  
 GRAHAM SMITH, G. S. (1905) *J. Hyg., Camb.* 5, 453.  
 HALL, M. W. (1926) *J. exp. Med.* 44, 539.  
 HIRST, L. F. (1917) *J. R. Army med. Cps* 29, 476.  
 JETTMAR, H. W. (1932) *Z. Parasitenk.* 4, 254.  
 JIMÉNEZ, J. F. (1940) *Proc. Soc. exp. Biol.* A 1 45, 40.  
 JIMÉNEZ, J. F. and BIDDINGH, G. J. (1940) *Proc. Soc. exp. Biol.* A 1 45, 546.  
 JONES, F. S. and LITTLE, R. B. (1923) *J. exp. Med.* 38, 139.  
 KESSLER, W. R. (1942) *Proc. Soc. exp. Biol.* A 1 49, 234.  
 KHALED, Z. (1923) *J. Hyg., Camb.* 21, 362.  
 KIKUTH, W. (1928) *Ann. Bact.* 7, 1729, (1929) *Zbl. Bakt.* 113, 1 (1931) *Z. Immunforsch.* 73, 1, (1934) *Proc. roy. Soc. Med.* 27, 1211.  
 LAUDA, E. and MARCUS, F. (1924) *Zbl. Bakt.* 107, 104.  
 LERMAN, H. and NEUMANN, R. (1896) 'Atlas und Grundlege der Bakteriologie, etc.' J. F. Lehmann, Munich.

- LWOFF A. (1930) *Ann. Inst. Pasteur* 62, 161.  
 LWOFF A. and PROUDOT A. (1929) *C. R. Soc. Biol.* 101, 9.  
 MCCLURE, J. A. W. and AYRES J. S. F. (1934) *J. Path. Bact.* 23, 121.  
 MARMONSTEIN, J. (1931) *J. infect. Dis.* 56, 142.  
 MARMONSTEIN-GOTTESMAN, J. and PELLER, D. (1932) *J. exp. Med.* 56, 763.  
 MATHE, M. (1921) *Arch. Sci. Exp. et Trévise* 25, 151.  
 MILLS, K. C., SHEDLEY G. S. and DOUGLAS, A. E. (1930) *J. exp. Med.* 4, 153.  
 MORAN, V. (1930) *Ann. Inst. Pasteur* 10, 23.  
 NOGUCHI, H. (1926) *J. exp. Med.* 44, 523, 67, 75, 79 (1927) *J. Amer. med. Ass.* 39, 79 (1928) Monograph on Trachoma *J. exp. Med.* 48, No. 2, 1928, No. 2.  
 NOGUCHI, H. and BARTISTE, T. S. (1926) *J. exp. Med.* 43, 531.  
 NYKING, C. (1935) *Zbl. Bakt.* 133, 443.  
 OAG, R. K. (1912) *J. Path. Bact.* 54, 121.  
 OLITSKY P. K. (1930) *Trans. 2nd ann. meeting Amer. Acad. Ophthalm. Otolaryng.* p. 223.  
 OLITSKY P. K. and GATES, F. L. (1921c) *J. exp. Med.* 33, 125 (1921d) *Ibid.* 33, 371 (1921e) *Ibid.* 33, 713 (1922a) *J. exp. Med.* 35, 913 (1922b) *Ibid.* 35, 931.  
 OLITSKY P. K. and McCARTHY J. E. (1923) *J. exp. Med.* 38, 42.  
 OLITSKY P. K., STURTEVANT J. T. and TYLER, J. R. (1933) *J. exp. Med.* 57, 571.  
 PIERARD, C. H. (1929) *C. R. Soc. Biol.* 103, 1111.  
 PETIT P. (1900) "Recherches critiques et bactériologiques sur les infections aigües de la corne" Thèse de la Faculté de Médecine de Paris. G. Sirey, Paris.  
 PETROVICH, J. (1906) *Zbl. Bakt.* 19, 177.  
 PINKERTON H. and WEINMAN D. (1937) *Proc. Soc. exp. Biol. & Med.* 5.  
 PIZZALUNA, G. (1931) *Bull. Inst. Pasteur* 28, 961.  
 REGENDAN, P. and REICHENOW E. (1932) *Arch. Sci. Exp. et Trévise* 36, 331.  
 SCHILLING, V. (1928) *Arch. u. Woch.* 7, 123.  
 SCHWITZ, J. (1934) *Zbl. Bakt.* 132, 111.  
 STRONG, R. P., TYLER, E. E., BROWN, C. T., SELLARD, A. W. and GARNHAMET J. C. (1915) *P. p. 1st Annual S. American* 1915. Harvard Univ. Press, Cambridge, Mass.  
 THOMPSON E. B. and TYLER, J. R. (1930) *J. exp. Med.* 52, 61.  
 VASSILIADIS, P. (1935) *Amer. Soc. exp. Biol.* 15, 272.  
 WEINMAN D. (1944) *Trans. Amer. microbiol. Soc.* 33, Pt. III, 443.

## CHAPTER 38

### THE SPIROCHÆTES

THE name Spirochæte was first given by Ehrenberg in 1833 to a large flexible motile organism occurring in water, it is now used as a general term for all elongated, motile, flexible organisms that are twisted spirally around their long axis. Though the spirochætes vary greatly in size, they all possess certain features in common: thus they possess no flagella, they exhibit no antero-posterior polarity (i.e. they can move either forwards or backwards), they contain no colouring matter and no cyanophycin granules, they show no definite localized nucleus, they divide transversely, division being either simple or multiple, and they exhibit no sexual phenomena of reproduction. These properties bring them closely into line with the Bacteria, to which they are more nearly related than to the Protozoa. Indeed as Dobell (1912) points out while there are many features in which they differ from the Protozoa, there is only one feature that differentiates them from the bacteria, namely motility without flagella. Even this difference is now subject to doubt, since studies by the electron microscope on some of the treponemata have revealed filamentous bodies that may be interpreted as flagella (Mudd, Plevitzky and Anderson 1943, Wile and Kearney 1943).

Without entering into the disputed question of their classification it is convenient for descriptive purposes to divide the spirochætes into four groups—*Spirochata*, *Cristispira*, *Treponema*, and *Leptospira*.

**Spirochaeta**—The members of this group possess an axial fibre, around which the body is twisted in a spiral manner, in just the same way as a spiral staircase is built round the newel. The organism possesses a series of regular primary spirals, during motion a series of secondary waves may be superimposed on these but whatever form the organism as a whole may assume, the primary spirals remain intact. Metachromatic granules of volutin are distributed uniformly throughout the length of the organism. The type species *Spirochata plicatilis* Ehrenberg, is usually 200–500  $\mu$  in length and 0.5–0.7  $\mu$  in thickness (Wenyon 1926). The number of primary spirals is 100 to 250, the distance between successive turns being about 2  $\mu$ . So far no members of this group have been found to be capable of causing disease.

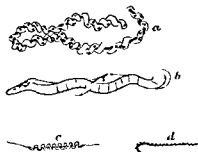


FIG 214—DIAGRAM OF THE SPIROCHÆTES

a *Spirochata*                      b *Cristispira*  
c *Treponema*                      d *Leptospira*  
(After Noguchi)

**Cristispira.**—The peculiar characteristic of members of this group is the possession of a band like membrane or *crista* which runs in a spiral manner along the organism. This membrane is extremely thin its width is much the same as that of the organism itself except at the ends where it narrows down to fuse with the surface pellicle. The body of the organism is divided into chambers by septa of thickened cytoplasm and on either side of each septum or sometimes distributed irregularly through the more fluid cytoplasm in the chambers are a number of metachromatic granules. The type species *Cristispira balbiani* is found in the crystalline style of the oyster it is about 45–100  $\mu$  long and 1–1.5  $\mu$  broad (Wenyon 1926). Other species have been found in a similar situation in other Mollusca.

**Treponema.**—Members of this group possess neither an axial fibre nor a *crista*. According to some observers the body is divided up into chambers like that of a *Cristispira* but this is by no means certain. The presence of metachromatic granules is doubtful. The organism shows a number of primary spirals which may be closely or loosely wound during movement secondary turns may develop but at rest these disappear so that the organism is straight. The ends may be rounded or pointed and in some members it is possible to distinguish a thin drawn out filament at the ends, this is probably not a true flagellum but the remains of the thin connecting bridge of cytoplasm that is seen during transverse division. Electron micrographs (Wile *et al* 1942 Wile and Kearney 1943 Madd Polvitzky and Anderson 1943) show the presence of a delicate cell wall or periplast enclosing the inner protoplasm. Dense granules 40–90  $m\mu$  in diameter are often seen in the protoplasm and large irregular spheroidal bodies 150–500  $m\mu$  in diameter may occur attached to the cell near the end or connected with it by a short stalk. What have been interpreted as true flagella may be seen often in groups of four, along the sides or near the ends of *Trep pallidum* and *Trep macrodentum*. Treponemata are widely distributed numerous species have been described in water in the gut of certain insects such as white ants and cockroaches and in the large gut of the toad in human beings they are found in the mouth, sometimes in the alimentary tract and the bronchi around the urethral orifice in certain ulcerating conditions of the skin in condylomata in the blood of patients with relapsing fever and in the manifold lesions of syphilis. They vary considerably in size thus *Treponema termitidis* Leidy is 20–60  $\mu$  long and 0.5  $\mu$  broad on the other hand *Treponema parvum* may be only 3  $\mu$  in length and 0.2  $\mu$  in thickness (Dobell 1912).

**Leptospira.**—The members of this group possess neither axial filament nor *crista* their cytoplasm is not obviously chambered. They show, however a large number of closely wound primary spirals this differentiates them from the treponemata, the spirals of which are fewer and less closely wound. Moreover the leptospiræ frequently have their ends turned round at a sharp angle to the rest of the body. When the organism is at rest the ends appear characteristically hooked but when it is in motion rotating round its long axis the ends take on the appearance of button holes the narrow pointed end of the button hole being attached to the body of the organism the wider and rounded end being free. During motion secondary curves often appear and disappear in rapid succession giving the organism a resemblance to a C O S or other curved letter. The primary spirals are absolutely regular and remain intact throughout all the various contractions executed by the organism as a whole. The leptospiræ are widely distributed in water and can be easily demonstrated in it by simple cultural methods.



(Hindle 1925) They are found in the tissues of patients with Weil's disease and other leptospiral infections. In length they vary from about  $6-85\ \mu$  but are generally about  $9-12\ \mu$  their thickness is only about  $0.1\ \mu$ . The length of the middle portion is variable but the hooked ends are always of about the same size suggesting that growth may occur only in the middle portion (Zielzer 1925). No internal structure can be distinguished in electron micrographs nor can flagellum-like bodies similar to those observed in some of the treponemata be seen (Morton and Anderson 1913).

The brief account that has been given of the different groups is necessarily dogmatic and it may well be that further work will necessitate a revision of our present classification.

### GENERAL CHARACTERISTICS OF THE SPIROCHÆTES

**Morphology**—With the exception of the structures seen in electron micrographs of some of the treponemata (see p. 908) spirochætes are generally regarded as being without flagella they are nevertheless motile. Three kinds of movement are generally described: (1) *Movements of flexion* in which the whole organism undergoes a change in shape. As a rule the natural shape of a spirochæte at rest is straight but during movement all sorts of twists and turns may develop one form following another in rapid succession but each one tending to return to the normal straight form. During these movements of flexion the primary spirals remain unaltered. (2) *Movements of rotation around the long axis* these are difficult to see unless the ends of the organism are bent at an angle to the main axis when the rotatory movement is especially apparent. When the rotation is very rapid and when as in *Leptospira* the ends are hooked the spirochæte may take on the appearance of a spiral thread with a button hole at each end. (3) *Movements of translation* by which the organism changes its position moving from one place to another. This change in place is probably dependent upon the rotatory movement which acts like a propeller driving the organism forwards or backwards according to the direction of the rotation.

Different spirochætes vary greatly in their activity some for example exhibit very active movements of flexion lashing furiously in various directions but making very little progress from their original position others dart rapidly hither and thither rendering their ocular pursuit almost impossible.

Multiplication occurs by transverse fission. It is not clear however whether the organism always divides in the middle giving rise to two shorter spirochætes of equal length or whether division occurs at times asymmetrically. According to Seguin's (1930) observations on *Trep. calligyrum* and *Trep. gallinarum* and Manouelian's (1940) observations on *Trep. pallidum* asymmetrical division at least in cultures is not uncommon the shorter cell seen after separation having only two one or even less than one complete spiral. Again whether the large knob-like bodies seen attached to some of the treponemata play any part in reproduction is still doubtful.

Generally speaking the spirochætes are more difficult to stain than the bacteria. Methylene blue which is usually a satisfactory bacterial stain leaves many of the spirochætes unstained. It has been suggested that the spirochætes are devoid of nucleoprotein and to the absence of this substance has been ascribed the failure of the treponemata and all the known leptospiræ to stain with this dye. On the other hand the fact that an organism does stain with methylene blue is no proof

that it contains nucleic acid (see Zuelzer 1925). The spirochætes that stain with methylene blue are generally coloured blue by Giemsa, whereas those that do not are generally coloured red. But the exact tint that results from Giemsa's stain depends to some extent on the medium in which the organism is grown, thus in the blood, *Trep. recurrentis* stains blue with occasional reddish granules, whereas in culture it stains red. In the larger spirochætes it is possible by such stains as Giemsa's and iron hæmatoxylin to bring out the finer details of structure, but in the smaller ones this is practically impossible. The reaction to Gram's stain is negative, though this stain is rarely used in practice.

For the demonstration of spirochætes in tissue-sections Levaditi's method is one of the most successful. It depends on the ability of the organisms, when treated with silver nitrate followed by reduction with a formal pyrogallie acid mixture to become impregnated with metallic silver, and therefore to appear black. For the demonstration of spirochætes in films the Fontana method of silver impregnation is most useful.

Many species, particularly the leptospiræ, are able to pass through the usual bacterial filter candles. This property they owe to their extreme tenuity. Exact measurements by Hindle and Elford (1933), made with graded collodion membranes show that the width of *Trep. pallidum* is about  $0.2 \mu$  and of *Lepto. biflexa*  $0.1 \mu$ . Use is often made of their filtrability to separate them from contaminating bacteria.

**Cultivation.**—The cultivation of spirochætes *in vitro* is not so simple as that of most bacteria. Nearly all the methods that have proved successful involve the use of a medium containing native animal protein, such as blood, serum or ascitic fluid. Whether this acts chemically as a nutrient material or physically as a protective colloid preventing the organisms from being poisoned by the products of their own metabolism, is not clear. A further requirement for many spirochætes is a low oxygen pressure: this is obtained by culturing them in narrow tubes containing a high column of medium, or by adding a piece of sterile kidney which produces a zone of anaerobiosis in its neighbourhood (Theobald Smith's method). It is usual to cover the medium with a layer of paraffin oil, this was at first believed to prevent the ingress of oxygen, but it appears now that this explanation is incorrect. The beneficial action of the oil probably depends partly on the prevention of evaporation of water from the medium, and partly on the prevention of the loss of  $\text{CO}_2$  from the medium, which would otherwise become progressively more alkaline (Gates and Olitsky 1921, Klingler and Robertson 1929). Spirochætes have been cultivated mainly in liquid or in semi-solid media. Colony production on the surface of solid media under aerobic or anaerobic conditions has been reported by a few workers (Twort 1921, Gates 1923, Aleksjanzew Malkin 1933, Seguin and Vincent 1933), but in general little success has been obtained with this method.

Multiplication occurs rather slowly, and may not be evident for a week or more. When a clear medium is used, growth may be evident from the appearance of a faint turbidity, but generally microscopical examination is necessary, particularly for motility and signs of transverse division. Most spirochætes seem to prefer a slightly alkaline medium, about pH 7.2-7.6. Subculture has to be performed every few days as a rule, the exact time depending on the particular organism. In young cultures the organisms are actively motile and under dark-ground illumination appear uniformly refractile, but in older cultures when degeneration sets in, they lose their motility, tend to agglutinate into clumps, and become granular,

the granules, some of which result from a change in the cytoplasm within the organism, and some of which are probably particles of the culture medium adhering to their exterior, are more highly refractile than the rest of the spirochæte, and show up as bright, glistening points. Some observers consider that these granules are not the result of degeneration, but are analogous to bacterial spores, affording a means of continuing life under unfavourable environmental conditions. The evidence in favour of this view is not convincing.

The isolation of spirochætes in pure culture from material in which they are accompanied by bacteria often presents great difficulty. Various methods may be tried, such as rapid subculture, making use of the ability of some species to grow out from the line of inoculation in stab cultures, picking deep colonies, choosing media made selective by the addition of a dye or some other substance, filtration, animal inoculation and so on (see Seguin and Vincent 1938, Schuffner 1940, Wichelhausen and Wichelhausen 1942).

With some organisms such as *Leptospira icterohæmorrhagiae* *in vitro* culture is remarkably successful, but with organisms such as *Trep pallidum* and *Trep recurrentis* it is not so satisfactory. For this reason these organisms are generally preserved by *in vivo* culture, that is to say, they are injected into a susceptible animal, which henceforth becomes a chronic carrier, and which can be drawn upon at will for a fresh supply of infective material. A modification of the animal inoculation method, which has proved successful in the cultivation of relapsing fever spirochætes and *Lepto icterohæmorrhagiae* is growth in the developing chick embryo (see Morrow *et al* 1938, Chabaud 1939, Oag 1939, Soule 1942).

**Resistance and Metabolism**—The resistance of spirochætes to inimical agencies is no greater and generally less than that of the vegetative bacteria. Dry heat, moist heat, and desiccation prove quickly fatal, as do comparatively low concentrations of the chemical disinfectants. It is generally stated that spirochætes are lysed by a 10 per cent solution of sodium taurocholate (see Prowazek 1907), and all, with the exception of the leptospiræ, by a 10 per cent solution of saponin (Noguchi 1917). Some of the highly parasitic members, such as *Trep pallidum*, are unable to survive outside the animal body for more than an hour or two. Indeed this particular organism is extremely susceptible to heat, being destroyed in an hour at 41.5° C. Advantage is now being taken of this property to sterilize the organisms in the tissues by exposure to fever heat temperatures (see Chapter 81). Spirochætes in infected tissues may be preserved alive for several months by keeping them in the frozen state, preferably at about -78° C (Jahnel 1937, Turner 1938).

Practically nothing is known about the metabolism of spirochætes. Scheff (1935), who made observations on two strains of *Trep pallidum*, one of *Trep recurrentis*, and one of *Trep anserinum*, stated that, when these organisms were grown in a medium containing glucose, they broke down the sugar to lactic acid and CO<sub>2</sub> without using up any of the oxygen.

**Antigenic Structure**—Antigenically, spirochætes behave very much as bacteria. They give rise on injection to agglutinins, spirochætoxicins, spirochætolysins, and protective antibodies. Agglutination tests are often conducted by the microscopic method, and observations made on the loss of motility and the clumping of the organisms, which may occur in radiate fashion. Similarly the destruction and lysis of the organisms that frequently follow contact with a highly immune serum are generally watched under the microscope. The macroscopic method, however,

is now used extensively for study of the leptospiræ, and antigenic types are determined by cross-agglutination, and sometimes cross absorption, tests. Protective bodies in serum are tested for in the usual way by animal inoculation.

A valuable method of study is afforded by the test described by Rieckenberg (1917), known sometimes as the *thrombocytobarin reaction* or the *adhesion phenomenon*. It depends on the fact that if a suspension of blood platelets or bacteria is added to a mixture of a spirochæte and its specific antiserum, and the preparation is observed under dark-ground illumination after preliminary incubation at 30° C for 20 minutes, the platelets or bacteria are seen to have become adherent to the spirochætes in the form of small clumps. In the presence of a non-specific serum no such clumping or adhesion occurs. By this reaction it is possible to distinguish rapidly between two such closely allied forms as *Lepto icterohæmorrhagæ* and *Lepto hebdomadis* (Brown and Davis 1927). Living motile spirochætes are required and fresh complement must be present. The specific antibody is destroyed by heating to 72° C for 30 minutes (Inoue 1930). Pfeiffer's test and cross immunity protection tests in animals are likewise of value in distinguishing between closely allied species, and even between variants of the same species.

**Pathogenicity**—The virulence of spirochætes appears to be subject to considerable variation. Many members are strictly parasitic and give rise to infections in man or animals, while others are saprophytic and appear to be devoid of any pathogenic effect. There is evidence, however, that the virulence of the parasitic members may undergo change as the result of residence in the body of the host. In relapsing fever, for example, the strains that appear in the blood at the second or third relapses may differ antigenically from the strain responsible for the original attack, and by virtue of this change are able to multiply in the tissues of a host that has become immunized to the original parent strain. Again, strains of certain spirochætes such as *Trep pallidum*, may be brought by passage to grow readily in an animal which at first resists their invasion. *Trep pertenue*, the organism that is responsible for yaws, is regarded by many observers as merely a variety of *Trep pallidum* which, by residence in the negro, has developed dermatropic affinities (Parham 1922). Residence outside the body of certain parasitic strains may apparently be accompanied by a fall in virulence, which renders them indistinguishable from naturally saprophytic strains. Thus *Lepto icterohæmorrhagæ*, if kept in water, may become indistinguishable from *Lepto biflexa* (Zuelzer 1925). Whether the virulence of naturally saprophytic species ever becomes increased so as to render them pathogenic for man is doubtful. Baermann and Zuelzer (1927, 1928) have brought a considerable amount of evidence to show that *Lepto biflexa* may be transformed by repeated animal passage into *Lepto icterohæmorrhagæ*. Their findings, however, are not in harmony with the experience of most other workers nor with the epidemiological picture of Weil's disease, and it seems probable that such instances are due to the recovery in virulence of a real *icterohæmorrhagæ* strain.

Though the spirochætes may be classified into the free-living, the commensal, and the pathogenic types, it must be realized that there is no sharp line of demarcation between the three groups. An organism that is pathogenic in one animal may be purely commensal in another, and an organism that is highly pathogenic at one time to a particular host may at another give rise to no more than a latent infection.

We append a description of some of the members that are of most interest to the student of medical and veterinary bacteriology.

*Treponema recurrentis*

*Isolation*—Observed by Obermeier (1873) in the blood of patients with European relapsing fever

*Morphology*—Actively motile spiral organisms varying considerably in length but usually 10–20  $\mu$  long. Series of 5–10 fairly regular but loose primary waves, each spiral is 2–3  $\mu$  long and about 1  $\mu$  in amplitude (Fig 215). The width is usually given as 0.2–0.3  $\mu$  (Wenyon 1926, Hindle 1931) but this is probably an under estimate. Personal observations on the organisms in blood have suggested that 0.4  $\mu$  more nearly represents their true diameter.<sup>1</sup> This is supported by the observations of Tilden (1937) who found that the limiting pore size for filtration through Eilford's gradocol membranes was 0.57  $\mu$ . After transverse fission the two new organisms may remain connected by a remnant of the periplast. Stains purplish red with Giemsa. Organisms are said to be shorter and thinner in young culture, thicker and longer in old (Plotz 1917).

*Cultivation*—First successful cultivation reported by Noguchi (1912c) who seeded a few drops of citrated blood from the heart of an infected mouse or rat into a tube containing 15 ml. of unheated and unfiltered aseptic or hydrocele fluid and a small piece of sterile rat bit's kidney. The blood was taken from the animal 48 to 72 hours after inoculation. Multiplication of the spirochaetes in the cultures was visible in 2 to 3 days and reached its maximum about the 7th to the 9th day. No change was noticeable in the medium, but actively motile spirochaetes could be found in every field arranged either singly in chains or in masses. After about the 9th day a sudden decrease in their numbers occurred and spherical bodies and irregular protoplasmic masses appeared, indicating that the organisms were undergoing degeneration. Subcultures were most successfully made on the 4th to the 9th days. Other workers (Plotz 1917, Kligler and Robertson 1922, Sinton 1924, Lapidari and Sparrow 1928, Yuan Po 1933, Scheff 1935) using Noguchi's technique or more often a modification of it have claimed to cultivate relapsing fever spirochaetes *in vitro* but the results appear to have been very irregular (see Moroder 1929), no one appears yet to have succeeded in establishing a culture that can be continued indefinitely in the laboratory (see Soule 1942). On the other hand growth can readily be obtained by inoculation of the chorio-allantoic membrane of the developing chick embryo (Chabaud 1939, Ong 1939, Soule 1942). The spirochaetes produce no change in the membrane itself but invade the blood of the embryo where they may be demonstrated by the usual methods. Some strains prove fatal to the embryo, others do not.

*Resistance and Metabolism*.—Resistance is apparently similar to that of the more susceptible vegetative bacteria. Said to remain viable in clotted blood for 6 days at room temperature and for at least 100 days at 0° C. (Wynns and Beek 1935). Little is known about metabolism. According to Scheff (1935) glucose is broken down with production of lactic acid and CO<sub>2</sub> but no oxygen is used up. A moderate partial pressure of oxygen however is required for growth, there is no multiplication under strict anaerobic conditions.

<sup>1</sup> Mr J. E. Barnard, F.P.S., has kindly measured for us a strain of *Trep. duttoni*, he finds it to be 0.35  $\mu$  in diameter.



FIG 215—*Treponema duttoni*

In film of blood. In one place the spirochaetes show a tendency to agglutination in rosette form. Giemsa ( $\times 1000$ ) [From specimen kindly supplied by the late Prof J. G. Thomson.]

invasive properties (Ellermann 1907) It is very frequently found in association with a characteristic fusiform bacillus likewise described by Vincent (1896 see Chapter 18) It has been suggested (Tunnichiff 1906) that *Trep vincenti* and the fusiform bacillus represent two phases of the same organism, but the balance of evidence is definitely against this view

### *Treponema pallidum*

*Isolation*—Described by Schaudinn and Hoffmann (1905) who observed it in chancres and inguinal glands of syphilitic patients (see Schauberg and Schlossberger 1930)

*Morphology*—Thin, delicate spirochæte with tapering ends Its length varies from 4–14  $\mu$  and its breadth is about 0.2  $\mu$  It contains a number of regular primary spirals which appear rather sharp and angular and each of which is a little over 1  $\mu$  in length During motion secondary curves may appear and disappear in rapid succession but the primary spirals remain undisturbed The organism is actively motile the movements were originally described by Schaudinn and Hoffmann (1905) as being of 3 types (1) rotation round

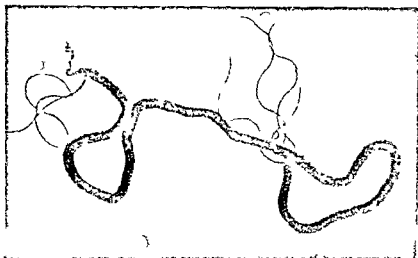


FIG. 216.—Electron micrograph of *Treponema pallidum* showing what appear to be flagella ( $\times 21,000$ ) [kindly supplied by Dr. Stuart Mudd]

the longitudinal axis, (2) backward and forward movements (3) flexion movements of the whole body resulting in the production of secondary waves The rotation or spinning movement is responsible for the backward or forward movements the primary spirals act like the blades of a propeller and drive the organism forward No flagella can be demonstrated by ordinary methods, but according to Mudd, Polevitzky and Anderson (1943) and Wile and Kearney (1943) flagella often four in number may be seen in electron micrographs distributed along the sides or near the ends of the organism In cultures the morphology is not so regular as in the animal body, Noguchi (1912c) has described three types of *pallidum*—the thicker the normal and the thinner type Whether these types are constant or merely represent fluctuations round a mean is not known, Noguchi favours the former view The organism stains rose red with Giemsa The organisms are held back by gradocol membranes having a pore size of 0.4  $\mu$  their narrowest diameter is therefore about 0.2  $\mu$  (Hindle and Elford 1933 Tilden 1937)

*Cultivation*—Schereschewsky (1909) was the first to cultivate *Trep pallidum* *in vitro* but he did not succeed in obtaining pure cultures Noguchi in 1911 was the first to do this He used a medium of serum water to which a piece of sterile rabbit tissue had been added,

the medium was seeded with a fragment of syphilitic rabbit's testicle, and the whole was covered with a layer of liquid paraffin. Incubation was carried out anaerobically at 37° C. The primary mixed culture was later purified by growth in serum agar slabs containing fresh rabbit tissue, in this medium a hazy zone around and above the tissue became perceptible in about 3 days due to proliferation of the spirochætes, by subculture from this zone he eventually succeeded in obtaining pure cultures of the organism. The following year Noguchi (1912b) succeeded in cultivating *Trep. pallidum* directly from human lesions. The medium he used consisted of a mixture of 2 parts of nutrient agar and 1 part of ascitic or hydrocele fluid, put up in tubes 20 X 2 cm in size, each containing at the bottom a piece of sterile rabbit kidney or testicle. Material from a chancre, condyloma, or skin papule was inoculated into the tube, and was covered with liquid paraffin. The organisms produced a slight haze round the kidney and could be picked off for purification. In fluid medium growth occurred very slowly and continued for several weeks. The pure cultures were inoculated into monkeys, and proved to be pathogenic. According to Gates (1923), surface colonies may be obtained on 6 per cent. rabbit blood agar plates incubated anaerobically. Colonies are said to be well developed in about a week at 37° C., and to be surrounded by a zone of complete hæmolysis. Other media have since been used with apparent success (Gates 1923, Weiss and Wilkes Weiss 1924, Hoder 1930, Aleksanzew Malkin 1933). It must be pointed out, however, that several reputable workers have entirely failed to cultivate *Trep. pallidum*. Among them is Jahnke (1934) who maintains that the so-called cultures of this organism are in fact cultures of a saprophytic spirochæte (see also Gohring 1940). According to Kast and Kolmer (1940), no one has yet succeeded in devising an *in vitro* method of cultivation in which the virulence of the organisms for the rabbit is preserved. The difficulty in practice of obtaining *in vitro* cultures is so great that for preserving the organism it is usual to employ *in vivo* methods. Brown and Pearce (1921b) found that if rabbits are infected with syphilis, the organisms are carried to the lymphatic glands and remain there indefinitely. When the strain is required for use, a popliteal gland is excised, ground up in a mortar with saline, and injected intratesticularly into fresh rabbits. Kolle and Schlossberger (1925) have moreover shown that *Trep. pallidum* remains alive in the tissues of mice for an indefinite period and can be recovered at any time from the glands spleen, or brain. Using this method they made three passages through mice in 19 months and found that the organisms remained fully virulent for rabbits.

**Resistance**—Very susceptible to heat. According to Boak, Carpenter, and Warren (1932), saline suspensions of infected rabbit testicle are sterilized by exposure to 39° C for 5 hours, 40° C for 3 hours, 41° C for 2 hours, and 41.5° C. for 1 hour. Dies out rapidly in stored blood unless frozen, so that the chances of transmitting syphilis with stored blood plasma or serum are very small (see Selbie 1933).

**Antigenic Structure**—Little known. Noguchi and Akatsu (1917) using agglutination and complement fixation, obtained evidence of an affinity of *Trep. pallidum* to *Trep. calligyrum*. They also observed a certain amount of heterogeneity between different strains of *pallidum*. There is said to be a strain specificity in cultures of *pallidum* (see Georgi et al. 1929), but in view of the serious criticisms that have been made on the supposed cultivation of this organism, it would be dangerous to lay too much stress on this statement.

### Pathogenicity of *Treponema pallidum* for Animals.

**RABBITS.**—Haensell in 1881 was the first to produce keratitis in rabbits by inoculation of syphilitic material into the anterior chamber of the eye. His observations were neglected for over 20 years, when they were confirmed by Bertarelli in 1906, the syphilitic nature of the lesions was proved by the demonstration in them of *Treponema pallidum*. The receptivity of the rabbit's eye has been confirmed by numerous workers. Bertarelli (1907, 1908) moreover showed that it was possible to carry over syphilis from one animal to another. According to Uhlenhuth and Mulzer (1913) inoculation of a small piece of

syphilitic rabbit's cornea into the anterior chamber of the eye of a fresh rabbit is followed by complete healing of the local wound in 5 to 10 days. After 3 to 6 weeks, as a rule, pericorneal congestion commences, followed by pannus and keratitis. The keratitis increases to an acme, after which retrogression and healing occur, this process may take weeks or months to complete, and may be interrupted by a relapse. Only a certain proportion of rabbits develop keratitis. The lesion is very much easier to produce by inoculation of rabbit than of human syphilitic material. Successive passages of the virus through the eye of rabbits resulted in an increase of virulence manifested by a reduction in the incubation period from 6 to 8 weeks to 4 to 5 weeks.

Syphilis may also be conveyed to rabbits by inoculation into the testicle, this method of transference was first successfully used by Parodi (1907). As with ocular injection, the implantation of human syphilitic material gives much less constant results than of that from the rabbit. Uhlenhuth and Mulzer (1913), for example, inoculated 27 rabbits intratesticularly with human syphilitic material—the juice from primary chancres—and obtained only 5 positive reactions. But after 15 passages through the rabbit the virulence had so increased that inoculation was almost uniformly successful, and the severity and extent of the disease were correspondingly greater. Brown and Pearce (1920a), using the method of intratesticular inoculation of ground up syphilitic rabbit testicle suspended in saline, were likewise uniformly successful in producing the disease. After an incubation period of about 3 to 4 weeks the testicle commences to swell, and soon reaches the size of a pigeon's egg, the inflammation also affects the epididymis and cord. Sometimes a small superficial erosion may develop at the site of inoculation, covered with a dry yellowish brown adhesive crust, or an actual chancre may appear. According to Brown and Pearce (1920a) the testicular reaction pursues a cyclic or relapsing course, periods of active progression alternating with periods of quiescence or retrogression, these phases apparently correspond with the variations in the number of spirochetes in the lesion. The length of time that the testicle is inflamed varies, the lesion may disappear in 6 weeks, or it may last for over a year.

The method of intracutaneous or subcutaneous injection of rabbit syphilitic material into the scrotum, introduced by Tomaszewski (1910), gives rise after an incubation period of about a fortnight to a typical primary chancre with a central necrotic area and indurated edges. Sometimes a diffuse lesion of the scrotum follows. These scrotal lesions are invariably accompanied by marked inguinal lymphadenitis. The scrotal infection may spread, and numerous secondary lesions develop, lasting from 1 to 16 months.

Following on scrotal or testicular infection, generalized lesions may develop affecting practically any structure of the body (Brown and Pearce 1920b, 1921a, Brown *et al.* 1921). Thus there may be papular or erythematous eruptions on the skin, sometimes appearing in successive crops, granulomatous lesions of the skin passing on to ulceration, alopecia, onychia, and paronychia, necrotic and ulcerative lesions of the mucosae and mucocutaneous borders, localized lesions of the periosteum, bone, cartilage, tendons, and tendon sheaths, including such typical manifestations as destruction of the nasal septum and separation of the epiphyses, conjunctivitis, keratitis, and iritis. Generalized lesions of syphilis may also be produced by the intravenous or intracardial injection of rabbits a few days old (Uhlenhuth and Mulzer 1913). For discussion of immunity in rabbits see p. 1165, Chapter 51.

**MONKEYS.**—The experiments of Metchnikoff and Roux (1903, 1904a, b, 1905) amplified the earlier observations of Klebs in 1875-77 (see Klebs 1932), and showed that syphilis might be transmitted to the anthropoid apes and with less certainty to monkeys. Of the apes the chimpanzee appeared to be the most susceptible. Altogether they inoculated 22 chimpanzees (*Troglodytes niger* and *T. cilius*) with syphilitic material, either of human origin or derived from experimental animals, and succeeded in producing disease in all of them. Inoculation was performed by scarification of the genitals, the thigh or the eyebrow. After an incubation period of 15 to 49 days, generally 4 weeks, a primary chancre developed at the site of inoculation, and was followed in a few days by swelling of the focal lymph glands. Many of the animals developed lesions of secondary syphilis.



3 to 9 weeks after the appearance of the chancre, these comprised a papular eruption on the skin, palmar psoanosis, mucous plaques of the lips, tongue, and palate and enlargement of the spleen. Occasionally very severe syphilis developed, accompanied by alopecia, skin eruptions, emaciation, paresis of the hind limbs, or even death. No lesions of tertiary syphilis were ever found, but it is to be noted that most of the animals died of bronchopneumonia before they had been many weeks under observation and in these tertiary lesions had no time to develop. In macaques secondary lesions were never observed. According to Uhlenhuth and Mulzer (1913) apes are more difficult to infect than rabbits. These workers were successful in conveying human syphilis to rabbits, from rabbits to monkeys, and from a monkey back to rabbits.

**OTHER ANIMALS.**—For studying syphilitic lesions, chimpanzees and rabbits are the most useful experimental animals. Infection can however be conveyed to certain other animals, such as pigs, guinea pigs, rats and mice. According to Tani, Kaki-hita, and Saito (1930) intratesticular inoculation of guinea pigs gives rise to no obvious lesions, but intracutaneous inoculation particularly into the penneal fold, is followed in about 11 days by the development of a swelling which persists for about 7 weeks (see also Mulzer and Hahn 1930). In rats and mice a symptomless infection is usually produced, similar to that often seen in guinea pigs. The spirochætes remain latent in the tissues for months, as can be shown by inoculation of rabbits (Kolle and Schlossberger 1926 1928). Occasionally however, a local chancre may be produced by inoculation of the scarified skin of the ano-scrotal region (Bessemans and de Potter 1930 1931).

### *Treponema cuniculi*

First observed by Bayon (1913). Responsible for a disease known as "rabbit syphilis." Morphologically very similar to *Trep. pallidum* but tends to be slightly longer and thicker. According to Noguchi (1922) dimensions are length ~30  $\mu$  average 13  $\mu$  width 0.25  $\mu$  length of spirals 1.12  $\mu$  amplitude of spirals 0.6-1.0  $\mu$ . Like *Trep. pallidum* it stains rose-red with Giemsa. Inoculation of infective material on to the scarified skin of the genital region is followed, after an incubation period of 2 to 8 weeks, by characteristic lesions (see Chapter 81).

### Notes on certain other *Treponemata* found in the Human or Animal Body

*Treponema pertenue*.—Described by Castellani (1905). Responsible for yaws. Morphologically indistinguishable from *Trep. pallidum*. Exact relation to this organism not yet fully understood.

*Treponema refringens*.—This organism was first described by Schaudinn and Hoffmann (1905) in their original report on the discovery of *Trep. pallidum*. It was observed in cases of syphilis complicated with such lesions as balanitis, ulcers, and papillomata, and in non-syphilitic lesions such as gonorrhœal papillomata. Noguchi (19123) cultivated it from a condyloma. According to him it grows luxuriantly in the deeper part of an ascitic agar tube, forming hazv colonies, denser than those of *Trep. pallidum*, which gradually extend from the deeper parts of the tube to the more superficial. It is an anaerobe—no growth occurs within 2 cm. of the surface. Growth becomes visible in 4 days at 3° C. and proceeds for some weeks. The addition of fresh tissue is not essential. In culture the organism is 6-24  $\mu$  long by 0.3-0.5  $\mu$  broad, the middle part of the organism is wavy but the two extremities are more regularly and deeply curved. The ends are pointed. It is non-pathogenic for rabbits and monkeys.

*Treponema calligyrum*.—This organism was observed by Noguchi in 1913 in two cases of condyloma, one syphilitic, the other not. Pure cultures were obtained by the ascitic agar stab method. In this medium growth is similar to that of *Trep. refringens*, the hazv colonies are more dense and diffuse than those of *Trep. pallidum*. In culture the organisms are 6-14  $\mu$  long by 0.3-0.4  $\mu$  wide. The primary spirals are regular and deep, the length of each spiral is 1-6  $\mu$ , and the amplitude 1 to 1.5  $\mu$ . The apex of the curve is not

sharp and pointed as in *pallidum*, but more or less rounded. It is non pathogenic for rabbits and monkeys.

*Treponema phagedenis*.—This organism was cultivated by Noguchi (1912f) from a phagedenic ulcer on the labium of a woman. Growth occurred in ascitic agar medium under anaerobic conditions in the absence of kidney tissue. In the original lesion the spirochaetes were 4–30  $\mu$  in length and 0.75  $\mu$  in thickness, in culture their length was less variable, being 10–15  $\mu$ . The number of spirals varies from one to eight, and there is great variation in the length of each spiral. Some organisms appear nearly straight. The ends are pointed, but not drawn out. Other spirochaetes have been described in ulcerative lesions round the genital regions, such as *Trep. balanitis*, *Trep. pseudo pallidum*, and *Trep. gangrenosa nosocomialis* (see Noguchi 1912f).

*Spirochaetes in the Human Mouth*.—Spirochaetes of different types have been described in the mouth, they can generally be seen in scrapings from between the teeth. Some times organisms morphologically indistinguishable from *Trep. pallidum* are found. Noguchi (1912a) succeeded in cultivating what he regards as two separate species. *Trep. microdentium* is a short spirochaete about 3–4  $\mu$  long by 0.25  $\mu$  wide, having shallow rectangular curves of constant size. The ends are drawn out and pointed. In culture it is said by Seguin and Vincent (1933) to be an actively motile organism, 4–7  $\mu$  long, having 6–12 well defined regular spirals. In serum agar tissue medium it forms a haze near the bottom of the tube, gradually becoming denser and spreading upwards till it is within 2–3 cm. of the surface. Growth is anaerobic. *Trep. macrodentium* is a larger organism, varying from 3–8  $\mu$  long by 0.7–1.0  $\mu$  broad in young cultures, and having 2–8 irregular shallow curves; the ends taper off abruptly. In older cultures the organisms are longer and thinner. In serum agar tissue medium growth occurs under anaerobic conditions in the form of a faint almost transparent haze. For methods of isolating and culturing the mouth spirochaetes, reference may be made to papers by Séguin and Vincent (1933), Kist and Kolmer (1940), and Wichelhausen and Wichelhausen (1942).

Vincent and Daufresne (1934), working mainly with pure cultures, have provisionally classified the mouth spirochaetes into groups, which they label A to G. Group B corresponds to *Trep. microdentium* and Group F to *Trep. macrodentium*.

*Treponema cobayae*.—Found by Knowles and Basu (1935) in the blood of guinea pigs. Blood parasite belonging to the relapsing fever group. Thin, delicate spirochaete, 13.5–23  $\mu$  in length, with finely tapering ends, average length of spirals 3.6  $\mu$ . Can be cultivated in Galloway's medium. Inoculation of guinea pigs with infected blood is followed, after an incubation period of 2 to 6 days, by a febrile disease accompanied by the presence of spirochaetes in the blood. Fully virulent strains kill 30–60 per cent. of inoculated animals. Relapses may occur in animals that recover from the first attack. White rats and rabbits are also susceptible to infection.

Blood spirochaetes have been described in other animals, such as the rabbit and the mouse (see Knowles and Basu 1935).

### *Leptospira icterohæmorrhagiæ*

*Isolation*.—First adequate description given in 1915 by Inada and his colleagues in Japan (see Inada *et al.* 1916), who observed it in the blood and tissues of patients with Weil's disease.

*Synonyms*.—*Spirochaeta icterohæmorrhagiæ*, *Spirochaeta icterogenes*.

*Morphology*.—Very delicate organism whose morphology can be studied satisfactorily only by dark ground illumination. The spirals are too fine to be properly resolved in stained preparations. In length it is about 6–12  $\mu$ , and 0.1–0.15  $\mu$  in thickness, forms as short as 4  $\mu$  and as long as 25  $\mu$  may sometimes be observed. It contains a number of perfectly regular closely wound spirals, each of which is about 0.5  $\mu$  long or even less, and has an amplitude of 0.5  $\mu$ . Near the extremities the spirals become even closer. Secondary waves commonly appear during motion, but the spirochaete has a marked

tendency to straighten itself out again. Apart from the primary spirals which are set more closely than in any other group of spirochaetes, the most characteristic feature of *Leptospira icterohaemorrhagiae* is its sharp, tapering, hooked ends, which are set at an angle to the main axis giving the whole organism a resemblance to the letter C or S. During the rotation that occurs when the organism is moving, these hooked ends are whirled round so rapidly that the organism appears to be furnished with a button hole or an eye splice at each extremity (Fig. 217). In fluid media the spirochaetes may become entangled with each other and give rise to the characteristic picture of a 'nest'. This appears as a highly refractile ball composed of hundreds of interlaced organisms, some of which project radially from the circumference (Taylor and Goyle 1931). In dry fixed films of blood or urine all sorts of forms may be seen, bearing a resemblance to the letters C, S, I, or b. Degeneration forms with thick, blunt, straight ends are not uncommon. According to Kaneko and Okuda (1917), the morphology of the spirochaetes in man is less regular than in the guinea pig, they are often shrunken and atrophic, of varying thickness with greater rigidity and less regular curves, they may show circumscribed thickenings at two or three points or they may resemble chains of granules. These irregular forms may perhaps result from the action of immune bodies on the organisms.

In suitable preparations the spirochaetes may be stained by Giemsa or by one of the silver impregnation methods. In stained films the primary spirals are not visible. Several observers have found that leptospirae may pass through Berkefeld candles (Inada *et al* 1916, Bauer 1927, Buchanan 1927, Dimitroff 1927) but the results are variable. According to Hindle and Elford (1933), the organisms are held back by collodion membranes

having a porosity of  $0.25 \mu$ , this suggests that their diameter is  $0.1 \mu$ .

**Cultivation.**—*Leptospira icterohaemorrhagiae* was first cultivated by the Japanese workers (Inada *et al* 1916) in Noguchi's ascitic fluid kidney medium. Subsequently Noguchi (1917) devised other media that were simpler to make and more satisfactory in practice. The first consists of rabbit serum 1 part, Ringer's solution 3 parts and citrated rabbit plasma 0.5 parts, the medium should be put up in tubes about  $\frac{1}{4}$  inch in diameter, and may or may not be covered with liquid paraffin. The second medium is similar to the first, but 0.5–1.0 parts of slightly alkaline 2 per cent agar are added, at a temperature of  $60$ – $65^{\circ}\text{C}$ , the whole medium being well mixed. From this semi-solid medium the citrated rabbit plasma may be omitted if necessary. The Noguchi-Wenyon medium is likewise satisfactory.



FIG. 217.—*Leptospira icterohaemorrhagiae*.

Diagrammatic drawing showing primary and secondary spirals, and hooked and button hole ends. (After Wenyon.)

it is made by mixing 9 parts of saline with 1 part of 2 per cent. nutrient agar, and adding 20 drops of fresh rabbit's blood to each 10 ml of the autoclaved medium cooled to  $50^{\circ}\text{C}$ , the tubes are not shaken. Very good results are obtained with Fletcher's (1928) medium. This is prepared by heating a 12 per cent. solution of rabbit serum in distilled water to  $50^{\circ}\text{C}$ , adding 6 ml of 2.5 per cent nutrient agar to every 100 ml of serum water mixture adjusting the reaction to pH 7.4 tubing in 5 ml quantities, and sterilizing at  $56^{\circ}\text{C}$  for 1 hour on 2 successive days. According to Gardner (1943a) the simplest and most satisfactory medium consists of a 12 per cent solution of heat inactivated Seitz filtered rabbit serum in glass-distilled water. For inoculation, 0.1 ml. of infected

guinea pig blood or liver, or rat kidney suspension should be used. In subculturing, 0.5-1.0 ml should be carried over to 5 ml of fresh medium. The tubes should be incubated at 25-30° C., growth occurs at 37° C., but degeneration rapidly sets in. Subcultures should be made every 4 to 6 weeks, and kept at 25° C. The optimum pH for growth is 7.6. The organism is aerobic, in Noguchi's serum media, growth occurs at the top, giving rise to a slight haze, which stops abruptly a few centimetres from the surface. *Lepto icterohæmorrhagiæ* can also be cultivated on the chorio allantoic membrane of the developing chick embryo (Morrow *et al* 1938, Chabaud 1939). The organisms become demonstrable in the blood of the allantoic artery in 4 or 5 days, the embryo generally dies within 7 days. Pure cultures of *Lepto icterohæmorrhagiæ* may be obtained from contaminated material by inoculating 0.5-1.0 ml intraperitoneally into a guinea pig, withdrawing blood by heart puncture ten minutes later, and culturing it in a suitable medium (Schöffner 1940), the leptospiræ invade the blood more rapidly than the accompanying bacteria.

**Resistance.**—*Leptospira icterohæmorrhagiæ* is killed by moist heat at 50-55° C in half an hour, it can withstand freezing. It is very sensitive to acid, being destroyed by human gastric juice in 30 minutes, it will not grow in an even slightly acid medium. The organisms are rendered motionless in 10 to 15 minutes by 1/2000 HgCl<sub>2</sub> and are gradually dissolved. They are rapidly destroyed by bile. In defibrinated blood kept at room temperature in the light, the organisms remained virulent for 7 days, and in decomposing liver for 27 hours (Uhlenhuth and Fromme 1916). In infected guinea pig liver kept in the ice-chest they remained virulent for 26 days (Buchanan 1927).

**Antigenic Structure.**—Though it was thought at one time that there was but a single species of *Leptospira*, namely *Lepto icterohæmorrhagiæ*, further experience has revealed the occurrence of several types differing in their antigenic structure, pathogenicity to guinea pigs and the nature and severity of the disease to which they give rise in man. Many of these types have been given specific names. The original *icterohæmorrhagiæ* species can be distinguished antigenically from other leptospiral strains that are pathogenic for human beings. Its relation to water spirochetes is still under discussion. Baermann and Zuelzer (1928) found that water strains were not agglutinated by the sera of convalescents from Weil's disease, or by the sera of animals inoculated experimentally with *Lepto icterohæmorrhagiæ*, and that sera prepared against avirulent water strains did not agglutinate Weil strains. Brown and Davis (1927), using the adhesion test (see p 912), found that *Lepto icterohæmorrhagiæ* from rats or man behaved alike, while *Lepto bifera* was antigenically distinct. Vaccination of guinea pigs with cultures of *Lepto bifera* failed to immunize them against *Lepto icterohæmorrhagiæ* (Uhlenhuth and Zuelzer 1921). According to Baermann and Zuelzer (1928), however, some water strains the virulence of which has been raised by animal passage, behave antigenically like *Lepto icterohæmorrhagiæ*. Some of the confusion is probably due to the lack of homogeneity among water strains. The majority of these are saprophytes and belong to the species *Lepto bifera*. Some of them, however, appear to be real *icterohæmorrhagiæ* strains, either in their normal virulent condition or in a degenerate avirulent condition. Passage through animals may succeed in restoring these to full virulence, rendering them indistinguishable antigenically from typical *icterohæmorrhagiæ* strains of parasitic origin. Since animals not infrequently act as leptospiral carriers, it is possible that apparent changes in antigenic structure and virulence brought about by passage are due to the isolation of an organism from the animal different from that which was inoculated. Caution must therefore be exercised in drawing conclusions from the type of evidence advanced by Baermann and Zuelzer (1928) and Zuelzer (1930).

### Pathogenicity of *Lepto. icterohæmorrhagiæ* for Animals

*Lepto icterohæmorrhagiæ* is highly pathogenic for young guinea pigs, whether given intraperitoneally, subcutaneously, cutaneously, or by the mouth. The golden hamster is likewise very susceptible (Morton 1912, Larson 1944). Rabbits, rats, and mice are only

slightly susceptible and usually remain perfectly well after inoculation (Martin and Pettit 1919). Cats, dogs, pigs, sheep, hens, pigeons, and monkeys are said to be refractory (Uhlenhuth and Fromme 1916; Martin and Pettit 1919) but later evidence suggests that *Leptospira sclero-haemorrhagiae* is responsible for some cases of infectious jaundice in the dog (Dhont *et al.* 1934).

Intraperitoneal injection of guinea pigs with 1 ml. of infected human blood or ground up rat's kidney is followed by an illness lasting for 5 to 12 days, and terminating in death. The chief symptoms of the disease are fever and jaundice. The fever commences the day after inoculation, reaches its acme in a few days, falls to normal, and finally to subnormal just before death. Jaundice first becomes visible when the temperature begins to fall—usually on the 4th or 5th day—it increases till death and is often accompanied by choluria. Anaemia and conjunctival congestion are frequent and external haemorrhages from the rectum, nose and genitals may occur. Blood counts reveal a lymphocytosis during the first few days of the disease and an anaemia (Buchanan 1937). Spirochaetes appear in the

blood about the 4th day but are not easy to find microscopically. Post mortem the animal shows generalized jaundice; there are haemorrhages into various parts of the body particularly the lungs, intestinal walls, retroperitoneal tissues and fatty tissues of the inguinal region. The haemorrhages in the lungs form irregular spots of varying size sharply demarcated from the surrounding tissue—giving the lungs a resemblance to the mottled wings of a butterfly (Inada *et al.* 1916). The spleen is enlarged and congested; the kidneys show an acute parenchymatous nephritis and capsular haemorrhages; the suprarenals are often enlarged and haemorrhagic. Histologically the chief lesions are cloudy swelling of the liver sometimes accompanied by focal necroses, acute parenchymatous nephritis, endothelial cell proliferation in the spleen and lymph glands, and haemorrhages in practically every structure of the body (Buchanan 1937). Spirochaetes are most numerous in the liver and are best demonstrated by dark-ground illumination. They occur in the spaces between the cells and when

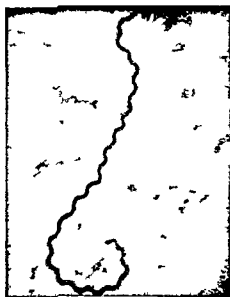


FIG. 16.—*Leptospira sclero-haemorrhagiae*.

Electron micrograph ( $\times 10,000$ ).

Kindly supplied by Dr H. E. Morton and Dr T. F. Anderson.

numerous are arranged about the cells like a garland. Their appearance is different from that seen under dark-ground illumination: they are short and thick; the primary spirals and tapering extremities are not evident and numerous irregular undulations are seen. They are found in smaller numbers in the kidneys and adrenals.

By passage from guinea pig to guinea pig the virulence of the organism can apparently be increased. Stokes (Stokes *et al.* 1911) for example found that the average time to death of animals inoculated intraperitoneally with human blood was 10 days but that when passage strains were used it was only 5 days. Noguchi (1911) likewise noted a reduction in survival time after passage of a strain through guinea pigs.

Notes on other species of *Leptospira*

*Leptospira*, *Rachmat* type—Found in cases of human infection in the Dutch East Indies. It is antigenically distinct from *Lepto icterohæmorrhagæ*, some strains are closely allied to *Lepto autumnalis* Akiyama A type (see Walch Sorgdrager 1939). It is virulent for guinea pigs, in which it usually causes jaundice. Its natural animal host remains unknown.

*Leptospira batavica*—This organism, which is sometimes referred to as the Swart van Tienen type, causes sporadic infections of human beings in Batavia, Borneo and the Celebes. It is antigenically distinguishable from other types. It is moderately virulent for the guinea-pig. Its natural host is *Rattus norvegicus*.

*Leptospira*, *Salinem* type—This type is also found in the Dutch East Indies where it causes sporadic cases of leptospiral infection in Sumatra. It is of moderate virulence for guinea pigs. It appears to be carried by the rat *Rattus brevicaudatus*.

*Leptospira hebdomadis*—Described by Ido, Ito and Wani (1918, 1919). Gives rise to the 7-day fever of Japan. Differs antigenically from *Lepto icterohæmorrhagæ*, and is less virulent for the guinea pig. It is avirulent for rats and mice in the laboratory. It is a natural parasite of the field mouse *Microtus montebellii*. Is apparently identical with the *akiyama* B type found in the autumn fever of Japan.

*Leptospira akiyama* and *Leptospira autumnalis*—The identity of these organisms is still under discussion. The first was isolated from cases of Autumn fever or *Akiyama* in Japan, the second from a disease known as *Hasamiyama*. Two types of organism from *Akiyama* have been described. The A type appears to belong to the *Rachmat* group, the B type is apparently identical with *Lepto hebdomadis* (Stefinopoulos and Hosoya 1928, Yang and Theiler 1930, Inoue 1939). The organism isolated from *Hasamiyama* is very similar to the *akiyama* A type. According to Walch-Sorgdrager (1939) *akiyama* A, *autumnalis* and *Rachmat* share a common antigen but are not identical. Both *akiyama* A and *autumnalis* are fairly virulent for guinea pigs, often producing jaundice. The natural host of *akiyama* A is a field mouse, *Microtus montebellii* of *Lepto autumnalis* a field mouse *Aperlemus speciosus*.

*Leptospira australis* A and B—These two types were isolated from cases of coastal fever in Queensland (Lumley 1937). The A type is antigenically distinct from other strains, the B type appears to be related to the *Salinem* type of the Dutch East Indies (Walch-Sorgdrager 1931). Both the Australian types are moderately pathogenic for guinea pigs and both are carried by rats—*R. culmorum*.

*Leptospira pomona*.—This organism was isolated from a patient affected with the 7-day fever of Queensland (Clayton and Derrick 1937). It is antigenically distinct from the other strains. Its virulence both for human beings and guinea pigs is low. It is said by Johnson (1942) to cause endemic infection of pigs and cattle in the coastal district of southern Queensland.

*Leptospira mitis*.—This organism was also isolated from a febrile patient in Queensland (Johnson 1942). It is antigenically distinct from other types, appears to be moderately virulent for guinea pigs and like *Lepto pomona*, causes an endemic infection of pigs and cattle. It is not to be confused with the *Leptospira mitis* which was described by Mino (1939) as being responsible for infection in rice field workers in Italy, and which was shown to be identical with *Lepto batavia*.

*Leptospira grippityphosa*.—This organism is the cause of a widespread disease in eastern Europe known as swamp or mud fever. It was cultivated by Korthof (1932) in a peptone rabbit serum salt mixture. It differs antigenically from other leptospiræ and is not very pathogenic to guinea-pigs, but it may infect mice on passage (Walch-Sorgdrager 1939). Its natural animal carrier is still unknown, though voles are suspected.

*Leptospira*, Andaman A and B types.—Organisms belonging to these two types were isolated by Taylor and Goyle (1931) from patients suffering from a febrile disease in the Andaman islands. Illness was confined mainly to adults working in mud or in marshy districts in the autumn. The A type is antigenically distinct and is of only moderate virulence for the guinea pig. The B type appears to be identical with *Lepto grippoliphosa* (Walch-Sorgdrager 1939).

*Leptospira sejroe*.—Isolated by Petersen and Christensen (1939) from the blood of a fisherman in the small Danish island of Sejro. It is antigenically related to *Lepto hebdomadis* but absorption experiments show it to be distinct. Its virulence for guinea pigs is low. Its natural host appears to be a mouse *Mus sp. ciliogus*.

*Leptospira oryzi*.—This organism was isolated from rice-field workers in Italy (Babudieri 1938, 1939). It appears to be antigenically related to *Lepto bataviae* but as it has not yet been properly studied its claim to specific rank is still in doubt.

*Leptospira canicola*.—This is a natural parasite of dogs in which it gives rise to a disease more often associated with uræmia than with jaundice (see Chapter 82). It was first distinguished from *Lepto icterohæmorrhagiae* by Klarenbeek and Schuffner (1933). It differs from *Lepto icterohæmorrhagiae* in its antigenic structure and its lower virulence for guinea pigs on first isolation. After passage through guinea pigs its virulence may increase. No jaundice is produced, but in animals that survive leptospira may be excreted in the urine for months. Occasionally it infects human beings but it is not a normal parasite of rats (see Walch-Sorgdrager and Schuffner 1938). Gardner (1943b) thinks that there may be a type of leptospira infecting dogs and man in Great Britain differing from *Lepto canicola* but the evidence on this point is not yet conclusive.

*Leptospira javanica*.—Found in field rats and cats in the Dutch East Indies (see Walch-Sorgdrager 1939). Has not so far been isolated from human beings.

*Leptospira biflexa*.—Usually referred to by German workers as *Sp. rochata pseudocitrogenes*. Widespread saprophyte found mainly in water. Described originally by Wolbach and Binger (1914). Often attached to other spirochætes and protozoa (Zuelzer 1923). Especially prevalent in the slime of ponds, lakes and rivers in the slime that collects on the ends of water taps and pipes, and in the roof slime of mines. Morphologically indistinguishable from *Lepto icterohæmorrhagiae* (Fig. 219). Is very easy to cultivate. Can thrive in tap or distilled water to which 0.1 per cent potassium nitrate has been added provided that the reaction is not acid (Zuelzer 1923). Grows readily in the media used for *Lepto icterohæmorrhagiae* but Ringer's solution must be replaced by tap or distilled water since *biflexa* is very susceptible to even low concentrations of sodium chloride (Uhlenhuth and Zuelzer 1921). The simplest and most effective medium is 10 per cent rabbit serum in distilled water. Isolation in pure culture is often



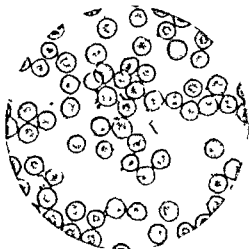
FIG. 219.—*Lepto biflexa*.

Dark ground illumination ( $\times 1500$  ca.)

difficult. Hindle (1925) found that if 20 ml. of water were added to a Petri dish containing a portion of human faeces about the size of a pea, and incubated at 25°–30° C. in the dark *Lepto biflexa* was generally observable microscopically in 10 days, and was abundant in 20 days. The leptospira were able to pass through an L5 candle. These observations formed the basis of several methods of isolation (Bauer 1927; Mochtar 1928; von Lagedes 1930). The general principle is to filter the water through a suitable candle and cultivate the filtrate. Is antigenically distinct from *Lepto icterohæmorrhagiae*. A specific precipitating substance of carbohydrate nature has been extracted from it by Hindle and White (1934). Is non-pathogenic for animals. Is believed by certain workers to be an avirulent form of *Lepto icterohæmorrhagiae* which can be rendered virulent by suitable animal passage. Balance of evidence is against this view.

## Spirillum minus

Sometimes referred to as *Spirocheta morsus muris* Described by Futaki and his colleagues (1916-1917) as the cause of rat bite fever in man According to Robertson (1924) it is a spirillum and not a spirochete, and its correct name is *Spirillum minus* Appears to be a natural parasite of rats, which act as healthy carriers of the organism Morphologically the spirillum is short, rather thick, and has tapering ends provided with one or more flagella It is 2-5  $\mu$  long, motile, and has regular rigid spirals, each of which is about 1  $\mu$  in length The movements are rapid—like those of a vibrio It is readily stained by ordinary aniline dyes, such as Loeffler's methylene blue, and by Giemsa Cultures may be obtained in Shimamine's medium, but successive transfers have not been successful The organism gives rise to one type of rat-bite fever in man Intrapertoneal inoculation of infective human material into mice is followed by no clinical evidence of disease, but spirilla appear in the blood after 5 to 14 days They are scarce at first, but later they increase though they never become numerous, it is uncommon to find two organisms in the same field (Theiler 1926) They persist indefinitely, though only in small numbers Rats behave like mice but the number of spirilla in the blood is fewer Intrapertoneal inoculation of guinea pigs produces a febrile disease After an incubation period of 6 to 15 days spirilla appear in small numbers in the blood, and pyrexia sets in accompanied by enlargement of the lymph glands There may be a marked inflammation of the subcutaneous tissue in the ano-genital region, involving the scrotal sacs, perianal tissue, and prepuce in males and the labia and perianal tissue in females Later, after 3 or 4 weeks alopecia ulceration of the skin and chronic conjunctivitis and keratitis may occur The disease is generally chronic lasting from about 2 to 4 months, but sometimes death occurs in the first 5 weeks (Ishiwara *et al* 1917) Spirilla can be demonstrated in the blood lymph glands spleen kidney adrenal and subcutaneous tissue In Robertson's (1924) experience spirilla were never demonstrable in the blood even by mouse inoculation nor did any of the guinea pigs die Rabbits may be infected but are less suitable for diagnostic purposes than mice or guinea pigs Monkeys are also susceptible (Inada *et al* 1916)

FIG. 220.—*Spirillum minus*

In film of blood of experimentally infected mouse  
Giemsa ( $\times 1000$ ) [From specimen kindly supplied by the late Prof J G Thomson]

## REFERENCES

- AKSJANZEW MALIN, S (1933) *Zbl Bakt*, 129, 405  
BABUDIERI B (1938) *Polichnico, Sez prat* 45, 1774 (1939) *Riv Parasitol* 3, 93  
BAERMANN, G and ZUEZLER, M. (1927) *Klin Wochr* 6, 979 (1928) *Zbl Bakt* 105, 345  
BAUER J H (1927) *Amer J trop Med*, 7, 177  
BAYON, H (1913) *Brit med J* 11, 1159  
BERTARELLI, E (1906) *Zbl Bakt* 41, 320. (1907) *Ibid*, 43, 238 448, (1908) *Ibid* 46, 51  
BESSEMAN, A. and POTTER, F DE (1930) *C R Soc Biol* 104, 818, (1931) *Ibid* 107, 279  
BOAK, R. A., CARPENTER, C. M and WARREN, S L. (1932) *J exp Med*, 56, 725  
BROWN H C and DAVIS, L J (1927) *Brit J exp Path*, 8, 397



- BROWN W. H. and PEARCE, L. (1920a) *J exp Med.*, 31, 473, 709 729 749 (1920b) *Ibid.*, 32, 445 473, 497. (1921a) *Ibid.*, 24, 167 (1921b) *Ibid.*, 34, 18a.
- BROWN, W. H., PEARCE, L., and WITHERELL, W. D. (1921) *J exp Med.*, 33, 495, 515, 52a.
- BUCHANAN, G. (1927) *Spec Rep Ser med Res Coun Lond.*, No 113
- CASTELLANI, A. (1905) *Brit med J.*, ii, 1250
- CHABAUD, A. (1939) *Bull Soc Path. exot.*, 32, 483.
- CLAYTON, G. E. B. and DERRICK, E. H. (1937) *Med J Aust.*, i, 647
- DRONT, C. M., KLARENBEK, A., SCHIFFNER, W. A. P., and VOLT J. (1934) *Ned Tijdschr Geneesk.*, 78, 5197
- DIMITROFF, V. T. (1927) *J infect Dis* 40, 508.
- DONELL, C. (1912) *Arch Protistenk.*, 26, 117
- EILERMANN, V. (1904) *Zbl. Bakt.*, 37, 729 (1907) *Z Hyg InfektKr.*, 58, 453
- FLETCHER, W. (1928) *Trans roy Soc trop Med.*, 21, 265.
- FUTANI, K., TAKAKI, I., TANIGUCHI, T., and OSUMI, S. (1916) *J exp Med.*, 23, 249. (1917) *Ibid* 25, 33
- GALLOWAY, I. A. (192a) *C P Soc Biol.*, 83, 1074.
- GARDNER, A. D. (1943a) *Mon. Bull. Emerg publ Hlth Lab Serv.*, 2, 40 (1943b) *Lancet*, ii, 157
- GATES, F. L. (1923) *J exp Med.*, 37, 311
- GATES F. L. and OLITSKY P. K. (1921) *J exp Med.* 33, 51
- GEORGI, F., PRACSNITZ, C., and FISCHER, O. (1929) *Klin. Wochr.*, 8, 2007
- GOERING G. (1940) *Z Immunforsch.*, 98, 90
- HAESVELL, P. (1881) *v Graefes Arch Ophthal.*, 27, 93
- HINDLE, E. (192a) *Brit med J.*, ii, 57 (1931) "A System of Bacteriology" *Med. Res. Coun.*, 8, 148
- HINDLE, E. and ELFORD W. J. (1933) *J Path. Bact.*, 37, 9
- HINDLE, E. and WHITE, P. B. (1934) *Proc. roy Soc., B* 114, 523
- HODER, F. (1930) *Z Immunforsch.*, 63, 256.
- IDO Y., ITO H., and WANI, H. (1918) *J exp Med.*, 28, 43a (1919) *Ibid.*, 29, 199
- INADA R., IDO Y., HOKI, P., KANEKO R., and ITO H. (1916) *J exp Med.*, 23, 377
- INOCK, S. (1930) *Zbl. Bakt.*, 117, 80
- ISHIWARA K., OHTAWARA, T., and TAMURA K. (1917) *J exp Med.*, 25, 45
- JAHNEL, F. (1934) *Klin. Wochr.*, 13, 500 (1937) *Ibid.*, 16, 1304
- JOHNSON D. W. (1942) *Med J Aust.* i, 431 (1943) *Brit med J.*, ii, 639
- KANEKO, R., and OKUDA, K. (1917) *J exp Med.*, 28, 32a
- KAST C. C. and HOLMER, J. A. (1940) *Am J Syph.*, 24, 671
- KLARENBEK A. and SCHIFFNER, W. A. P. (1933) *Ned Tijdschr Geneesk.*, 77, 4271
- KLEBS, A. C. (1932) *Science*, 75, 191
- KRIGLER, I. J., HEENOVITZ, D., and PEREK, M. (193a) *J comp Path. Therap.*, 51, 206.
- KRIGLER, I. J. and ROBERTSON O. H. (1922) *J exp Med.*, 35, 303.
- KNOWLES, R. and BASU B. C. (193a) *Indian J med Res.*, 22, 449
- KNOWLES, R. GUPTA, B. M. D., and BASU, B. C. (1912) *Indian J med. Res.*, Memoir No 22
- KOLLE, W. and SCHLOSSBERGER, H. (1926) *Dtsch med Wochr.*, 52, 1245 (1927a) *Ibid.*, 53, 129
- KORTHOFF G. (1932) *Zbl. Bakt.*, 125, 429
- LANDAUER, E. (1931) *Ann. Inst. Pasteur*, 47, 667
- LAPIDARI, M. and SPARROW H. (192a) *Arch. Inst. Pasteur Tunis* 17, 191
- LARSON C. L. (1944) *Publ. Hlth. Exp. Wsch* 59, 522.
- LUMLEY G. F. (1937) *Med. J Aust.*, i, 604.
- MANOUELIAN V. (1940) *Ann. Inst Pasteur* 64, 439
- MARTIN L. and PETTIT A. (1919) "Spirochétose icterohémorragique" Paris.
- METCHNIKOFF, E. and ROUX, E. M. (1903) *Ann. Inst. Pasteur*, 17, 809 (1904a) *Ibid.*, 18, 1. (1904b) *Ibid.*, 18, 657. (1905) *Ibid.*, 19, 673.
- MINO P. (1939) *Policlinico Sez. med* 46, 410
- MOCHTAR, A. (192a) *Zbl. Bakt.*, 107, 374
- MORDEK, J. (1929) *Arch. Schiff's Tropenhyg.*, 33, 603
- MORROW G., STVERTON J. T., STILES, W. W., and BERRY G. P. (193a) *Science*, 83, 374.
- MORTON H. E. (1942) *Proc. Soc. exp Biol.*, N Y 49, 566.
- MORTON H. E. and ANDERSON, T. F. (1943) *J Bact* 45, 143.
- MUDD S. POLKOVITZKY, K., and ANDERSON T. F. (1943) *J Bact.*, 46, 1a.
- MÜLLER, P. and HART C. F. (1930) *Arch. Hyg.*, 103, 95.
- NOGUCHI, H. (1911) *J exp Med.*, 14, 99 (1912a) *Ibid.*, 15, 81. (1912b) *Ibid.*, 15, 90. (1912c) *Ibid.*, 15, 201. (1912d) *Ibid.*, 15, 466. (1912e) *Ibid.*, 16, 199 (1912f) *Ibid.*, 16, 261 (1912g) *Ibid.*, 16, 620. (1913) *Ibid.*, 17, 89. (1917) *Ibid.*, 25, 735. (1922) *Ibid.*, 35, 391
- NOGUCHI, H. and AKATSU, S. (1917) *J exp Med.*, 25, 763.
- NORRIS, C., PAPFENHIMER, A. M., and FLOTANOV, T. (1906) *J infect Dis.*, 3, 266

- NOVY, F. G and KNAPP, R. E (1906) *J. infect. Dis.*, 3, 291  
 OAG, R. K (1939) *J. Path. Bact.*, 49, 339  
 OBERMEIER, O (1873) *Berl. klin. Wochr.*, 10, 152, 378, 391, 455  
 PARRAM, J. C (1922) *Amer. J. trop. Med.*, 2, 341  
 PARODI, U (1907) *Zbl. Bakt.*, 44, 428  
 PETERSEN, B and CHRISTENSEN, H. I. (1939) *C. R. Soc. Biol.*, 130, 1507  
 PLOTZ, R (1917) *J. exp. Med.*, 26, 37  
 PROWAZEK, S. von (1907) *Arch. Gesundheitsamt, Berl.*, 26, 23  
 RIECKENBERG, H (1917) *Z. Immunforsch.*, 26, 53  
 ROBERTSON, A (1924) *Ann. trop. Med. Parasit.*, 18, 157  
 SAKHAROFF, N (1891) *Ann. Inst. Pasteur*, 5, 564  
 SCHAUDINNY, F and HOFFMANN, E (1905) *Arch. Reichsgesundh. Amt*, 22, 527  
 SCHEFF, G (1935) *Zbl. Bakt.*, 134, 35  
 SCHERESCHESKY, J (1909) *Dtsch. med. Wochr.*, 35, 835, 1260, 1652  
 SCHUBERT, A and SCHLOSSBERGER, H (1930) *Klin. Wochr.*, 8, 499  
 SCHÜFFNER, W (1940) *Zbl. Bakt.*, 145, 341  
 SÉGUIN, P (1930) *C. R. Soc. Biol.*, 104, 247, 836  
 SÉGUIN, P and VINCENT, R (1938) *Ann. Inst. Pasteur*, 61, 200  
 SELBIE, F. B (1943) *Brit. J. exp. Path.*, 24, 150  
 SINTON, J. A (1924) *Indian J. med. Res.*, 11, 825  
 SOULE, M. H (1942) *Publ. Amer. Ass. Advanc. Sci.*, No 18, p. 53  
 STÉFANOPOULOU, G. J and HOSOYA, S (1928) *C. R. Soc. Biol.*, 98, 1317  
 STOKES, A., RYLE, J. A., and TYTLER, W. H (1917) *Lancet*, i, 142  
 TANI, KAKISHITA, M., and SAITO, K. (1930) *Zbl. Bakt.*, 117, 73  
 TAYLOR, J. and GOYLE, A. N (1931) *Indian med. Res. Memoirs*, No 20  
 TREILER, M (1926) *Amer. J. trop. Med.*, 6, 131  
 TILDEN, E. B (1937) *J. Bact.*, 33, 307  
 TOMASZCZEWSKI (1910) *Dtsch. med. Wochr.*, 36, 1025  
 TUNNICLIFF, R (1906) *J. infect. Dis.*, 3, 148  
 TURNER, T. B (1938) *J. exp. Med.*, 67, 61  
 TWORT, F. W (1921) *Lancet*, ii, 798  
 UHLENHUTH and FROMME (1916) *Berl. klin. Wochr.*, 53, 269  
 UHLENHUTH, P. and MÜLLER, P (1913) *Arch. Reichsgesundh. Amt*, 44, 307  
 UHLENHUTH and ZUELZER (1921) *Zbl. Bakt.*, 85, Beiheft, 141  
 VAGEDES, K. von (1930) *Zbl. Bakt.*, 133, 401  
 VINCENT, H (1896) *Ann. Inst. Pasteur*, 10, 488. (1899) *Ibid.*, 8, 609  
 VINCENT, R. and DAUFRESNE, M (1934) *C. R. Soc. Biol.*, 116, 490  
 WALCH-SORGDORFER, B (1939) *Bull. Hlth. Org., L. O. N.*, 8, 143  
 WALCH-SORGDORFER, B and SCHÜFFNER, W (1938) *Zbl. Bakt.*, 141, 97  
 WEISS, C. and WILKES WEISS, D (1924) *J. infect. Dis.*, 34, 212  
 WENYON, C. M (1926) "Protozoology," in London  
 WICHELHAUSEN O. W. and WICHELHAUSEN, R. H (1942) *J. dent. Res.*, 21, 543  
 WILE, U. J. and HEARNEY, E. B (1943) *J. Amer. med. Ass.*, 122, 167  
 WILE, U. J., PICARD, R. G., and KEARNEY, E. B (1942) *J. Amer. med. Ass.*, 119, 880  
 WOLBACH, S. B. and BINGEN, C. A. L. (1914) *J. med. Res.*, 30, 23  
 WYNN, H. L. and BECK, M. D (1935) *Amer. J. publ. Hlth.*, 25, 270  
 YANG, K. and TREILER, M (1930) *Amer. J. trop. Med.*, 10, 407  
 YEAN PO, L. (1933) *Kelasaato Arch.*, 10, 78  
 ZUELZER, M (1925) "Die Spirochäten" "Handbuch der pathogenen Protozoen"  
 Band 3, 1627 Leipzig. (1928) *Zbl. Bakt.*, 105, 384, (1930) *Arch. Hyg.*, 103, 282

## CHAPTER 39

### RICKETTSIA

#### DEFINITION—*Rickettsia*

Small, Gram negative, bacterium like organisms, usually less than half a micron in diameter. More or less pleomorphic. Stain rather poorly with aniline dyes, but well with Giemsa. Natural inhabitants of intestinal canal of arthropods, usually occupy an intracellular position. Some species are parasitic in higher animals and are pathogenic for man. The type species is *Rickettsia prowazekii*.

*Rickettsia* is the name given to certain small bacteria like bodies which are found in the alimentary canal of insects and other arthropods, and which are frequently associated with disease in man and animals. Definition of the group presents several difficulties. Some workers, like Zinsser (1937), would automatically exclude any organism such as *R. melophagi* that has been cultivated on artificial media. Further, they would insist that, when growing in the animal body or in tissue cultures only such organisms as multiply intracellularly should be regarded as true rickettsiae. Most species appear to be unable to pass through the ordinary bacterial filters, though there are exceptions, as with *R. burnetii*. Since, however, it is often difficult to obtain a homogeneous suspension of the organisms free from cellular material, too much weight should not be placed on this characteristic. It seems clear that the rickettsiae occupy a position in between the smallest bacteria like *Bartonella*, and the filtrable viruses. The fact that they can be resolved microscopically by visible light, and that they are held back by membranes which allow most of the filtrable viruses to pass through, brings them into line with the bacteria, but their failure to grow on ordinary culture media and their predilection for intracellular multiplication, show that their metabolic requirements are more akin to those of the filtrable viruses.

We shall not attempt to define the genus too closely, nor shall we exclude from the present chapter organisms, such as *R. quintana*, merely because they appear to grow extracellularly. Mention will be made of *R. conjunctivae*, *R. canis*, *R. bovis* and *R. ovina*, even though it is doubtful whether these organisms are natural inhabitants of arthropods, or even whether they are living organisms at all.

The first-named species was described by da Rocha Lima in 1916, who found these bodies in lice taken from patients with typhus fever, he proposed the name of *Rickettsia prowazekii* in honour of Ricketts and of Prowazek, both of whom died of typhus fever while investigating the disease. The second species was described by Topfer (1916) also in 1916, in lice taken from patients suffering from Wolhynian fever—better known as Trench fever, this species has been given the alternative names of *Rickettsia quintana*—on account of the 5-day febrile parox-

ysms characteristic of this disease—and *R. wolhynica*, it appears to be closely related to if not identical with, *R. pediculi*, which was found by da Rocha Lima in the apparently normal human body louse. A third species which was first described by Ricketts as far back as 1909 in Rocky Mountain spotted fever, has been called *Derma-centrozetes rickettsi* by Wolbach, as this organism appears to belong to the *Rickettsia* group, we shall refer to it as *Rickettsia rickettsi*. Sellards in 1923 claimed to have cultivated a fourth species, *R. nipponica*, from animals experimentally infected with tsutsugamushi fever in Japan. This species has been given the alternative names of *R. tsutsugamushi*, *R. akamushi* and *R. orientalis*, but, in spite of Philip's (1943) argument that the name *R. nipponica* is ruled out because of the reported cultivation of this organism on artificial media there still seems insufficient reason for discarding it. If every organism on which a fallacious observation has been made must be re-named, bacterial taxonomy will become even more perplexing than it is already.

Cowdry (1925) described a fifth species *R. ruminantium*, as the cause of heart water of sheep, goats and cattle in South Africa. A murine type of typhus virus, sometimes referred to as *R. mooseri*, was discovered as the result of epidemiological and bacteriological studies undertaken by American and Mexican workers (for references see p. 1845). More recently, the observations of Derrick (1937) and Burnet and Freeman (1937) in Australia and of Davis and Cox (1938) in the United States have revealed a rickettsia *R. burneti* (*R. diaporica*) as the cause of a febrile disease known as Q fever.

Besides these six or seven species, which have been found in association with disease of mammals, over forty other, apparently non pathogenic, species of *Rickettsia* have been described in various insects and other arthropods. One or two of these will receive brief mention at the end of this chapter, together with a few pathogenic organisms whose relationship to the *Rickettsia* group is still in doubt.

As the rickettsias have been found both in blood sucking and in non blood sucking insects it seems probable that they are primarily inhabitants of the alimentary canal of insects, and that infection of insects occurs by contamination with infected excreta (Hindle 1921). Some species are found not only in the lumen of the alimentary canal, but also in the epithelial cells lining the canal. A further invasion of the body may occur, leading to infection of the salivary glands and other tissues. Most species appear to be transmitted hereditarily to successive generations through infection of the eggs. There is a marked host specificity. Some species appear to have reached a perfect equilibrium with their insect host, and sometimes with the animal on which the insect is a parasite, such as in the system comprised by the murine typhus virus, the rat flea and the rat. Others however, which by some workers (see Zinsser 1937, Burnet 1942) are supposed to have developed later in the evolutionary period, are pathogenic for their insect hosts, killing a high proportion of them—as for instance, the classical typhus rickettsia and the louse. A few species appear to have become adapted to an alternate



FIG. 221.—*R. melophagi*  
Smear preparation from gut of sheep-  
ked. Giemsa ( $\times 1000$ )

existence in insects and in animals, infection of insects occurs in these cases by blood sucking. There is, however, no evidence to suggest that a separate cycle of development occurs in either host, as is so frequently observed with the protozoa.

Morphologically, in the gut of the louse rickettsiae appear like very small cocci, diplococci, or short bacilli. Their size is generally given as  $0.3-0.5 \mu$  long by  $0.3 \mu$  broad but the bacillary forms may reach a length of 1.5 or even  $2.0 \mu$ . According to Elford and van den Ende (1944) the size of *R. prowazeki*, judged by ultra violet photomicrographs, is  $0.6-1.8 \mu$  long by  $0.45 \mu$  broad. Gradocol membrane experiments likewise suggest that the width is not less than  $0.4-0.5 \mu$ —a value greater than that which has previously been quoted. Their arrangement is variable, single forms may occur, diploid forms are very common, and the small coccoid forms are often grouped in dense masses. Under

certain conditions, they may form intracellular homogeneous or granular inclusion bodies not unlike those seen in psittacosis (Pinkerton and Hass 1932, Begg *et al* 1944). By the electron microscope they appear, like bacteria, to possess a limiting membrane, enclosing a substance of moderate opacity, dense granules may be seen in some forms (Plotz *et al* 1943, Weiss 1943). Most species are non motile. On the whole they stain poorly with the ordinary aniline dyes, but Castaneda (1930), Lepine (1932), Laigret and Auburtin (1938), and Macchiavello (1941) have shown

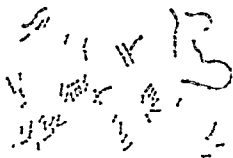


FIG 222.—*Pickettsia prowazeki*

Smear preparation from louse's gut, showing bacillary forms and occasional thread like chains. Magnification 2 000 diameters (approx). (After Wolbach, Todd and Palfrey.)

that good results may be obtained with methylene blue, thionin, and fuchsin, provided the stain is made up with a suitable buffer or mordant. Macchiavello's stain, in particular, gives excellent results with smears, though it is not so successful for tissue sections. For general purposes, Giemsa is probably the best stain, with this the rickettsiae appear as purple cocco-bacilli, or frequently as bipolar staining rods, sometimes they seem to be surrounded by a paler staining substance. They are uniformly Gram negative. When very few in number, they cannot be recognized with certainty, since their resemblance to minute particles of detritus is too close, but when they are present in large numbers, their recognition is comparatively easy. It is by reason of the small numbers in which they occur in the blood of infected men and animals, that their demonstration in this medium has only rarely been accomplished. Arkwright, Bacot and Duncan (1919-20) lay down the following criteria for the recognition of *Rickettsia* in the louse's gut: (1) its minute size,  $0.3-0.5 \mu \times 0.3 \mu$ , (2) its irregularity in shape, round, oval, diplococcal, and bipolar-stained bacillary forms being seen, (3) its occurrence in very large numbers or even masses, especially on flakes of solid material in the excreta, (4) its well stained appearance and purple colour with Giemsa.

**Cultivation and Metabolism**—None of the pathogenic species has yet been cultivated apart from living cells. Of the commensal species, *R. melophagi*, found in the sheep ked, is said to have been grown on blood agar. Noguchi (1926) claimed to have cultivated some of the commensal rickettsiae, found in ticks, on leptospiral medium containing 0.2 per cent of a carbohydrate, when 1 per cent of agar was added, and the medium was slanted, surface colonies were obtained. Working with the rickettsiae from Rocky Mountain fever and from typhus, Wolbach and Schlesinger (1923-24) succeeded in obtaining growth in tissue cultures. The organisms survived and multiplied only in the endothelial cells. Primary cultures remained alive and virulent for 1 to 2 weeks as a rule, and later generation cultures for 2 to 4 weeks. Nigg and Landsteiner (1930) showed that cultivation of the typhus virus could be accomplished in a medium, similar to that described by Maitland and Maitland (1928) for vaccinia virus, which contains living but not actively proliferating cells. A study by Zinsser and Schoenbach (1937) of the growth of rickettsiae in such a medium showed that multiplication of these organisms did not begin till after the tissue cells had ceased growing actively. This was different from the behaviour of viruses, which multiply most during the stage of tissue cell growth. Burnet (1938) has made similar observations and would regard this difference in metabolism between the rickettsiae and filtrable viruses as of classificatory value. Another method used successfully in the cultivation of the filtrable viruses, namely growth on the chorio allantoic membrane of the developing chick embryo, was found by da Cunha (1934) to be applicable to rickettsiae. An even more successful method is that described by Cox (1941) of growing rickettsiae in the yolk sac of the developing chick embryo. Pure or practically pure strains of *R. prowazeki* may be obtained by Weigl's method of intrarectal injection of body lice with infective material. The intestine of the louse is practically free from ordinary bacteria so that it serves as an almost sterile medium for the cultivation of rickettsiae. In practice *in vivo* cultivation in the tissues of a susceptible animal, such as the testicle of the guinea pig or rabbit is frequently employed for preserving strains of typhus virus, according to Kodama and Takahashi (1931), viruses kept in this way undergo no change in antigenic structure or pathogenicity. Certain rickettsial strains, such as *R. nipponica*, can be cultivated in the anterior chamber of the rabbit's eye (Nagayo *et al.* 1930). After an incubation period of 4 to 8 days iritis develops similar to that occurring naturally in tsutsugamushi fever. Histological examination reveals the presence of peculiar corpuscles in the endothelial cells of Descemet's membrane, consisting apparently of colonies of rickettsiae. The animal method of cultivation has been applied particularly to *R. prowazeki* and *R. mooseri* in an effort to provide heavy suspensions of rickettsiae for serological and vaccination purposes. The mouse's lung has proved particularly useful for this purpose, the animals being inoculated intranasally (Okamoto 1937, Castaneda 1939, Durand and Sparrow 1940, Giroud and Panther 1942).

Little is yet known about the growth requirements of *Rickettsia*, but interesting observations have been made. Pinkerton and Hass (1932) found that in plasma tissue cultures *R. prowazeki* grew best at 32° C. This behaviour may be related to the preference that some species of rickettsiae show, after inoculation into the peritoneal cavity of the guinea pig for growth in the scrotal sac, where the temperature is lower, rather than for the general peritoneal cavity. In the Maitland medium growth occurs equally well at 32° C and at 37° C. This apparent dis

crepancy may be due to the circumstance that in plasma tissue cultures the cells are multiplying actively, whereas in the Mantland medium mitotic division is rarely seen. From these observations Pinkerton (1934) drew the conclusion that typhus rickettsiae grow best in cells which are metabolizing slowly. That this explanation is correct is rendered probable by the work of Zinsser and Schoenbach (1937) and Burnet (1938). Thus Zinsser and Schoenbach found that in tissue culture rickettsiae underwent no appreciable multiplication till the tissue cells had ceased to grow actively. Viruses, on the other hand, multiplied most abundantly during the stage of tissue cell growth. Burnet (1938) made similar observations on the rickettsia of Q fever. It would appear that, in this respect, there is an important difference between the metabolism of *Rickettsia* and of filtrable viruses. The reason why rickettsiae grow better at 32° C than at 37° C in plasma tissue cultures and in the scrotal sac than in the peritoneal cavity, is probably not because they prefer the lower temperature in itself, but because tissue cell metabolism is less rapid at this temperature than at 37° C. Further observations by Burnet and Freeman (1941) on egg membrane cultures suggest that rickettsiae grow only in regions where there is an abundant supply of oxygen. The authors recall the fact that the rickettsiae are essentially parasites of the vascular endothelium which is rich in oxygen, and explain their better growth in the later than in the early stages of tissue cultures on the ground that, not till active tissue growth is over, does the oxidation reduction potential of the cells rise sufficiently high to enable rickettsiae to multiply. Other factors that favour growth of rickettsiae in the animal body, such as riboflavin deficiency, benzol poisoning and X ray irradiation possibly act in the same way, namely by lowering intracellular metabolism.

Though most of the pathogenic species of *Rickettsia* seem to require intracellular conditions for growth *R. nipponica* and *R. burneti* are both said to be capable of growth outside the cells though not, of course, in a cell free medium. A further difference in the metabolism of different species is shown by the fact that the typhus group of rickettsiae multiply exclusively in the cytoplasm, whereas the spotted fever group (*R. rickettsi*) grow best within the nucleus.

The Resistance of *Rickettsia* has not been fully studied. The rickettsiae of typhus, Rocky Mountain fever, and heartwater are said to be easily inactivated by heat drying and chemical disinfectants, but *R. quintana* is said to be more resistant (Cowdry 1926). The Trench fever Committee (Bruce 1921), however, found that the infectivity of louse excreta was destroyed by exposure to moist heat for 20 minutes at 60° C, and to dry heat for the same time at 100° C. Arkwright and Bacot (1923) found that *R. prowazeki* remained virulent for 11 days in louse excreta which had been kept dry at room temperature. The viability of rickettsiae in infected tissues and in tissue cultures, as judged by their infectivity, seems to be considerably affected by the temperature at which they are kept. Spencer and Parker (1924) working with *R. rickettsi*, found that certain tissues remained infective in pure glycerol for as long as 10 months if preserved at -10° C. Nigg (1930) working with *R. mooseri*, found that tissue cultures in a serum Tyrode mixture remained alive and virulent for several months at 37° C and at -20° C, but generally died out in a week or two at the intermediate temperatures of 20° C and -4° C. Sterile skim milk is said to be a good suspending agent for rickettsiae that are to be preserved in the dried state (Topping 1940), and used as a diluent it maintains the virulence of *R. prowazeki* at 26°-28° C. for 24 hours (Anderson

1944) In serum broth at pH 6.0-8.5 rickettsial suspensions may be stored for months at  $-77^{\circ}\text{C}$  with little alteration in potency (Elford and van den Ende 1944)

**Antigenic Structure**—The difficulty of obtaining suspensions of rickettsiae free from admixture with cells and other bacteria has rendered the study of the antigenic structure of these organisms peculiarly difficult. Ledingham (1940) and others have shown that inoculation of infective material or of rickettsial suspensions into rabbits is followed by the appearance of specific agglutinins. Advantage has been taken of this circumstance to study the relationship of the pathogenic rickettsiae to *Proteus*  $\lambda$  strains. Without entering here into the practical performance of the Weil-Felix test (see Chapter 83) it may be mentioned that the serum of patients suffering from typhus and typhus-like diseases frequently agglutinates *Proteus* OX 19 or one of its variant strains OX 2 or OX K. Castaneda and Zia (1933) studying *R. prowazekii* and *Proteus*  $\lambda$  19 by the agglutination and absorption of agglutinins technique found that these organisms behaved as if each possessed a specific and a group somatic antigen. White (1933) using in addition the precipitation test obtained evidence of the existence in  $\lambda$  19 of two distinct heat stable somatic receptors: (1) an alkali labile receptor (Castaneda's P factor) which is mainly responsible for the agglutination of this organism by its own antiserum; (2) an alkali stable receptor (Castaneda's  $\lambda$  factor) which is responsible for the reaction of this organism with the sera of typhus patients. White's conclusion received confirmation from the further work of Castaneda (1934, 1935) who was successful in extracting specific soluble substances of polysaccharide nature from  $\lambda$  19 and *R. prowazekii*. These substances have already been referred to as P and  $\lambda$ . It appears therefore as if *Proteus*  $\lambda$  19 and *R. prowazekii* possess a common alkali stable antigenic factor ( $\lambda$ ) of polysaccharide nature which is responsible for the Weil-Felix reaction. In addition *Proteus*  $\lambda$  19 contains a specific alkali labile receptor also apparently of polysaccharide nature which plays no part in this reaction (see Chapter 27). Whether *R. prowazekii* contains a specific receptor of its own similar to the P factor of *Proteus*  $\lambda$  19 is not yet clear but there seems little doubt from Castaneda and Zia's work that it possesses a heat labile antigen behaving in much the same way towards the heat stable antigen as the  $\lambda$  1 antigen of the typhoid bacillus does to the O antigen. Topping (1941) has reported the existence of a soluble substance present in the supernatant fluid after ether extraction and centrifugation of yolk sac cultures of *R. prowazekii* possessing apparently the same antigenic properties as those of the intact organisms.

The relationship of the different types of typhus virus to each other and to the viruses of Rocky Mountain spotted fever, tsutsugamushi fever, fièvre boutonneuse, tick bite fever and Q fever has been studied partly by serological methods and partly by cross protection tests in living animals. The interpretation of the results is so closely bound up with the Weil-Felix reaction and with the clinical and epidemiological characteristics of these diseases that it is proposed to defer further discussion of this subject to Chapter 83. Suffice it to say that there appear to be at least three major receptors in *Proteus*  $\lambda$  strains represented by the OX 19, OX 2 and OX K types which correspond to similar receptors in rickettsial strains isolated from different typhus and typhus-like diseases. Ciucu and his colleagues (1933) have shown that all three receptors are of glycolipid nature and can be extracted from the bodies of the bacilli by the trichloroacetic acid method.

**Pathogenicity**—As already mentioned there are five or six known pathogenic species for man and one for cattle. Leaving aside this last species *R. ruminantium*



about which comparatively little is known, we may refer briefly to the reproduction of the various rickettsial diseases in animals

**Pathogenicity of *R. prowazeki* for Animals**—A febrile disease simulating typhus can be reproduced in apes, monkeys, and guinea pigs by the classical *louse-borne* type of virus, rabbits and rats are relatively resistant to inoculation. According to Nicolle, Conr and Conseil (1911) chimpanzees are more sensitive than macaques, subcutaneous inoculation of 1 ml. of human blood is generally sufficient to infect chimpanzees but for macaques 4-5 ml intraperitoneally are required. The blood of human patients is most virulent towards the end of the fever, but it is said to be virulent from 2 days before the onset to 2 days after the decline of the fever (Arkwright *et al* 1919 20)

After inoculation of typhus blood into monkeys there is an incubation period of about a week followed by a rise of temperature, which continues to ascend gradually for some days just as in man, the temperature is maintained for 7 to 10 days and then falls rapidly. A period of hypothermia may succeed, followed by a return to normal temperature. Accompanying the fever there are general constitutional symptoms, such as anorexia, ruffled coat and conjunctival congestion, on the 3rd or 4th day a rash sometimes breaks out on the face. Death may occur. During the early part of the fever there is a leucopenia, followed by a return to normal, the leucocytes continue to rise, passing above normal during convalescence and not returning to normal till about a month after inoculation. The disease can be passed indefinitely through monkeys. A single attack provided it is severe produces a solid immunity, but after a mild attack the immunity is less marked. Instead of typhus blood monkeys can be infected with a suspension of guinea pig brain tissue or with ground up lice or louse excreta. Arkwright Bacot and Duncan (1919) brought evidence to show that the monkey louse, *Pediculus longiceps*, became infected by feeding on typhus monkeys or after rectal injection of typhus blood and was able to transmit the disease to normal monkeys. The infected lice were found to contain rickettsiae, *Pediculus* from non inoculated monkeys never contained rickettsiae.

Guinea pigs can be infected by virus from man the louse or infected guinea pigs or monkeys. The incubation period is generally 6 to 14 days, but it may extend to 26 days (da Rocha Lima 1920a), it is longer after subcutaneous than after intraperitoneal injection. The disease is characterized mainly by fever. The rectal temperature rises from 102° to 103° F at the end of the incubation period remains at between 103° and 106° F for 3 to 14 days and then falls to normal. According to Grünfeld Serebrjannaja, and Neumann (1933) there is a mononuclear leucocytosis reaching its maximum as the fever declines, the mononuclear cells rise from 2 per cent. to between 6 and 14 per cent. The animals recover and are subsequently immune to a fresh inoculation. If killed, there is little to be seen macroscopically beyond slight enlargement and darkening of the spleen and sometimes slight congestion of the testicles, which may be covered with a gelatinous exudate. Microscopically, both in man and in guinea pigs the main lesions are found in the blood capillaries, especially those in the skin skeletal muscles and central nervous system. They consist of thromboses with perivascular accumulations of cells, often accompanied by small hemorrhages. In the central nervous system characteristic nodules are found simulating tubercles. The primary lesion is in the endothelial cells lining the walls of the capillaries. Rickettsiae have been demonstrated in the lesions of the skin kidneys, testicles brain and other organs in man (Wolbach *et al* 1922). The height and duration of the fever in guinea-pigs is variable and great care should be taken before concluding that it is definitely caused by the typhus virus. Ecker and Weed (1932) and Badger (1933a b) point out that symptoms very suggestive of infection with *R. prowazeki* or *P. rickettsii* may be produced in guinea pigs by certain organisms of the *Proctus* and *Salmonella* groups. Cultural, serological and cross-immunity tests may all be required to establish the real causative agent in any given febrile condition. According to Arkwright and Bacot (1923),

the most certain way of establishing that an attack of fever in the guinea pig is really due to the typhus virus is to inject live intra rectally with a suspension of the guinea pig's platelets and observe the development of rickettsiae in the excreta. Infected guinea pigs, it may be noted, do not give a positive Weil-Felix reaction though natural agglutinins to *Proteus* OX 19 are sometimes present in a titre of 1/20 or less.

The typhus virus can be passed from man to monkeys from monkeys to guinea pigs and from guinea pigs to monkeys.

**Pathogenicity of *R. mooseri* for Animals**—The murine typhus virus gives rise in guinea pigs to a disease differing in certain respects from that caused by the classical louse-borne virus (Pinkerton 1929, 1931, Zinsser and Castaneda 1930). After intraperitoneal inoculation with the murine type the temperature rises rather earlier, about the 4th to 6th day though the actual height reached may be less than with the louse-borne type. The scrotal and testicular reaction caused by the murine type first described by Neill (1917) when investigating Mexican typhus is much more intense and microscopic examination reveals the presence of large numbers of rickettsiae—sometimes known as Mooser (1928) bodies—in the tunica vaginalis. On the other hand nodular lesions in the brain are more frequent in louse-borne than in murine type infections. *P. mooseri* is further distinguished from *R. prowazeki* by its ability to give rise after intraperitoneal inoculation to a febrile disease in rats accompanied by proliferation of rickettsiae in the scrotal sac; *R. prowazeki* causes a completely inapparent infection in these animals. Moreover, *R. mooseri* causes a heavy infection of the lung after intranasal inoculation into rats or mice (see Castaneda 1930) whereas *R. prowazeki* usually grows much less abundantly. Incidentally it may be noted that when growing in the lungs of rats or rabbits the murine virus may give rise to intracellular inclusion bodies of the morula type consisting of colonies of rickettsiae (Begg *et al.* 1944). Cross immunity tests indicate that the two types of virus are very closely related (Mooser and Dummer 1930, Nicolle and Laigret 1932, Zinsser and Castaneda 1934). More recent work—mainly unpublished—suggests, however, that they are not antigenically identical.

**Pathogenicity of *R. rickettsii* for Animals**—The disease produced in guinea pigs by inoculation of Rocky Mountain spotted fever virus is similar to that caused by the typhus virus but is much more severe. After intraperitoneal inoculation with the Western type the incubation period is usually only 2 to 4 days. The temperature rises rapidly to about 106°F, and death usually occurs within a week. From the 3rd or 4th day of the fever swellings and hemorrhages of the scrotum and ears occur which may go on to necrosis. Post mortem examination shows a considerable enlargement of the spleen and frequently a marked scrotal reaction with rickettsial bodies in the tunica vaginalis. The Eastern type is said to be less virulent but both viruses produce a characteristic rash in the monkey (Badger 1933c).

Rabbits can be infected with the Rocky Mountain virus—they develop a febrile disease; rabbits inoculated with the typhus virus do not react at all. It is interesting to note as in indicating the closeness of the relationship between the two viruses, that rabbits experimentally infected with the Rocky Mountain virus may develop agglutinins to *Proteus* OX 19 and give a positive Weil-Felix reaction; rabbits inoculated with the typhus virus likewise develop agglutinins—usually to a rather higher titre (Munter 1929). But inoculation of a rabbit with typhus is said not to protect it against subsequent inoculation with Rocky Mountain virus indicating that though both viruses closely resemble each other antigenically they are distinguishable by their virulence and by their immunizing properties. Experiments on guinea pigs, however, indicate that inoculation with either virus provides a certain amount of protection against subsequent inoculation with the other (Breml 1928). White mice and rats are said to develop a symptomless infection after intraperitoneal inoculation with Rocky Mountain spotted fever virus (Fukuda 1929) but after intranasal inoculation they develop pulmonary lesions in which considerable numbers of rickettsiae are present (Durand and Crouse 1940, Durand and Sparrow 1940).

**Pathogenicity of *R. nipponica* for Animals.**—The viruses of tsutsugamushi fever, mite fever and scrub typhus appear to be closely related. They are not, as a rule very infective for guinea-pigs, though occasional strains may prove highly fatal, giving rise to ascites and splenic enlargement after intraperitoneal inoculation. In white rats they produce an unapparent infection. One of their most striking properties is their ability to give rise to an acute reaction, characterized by circum-corneal injection, iritis, turbidity of the aqueous humour, pannus, and the presence of rickettsial bodies in Descemet's membrane on inoculation into the anterior chamber of the eye of rabbits (Nagao *et al.* 1931; Lewthwaite and Savory 1934, 1935). The serum of the animals of an acclimated *Peromyscus* O.V.K. 10 days or so after inoculation. The viruses of this group are further distinguished by the ulceration and bubo formation which they cause on intracutaneous injection of monkeys. The virus of *fevre boutonneuse* appears to be the only other known *P. nipponica* that can produce a marked local lesion in monkeys. After intraperitoneal inoculation with infective material from the rabbit *mouse* may die in a fortnight or so post mortem, there is a sticky fluid in the peritoneum containing large numbers of rickettsiae.

**Pathogenicity of *R. burneti* for Animals.**—This organism may cause a febrile reaction in monkeys inoculated intracutaneously or subcutaneously. No local lesion occurs in the skin similar to that seen after intracutaneous inoculation with *R. nipponica* (Burnet and Freeman 1935). Inoculated intraperitoneally into guinea-pigs it gives rise after an incubation period of 2 to 14 days—depending on the dose—to a febrile non-fatal disease of 4 to 6 days' duration. If the animal is killed during the height of the infection, the spleen is found to be enlarged. The serosal sac is seldom affected. Strains of American origin are more virulent than those of Australian origin, frequently producing a fibinous exudate around the spleen in which large numbers of rickettsiae can be demonstrated microscopically. The liver of guinea-pigs inoculated with either strain can be shown by inoculation to be highly infective (Derrick 1937; Dyer 1939; Burnet and Freeman 1939).

*Mouse* rats and rabbits are all susceptible but suffer from an unapparent infection. In *mouse* killed 10 to 14 days after a heavy intraperitoneal inoculation, the liver is enlarged and pale and the spleen is often greatly enlarged, tense and of a uniform deep red colour. Rickettsiae are present in smears from both organs (Burnet and Freeman 1937, 1939). *Mouse* inoculated intranasally may develop pneumonia, characterized by irregularly distributed nodules of consolidation, in which large numbers of rickettsiae can be demonstrated (Findlay 1940).

**Pathogenicity of *R. quintana* for Animals.**—We have little exact knowledge of the behaviour of the virus of trench fever in laboratory animals. Da Rocha Lima (1939) states that a small proportion of guinea-pigs may develop a low undulating fever after inoculation with material containing this organism, but that infection cannot be transmitted by passage to fresh animals.

#### Notes on a few miscellaneous organisms.

***R. rocha-limae*.**—This organism was described by Weigl in 1921. It is apparently a parasite of the body louse. It is distinguished from *P. pediculi* by morphological peculiarities and its variability (see also p. 1852). Introduced into a flock of lice it may spread with extraordinary rapidity (Weigl 1939). There is reason to believe that it may give rise to a bacteraemia in man. It has been regarded by some authors as the cause of trachoma, but the evidence in favour of this is unconvincing (see Chapter 6). What relation *P. rocha-limae* has to similar rickettsiae found in lice such as *P. urens* and others, is still doubtful (see Herzig 1939).

***R. conjunctivae*.**—Coles (1931, 1935, 1936) in South Africa described a rickettsia-like organism in the conjunctival epithelium of sheep, goats and cattle. Sometimes its presence was accompanied by inflammation. A similar organism was

met with by Donatien and Lestoquard (1937a) in their study of an epidemic of conjunctivitis among sheep in Algeria. The organism has not been fully examined, from its description it appears rather large, no arthropod vector is known, and it is doubtful whether it belongs to the *Rickettsia* group. According to Giroud and Panthier (1939) there is no such organism as *R. conjunctivæ*; the bodies that have been mistaken for it are the result of bacteria undergoing phagocytosis, and can be found in the conjunctiva of normal cattle.

*R. canis*, *R. ovina*, and *R. bovis*.—These organisms, which were described by Donatien and Lestoquard (1935, 1936a, b, 1937b) in Algeria, are believed to be parasites of the mononuclear cells. *R. canis* is said to be responsible for a severe disease of dogs, the other two species are said to give rise to a relatively mild disease in their respective hosts. There is evidence that infection is transmitted by ticks. Whether they are true micro-organisms and, if so, whether they belong to the *Rickettsia* group, or should be classified in some other genus, must await further study. Again, their relation to the intracellular bodies found by Kurloff in the mononuclear cells of the guinea pig's blood is doubtful (see Mochowski 1937).

**Classification.**—Most of our knowledge on the relationship of the pathogenic rickettsiæ to each other has been gleaned from an examination of the sera of patients and experimentally inoculated rabbits and from cross immunity experiments in animals. The results of these will be more conveniently dealt with in Chapter 83. Suffice it to say here that, excluding *R. quintana* about which our information is very slight, the rickettsiæ pathogenic to man may be classified broadly into four groups: the typhus group, the spotted fever group, the tsutsu gamushi group and the Q fever group. Within each of these groups there are types of *Rickettsia*, sometimes with a circumscribed geographical distribution, that differs from each other in their insect host, in their virulence for man and for experimental animals, and in their apparent antigenic structure. Whether these types should be regarded as species or as varieties of the main type it is at present impossible to decide. (For general reviews on the rickettsiæ see Arkwright 1921, Wolbach 1925, 1941, Cowdry 1926, Pinkerton 1942, and for useful technical information on their study see Clavero and Gallardo 1943).

## REFERENCES

- ANDERSON, C. R. (1944) *J. Bact.* **47**, 519.  
 ARKWRIGHT, J. A. (1924) *J. R. Army med. Cps.* **42**, 447.  
 ARKWRIGHT, J. A. and BACOT, A. W. (1923) *Brit. J. exp. Path.* **4**, 70.  
 ARKWRIGHT, J. A., BACOT, A., and DUNCAN, F. M. (1919) *Trans. Soc. trop. Med. Hyg.* **12**, 61. (1919-20) *J. Hyg., Camb.*, **18**, 76.  
 BADGER, L. F. (1933a) *Amer. J. trop. Med.* **12**, 179. (1933b) *Publ. Hlth. Rep., Wash.*, **48**, 677. (1933c) *Amer. J. publ. Hlth.*, **23**, 19.  
 BOGG, A. M., FLETCHER, F. and EADE, M. VAN DER. (1944) *J. Path. Bact.* **58**, 100.  
 BREINL, F. (1928) *J. infect. Dis.*, **42**, 48.  
 BRUCE, D. (1921) *J. Hyg., Camb.* **20**, 208.  
 BURNET, F. M. (1938) *Aust. J. exp. Biol.* **16**, 219. (1942) *Med. J. Aust.* **1**, 129.  
 BURNET, F. M. and FREEMAN, M. (1937) *Med. J. Aust.* **1**, 299. (1938) *Ibid.*, **1**, 1114. (1939) *Ibid.*, **1**, 887. (1941) *J. Immunol.* **40**, 405.  
 CASTANEDA, M. R. (1930) *J. infect. Dis.*, **47**, 416. (1934) *J. exp. Med.*, **60**, 119. (1935) *Ibid.*, **62**, 289. (1939) *Amer. J. Path.* **15**, 467.  
 CASTANEDA, M. R. and ZIA, S. (1933) *J. exp. Med.*, **58**, 55.  
 CICCA, M., MESROBIAN, L., BADENSKI, A., and MCNEILAN, G. (1935) *C. R. Soc. Biol.*, **127**, 1414.  
 CLAVERO, G. and GALLARDO, F. P. (1943) "Técnicas de Laboratorio en el Tifus exantemático." Prensa española, Madrid.  
 COLES, J. D. W. A. (1931) *17th Rep., Director vet. Serv., South Africa*, p. 175. (1935) *Onderstepoort J.*, **4**, 389. (1936) *J. S. Afr. vet. med. Ass.*, **7**, 221.

- COWDREY, E. V. (1925) *J exp Med*, 42, 231-233. (1926) *Arch Path Lab Med*, 2, 59
- COX, H. R. (1941) *Science* 94, 399
- CUNHA, A. M. DA (1934) *C R Soc Biol*, 117, 392
- DAVIS, G. E. and COX, H. R. (1935) *Publ Hlth Rep*, Wash., 53, 2209
- DEBBICK, E. H. (1937) *Med J Aust*, ii, 281
- DOXATYEN, A. and LESTOQUARD, F. (1935) *Bull. Soc. Path exot*, 28, 418. (1936a) *Ibid.*, 29, 105. (1936b) *Ibid.*, 29, 105-7. (1937a) *Ibid.*, 30, 18. (1937b) *Arch Inst Pasteur Alger*, 15, 142
- DURAND, P. and GIBOUT, P. (1940) *Arch Inst Pasteur Tunis* 29, 234
- DURAND, P. and SPARROW, H. (1940) *Arch Inst Pasteur Tunis* 29, 1
- DYER, R. E. (1939) *Publ. Hlth Rep*, Wash., 54, 1229
- ECKER, E. E. and WEED, L. A. (1932) *J infect Dis*, 50, 484
- FINDLAY, G. M. (1942) *Trans R Soc trop Med Hyg*, 35, 213
- FUKUDA, Y. (1929) *Zbl Bakt*, 111, 403
- GIBOUT, P. and PANTHIER, R. (1939) *Bull. Soc. Path exot*, 32, 464. (1942) *Ann Inst Pasteur*, 68, 137
- GRUNFELD, A. A., SERZEBJANNAYA, A. I., and NEUMANN, M. W. (1933) *Zbl Bakt* 129, 56
- HERZIG, A. (1939) *Zbl Bakt*, 143, 299-303
- HINDLE, E. (1921) *Parasitology* 13, 152
- KODAMA, M. and TAKAHASHI, K. (1931) *Zbl Bakt*, 119, 311
- LAURET, J. and AUBURN, P. (1935) *Bull. Soc. Path exot*, 31, 790
- LEDINGHAM, J. C. G. (1920) *Lancet*, i, 1264
- LEFAYE, P. (1932) *C P Soc Biol*, 109, 1162
- LEWTHWAITE, R. and SATOOR, S. R. (1934) *Trans 9th Congr., Far East Assoc. trop Med.*, Nanking 1, 249. (1936) *Brit J exp Path.*, 17, 1
- MACCHIARELLO, A. (1941) See Zinsser (1941) p. 896
- MAITLAND, H. B. and MAITLAND, M. C. (1928) *Lancet* ii, 596
- MOCHKOVSKI, C. (1937) *C R Soc Biol*, 126, 379
- MOOSER, H. (1928) *J infect Dis*, 43, 241, 261
- MOOSER, H. and DEMMER, C. (1930) *J exp Med*, 51, 189
- MUSTER, H. (1928) *Z Hyg Infektkr*, 103, 124
- NAGAYO, M., MITAGAWA, Y., MITAMURA, T., TAMIYA, T., SATO, K., HAZATO, H., and IMAMURA, A. (1931) *Jap J exp Med*, 9, 87
- NAGAYO, M., TAMIYA, T., MITAMURA, T., and SATO, K. (1930) *C R Soc Biol*, 104, 637
- NEILL, M. H. (1917) *Publ. Hlth Rep*, Wash., 32, 1103
- NEITZ, W. O. (1940) *J S Afr vet med Ass*, 11, 10
- NICOLLE, C., CONOR, A., and COSSELL, E. (1911) *Ann Inst Pasteur*, 25, 97
- NICOLLE, C. and LAURET, J. (1937) *Arch Inst. Pasteur, Tunis*, 21, 251
- NIGO, C. (1933) *J exp Med*, 61, 17
- NIGO, C. and LANDSTEINER, K. (1930) *Proc. Soc. exp Biol.*, N. Y., 23, 3.
- NOGUCHI, H. (1926) *J exp Med*, 43, 515
- OKAMOTO, Y. (1937) *Kiasso Arch*, 14, 23-99-113.
- PHILIP, C. B. (1943) *Amer J Hyg*, 37, 301
- PINKERTON, H. (1929) *J infect Dis*, 44, 337. (1931) *J exp Med*, 54, 181 (1934) *Arch exp Zellforsch* 15, 425. (1942) *Bact. Rev.*, 6, 37
- PINKERTON, H. and HASS, G. M. (1932) *J exp. Med*, 56, 131, 145, 151
- PLOTT, H. SMADEN, J. E., ANDERSON, T. F., and CHAMBERS, L. A. (1943) *J exp Med*, 77, 300
- RICKETTS, H. T. (1909) *J Amer med Ass*, 52, 379
- ROCHA LIMA, H. DA (1916) *Berl Klin Wochr*, 53, 56" (1909a) Prowazek's "Handbuch der pathogenen Protozoen" ii, 990. (1920b) *Ibid*, 1031
- SEILLARDS, A. W. (1923) *Amer J trop Med*, 3, 529
- SPENCER, P. R. and PARKER, P. R. (1924) *Publ Hlth Rep*, Wash., 39, 55
- TÖFFER, H. (1916) *Munch med Wochr*, 63, 1495
- TOPPING, A. H. (1940) *Publ. Hlth. Rep*, Wash., 55, 545. (1944) *Ibid.* 59, 16-1
- WEISS, L. J. (1943) *J Immunol*, 47, 353.
- WEIGL, R. (1939) *Zbl Bakt*, 143, 291
- WHITE, P. B. (1933) *Brit J. exp Path.*, 14, 145
- WOLBACH, S. B. (1920) *J Amer med. Ass*, 84, 723. (1941) "Virus and Rickettsial Diseases. Harvard Univ Press, Camb., Mass., p. 789
- WOLBACH, S. B. and SCHLESINGER, M. J. (1923-24) *J med Res*, 44, 231
- WOLBACH, S. B., TODD, J. L., and PALFREY, F. W. (1922) "Report of Typhus Research Commission of League of Red Cross Societies to Poland." Harvard Univ Press, Cambridge Mass
- ZINSSER, H. (1937) *Amer J Hyg*, 25, 430. (1941) "Virus and Pickettsial Diseases." p. 872. Harvard Univ Press, Cambridge, Mass.
- ZINSSER, H. and CASTAYEDA, M. R. (1930) *J exp. Med*, 52, 649. (1934) *Ibid.*, 59, 471
- ZINSSER, H. and SCHROENBACH, E. B. (1937) *J exp Med*, 68, 207

## CHAPTER 40

### THE PLEUROPNEUMONIA GROUP OF ORGANISMS

#### TENTATIVE DEFINITION

Microscopically visible, extremely pleomorphic organisms showing granules rings coccoid forms, filaments and other bizarre forms. Some forms can pass coarse bacterial filters. Size of smallest elements varies from about 125 to 250  $\mu$ . Non motile. Stain poorly with ordinary bacterial stains, but well with Giemsa. Gram negative. Grow in nutrient media in absence of living tissue cells. Facultative anaerobes. Form characteristic minute colonies on suitable solid media. Parasitic species require a high concentration of animal protein in the medium. Readily destroyed by heat. Apparently no special resistance to glycerol. Some species are bile soluble. Antigenic specificity is usual, but not complete. Ability to give rise to inclusion bodies in tissues very doubtful. Considerable degree of host specificity. Immunity following disease does not appear to be specially lasting.

Though the organism responsible for pleuropneumonia of cattle (see Chapter 84) was recognized and cultivated by Nocard and Roux as long ago as 1898, it is only within the past few years that a number of closely related organisms, some pathogenic, some saprophytic, have been described, and that the importance of a large group of organisms possessing unusual and distinctive properties has been realized. The complex morphology of the pleuropneumonia organism was described by Bordet (1910) and by Borrel and his colleagues (1910). The fact that Berkefeld filtrates often proved infective afforded ground for the belief that it was a filtrable virus. The more recent observations, however, of Barnard (1926), Smiles (1926), Ørskov (1927), Nowak (1929), Wroblewski (1931), Ledingham (1933), Kheneberger (1934), Tang *et al* (1935, 1936), Turner (1935), and Merling Eisenberg (1935) have rendered it probable that only the tiny granular or elementary forms, and the plastic filamentous forms, are capable of passing through coarse filters.

The second organism of this group was described by Bridre and Donatien (1923, 1925), who isolated it from sheep infected with contagious agalactia (see Chapter 84). In 1935 Kheneberger reported the occurrence in cultures of *Streptobacillus moniliformis* (*Actinomyces muris*) of a pleuropneumonia like organism now referred to as L1, living apparently in symbiosis with the bacillus. This discovery formed the start of a fruitful series of investigations. By means of the technique that Kheneberger described for their cultivation, she and numerous other workers during the following years succeeded in isolating, mainly from rats and mice, a number of different species of pleuropneumonia like organisms. The existence of similar saprophytic organisms was demonstrated in sewage by Laidlaw and Elford (1935), and confirmed by Seiffert (1937a, b).

The systematic position of these organisms has given rise to much discussion. The curious association of L1 with *Streptobacillus moniliformis* has been in particular the subject of wide speculation. Though Klieneberger herself regards it as a symbiont, other workers, notably Dienes (1939) and Heilman (1941) maintain that it is merely a variant form of the bacillus. It is too early as yet to form any sound judgment on these conflicting views. The fact however that none of the numerous other pleuropneumonia like organisms has been found accompanying a given bacillus does tend in our opinion to support Klieneberger's contention. In this regard the close association of the genetically distinct fusiform bacilli and spirochaetes found in Vincent's angina will be recalled to mind.

Apart from their peculiar mode of reproduction the pleuropneumonia group of organisms seems to fall in between the bacteria on the one hand and the *Rickettsia* and filtrable virus group of organisms on the other. They are distinguished from *Rickettsia* mainly by their extreme pleomorphism and by their ability to grow on nutrient media in the absence of living cells. From bacteria they seem to differ less fundamentally but the filtrability of their smallest elements and their limited metabolic powers assign them a place at the lowest end of the bacterial scale. What relation they bear to the group of cocco-bacilliiform bodies believed by Nelson (1936a b 193 1940) to be responsible for fowl coryza and for infectious catarrh of mice and rats it is impossible to say though it may be noted that the organisms described by Nelson were less pleomorphic and grew only in the presence of living cells.

The nomenclature and classification of the pleuropneumonia group of organisms present grave difficulties. Ledingham (1933) would place them in the family *Actinomycetaceae*. Turner (1935) in a new order of *Borrelomycetales* and Sabin (1941) in a new class of *Paramycetes*. Klieneberger and Smiles (1940) agree with Sabin that their inclusion among the *Schizomycetes* is hardly justifiable. Until we know more about the genetic relationship of these organisms to the common bacteria it seems wiser to refrain from premature commitments and to suffer the discomfort of using a clumsy though comprehensive term like the pleuropneumonia group.

In the remainder of this chapter we shall give a fairly full description of the original type species, and brief accounts of the others. Those who wish for further information should consult the review by Sabin (1941) to which we ourselves are indebted.

### The Organism of Pleuropneumonia

**Cultivation.**—This organism which is sometimes referred to as *Asterococcus mycoides* (Borrel *et al* 1910) can be cultivated on a number of different media, but most workers have used serum broth or serum agar. Growth is said to occur under both aerobic and anaerobic conditions according to Turner (1935) micro-aerophilic conditions are most suitable. The optimum temperature for development is 34° C no growth occurs below 30° C (Tang *et al* 1935). With freshly isolated strains 2-3 day serum broth cultures often contain distinctive mucoid islands and threads visible to the naked-eye while dark-ground examination may reveal the presence of minute colonies the smallest of which is about 1.0  $\mu$  in diameter (Tang *et al* 1936). With older strains only a general cloudiness of the medium is seen. On solid media dew-drop colonies appear in 5 or 6 days. Under the microscope these are often umbonate and consist of a yellowish brown granular

centre surrounded by a smooth transparent peripheral extension (Fig 223) Well developed colonies may reach a diameter of 2 mm Cultivation is also successful on the chorio allantoic membrane of the developing chick embryo (Tang *et al* 1936) Swift (1911), however, found that both the organism of pleuropneumonia itself and seven other strains of the same group grew better on dead membranes, prepared by freezing the embryo for an hour with dry ice, than on living membranes—a point of possibly some differential importance from *Rickettsia* and the filtrable viruses On living membranes no constant macroscopic lesions were produced, and the embryo was not killed.

**Morphology**—The morphology of the organism is influenced by a number of factors, particularly the age of the culture and the method of examination Growth of organisms of the pleuropneumonia group appears to be accompanied by the



FIG 223—ORGANISM OF PLEUROPNEUMONIA

Surface colonies on serum agar ( $\times 140$ )  
(After Tang *et al*)



FIG 224—ORGANISM OF PLEUROPNEUMONIA

Elementary bodies, granules or conchoids Dark ground illumination ( $\times 3600$ )  
(After Turner)



FIG 225—ORGANISM OF PLEUROPNEUMONIA

Spheroid showing unipolar germination Dark ground illumination ( $\times 3600$ )  
(After Turner)



FIG 226 (Inset)—ORGANISM OF PLEUROPNEUMONIA

Spheroid showing multipolar germination Dark ground illumination ( $\times 3600$ )  
(After Turner)

FIG 227—ORGANISM OF PLEUROPNEUMONIA

Multipolar germination showing how the buds have moved away from the parent spheroid and are developing into filaments Dark ground illumination ( $\times 3600$ ) (After Turner)

formation of large amounts of cholesterol and cholesterol esters from the serum in the medium (Partridge and Kheneberger 1941) These bodies are present as the myelin forms of lecithin, and assume the most bizarre shapes (Williams 1941) In addition there is reason to believe that the protoplasm of the organism itself is in some stages of its development peculiarly plastic, and is readily distorted by external pressure or tension Since many workers have used different methods of examining cultures, and since most of these methods have entailed a risk of distorting the microbial elements, it is not surprising that the observations recorded have often been diverse, conflicting, and difficult to interpret More recently Kheneberger and Smiles (1942) and Kheneberger (1942) have described methods for examining cultures *in situ*, either by reflected light on a dark ground using annular oblique incident illumination, or by fixation and staining The use of these methods suggests that the developmental process of the pleuropneumonia group of organisms is simpler than had formerly been believed, and that the



majority of bizarre forms described by previous observers (see Ledingham 1933, Turner 1935, Tang *et al* 1935) were not essential morphological stages of growth, but were either myelin forms or the result of distortion of the plastic elements of the organism. Agreement, however, has not yet been reached on the true morphology and mode of development of the pleuropneumonia organism, and we shall therefore give (a) one account based on the older methods of examination of slide preparations by transmitted light on a dark ground or by the staining of impression films, and (b) another based on the more recent methods of examination of the organisms *in situ*, either by reflected light on a dark ground or by fixation and staining.

(a) *By Older Methods of Examination*—At the risk of undue simplification, we



FIG. 228—ORGANISM OF PLEUROPNEUMONIA

Stage of ramification showing nodes of protoplasmic condensation which form the starting point for the outgrowth of fresh filaments. Dark ground illumination ( $\times 1114$ ) (After Tang *et al*)

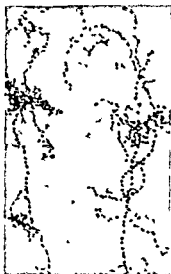


FIG. 229—ORGANISM OF PLEUROPNEUMONIA

Stage of chain formation, showing condensation of protoplasm at multiple points of filament. Giemsa ( $\times 1640$ ) (After Tang *et al*)

shall follow Tang and his colleagues (1935) in describing five morphological stages in the growth of this organism. (a) *Granular stage* Small granules, coccoid, diplococcoid, and cocco bacillary bodies are seen, usually  $0.15-0.4 \mu$  in diameter. Turner refers to them as "conidioids." They stain deeply with Giemsa, and may be regarded as a resting stage (Fig. 224). (b) *Filamentous stage* On inoculation into a fresh medium the granular bodies grow into spheroids about  $0.4-0.8 \mu$  in diameter, and develop on their periphery one or more spherical buds (Figs. 225, 226). These gradually move away from the parent body, but remain attached to it by a filament (Fig. 227). In old strains this filament is usually very short, and resembles with its terminal bud a sporing tetanus bacillus. In freshly isolated strains, however, enormously long filaments develop, sometimes crossing several fields of the microscope. Endomycelial protoplasmic streaming is often notice

able. The filaments stain very poorly with Giemsa. (c) *Stage of ramification.* During the process of streaming, areas of protoplasmic condensation appear at various points in the filament, and form the starting point for the outgrowth of fresh filaments. A tangled branching mycelium is the result (Fig. 228). In older strains this stage is lacking. (d) *Stage of chain formation.* In this stage the streaming protoplasm condenses rapidly at multiple points, so that the filament takes on a streptococcal appearance (Fig. 229). (e) *Stage of disintegration.* The chains break up, and set free a multitude of granular and coccoid forms, thus completing the cycle of development. In old cultures, in which the filamentous stage is lacking, the buds in the second stage become detached from the parent body and either grow out again into spheroids which themselves start budding, or remain in the granular phase.

It will be seen that reproduction occurs partly by budding and partly by fragmentation. Turner (1930) has described no fewer than five different methods of reproduction, and the reader who is interested will do well to refer to his beautifully illustrated paper. Again, reference must be made to Ledingham (1933) and

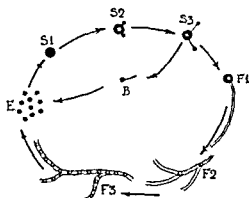


FIG. 230.—ORGANISM OF PLEUROPNEUMONIA.

Diagrammatic representation of developmental cycle according to Ledingham (1933), Klieneberger (1934), Turner (1935) and Tang and his colleagues (1936). Outer circle represents freshly isolated strain forming filaments. Inner circle represents old laboratory strain not forming filaments.

- F Elementary bodies granules or comidioids.
- S Spheroid.
- S<sub>1</sub> Bipolar germination of spheroid.
- S<sub>2</sub> Spheroid with buds still attached to it by very short filaments—drumstick appearance.
- F<sub>1</sub> Long filament growing out from spheroid.
- F<sub>2</sub> Filament showing ramification.
- F<sub>3</sub> Filament showing protoplasmic condensation preparatory to liberation of condensed particles as elementary bodies.
- B Buds detached from spheroid and becoming elementary bodies.

Klieneberger (1934) for the mode of origin of the vibronic forms, chromatic nodes, large oval swollen bodies, and other elements that are seen in preparations from colonies on solid media, and to Tang and his colleagues (1936) for a description of the apparently rare ameboid and giant ring forms. Fig. 230 represents diagrammatically the apparent sequence of development.

(b) *By Newer Methods of Examination.*—The picture obtained by the newer methods of examination referred to on p. 911, is much simpler. Following Klieneberger and Similes (1942), we may describe two methods of multiplication

(1) segmentation and (2) the formation of elementary corpuscles within a body surrounded by a limiting membrane (1) Starting as an elementary corpuscle or granule (Fig 231*a*) the organism grows into a small re-ovate sphere which may take on an irregular (*b*) or even filamentous (*c*) shape This then divides into a variable number of segments (*d* *e*) which may or may not separate from each other (2) Starting in the segments resulting from (1*b*) darkly stained bodies (*f*) probably consisting of nuclear material (see Klieneberger 1942) appear These then divide forming multiple elementary corpuscles within a body surrounded by a limiting membrane (*g*) Later each of these corpuscles is liberated apparently surrounded by a small portion of cytoplasm (*h*) This stage it may be remarked, recalls to mind the formation of merozoites in the developmental cycle of the malarial parasite Later still the cytoplasmic sheath is lost and the elementary corpuscles remaining (*a*) are indistinguishable from those from which the original cycle described in (1) began

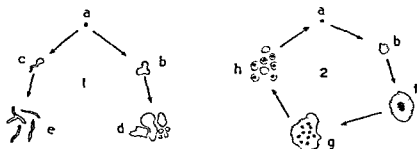


FIG 231—ORGANISM OF PLEUROPNEUMONIA

Diagrammatic representation of developmental cycle according to Klieneberger and Smiles (1942) and Klieneberger (1942)

1 Proliferation by segmentation.

Proliferation by formation of elementary corpuscles.

(a) Elementary granule

(b) Sphere

(c) Small filament

(d) Segments resulting from division of (b)

(e) Filaments resulting from division of (c)

(f) Nucleated bodies.

(g) Multiple granules within a limiting membrane resulting from division of (f)

(h) Liberated granules still surrounded by cytoplasm

**Resistance, Metabolism, Biochemical Reactions and Antigenic Structure**—According to Tang and his colleagues (1933), serum broth cultures may remain viable for 45 days at 37° C, and for 98 days at 0°–5° C. The organisms are bile-soluble, particularly in the filamentous stage, but are very resistant to ultra violet irradiation and to the photodynamic action of methylene blue (Tang *et al* 1936). They ferment glucose, maltose, and dextrin, and to a less extent sucrose, with the production of acid but not mannitol lactose, or salicin. Hemoglobin is reduced by freshly isolated strains. Warren (1942) states that growth is prevented by 10 per cent CO<sub>2</sub>, that a hemolysin is produced, that methylene blue is decolorized in the presence of lactate and that the time-potential curve differs from that of the L group of organisms. Little is known of the antigenic structure of this organism but there is some evidence, based mainly on cross protection tests of the existence of more than one immunological type.

**Pathogenicity**—The organism is naturally pathogenic to cattle. Experimentally the subcutaneous inoculation of 0.5–1.0 ml. of infected lymph or a virulent culture produces in 8 to 25 days a tense hot painful inflammatory swelling accompanied by high fever and often followed by death. Incision of the skin over the affected part is followed by the exudation of a clear straw coloured fluid often amounting to several litres. Post mortem the connective tissue meshes of the lungs are distended with an immense quantity of clear yellow fluid which is here and there coagulated into gelatinous trembling masses. Microscopical examination of the freshly collected fluid reveals the presence of forms similar to those seen in culture. The mycelial phase appears to predominate (Turner 1935). It is interesting in this connection to note that only freshly isolated strains showing the filamentous phase of development are fully virulent. There may be a little serous exudate in the pleural cavity and the thoracic and inguinal lymphatic glands may be affected. According to Daubney (1935) the typical disease can be reproduced by inoculation into the jugular vein of lymph or culture mixed with a few millilitres of 10 per cent agar. The emboli are held up in the lungs and form the starting point of the disease. Goats and buffaloes appear to be susceptible to experimental inoculation but laboratory animals are resistant (Nocard and Roux 1898 Tang *et al.* 1935).

#### The Organism of Contagious Agalactia

This organism was isolated by Bridré and Donat en (1923, 1925) from infected sheep (see Chapter 84). It has been studied by Nowak and Wroblewski (1930), Wroblewski (1931), Ledingham (1933) and Nowak and Lominski (1934). Its general characters are so similar to those of the pleuropneumonia organism that we do not propose to describe them separately. The disease can be produced experimentally by inoculation with pure cultures. The goat is more susceptible than the sheep. The subcutaneous inoculation of 0.5–1.0 ml. of a pure culture is followed in 4 to 7 days by the appearance of a small local swelling which disappears during the following week. After a further incubation period of 1 to 4 weeks localizing lesions appear in the joints, cornea and in lactating females the udder. The amount of milk secreted diminishes and a yellowish purulent fluid takes its place. Laboratory animals appear to be insusceptible.

#### Other Organisms of the Pleuropneumonia Group

It is unnecessary to describe in detail the individual features of the numerous other members of the pleuropneumonia group that have been isolated. All of them agree in being pleomorphic though differences are evident in the range and shape of the elements that are formed. All the parasitic species require for growth particularly on solid media a high proportion of animal protein. For this purpose a 30 per cent serum broth or agar of pH 7.6–8.0 to which 5 per cent of boiled blood may be added is generally suitable. The optimum reaction for growth appears to be about pH 7.8–8.0 below pH 7.0 growth generally ceases. The appearance and rate of development of colonies on solid media differ to some extent with different species. Most members grow better aerobically than anaerobically but the organism isolated from dogs and the L4 strain from rats are exceptions. Incubation in 10 per cent CO<sub>2</sub> is said to enhance the growth of L1 but to inhibit that of other L organisms (Warren 1949). Fermentation of sugars is very weak and the final pH reached is seldom lower than 7.0. Haemolysis is

reduction of hæmoglobin result from the growth of some strains. Antigenically there is a considerable degree of specificity though there is evidence of some group relationship among many of the members that have been studied (Kieneberger 1940). The parasitic species often seem to lead a harmless commensal existence in the body of their host though under certain conditions they may give rise to lesions of various types of which arthritis is one of the commonest. At least one species L5 forms an exotoxin acting on the central nervous system. Virulence is apt to decline rapidly in artificial culture. Pathogenicity is limited usually to a single host species. Some members are susceptible in the tissues to organic gold salts (Collier 1939a, Findlay *et al.* 1939).

The saprophytic species isolated from sewage earth and other situations differ from the parasitic species mainly in their simpler growth requirements—a high proportion of animal protein being unnecessary for their development—in their ability to multiply at 22° C. in their antigenic structure and in their absence of pathogenicity. We append brief notes on some of the commoner strains.

#### Pleuropneumonia Strains in Dogs

Shoetensack (1934) isolated in pure culture an organism sometimes referred to as *Asterococcus canis* from nasal secret on lung and liver of dogs suffering from distemper. Two antigenic types are recognized (Kieneberger 1938, 1940). The pathogenicity of these strains to dogs and their relationship to distemper are in doubt.

#### Pleuropneumonia Strains in Rats

**L1 strains**—Kieneberger (1935) described the isolation of a pleuropneumonia like organism from cultures of *Streptobacillus moniliformis* which is a normal parasite of the nasopharynx of rats (see p. 385). She obtained it in pure culture and showed that it had the same colony type characterized by a central granular part embedded in the agar medium and a flatter peripheral zone as the organism of pleuropneumonia. It was filtrable through Berkefeld V candles. By itself it appeared to be non pathogenic. From the lung of one rat it was isolated independently of *Streptobacillus moniliformis* (Kieneberger 1938). Though Kieneberger maintained that it was a symbiont, Dienes (1939, 1942), Heilman (1941), Smith (1941) and Brown and Vanemaker (1942) regard it as a variant of the bacillary organism. Kieneberger's (1942) most recent observations bring further evidence in favour of the symbiotic view. She shows that the developmental cycle of the L1 organism appears to be almost identical with that of the organism of pleuropneumonia as described by Kieneberger and Smiles (1942) and that the results of cross absorption tests made between *Streptobacillus moniliformis* and L1 are most easily explicable on the assumption that the two organisms are antigenically distinct.

**L3 strains**—Kieneberger and Steabben (1937) isolated this organism from the bronchiectatic lesions that are so common in the lungs of old rats. It is antigenically distinct from L1 (Kieneberger 1940). Though abscess formation results from subcutaneous inoculation of pure cultures into half grown mice it has so far proved impossible to reproduce the natural disease in rats (Kieneberger and Steabben 1940).

**L4 strains**—This organism was isolated first by Woglom and Warren (1933) from subcutaneous abscesses in rats following inoculation with a rat sarcoma strain. Its relation to the pleuropneumonia group was not recognized till, later in the same year, Kieneberger (1933) reported the isolation of an organism which she called L4 from the swollen submaxillary gland of a rat. It is antigenically distinct from L1 or L3. When inoculated subcutaneously or intraperitoneally into rats it gives rise to abscess formation. Inoculated intravenously with cells or agar it produces a severe arthritis, particularly in young rats. An organism labelled L7 was isolated by Findlay, Mackenzie, MacCallum and Kieneberger (1939) from spontaneous polyarthritis in the rat but this organism was later shown by Kieneberger (1939a) to be identical with L4. Its relation to the organisms described by Collier (1939b) and Beeuwkes and Collier (1942) which were like

were isolated from polyarthritis in the rat and to the organisms cultivated by Preston (1942) from the swollen joints and middle ear of rats is still in doubt. Gold salts appear to be of prophylactic value in protecting against arthritis caused by the inoculation of L4 into rats (Collier 1939a; Findlay *et al* 1940).

#### Pleuropneumonia Strains in Mice

Pleuropneumonia like organisms in the mouse's brain were demonstrated independently by Findlay and his collaborators (1938) in this country while investigating lymphocytic choromeningitis and by Sabin (1938) in the United States during the course of experiments with *Toxoplasma*. Several other strains biologically and immunologically distinct have since been isolated. Except in very young mice they seem to be normal parasites of the conjunctiva, nose and brain (Sabin and Johnson 1940).

*L<sub>0</sub> and Type A strains*—L5 was isolated by Findlay and his colleagues (1938) from the brains of mice suffering from rolling disease. A similar if not identical organism was isolated by Sabin (1938) who showed that it produced a true exotoxin having strongly neurolytic properties. Intracerebral inoculation of the organism gives rise usually after an incubation period of 2-3 days to a disease characterized by rolling movements on the long axis of the body. Some mice die and at post mortem extensive necrosis and lysis are found of the posterior pole of the cerebellum. Apparently the same organisms have been isolated from the lungs by Sullivan and Dienes (1939).

*L6 strains*—These strains were isolated by Findlay and his colleagues (1939) from the brains of mice that had been inoculated intracerebrally with the blood of splenectomized mice containing *Eperythrozoon coccoides*. They differ colonially and antigenically from L5 strains.

*M55 strain*—This organism was isolated from the swollen joint of a mouse by Jahn (see Kheneberger 1940). It is responsible for one type of mouse arthritis.

*B, C, D and E strains*—These organisms were isolated by Sabin (1938, 1939a, b) and by Sabin and Johnson (1940) from the conjunctiva, respiratory tract and brain of mice. All four types are able to give rise to arthritis on intravenous inoculation. What relation they bear to the L6 and M55 strains is not yet known.

*Edward's strains*—Edward (1940) isolated 7 strains from the lungs of normal mice. Some evidence was obtained that they might give rise to pneumonic lesions after nasal instillation but they did not produce arthritis.

#### Pleuropneumonia like Strains in Guinea pigs

The guinea pig so far has not been a fruitful source of pleuropneumonia like organisms. Kheneberger (1940) isolated a strain in conjunction with *Streptobacillus moniliformis* from abscesses in the neck but circumstances prevented its proper study. Smith (1941) likewise reported the isolation of *Streptobacillus moniliformis* from abscesses in guinea pigs but was unable to separate a pleuropneumonia like strain from the bacillary organisms.

#### Pleuropneumonia like Strains in Man

Dienes and Edsall (1937) isolated a strain from a suppurating Bartholin's gland in a woman who was working in the laboratory with rats. Subsequently Dienes (1940) demonstrated pleuropneumonia like strains in the cervical secretion of five patients suffering from pelvic infections. Whether these organisms are pathogenic or not is still undetermined. The frequent association of pleuropneumonia like organisms with arthritis in rats and mice has naturally raised hopes that they may be responsible for human rheumatism. In spite however of preliminary suggestive findings by Swift and Brown (1939) practically all attempts to demonstrate such a relationship have proved unsuccessful (see Sabin 1941).

#### Saprophytic Pleuropneumonia like Strains

Laidlaw and Elford (1936) described the isolation of three pleuropneumonia like strains A, B and C from London sewage by the inoculation of suitable filtrates into Fildes broth. The organisms grew best at 30°C in Hartley's horse digest broth pH 8.0 to which Fildes' peptic digest of red cells had been added. They fermented no sugars.

and were non pathogenic to animals. Strains A and C were antigenically distinct, strain B was more closely related to A than to C. Pirie (1937) found certain metabolic differences between strains A and C. Similar organisms have been isolated by Seiffert (1937a, b) from soil, manure and related substances. These saprophytic forms can grow in the absence of high concentrations of natural animal proteins.

## REFERENCES

- BARNARD J. E. (1936) *J. P. micro Soc.* p. 2,3.  
 BEEUWES H. and COLLIER, W. A. (1949) *J. infect. Dis.*, 70, 1.  
 BORDET J. (1910) *Ann. Inst. Pasteur* 24, 161.  
 BORDET, DUJARDIN BEAUMETZ, JEANTET and JOURAN (1910) *Ann. Inst. Pasteur* 24, 163.  
 BRIDGES J. and DONATIEN A. (1923) *C. P. Acad. Sci.*, 177, 841. (1925) *Ann. Inst. Pasteur* 39, 925.  
 BROWN T. M. and NICHOLSON, J. C. (1942) *Johns Hopkins Hosp. Bull.* 70, 201.  
 COLLIER, W. A. (1939a) *Z. Immunforsch.*, 95, 13. (1939b) *J. Path. Bact.* 48, 59.  
 DACHNEY P. (1935) *J. comp. Path.*, 48, 83.  
 DIENES, L. (1939) *J. infect. Dis.* 65, 24. (1940) *Proc. Soc. exp. Biol., N. Y.*, 44, 468. (1942) *J. Bact.* 44, 37.  
 DIENES, L. and EDGALL, G. (1935) *Proc. Soc. exp. Biol., N. Y.*, 36, 740.  
 EDWARD D. G. H. (1940) *J. Path. Bact.*, 50, 409.  
 ELFORD W. J. and ENDE, M. VAN DEN (1944) *Brit. J. exp. Path.* 25, 213.  
 FINDLAY G. M., MACKENZIE, P. D., MACCALLUM F. O., and KLIENEBERGER, E. (1935) *Lancet* ii, 1511. (1939) *Lancet* ii, 7.  
 FINDLAY G. M., MACKENZIE, R. D., and MACCALLUM F. O. (1940) *Brit. J. exp. Path.* 21, 13.  
 HEILMAN F. R. (1941) *J. infect. Dis.*, 63, 32, 45.  
 KLIENEBERGER, E. (1934) *J. Path. Bact.*, 39, 409. (1935) *Ibid.* 40, 93. (1937) *Ibid.*, 42, 387. (1938) *J. Hyg., Camb.* 38, 458. (1939a) *Ibid.*, 39, 260. (1939b) *J. Path. Bact.* 49, 451. (1940) *J. Hyg., Camb.*, 40, 204. (1942) *Ibid.*, 42, 485.  
 KLIENEBERGER, E. and SMILES J. (1942) *J. Hyg., Camb.*, 42, 110.  
 KLIENEBERGER, E. and STEARNS D. B. (1937) *J. Hyg. Camb.*, 37, 143. (1940) *Ibid.*, 40, 223.  
 LAIDLAW P. P. and ELFORD W. J. (1936) *Proc. roy. Soc., B* 120, 209.  
 LEDINGHAM J. C. G. (1933) *J. Path. Bact.*, 37, 393.  
 MERLING EISENBERG K. B. (1935) *Brit. J. exp. Path.*, 16, 411.  
 NELSON J. B. (1936a) *J. exp. Med.* 63, 515. (1936b) *Ibid.*, 64, 449. (1937) *Ibid.* 65, 501. (1940) *Ibid.* 72, 655.  
 NOCARD and ROUX. (1898) *Ann. Inst. Pasteur* 12, 240.  
 NOWAK, J. (1939) *Ann. Inst. Pasteur* 43, 1330.  
 NOWAK J. and LOMINSKI, I. (1934) *Ann. Inst. Pasteur* 53, 423.  
 NOWAK J. and WROBLEWSKI W. (1930) *Trans. Congr. int. Microbiol.*, 1, 619.  
 ORSKOV, J. (1927) *Ann. Inst. Pasteur* 41, 473.  
 PARTRIDGE, M. and KLIENEBERGER, E. (1941) *J. Path. Bact.*, 52, 219.  
 PIRIE, A. (1937) *Brit. J. exp. Path.*, 18, 96.  
 PRESTON W. S. (1942) *J. infect. Dis.*, 70, 150.  
 SARIN A. B. (1938) *Science* 83, 159-575. (1939a) *Ibid.* 83, 225. (1939b) *Ibid.*, 80, 18. (1941) *Bact. Rev.* 5, 1-331.  
 SARIN A. B. and JOHNSON B. (1940) *Proc. Soc. exp. Biol., N. Y.*, 44, 569.  
 SEIFFERT G. (1937a) *ZM. Bakt.* 129, 33. (1937b) *Ibid.*, 140, Beheft. p. 165.  
 SHOOTENACK, M. (1934) *Kansas Arch.* 11, 500.  
 SMILES J. (1936) *J. E. micro Soc.*, p. 25.  
 SMITH, W. (1941) *J. Path. Bact.* 53, 29.  
 SULLIVAN E. R. and DIENES L. (1939) *Proc. Soc. exp. Biol., N. Y.*, 41, 620.  
 SWIFT H. F. (1941) *J. exp. Med.*, 74, 557.  
 SWIFT H. F. and BROWN T. M. (1939) *Science* 89, 271.  
 TANG F. F., WEI H. and EDGAR J. (1936) *J. Path. Bact.* 42, 4.  
 TANG F. F., WEI H., McWHIRTER, D. L., and EDGAR, J. (1935) *J. Path. Bact.* 40, 391.  
 TURNER, A. W. (1935) *J. Path. Bact.*, 41, 1.  
 WARREN J. (1942) *J. Bact.* 43, 911.  
 WILLIAMS, S. (1941) *Aust. J. exp. Biol.*, 19, 200.  
 WOGLON, W. H. and WARREN J. (1938) *Nature* 87, 370.  
 WROBLEWSKI, W. (1931) *Ann. Inst. Pasteur* 47, 94.

serve to influence the results of all filtration experiments." The thickness of the filter also makes a considerable difference

The ordinary porous bacterial filter consists of a positively charged alkaline earth cation, and a negatively charged silicate anion. If a simple basic dye, such as methylene blue, which consists of an organic coloured cation united to an inorganic anion, is passed through a filter, a large amount of the dye will be adsorbed. The explanation of this is that the organic coloured cation enters into combination with the silicate anion in the filter forming an insoluble dye-silicate, which is retained; the soluble salt, such as NaCl or KCl, formed by the union of the alkaline earth cation in the filter with the inorganic anion of the dye, passes through. A similar phenomenon is observed in protein solutions. In solutions more acid than the isoelectric point of the protein, the dissociated protein is chiefly in the form of multivalent cations capable of entering into combination with the silicate anions in the filter, and forming an insoluble compound, which is retained. On the other hand, in solutions more alkaline than the isoelectric point the dissociated protein is chiefly in the form of multivalent anions, capable of entering into combination with the alkaline earth cations in the filter with the formation of soluble salts which pass through. This is probably why enzymes, toxins, and viruses appear to pass more readily through filters in weakly alkaline than in acid solutions (see Mudd 1922 23, 1923)

The nature of the suspending fluid plays an important part in determining the result. Several workers have noted that viruses pass much more readily through filters when the suspension is made up with broth or serum than with saline or phosphate buffer (Grinnell 1929, Ward 1929, Sawyer and Froehner 1929, Tallerman 1929, Marie and Urbain 1930, Galloway and Elford 1931). The mode of action of the broth is not known with certainty, but according to Elford (1933) it appears to be closely related to the ability of this medium to stabilize the dispersion of a lyophilic colloid. Soap has the opposite effect. Another factor, which is of special importance in comparing the filtrability of two different strains of virus, is the initial concentration of virus. The greater the number of virus particles present in the suspension, the more likely is virus to be found in the filtrate (Galloway and Elford 1931)

Cataphoresis experiments on such viruses as vaccinia, fowl pox, foot and mouth, rabies, yellow fever, myxoma, and Rous sarcoma have agreed in showing that most viruses carry a negative charge in neutral or nearly neutral suspensions (Douglas and Smith 1928, Findlay 1930, Hindle and Findlay 1930, Poppe and Busch 1930, Natarajan and Hyde 1930, Siebert Modrow 1930, Sankaran *et al* 1934). It is true that Oltzky and Boez (1927) stated that the foot and mouth virus carried a positive charge up to pH 8.0, but the results of these workers have not been confirmed. Most viruses have been studied over a range of about pH 5.0 to 9.0, they have been found to be negatively charged up to about pH 7.6, though the exact location of the isoelectric point has varied from pH 7.0 with the yellow fever virus to pH 9.3 with the virus of myxoma. Not too much attention however, should be paid to these measurements, since most of them have been carried out in the presence of tissue protein. Beard, Finkelstein and Wyckoff (1933), who worked with a relatively pure suspension of vaccinal elementary bodies, found their isoelectric point to be between pH 4.6 and 4.3. The nature of the charge carried will affect, to some extent, the passage of the virus through a filter. Incidentally, use may be made of the electric charge carried by the virus to free it from other material in a tissue suspension, or at any rate to obtain it in a more concentrated form. (Douglas and Smith 1928, Sankaran *et al* 1934)



It is important to realize that the mere passage of an organism through a filter candle does not justify its inclusion in the group of filtrable viruses. Even under conditions of careful experimentation small organisms particularly slender flexible and motile organisms such as spirochaetes frequently appear in small numbers in the filtrate, and conversely, the mere failure of an organism to pass through a filter candle does not justify its exclusion from the group of filtrable viruses. Some viruses for example such as those of varicella and herpes zoster, have not yet been shown to be filtrable, yet there is little doubt from what is known of their other properties that they should be included in this group. The term filtrable virus is one connoting a number of properties the most important of which have already been defined at the head of this chapter.

**Ultrafiltration**—In recent years ultrafiltration has been introduced. In this process thin collodion membranes are prepared with a given size of pore the size being determined largely by the concentration of collodion used. In the development of these filters Elford (1931, 1933) has played a prominent part. Starting from the earlier work of Bechhold, he has been able by the use of appropriate solvent mixtures and by the careful standardization of his technique to prepare a series of membranes of very regular and accurately graded porosities by means of which determinations of the size of many of the commoner viruses have been successfully made, and subsequently confirmed by other workers (see also Elford, Grabar and Ferry, 1935; Duclaux and Amat, 1938). These filters—*Gradocol membranes*—approach nearer to the mechanical sieve than do ordinary filter candles; they appear to be less influenced by the various secondary factors which we have mentioned and to be capable when properly used of sorting out particles very largely according to their size, though the influence of the pH of the suspending fluid and of the electrical charge carried must still be taken into account.

In calculating the size of the particle from the average pore diameter through which it just fails to pass the effect of adsorption has to be considered. This effect is most influential in membranes with very small pores (Table 58). Thus a particle held back by a membrane with an average pore diameter of 30  $m\mu$  probably has a diameter of 10–15  $m\mu$  while one held back by a membrane of 1 000  $m\mu$  probably has a diameter of 0.75–1.0  $\mu$  (Elford, 1933). This relationship however is disputed by Markham, Smith and Lea (1942) (see also Cox and Hyde, 1932; Asheshov, 1933 on ultrafiltration).

TABLE 58

RELATION OF SIZE OF RETAINED PARTICLE TO AVERAGE PORE DIAMETER OF GRADOCOL MEMBRANES (Elford, 1933)

Membrane average pore diameter	Size of Retained Particle
$m\mu$	
10–100	(0.33–0.5) $d$
100–500	(0.5–0.75) $d$
500–1 000	(0.75–1.0) $d$

$d$  = average pore diameter of limiting membrane for optimum filtration conditions

**Microscopical Examination**—There are difficulties in the microscopical examination of viruses. It has already been pointed out in Chapter 2 that under ordinary

conditions of examination it is impossible to resolve particles less than  $0.2 \mu$  in diameter. Resolution it will be remembered is limited by the numerical aperture of the objective and the wave-length of the light used. Since there are serious difficulties in increasing the numerical aperture of the objective it follows that the only way to resolve very small bodies is to use a wave-length shorter than any present in the visible spectrum. Resolution however is not always required, and considerable attention has been devoted of recent years to methods for rendering small bodies visible. Though certain filtrable viruses may be demonstrated in sections or smear preparations their study is greatly facilitated by obtaining them in a suspension relatively free from tissue cells and other gross matter. Usually this is done by differential centrifugation sometimes accompanied by filtration. The suspension can then be examined by one of the following methods.

I *Fixing and staining with a suitable dye*—Numerous workers, among whom Ledingham (1931) has been one of the foremost, have used this method. The dyes chosen are most frequently Giemsa's stain, or one of its modifications. By this means minute particles—the so-called elementary bodies—may be rendered visible in appropriate preparations. Since, however it is impossible to demonstrate very small particles by transmitted light, even when they are deeply stained it follows that this method is limited to the larger viruses. Its most conspicuous success has, in fact, been achieved hitherto with the virus of vaccinia, the diameter of which is about  $0.15 \mu$ . It has proved of particular value in the microscopical observation of agglutination where of course visibility and not resolution, of the aggregating particles is alone required.

II. *Dark-ground examination using visible light*.—Provided the particles under examination can scatter enough light and there is a sufficient difference of refractive index between them and the medium in which they are suspended, this method enables very small particles to be rendered visible even though they are incapable of resolution. It provides a useful means of direct microscopical observation of virus particles.

III *Photography in ultra violet light*.—Barnard (1925) has been the chief exponent of this method. After preliminary examination by method II, photographs are taken at particular wave-lengths in the ultra violet spectrum (see Chapter 2). Either transmitted or dark-ground illumination may be used. The former suffers from the disadvantage that the ability of viruses to absorb light is very low and the image so obtained is smaller than it otherwise would be. With dark-ground illumination there is strong contrast, and sharply defined images are obtainable though their size tends to be slightly too large. With a wave-length of  $257 m\mu$  particles as small as  $75 m\mu$  can be actually resolved. Their approximate size can then be determined from the mean of the images given by transmitted and dark-ground illumination. Theoretically, this method is open to almost unlimited extension, but in practice great technical difficulties are encountered.

*Fluorescence microscopy*.—This makes use of the ability of certain bodies to transmute the short invisible waves of ultra violet light to longer visible waves (see Claiberg 1939).

V *Annular oblique incident illumination*.—In this method a relatively opaque body like the chorio-allantoic membrane, can be examined by oblique illumination from above (see Himmelweit 193).

VI *Electron microscope*.—More recently this has provided a tool that bids fair to outstrip any method hitherto known. Though technical defects prevent

the realization of the full potentialities of this instrument, it is said to be possible now, under optimal conditions, to resolve particles as small as  $1 \mu\mu$  and to use magnifications up to 200 000 times (see Marton 1941, Stanley and Anderson 1941, Sharp *et al* 1942, 1943, 1944, Luria *et al* 1943, Taylor *et al* 1943)

**Centrifugalization**—There are considerable mechanical difficulties in constructing a machine that is sufficiently powerful to throw down very fine suspended particles, this difficulty is increased if the suspending fluid as is usually the case, has a specific gravity greater than water. The centrifugal force of a machine varies with the square of the rate of rotation, and directly with the distance of the centrifuged material from the centre of the plate. But neither of these factors can be increased indefinitely, because with increasing rate of rotation and with increasing diameter of the plate, a vibration develops that very largely counteracts the centrifugal force. Numerous other mechanical factors, such as the air resistance and the heat generated in the machine, come into play when high speeds are developed, and limit the rate and time during which the machine may be run. Nevertheless, serious attempts have been made in recent years to overcome these

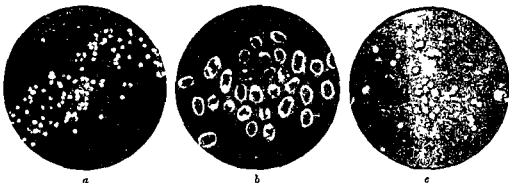


FIG 232—ON LEFT (a) ELEMENTARY BODIES OF VACCINIA FROM RABBIT TESTIS AND ON RIGHT (c) ELEMENTARY BODIES OF CANARY POX IN THE MIDDLE (b) *Chromo prodigiosum* FOR COMPARISON. PHOTOGRAPHED IN ULTRA VIOLET LIGHT ( $\times 3200$ ) (After Barnard)

difficulties, and very real progress has been registered. The introduction of higher speed electric motors, of the Lundgren angle centrifuge and of the spinning top centrifuge of Henriot and Huguenard (1925, 1927) (see also McIntosh 1935, McIntosh and Selbie 1937) have each contributed to this end. Indeed with the spinning top centrifuge, in which friction is diminished to a minimum, speeds of 80 000 r.p.m. have been reported. The greatest advance however has been made by Svedberg and his colleagues (1934) (see also Svedberg 1937), who have devised a centrifuge capable of revolving at 160 000 r.p.m. In this machine the rotor carries a cell which contains a column, 8 mm in height, of the fluid to be centrifuged situated 36 mm from the centre. The cell has windows of crystalline quartz to allow of serial photographs being taken to register the progress of sedimentation. The rotor is driven by two twin turbines fed with oil at a pressure of 15 kgm per sq. cm. Rotation takes place in an atmosphere of hydrogen at 25 mm pressure, so as to limit air friction and convection currents. In the original design (Svedberg and Nichols 1927) 240 litres of oil were required per minute to drive the turbines, and 7 litres of oil per minute to lubricate and cool the bearings. The cost of this machine has so far prohibited most laboratories from testing it, but in

Svedberg's hands it has been used with conspicuous success in estimating the molecular weight of proteins

Bauer and Pickels (1936-1937) have devised a high speed centrifuge combining some of the principles of the Henriot and Huguenard and some of the Svedberg model. Elford (1936) has shown that by introducing a capillary tube into the fluid to be centrifuged and collecting the virus particles on blotting paper at the bottom much of the difficulty caused by mixing can be overcome. Schlesinger (1936) has adapted the Sharples centrifuge for the concentration of virus by using a thin layer of suspension in a hollow cylinder rotating vertically so that the total distance the virus particles have to travel is a fraction of a millimetre. McIntosh and Sells (1940) have modified the Sharples centrifuge so as to permit the collection of the virus particles on a sheet of cellophane. This method has the advantage that a continuous stream of the virus to be purified can be fed to the machine. The preparation of pure virus suspensions is sometimes facilitated by preliminary tryptic digestion of the matrix in which they are embedded.

Not only is it possible now with some of the high speed centrifuges to throw down completely the larger viruses but their approximate size can be calculated from measuring their rate of sedimentation. Bechhold and Schlesinger (1931) have worked out a formula from which the size of evenly dispersed spherical particles submitted to a constant centrifugal force may be determined. Further by measuring the rate of concentration it can be ascertained whether the particles are of uniform size. It can be shown for instance, that the logarithm of the concentration of particles in the supernatant fluid is proportional to the length of time of centrifugation. If they are of unequal size the larger particles will be thrown down rapidly and the curve formed by plotting the logarithms of the concentrations against time will not be a straight line.

The larger viruses of  $0.1-0.2 \mu$  in diameter, can be thrown down completely under suitable conditions in about half an hour by a centrifuge revolving at 10 000 r.p.m. (see Armes 1933) while particles of about  $60 m\mu$  such as the staphylococcal bacteriophage require a speed of 40 000 r.p.m. maintained for 1 to  $1\frac{1}{2}$  hours (McIntosh 1935). Where centrifuges of only 3 000 r.p.m. are available and it is desired to concentrate the suspension the virus may sometimes be adsorbed on to kaolin, animal charcoal or blood corpuscles and the deposit subsequently suspended in a protein free medium (see Levaditi and Nicolau 1923, Gins and Krause 1923, Tang 1932, Francis and Salk 1942). Viruses vary however in their reaction to different adsorbing agents (Lewis and Andervont 1927) and this method is therefore not always successful. The purity of centrifuged virus suspensions is dependent on the number of particles of foreign matter present resembling in sedimentation rate that of the virus which is being concentrated. Methods for determining the degree of purity have been suggested by Smadel, Rivers and Pickels (1939) and Luria (1940).

**Morphology**—Information on the shape of filtrable virus particles has been furnished by Barnard who has been successful in photographing some of the larger viruses in ultra violet light. One of the most carefully studied is that of ectromelia, a virus with a diameter of about  $120 m\mu$  (Barnard and Elford 1931). This organism is coccoid and frequently occurs in pairs. Isolated organisms are spherical and highly refractile, the refractivity apparently decreasing with shortening of the wave length used for illumination. Reproduction is by binary fission and elongation is evident before division. The final separation of the two organisms takes place quickly, but a very fine connecting filament may be

left between them. Ultra violet photographs show a more highly refractive outline corresponding to the periphery of the cell and often an increased density at the poles. Whether these appearances are due to the presence of a cell wall and to polar condensation of the cytoplasm respectively or are merely the effects of interfacial phenomena it is impossible to say. The foot and mouth virus which is among the smallest of the viruses is according to Barnard (1937) rod shaped the length being sometimes as much as three times the breadth.

TABLE 59  
APPROXIMATE SIZES OF FILTRABLE VIRUSES

	mμ		mμ
Staphylococcus	1 000	Staphylococcus phage	} 50
Bovine pleuropneumonia spheres	} 300	Lymphocytic choriomeningitis	
Rickettsia		Durand's disease	
Psittacosis	275	Coli phage	} 40
Pseudo lymphocytic choriomeningitis	190	Shiga dysentery phage D <sub>54</sub>	
Vaccinia	} 150	Salmonella phage S <sub>41</sub>	
Canary pox		Rabbit papilloma	
Bovine pleuropneumonia particles		Megatherium phage	} 35
Rabbit fibroma		Infectious anaemia of horses	
Rabbit myxoma		Tobacco mosaic (long diameter)	33
Lymphogranuloma	} 100	Rift Valley fever	} 30
Sandfly fever*		American equine encephalomyelitis	
Herpes	} 105	Coli phage C <sub>36</sub>	} 25
Ectromelia		Flexner dysentery phages D <sub>13</sub> D <sub>10</sub> D <sub>48</sub>	
Rabies		St. Louis encephalitis	
Pseudorabies		Japanese encephalitis	
Borna disease	} 100	West Nile encephalitis	} 22
Newcastle disease		Louping ill	
Influenza B		Hemocyanin molecule (H <sub>1</sub> x)	} 18
Russian spring summer encephalitis	} 90	Coli phage C <sub>13</sub>	
Staphylococcus phage		Salmonella phage S <sub>13</sub>	
Influenza A	} 85	Yellow fever	} 10
Swine influenza (American)		Foot and mouth disease	
Vesicular stomatitis	} 75	Poliomyelitis	} 8
Rous sarcoma		Mouse encephalomyelitis	
Fowl plague	} 60	Edestin molecule	} 5.6
Staphylococcus $\phi$ phage		Serum globulin molecule	
Flexner dysentery phages D <sub>4</sub> D <sub>19</sub>		Oxyhaemoglobin molecule	} 4
		Egg albumin molecule	

\* Alternative size given as 90-140 mμ.

Further information on the shape size and structure of the viruses is being afforded by the electron microscope. By its use the elementary bodies of vaccinia appear to be brick shaped to possess some sort of limiting membrane and to contain fine circumscribed areas of greater density arranged like the spots on dice (Green *et al.* 1942) (see Fig. 233).

The size of many viruses has now been calculated mainly from data based on the use of Elford's gradocol membrane technique and to a less extent from the microscopical photographs of Barnard the electron micrographs of American workers high speed centrifugation and the rate of diffusion of virus particles in a suitable medium. These different methods do not always agree in their results. By the filtration method for example the figure reached tends to correspond to the size of the smaller particles, by the centrifugation technique to the

size of the larger particles. The size of the smaller viruses may be overestimated by ultra violet microscopy because of failure of perfect resolution (see Barnard 1937). Electron micrographs vary somewhat according to the mode of preparation of the film. Conclusions drawn from the rate of sedimentation in a gravitational field are affected by the density of the virus particles, and density appears to be variable and is difficult to determine with accuracy (see Smadel *et al* 1935). Nevertheless the degree of concordance is sufficient to justify us in assigning with some confidence mean particle diameters to the more important viruses. In Table 59 the size of the commoner viruses and some of the bartenophages is

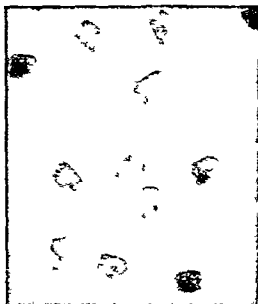


FIG. 233.—ELECTRON MICROGRAPH OF VACCINIA VIRUS ELEMENTARY BODIES ( $\times 22,000$ )  
[From Green *et al.*, 1942.]

lipin arranged around the particle in two or three loose layers, but no real membrane. These results suggest that the composition of the larger virus particles is essentially similar to that of ordinary bacteria. The nature of the smaller viruses is in greater doubt, Stanley's work (see p. 205) indicating that they may be no more in fact, than macromolecules of protein. (For a general review of the properties of vaccinal elementary bodies, see Smadel and Hoagland 1942.)

**Habitat.**—With the exceptions noted below all the filterable viruses at present known are associated with living cells whether in the animal or the vegetable kingdom. This does not mean that they are never found apart from disease processes, for their presence has been demonstrated in healthy carriers. But it does mean that they are essentially parasitic. The existence however of saprophytic viruses is suggested by the work of Barnard (1935). Hitherto the only satisfactory criterion of the presence of a virus has consisted in the production of characteristic lesions in a susceptible animal by a suitably prepared filtrate—a technique which may be remarked, that automatically excludes the discovery of a saprophytic virus

compared with *Staphylococcus* on the one hand and the larger protein molecules on the other.

The method of reproduction of the filterable viruses is still in doubt. The evidence so far obtained seems to favour binary fission. Eisenberg Merling (1943) has described a complex life cycle for the vaccinia virus but his observations and the conclusions he draws from them must await confirmation.

**Microchemical analysis** of vaccinal elementary bodies has revealed the presence of a h. carbohydrate fat and nitrogen, a part of which is undoubtedly in the form of protein (Hughes *et al* 1935). Further observation by Hoagland, Smadel and Pivens (1946) have led to the identification of neutral fat, phospholipin, reducing sugar after hydrolysis and thymonucleic acid. McFarlane and his colleagues (1939) believe that there is a shell

or of a completely avirulent variant of a parasitic virus. Barnard, however, by ultra violet photography has been able to detect the presence of minute cultivable bodies, about 150 m $\mu$  in diameter, in sterile tubes of serum broth.

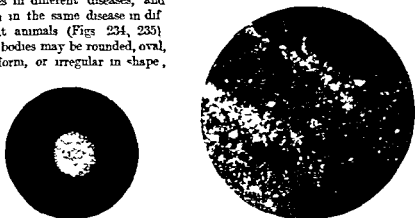
Many of the viruses in the animal body appear to show a particular affinity for special tissues such as nervous tissue or skin, this resembles the affinity manifested by many of the known bacteria for special tissues. Even however, when the lesions are confined to one tissue, the virus can frequently be demonstrated in other parts of the body. There is evidence, too, that the tissue localization is more apparent than real, depending on the mode of infection. Thus Ledingham (1924) found that in rabbits the virus of vaccinia, which usually affects the skin, was able to give rise to nodules on the peritoneum after direct inoculation into the spleen or the abdominal cavity. As well as their selective tissue localization many viruses exhibit a species specificity, giving rise to lesions only in one particular species of animal. Thus Cole and Kuttner's salivary gland virus is active only in guinea pigs, Virus m only in rabbits, and so on. On the other hand there are viruses, such as those of rabies and foot and mouth disease which are pathogenic not only to different species but also to widely separate groups of animals. Possibly too much weight has been laid in the past on the species specificity of the viruses. It is now clear that most of the viruses are capable of infecting several different species of animal under experimental conditions.

Apart from the presence of a virus in a healthy carrier free from all clinical symptoms of disease, it has been shown that a virus may remain latent in the tissues after causing an initial infection. Thus, according to Gastinel and Reilly (1928) the herpes virus can sometimes be demonstrated in the brain of guinea pigs that have recovered from a keratitis caused by inoculation of the cornea. Its presence gives rise to no symptoms, but can be shown by inoculation of the brain on to the cornea of a normal guinea pig. It is possible that an attack of intercurrent disease, or some artificial procedure such as vaccination may activate such a latent virus, and cause it to give rise to clinical disease. In some virus infections, such as yellow fever, there is reason to believe that the virus after causing an attack of the disease, remains latent in the tissues for years if not for the patient's whole life. By this means a lasting immunity is maintained (see Rivers 1943).

Whether the filtrable viruses occupy an intra or an extracellular position in the body is not certainly known but the indirect evidence so far accumulated suggests that their growth and multiplication occurs actually within the cells. The rinderpest virus for example, appears to be contained within the leucocytes by centrifugalization of the blood, the virus is found to be concentrated mainly in the leucocytic layer. Similarly with the virus of fowl plague Todd (1928) found that in centrifuged blood the concentration of the virus was 100 times greater in the leucocytic layer than in the clear plasma or the washed red cells. Moreover there is evidence that for their multiplication the filtrable viruses often prefer young newly formed cells, many of the viruses acting on the skin, for example give rise to lesions first along the lines of scarification where repair is taking place. Further evidence in favour of this view is that certain viruses e.g. Virus m and vaccinia have been found to grow in a transplantable rabbit tumour, in which the cells are in process of active multiplication, and to survive longer in the tumour than in the healthy tissues of the rabbit (Rivers and Pearce 1925). The observations of Perdrau and Todd (1936) on the lower susceptibility

of viruses to ultra violet light in the presence of cells, particularly dividing cells than in their absence also suggest that viruses occupy an intracellular position and multiply most readily in growing cells. How far, in fact, the viruses are *cytotropic* and how far they are *cytotrophic* is a matter for dispute—if in fact any clear distinction can be drawn between these two properties. Goodpasture (1930) believes that actual growth occurs only in the living cells of the body, while Ledingham (1932) is not prepared to go to this length. Probably they take advantage of ferment action in the body cells, and receive their nutritive material in a partly digested state. If this is so, then viruses, including the bacteriophage, must be among the most dependent parasites of which we have knowledge in the unicellular world.

**Inclusion Bodies**—Histological examination of the lesions occurring in filtrable virus diseases often reveals the presence within the cytoplasm or the nucleus, or sometimes both, of peculiar bodies whose nature is at present unknown, and which are usually referred to as "inclusion bodies." The appearance of these bodies varies in different diseases, and often in the same disease in different animals (Figs 234, 235). The bodies may be rounded, oval, pyriform, or irregular in shape,



Ectromelia virus on left (Fig. 234) inclusion body from foot of mouse, on right (Fig. 235) inclusion body after maceration, showing the liberated elementary bodies. Photographed in visible light ( $\times 1200$ ) (After Barnard)

their substance may be hyaline or granular, in structure they may be homogeneous, or they may contain one or more, often several, elementary corpuscles, in their staining reactions they may be basophilic or acidophilic, and within the same inclusion body the granules or elementary corpuscles may stain differently from the ground substance, lipid substances staining with osmic acid are sometimes found. In many diseases affecting the skin, such as fowl pox, human variola, and the common wart, the formation of inclusion bodies is restricted to the epidermis, but in others, such as zoster, varicella, and venereal herpes, they are found both in the epidermis and in the corium. Moreover, according to Lipschütz (1920), only certain layers of the epidermis may be affected, thus in the common wart, inclusion bodies are found in the prickle- and horn-cell layers but not in the basal cell layer. Inclusion bodies can be produced experimentally only by the inoculation of living viruses, they are not formed after inoculation of dead viruses, even though the latter have immunizing properties, e.g. vaccinia and herpes. After inoculation of the virus, the inclusion bodies appear at different times in different infections. Thus in common warts, the nuclear inclusion bodies are demonstrable



only in the earliest stages, in herpetic keratitis of rabbits the inclusion bodies appear within the first 24 hours, in venereal herpes they are best seen on the third day, and so on. There appears to be some relationship between the presence of inclusion bodies and the infectivity of the tissue, in herpetic keratitis of rabbits for example it is said that with the disappearance of the nuclear inclusion bodies the disease can be no longer propagated. Experimentally the formation of inclusion bodies can be stimulated by the inoculation not only of infected tissue extracts, but often of filtered cell free material. They can moreover often be demonstrated in tissue cultures (Andrews 1929 Rivers *et al.* 1929).

The earlier workers regarded these inclusion bodies as protozoa and pictured them as varying stages of an elaborate life-cycle. Subsequently they were believed to represent cellular degeneration products due to nucleolar extrusion, vacuolation of the cytoplasm and other processes consequent on the attack of the virus. Von Prowazek regarded them as of a dual nature consisting of micro-organisms embedded in material deposited around them as the result of a reaction of the cell protoplasm, for these bodies the term *Chlamydozoa*—literally cloak animals—was proposed. It is now however becoming increasingly clear that intracellular inclusion bodies are essentially colonies of the infecting virus. Since Woodruff and Goodpasture (1929, 1930) showed that the Bollinger inclusion body of fowl pox consisted of 10 000–20 000 minute Borrel bodies and that a single Bollinger body, washed free from surrounding virus, was capable of giving rise to a typical fowl pox lesion on skin inoculation, it has been difficult to regard inclusion bodies as other than intracellular aggregations of elementary virus particles. By tryptic digestion, by maceration by surface tension or other means it has now been shown that the inclusion bodies of ectromelia (Barnard and Elford 1931), vaccinia (Ledingham 1931, Paschen 1932) and psittacosis (Bedson and Bland 1932, 1934) contain masses of elementary bodies which are apparently responsible for giving rise to characteristic intracellular changes. Moreover the formation of inclusion bodies from elementary bodies has now been watched experimentally in the chorio-allantoic membrane and the rabbit's cornea after inoculation with vaccinia virus (Herzberg 1936, Tang and Wei 1937, Himmelweit 1938, Bland and Robinow 1939). Whether the intranuclear acidophilic bodies which are so common in infections caused by neutrotropic viruses are of the same nature as the intracellular but extranuclear bodies has not yet been made clear either by morphological study or by the micro-encimeration technique (see Cowdry 1933). There is some evidence that they result from flocculation of the nuclear colloids (Findlay 1939). Whatever the structure of inclusion bodies may be however there is no doubt whatever of their significance: their presence in the tissue is a sure sign of infection and is made use of in the routine diagnosis of certain of the filtrable virus diseases such as rabies. (For a pictorial review of inclusion bodies see Findlay and Ludford 1926 and for a general account of their properties see Goodpasture 1929, 1929–30, Ledingham 1935. See also Figs 298–302.)

**Cultivation.**—With the possible exception of saprophytic viruses cultivable in serum broth the filtrable viruses have proved refractory to cultivation in the absence of living cells. In 1915 Noguchi succeeded in obtaining pure cultures *in vivo* of vaccinia virus by growing it in the testicles of rabbits and bulls. The virus obtained from skin scrapings was first freed from bacteria by suitable means and was then inoculated intratesticularly into rabbits. Transfers were made every four days. Several passages were necessary before the virus became adapted to growth

in the testicle, but subsequently transfers were made without difficulty, and the virus reached its maximum multiplication in the testicle after 4 or 5 days, it remained stationary in amount till the 8th day, and then decreased till after 5 weeks its presence could no longer be detected. By testicular cultivation, the virus did not lose its affinity for the skin, both the testicular and the skin strains gave similar reactions in the skin, cornea, and testicles of rabbits, and in the skin of human beings. In 1925 Parker and Nye succeeded in growing the vaccinia and herpes viruses in tissue cultures prepared with normal rabbit testis and plasma. This was confirmed by Carrel and Rivers in 1927 working with the vaccinia virus. Infected rabbit testicle was ground up in a mortar, added to chick-embryo pulp, left for 24 hours in the ice-chest, and then inoculated into a Carrel flask containing a coagulum of hen plasma. The cultures were washed every 2 or 3 days with Tyrode's solution, and were nourished with a dilute fowl-embryo extract. After periods varying from 1 to 4 weeks the contents of the flasks were withdrawn, ground up in a mortar, and titrated on rabbits by the intradermal method. It was found that the cultures, which were seeded with 25 to 250 intradermal units of virus per ml., contained after incubation for a week between 10,000 and 100,000 units per ml. showing that actual multiplication of the virus had occurred. Haagen (1926) reported a modified method of *in vitro* cultivation of vaccinia virus in the presence of rabbit testicle, rabbit plasma and rabbit spleen extract. Using this method, he carried the virus through 37 passages in a period of about 8 months. During the first 5 days of each subculture the virus multiplied about 1,000 times, and during the whole period its virulence remained approximately constant. Findlay (1928) reported similar success in the cultivation of the fowl pox virus by Carrel's method. Cracium and Oppenheimer (1926) cultivated vaccinia virus in association with embryonic guinea pig's cornea, and carried through nine successive transfers during a period of 71 days. Finally the *in vitro* cultivation of vaccinia virus in the apparent absence of proliferating cells was reported by the Matlands (1928). Infected rabbit testicle was ground up, diluted with Tyrode's solution, added to minced hen's kidney, placed in the cold room for 4 hours, and then cultivated in a Carrel flask containing hen's serum. The flasks were incubated at 37° C., and subcultures made about once a week. Four passages were made in all. A  $1/625$  dilution of the last subculture, which represented a dilution of  $1/625 \times 10^8$  of the primary inoculum, produced vaccinia on inoculation into rabbits, the original inoculum was active only in a  $1/2500$  dilution. Living tissue was, of course, not excluded, but no evidence of its multiplication was obtained. Various modifications of the Matland technique, such as cultivation in agar slant cultures (Kurotchkin 1939), wide tubes (Findlay and MacCallum 1940), and roller tubes (Feller *et al.* 1940) have been successfully introduced.

The function of the tissue cells in the cultivation of viruses has not been established, though much is known about the various factors that influence growth (see Hallauer 1935). Zinsser and Schoenbach (1937) observed that development of the equine encephalitis virus reached its maximum in 3 to 4 days, whereas that of *Rickettsia* was considerably later—6th to 8th day. They therefore drew the tentative conclusion that the metabolism of viruses and of rickettsiae differed, the former multiplying best during the period of active growth of the tissue cells, the latter after the cells had begun to die off. Matland and Laing (1941) have brought evidence to suggest that with vaccinia virus the cells exert two separate functions, one to initiate growth, the other to maintain growth once the virus

has started to multiply. That the growth factors supplied by the cells are relatively specific for different viruses is suggested by Andrewes' (1942) observation that growth of one strain of a given virus may render the tissue culture medium unsuitable for growth of another strain of the same virus added 24 hours later, though not for a strain of a different virus. This is analogous to experience in the growth of bacteria on lifeless media, and suggests that some substance required for the growth of a particular species is soon exhausted. As Rivers (1932) points out, many viruses in tissue culture exhibit both a species and a cellular specificity. Fowl plague virus multiplies only in the presence of chick embryo skin and brain, not in cultures of fibroblasts, moreover, avian tissue appears to be essential.

None of the filtrable viruses has yet been cultivated in the absence of living cells. It is true that Eagles and McClean (1931) and Eagles (1935) state that they have grown vaccinia virus in a cell free medium, but their results have not so far been confirmed (Matland *et al.* 1932, Rivers and Ward 1933). It may indeed be questioned whether such highly parasitic organisms as the viruses appear to be are provided with sufficient enzyme systems to enable them to grow in the absence of the cellular activity of their host. The successful cultivation of the pathogenic viruses on lifeless media may well have to await the reproduction *in vitro* of the complete ferment mechanism of the living cell.

Most of the common viruses have now been grown in tissue culture. Another method, which has come into increasing prominence of late years, makes use of the developing hen's egg. It was introduced by Ogston in 1881 for the cultivation of bacteria, and was re-discovered by Woodruff and Goodpasture (1931) fifty years later. Inoculation of the chorio-allantoic membrane is the most generally useful way of applying this technique. Some viruses, like vaccinia and psittacosis, produce characteristic lesions or pocks on the membrane; others, like fowl plague and vesicular stomatitis, kill the embryo before local lesions have had time to develop; others like Rift Valley fever and influenza, produce both local membrane lesions and characteristic effects on the embryo; and others, like poliomyelitis, rabies, and foot and mouth disease viruses, fail to grow on the egg membrane at all. Burnet has been particularly fertile in recognizing the potentialities of this method, and readers who are interested in its technical details and general application would do well to consult his monograph (1936). (See also Stevenson and Butler 1939, Burnet and Faris 1942, Dunham and MacNeal 1942.) Inoculation into the yolk sac or the amniotic sac, or even directly into the embryo, may be used for certain purposes.

**Resistance**—The filtrable viruses vary considerably in their resistance to noxious agencies. Generally speaking, they resemble the vegetative bacteria more closely than the spore bearing organisms, that is to say they are generally destroyed by exposure to moist heat at 55–60° C within half an hour, and succumb to fairly low concentrations of chemical disinfectants. On the whole, they appear to be more resistant than the vegetative bacteria to chemical agencies, but it must be remembered that experiments can never be performed in the complete absence of cellular, or at any rate, of protein material, their apparently greater resistance may, therefore, be due to the protective action of substances in the medium.

The effect of desiccation varies, partly with the method employed, and partly with the particular virus in question. The Foot and Mouth Disease Research Committee (Report 1927) found that filtered vesicle fluid from the guinea pig if dried rapidly on slides at 37° C, was often inactivated immediately, on the other

hand if dried slowly at room temperature and kept at room temperature over  $H_2SO_4$ , it survived for 3 to 6 months. Noguchi (1918) found that when dried vaccinia virus remained alive for over a year but its virulence was considerably decreased. Haagen (1939) on the other hand observed no loss of virulence in a year when mouse brain infected with vaccinia was dried over calcium chloride and kept in sealed glass vessels in the ice-chest. One hour's exposure to the August sun of the foot-and-mouth virus, dried on a glass slide inactivated it. Most viruses appear to be very resistant to cold. Vaccinia virus withstands a temperature of  $-180^\circ C$  for months and even repeated freezing and thawing fails to destroy it. Frozen and dried they may live for months (Sawyer *et al* 1929, Wooley 1939). Vaccinia virus in dry powdered form withstands dry heat at  $100^\circ C$  for 5 to 10 minutes. Most heat at  $55-60^\circ C$  for half to one hour is fatal to most viruses but blood from swine fever is said to withstand a temperature of  $58^\circ C$  for at least 2 hours. It is inactivated however within an hour by a temperature of  $78^\circ C$ . Some viruses, like the poliomyelitis virus, are rapidly killed by ultra violet light. Many are also susceptible to photodynamic action (see Chapter 5) and succumb in a few minutes when exposed to a concentration of about 1/100 000 methylene blue in the presence of daylight (Perdrau and Todd 1933, Herzberg 1933, Shortt and Brooks 1934). Alpha rays, X rays and gamma rays are all lethal (Lea and Salaman 1942).

Most viruses exhibit a fairly high resistance to glycerol and one of the best methods of preserving infectious tissue is to suspend it in 50 per cent glycerolated saline, cover it with liquid paraffin and store it in the ice-chest (Perdrau 1927). Pure glycerol destroys the viruses fairly rapidly as a rule, thus Noguchi (1918) found that vaccinia virus was destroyed by pure glycerol at  $4^\circ C$  within 24 hours, though in 40 per cent glycerol it survived for about 6 months. The preservative action of glycerol probably depends on the inhibition it exerts on autolysis of the infected tissue (Rivers 1928).

Survival in distilled water, saline or Ringer's solution varies considerably. In the ice-chest many viruses will survive for a long time but most of them perish rapidly if kept at room temperature or  $37^\circ C$ . The foot and mouth virus in saline rarely survives at  $3^\circ C$  for more than 24 hours, and the lymphocytic choriomeningitis virus is non-infective within 3 hours at  $20^\circ C$  (Lepine *et al* 1931). Susceptibility to oxidation may be chiefly responsible for this behaviour. At low temperatures the presence of tissue cells seems to be beneficial to the survival of viruses but at higher temperatures the reverse is probably true. Ames (1931) for example found that vaccinia virus remained virulent much longer at  $31^\circ C$  when stored in the form of a suspension of elementary bodies than in tissue culture. In general the presence of serum or 0.5 per cent agar is beneficial for survival, as is also storage under anaerobic conditions (Zinsser and Tang 1929, Zinsser and Seastone 1930, McClean and Eagles 1931). The optimum H ion concentration for survival of the foot and mouth virus and the vaccinia virus is pH 7.6 between pH 4.0 and 3.0 vaccinia virus is rendered non-infective within about an hour (Beard *et al* 1933).

*Disinfectants*—Vaccinia virus in testicular suspension is said to survive in 0.5 and 1.0 per cent phenol solutions for over a year at  $4^\circ C$ , but to be destroyed by 2 per cent phenol within 24 hours and by a 1/30 000 solution of iodine within 1 hour at  $37^\circ C$  (Noguchi 1918). Likewise ectromelia virus in a suspension of mouse liver will remain virulent in 0.5 per cent phenolized saline at  $4^\circ C$  for

several months. Purified influenza virus on the other hand, is destroyed by 0.5 per cent phenol at 4° C within a week (Knight and Stanley 1944). Gordon (1925) found that vaccinia virus was destroyed by 50 per cent ethyl alcohol, 50 per cent methyl alcohol, and 50 per cent acetone within an hour at room temperature, 20 per cent ethyl alcohol, 10 per cent methyl alcohol, 10 per cent acetone, and 20 per cent ether failed to destroy it in 24 hours, even 50 per cent ether did not destroy it completely in this time. Potassium permanganate was found to be extremely viricidal, destroying it even in a 1/10,000 solution within an hour at room temperature. Chloroform is said to be very much more destructive than ether, alcohol, or acetone (Reynolds 1928). The foot and mouth virus is resistant to concentrations of phenol, lysol, toluol, hydrogen peroxide, chlorine, iodine, acetone, and chloropicrin that rapidly destroy vegetative bacteria, but it is killed by 0.1 per cent formal at 26–27° C in 24 hours, and by 2 per cent. antiformin or 0.4 per cent HgCl<sub>2</sub> within 24 hours. The effect of bile salts varies according to the species of virus (Smith 1939). Influenza A and louping ill viruses are inactivated almost instantaneously by exposure at room temperature to a final concentration of 1/1,000 sodium deoxycholate, whereas vaccinia and ectromelia viruses are unaffected after 2 hours. Inactivation of susceptible viruses is thought to be due to lysis. Certain soaps and unsaturated fatty acids, and some synthetic detergents, have been found to have a strong destructive action on influenza virus (Stock and Francis 1940, Knight and Stanley 1944).

With the very doubtful exception of the lymphogranuloma virus (MacCallum and Findlay 1938, Rodaniche 1942), the mouse pneumonia virus and the viruses of trachoma and inclusion blennorrhoea (Rake *et al* 1942), the viruses are insusceptible to sulphonamides. Penicillin likewise appears to be without action.

**Metabolism**—Practically nothing is known about the metabolism of the filtrable viruses, one of the great hindrances being the impossibility of cultivating them in the absence of tissue cells. In purified suspensions of vaccinia virus Parker and Smythe (1937) could demonstrate no oxygen consumption. On the other hand Macfarlane and Salaman (1938) and Macfarlane and Dolby (1940) though finding no evidence of dehydrogenase activity, were able to demonstrate the presence of phosphatase and catalase. Both ribonucleic acid and adenylic acid were rapidly hydrolysed. It is not yet clear, however, whether these enzymes form an integral part of the elementary bodies or are derived from the host tissues (see Smadel and Hoagland 1942). The ability to oxidize cysteine has been traced to the presence of copper in vaccinia elementary bodies (Hoagland *et al* 1941). Flavin and biotin have also been found (see Rivers 1943). What part these various enzymes play in the metabolism of the larger viruses under natural conditions, it is too early to say.

**Antigenic Structure**—Although the study of the antigenic structure of viruses is as yet in its infancy, enough has been learned to show that, in this respect, viruses differ in no essential way from bacteria. The presence of precipitins in the blood serum of animals inoculated with vaccinia virus has been reported by several workers (see Sobernheim 1925). Gordon (1925) found agglutinins and complement fixing bodies in the serum of rabbits inoculated with vaccinia, active up to a dilution of 1/100–1/200, both antibodies were specific in the sense that they reacted solely with vaccinia and variola virus suspensions, and gave no reaction with variella virus, sterile pus, or brain suspensions from encephalitis lethargica. In general

precipitins have been demonstrated more frequently than agglutinins or complement fixing bodies

Recent work, particularly with vaccinia, has revealed an antigenic complexity in the viruses similar to that present in many bacteria. The elementary bodies of vaccinia appear to contain two agglutinogens, one of which L is destroyed by exposure to 56° C for one hour, the other of which S, withstands a temperature of at least 95° C for this time. By suitable methods precipitinogens can be extracted from infected material and from tissue cultures which appear to correspond to the agglutinogens in the elementary bodies (Craigie 1932, 1935, Smith 1932, Craigie and Wishart 1934a, b, 1936, Ch'en 1934, Salaman 1934, Parker and Rivers 1937, Smadel *et al* 1940a). It was thought at first that the two antigens were quite separate and that the heat stable precipitinogen was a polysaccharide hapten, but more recent work has shown that both the L and the S substances are contained in a single protein molecule (Shedlovsky and Smadel 1942, Shedlovsky *et al* 1943, Smadel *et al* 1943). In addition, two further antigens have been demonstrated in vaccinal elementary bodies. One is the so-called NP or nucleoprotein antigen described by Smadel, Rivers and Hoagland (1942), and demonstrable by precipitation. The other is the X antigen, which can be demonstrated by agglutination using a serum from which the LS and NP antibodies have been removed by absorption (Craigie and Wishart 1936, 1938, Smadel *et al* 1942). Soluble antigens capable of reacting in complement fixation or precipitin tests, have been demonstrated in other viruses, such as the viruses of lymphocytic choriomeningitis and lymphogranuloma.

A further analogy with bacteria is afforded by the demonstration of multiple antigenic types in a single species of virus. For example, at least three distinct types of foot and mouth virus, differing in their infectivity, have been found (Vallee and Carre 1922, Waldmann and Trautwein 1926). Several immunologically distinct types of poliomyelitis have now been differentiated, some of these are so sharply defined that the serum of monkeys convalescent from infection with one type will not protect against another (see Sabin 1941).

The existence of a group antigenic relationship such as that demonstrated between the viruses of influenza and swine influenza (see Chapter 74) is again reminiscent of the antigenic morphology of bacteria.

The inoculation of many viruses into suitable animals is followed by the appearance of neutralizing antibodies in the serum. The mode of action of these antibodies forms a constant subject of discussion. Whether they destroy the virus, whether they merely inactivate it, whether they sensitize it, or whether they fail even to combine with it, is still not known with certainty. Most of the evidence seems to be in favour of the occurrence of a slow union between virus and antibody leading to sensitization or actual destruction (see Chapter 55).

The type of antibody called forth by different viruses varies. The only antibody to poliomyelitis virus of which we have knowledge is a neutralizing antibody. On the other hand, vaccinia virus stimulates the production of agglutinins, precipitins, and complement-fixing bodies as well as neutralizing antibodies. In some instances, as in the vaccinia and lymphocytic choriomeningitis viruses, the neutralizing antibodies appear to be distinct from the other antibodies (Salaman 1937, Smadel *et al* 1940b), and we are still ignorant of the nature of the antigen against which the neutralizing antibody is active. In general, neutralizing antibodies are of most help in revealing antigenic affinities and differences between

closely related viruses, but there are exceptions. The relationship, for example, between the Eastern and Western types of *equine encephalomyelitis* viruses is said to be brought out by the complement fixation, but not by the neutralization test (Havens *et al* 1943).

**Pathogenicity**—The pathogenicity of different viruses for different hosts varies greatly. Some, like the *vaccinia* and the *rabies* virus, have a wide range of pathogenicity, others, like the foot and mouth and the *encephalitis* viruses have a narrow range, and still others, like the *measles* and *mumps* viruses seem to be pathogenic for one species alone. Some attack only mammals, some only birds, and some both mammals and birds. One curious feature of most of the viruses is their ability to grow in the embryonic or associated cells of the developing hen's egg, in spite of the fact that the chicken, once it is hatched is resistant to all the pure mammalian viruses. In becoming adapted to different hosts viruses often undergo minor variations, there is evidence for example that the numerous animal poxes, with the possible exception of fowl pox, are due to varieties of one and the same virus (Zwick 1924). On the other hand, the properties of a virus may be considerably altered. Thus inoculation of the calf with *variola* virus and subsequent transference by passage through calves, modifies the virus in such a way that when reinoculated into human beings it gives rise not to smallpox but to *vaccinia*. Passage of the street virus of *rabies* through the brain of rabbits gives rise to the production of a fixed virus which, though it kills rabbits on intra cerebral inoculation more rapidly than the street virus, is yet much less virulent than the street virus on subcutaneous inoculation (Levaditi *et al* 1924). This example illustrates another characteristic that is frequently observed in the study of viruses, namely, their adaptation not only to one particular host but to one special tissue or route of inoculation. Findlay (1936), whose review on variation in the animal viruses should be consulted, is of opinion that variants are of two types. (a) variants associated with pathological lesions unlike those produced by the parent strains, but without any great antigenic difference. (b) variants associated with pathological lesions like those produced by the parent strains but with considerable antigenic difference, more often quantitative than qualitative in character. Some changes, such as the conversion of rabbit fibroma into myxoma, may justly be regarded as mutations, the majority are to be regarded as changes of the environmental type.

Infection in most of the filtrable virus diseases appears to occur by direct contagion, the infective material gaining access to the body either by the nasopharynx or sometimes by the skin as in *rabies*. In certain diseases the virus is inoculated into the blood stream by an insect vector, yellow fever, for example, is carried by the mosquito *Aedes aegypti*, dengue fever by *Aedes aegypti* and *Aedes albopictus*, and Pappataci fever by the sandfly, *Phlebotomus papatasi*. Laboratory infections are not infrequent, especially with yellow fever, psittacosis, Rift Valley fever and louping ill. Little is yet known about the mechanism of infection, but studies of the *vaccinia* virus suggest strongly that the elementary bodies constitute the infecting agents, and that in virulent strains even one elementary body may suffice to cause infection (Parker 1933, Parker *et al* 1941, Smadel *et al* 1939).

In general, the cellular lesions produced by viruses are of one or other of two kinds. Either the cells are stimulated so that the tissues become hyperplastic as in the fibromata, papillomata, warts and poxes, or they may be damaged so severely that they die, as in the hepatic necrosis of yellow fever and Rift

Valley fever. A preliminary hyperplasia may be succeeded as in the pores, by necrosis.

**The Nature of Viruses.**—Up till a few years ago the evidence was becoming increasingly stronger that the animal viruses were essentially minute micro-organisms. Microscopical filtration and centrifugal observations left little doubt that they consisted of relatively large particles which in each virus, were of fairly uniform size. The agreement between the results of these three methods of examination was, in fact, so good as to make it necessary to conclude that, if the visible coccoïd bodies were not the infective units of the virus then the virus must consist of particles of the same size which for some reason or other were invisible either by direct observation or by ultra violet photography (Dale 1930). In their morphological appearance in their formation of discrete colonies under suitable conditions—as on the chorio-allantoic membrane of the developing chick embryo—in their complex antigenic structure in their ability to stimulate the production of different types of antibody in their pathogenicity and their selective tissue localization in their capacity for variation, in their neutralizability by specific antiserum and in their power to give rise to immunity to fresh infection, some of the better studied viruses were apart from their small size and their failure to grow on lifeless media indistinguishable from ordinary bacteria.

In 1930 however just when this view was gaining general acceptance, Stanley in the United States made the startling announcement that he had succeeded in crystallizing the tobacco mosaic virus. From subsequent work carried out in the United States and in Great Britain (see Stanley 1930a b c d 1941 Bawden 1943) it became clear that this virus, which produces a disease in the Turkish tobacco plant, apparently consists of large nucleoprotein molecules about ten times as long as they are wide having a molecular weight of about 17 million, and capable of fitting together lengthwise to form needle-shaped crystals 20–30  $\mu$  long. The high molecular weight of this substance—greater than even the largest hæmocyanin molecules—its ability to produce disease in a quantity as small as one thousand millionth of a gram combined with the almost perfect parallelism between the amount of protein estimated chemically and the virus activity estimated biologically set it apart from any other known protein. It was further shown that variants of this virus had a slightly different chemical structure from the parent virus and that chemical alterations brought about in the molecule by laboratory methods led to changes in its pathogenic properties.

The shock produced by Stanley's discovery was somewhat mitigated when it was pointed out (Takahashi and Rawlins 1935 Bawden *et al.* 1936 Bernal and Fankuchen 1937) that the crystals were not true crystals but two-dimensional liquid crystals formed as the result of linear aggregation of smaller thread like bodies showing anisotropy of flow. Under these conditions it did not appear necessary to abandon the conception of viruses as living micro-organisms merely because some of them could arrange themselves in orderly rows (Andrews 1937). When it was shown however by Bawden and Pirie (1937) that the bull's head virus of tomatoes formed typical three-dimensional dodecahedral crystals, apparently composed of spherical isotropic particles this way of escape from the dilemma proposed by Stanley looked less promising.

How are we to reconcile the complex bacteria like structure and behaviour of the larger animal viruses with the apparently pure nucleoprotein molecules of the plant viruses? The explanation favoured by many workers (Green 1930



1938, Landlaw 1938, Gortner 1938) is that the viruses represent decadent forms of organisms that have become progressively degraded through long persistent parasitism. At the upper end of the scale are representatives such as the psittacosis and the vaccinia viruses that contain protein, fat and carbohydrate and that differ from bacteria chiefly in their loss of power to synthesize some factor or factors essential for their growth and multiplication. At the lower end of the scale are representatives, such as the tobacco mosaic virus that consist of pure nucleoprotein, and that are entirely dependent on living cells for all their ferment and autotrophic activities—inert chemical complexes which become living only when bathed in functioning protoplasm. This view has something to be said for it: it serves to reconcile the apparent break between the living and the non living. It is, however, not without difficulties of its own. Since the tobacco mosaic virus can grow in a number of plants, belonging even to different families in which the composition of the cell protoplasm is known to be different, it follows that its huge nucleoprotein molecule must be built up from relatively simple chemical substances. This presupposes a degree of synthetic organizing ability hitherto associated with living cells alone. Acceptance of this view opens up once again the whole question of spontaneous generation. If a protein molecule can in contact with living matter, reproduce itself and undergo variations each of which is attended by specific biological changes and each of which is genetically transmissible, it is difficult to avoid the conclusion that new bodies presenting the characters of living matter, must be constantly appearing in Nature. How also are we to regard structure in relation to size? The foot and mouth virus for example, which is about 12  $m\mu$  in diameter is so similar in almost every respect to the virus of vesicular stomatitis which has a particle size of 85  $m\mu$  that it is difficult to avoid the conclusion that they are structurally alike. Yet the foot and mouth virus is smaller—at least in one diameter—than the tobacco mosaic virus, the length of which is estimated to be about 30  $m\mu$ . Must we assume that the foot-and-mouth virus which incidentally has room for only about ten protein molecules, is organized like one of the larger animal viruses in spite of its being smaller than a plant virus which is known to consist of a single macromolecule?

One thing is clear. Before we can reach any conclusion on the nature of filtrable viruses, we shall have to re-define our terms. At the larger protein level the words "living" and "non living" have lost their conventional meaning (see Pirie 1937). It is difficult, even in Science, to avoid the common solecism of attempting to force new facts into a conception that has no reality as such, but has been formed merely as an abstraction from other previously known facts—and it is time for us to realize that our concept of 'life' is too crude to be used in relation to the infinitely small. Whatever the viruses are—micro-organisms reproducing themselves by binary fission, huge nucleoprotein molecules multiplying by autocatalysis or something else still—there is no doubt that they present us at the moment with one of the most fascinating and fundamental problems of the biochemical world.

We have said nothing in this chapter about the filtrable tumours. Here again we are on very difficult ground. There is evidence suggesting that virus particles are essential to their reproduction and that tumours both of avian and mammalian origin are caused to some extent at least, by infecting particles having many of the characteristics of the known filtrable viruses (see Gye 1925, Andrewes 1934, Ledingham and Gye 1935, and Chapter 89).

## REFERENCES

- AMIES C P (1933) *Lancet* 1 1015 (1934) *Brit J exp Path* 15 180  
 ANDREWES C H (1929) *Brit J exp Path.*, 10 188 (1934) *Lancet* : 63 117 (1935)  
*Proc roy Soc Med* 31, '03 (1942) *Brit J exp Path.*, 23, '14  
 ASHLEHOV I (1933) *J Bact* 25 323 339  
 BARNARD J E. (1925) *Lancet* 117 (1935) *Brit J exp Path* 16, 129 (1937)  
*Proc roy Soc. B.*, 124, 10"  
 BARNARD J E. and ELFORD W J (1931) *Proc roy Soc B* 109 360  
 BAUER, J H and PICKELS F G (1936) *J exp Med.*, 64, 503 (1937) *Id.*, 65 565  
 BAWDEN F C (1943) *Plant Viruses and Virus Diseases* 2nd edn. Chronica Botanica  
 Co Waltham Mass.  
 BAWDEN F C and PIRIE W (1938) *Brit J exp Path* 19 251  
 BAWDEN F C PIRIE, W BERNAL, J D and FANKUCHEN I. (1936) *Nature, Lond.*,  
 138 1051  
 BEARD J W FINKELSTEIN H and WYCKOFF R. W G (1933) *J Immunol.* 35, 415  
 BECHHOLD H and SCHLESINGER, M (1931) *Biochem Z* 238, 387  
 BEDSON S P and BLAND J O W (193 ) *Brit J exp Path.*, 13, 461 (1934) *Ibid.*, 15  
 '43  
 BERNAL, J D and FANKUCHEN I. (1937) *Nature Lond.*, 139 9-3.  
 BLAND J O W and POWISOW C. F (1939) *J Path Bact* 48, 331  
 BURNET F M (1936) *Spec. Pep Ser med Res Coun* No. 220  
 BURNET F M and FARIS D D (1942) *J Bact* 44, '41  
 CARREL, A. and RIVERS T M (1927) *C P Soc. Bd* 96, 848  
 CHEN W K. (1934) *Proc Soc exp Biol. N Y.*, 32, 491  
 CLAIBERG K W (1939) *Klin Wochr* 18, 632.  
 COWDREY E V (1933) *Amer J Path* 9 149  
 COX H R and HYDE, P P (1937) *Amer J Hyg* 18, 667  
 CRACUT E. C. and OFFENHEIMER E. H (1926) *J exp Med.*, 43, 815  
 CRAIGIE J (1937) *Brit J exp Path.*, 13, '39 (1935) *J Immunol.*, 29 '70.  
 CRAIGIE, J and WISHART F O (1934a) *Brit J exp Path.*, 15 390 (1934b) *Trans roy  
 Soc Can.*, Sect on 91 (1936) *J exp Med* 64, 803 819 (1935) *J Bact* 35 2.  
 DALE H H (1935) Huxley Memorial Lecture Macmillan & Co., Ltd., London.  
 DOUGLAS, S P and SMITH W (1928) *Brit J exp Path* 9 '13.  
 DUCLAUX J and AMAT M (1938) *C R Acad Sc* 206, 145  
 DUNHAM W B. and MACNEAL, W J (1942) *J Bact* 44, 413  
 EAGLES, G H (1935) *Brit J exp Path* 16, 188  
 EAGLES, G H and McCLEAN D (1931) *Brit J exp Path.* 12, 97  
 EISENBERG-MERLING K B (1943) *Brit J exp Path.*, 24 '40  
 ELFORD W J (1931) *J Path Bact.*, 34, 505 (1933) *Proc roy Soc. B* 112, 324  
 (1936) *Brit J exp Path* 17 399  
 ELFORD W J GRABAR, P., and FERRY J D (1935) *Brit J exp Path.*, 16, 583  
 FELLER, A E., ENDERS J F and WELLES, T H (1940) *J exp Med* '2, 36  
 FINDLAY G M (1928) *Brit J exp Path* 9 28 (1930) *Brit J exp Path* 11, 109  
 (1936) *J R microscop Soc.*, 56, '13 (1939) *Brit med J* 1 25  
 FINDLAY G M and LUDFORD R. J (1936) *Brit J exp Path* 7 '23  
 FINDLAY G M and MACCALLUM, F O (1940) *Lancet* ii 163.  
 FRANCIS T and SALK, J E (194 ) *Science* 96 499  
 GALLOWAY I. A and ELFORD W J (1931) *Brit J exp Path.*, 12, 40  
 GASTINEL, P and REILLY J (1928) *Bull. Med.*, 42, 839  
 GINS H A and KRAUSE, C. (1923) *Ergebn. allg Path path. Anat.* 20 : 805  
 GOODPASTURE, E. W (1929) *Arch Path.*, 7 114 (1929-30) *Harvey Lectures* (1930)  
*Zbl Ges Neurol Psych* at 129 599  
 GORDON M H. (1925) *Spec Pep Ser med Res Coun. Lond.*, No. 98  
 GORTNER, R. A (1938) *Science* 87 579  
 GREEN R. G (1935) *Science* 82, 443 (1935) *Biodynamica*, No. 39 1  
 GREEN R H ANDERSON T F., and SMADEL, J E. (1942) *J exp Med.*, '5 651  
 GRIFFELL, F B (1929) *J Bact.*, 18, 15  
 GYK, W E (1925) *Lancet* ii 109  
 HAAGEN E. (1925) *Zbl Bakt.* 109 31 (1939) *Ibid* 143, 283  
 HALLAUER C. (1938) Doerr and Hallauer's *Handbuch der Virusforschung* : 369  
 HAYES W P WATSON D W., GREEN R. H LAVIN G I and SMADEL, J E. (1943)  
*J exp Med* '7 139  
 HENRIOT E. and HOUENARD E. (1925) *C R Acad Sc.*, 180 1369 (1927) *J Phys  
 Pod um* 8, 433  
 HERRING K. (1933) *Z Immunforsch* 80 507 (1936) *Zbl Bakt.*, 138, 25  
 HIMMELWEIT F (1938) *Brit J exp Path.*, 19 105.

- HINDLE, E and FINDLAY, G M (1930) *Brit J exp Path*, 11, 131  
 HOAGLAND, C L, SMADEL, J E, and RIVERS, T M (1940) *J exp Med*, 71, 377  
 HOAGLAND, C L, WARD, S M, SMADEL, J E, and RIVERS, T M. (1941) *J exp Med*, 74, 69  
 HUGHES, T P, PARKER, R F, and RIVERS, T M (1935) *J exp Med*, 62, 349  
 KNIGHT, C A and STANLEY, W M (1944) *J exp Med*, 79, 291  
 KUROTSCHKIN, T J (1939) *Proc Soc. exp Biol*, N Y, 41, 407  
 LAIDLAW, P P (1938) 'Virus Diseases and Viruses' Camb Univ Press  
 LEA, D E and SALAMAN, W H (1942) *Brit J exp Path*, 23, 27  
 LEDINGHAM, J C G (1924) *Brit J exp Path*, 5, 332, (1931) *Lancet*, ii 525 (1932) *Brit med J*, ii 953, (1935) *Johns Hopkins Hosp Bull*, 56, 247, 337, *Ibid*, 57, 32  
 LEDINGHAM, J C G and GYE, W E (1935) *Lancet*, i 376  
 LÉPINE, P, SAUTTER, V, and KREIS, B (1937) *C R Soc Biol*, 124, 422  
 LEVADITI, C and NICOLAU, S (1923) *C R Soc Biol*, 88, 66  
 LEVADITI, C, NICOLAU, S, and SCHOEN, R (1924) *C R Soc Biol*, 91, 423  
 LEWIS, M R and ANDERSON, H B (1927) *Amer J Hyg*, 7, 505  
 LIPSCHÜTZ, B (1925) *Zbl Bakt*, 96, 222  
 LURIA, S (1940) *Ann Inst Pasteur*, 64, 415  
 LURIA, S E, DELBRÜCK, M, and ANDERSON, T F (1943) *J Bact* 46, 57  
 MACCALLUM, F O and FINDLAY, G M (1938) *Lancet* ii 136  
 McCLEAN, D and EAGLES, G H (1931) *Brit J exp Path* 12, 103  
 MACFARLANE, A S, MACFARLANE M G, AMES, C R, and EAGLES, G H (1939) *Brit J exp Path*, 20, 485  
 MACFARLANE, M G and DOLBY, D E (1940) *Brit J exp Path*, 21, 219  
 MACFARLANE M G and SALAMAN, M H. (1938) *Brit J exp Path* 19, 184  
 McINTOSH, J (1935) *J Path Bact*, 41, 215  
 McINTOSH, J and SELMIE, F R (1937) *Brit J exp Path*, 18, 162 (1940) *Ibid*, 21, 133  
 MAITLAND, H B and LAING, A W (1941) *J Path Bact*, 53, 419  
 MAITLAND, H B, LAING, A W, and LYTH, R (1932) *Brit J exp Path* 13, 90  
 MAITLAND, H B and MAITLAND, M C (1928) *Lancet*, ii 596.  
 MARIE, A C and URBAN, A. (1930) *C R Soc Biol*, 103, 866  
 MARRHAM, R, SMITH, K M and LEA, D (1942) *Parasitology* 34, 315  
 MARTON, L. (1941) *J Bact*, 41, 397  
 MCDON, S (1922-3) *Amer J Physiol*, 63, 429, (1928) "Filterable Viruses" T M Rivers p 55 Baulhière, Tindall & Cox, London  
 NATARAJAN, C V and HYDE R R (1930) *Amer J Hyg*, 11, 652  
 NOGUCHI, H (1915) *J exp Med*, 21, 539, (1918) *Ibid*, 27, 425  
 OOSTON, A. (1931) *Brit med J*, i 369  
 OLITSKY, P K, and BOËZ, L. (1927) *J exp Med*, 45, 685  
 PARKER, F. and NYE, R V (1925) *Amer J Path*, 1, 325, 337  
 PARKER, R F (1939) *J exp Med* 67, 725  
 PARKER, R F, BRONSON, L H, and GREEN, R H (1941) *J exp Med*, 74, 963  
 PARKER, R F and RIVERS, T M (1937) *J exp Med*, 65, 243  
 PARKER, R F and SMITHE C V (1937) *J exp Med*, 65, 109  
 PASCHKE, E (1932) *Zbl Bakt*, 124, 89  
 PERDRAU, J R (1927) *Brit J exp Path*, 8, 167  
 PERDRAU, J R and TONN, C (1933) *Proc roy Soc. B*, 112, 288, (1936) *Ibid* 121, 253  
 PIRIE, N W (1937) "Perspectives in Biochemistry," Camb Univ Press  
 POPPE, K. and BUSCH, G (1930) *Z Immunforsch*, 68, 510  
 RAKE, G, SHAFFER M I, and THYGESON, P (1942) *Proc Soc exp Biol*, A 1 49, 545  
 Report (1927) 2nd Progr Rep Foot and Mouth Dis Res Comm Ministry Agric Fish, Lond  
 REYNOLDS, F D (1928) *J. exp Med*, 47, 389  
 RIVERS, T M (1928) 'Filterable Viruses' p 3 Baulhière, Tindall & Cox London  
 (1932) *Physiol Rev* 12, 423, (1943) 'Virus Diseases' p 3 Cornell Univ Press Ithaca NY  
 RIVERS, T M, HAAOEN, E, and MCCREYNOLDS, R S (1929) *J exp Med*, 50, 665  
 RIVERS, T M and PEARCE L. (1925) *J exp Med*, 42, 523  
 RIVERS, T M and WARD, S M (1933) *J exp Med*, 57, 51, 741  
 RODANICHE, E C (1942) *J infect Dis*, 70, 58  
 SABIN, A B (1941) *J Amer med Ass*, 117, 267  
 SALAMAN, M H (1934) *Brit J exp Path*, 15, 351, (1937) *Ibid*, 18, 245  
 SANKARAN, G, IYENGAR K R K, and BEER, W A (1934) *Indian J med Res*, 21, 909  
 SAWYER, W A. and FROBISHER, M (1929) *J exp Med*, 50, 713

- SAWYER, W. A., LLOYD, W. D. M., and KITCHEN, S. F. (1929) *J exp Med.*, 50, 1.  
 SCHLESINGER, M. (1935) *Nature*, 133, 549.  
 SHARP, D. G., TAYLOR, A. R., BEARD, D., and BEARD, J. W. (1942) *Proc Soc exp Biol.*, 1 J., 50, 205. (1943) *Arch Path.*, 38, 167.  
 SHARP, D. G., TAYLOR, A. R., McLEAN, I. W., BEARD, D., BEARD, J. W., FELLER, A. E., and DINGLE, J. H. (1944) *J Immunol.*, 48, 129.  
 SHEDLOVSKY, T., POTHEV, A., and SMADEL, J. E. (1943) *J exp Med.*, 77, 155.  
 SHEDLOVSKY, T. and SMADEL, J. E. (1942) *J exp. Med.*, 75, 162.  
 SHORTT, H. E. and BROOKS, A. G. (1934) *Indian J med Res.*, 21, 531.  
 SICHERT MODROW, I. (1930) *Zbl Bakt.*, 119, 12.  
 SMADEL, J. E. and HOAGLAND, C. L. (1942) *Bact Rev.*, 6, 79.  
 SMADEL, J. E., HOAGLAND, C. L., and SHEDLOVSKY, T. (1943) *J exp. Med.*, 77, 165.  
 SMADEL, J. E., LAVIN, G. I., and DUBOS, R. J. (1940a) *J exp Med.*, 71, 373.  
 SMADEL, J. E., WALL, M. J., and BAKER, R. D. (1940b) *Ibid.*, 71, 43.  
 SMADEL, J. E., PICKELS, E. G., and SHEDLOVSKY, T. (1938) *J exp Med.*, 68, 607.  
 SMADEL, J. E., PIVERS, T. M., and HOAGLAND, C. L. (1942) *Arch. Path.*, 34, 275.  
 SMADEL, J. E., PIVERS, T. M., and PICKELS, E. G. (1939) *J exp Med.*, 70, 379.  
 SMITH, W. (1932) *Brit J exp Path.*, 13, 434. (1939) *J Path Bact.*, 48, 557.  
 SOBERNHEIM, G. (1925) *Ergebn Hyg.*, 7, 133.  
 STANLEY, W. M. (1938a) *J phys Chem.*, 42, 55. (1938b) *Bull New York Acad Med.*, 14, 398. (1938c) *J appl. Phys.*, 9, 148. (1938d) *Amer Nat.*, 72, 110. (1941) "Virus Diseases," p. 33. Cornell Univ Press, Ithaca, N Y.  
 STANLEY, W. M. and ANDERSON, T. F. (1941) *J biol Chem.*, 139, 322.  
 STEVENSON, W. D. H. and BUTLER, C. G. (1939) *Rep publ. Hlth med. Subj., Min Hlth, Lond.*, No. 87.  
 STOCK, C. C. and FRANCIS, T. (1940) *J exp Med.*, 71, 661.  
 SVEDBERG, T. (1937) *Nature, Lond.*, 129, Suppl., 1051.  
 SVEDBERG, T., BOESTAD, G., and ERIKSSON QUENSTEDT, I. B. (1934) *Nature*, 134, 98.  
 SVEDBERG, T. and NICHOLS, J. B. (1927) *J Amer chem. Soc.*, 49, 2920.  
 TAKAHASHI, W. and RAWLINS, T. E. (1930) *Science*, 81, 299.  
 TALLERMAN, K. H. (1929) *Brit J exp Path.*, 10, 360.  
 TANG, F. F. (1932) *J Bact.*, 24, 133.  
 TANG, F. F. and WEI, H. (1937) *J Path. Bact.*, 45, 317.  
 TAYLOR, A. P., SHARP, D. G., BEARD, D., BEARD, J. W., DINGLE, J. H., and FELLER, A. E. (1943) *J Immunol.*, 47, 261.  
 TODD, C. (1928) *Brit J exp Path.*, 9, 19.  
 VALLÉE, H. and CARRÉ, H. (1929) *C P Acad Sci.*, 174, 1498.  
 WALDMAN, O. and TRAUTWEIN, K. (1926) *Berl. tierarztl. Wochr.*, 42, 569.  
 WARD, H. K. (1929) *J exp Med.*, 50, 31.  
 WOODRUFF, A. M. and GOODPASTURE, E. W. (1931) *Amer J Path.*, 7, 209.  
 WOODRUFF, C. E. and GOODPASTURE, E. W. (1929) *Amer J Path.*, 5, 1. (1930) *Ibid.*, 6, 713.  
 WOOLEY, J. G. (1939) *Publ. Hlth Rep. Wash.*, 54, 1077.  
 ZINSSER, H. and SCHÖENBECH, E. B. (1937) *J exp Med.*, 66, 207.  
 ZINSSER, H. and SEASTOVE, C. V. (1930) *J Immunol.*, 18, 1.  
 ZINSSER, H. and TANG, F. (1929) *J Immunol.*, 17, 343.  
 ZWICK. (1924) *Dtsch. tierarztl. Wochr.*, 32, 643.

# INDEX

All organisms are listed under the generic names given in the Classificatory Chart on p 319. Since many rod shaped organisms have in the past been known under the generic name of *Bacillus*, irrespective of whether or not they belonged to this genus, we have for the convenience of the reader included most of them under the heading *Bacillus* (non italicized). This heading should therefore be consulted by those who are doubtful to what genus a given organism belongs.

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